

## Electronic Supplementary Information

# A mitochondria-targeted near-infrared probe for colorimetric and ratiometric fluorescence detection of hypochlorite in living cells

Junchao Xu,<sup>ac</sup> Houqun Yuan,<sup>b</sup> Caiqin Qin,<sup>a</sup> Lintao Zeng<sup>\*ac</sup> and Guang-Ming Bao<sup>\*b</sup>

<sup>a</sup> Department of Chemistry and Materials Sciences, Hubei Engineering University, Xiaogan 432100, P.R. China. E-mail: zlt1981@126.com (L. Zeng).

<sup>b</sup> College of Animal Science and Technology, Jiangxi Agricultural University, Nanchang 330045, P.R. China. Email: bycb2005@gmail.com (G.-M. Bao).

<sup>c</sup> School of Chemistry and Chemical Engineering, Tianjin University of Technology, Tianjin 300384, P.R. China.

## Contents

1. <sup>1</sup>H NMR, <sup>13</sup>C NMR and HR-MS spectra of **CMBI**
2. Summary of photophysical properties
3. Ratiometric response of **CMBI** toward ClO<sup>-</sup> at low concentrations
4. Selectivity of **CMBI** toward ClO<sup>-</sup>
5. Kinetics research for **CMBI** towards ClO<sup>-</sup>, HSO<sub>3</sub><sup>-</sup> and HS<sup>-</sup>
6. Proposed sensing mechanism
7. Cell toxicity test of **CMBI**
8. Comparison of ratiometric fluorescent probes for ClO<sup>-</sup>
9. <sup>1</sup>H NMR spectra of compounds **1** and **2**

# 1. <sup>1</sup>H NMR, <sup>13</sup>C NMR and HR-MS spectra of CMBI

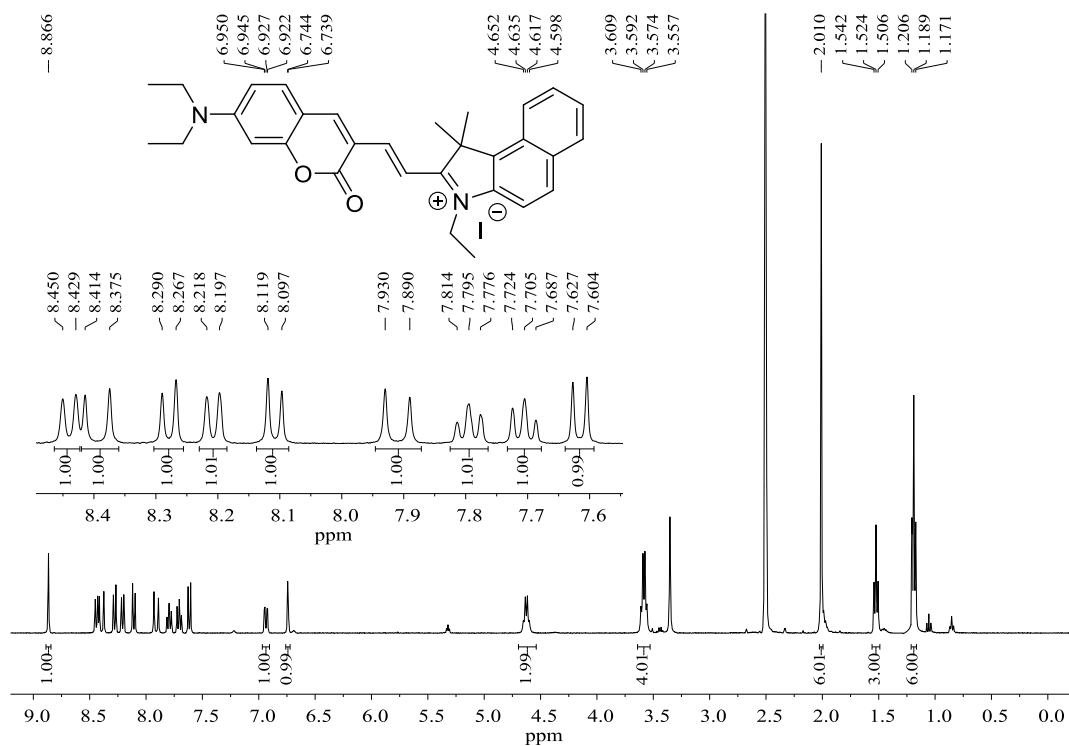


Fig. S1 <sup>1</sup>H NMR spectrum of CMBI in DMSO-*d*<sub>6</sub> (400 MHz).

