Raw	60 ºC 30 min	60 ºC 60 min	60 ºC 90 min	60 ºC 120 min
Raw	80 °C 30 min	80 ºC 60 min	80 ºC 90 min	80 ºC 120 min
Raw	100 °C 30 min	100 °C 60 min	100 °C 90 min	100 °C 120 min

Electronic Supplementary Information

Fig. S1. Dissolution of ball-milled wood in DMSO/[Emin]OAc solvent system under different temperatures (Optical microscopy).



Fig. S2. ³¹P-NMR spectra of the lignin fractions isolated from poplar wood

Table S1

Samples	Crystalline region	Amorphous region	CrI ^a (%)
Raw	1.00	4.08	19.68
R-raw-60 °C	1.00	3.15	24.10
R-raw-80 °C	1.00	2.81	26.26
R-raw-100 °C	1.00	3.68	21.36

Quantification of the raw and regenerated poplar wood by CP/MAS ¹³C-NMR

^a Crystallinity index

 $CrI~(\%) = I_{Crystalline~region} / ~I_{Crystalline~region+Amorphous~region}$

Table S2

Labels	$\delta_C / \delta_H (ppm)$	Assignment
C _β	52.9/3.45	C_{β} -H _{β} in phenylcomaran structures(C)
\mathbf{B}_{β}	53.5/3.05	C_{β} -H _{β} in resinol substructures(B)
-OCH ₃	55.6/3.72	C-H in methoxyls
A_{γ}	59.5/3.70 and 3.39	C_{γ} -H _{γ} in β -O-4' substructures(A)
D_{β}	59.7/2.73	C_{β} -H _{β} in spirodienone substructures(D)
I_{γ}	61.4/4.10	C_{γ} -H _{γ} in cinnamyl(sinapyl/coniferyl) alcohol end groups(I)
C_{γ}	62.3/3.73	C_{γ} -H _{γ} in phenylcomaran structures(C)
X_5	62.5/3.39	C_5 - $H_5 \beta$ -D-xylopyranoside substructures(X)
A'_{γ}	63.1/4.29	C_{γ} -H _{γ} in γ -acylated β -O-4' substructures(A')
\mathbf{B}_{γ}	71.1/3.81 and 4.17	C_{γ} -H _{γ} in resinol substructures(B)
A_{α}	71.9/4.86	C_{α} -H _{α} in β -O-4' substructures(A)
D'_{α}	79.4/4.10	C_{α} -H _{α} in spirodienone substructures(D')
BE	81.0/4.65	C_{α} -H _{α} in benzyl-ether (BE) linkage
D_{α}	81.1/5.09	C_{α} -H _{α} in spirodienone substructures(D)
$A_{\beta}(G/H)$	83.4/4.43	C_{β} -H _{β} in β -O-4' linked to a G/H unit(A)
\mathbf{B}_{α}	84.8/4.65	C_{α} -H _{α} in resinol substructures(B)
$A_{\beta}(S)$	85.8/4.11	C_{β} -H _{β} in β -O-4' linked to a S unit(A)
C_{α}	86.79/5.46	C_{α} -H _{α} in phenylcomaran structures(C)
S _{2,6}	103.9/6.71	C _{2,6} -H _{2,6} in syringyl units(S)
S' _{2,6}	106.2/7.32	C _{2,6} -H _{2,6} in oxidized(C=O) phenolic syringyl units(S')
G ₂	110.9/6.96	C ₂ -H ₂ in guaiacyl units(G)
G ₅	114.9/6.76	C_5 - H_5 in guaiacyl units(G)
G ₆	119.0/6.78	C ₆ -H ₆ in guaiacyl units(G)
H _{2,6}	127.8/7.22	C _{2,6} -H _{2,6} in <i>p</i> -hydroxyphenyl units(S)
PB _{2,6}	131.2/7.66	C _{2,6} -H _{2,6} in <i>p</i> -hydroxybenzoate units(S)

Assignments of ¹³C-¹H correlation signals in the HSQC spectra of the lignin from poplar wood (according to the previous publications¹⁻⁴).

