

-SUPPLEMENTARY INFORMATION-

Advances and mechanistic insight on the catalytic Mitsunobu reaction using recyclable azo reagents

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General remarks

All reactions were performed in oven-dried glassware. All reagents purchased commercially were used without further purification unless otherwise noted. Iron phthalocyanine was purchased from Tokyo Chemical Industry Co., Ltd. Dehydrated THF, toluene and CH₂Cl₂ were purchased from Kanto Chemical Co., Inc. Other solvents were dried with activated molecular sieves. Molecular sieves were activated by heating with a heat gun (ISHIZAKI ELECTRIC MFG PJ-208A, ca. 450 °C) in vacuo (ca. 0.1 mmHg) for 5 min unless otherwise noted. Thin-layer chromatography (TLC) analysis was performed by illumination with a UV lamp (254 nm) or staining with PMA and heating. Silica gel column chromatography was carried out on silica gel 60N (Kanto Chemical Co., Inc., spherical, neutral, 40–50 µm). The chromatographic separation of enantiomers was performed using a JASCO PU-2080 Plus liquid chromatography equipped with Multi UV-Vis (JASCO MD-910). Melting points were recorded on Yanako melting point apparatus or on a Kofler micro hot stage and were uncorrected.

¹H and ¹³C NMR spectra were recorded with JEOL JNM ECS400 (400 MHz and 100 MHz), JEOL JNM ECS500 (500 MHz and 125 MHz) and JEOL JNM ECA600 (600 MHz and 150 MHz) spectrometers at 293–295 K in Japan. ³¹P NMR spectra were recorded with a JEOL JNM ECA600 (243 MHz) spectrometer at 298 K in Japan. Some ¹H, ³¹P, ¹³C and ¹⁵N NMR spectra were recorded with a Bruker Avance III 500 MHz NMR (500 MHz, 202 MHz, 126 MHz and 51 MHz) instrument at 296 K in Slovenia. Proton spectra were referenced to TMS as an internal standard. Carbon chemical shifts were determined relative to the ¹³C signal of CDCl₃ (77.0 ppm). ³¹P NMR spectra were referenced to external 85% phosphoric acid (δ = 0 ppm). ¹⁵N chemical shifts were extracted from ¹H–¹⁵N gs-HMBC spectra (with 20 Hz digital resolution in the indirect dimension and the parameters adjusted for a long-range ¹H–¹⁵N coupling constant of 5 Hz), determined with respect to external nitromethane and corrected to external ammonia by addition of 380.5 ppm. Assignments of some proton, carbon, phosphorous, and nitrogen resonances were performed by 2D NMR techniques (¹H–¹H gs-COSY, ¹H–¹³C gs-HSQC, ¹H–¹³C gs-HMBC, ¹H–³¹P gs-HMBC, ¹H–¹⁵N gs-HMBC). Coupling constants (*J*) are given in Hz. Multiplicities are indicated as follows: s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet) or br (broadened).

IR spectra were recorded with a JASCO Fourier Transform IR-460 spectrometer or with a Perkin-Elmer Spectrum 100 (equipped with a Specac Golden Gate Diamond ATR as a solid sample support). Mass spectra were recorded on JEOL JMS-T100TD (direct analysis in real time, DART or electrospray ionization, ESI). A time-of-flight (TOF) mass spectrometer equipped with a double orthogonal electrospray source at atmospheric pressure ionization (ESI) coupled to an HPLC instrument was used for recording HRMS spectra. Optical rotations were measured on a JASCO P-1030 polarimeter. Absorption spectra were measured in a 0.1 mm quartz cell on a JASCO V-570 spectrophotometer. Thermogravimetry-Differential thermal analysis (TG-DTA) was performed on

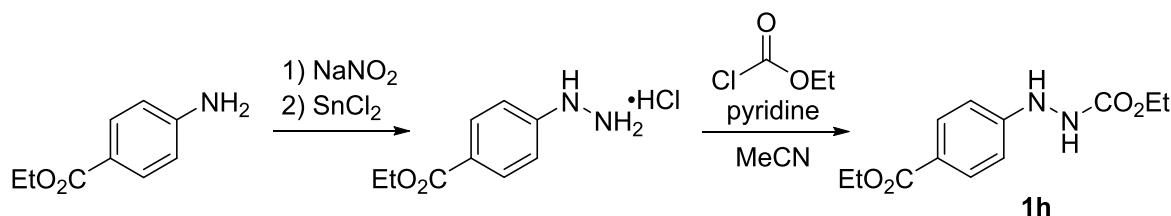
EXSTAR TG/DTA-7300 (heating rate: 10.0 °C/min) under a nitrogen atmosphere. Differential scanning calorimetric (DSC) analysis was performed on Perkin Elmer Jade DSC (heating rate of 10.0 °C/min) under a nitrogen atmosphere.

Experimental details

1. Synthetic procedure and analytical data for products

Ethyl 2-arylhydrazinecarboxylates (**1**) and ethyl 2-ary lazocarboxylates (**2**) except for ethyl 2-[(4-ethoxycarbonyl)phenyl]hydrazinecarboxylate (**1h**) and 2-[(4-ethoxycarbonyl)phenyl]azocarboxylate (**2h**) were known compounds and prepared according to previous reports.^{1,2}

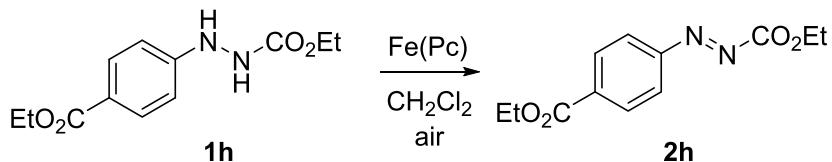
*Procedure for preparation of ethyl 2-[4-(ethoxycarbonyl)phenyl]hydrazinecarboxylate (**1h**):*



A suspension of ethyl 4-aminobenzoate (1.65 g, 10 mmol) in concentrated HCl aq. (10 mL) was cooled to 0 °C. A solution of NaNO₂ (759 mg, 11 mmol) in water (10 mL) was added dropwise to the suspension. After the mixture was stirred for 10 min at 0 °C, a solution of SnCl₂ (5.88 g, 31 mmol) in concentrated HCl aq. (30 mL) was slowly added to the mixture. The mixture was stirred for 30 min at room temperature, and the formed precipitate was collected by filtration. The precipitate was washed with concentrated HCl aq. and dried under vacuum. To a solution of the obtained crude product and pyridine (3.96 g, 50 mmol) in CH₃CN (10 mL) was added dropwise ethyl chloroformate (1.19 g, 11 mmol) at 0 °C. The mixture was stirred for 30 min at 0 °C and allowed to warm to room temperature, and further stirred for 13 h. The reaction mixture was poured into water and extracted with ethyl acetate. The combined organic layers were washed with brine and dried with Na₂SO₄. After the solvent was evaporated, the crude product was washed by hexane and dried under vacuum to afford ethyl 2-[4-(ethoxycarbonyl)phenyl]hydrazinecarboxylate (**1h**: 1.45 g, 5.74 mmol, 57% yield) as a white solid. Recrystallization (hexane/EtOAc) of this product gave highly pure form as colorless needles.

Mp 81.5–82.0 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.94 (2H, app. d, *J* = 9.2 Hz), 6.81 (2H, d, *J* = 8.7 Hz), 6.58 (1H, br-s), 6.04 (1H, br-s), 4.33 (2H, q, *J* = 7.2 Hz), 4.21 (2H, q, *J* = 7.0 Hz), 1.37 (3H, t, *J* = 7.1 Hz), 1.28 (3H, br-s); ¹³C NMR (101 MHz, CDCl₃) δ 166.5, 156.8, 151.8, 131.3, 122.7, 111.8, 62.2, 60.5, 14.5, 14.4; IR (KBr, cm⁻¹) ν 2982, 1694, 1608, 1539, 1282, 1171, 1108; HRMS (DART+) (*m/z*): calcd for C₁₂H₁₇N₂O₂ ([M+H]⁺): 253.1188, found: 253.1197.

*Procedure for preparation of ethyl 2-[4-(ethoxycarbonyl)phenyl]azocarboxylate (**2h**):*

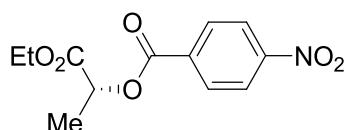


According to the reported procedure,² a mixture of ethyl 2-[4-(ethoxycarbonyl)phenyl]hydrazinecarboxylate (**1h**) (1.01 g, 4.0 mmol) and iron phthalocyanine (227 mg, 0.40 mmol) in CH_2Cl_2 (4.0 mL) was stirred for 20 h at room temperature under air. After the solvent was removed under reduced pressure, the residue was purified by flash chromatography (silica gel, hexane/EtOAc=5:1) to give ethyl 2-[4-(ethoxycarbonyl)phenyl]azocarboxylate (**2h**: 1.00 g, 4.0 mmol, 100% yield) as a red oil.

¹H NMR (400 MHz, CDCl_3) δ 8.21 (2H, app. d, J = 6.9 Hz), 7.97 (2H, app. d, J = 6.9 Hz), 4.54 (2H, q, J = 7.2 Hz), 4.42 (2H, q, J = 7.2 Hz), 1.48 (3H, t, J = 7.1 Hz), 1.43 (3H, t, J = 7.3 Hz); ¹³C NMR (101 MHz, CDCl_3) δ 165.5, 161.9, 153.7, 134.6, 130.6, 123.4, 64.7, 61.5, 14.3, 14.1; IR (neat, cm^{-1}) ν 2985, 1760, 1722, 1277, 1246, 1188, 1106; HRMS (DART+) (m/z): calcd for $\text{C}_{12}\text{H}_{15}\text{N}_2\text{O}_2$ ($[\text{M}+\text{H}]^+$): 251.1032, found: 251.1025.

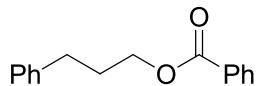
General procedure for catalytic Mitsunobu reactions:

A mixture of alcohol (1.0 mmol), nucleophile (1.1 mmol), triphenylphosphine (525 mg, 2.0 mmol), a hydrazine catalyst [ethyl 2-(3,4-dichlorophenyl)hydrazinecarboxylate (**1a**): 24.9 mg, 0.1 mmol or ethyl 2-(4-cyanophenyl)hydrazinecarboxylate (**1j**): 20.5 mg, 0.1 mmol], iron phthalocyanine (56.8 mg, 0.1 mmol) and powder of activated molecular sieves 5 \AA (500 mg) in toluene (250 μL) was irradiated for ca. 1 minute in an ultrasound bath and vigorously stirred under air atmosphere (balloon) at room temperature until disappearance of alcohol or triphenylphosphine. After the reaction mixture was filtrated through a filter paper or a pad of Celite[®], the solvent was removed under reduced pressure. The crude material was purified by silica gel chromatography (*n*-hexane/EtOAc) to give the corresponding product.

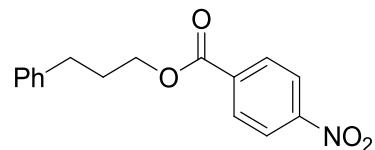


(R)-1-Ethoxycarbonylethyl 4-nitrobenzoate (5).¹ (–)-(S)-Ethyl lactate (**3**) (118 mg, 1.0 mmol, 99:1 er) and 4-nitrobenzoic acid (**3**) (184 mg, 1.1 mmol) were used as substrates. 12 h. 93% yield (249 mg, 0.93 mmol). 99:1 er [Fe(Pc) (1 mol%), PPh₃ (1.5 equiv): 18 h. 81% yield (215 mg, 0.81 mmol), 99:1 er]. White solid. Elute: Hex/AcOEt = 6:1. ¹H NMR (500 MHz, CDCl_3) δ 8.31 (2H, app. d, J = 9.5 Hz), 8.27 (2H, app. d, J = 9.5 Hz), 5.36 (1H, q, J = 7.0 Hz), 4.26 (2H, q, J = 7.0 Hz), 1.67 (3H, d, J = 7.0 Hz), 1.30 (3H, t, J = 7.0 Hz); ¹³C NMR (125 MHz, CDCl_3) δ 170.2, 164.0, 150.7, 134.8, 130.9, 123.5, 69.9, 61.6, 16.9, 14.0; The enantiomeric ratio was determined by HPLC analysis

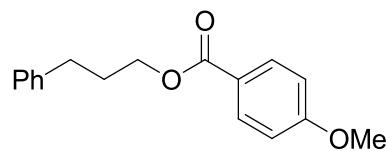
using a chiral column. Chiral HPLC: Daicel-Chiraldpak AD-H 46 × 150 mm, multi UV-Vis detector (230 nm), room temperature eluent: (*n*-hexane/*i*-PrOH) 5:1, flow rate: 0.5 mL/min, retention time (min) 14.0 (*R* isomer), 17.5 (*S* isomer). $[\alpha]_D^{25} = -18.5$ (*c* 1.00, CHCl₃) [lit.¹ (*S*)-1-ethoxycarbonylethyl 4-nitrobenzoate (99:1 er), $[\alpha]_D^{30} = +13.6$ (*c* 1.00, CHCl₃)].



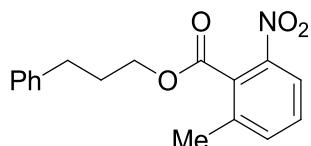
3-Phenylpropyl benzoate (11).¹ 3-Phenylpropanol (**6**) (136 mg, 1.0 mmol) and benzoic acid (134 mg, 1.1 mmol) were used as substrates. 18 h. 93% yield (222 mg, 0.93 mmol) [Fe(Pc) (1 mol%), PPh₃ (1.5 equiv): 36 h. 93% yield (223 mg, 0.93 mmol)]. Colorless oil. Elute: Hex/AcOEt = 30:1. ¹H NMR (600 MHz, CDCl₃) δ 8.04 (2H, d, *J* = 7.2 Hz), 7.55 (1H, app. t, *J* = 7.6 Hz), 7.43 (2H, app. t, *J* = 7.7 Hz), 7.29 (2H, app. t, *J* = 7.6 Hz), 7.22–7.18 (3H, m), 4.34 (2H, t, *J* = 6.6 Hz), 2.78 (2H, t, *J* = 7.6 Hz), 2.10 (2H, tt, *J* = 7.6, 6.6 Hz); ¹³C NMR (150 MHz, CDCl₃) δ 166.5, 141.1, 132.8, 130.3, 129.5, 128.43, 128.39, 128.30, 126.0, 64.2, 32.2, 30.2.



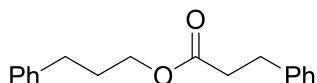
3-Phenylpropyl 4-nitrobenzoate (12).¹ 3-Phenylpropanol (**6**) (136 mg, 1.0 mmol) and 4-nitrobenzoic acid (**4**) (184 mg, 1.1 mmol) were used as substrates. 12 h. 91% yield (257 mg, 0.91 mmol) [**1a** (3 mol%), Fe(Pc) (3 mol%), PPh₃ (1.5 equiv): 23 h. 88% yield (250 mg, 0.88 mmol)]. Pale yellow oil. Elute: Hex/AcOEt = 20:1. ¹H NMR (600 MHz, CDCl₃) δ 8.28 (2H, app. d, *J* = 8.9 Hz), 8.16 (2H, app. d, *J* = 8.9 Hz), 7.30 (2H, app. t, *J* = 7.6 Hz), 7.22–7.20 (3H, m), 4.40 (2H, t, *J* = 6.6 Hz), 2.80 (2H, t, *J* = 7.2 Hz), 2.15 (2H, tt, *J* = 7.2, 6.6 Hz); ¹³C NMR (150 MHz, CDCl₃) δ 164.6, 150.5, 140.9, 135.6, 130.6, 128.5, 128.4, 126.1, 123.5, 65.3, 32.3, 30.0.



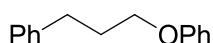
3-Phenylpropyl-4-methoxybenzoate (13).³ 3-Phenylpropanol (**6**) (136 mg, 1.0 mmol) and 4-methoxybenzoic acid (167 mg, 1.1 mmol) were used as substrates. 18 h. 89% yield (238 mg, 0.89 mmol). Colorless oil. Elute: Hex/AcOEt = 20:1. ¹H NMR (600 MHz, CDCl₃) δ 7.99 (2H, app. d, *J* = 8.9 Hz), 7.29 (2H, app. t, *J* = 7.6 Hz), 7.22–7.18 (3H, m), 6.92 (2H, app. d, *J* = 8.9 Hz), 4.31 (2H, t, *J* = 6.6 Hz), 3.85 (3H, s), 2.78 (2H, t, *J* = 7.6 Hz), 2.09 (2H, tt, *J* = 7.6, 6.6 Hz); ¹³C NMR (150 MHz, CDCl₃) δ 166.3, 163.3, 141.2, 131.5, 128.4, 126.0, 122.8, 113.6, 63.9, 55.4, 32.3, 30.3.



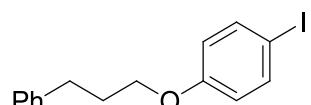
3-Phenylpropyl-2-methyl-6-nitrobenzoate (14). 3-Phenylpropanol (**6**) (136 mg, 1.0 mmol) and 2-methyl-6-nitrobenzoic acid (199 mg, 1.1 mmol) were used as substrates. 18 h. 89% yield (265 mg, 0.89 mmol). Pale yellow oil. Elute: Hex/AcOEt=10:1. ^1H NMR (600 MHz, CDCl_3) δ 7.99 (1H, d, J = 8.2 Hz), 7.54 (1H, d, J = 7.6 Hz), 7.47 (1H, t, J = 7.9 Hz), 7.29 (2H, app. t, J = 7.6 Hz), 7.22–7.19 (3H, m), 4.40 (2H, t, J = 6.6 Hz), 2.73 (2H, t, J = 7.8 Hz), 2.43 (3H, s), 2.08 (2H, tt, J = 7.8, 6.6 Hz); ^{13}C NMR (150 MHz, CDCl_3) δ 166.5, 146.2, 141.0, 137.5, 135.9, 129.66, 129.57, 128.4, 126.0, 121.8, 65.6, 32.1, 30.0, 19.1; IR (neat, cm^{-1}) ν 2928, 1734, 1539; HRMS (DART+) (m/z): $[\text{M}+\text{NH}_4]^+$ calcd for $\text{C}_{17}\text{H}_{21}\text{N}_2\text{O}_4$: 317.1501, found: 317.1504.