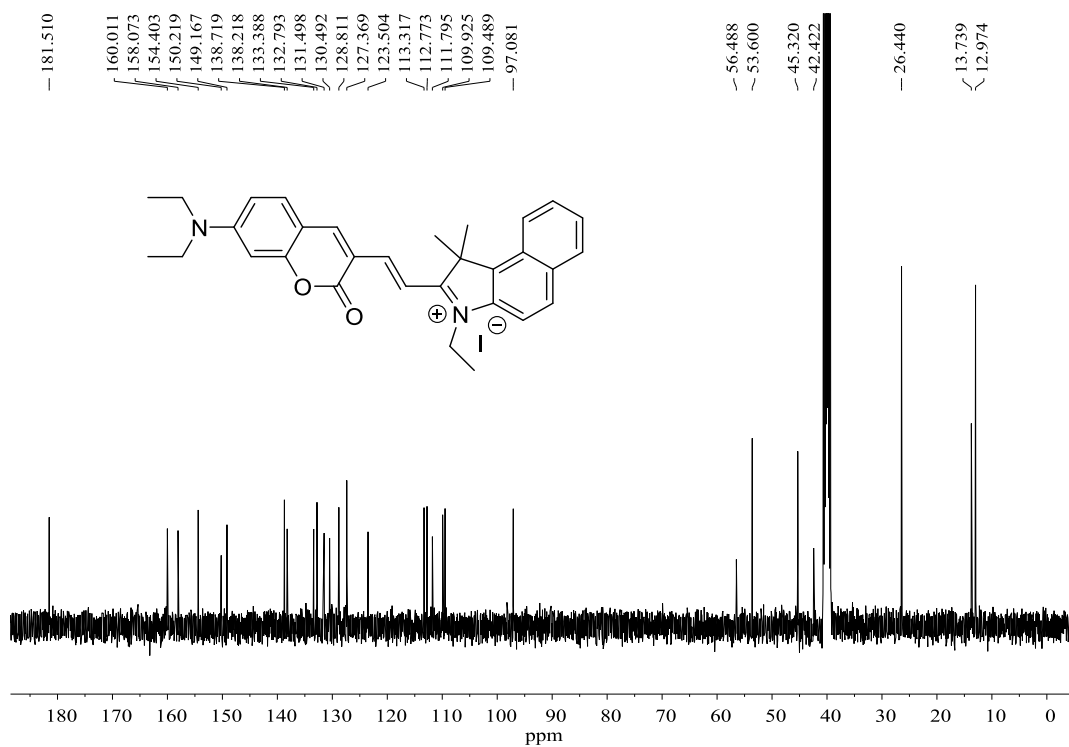
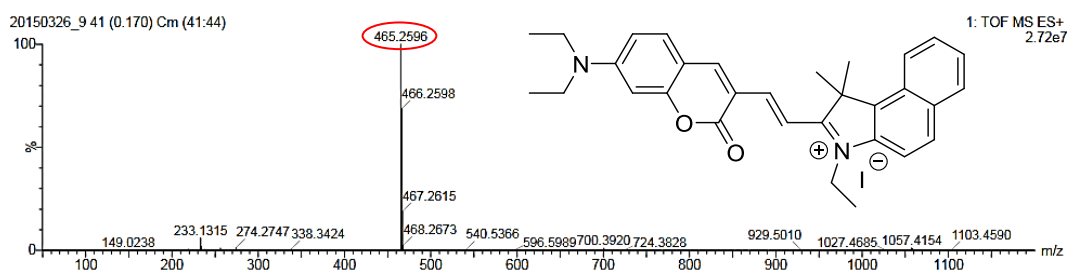


Fig. S2 <sup>13</sup>C NMR spectrum of CMBI in DMSO-*d*<sub>6</sub> (100 MHz).



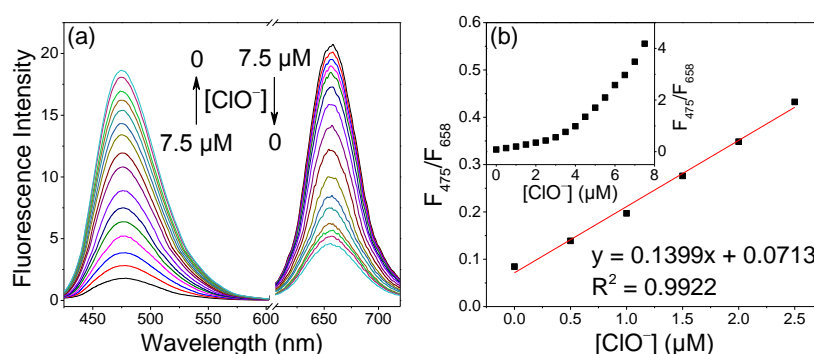
**Fig. S3** HR-MS (ESI) spectrum of **CMBI**.