Reference

- 1 S. Ralph, J. Ralph, L. Landucciand L. Landucci. US Forest Prod. Lab., Madison, WI (http://ars. usda. gov/Services/docs. htm. 2004.
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2. Methods

2.1. Structure elucidation of CELs and RCELs

Acetylation of Lignin Samples

About 50 mg of CEL and RECL were dissolved in a solution of dimethyl sulfoxide: 1-methylimidazole (2:1, v/v) 3mL and stirred without direct light at room temperature for 12 h. Acetic anhydride (1 mL) was added to the reaction mixture and continued reacting for 2 h. The reaction mixture was dropped slowly into 100 mL acid water (pH=2) to induce precipitation followed by centrifugation. The acetylated lignin was dissolved in tetrahydrofuran (THF) (2 mg/mL), and the solution was filtered through a 0.22 μ m filter. The filtered solution (20 μ L) was injected into the HPLC system and detected using an UV detector setting at 280 nm. THF was used as the mobile phase and the flow rate was 0.8 mL/min. Standard PL polystyrene samples were used for calibration. GPC analyses were run at least twice.

2D-HSQC NMR

About 50 mg of lignin was dissolved in 0.5 mL of DMSO- d_6 (99.8% D). For quantitative 2D-HSQC spectra, the Bruker standard pulse program hsqcetgpsi2 was used for HSQC experiments. The spectral widths were 5000 Hz and 20000 Hz for the ¹H- and ¹³C-dimensions, respectively. The number of collected complex points was 1024 for ¹H-dimension with a recycle delay of 1.5 s. The number of transients was 64, and 256 time increments were always recorded in the ¹³C-dimension. The ¹J_{CH} used was 145 Hz. Prior to Fourier transformation, the data matrixes were zero filled up to 1024 points in the ¹³C-dimension. Data processing was performed using standard Bruker Topspin-NMR software.

A quantitative analysis of the intensities of the HSQC cross-signal was performed according to the following formula: ¹⁻²

 $I(C_9)$ units = $0.5I(S_{2,6}) + I(G_2)$

Where $I(S_{2,6})$ is the integration of $S_{2,6}$, including S and S', $I(G_2)$ is the integral value of G_2 . $I(C_9)$ represents the integral value of the aromatic ring. According to the internal standard ($I(C_9)$), the amount of I(X)% could be obtained by the following formula:

 $I(X)\% = I(X)/I(C_9) \times 100\%$

Where I(X) is the integral value of the α -position of A (β -O-4'), B (β - β), and C (β -5), the integration should be in the same contour level.

In the aromatic region, $C_{2,6}$ -H_{2,6} correlations from S units and C_2 -H₂ correlation from G units were used to estimate the S/G ratio of lignin. The S/G ratio could be obtained by the following formula: $S/G = 0.5I(S_{2/6})/I(G_2)$

Quantitative ³¹P NMR

20 mg lignin was dissolved in 500 μ L of anhydrous pyridine and deuterated chloroform (1.6:1, v/v) under stirring. This was followed by the addition 100 μ L of cyclohexanol (10.85 mg/mL in anhydrous pyridine and deuterated chloroform 1.6:1, v/v) as an internal standard and 100 μ L of Chromium (III) acetylacetonate solution (5 mg mL⁻¹ in anhydrous pyridine and deuterated chloroform 1.6:1, v/v) as relaxation reagent. The mixture was reacted with 100 μ L of phosphitylating reagent (2-chloro-4,4,5,5-tetramethyl-1,3,2-dioxaphospholate, TMDP) for about 10 min and was transferred into a 5 mm NMR tube for subsequent NMR analysis.

Reference

M. Sette, R. Wechselbergerand C. Crestini. *Chem–Eur.* 2011,17,9529-9535.
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