

SUPPORT INFORMATION

Novel Ibuprofenate- and Docusate-Based Ionic Liquids: Emergence of Antimicrobial Activity

Clarissa P. Frizzo*, Keli Wust, Aniele Z. Tier, Thaíssa Beck, Letícia V. Rodrigues, Rodrigo A. Vaucher, Leandro Bolzan, Silvio Terra, Felix Soares, and Marcos A.P. Martins

*e-mail: clarissa.frizzo@gmail.com

Index

Table S1. Molecular formula, molecular weight, monoisotopic mass and mass spectrometry data^a for the IL-APIs.

Table S2. Solubility of IL-APIs and starting materials in water ^a

Table S3. Values of inhibition halos (mm)^a for Ionic Liquids and starting materials.

Table S4. Values of inhibition halos (mm)^a for Ionic Liquids and starting materials.

Table S5. Values of inhibition halos (mm)^a for Ionic Liquids and starting materials.

Table S6. MIC^a (mM) values of antifungal assay of IL-API derivative of ibuprofenate anion.

Table S7. MIC^a (mM) values of antifungal assay of IL-API derivative of docusate anion.

Table S8. MIC^a (mM) values of antibacterial assay of IL-API.

Table S9. Total antioxidant capacity (%) of the compounds by phosphomolybdenum method presented as mean ± S.E.M.

Table S1. Molecular formula, molecular weight, monoisotopic mass and mass spectrometry data^a for the IL-APIs.

IL-API	Molecular Formula	Molecular Mass (g Mol ⁻¹)	Monoisotopic Mass (g Mol ⁻¹)	Molecular Ion (m/z) ^b	Molecular Ion (m/z) ^c
[Ran][Ibu]	C ₂₆ H ₄₀ N ₄ O ₅ S	520.68	520.68	315.2	205.1
[Bup][Ibu]	C ₃₁ H ₄₆ N ₂ O ₃	494.71	494.35	289.3	205.1
[Lid][Ibu]	C ₂₇ H ₄₀ N ₂ O ₃	440.62	440.30	235.2	205.1
[Dif][Ibu]	C ₃₀ H ₃₉ NO ₃	461.64	461.29	256.2	205.1
[Bup][Doc]	C ₃₈ H ₆₆ N ₂ O ₈ S	711.00	710.45	289.3	421.2
[Gli][Doc]	C ₂₂ H ₄₃ NO ₉ S	497.64	497.27	74.1	421.2
[EGli][Doc]	C ₂₄ H ₄₇ NO ₉ S	525.70	525.30	104.1	421.2
[Dif][Doc]	C ₃₇ H ₅₉ NO ₈ S	677.63	677.40	256.2	421.2
[Ran][Sulf]	C ₂₁ H ₃₂ N ₆ O ₆ S ₂	528.65	528.18	315.2	213.00

^aData from Mass Spectrometer Agilent 6460 Triple Quadrupole 6460 (LC-MS/MS) ^bPositive Mode. ^cNegative Mode.

Table S2. Solubility of IL-APIs and starting materials in water ^a.

IL-API	Solubility (mg·mL ⁻¹) (± Sdev)	API precursor	Solubility (mg·mL ⁻¹) (± Sdev)
[Ran][Ibu]	145 ± 7	[Na][Ibu] ^b	140 ± 10
[Dip][Ibu]	30± 0	[Ran][Cl]	20± 0
[Gly][Doc]	65 ± 7	[EGly][Cl]	490 ± 14
[EGly][Doc]	20 ± 0	[Gly][Cl]	520 ± 21
[Dip][Doc]	10± 0	[Lid][Cl]	395 ± 21
[Ran][Sulf]	30± 0	[Dip][Cl]	320 ± 10

^aThe solubility of the solute was determined by the “flask method”¹ in triplicate. ^bThe solubility of the ibuprofen free acid is 0.02–0.08 mg·mL⁻¹.²

Table S3. Values of inhibition halos (mm)^a for Ionic Liquids and starting materials.