## 2. Summary of photophysical properties

**Table S1** Spectroscopic properties of **CMBI** (10  $\mu\text{M}$ ) and **CMBI** (10  $\mu\text{M}$ ) with  $\text{ClO}^-$  (100  $\mu\text{M}$ ) in EtOH/PBS solution ( $v/v = 1/9$ , pH 7.4, 10 mM).

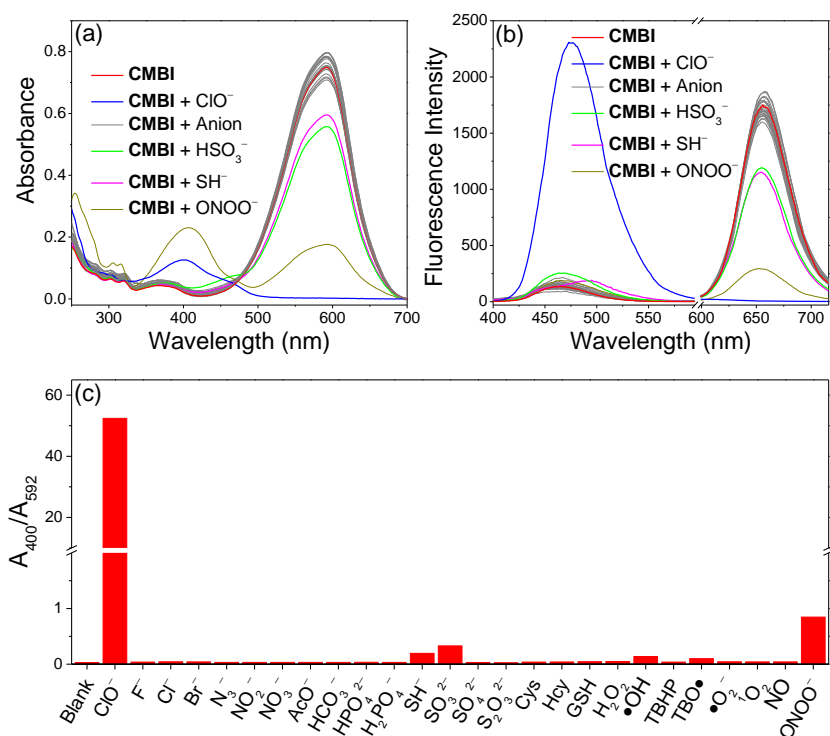
Compound	$\lambda_{\text{abs}}$ (nm)	$\lambda_{\text{em}}$ (nm)	$\Phi$	$\Delta\nu$ ( $\text{cm}^{-1}$ )
<b>CMBI</b>	592	658	0.012	1694
<b>CMBI</b> + $\text{ClO}^-$	400	475	0.141	3947