3-Phenylpropyl-3-phenylpropanoate (15).³ 3-Phenylpropanol (**6**) (136 mg, 1.0 mmol) and 3-phenylpropionic acid (165 mg, 1.1 mmol) were used as substrates. 18 h. 91% yield (244 mg, 0.91 mmol). Colorless oil. Elute: Hex/AcOEt = 30:1. ^1H NMR (600 MHz, CDCl_3) δ 7.30–7.27 (4H, m), 7.22–7.18 (4H, m), 7.14 (2H, app. d, J = 7.6 Hz), 4.08 (2H, t, J = 6.6 Hz), 2.95 (2H, t, J = 7.7 Hz), 2.631 (2H, t, J = 7.2 Hz), 2.627 (2H, t, J = 7.8 Hz), 1.92 (2H, tt, J = 7.2, 6.6 Hz); ^{13}C NMR (150 MHz, CDCl_3) δ 172.9, 141.1, 140.5, 128.46, 128.38, 128.35, 128.24, 128.21, 125.9, 63.8, 35.8, 32.1, 30.9, 30.1.

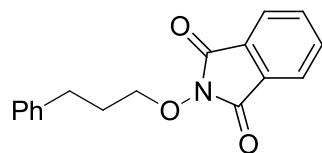


(3-Phenylpropoxy)benzene (9).¹ 3-Phenylpropanol (**6**) (136 mg, 1.0 mmol) and phenol (**7**) (104 mg, 1.1 mmol) were used as substrates. 18 h. 87% yield (184 mg, 0.87 mmol) [Fe(Pc) (1 mol%), PPh₃ (1.5 equiv): 36 h. 76% yield (161 mg, 0.76 mmol)]. Colorless oil. Elute: Hex/AcOEt = 20:1. ^1H NMR (600 MHz, CDCl_3) δ 7.30–7.25 (4H, m), 7.22–7.19 (3H, m), 6.93 (1H, app. t, J = 7.2 Hz), 6.90–6.89 (2H, m), 3.95 (2H, t, J = 6.6 Hz), 2.80 (2H, t, J = 7.7 Hz), 2.10 (2H, tt, J = 7.7, 6.6 Hz); ^{13}C NMR (150 MHz, CDCl_3) δ 159.0, 141.5, 129.4, 128.5, 128.4, 125.9, 120.5, 114.5, 66.7, 32.1, 30.8.

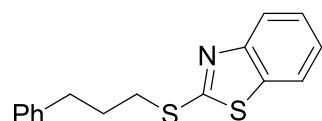


1-Iodo-4-(3-phenylpropoxy)benzene (16).⁴ 3-Phenylpropanol (**6**) (136 mg, 1.0 mmol) and 4-iodophenol (242 mg, 1.1 mmol) were used as substrates. 24 h. 80% yield (270 mg, 0.80 mmol). Colorless oil. Elute: Hex/AcOEt = 20:1. ^1H NMR (600 MHz, CDCl_3) δ 7.53 (2H, app. d, J = 9.7, Hz), 7.28 (2H, app. t, J = 7.6 Hz), 7.20–7.18 (3H, m), 6.65 (2H, app. d, J = 9.6 Hz), 3.90 (2H, t,

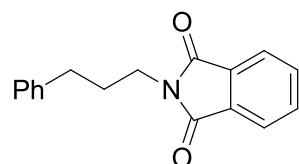
J = 6.6 Hz), 2.79 (2H, t, *J* = 7.6 Hz), 2.08 (2H, tt, *J* = 7.6, 6.6 Hz); ^{13}C NMR (150 MHz, CDCl_3) δ 158.8, 141.3, 138.1, 128.5, 128.4, 126.0, 116.9, 82.5, 66.9, 32.0, 30.6.



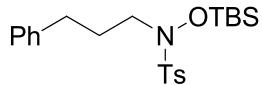
2-(3-Phenylpropoxy)isoindoline-1,3-dione (17).¹ 3-Phenylpropanol (**6**) (136 mg, 1.0 mmol) and *N*-hydroxyphthalimide (179 mg, 1.1 mmol) were used as substrates. 48 h. 85% yield (237 mg, 0.85 mmol) [0.5 M at 65 °C: 12 h. 85% yield (238 mg, 0.85 mmol)]. Pale yellow solid. Elute: Hex/AcOEt = 6:1. ^1H NMR (600 MHz, CDCl_3) δ 7.83–7.82 (m, 2H), 7.74–7.73 (m, 2H), 7.31–7.26 (4H, m), 7.19 (1H, app. t, *J* = 7.0 Hz), 4.22 (2H, t, *J* = 6.6 Hz), 2.87 (2H, t, *J* = 7.8 Hz), 2.10 (2H, tt, *J* = 7.8, 6.6 Hz); ^{13}C NMR (150 MHz, CDCl_3) δ 163.6, 141.1, 134.4, 128.9, 128.6, 128.4, 126.0, 123.4, 77.5, 31.7, 29.9.



2-((3-Phenylpropyl)thio)benzo[d]thiazole (18). 3-Phenylpropanol (**6**) (136 mg, 1.0 mmol) and 2-mercaptopbenzothiazole (184 mg, 1.1 mmol) were used as substrates. 24 h. 89% yield (253 mg, 0.89 mmol). Colorless oil. Elute: Hex/AcOEt=20:1. ^1H NMR (600 MHz, CDCl_3) δ 7.85 (1H, app. d, *J* = 7.9 Hz), 7.73 (1H, app. d, *J* = 7.9 Hz), 7.39 (1H, app. t, *J* = 7.7 Hz), 7.30–7.26 (3H, m), 7.25–7.18 (3H, m), 3.33 (2H, t, *J* = 7.2 Hz), 2.80 (2H, t, *J* = 7.2 Hz), 2.15 (2H, quint, *J* = 7.2 Hz); ^{13}C NMR (150 MHz, CDCl_3) δ 166.9, 153.3, 140.8, 135.1, 128.46, 128.42, 126.1, 125.9, 124.1, 121.4, 120.9, 34.6, 32.8, 30.7; IR (neat, cm^{-1}) ν 2928, 1456, 1423; HRMS (DART+) (*m/z*): $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{16}\text{H}_{16}\text{NS}_2$: 286.0724, found: 286.0720.

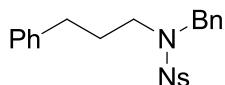


2-(3-Phenylpropyl)isoindole-1,3-dione (10).¹ 3-Phenylpropanol (**6**) (136 mg, 1.0 mmol) and phthalimide (**8**) (162 mg, 1.1 mmol) were used as substrates. 12 h (0.5 M at 65 °C). 89% yield (234 mg, 0.89 mmol) [General procedure: 48 h. 84% yield (221 mg, 0.84 mmol)]. Pale yellow oil, Elute: Hex/AcOEt = 10:1. ^1H NMR (500 MHz, CDCl_3) δ 7.82–7.81 (m, 2H), 7.70–7.68 (m, 2H), 7.24 (2H, app. t, *J* = 7.4 Hz), 7.19 (2H, app. d, *J* = 6.9 Hz), 7.14 (1H, app. t, *J* = 7.2 Hz), 3.74 (2H, t, *J* = 7.5 Hz), 2.68 (2H, t, *J* = 8.0 Hz), 2.03 (2H, tt, *J* = 8.0, 7.5 Hz); ^{13}C NMR (125 MHz, CDCl_3) δ 168.3, 141.0, 133.8, 132.0, 128.3, 128.2, 125.9, 123.1, 37.7, 33.1, 29.8.

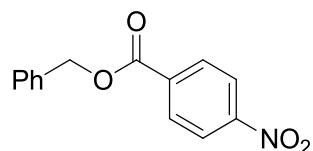


3-Phenylpropyl-*O*-(*tert*-butyldimethylsilyl)-*N*-tosylhydroxylamine (19).⁵

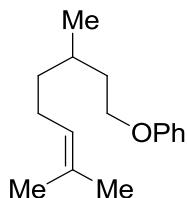
3-Phenylpropanol (**6**) (136 mg, 1.0 mmol) and *O*-(*tert*-butyldimethylsilyl)-*N*-tosylhydroxylamine (332 mg, 1.1 mmol) were used as substrates. 60 h. 87% yield (364 mg, 0.87 mmol). Colorless oil. Elute: Hex/AcOEt = 20:1. ¹H NMR (600 MHz, CDCl₃) δ 7.69 (2H, app. d, *J* = 8.2 Hz), 7.32 (2H, app. d, *J* = 8.2 Hz), 7.27 (2H, app. t, *J* = 7.6 Hz), 7.18 (1H, app. t, *J* = 7.9 Hz), 7.14–7.13 (2H, m), 2.94 (2H, t, *J* = 7.0 Hz), 2.62 (2H, t, *J* = 7.8 Hz), 2.44 (3H, s), 1.89 (2H, tt, *J* = 7.8, 7.2 Hz), 0.91 (9H, s), 0.28 (6H, s); ¹³C NMR (150 MHz, CDCl₃) δ 144.5, 141.1, 129.9, 129.8, 129.2, 128.39, 128.37, 126.0, 55.4, 33.1, 28.5, 26.0, 21.6, 18.2, –4.3.



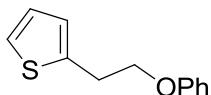
N-Benzyl-4-nitro-*N*-(3-phenylpropyl)benzenesulfonamide (20).¹ 3-Phenylpropanol (**6**) (136 mg, 1.0 mmol) and *N*-benzyl-2-nitrobenzenesulfonylamide (321 mg, 1.1 mmol) were used as substrates. 12 h (0.5 M at 65 °C). 90% yield (367 mg, 0.90 mmol) [General procedure: 48 h. 60% yield (245 mg, 0.60 mmol)]. Pale yellow oil. Elute: Hex/AcOEt = 5:1. ¹H NMR (600 MHz, CDCl₃) δ 7.91 (d, *J* = 7.9 Hz, 1H), 7.68–7.58 (m, 3H), 7.30–7.15 (m, 8H), 6.98 (d, *J* = 7.6 Hz, 2H), 4.51 (s, 2H), 3.25 (t, *J* = 7.6 Hz, 2H), 2.43 (t, *J* = 7.8 Hz, 2H), 1.70 (tt, *J* = 7.8, 7.6 Hz, 2H); ¹³C NMR (150 MHz, CDCl₃) δ 147.9, 140.9, 135.6, 133.7, 133.4, 131.6, 130.9, 128.7, 128.4, 128.3, 128.2, 127.9, 126.0, 124.2, 51.3, 46.7, 32.6, 29.0. [Note: One peak (46.7 ppm) has been inadvertently omitted from the list of the ¹³C NMR data in the previous report¹]. HRMS (DART+) (*m/z*): [M+H]⁺ calcd for C₂₂H₂₃N₂O₄S: 411.1379, found: 411.1369. [Note: Since the HRMS data was incorrect in the previous report (the value based on [M–Ns]⁺ was inadvertently described),¹ reanalysis of HRMS was conducted]



Benzyl-4-nitrobenzoate (21).³ Benzyl alcohol (108 mg, 1.0 mmol) and 4-nitrobenzoic acid (**4**) (184 mg, 1.1 mmol) were used as substrates. 12 h. 90% yield (230 mg, 0.90 mmol). Pale yellow oil. Elute: Hex/AcOEt = 10:1. ¹H NMR (600 MHz, CDCl₃) δ 8.28 (2H, app. d, *J* = 9.0 Hz), 8.24 (2H, app. d, *J* = 9.0 Hz), 7.46 (2H, d, *J* = 7.2 Hz), 7.43–7.38 (3H, m), 5.41 (2H, s); ¹³C NMR (150 MHz, CDCl₃) δ 164.5, 150.6, 135.48, 135.22, 130.8, 128.73, 128.65, 128.44, 123.5, 67.6.



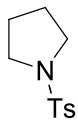
((3,7-Dimethyloct-6-en-1-yl)oxy)benzene (22). (\pm)-Citronellol (156 mg, 1.0 mmol) and phenol (7) (104 mg, 1.1 mmol) were used as substrates. 24 h. 77% yield (177 mg, 0.77 mmol). Colorless oil. Elute: Hex/AcOEt = 15:1. ^1H NMR (600 MHz, CDCl_3) δ 7.28–7.25 (2H, m), 6.94–6.89 (3H, m), 5.11 (1H, app. t, J = 7.2 Hz), 4.02–3.95 (2H, m), 2.07–1.95 (2H, m), 1.87–1.81 (1H, m), 1.71–1.68 (1H, m), 1.69 (3H, s), 1.62–1.58 (1H, m), 1.61 (3H, s), 1.41–1.38 (1H, m), 1.25–1.19 (1H, m), 0.95 (3H, d, J = 6.5 Hz); ^{13}C NMR (150 MHz, CDCl_3) δ 159.1, 131.3, 129.4, 124.7, 120.4, 114.5, 66.1, 37.1, 36.1, 29.5, 25.7, 25.4, 19.5, 17.7; IR (neat, cm^{-1}) ν 2923, 1600, 1498, 1243; HRMS (DART+) (m/z): [M+H] $^+$ calcd for $\text{C}_{16}\text{H}_{25}\text{O}$: 233.1905, found: 233.1894.



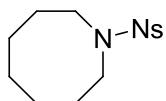
2-(2-Phenoxyethyl)thiophene (23). 3-Thiopheneethanol (128 mg, 1.0 mmol) and phenol (7) (104 mg, 1.1 mmol) were used as substrates. 18 h. 82% yield (167 mg, 0.82 mmol). colorless oil. Elute: Hex/AcOEt = 30:1. ^1H NMR (600 MHz, CDCl_3) δ 7.29–7.26 (2H, m), 7.15 (1H, dd, J = 5.2, 1.4 Hz), 6.96–6.91 (5H, m), 4.18 (2H, t, J = 6.7 Hz), 3.30 (2H, app. t, J = 6.9 Hz); ^{13}C NMR (150 MHz, CDCl_3) δ 158.6, 140.4, 129.5, 126.8, 125.5, 123.9, 120.9, 114.6, 68.3, 30.0; IR (neat, cm^{-1}) ν 2924, 1596, 1497, 1243; HRMS (DART+) (m/z): [M+H] $^+$ calcd for $\text{C}_{12}\text{H}_{13}\text{OS}$: 205.0687, found: 205.0689.



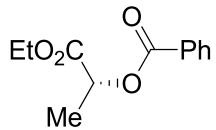
2-((2-(1H-Indol-3-yl)ethyl)thio)benzo[d]thiazole (24).⁶ 3-Indoleethanol (161 mg, 1.0 mmol) and 2-mercaptopbenzothiazole (184 mg, 1.1 mmol) were used as substrates. 24 h. 96% yield (296 mg, 0.96 mmol). White solid. Elute: Hex/AcOEt = 10:1. ^1H NMR (600 MHz, CDCl_3) δ 8.07 (1H, br s), 7.90 (1H, d, J = 8.6 Hz), 7.75 (2H, d, J = 7.6 Hz), 7.41 (1H, app. t, J = 7.7 Hz), 7.35 (1H, d, J = 7.9 Hz), 7.29 (1H, app. t, J = 7.6 Hz), 7.21 (1H, app. t, J = 7.6, Hz), 7.17–7.15 (1H, m), 7.07 (1H, d, J = 2.4 Hz), 3.66 (2H, t, J = 7.8 Hz), 3.30 (2H, t, J = 7.8 Hz); ^{13}C NMR (150 MHz, CDCl_3) δ 167.1, 153.3, 136.3, 135.2, 127.1, 126.0, 124.1, 122.2, 122.1, 121.4, 120.9, 119.5, 118.9, 114.1, 111.2, 34.2, 25.5.



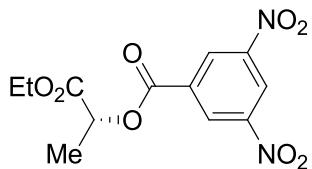
N-(*p*-Toluenesulfonyl)pyrrolidine (25).⁷ *N*-(4-Hydroxybutyl)-toluenesulfonamide⁸ (243 mg, 1.0 mmol) was used as substrates in 0.5 M. 18 h. 94% yield (211 mg, 0.94 mmol) [Fe(Pc) (1 mol%), PPh₃ (1.5 equiv): 36 h. 87% yield (196 mg, 0.87 mmol)]. White solid. Elute: Hex/AcOEt = 5:1. ¹H NMR (600 MHz, CDCl₃) δ 7.72 (2H, app. d, *J* = 8.2 Hz), 7.32 (2H, app. d, *J* = 8.2 Hz), 3.24–3.22 (4H, m), 2.43 (3H, s), 1.76–1.74 (4H, m); ¹³C NMR (150 MHz, CDCl₃) δ 143.3, 133.9, 129.6, 127.5, 47.9, 25.2, 21.5.



1-(2-Nitrobenzenesulfonyl)azocane (26).⁹ *N*-(7-Hydroxyheptyl)-2-nitrobenzenesulfonamide⁹ (316 mg, 1.0 mmol) was used as substrates in 0.05 M. 36 h. 40% yield (118 mg, 0.40 mmol). White solid. Elute: Hex/AcOEt = 5:1. ¹H NMR (500 MHz, CDCl₃) δ 7.93–7.92 (1H, m), 7.68–7.66 (2H, m), 7.59–7.58 (1H, m), 3.33 (4H, t, *J* = 6.0 Hz), 1.79–1.77 (4H, m), 1.67–1.65 (6H, m); ¹³C NMR (125 MHz, CDCl₃) δ 148.3, 133.2, 132.5, 131.3, 130.5, 123.9, 49.2, 27.7, 26.5, 24.7.

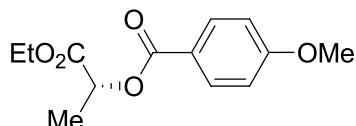


(R)-1-Ethoxycarbonylethyl benzoate (34).¹⁰ (−)-(S)-Ethyl lactate (**3**) (118 mg, 1.0 mmol) and benzoic acid (134 mg, 1.1 mmol) were used as substrates. 36 h. 84% yield (186 mg, 0.84 mmol) 99:1 er. Pale yellow solid. Elute: Hex/AcOEt=20:1. ¹H NMR (600 MHz, CDCl₃) δ 8.09 (2H, app. d, *J* = 8.2 Hz), 7.58 (1H, app. t, *J* = 7.4 Hz), 7.45 (2H, app. t, *J* = 7.7 Hz), 5.31 (1H, q, *J* = 7.0 Hz), 4.23 (2H, q, *J* = 7.0 Hz), 1.63 (3H, d, *J* = 7.0 Hz), 1.28 (3H, t, *J* = 7.0 Hz); ¹³C NMR (150 MHz, CDCl₃) δ 170.8, 165.9, 133.2, 129.8, 129.5, 128.3, 69.1, 61.3, 17.0, 14.1; The enantiomeric ratio was determined by HPLC analysis using a chiral column. Chiral HPLC: Daicel-Chiraldak OJ-H 46 × 150 mm, multi UV-Vis detector (230 nm), room temperature eluent: (*n*-hexane /*i*-PrOH) 99.5:0.5, flow rate: 1.0 mL/min, retention time (min) 22.8 (*R* isomer), 25.1 (*S* isomer). [α]_D²⁵ = −12.6 (*c* 1.00, CHCl₃) [lit.¹¹ (*R*)-1-Ethoxycarbonylethyl benzoate; [α]_D²⁵ = −13.8 (*c* 1.00, CHCl₃)], [Authentic sample (retention): (*S*)-1-Ethoxycarbonylethyl benzoate (99:1 er), [α]_D²⁵ = +16.1 (*c* 1.00, CHCl₃)].