Fungi	[Dip][Ibu]	[Ran][Ibu]	[Ibu][Na]	[Dip][Cl]
<i>C. kefyr</i>	14	10	9	24
<i>C. guilhermondii</i>	18	-	12	18
<i>C. albicans</i>	14	8	10	12
<i>C. dubliniensis</i>	20	10	10	18
<i>C. glabrata</i>	14	8	9	12
<i>C. parapsilosis</i>	12	-	13	10
<i>C. lusitaneae</i>	14	-	-	14
<i>C. krusei (clinical isolated)</i>	12	12	-	14
<i>C. tropicalis</i>	14	-	-	10
<i>C. catenulata</i>	14	-	-	14
<i>C. geocharles</i>	20	-	-	-
<i>Cryptococcus neoformans</i>	-	12	19	-

^aThe inhibition halos values were obtained by using disk diffusion technique as culture broth Mueller Hinton according to CLSI M44-A2 protocol (2008). As a control of inhibition was used 100 µg of Amphotericin B (Biorard) having greater than 10 mm halo. Considered susceptible by the CLSI (2008). ^b[Ran][Cl] was not active against any tested fungus.

Table S4. Values of inhibition halos (mm)^a for Ionic Liquids and starting materials.

Fungi	[Gly][Doc]	[EGly][Doc]	[Dip][Doc]	[Na][Doc]	[Dip][Cl]
<i>C. kefyr</i> ^b	24	19	18	12	24
<i>C. guilhermondii</i>	12	9	10	-	18
<i>C. albicans</i>	20	9	18	11	12
<i>C. membrana faciens</i>	-	10	-	11	-
<i>C. dubliniensis</i>	10	9	-	11	18
<i>C. glabrata</i>	10	12	-	8	12
<i>C. parapsilosis</i>	12	-	12	-	10
<i>Cryptococcus neoformans</i>	-	12	-	12	-
<i>C. lusitaneae</i>	-	-	14	-	14
<i>C. krusei</i>	16	-	12	-	14
<i>C. tropicalis</i>	12	-	14	-	10
<i>C. catenulata</i>	20	-	18	-	14
<i>C. geocharles</i>	10	-	8	-	-

^aThe inhibition halos values were obtained by using disk diffusion technique as culture broth Mueller Hinton according to CLSI M44-A2 protocol (2008). As a control of inhibition was used 100 µg of Amphotericin B (Biorard) having greater than 10 mm halo. Considered susceptible by the CLSI (2008). ^b[EGly][Cl] was active only against *C. kefyr* with a inhibition halos of 10 mm. ^c[Gly][Cl] was not active.

Table S5. Values of inhibition halos (mm)^a for Ionic Liquids and starting materials.

Bacteria	[Dip][Ibu]	[Egly][Doc]	[Dip][Doc]	[Ibu][Ran]	[Ibu][Na]	[Dip][Cl]	[Doc][Na]
<i>Enterobacter aerogenes</i> ATCC 13048	12	8	-	-	-	-	-
<i>Escherichia coli</i> ATCC 25922	16	-	-	-	-	-	-
<i>Shigella boydii</i> sorotipo 10 NCTC 9358 ATCC-IAL 07199	14	-	-	9	18	-	-
<i>A. baumannii</i> ATCC 19606	-	-	10	-	-	-	-
<i>Proteus mirabilis</i> ATCC 25933	16	-	-	-	10	-	-
<i>Pseudomonas Aeruginosa</i> PNQC proex 340	10	-	-	-	10	-	-
<i>Proteus vulgaris</i> ATCC 39882	14	-	-	-	-	-	-
<i>Salmonella typhimurium</i> ATCC14028	16	-	-	-	-	-	-
<i>Escherichia coli</i> ATCC35218	10	-	-	-	12	-	-
<i>Escherichia coli</i> ATCC8739	14	-	-	-	10	-	-
<i>Streptococcus sp</i> (isolado clínico)	-	10	-	-	-	-	12
<i>Staphylococcus aureus</i> PNCQ	-	8	-	-	-	-	-
<i>Staphylococcus aureus</i> ATCC 00039	-	12	-	10	-	-	10
<i>S. sonneii</i> (clinical isolate)	-	-	-	10	12	-	-
<i>Listeria monocytogenes</i> ATCC 7644	-	10	-	-	-	-	12
<i>Critobacter freundii</i> ATCC 8090	-	-	12	-	-	10	-
<i>Klebsiella pneumoniae</i> ATCC 700603	-	-	10	-	-	-	-