## 3. Ratiometric response of **CMBI** toward $\text{ClO}^-$ at low concentrations



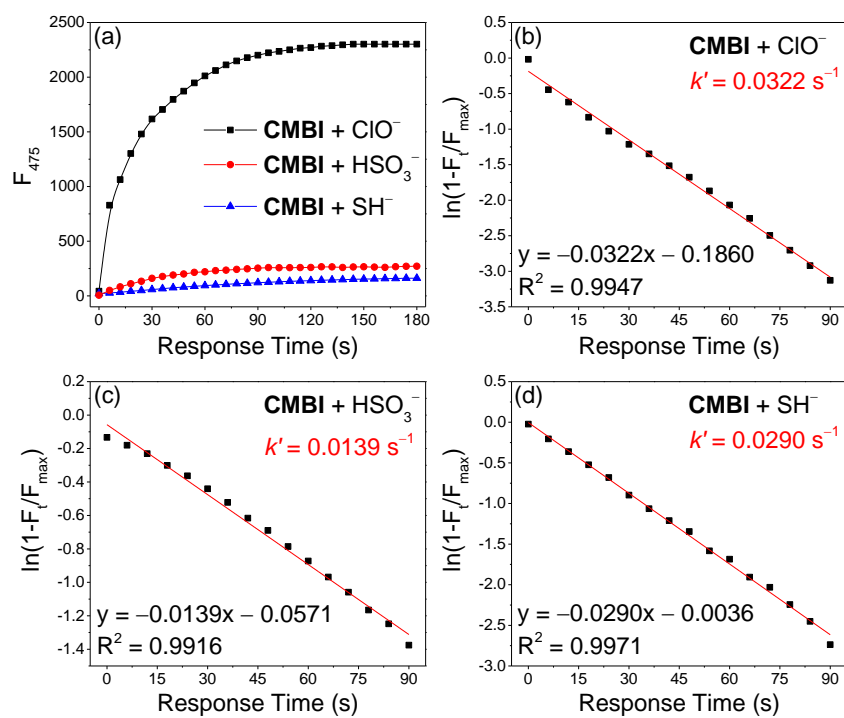
**Fig. S4** (a) Fluorescence spectra changes of 1  $\mu\text{M}$  **CMBI** in EtOH/PBS solution ( $v/v = 1/9$ , pH 7.4, 10 mM) upon addition of increasing amount of  $\text{ClO}^-$  (0 – 7.5  $\mu\text{M}$ ). (b) Linear relationship between fluorescence intensity of 1.0  $\mu\text{M}$  **CMBI** *versus* concentrations of  $\text{ClO}^-$  in EtOH/PBS solution ( $v/v = 1/9$ , pH 7.4, 10 mM). Each spectrum was recorded after 3 min.

#### 4. Selectivity of CMBI toward $\text{ClO}^-$



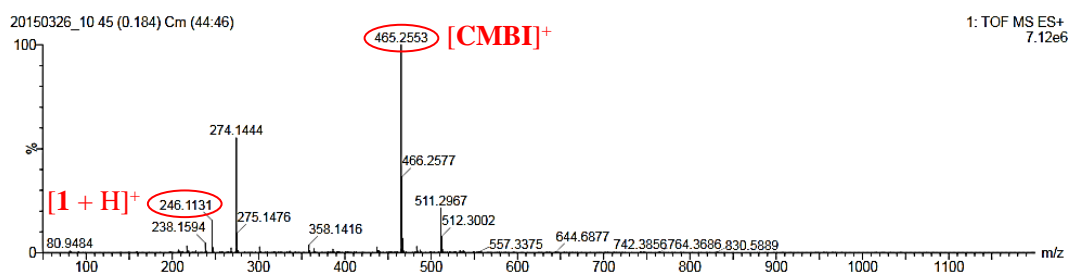
**Fig. S5** (a) UV-vis absorption, (b) fluorescence spectra changes and (c) UV-vis absorption ratios ( $A_{400}/A_{592}$ ) of 10  $\mu\text{M}$  CMBI in EtOH/PBS solution ( $v/v = 1/9$ , pH 7.4, 10 mM) in the presence of various small molecular species (500  $\mu\text{M}$ ) and ROS/RNS (100  $\mu\text{M}$ ). Each spectrum was recorded after 3 min.

## 5. Kinetics research for CMBI towards $\text{ClO}^-$ , $\text{HSO}_3^-$ and $\text{HS}^-$



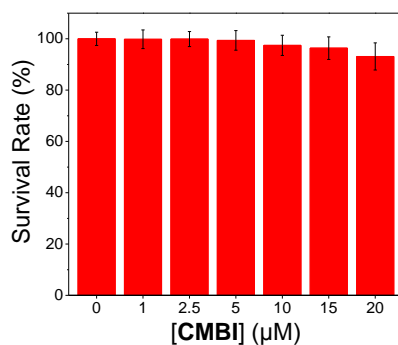
**Fig. S6** Time-resolved fluorescence responses (a) and Pseudo-first-order kinetic plot of 10  $\mu\text{M}$  CMBI towards 100  $\mu\text{M}$   $\text{ClO}^-$  (b),  $\text{HSO}_3^-$  (c) and  $\text{SH}^-$  (d) in EtOH/PBS solution (v/v = 1/9, pH 7.4, 10 mM).

## 6. Proposed mensing mechanism



**Fig. S7** HR-MS (ESI) spectrum of **CMBI** with  $\text{ClO}^-$ .

## 7. Cell toxicity test of CMBI



**Fig. S8** Cell toxicity effect of **CMBI**. HeLa cells were incubated with **CMBI** (0–20  $\mu\text{M}$ ) for 24 h. Results are mean  $\pm$  SD,  $n = 3$ .