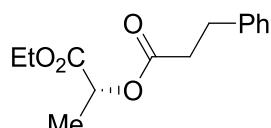


(R)-1-Ethoxycarbonylethyl 3,5-dinitrobenzoate (35).¹ (−)-(S)-Ethyl lactate (**3**) (118 mg, 1.0 mmol) and 3,5-dinitrobenzoic acid (233 mg, 1.1 mmol) were used as substrates. 18 h. 89% yield (278 mg, 0.89 mmol) 87:13 er. Pale yellow solid. Elute: Hex/AcOEt = 6:1. ¹H NMR (500 MHz, CDCl₃) δ 9.26 (1H, t, *J* = 2.0 Hz), 9.21 (2H, d, *J* = 2.3 Hz), 5.42 (1H, q, *J* = 7.0 Hz), 4.27 (2H, q, *J* = 7.2 Hz), 1.73 (3H, d, *J* = 7.5 Hz), 1.32 (3H, t, *J* = 7.2 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 169.7, 162.0, 148.6, 133.2, 129.6, 122.7, 70.8, 61.9, 16.9, 14.1; The enantiomeric ratio was determined by HPLC analysis using a chiral column. Chiral HPLC: Daicel-Chiraldak OJ-H 46 × 150 mm, multi UV-Vis detector (210 nm), room temperature eluent: (*n*-hexane /*i*-PrOH) 1:5, flow rate: 0.25 mL/min, retention time (min) 51.0 (*S* isomer), 61.1 (*R* isomer). [α]_D²⁵ = −3.5 (c 1.00, CHCl₃) [lit.¹ (*S*)-1-ethoxycarbonylethyl 3,5-dinitrobenzoate (>99:1 er), [α]_D²² = +8.8 (c 1.00, CHCl₃)].

For data of the reaction at 65 °C in THF (0.5 M), see the Supporting Information in ref. 1.

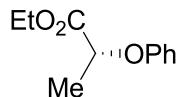


(R)-1-Ethoxycarbonylethyl 4-methoxybenzoate (36). (−)-(S)-Ethyl lactate (**3**) (118 mg, 1.0 mmol) and 4-methoxybenzoic acid (167 mg, 1.1 mmol) were used as substrates. 36 h. 79% yield (197 mg, 0.79 mmol) 99:1 er. Colorless oil. Elute: Hex/AcOEt = 10:1. ¹H NMR (600 MHz, CDCl₃) δ 8.04 (2H, app. d, *J* = 8.9 Hz), 6.93 (2H, app. d, *J* = 8.9 Hz), 5.28 (1H, q, *J* = 6.6 Hz), 4.23 (2H, q, *J* = 7.2 Hz), 3.86 (3H, s), 1.61 (3H, d, *J* = 6.6 Hz), 1.28 (3H, t, *J* = 7.2 Hz); ¹³C NMR (150 MHz, CDCl₃) δ 171.0, 165.6, 163.6, 131.9, 121.8, 113.6, 68.9, 61.3, 55.4, 17.1, 14.1; IR (CHCl₃, cm^{−1}) ν 2987, 1717, 1608, 1513, 1457, 1259; HRMS (DART+) (*m/z*): [M+H]⁺ calcd for C₁₃H₁₇O₅: 253.1076, found: 253.1076. The enantiomeric ratio was determined by HPLC analysis using a chiral column. Chiral HPLC: Daicel-Chiraldak OD-H 46 × 150 mm, multi UV-Vis detector (250 nm), room temperature eluent: (*n*-hexane /*i*-PrOH) 99.5:0.5, flow rate: 1.0 mL/min, retention time (min) 30.2 (*S* isomer), 34.0 (*R* isomer). [α]_D²⁵ = −29.4 (c 1.00, CHCl₃) [Authentic sample (retention): (*S*)-1-Ethoxycarbonylethyl 4-methoxybenzoate (99:1 er), [α]_D²⁵ = +32.6 (c 1.00, CHCl₃)].

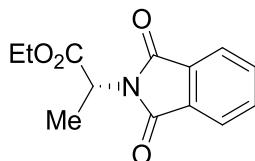


(R)-1-Ethoxy-1-oxopropan-2-yl 3-phenylpropanoate (37). (−)-(S)-Ethyl lactate (**3**) (118 mg, 1.0 mmol) and 3-phenylpropionic acid (165 mg, 1.1 mmol) were used as substrates, and the reaction was performed at 0 °C. 96 h. 61% yield (185 mg, 0.75 mmol) 90:10 er [General

procedure: 42 h. 75% yield (185 mg, 0.75 mmol), 78:22 er]. Colorless oil. Elute: Hex/AcOEt = 10:1. ^1H NMR (600 MHz, CDCl_3) δ 7.29 (2H, app. t, J = 7.4 Hz), 7.22–7.19 (3H, m), 5.07 (1H, q, J = 7.2 Hz), 4.19 (2H, q, J = 7.2 Hz), 2.98 (2H, t, J = 7.4 Hz), 2.77–2.67 (2H, m), 1.47 (3H, d, J = 7.2 Hz), 1.26 (3H, t, J = 7.2 Hz); ^{13}C NMR (150 MHz, CDCl_3) δ 172.2, 170.8, 140.3, 128.4, 128.3, 126.2, 68.6, 61.3, 35.5, 30.7, 16.9, 14.0; IR (neat, cm^{-1}) ν 2987, 1734, 1455, 1374; HRMS (DART+) (m/z): $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{14}\text{H}_{19}\text{O}_4$: 251.1283, found: 251.1284. The enantiomeric ratio was determined by HPLC analysis using a chiral column. Chiral HPLC: Daicel-Chiraldak OD-H 46 × 150 mm, multi UV-Vis detector (210 nm), room temperature eluent: (*n*-hexane/*i*-PrOH) 99.5:0.5, flow rate: 1.0 mL/min, retention time (min) 32.8 (*S* isomer), 41.1 (*R* isomer). $[\alpha]_D^{25} = +26.6$ (c 1.00, CHCl_3) [Authentic sample (retention): (*S*)-1-Ethoxy-1-oxopropan-2-yl 3-phenylpropanoate (99:1 er), $[\alpha]_D^{25} = -29.5$ (c 1.00, CHCl_3)].

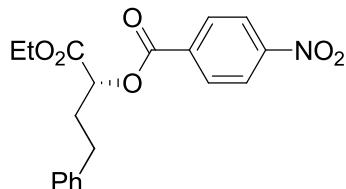


Ethyl (*R*)-2-phenoxypropanoate (38).¹² (–)(*S*)-Ethyl lactate (**3**) (118 mg, 1.0 mmol) and phenol (**7**) (104 mg, 1.1 mmol) were used as substrates. 48 h. 63% yield (121 mg, 0.63 mmol) 99:1 er. Colorless oil. Elute: Hex/AcOEt = 8:1. ^1H NMR (600 MHz, CDCl_3) δ 7.28–7.25 (2H, m), 6.97 (1H, app. t, J = 7.4 Hz), 6.88 (2H, app. d, J = 7.9 Hz), 4.74 (1H, q, J = 7.2 Hz), 4.22 (2H, q, J = 7.2 Hz), 1.62 (3H, d, J = 6.6 Hz), 1.24 (3H, t, J = 7.2 Hz); ^{13}C NMR (150 MHz, CDCl_3) δ 172.2, 157.5, 129.5, 121.5, 115.0, 72.5, 61.2, 18.5, 14.1; The enantiomeric ratio was determined by HPLC analysis using a chiral column. Chiral HPLC: Daicel-Chiraldak OD-H 46 × 150 mm, multi UV-Vis detector (275 nm), room temperature eluent: (*n*-hexane/*i*-PrOH) 90:10, flow rate: 1.0 mL/min, retention time (min) 4.7 (*S* isomer), 7.3 (*R* isomer). $[\alpha]_D^{25} = +50.9$ (c 1.00, MeOH) [lit.¹² ethyl (*R*)-2-phenoxypropanoate, $[\alpha]_D^{18} = +47.2$ (c 0.50, MeOH)].

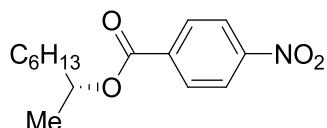


Ethyl (*R*)-2-(1,3-dioxoisodolin-2-yl)propanoate (39).¹³ (–)(*S*)-Ethyl lactate (**3**) (118 mg, 1.0 mmol) and phthalimide (**8**) (162 mg, 1.1 mmol) were used as substrates in 0.5 M. 48 h. 49% yield (121 mg, 0.49 mmol) 99:1 er. white solid. Elute: Hex/AcOEt=10:1. ^1H NMR (500 MHz, CDCl_3) δ 7.87 (2H, app. dd, J = 5.0, 3.5 Hz), 7.75 (2H, app. dd, J = 5.0, 3.5 Hz), 4.97 (1H, q, J = 7.5 Hz), 4.26–4.17 (2H, m), 1.70 (3H, d, J = 6.5 Hz), 1.24 (3H, t, J = 7.5 Hz); ^{13}C NMR (125 MHz, CDCl_3) δ 169.7, 167.4, 134.1, 131.9, 123.5, 61.8, 47.5, 15.2, 14.1; The enantiomeric ratio was determined by HPLC analysis using a chiral column. Chiral HPLC: Daicel-Chiraldak OD-H 46 × 150

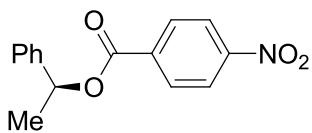
mm, multi UV-Vis detector (210 nm), room temperature eluent: (*n*-hexane/*i*-PrOH) 99.5:0.5, flow rate: 0.5 mL/min, retention time (min) 38.6 (*S* isomer), 40.9 (*R* isomer). $[\alpha]_D^{25} = +18.4$ (*c* 1.00, MeOH) [lit.¹³ ethyl (*R*)-2-(1,3-dioxoisooindolin-2-yl)propanoate, $[\alpha]_D^{25} = +18.2$ (*c* 1.00, MeOH)].



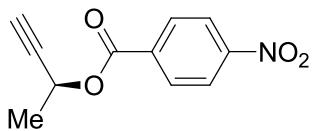
(*S*)-1-Ethoxy-1-oxo-4-phenylbutan-2-yl 4-nitrobenzoate (40). Ethyl (*–*)(*R*)-2-hydroxy-4-phenylbutyrate (**27**) (208 mg, 1.0 mmol, 99:1 er) and 4-nitrobenzoic acid (**4**) (184 mg, 1.1 mmol) were used as substrates. 24 h. 93% yield (332 mg, 0.93 mmol) 99:1 er. Pale yellow oil. Elute: Hex/AcOEt = 15:1. ^1H NMR (600 MHz, CDCl_3) δ 8.29 (2H, app. d, *J* = 8.9 Hz), 8.17 (2H, app. d, *J* = 8.9 Hz), 7.31–7.29 (2H, m), 7.23–7.20 (3H, m), 5.28 (1H, t, *J* = 6.6 Hz), 4.24 (2H, q, *J* = 7.2 Hz), 2.85 (2H, t, *J* = 7.8 Hz), 2.36 (2H, dt, *J* = 7.8, 6.6 Hz), 1.29 (3H, t, *J* = 7.2 Hz); ^{13}C NMR (150 MHz, CDCl_3) δ 169.5, 164.1, 150.7, 140.2, 134.8, 131.0, 128.6, 128.4, 126.3, 123.5, 73.1, 61.7, 32.5, 31.5, 14.1; IR (neat, cm^{-1}) ν 2983, 1733, 1530, 1276; HRMS (DART+) (*m/z*): calcd for $[\text{M}+\text{NH}_4]^+$ $\text{C}_{15}\text{H}_{21}\text{N}_2\text{O}_6$: 375.1556, found: 375.1563; The enantiomeric ratio was determined by HPLC analysis using a chiral column. Chiral HPLC: Daicel-Chiraldak OJ-H 46 × 150 mm, multi UV-Vis detector (210 nm), room temperature eluent: (*n*-hexane/*i*-PrOH) 5:1, flow rate: 1.0 mL/min, retention time (min) 52.4 (*S* isomer), 25.7 (*R* isomer). $[\alpha]_D^{25} = -16.5$ (*c* 1.00, CHCl_3) [Authentic sample (retention): (*R*)-1-ethoxy-1-oxo-4-phenylbutan-2-yl 4-nitrobenzoate (99:1 er), $[\alpha]_D^{25} = +19.6$ (*c* 1.00, CHCl_3)].



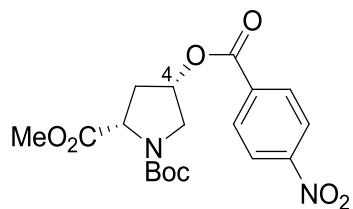
(*R*)-2-Octyl 4-nitrobenzoate (41).¹⁴ (+)-(S)-2-Octanol (**28**) (130 mg, 1.0 mmol, 99:1 er) and 4-nitrobenzoic acid (**4**) (184 mg, 1.1 mmol) were used as substrates. 12 h. 91% yield (252 mg, 0.91 mmol) 99:1 er. Pale yellow oil. Elute: Hex/AcOEt = 30:1. ^1H NMR (600 MHz, CDCl_3) δ 8.29 (2H, app. d, *J* = 8.9 Hz), 8.21 (2H, app. d, *J* = 8.9 Hz), 5.20–5.18 (1H, m), 1.79–1.74 (1H, m), 1.67–1.60 (1H, m), 1.42–1.26 (8H, m), 1.38 (3H, d, *J* = 7.2 Hz), 0.88 (3H, t, *J* = 7.2 Hz); ^{13}C NMR (150 MHz, CDCl_3) δ 164.3, 150.4, 136.3, 130.6, 123.4, 73.1, 35.9, 31.7, 29.1, 25.4, 22.5, 20.0, 14.0; The enantiomeric ratio was determined by HPLC analysis using a chiral column. Chiral HPLC: Daicel-Chiraldak OJ-H 46 × 150 mm, multi UV-Vis detector (230 nm), room temperature eluent: (*n*-hexane/*i*-PrOH) 99.5:0.5, flow rate: 0.5 mL/min, retention time (min) 15.7 (*S* isomer), 19.9 (*R* isomer). $[\alpha]_D^{25} = -33.8$ (*c* 1.00, CHCl_3) [lit.¹⁵ (*R*)-2-Octyl 4-nitrobenzoate, $[\alpha]_D^{23} = -34.2$ (*c* 2.21, CHCl_3); Authentic sample (retention): (*S*)-2-Octyl 4-nitrobenzoate (99:1 er), $[\alpha]_D^{25} = +39.6$ (*c* 1.00, CHCl_3)].



(S)-1-Phenylethyl 4-nitrobenzoate (42).¹⁶ (+)-(R)-1-Phenyl-1-ethanol (**29**) (122 mg, 1.0 mmol, 98:2 er) and 4-nitrobenzoic acid (**4**) (184 mg, 1.1 mmol) were used as substrates. 12 h. 89% yield (241 mg, 0.89 mmol) 92:8 er. Colorless oil. Elute: Hex/AcOEt = 30:1. ¹H NMR (600 MHz, CDCl₃) δ 8.28 (2H, app. d, *J* = 8.9 Hz), 8.23 (2H, app. d, *J* = 8.9 Hz), 7.45–7.44 (2H, m), 7.39 (2H, app. t, *J* = 7.6 Hz), 7.34 (1H, app. t, *J* = 7.2 Hz), 6.16 (1H, q, *J* = 6.5 Hz), 1.71 (3H, d, *J* = 6.5 Hz); ¹³C NMR (150 MHz, CDCl₃) δ 163.9, 150.5, 140.9, 135.8, 130.7, 128.7, 128.2, 126.1, 123.5, 74.2, 22.2; The enantiomeric ratio was determined by HPLC analysis using a chiral column. Chiral HPLC: Daicel-Chiralpak OJ-H 46 × 150 mm, multi UV-Vis detector (210 nm), room temperature eluent: (*n*-hexane/*i*-PrOH) 1:1 flow rate: 0.5 mL/min, retention time (min) 22.3 (*R* isomer), 26.5 (*S* isomer). [α]_D²⁵ = +43.9 (*c* 1.00, CHCl₃) [Authentic sample (retention): (*R*)-1-Phenylethyl 4-nitrobenzoate (98:2 er), [α]_D²⁵ = −49.0 (*c* 1.00, CHCl₃); lit.¹⁵ (*R*)-1-Phenylethyl 4-nitrobenzoate, [α]_D²³ = −50.5 (*c* 1.29, CHCl₃)].



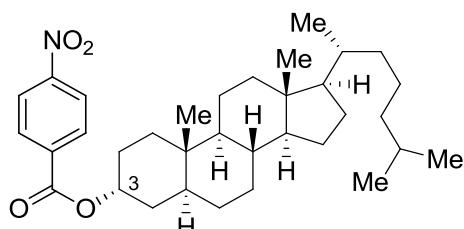
(S)-3-Butynyl 4-nitrobenzoate (43).¹⁷ (+)-(R)-3-Butyn-2-ol (**30**) (70.1 mg, 1.0 mmol, 99:1 er) and 4-nitrobenzoic acid (**4**) (184 mg, 1.1 mmol) were used as substrates. 12 h. 87% yield (189 mg, 0.87 mmol) 96:4 er. White solid. Elute: Hex/AcOEt = 30:1. ¹H NMR (600 MHz, CDCl₃) δ 8.30 (2H, app. d, *J* = 9.3 Hz), 8.24 (2H, app. d, *J* = 9.3 Hz), 5.71 (1H, qd, *J* = 6.6, 1.8 Hz), 2.54 (1H, d, *J* = 1.8 Hz), 1.68 (3H, d, *J* = 6.6 Hz); ¹³C NMR (150 MHz, CDCl₃) δ 163.6, 150.7, 135.1, 130.9, 123.6, 81.4, 73.7, 61.7, 21.2; The enantiomeric ratio was determined by HPLC analysis using a chiral column. Chiral HPLC: Daicel-Chiralpak OJ-H 46 × 150 mm, multi UV-Vis detector (230 nm), room temperature eluent: (*n*-hexane/*i*-PrOH) 5:1, flow rate: 1.0 mL/min, retention time (min) 12.4 (*R* isomer), 25.4 (*S* isomer). [α]_D²⁵ = −9.2 (*c* 1.00, CHCl₃) [Authentic sample (retention): (*R*)-3-Butynyl 4-nitrobenzoate (99:1 er); [α]_D²⁵ = +14.4 (*c* 1.00, CHCl₃)].