^aThe inhibition halos values were obtained by using disk diffusion technique as Mueller Hinton broth culture medium according to the CLSI protocol (2012). As a positive growth control was used culture medium more microbial inoculum. As negative control for Gram positive bacteria was used doxycycline 30 µg showing greater than 16 mm halo. As a negative control for Gram negative bacteria was used Amikacin 30 µg presenting higher halo to 17 mm. Susceptible considered by the CLSI (2012).

Table S6. MIC^a (mM) values of antifungal assay of IL-API derivative of ibuprofenate anion.

Fungui	[Dip][Ibu]	[Ran][Ibu] ^b	[Ibu][Na]	[Dip][Cl]
<i>C. kefyr</i>	7	6	27	4
<i>C. guilhermondii</i>	7	-	14	4
<i>C. albicans</i>	3	12	27	17
<i>C. dubliniensis</i>	7	2	5	4
<i>C. glabrata</i>	7	12	27	17
<i>C. parapsilosis</i>	3	-	27	9
<i>C. lusitaneae</i>	13	-	-	17
<i>C. krusei</i>	7	5	-	7
<i>C. tropicalis</i>	3	-	-	7
<i>C. catenulata</i>	3	-	-	7
<i>C. geocharles</i>	2	-	-	-
<i>Cryptococcus neoformans</i>	-	0.4	2	

^aMIC values were obtained by microdilution using a culture medium Mueller Hinton broth according to the CLSI protocol 2010. Testing was performed in triplicate. As a positive control the growth culture medium over the microbial inoculum was used. How to control inhibition were used to Amphotericin B 100 µg (Biorard).^b [Ran][Cl] was not active against any tested fungus.

Table S7. MIC^a (mM) values of antifungal assay of IL-API derivative of docusate anion.

Fungi	[Gly][Doc] ^c	[EGly][Doc]	[Dip][Doc]	[Na][Doc]	[Dip][Cl]
<i>C. kefyr</i> ^b	11	0.17	8	0.2	4
<i>C. guilhermondii</i>	22	48	8	-	4
<i>C. albicans</i>	11	12	4	4	17
<i>C. membrana faciens</i>	-	24	-	28	-
<i>C. dubliniensis</i>	22	24	-	28	4
<i>C. glabrata</i>	10	3	-	28	17
<i>C. parapsilosis</i>	11	-	15	-	9
<i>Cryptococcus neoformans</i>	-	25	-	0.2	-
<i>C. lusitaneae</i>	11	-	-	12	17
<i>C. krusei</i>	6	-	-	23	9
<i>C. tropicalis</i>	11	-	-	23	9
<i>C. catenulata</i>	11	-	-	23	9
<i>C. geocharles</i>	11	-	-	47	-

^aMIC values were obtained by microdilution using a culture medium Mueller Hinton broth according to the CLSI protocol 2010. Testing was performed in triplicate. As a positive control the growth culture medium over the microbial inoculum was used. How to control inhibition Polymyxin 300 units were used to bactetrias Gram negative and Vancomycin 30 mg for Gram positive. ^b[EGly][Cl] was active only against *C. kefyr* with a MIC of 11 mM L⁻¹. ^c[Gly][Cl] was not active.

Table S8. MIC^a (mM) values of antibacterial assay of IL-API.