## 8. Comparison of ratiometric fluorescent probes for ClO<sup>-</sup>

**Table S2** Comparison of ratiometric fluorescent probes for ClO<sup>-</sup>

Probes	$\lambda_{\text{ex}}$ /nm	$\lambda_{\text{em}}$ /nm	Reagents	Detection limit/nM	Response time/s	Application
Phenanthroimidazole Probe <b>1</b> <sup>[1]</sup>	394	439 509	pH 9.0 PBS (80% DMF)	—	Within seconds	—
<b>1b</b> <sup>[2]</sup>	540 464	585 505	pH 7.4 PBS (20% DMF)	200	< 600	MCF-7 cells (Ex) Ex = exogenous
<b>Cou-Rho-HOCl</b> <sup>[3]</sup>	410	473 594	pH 7.4 PBS (50% DMF)	52	< 60	Bel 7702 cells (Ex) RAW 264.7 cells (En) En = endogenous
BODIPY-DCDHF Probe <b>1</b> <sup>[4]</sup>	465	629 520	pH 7.4 PBS (1% EtOH, 0.1 % Triton X-100)	500	~ 400	RAW 264.7 cells (Ex, En)
<b>Cy7-NR1</b> <sup>[5]</sup>	540	780 566	pH 7.4 PBS (0.5% DMF)	19.5	~ 360	HeLa cells (Ex)
<b>Z2</b> <sup>[6]</sup>	460	509 600	pH 7.4 PBS	63	< 120	Test paper
<b>CDH</b> <sup>[7]</sup>	316	376 456	pH 7.4 HEPES (40% CH <sub>3</sub> CN)	1070	< 60	Water samples TLC plate strips
<b>HRS1</b> <sup>[8]</sup>	410 554	501 578	pH 8.5 PBS (40% DMF)	24	< 60	Water samples
<b>DPNO</b> <sup>[9]</sup>	305	354 430	pH 7.4 HEPES (60% CH <sub>3</sub> CN)	56	< 60	Water samples TLC plate strips
<b>DPH</b> <sup>[10]</sup>	344	495 375	pH 7.4 HEPES (50% EtOH)	350	—	Water samples
<b>CMCY</b> <sup>[11]</sup>	460	631 480	pH 7.4 PBS	80	< 120	HeLa cells (Ex)
Coumarin-Pyridinium Probe <b>1</b> <sup>[12]</sup>	420	631 488	pH 7.4 PBS (75% THF)	93	~ 20	Water samples RAW 264.7 cells (En)
1,8-diaminonaphthalene <sup>[13]</sup>	325	440 518	pH 7.0 HEPES	20	< 20	Water samples HepG2 cells (Ex)
Coumarin-Rhodamine Probe <b>2</b> <sup>[14]</sup>	410	580 470	pH 5.0 PBS (40% DMF)	—	< 50	RAW 264.7 cells (En)