1-(tert-butyl) 2-methyl (2S,4S)-4-((4-nitrobenzoyl)oxy)pyrrolidine-1,2-dicarboxylate (44).¹⁸ (2S,4R)-4-Hydroxypyrrolidine-1,2-dicarboxylic acid 1-*tert*-butyl 2-methyl ester (**31**) (245 mg, 1.0

mmol) and 4-nitrobenzoic acid (**4**) (184 mg, 1.1 mmol) were used as substrates. 16 h. 88% yield (346 mg, 0.88 mmol, as a single diastereomer in NMR analysis). [Fe(Pc) (3 mol%), PPh₃ (1.5 equiv); 36 h. 91% yield (359 mg, 0.91 mmol), as a single diastereomer in NMR analysis]. White solid. Elute: Hex/AcOEt=3:1. Rotamers were observed in NMR spectra. ¹H NMR (600 MHz, CDCl₃) δ 8.29 (2H, app. d, *J* = 8.6 Hz), 8.16 (2H, app. d, *J* = 8.6 Hz), 5.58 (1H, app. d, *J* = 4.1 Hz), 4.62 and 4.56 (total 1H, both d, both *J* = 8.4 Hz, based on rotamers), 3.85–3.82 (2H, m), 3.71 and 3.70 (total 3H, both s, based on rotamers), 2.64–2.53 (1H, m), 2.48 and 2.45 (total 1H, both s, based on rotamers), 1.49 and 1.45 (total 9H, both s, based on rotamers); ¹³C NMR (150 MHz, CDCl₃) δ 172.3, 172.1, 164.0, 163.8, 154.0, 153.6, 150.7, 135.0, 130.8, 123.5, 80.6, 80.5, 74.5, 73.4, 57.7, 57.4, 52.5, 52.3, 52.2, 52.1, 36.5, 35.6, 28.3, 28.2.

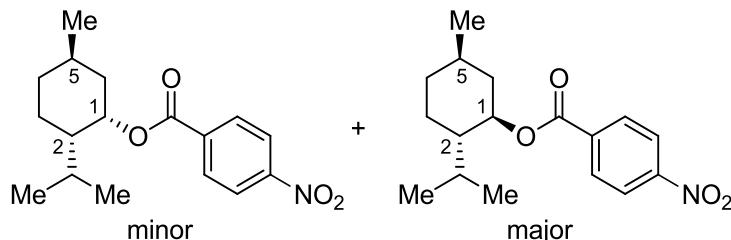
[The authentic sample of 1-(*tert*-butyl)-2-methyl-(2*S*,4*R*)-4-((4-nitrobenzoyl)oxy)pyrrolidine-1,2-dicarboxylate (*epi*-**44**) was prepared by condensation between (2*S*,4*R*)-4-hydroxypyrrrolidine-1,2-dicarboxylic acid 1-*tert*-butyl 2-methyl ester (**31**) and 4-nitrobenzoic acid (**4**) with EDC·HCl. ¹H NMR (600 MHz, CDCl₃) δ 8.30 (2H, app. d, *J* = 9.0 Hz), 8.19 (2H, app. d, *J* = 9.0 Hz), 5.58–5.57 (1H, m), 4.54 and 4.45 (total 1H, both t, both *J* = 7.8 Hz, based on rotamers), 3.88, 3.86 and 3.71 (total 2H, dd, *J* = 6.6, 4.2 Hz, d, *J* = 3.0 Hz and d, *J* = 12.6 Hz, based on rotamers), 3.79 and 3.78 (total 3H, both s, based on rotamers), 2.60–2.53 (1H, m), 2.38–2.34 (1H, m), 1.47 and 1.44 (total 9H, both s, based on rotamers). No peak on C3 (δ 2.38–2.34 (1H, m)) of 1-(*tert*-butyl)-2-methyl-(2*S*,4*R*)-4-((4-nitrobenzoyl)oxy)pyrrolidine-1,2-dicarboxylate was detected in ¹H NMR of the crude product including **44**]



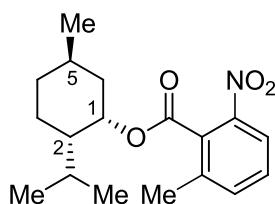
(3S)-5α-Cholestan-3-yl 4-nitrobenzoate (45).¹⁴ Dihydrocholesterol (**32**) (389 mg, 1.0 mmol) and 4-nitrobenzoic acid (**4**) (184 mg, 1.1 mmol) were used as substrates. 24 h. 82% yield (439 mg, 0.82 mmol, as a single diastereomer). Pale yellow solid. Elute: Hex/AcOEt = 25:1. ¹H NMR (600 MHz, CDCl₃) δ 8.31 (2H, app. d, *J* = 8.9 Hz), 8.22 (2H, app. d, *J* = 8.9 Hz), 5.32–5.31 (1H, m), 1.99 (1H, dt, *J* = 12.5, 3.2 Hz), 1.88 (1H, br d, *J* = 12.6 Hz), 1.86–1.80 (2H, m), 1.70–1.50 (8H, m), 1.40–0.92 (18H, m), 0.91 (3H, d, *J* = 6.6 Hz), 0.87 (3H, d, *J* = 6.6 Hz), 0.86 (3H, d, *J* = 7.2 Hz), 0.85 (3H, s), 0.78 (1H, dt, *J* = 16.2, 6.3 Hz), 0.67 (3H, s); ¹³C NMR (150 MHz, CDCl₃) δ 164.0, 150.4, 136.6, 130.6, 123.5, 72.2, 56.5, 56.3, 54.4, 42.6, 40.5, 40.0, 39.5, 36.2, 35.9, 35.8, 35.4, 33.2, 32.9, 31.9, 28.4, 28.2, 28.0, 26.2, 24.1, 23.8, 22.8, 22.5, 20.8, 18.7, 12.1, 11.4.

[The authentic sample of (3*R*)-5α-cholestan-3-yl 4-nitrobenzoate (*epi*-**45**) was prepared by condensation of dihydrocholesterol (**32**) and 4-nitrobenzoic acid (**4**) with EDC·HCl. ¹H NMR

(600 MHz, CDCl₃) δ 8.27 (2H, app. d, *J* = 8.4 Hz), 8.21 (2H, app. d, *J* = 8.4 Hz), 5.02–4.94 (1H, m), 1.99–1.97 (2H, m), 1.82–1.79 (2H, m), 1.73–1.66 (3H, m), 1.57–1.49 (3H, m), 1.37–1.23 (9H, m), 1.16–0.98 (9H, m), 0.91 (3H, d, *J* = 6.6 Hz), 0.89–0.86 (9H, m), 0.69–0.68 (1H, m), 0.66 (3H, s). No peak on C3 (δ 5.02–4.94 (1H, m)) of (3*R*)-5α-cholestane-3-yl 4-nitrobenzoate was detected in ¹H NMR of the crude product including **45**]



(1S,2S,5R)-Menthyl 4-nitrobenzoate (46).¹⁴ (−)-Menthol (**33**) (156 mg, 1.0 mmol) and 4-nitrobenzoic acid (**4**) (184 mg, 1.1 mmol) were used as substrates. 48 h. 76% yield (230 mg, 0.76 mmol, as an inseparable mixture of two diastereomers, 11:89 dr in NMR analysis). White solid. Elute: Hex/AcOEt = 30:1. ¹H NMR (600 MHz, CDCl₃, for major isomer) δ 8.29 (2H, app. d, *J* = 8.9 Hz), 8.21 (2H, app. d, *J* = 8.9 Hz), 4.98 (1H, td, *J* = 10.8, 4.2 Hz), 2.15–2.12 (1H, m), 1.95–1.90 (1H, m), 1.77–1.73 (2H, m), 1.62–1.55 (2H, m), 1.19–1.11 (2H, m), 0.95 (3H, d, *J* = 6.6 Hz), 0.93 (3H, d, *J* = 7.2 Hz), 0.80 (3H, d, *J* = 6.6 Hz); ¹H NMR (600 MHz, CDCl₃, for partial peaks of minor isomer) δ 5.50 (1H, app. d, *J* = 2.4 Hz); ¹³C NMR (150 MHz, CDCl₃) δ 164.2 (major), 164.0 (minor), 150.4 (major), 136.3 (minor), 136.2 (major), 130.62 (major), 130.58 (minor), 123.53 (minor), 123.46 (major), 76.1 (major), 73.1 (minor), 47.1 (major), 46.9 (minor), 40.8 (major), 39.1 (minor), 34.7 (minor), 34.2 (major), 31.4 (major), 29.4 (minor), 26.8 (minor), 26.5 (major), 25.3 (minor), 23.5 (major), 22.1 (minor), 22.0 (major), 20.9 (minor), 20.74 (minor), 20.70 (major), 16.4 (major); The diastereomeric ratio was determined from integration values of peaks of C1 [4.98 (1H, td, *J* = 10.8, 4.2 Hz, for major isomer), 5.50 (1H, app. d, *J* = 2.4 Hz, for minor isomer)].



(1S,2S,5R)-Menthyl 2-methyl-6-nitrobenzoate (47). (−)-Menthol (**33**) (156 mg, 1.0 mmol) and 2-methyl-6-nitrobenzoic acid (199 mg, 1.1 mmol) were used as substrates. 96 h. 66% yield (195 mg, 0.66 mmol, as a single diastereomer in NMR analysis). Colorless crystals, mp 66.5–67.0 °C (n-hexane/EtOAc). Elute: Hex/AcOEt = 15:1. ¹H NMR (600 MHz, CDCl₃) δ 7.98 (1H, d, *J* = 7.8 Hz), 7.53 (1H, d, *J* = 7.2 Hz), 7.45 (1H, t, *J* = 7.8 Hz), 5.567 (1H, app. d, *J* = 2.1 Hz), 2.43 (3H, s), 2.37–2.32 (1H, m), 1.75–1.69 (2H, m), 1.62–1.57 (1H, m), 1.51–1.45 (1H, m), 1.22 (1H, qd, *J* =

12.6, 3.6 Hz), 1.14 (1H, ddd, J = 15.0, 13.2, 2.4 Hz), 1.08–1.03 (1H, m), 1.01 (3H, d, J = 6.6 Hz), 0.96–0.90 (1H, m), 0.91 (3H, d, J = 6.6 Hz), 0.88 (3H, d, J = 6.6 Hz); ^{13}C NMR (150 MHz, CDCl_3) δ 166.1, 146.0, 137.2, 136.0, 130.3, 129.3, 121.8, 73.7, 47.1, 38.4, 34.6, 28.7, 26.6, 24.9, 22.1, 20.99, 20.95, 19.1; IR (CHCl_3 , cm^{-1}) ν 2954, 1736, 1536, 1456, 1347, 1270; HRMS (DART+) (m/z): $[\text{M}+\text{NH}_4]^+$ calcd for $\text{C}_{18}\text{H}_{29}\text{N}_2\text{O}_4$: 337.2127, found: 337.2137.

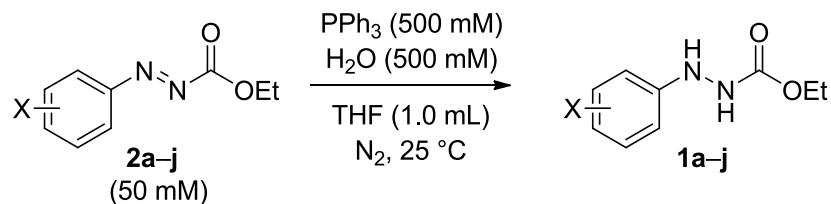
[The authentic sample of (*1R,2S,5R*)-menthyl 2-methyl-6-nitrobenzoate (*epi-47*) was preparation by acylation of (–)-menthol (**33**) and 2-methyl-6-nitrobenzoic anhydride. ^1H NMR (600 MHz, CDCl_3) δ 8.00 (1H, d, J = 8.4 Hz), 7.53 (1H, d, J = 7.8 Hz), 7.45 (1H, t, J = 7.8 Hz), 4.99 (1H, dt, J = 12.8, 4.2 Hz), 2.43 (3H, s), 2.41–2.37 (1H, m), 2.02–2.00 (1H, m), 1.73 (2H, app. d, J = 12.8 Hz), 1.59–1.56 (1H, m), 1.44 (1H, app. t, J = 6.6 Hz), 1.15–1.05 (2H, m), 0.97 (3H, d, J = 6.0 Hz), 0.93–0.91 (1H, m), 0.89 (3H, d, J = 6.6 Hz), 0.86 (3H, d, J = 6.6 Hz). No peak on C1 (δ 4.99 (1H, td, J = 12.8, 4.2 Hz)) of (*1R,2S,5R*)-menthyl 2-methyl-6-nitrobenzoate was detected in ^1H NMR of the crude product including **47**]

2. Kinetic studies for developed Mitsunobu reagents

Procedure for kinetic studies of the reaction of ethyl 2-arylazocarboxylates with triphenylphosphine and water:

A solution of ethyl 2-arylazocarboxylate (50 mM), triphenylphosphine (500 mM) and water (500 mM) in THF (1.0 mL) was prepared in a 10 mL round-bottom flask at 25 °C under a nitrogen atmosphere, and a small amount (ca. 0.5 mL) of a sample was taken from the reaction mixture, and absorption spectra of the sample continuously were measured in a 0.1 mm quartz cell. The first measurement was defined as 0 min for convenience (ca. 2 minutes from addition of reagents). Concentrations of the azo compound at each time were estimated from the absorption intensity based on calibration curves.

Table S1. The kinetic data of substituted ethyl 2-arylazocarboxylates



Entry	X	σ	$k_{\text{obs}} (\text{min}^{-1})$	$\log(k_X/k_H)$
1	<i>p</i> -OMe (b)	-0.28	0.0005	-1.09342
2	<i>p</i> -Me (c)	-0.14	0.0021	-0.47017
3	<i>p</i> -H (d)	0	0.0062	0
4	<i>p</i> -F (e)	0.06	0.0064	0.013788
5	<i>p</i> -Cl (f)	0.22	0.0195	0.497643
6	<i>m</i> -Cl (g)	0.37	0.0375	0.809668
7	<i>p</i> -CO ₂ E _t (h)	0.44	0.0639	1.013109
8	<i>p</i> -CF ₃ (i)	0.53	0.1218	1.293256
9	<i>p</i> -CN (j)	0.71	0.3224	1.716003
10	3,4-diCl (a)	—	0.0851	1.137538