Bacteria		[Dip]	[Ibu]	[Egly]	[Doc]	[Ibu][Na]	[Dip][Cl]	[Doc][Na]
<i>Enterobacter aerogenes</i> ATCC 13048		13		24		-	-	-
<i>Escherichia coli</i> ATCC 25922		4		-		-	-	-
<i>Shigella boydii sorotipo</i> 10 NCTC 9358 ATCC-IAL 07199		7		-	14	4		
<i>Proteus mirabilis</i> ATCC 25933		13		-	55	17		
<i>Pseudomonas Aeruginosa</i> PNQC proex 340		7		-	-	9		
<i>Proteus vulgaris</i> ATCC 39882		13		-	-	-		
<i>Salmonella typhimurium</i> ATCC14028		7		-	-	-		
<i>Escherichia coli</i> ATCC35218		13		10	-	9		
<i>Escherichia coli</i> ATCC8739		7		-	-	17		
<i>Estreptococcus sp</i> (isolado clínico)		-		12	-	-		4
<i>Staphylococcus aureus</i> PNCQ		-		24	-	-		
<i>Staphylococcus aureus</i> ATCC 00039		-		3	-	-		4
<i>Listeria monocytogenes</i> ATCC 7644		-		12	-	-		4
<i>Critobacter freundii</i> ATCC 8090		-		10	-	17		
<i>Klebsiella pneumoniae</i> ATCC 700603		-		5	-	-		

^aMIC values were obtained by microdilution using a culture medium Mueller Hinton broth according to the CLSI protocol 2010. Testing was performed in triplicate. As a positive control the growth culture medium over the microbial inoculum was used. How to control inhibition Polymyxin 300 units were used to bactetrias Gram negative and Vancomycin 30 mg for Gram positive.

Table S9. Total antioxidant capacity (%) of the compounds by phosphomolybdenum method presented as mean ± S.E.M.

[] μM Compds	100	250	500	1000
[Na][Doc]	1.49 ± 0.6	1.61 ± 0.1	2.07 ± 1.1	1.30 ± 1.0
[Na][Ibu]	4.18 ± 3.3	0.87 ± 0.7	12.10 ± 6.0	54.18 ± 5.9
[Na][Sulf]	2.42 ± 0.5	1.41 ± 0.9	1.61 ± 0.6	3.91 ± 0.3
[Ran][Cl]	14.18 ± 0.9	17.06 ± 3.4	30.09 ± 1.1	17.12 ± 2.6
[Dip][Cl]	3.91 ± 1.6	2.53 ± 0.3	8.30 ± 1.0	7.72 ± 1.7
[EGly][Cl]	2.19 ± 0.8	2.07 ± 1.2	3.80 ± 2.9	2.88 ± 0.8
[Gly][Cl]	5.99 ± 2.9	5.30 ± 2.9	5.30 ± 0.4	3.68 ± 0.6
[Ran][Sulf]	14.06 ± 0.2	17.07 ± 3.1	29.28 ± 2.1	41.04 ± 2.3
[Ran][Ibu]	20.17 ± 0.5	21.21 ± 1.0	29.74 ± 4.5	22.25 ± 2.7
[EGly][Doc]	3.98 ± 1.9	3.11 ± 0.7	2.77 ± 0.5	6.45 ± 0.8
[Gly][Doc]	2.99 ± 1.5	8.53 ± 0.7	2.77 ± 0.3	40.81 ± 6.5

[Dip][Ibu]	4.03 ± 1.4	3.11 ± 1.1	5.76 ± 2.2	128.89 ± 12.8
[Dip][Doc]	2.65 ± 0.6	4.03 ± 2.1	12.10 ± 6.0	45.99 ± 12.1

Referencens

- 1 OECD, 1995. OECD Guideline for the testing of chemical: 105 Water solubility: Flask Method, Paris.
- 2 U. Domanska, A. Pobudkowska, A. Pelczarska P.J. Gierycz, *Phys. Chem., B.* 113. 2009, 8941-8947.