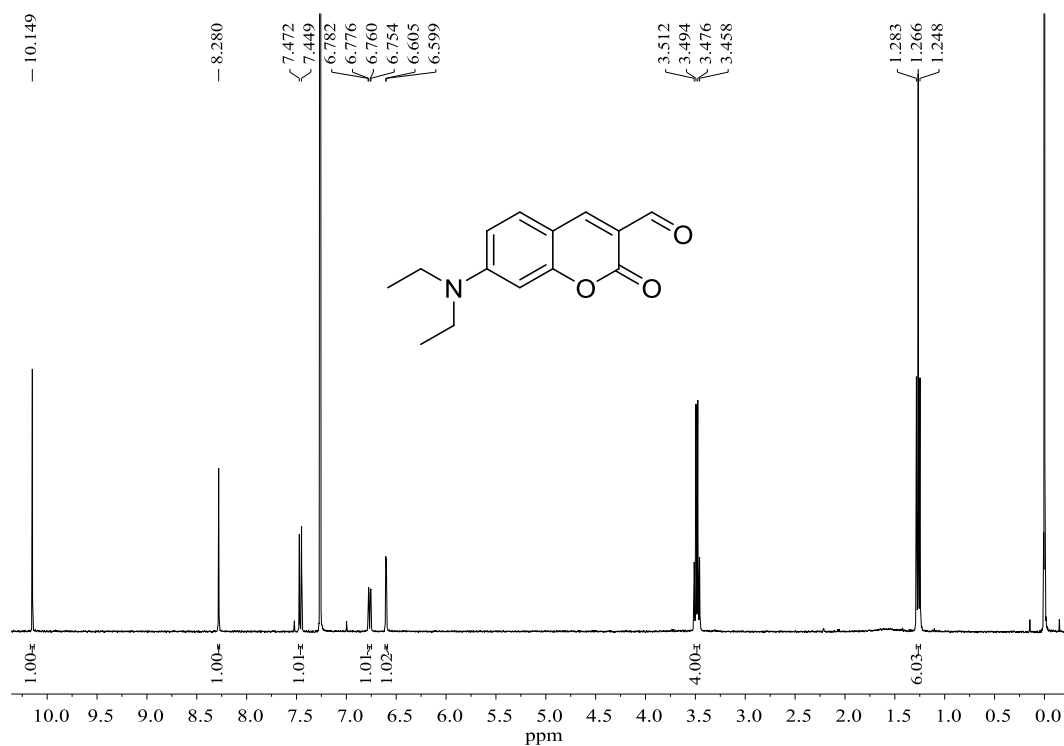
<b>BTHT</b> <sup>[15]</sup>	365	562 476	pH 7.4 HEPES (50% CH <sub>3</sub> CN)	140	< 480	Water samples TLC plate strips
<b>NIR-1</b> thiospirolactone	430	486	pH 7.4 PBS	40	Within seconds	Water samples
Probe <b>1</b> <sup>[16]</sup>	650	707	(30% CH <sub>3</sub> CN)			HeLa cells (Ex) RAW 264.7 cells (En)
<b>TAM</b> <sup>[17]</sup>	430	630 485	pH 7.1 H <sub>2</sub> O (40% THF)	70	~ 100	TLC plate strips PBMCs (Ex)
Dansyl-Rhodamine	400	494	pH 8.0 PBS	1130	< 120	Water samples
Probe <b>1</b> <sup>[18]</sup>		578	(10% CH <sub>3</sub> CN)			Test paper
<b>PMN-TPP</b> <sup>[19]</sup>	410	522 640	pH 7.3 PBS (0.5% DMSO, 1 mM Triton X-100)	430	Few seconds	RAW 264.7 cells (En) Nude mouse (En)
<b>Naph-Rh</b> <sup>[20]</sup>	350	440 585	pH 6.0 PBS (30% EtOH)	100	< 50	RAW 264.7 cells (En)
<b>RMClO-2</b> <sup>[21]</sup>	412	476	pH 7.4 PBS	0.84	Within seconds	Normal and cancer cells
	540	570	(0.1% DMSO)			
<b>HPQ-Cy<sub>2</sub></b> <sup>[22]</sup>	365	575 435	pH 7.4 PBS (0.5% EtOH)	28	< 50	A375 cells (En)
<b>CRSH</b> <sup>[23]</sup>	350	470 580	pH 5.0 PBS (50% EtOH)	210	~ 60	RAW 264.7 cells (En)
<b>STBR</b> <sup>[24]</sup>	370	514 595	pH 7.5 PBS (20% CH <sub>3</sub> CN)	220	—	TLC plate strips Baker's yeast (Ex)
<b>AETU-HOCI</b> <sup>[25]</sup>	380	533 473	pH 7.4 PBS (50% EtOH)	3.9	~ 60	RAW 264.7 cells (En)
<b>QC1</b> <sup>[26]</sup>	400	482 381	pH 7.4 HEPES (1% DMSO)	410	—	—
Naphthalimide-Indolium	430	620	pH 7.0 HEPES	53	~ 50	HepG2 cells (Ex)
Probe <b>1</b> <sup>[27]</sup>		515	(50% MeOH)			
<b>HES-BODIPY</b> <sup>[28]</sup>	480	562 532	pH 7.1 PBS (10% EtOH)	430	Within seconds	RAW 264.7 cells (En)
<b>BRT</b> <sup>[29]</sup>	525	540 580	pH 5.0 PBS (60% EtOH)	38	~ 15	RAW 264.7 cells (En)
<b>CARSH</b> <sup>[30]</sup>	350	490 580	pH 5.0 PBS (50% EtOH)	200	~ 60	RAW 264.7 cells (En)
<b>Lyso-HA</b> <sup>[31]</sup>	420	450 586	pH 5.0 PBS (30% DMSO)	110	Within seconds	HeLa cells (Ex) RAW 264.7 cells (En)