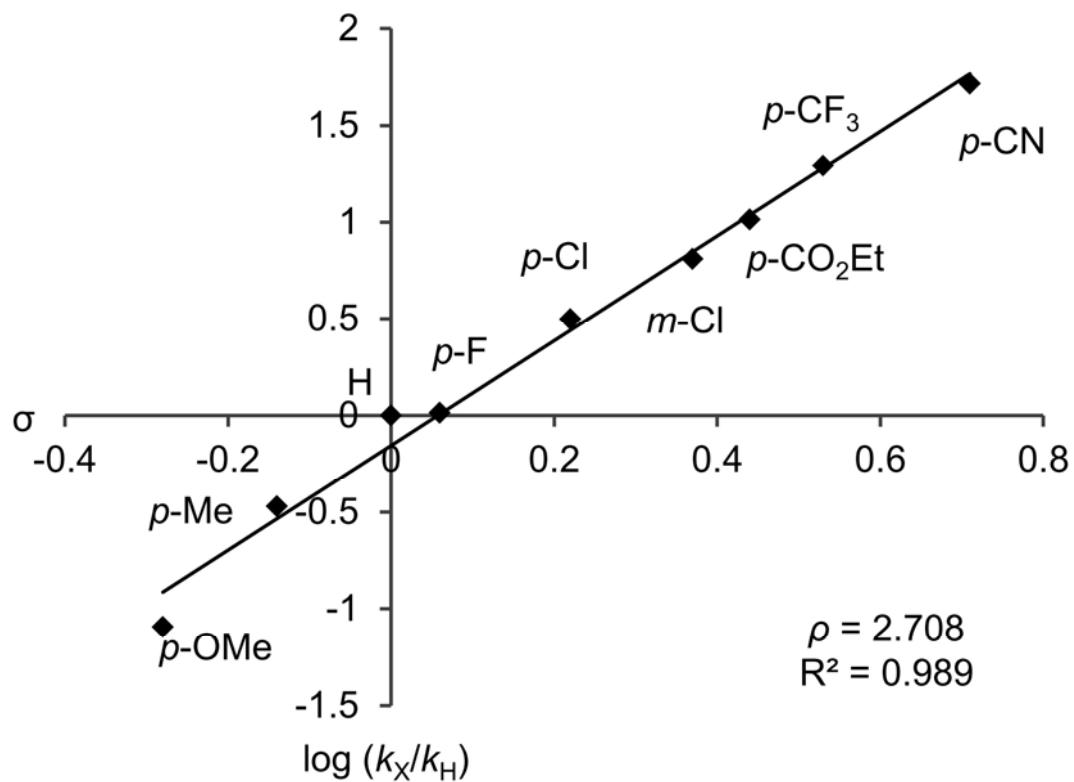
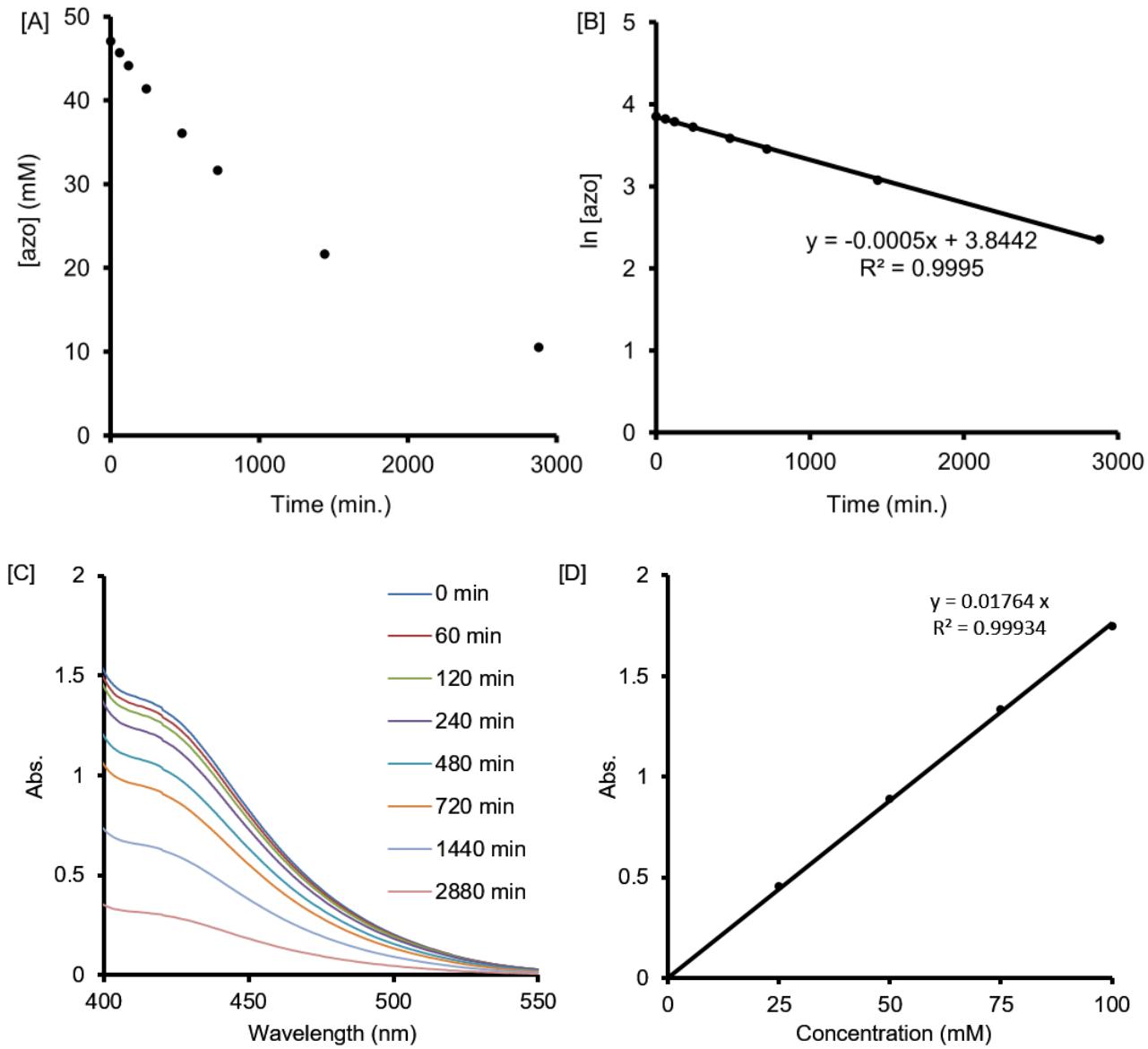
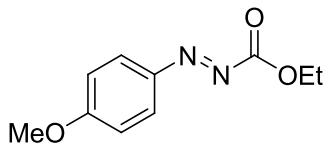
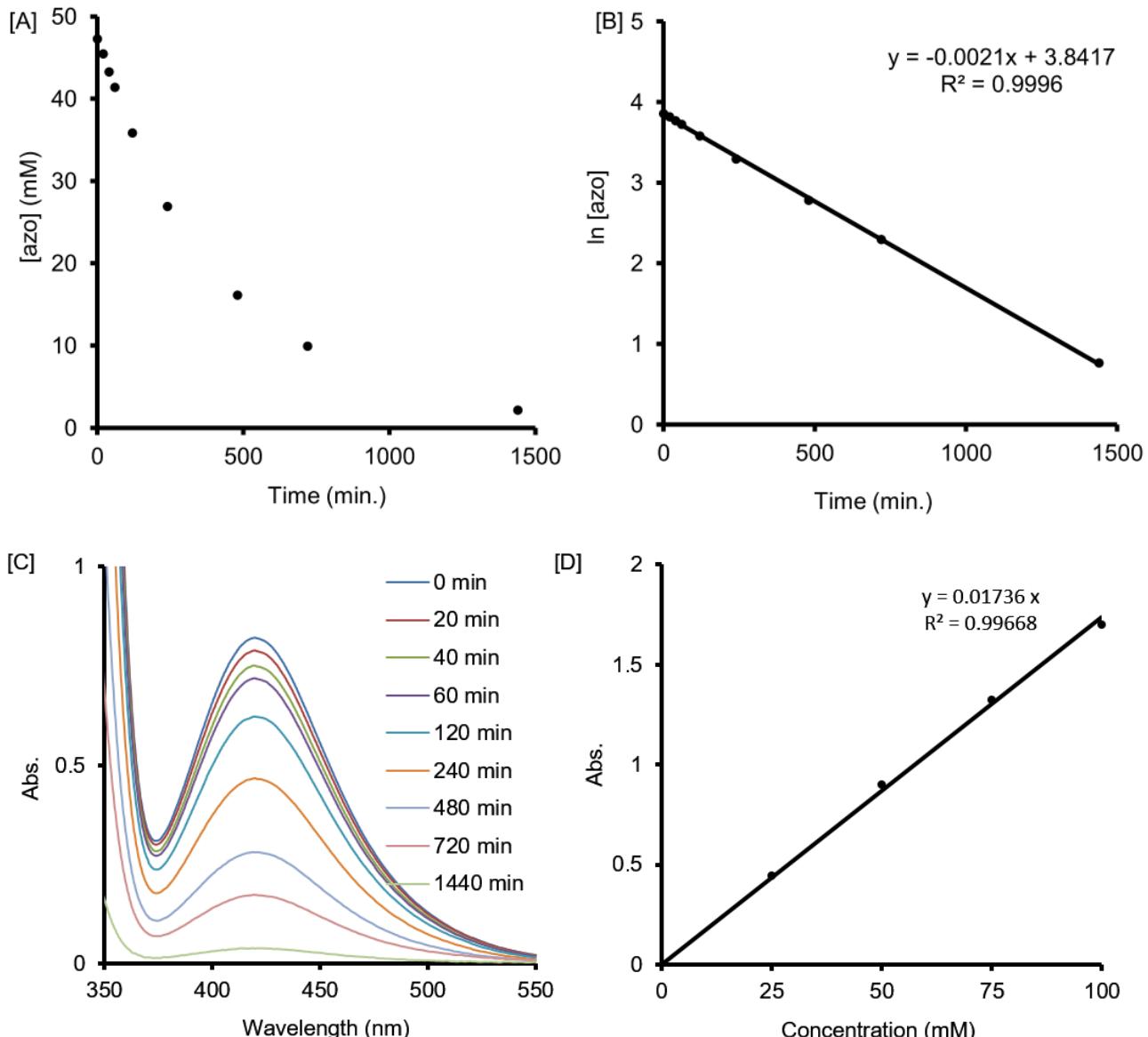


Figure S1. The Hammett plot of *p*-substituted ethyl arylazocarboxylates



Time (min)	0	60	120	240	480	720	1440	2880
Abs (450nm)	0.83027	0.80586	0.77874	0.72994	0.63628	0.55819	0.38182	0.18547
Conc (mM)	47.06746	45.68367	44.14626	41.37982	36.07029	31.64342	21.64512	10.51417

Figure S2. [A] Plot of the concentration of ethyl 2-(4-methoxyphenyl)azocarboxylate (**2b**) against time. [B] Semi-log plot of [A]. [C] Visible light absorption spectra.. [D] Calibration curve.



Time (min)	0	20	40	60	120	240	480	720	1440
Abs (420nm)	0.82056	0.78905	0.75101	0.71838	0.62215	0.46707	0.27989	0.17218	0.03721
Conc (mM)	47.26728	45.45219	43.26094	41.38134	35.83813	26.90495	16.1227	9.918203	2.143433

Figure S3. [A] Plot of the concentration of ethyl 2-(4-methylphenyl)azocarboxylate (**2c**) against time. [B] Semi-log plot of [A]. [C] Visible light absorption spectra. [D] Calibration curve.

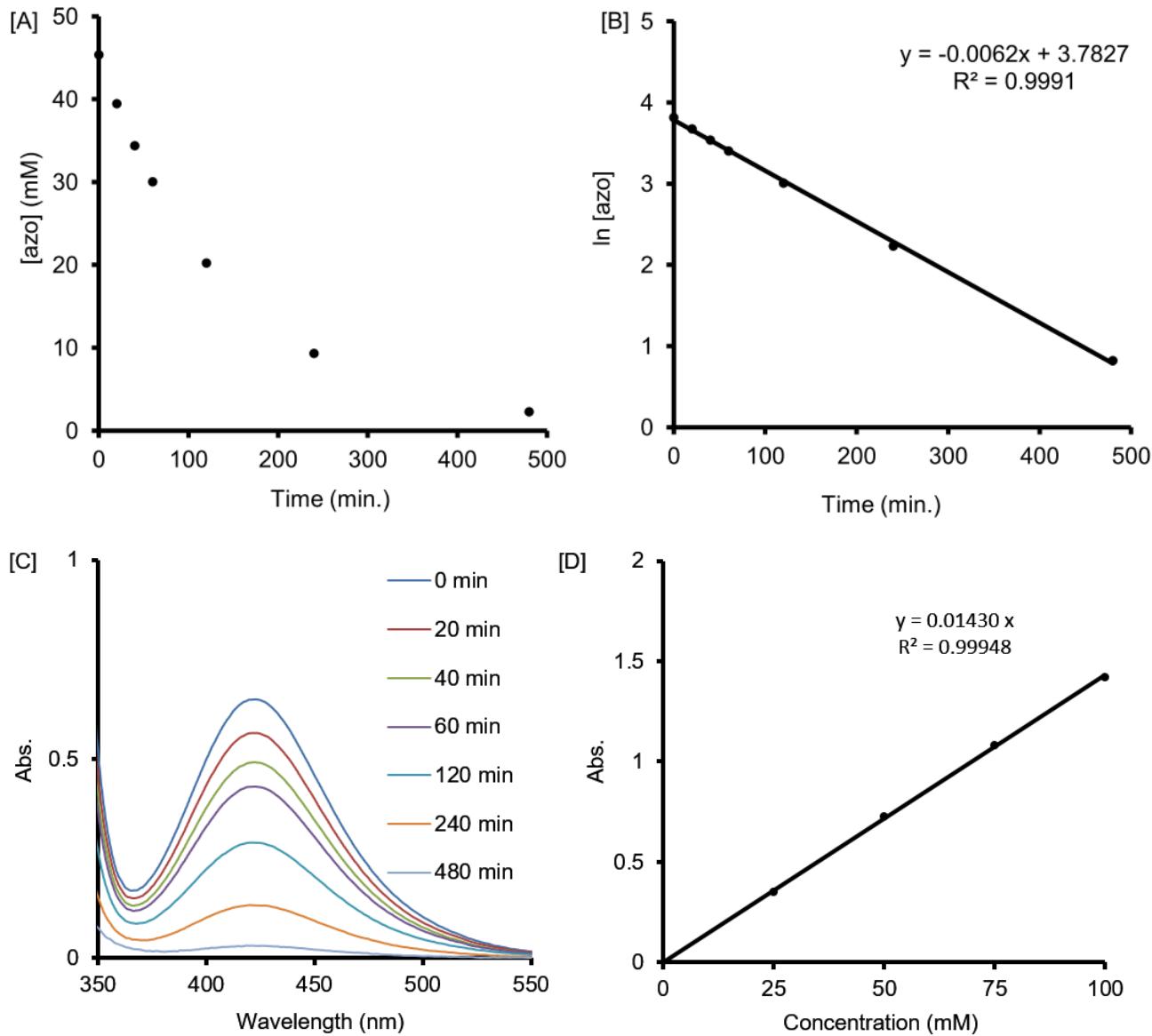
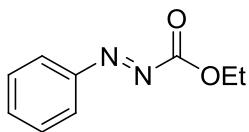
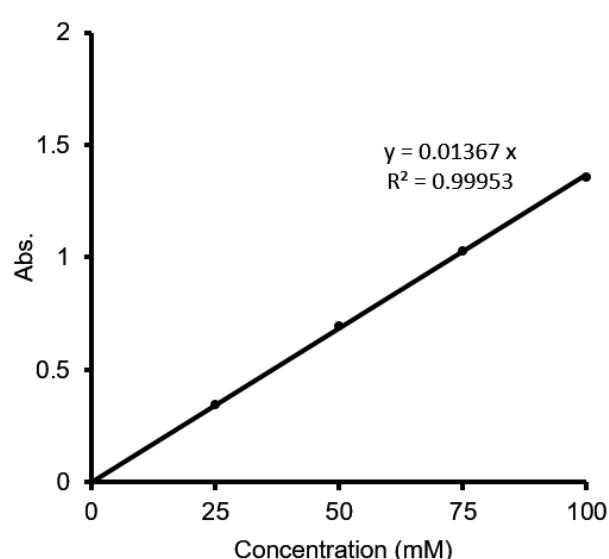
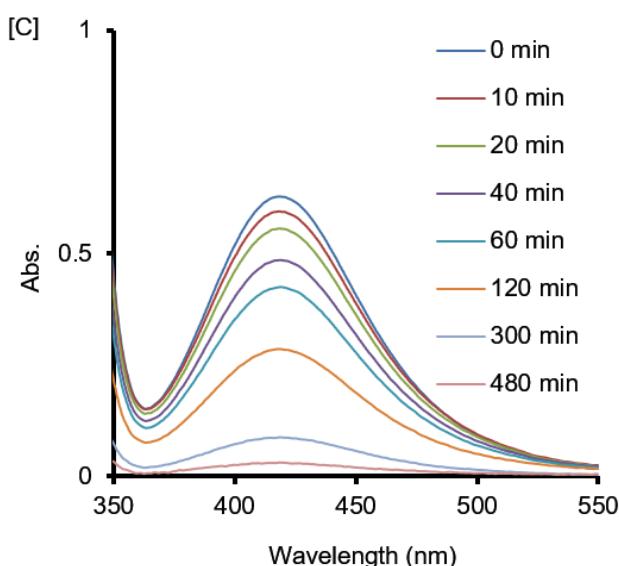
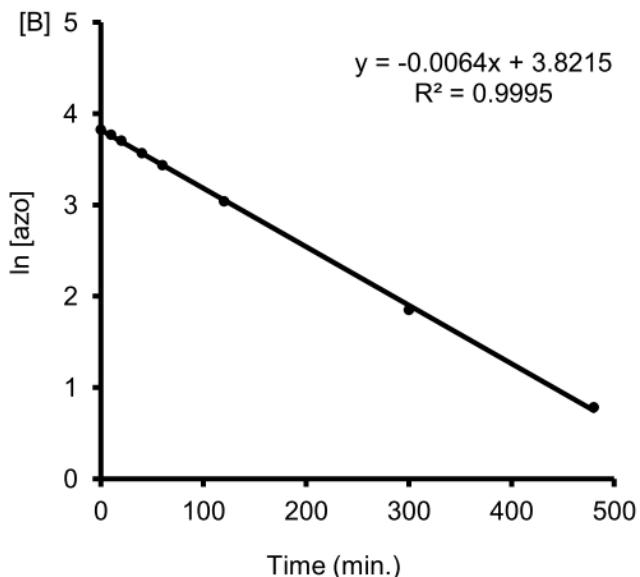
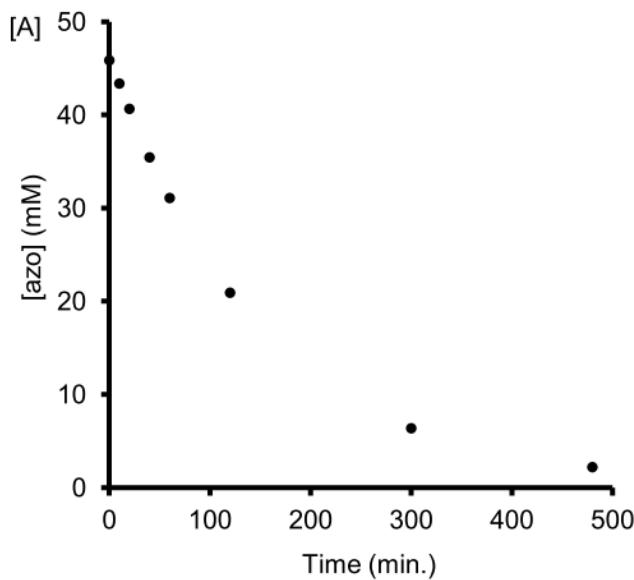
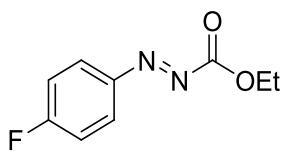


Figure S4. [A] Plot of the concentration of ethyl 2-phenylazocarboxylate (**2d**) against time. [B] Semi-log plot of [A]. [C] Visible light absorption spectra. [D] Calibration curve.



Time (min)	0	10	20	40	60	120	300	480
Abs (419nm)	0.62695	0.59284	0.55568	0.48446	0.42485	0.28585	0.08697	0.02991
Conc (mM)	45.8632	43.36796	40.6496	35.43965	31.07901	20.91075	6.362107	2.188003

Figure S5. [A] Plot of the concentration of ethyl 2-(4-fluorophenyl)azocarboxylate (**2e**) against time. [B] Semi-log plot of [A]. [C] Visible light absorption spectra. [D] Calibration curve.

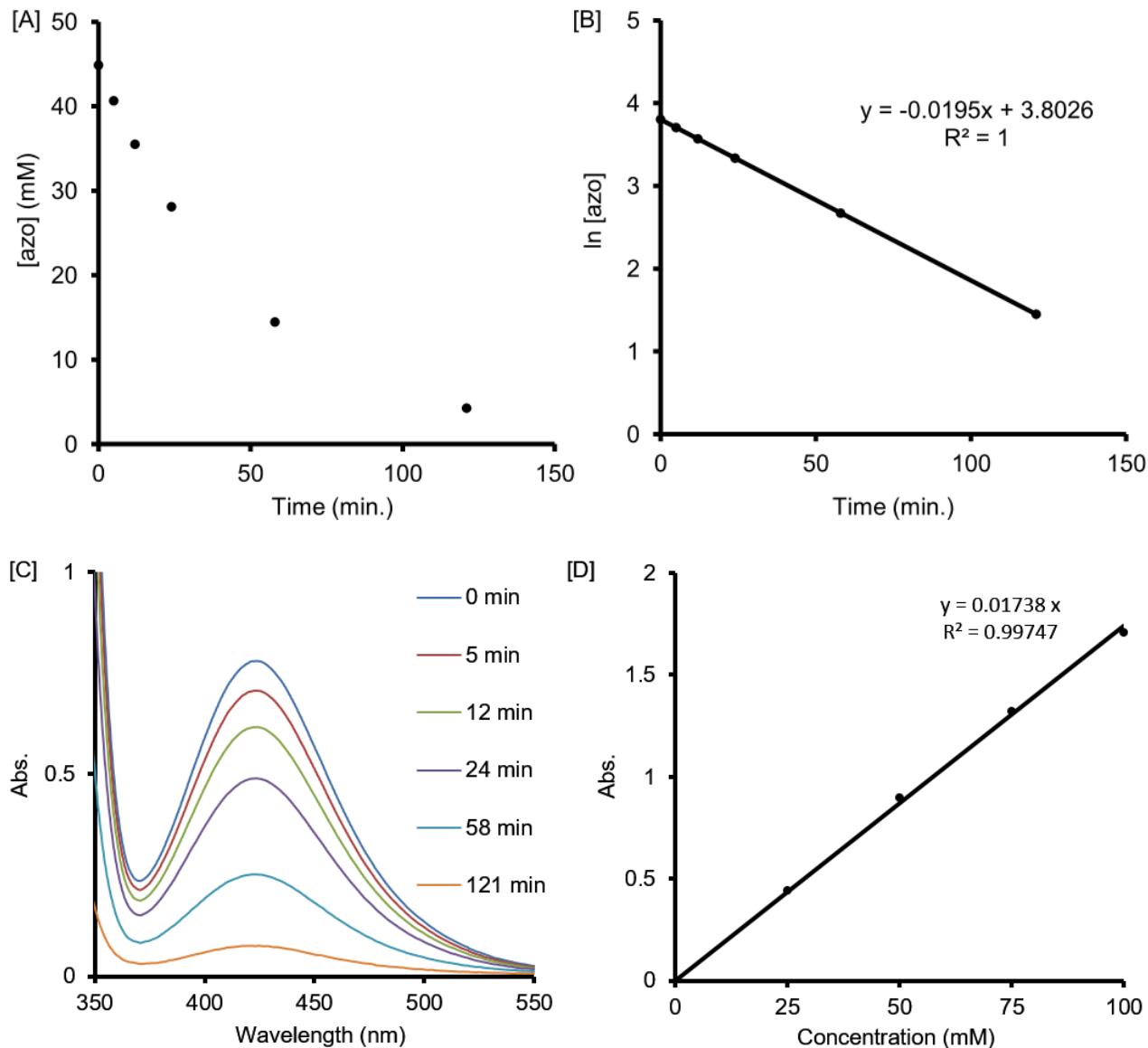
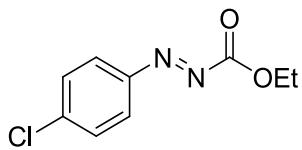
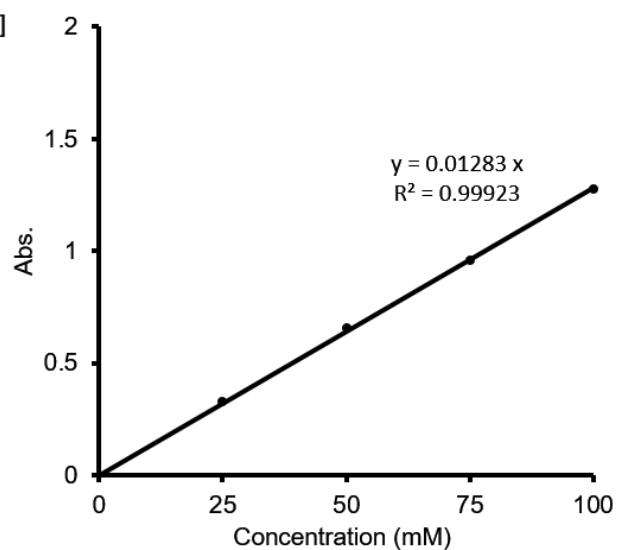
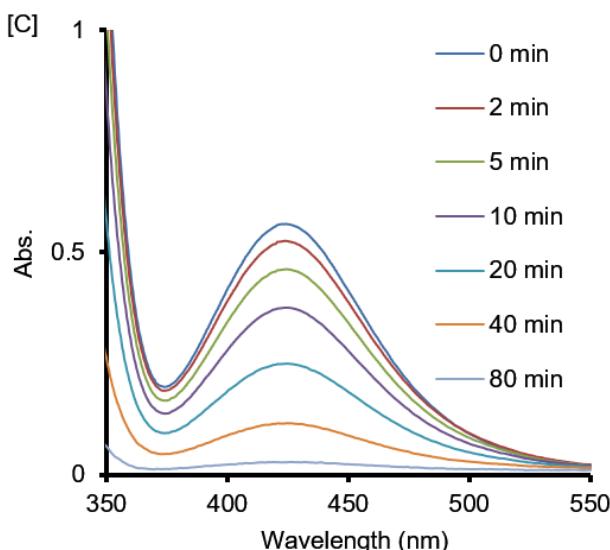
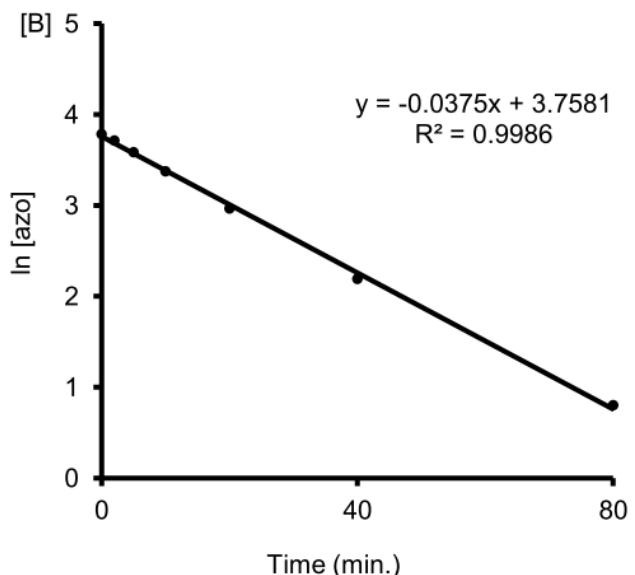
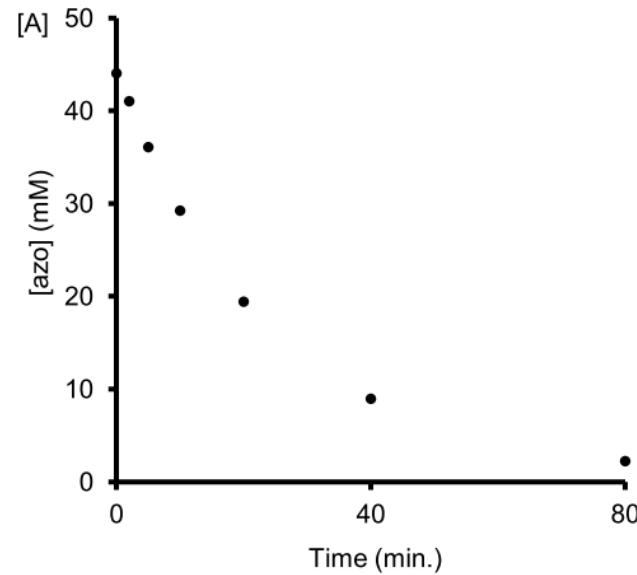
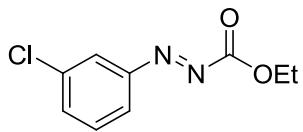
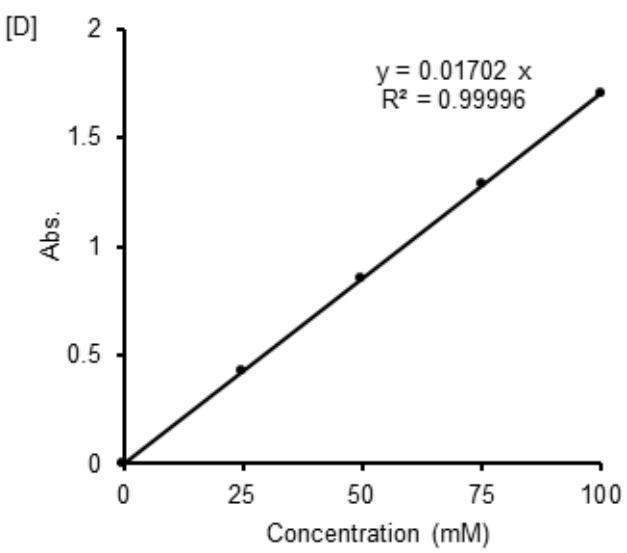
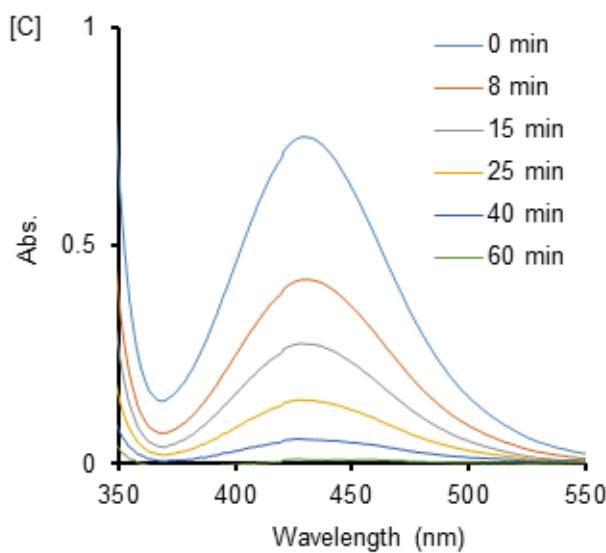
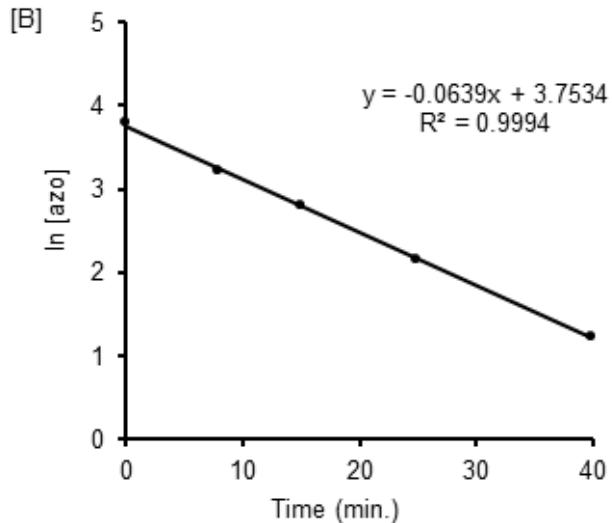
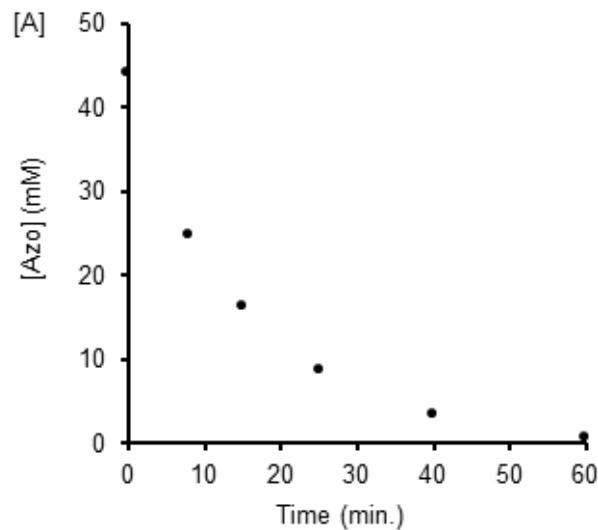
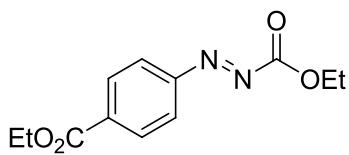


Figure S6. [A] Plot of the concentration of ethyl 2-(4-chlorophenyl)azocarboxylate (**2f**) against time. [B] Semi-log plot of [A]. [C] Visible light absorption spectra. [D] Calibration curve.



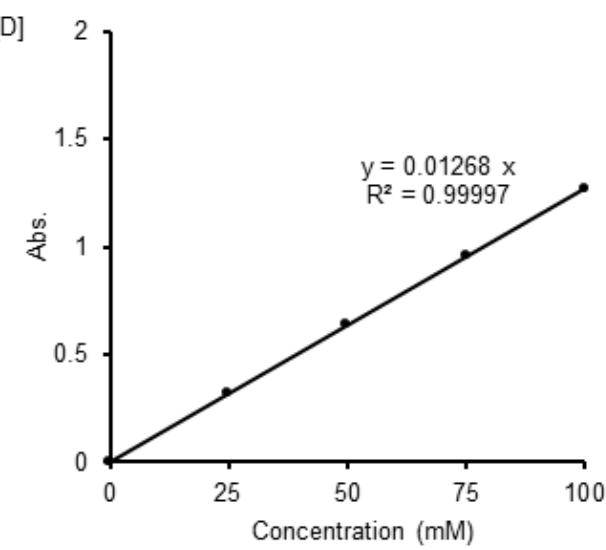
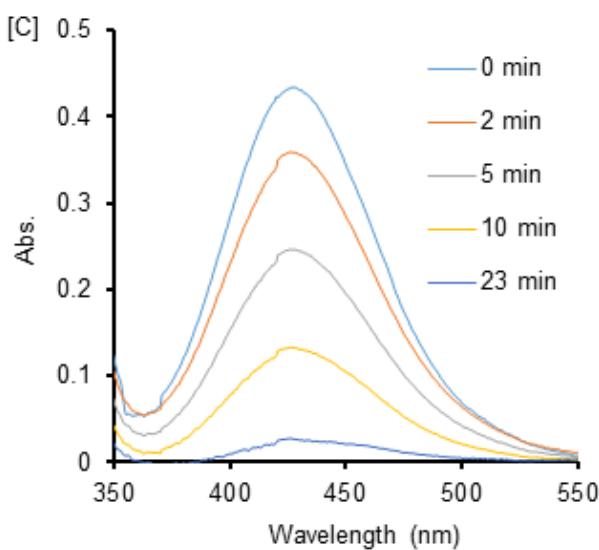
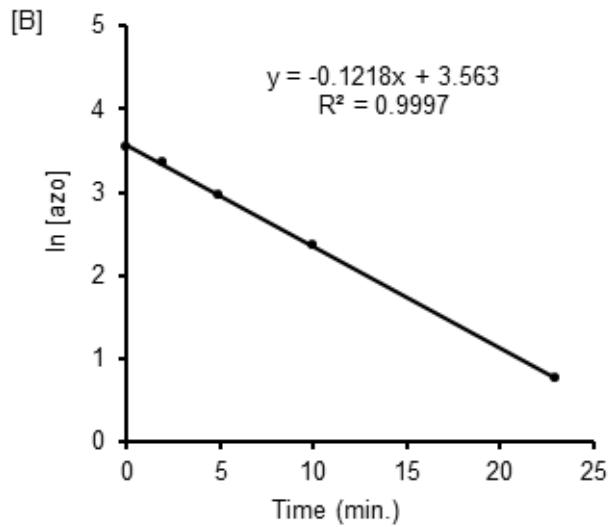
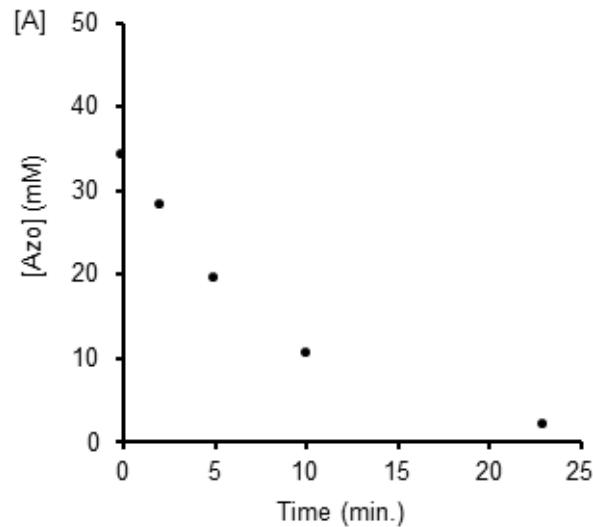
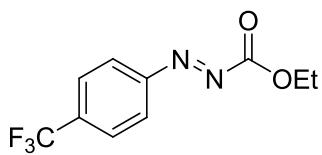
Time (min)	0	2	5	10	20	40	80
Abs (424nm)	0.56505	0.52644	0.46294	0.37509	0.24915	0.11483	0.0286
Conc (mM)	44.04131	41.03196	36.08262	29.23539	19.41933	8.950117	2.22915

Figure S7. [A] Plot of the concentration of ethyl 2-(3-chlorophenyl)azocarboxylate (**2g**) against time. [B] Semi-log plot of [A]. [C] Visible light absorption spectra. [D] Calibration curve.