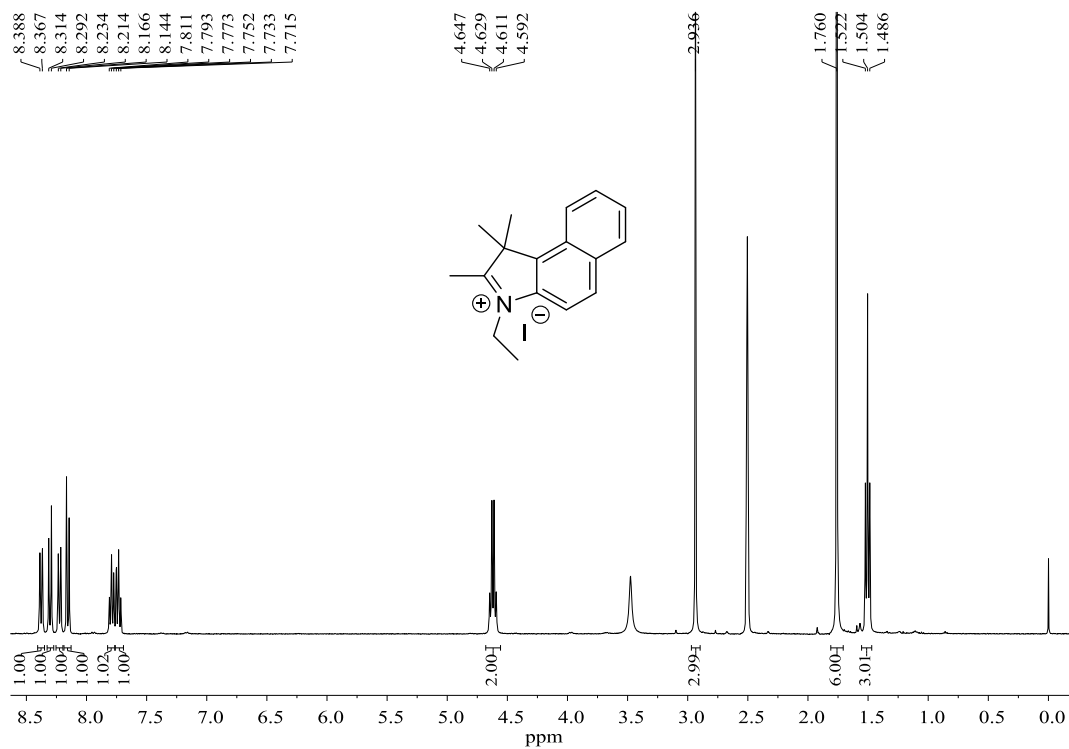
<b>MTPE-M</b> <sup>[32]</sup>	340	595	pH 7.4 PBS (1% DMSO, 1mM CTAB)	470	~ 180	RAW 264.7 cells (En) Zebrafish (En)
		498				
<b>Py-Cy</b> <sup>[33]</sup>	430	613	pH 7.4 PBS (5% DMSO)	350	4200	Water samples HeLa cells (Ex) RAW 264.7 cells (En)
		470				
<b>CMBI</b> (This work)	390	585	pH 7.4 PBS (10% EtOH)	33	~ 90	HeLa cells (Ex)
		475				