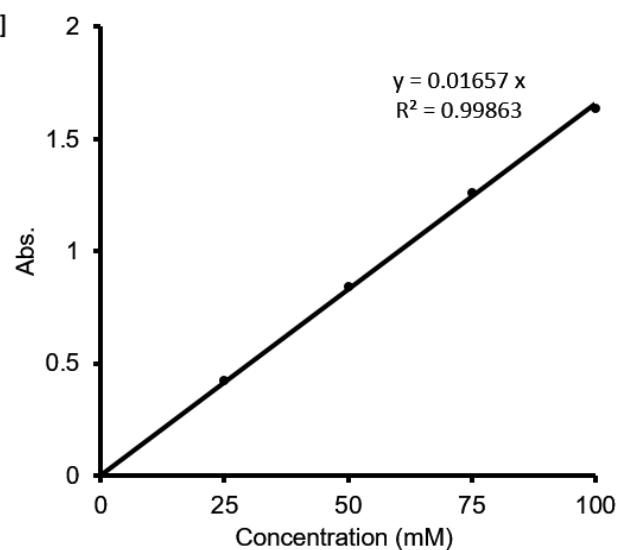
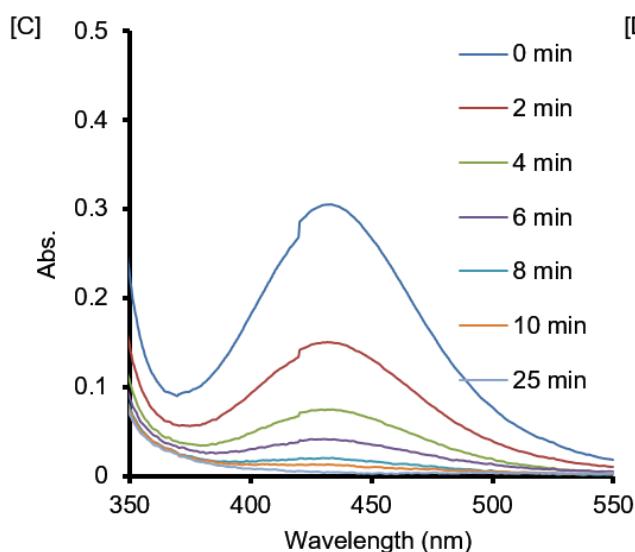
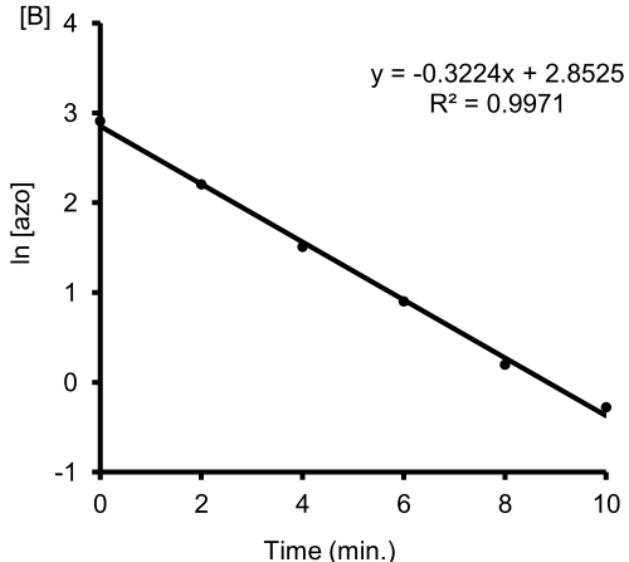
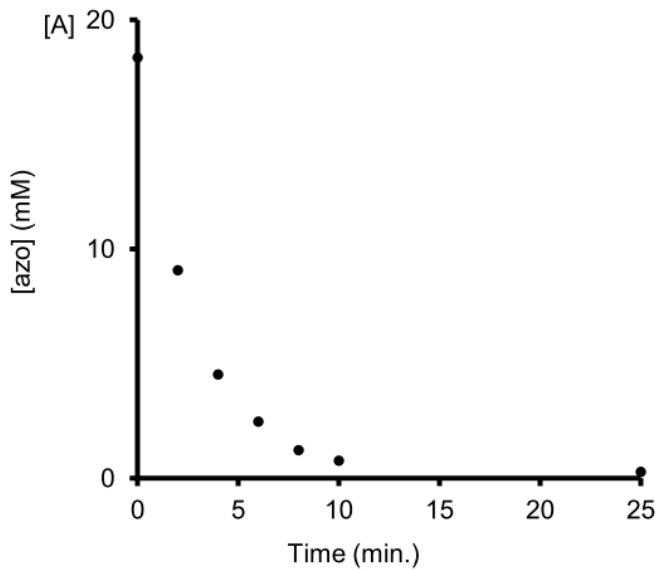
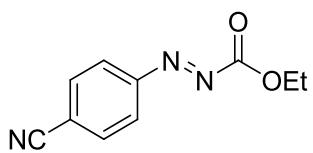
Time (min)	0	8	15	25	40	60
abs (429 nm)	0.74995	0.42258	0.27573	0.1463	0.05711	0.01066
Conc (mM)	44.06287	24.82844	16.20035	8.59577	3.355464	0.626322

Figure S8. [A] Plot of the concentration of ethyl 2-[4-(ethoxycarbonyl)phenyl]azocarboxylate (**2h**) against time. [B] Semi-log plot of [A]. [C] Visible light absorption spectra. [D] Calibration curve.



Time (min)	0	2	5	10	23
abs (427 nm)	0.43355	0.35835	0.24598	0.13274	0.02699
Conc (mM)	34.19164	28.26104	19.39905	10.46845	2.128549

Figure S9. [A] Plot of the concentration of ethyl 2-[4-(trifluoromethyl)phenyl]azocarboxylate (**2i**) against time. [B] Semi-log plot of [A]. [C] Visible light absorption spectra. [D] Calibration curve.



Time (min)	0	2	4	6	8	10	25
Abs (434nm)	0.30405	0.15027	0.07488	0.04079	0.02017	0.01255	0.00448
Conc (mM)	18.34943	9.068799	4.51901	2.461678	1.21726	0.757393	0.270368

Figure S10. [A] Plot of the concentration of ethyl 2-(4-cyanophenyl)azocarboxylate (**2j**) against time. [B] Semi-log plot of [A]. [C] Visible light absorption spectra. [D] Calibration curve.

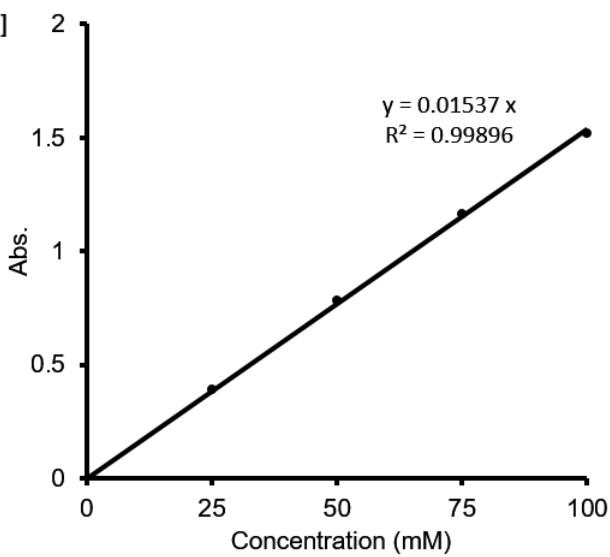
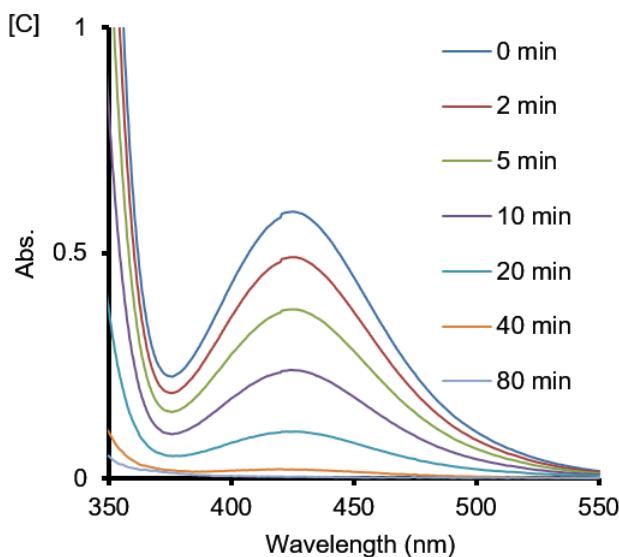
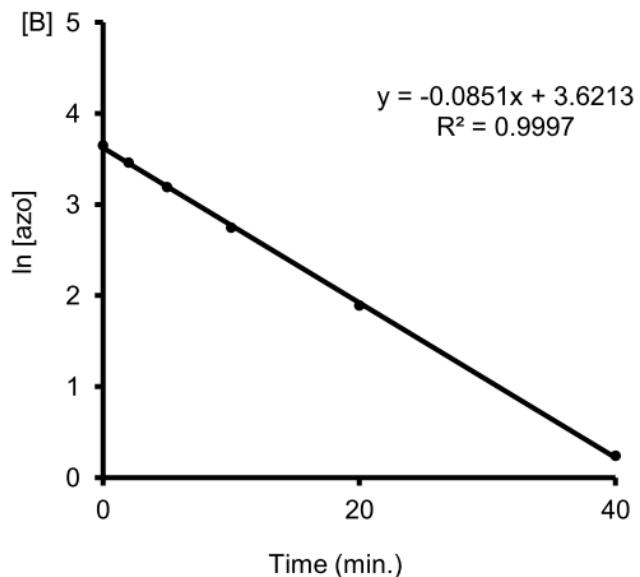
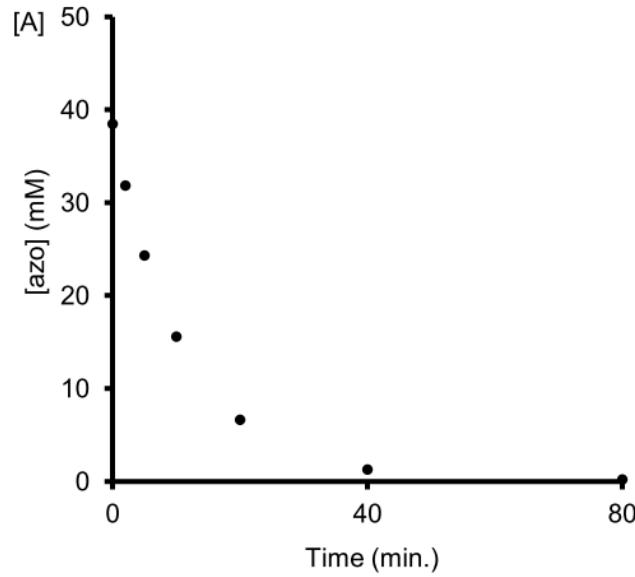


Figure S11. [A] Plot of the concentration of ethyl 2-(3,4-dichlorophenyl)azocarboxylate (**2a**) against time. [B] Semi-log plot of [A]. [C] Visible light absorption spectra. [D] Calibration curve.

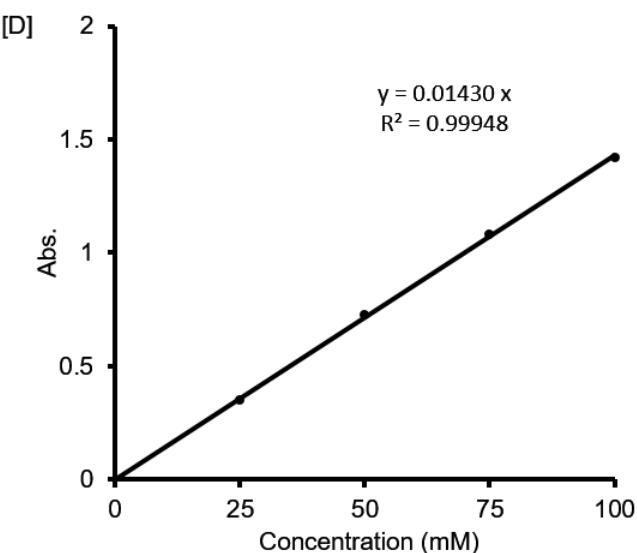
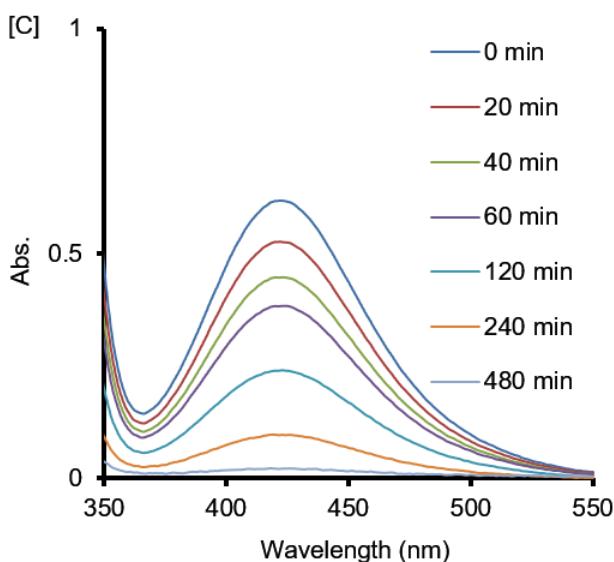
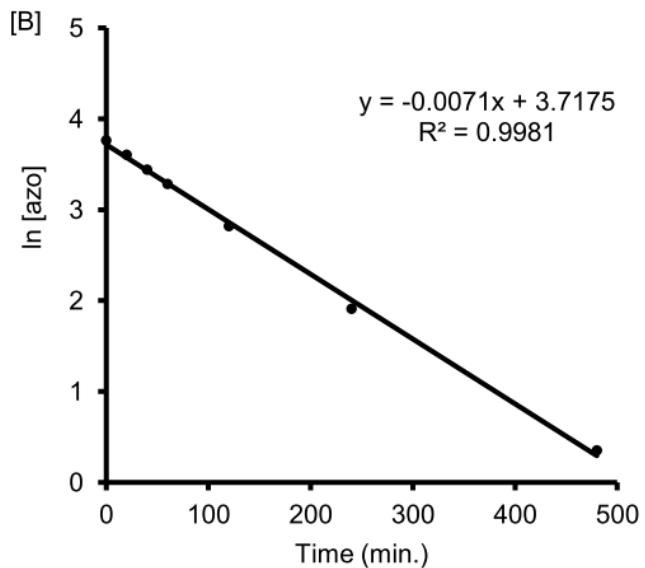
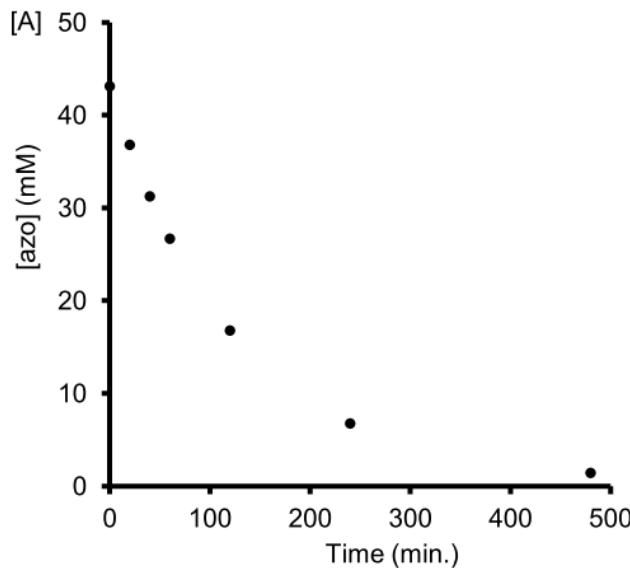
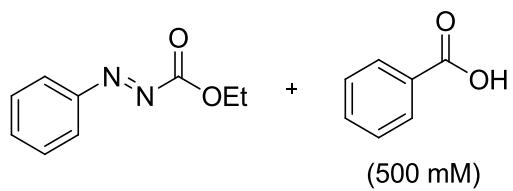
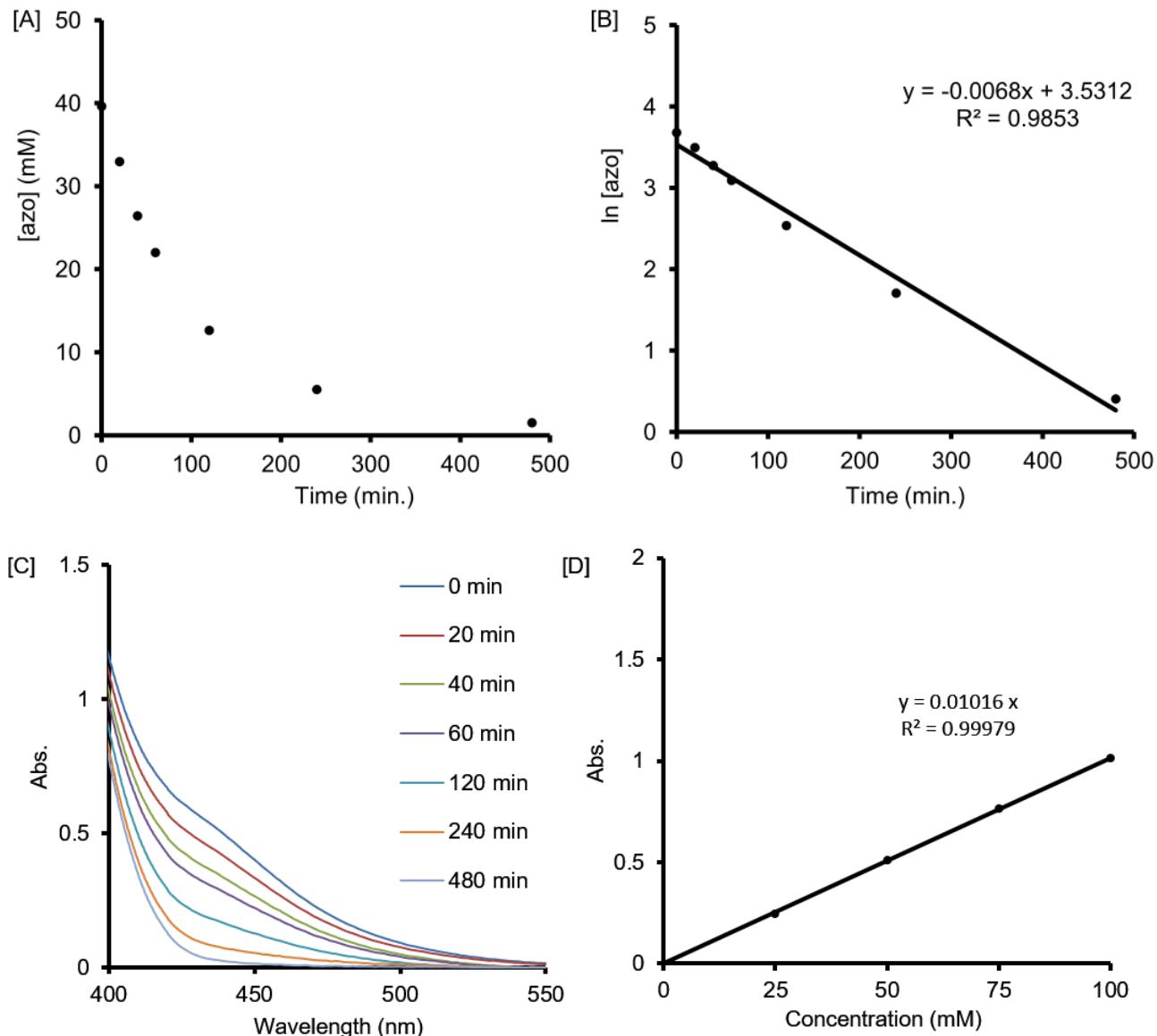
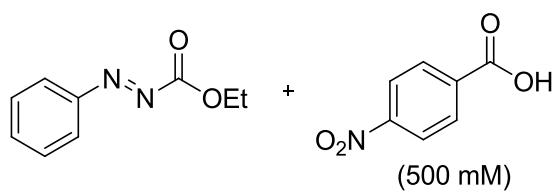


Figure S12. [A] Plot of the concentration of ethyl 2-phenylazocarboxylate (**2d**) against time in the presence of benzoic acid. [B] Semi-log plot of [A]. [C] Visible light absorption spectra. [D] Calibration curve.