- 1 W. Lin, L. Long, B. Chen and W. Tan, *Chem. Eur. J.*, 2009, **15**, 2305.
- 2 L. Yuan, W. Lin, J. Song and Y. Yang, *Chem. Commun.*, 2011, **47**, 12691.
- 3 L. Yuan, W. Lin, Y. Xie, B. Chen, J. Song, *Chem. Eur. J.*, 2012, **18**, 2700.
- 4 J. Park, H. Kim, Y. Choi and Y. Kim, *Analyst*, 2013, **138**, 3368.
- 5 Z. Lou, P. Li, P. Song and K. Han, *Analyst*, 2013, **138**, 6291.
- 6 Q. Wang, C. Liu, J. Chang, Y. Lu, S. He, L. Zhao and X. Zeng, *Dyes Pigm.*, 2013, **99**, 733.
- 7 S. Goswami, S. Paul and A. Manna, *Dalton Trans.*, 2013, **42**, 10097.
- 8 L. Long, D. Zhang, X. Li, J. Zhang, C. Zhang and L. Zhou, *Anal. Chim. Acta*, 2013, **775**, 100.
- 9 S. Goswami, A. Manna, S. Paul, C. K. Quah and H.-K. Fun, *Chem. Commun.*, 2013, **49**, 11656.
- 10 C.-C. Zhang, Y. Gong, Y. Yuan, A. Luo, W. Zhang, J. Zhang, X. Zhang and W. Tan, *Anal. Methods*, 2014, **6**, 609.
- 11 J. Zha, B. Fu, C. Qin, L. Zeng and X. Hu, *RSC Adv.*, 2014, **4**, 43110.
- 12 L. Wang, L. Long, L. Zhou, Y. Wu, C. Zhang, Z. Han, J. Wang and Z. Da, *RSC Adv.*, 2014, **4**, 59535.
- 13 Y. Yang, C. Yin, F. Huo, J. Chao, Y. Zhang and S. Jin, *Sens. Actuators, B*, 2014, **199**, 226.
- 14 Y.-R. Zhang, X.-P. Chen, J. Shao, J.-Y. Zhang, Q. Yang, J.-Y. Miao and B.-X. Zhao, *Chem. Commun.*, 2014, **50**, 14241.
- 15 A. Manna and S. Goswami, *New J. Chem.*, 2015, **39**, 4424.
- 16 S. Ding, Q. Zhang, S. Xue and G. Feng, *Analyst*, 2015, **140**, 4687.
- 17 S. Goswami, K. Aich, S. Das, B. Pakhira, K. Ghoshal, C. K. Quah, M. Bhattacharyya, H.-K. Fun and S. Sarkar, *Chem. Asian J.*, 2015, **10**, 694.
- 18 H. J. Lee, M. J. Cho and S.-K. Chang, *Inorg. Chem.*, 2015, **54**, 8644.
- 19 H. Xiao, J. Li, J. Zhao, G. Yin, Y. Quan, J. Wang and R. Wang, *J. Mater. Chem. B*, 2015, **3**, 1633.
- 20 Y.-R. Zhang, N. Meng, J.-Y. Miao and B.-X. Zhao, *Chem. Eur. J.* 2015, **21**, 19058.
- 21 J.-T. Hou, K. Li, J. Yang, K.-K. Yu, Y.-X. Liao, Y.-Z. Ran, Y.-H. Liu, X.-D. Zhou and X.-Q. Yu, *Chem. Commun.*, 2015, **51**, 6781.
- 22 L. Zhou, D.-Q. Lu, Q. Wang, S. Hu, H. Wang, H. Sun and X. Zhang, *Spectrochim. Acta A*, 2016, 166, 129.
- 23 Y.-R. Zhang, Z.-M. Zhao, L. Su, J.-Y. Miao and B.-X. Zhao, *RSC Adv.*, 2016, **6**, 17059.
- 24 A. Manna, D. Sarkar, S. Goswami, C. K. Quah and H.-K. Fun, *RSC Adv.*, 2016, **6**, 57417.
- 25 W. Shu, P. Jia, X. Chen, X. Li, Y. Huo, F. Liu, Z. Wang, C. Liu, B. Zhu, L. Yan and B. Du, *RSC Adv.*, 2016, **6**, 64315.
- 26 L. Wang, Y. Hu, Y. Qu, J. Xu and J. Cao, *Dyes Pigm.*, 2016, **128**, 54.
- 27 J. Li, P. Li, F. Huo, C. Yin, T. Liu, J. Chao and Y. Zhang, *Dyes Pigm.*, 2016, **130**, 209.
- 28 X. Wang, L. Zhou, F. Qiang, F. Wang, R. Wang and C. Zhao, *Anal. Chim. Acta*, 2016, **911**, 114.
- 29 Y. Liu, Z.-M. Zhao, J.-Y. Miao and B.-X. Zhao, *Anal. Chim. Acta*, 2016, **921**, 77.
- 30 Y.-R. Zhang, Z.-M. Zhao, J.-Y. Miao and B.-X. Zhao, *Sens. Actuators, B*, 2016, **229**, 408.
- 31 M. Ren, B. Deng, K. Zhou, X. Kong, J.-Y. Wang, G. Xu and W. Lin, *J. Mater. Chem. B*, 2016, **4**, 4739.
- 32 Y. Huang, P. Zhang, M. Gao, F. Zeng, A. Qin, S. Wu and Z. Tang, *Chem. Commun.*, 2016, **52**, 7288.
- 33 Y. Wu, J. Wang, F. Zeng, S. Huang, J. Huang, H. Xie, C. Yu and S. Wu, *ACS Appl. Mater. Interfaces*, 2016, **8**, 1511.

## 9. $^1\text{H}$ NMR spectra of compounds 1 and 2



**Fig. S9**  $^1\text{H}$  NMR spectrum of compound 1 in  $\text{CDCl}_3$  (400 MHz).



**Fig. S10**  $^1\text{H}$  NMR spectrum of compound 2 in  $\text{DMSO}-d_6$  (400 MHz).