Time (min)	0	20	40	60	120	240	480
Abs (450nm)	0.4026	0.33481	0.26844	0.22353	0.12827	0.05588	0.01521
Conc (mM)	39.62598	32.95374	26.42126	22.00098	12.625	5.5	1.497047

Figure S13. [A] Plot of the concentration of ethyl 2-phenylazocarboxylate (**2d**) against time in the presence of 4-nitrobenzoic acid. [B] Semi-log plot of [A]. [C] Visible light absorption spectra. [D] Calibration curve.

*Monitoring aerobic oxidation of ethyl 2-(3,4-dichlorophenyl)hydrazinecarboxylate (**1a**):*

A solution of ethyl 2-(3,4-dichlorophenyl)hydrazinecarboxylate (**1a**) (1.0 mmol) and dimethyl sulfone (94.1 mg, 1.0 mmol, internal standard) in CDCl_3 (5.0 mL) was prepared in a 10 mL round-bottom flask. Iron phthalocyanine (28.4 mg, 50 μmol) was added to the stirred solution at 22 °C under air atmosphere (defined as 0 min). A small amount (ca. 0.1 mL) of a sample was taken from the reaction mixture every 10 min while mixture was stirred, the sample was rapidly filtered through Celite®. An obtained clear solution was diluted with CDCl_3 and the solution was analyzed by ^1H NMR. The amount of the azo product was estimated from the integration value (full relaxation was confirmed).

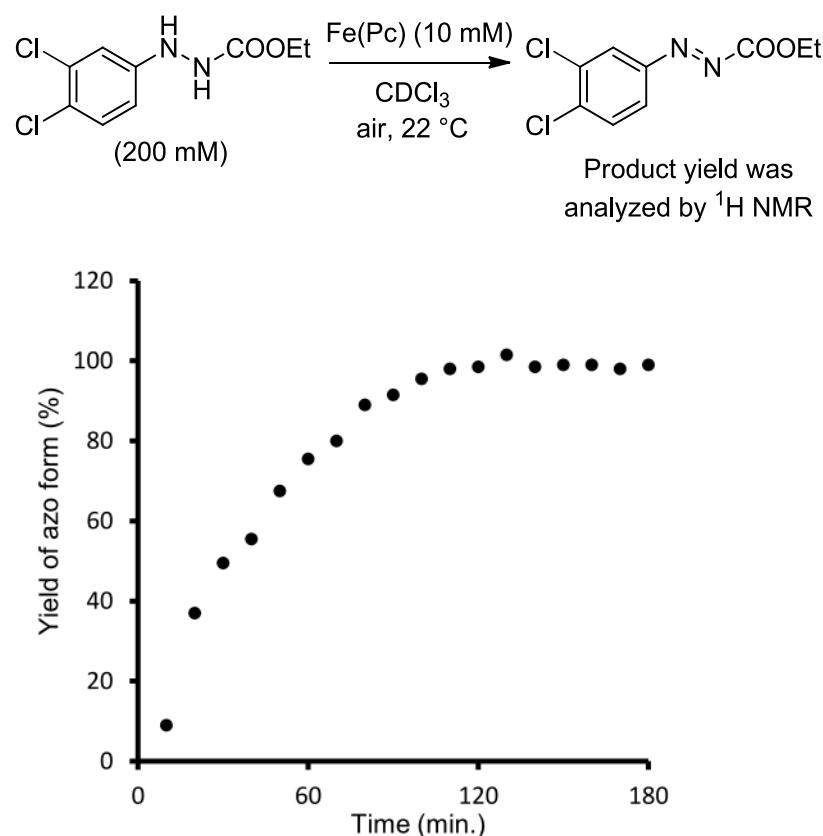
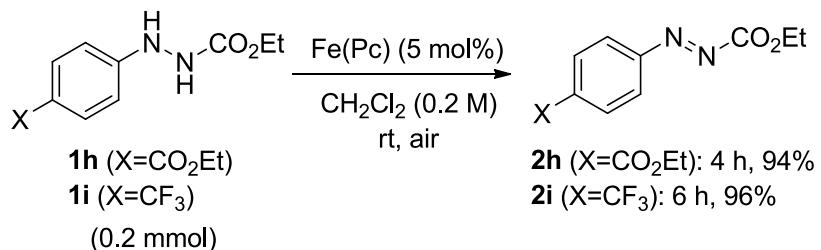


Figure S14. The kinetic plot of the reaction of aerobic oxidation of ethyl 2-(3,4-dichlorophenyl)hydrazinecarboxylate (**1a**) (200 mM) with $\text{Fe}(\text{Pc})$ (10 mM) in CDCl_3 at 22 °C.

*Aerobic oxidation of 2-[4-(ethoxycarbonyl)phenyl]hydrazinecarboxylate (**1h**) and 2-[4-(trifluoromethyl)phenyl]hydrazinecarboxylate (**1i**) on 0.2 mmol scale:*



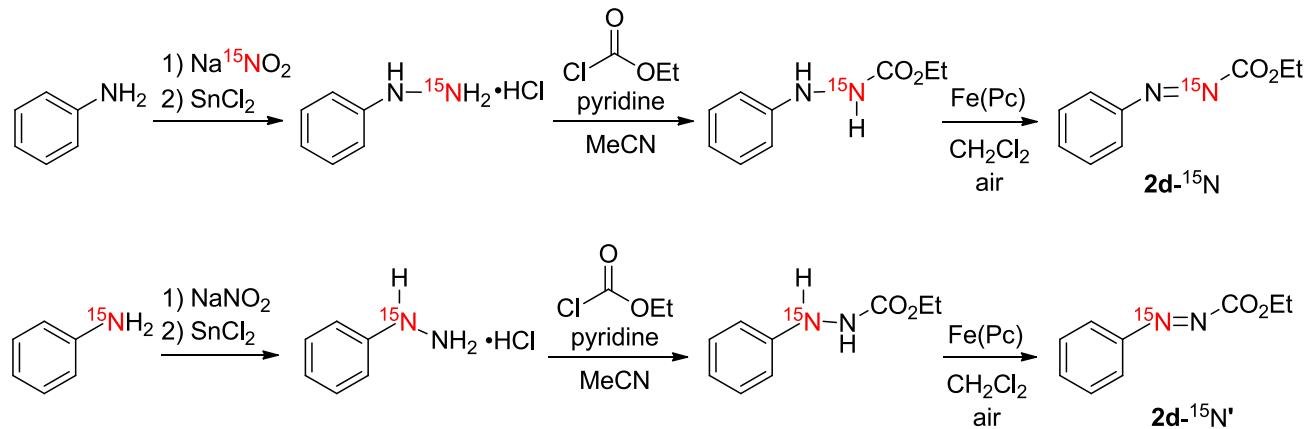
In the previous study,² iron-catalyzed aerobic oxidation reactions of 2-[4-(ethoxycarbonyl)phenyl]hydrazinecarboxylate (**1h**) and 2-[4-(trifluoromethyl)phenyl]hydrazinecarboxylate (**1i**) were not tested. Therefore, preparative reactions of **1h** and **1i** with iron phthalocyanine were performed under some conditions (0.2 mmol scale in CH₂Cl₂).

According to the reported procedure,² **1h** (50.5 mg, 0.20 mmol) and **1i** (46.9 mg, 0.20 mmol) was treated with iron phthalocyanine (5.7 mg, 0.01 mmol) in CH₂Cl₂ (1 mL) under air, respectively. The reaction time was carefully monitored by TLC analysis (**1h**: 4 h; **1i**: 6 h). After the solvent was removed under reduced pressure, the residue was purified by flash chromatography (silica gel, hexane/EtOAc = 5:1) to give 2-[4-(ethoxycarbonyl)phenyl]azocarboxylate (**2h**, 47.1 mg, 94% yield, red oil) and 2-[4-(trifluoromethyl)phenyl]azocarboxylate (**2i**, 47.3 mg, 96% yield, red oil), respectively.

Thus, these results imply that a trend in reactivity of **1h** and **1i** is roughly similar to that of ethyl 2-phenylhydrazinecarboxylate (**1d**) or ethyl 2-(4-cyanophenyl)hydrazinecarboxylate (**1j**).²

3. NMR experiments of intermediates in the Mitsunobu reactions with ethyl 2-arylazocarboxylates

*Procedure for preparation of ethyl 2-phenylazocarboxylate-1-¹⁵N (**2d-¹⁵N**) and ethyl 2-phenylazocarboxylate-2-¹⁵N (**2d-¹⁵N'**):*

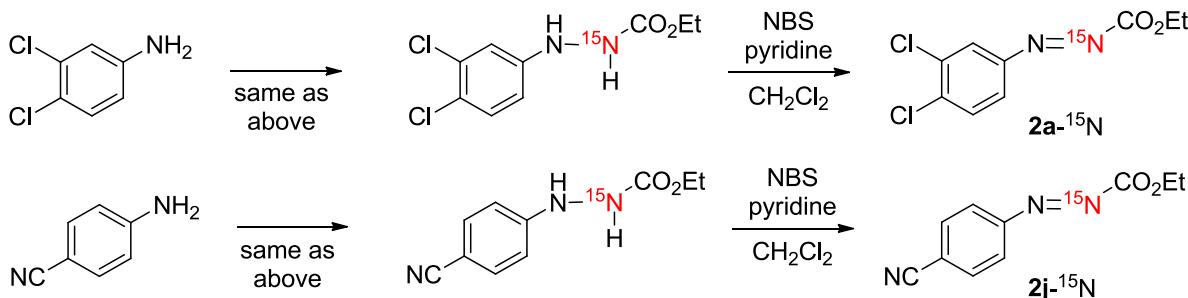


A suspension of aniline (186 mg, 2.0 mmol) or aniline-¹⁵N (189 mg, 2.0 mmol, 98.8% ¹⁵N incorporation) in concentrated HCl (2.0 mL) was cooled to 0 °C. A solution of Na¹⁵NO₂ (154 mg, 2.2 mmol, 99.2% ¹⁵N incorporation) or NaNO₂ (152 mg, 2.2 mmol) in water (2.2 mL) was added dropwise to the suspension. After the mixture was stirred for 30 min at 0 °C, a solution of SnCl₂ (1.18 g, 6.2 mmol) in concentrated HCl aq. (6.2 mL) was slowly added to the mixture. The mixture was stirred for 1 h at 0 °C, and the formed precipitate was collected by filtration. The precipitate was washed with diluted HCl aq. and dried under vacuum. To a solution of the obtained crude products and pyridine (791 mg, 10 mmol) in CH₃CN (10 mL) was added dropwise ethylchloroformate (260 mg, 2.4 mmol) at 0 °C. The mixture was stirred for 10 min at 0 °C and for 4 h at room temperature. The reaction mixture was poured into water and extracted with ethyl acetate. The combined organic layers were washed with brine, and dried with Na₂SO₄. After the solvent was evaporated, the crude product was washed by hexane, and dried under vacuum. A mixture of the crude products and iron phthalocyanine (57 mg, 0.1 mmol) in CH₂Cl₂ (10 mL) was stirred for 15 h at room temperature under air. After the solvent was removed under reduced pressure, the residue was purified by flash chromatography (silica gel, hexane/EtOAc, 5:1) to give ethyl 2-phenylazocarboxylate-1-¹⁵N (**2d-¹⁵N**) (98.6 mg, 0.55 mmol, 28% yield from aniline) or ethyl 2-phenylazocarboxylate-2-¹⁵N (**2d-¹⁵N'**) (79 mg, 0.44 mmol, 23% yield from aniline-¹⁵N) as a red oil.

2d-¹⁵N: ¹H NMR (500 MHz, CDCl₃): δ 7.96–7.91 (m, 2H, H-2 and H-6), 7.61–7.56 (m, 1H, H-4), 7.56–7.50 (m, 2H, H-3 and H-5), 4.52 (q, *J* = 7.1 Hz, 2H, CH₂), 1.47 (t, *J* = 7.1 Hz, 3H, CH₃); ¹³C NMR (126 MHz, CDCl₃): δ 162.2 (CO), 151.6 (d, *J* = 4.9 Hz, C-1), 133.8 (C-4), 129.3 (C-3 and C-5), 123.7 (d, *J* = 3.8 Hz, C-2 and C-6), 64.4 (CH₂), 14.1 (CH₃); ¹⁵N NMR (51 MHz, CDCl₃) δ 149 (ArN), 107.2 (¹⁵N-CO); IR (neat, cm⁻¹) ν 3064, 2983, 1751, 1588, 1489, 1473, 1449, 1234; HRMS (ESI+)(*m/z*): [M+H]⁺ calcd for C₉H₁₁N(¹⁵N)O₂: 180.0785; found: 180.0787.

2d-¹⁵N: ¹H NMR (500 MHz, CDCl₃): δ 7.96–7.92 (m, 2H, H-2 and H-6), 7.62–7.57 (m, 1H, H-4), 7.56–7.50 (m, 2H, H-3 and H-5), 4.53 (q, J = 7.1 Hz, 2H, CH₂), 1.48 (t, J = 7.1 Hz, 3H, CH₃); ¹³C NMR (126 MHz, CDCl₃): δ 162.2 (d, J = 6.3 Hz, CO), 151.6 (d, J = 2.8 Hz, C-1), 133.8 (C-4), 129.3 (d, J = 1.8 Hz, C-3 and C-5), 123.7 (d, J = 4.3 Hz, C-2 and C-6), 64.5 (CH₂), 14.1 (CH₃); ¹⁵N NMR (51 MHz, CDCl₃): δ 149 (Ar-¹⁵N); IR (neat, cm⁻¹) ν 3063, 2983, 2939, 1750, 1587, 1490, 1471, 1459, 1233; HRMS (ESI+) (*m/z*): [M+H]⁺ calcd for C₉H₁₁N(¹⁵N)O₂: 180.0785; found: 180.0785.

Procedure for preparation of ethyl 2-(3,4-dichlorophenyl)azocarboxylate-1-¹⁵N (2a-¹⁵N) and ethyl 2-(4-cyanophenyl)azocarboxylate-1-¹⁵N (2j-¹⁵N):



According to the procedure for preparation of 2d-¹⁵N, ethyl 2-(3,4-dichlorophenyl)hydrazinecarboxylate-1-¹⁵N and ethyl 2-(4-cyanoaniline)hydrazine-1-¹⁵N were prepared from 3,4-dichloroaniline and 4-cyanoaniline. To a solution of an appropriate hydrazide (0.5 mmol) and pyridine (101 μ L, 99 mg, 1.25 mmol) in dichloromethane (4 mL), *N*-bromosuccinimide (98 mg, 0.55 mmol) was added portion-wise at room temperature. After stirring for 30 min at room temperature, the reaction mixture was diluted with dichloromethane (15 mL), and the organic layer was successively washed with 1 M solution of hydrochloric acid (10 mL), 1.5% solution of sodium thiosulfate (5 mL), a saturated solution of sodium bicarbonate (10 mL) and brine (10 mL), and dried over anhydrous sodium sulfate. After the solvent was evaporated, the crude product was purified by flash chromatography (silica gel, hexanes/EtOAc, 6:1) to give the corresponding azo compound.

2a-¹⁵N: 92% yield (114 mg, 0.46 mmol). Red solid, mp 49–50 °C.

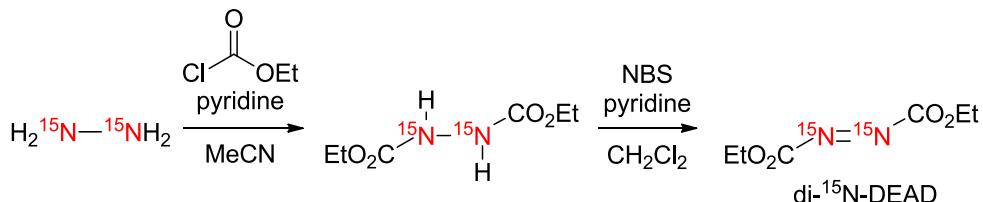
¹H NMR (500 MHz, CDCl₃): δ 8.02 (d, J = 2.2 Hz, 1H, H-2), 7.81 (dd, J ₁ = 8.6 Hz, J ₂ = 2.2 Hz, 1H, H-6), 7.63 (d, J = 8.6 Hz, 1H, H-5), 4.53 (q, J = 7.1 Hz, 2H, CH₂), 1.47 (t, J = 7.1 Hz, 3H, CH₃); ¹³C NMR (126 MHz, CDCl₃): δ 161.6 (CO), 150.3 (d, J = 5.3 Hz, C-1), 138.1 (C-4), 134.0 (C-3), 131.2 (C-5), 124.6 (d, J = 4.1 Hz, C-2), 123.6 (d, J = 3.9 Hz, C-6), 64.8 (CH₂), 14.1 (CH₃); ¹⁵N NMR (51 MHz, CDCl₃): δ 114 (¹⁵N-CO); IR (neat, cm⁻¹) ν 3351, 3094, 2983, 1755, 1586, 1520, 1471, 1383, 1234; HRMS (ESI+) (*m/z*): [M+H]⁺ calcd for C₉H₉Cl₂N(¹⁵N)O₂: 248.0006; found: 248.0006.

2j-¹⁵N: 95% yield (97.0 mg, 0.475 mmol). Red solid, mp 45–46 °C.

¹H NMR (500 MHz, CDCl₃): δ 8.00 (d, J = 8.5 Hz, 2H, H-2 and H-6), 7.85 (d, J = 8.5 Hz, 2H, H-3

and H-5), 4.55 (q, $J = 7.2$ Hz, 2H, CH₂), 1.48 (t, $J = 7.2$ Hz, 3H, CH₃); ¹³C NMR (126 MHz, CDCl₃): δ 161.6 (CO), 153.1 (d, $J = 5.3$ Hz, C-1), 133.4 (C-3 and C-5), 124.0 (d, $J = 4.1$ Hz, C-2 and C-6), 117.8 (CN), 116.7 (C-4), 65.0 (CH₂), 14.1 (CH₃); ¹⁵N NMR (51 MHz, CDCl₃): δ 145 (Ar-N), 125 (¹⁵N-CO); IR (neat, cm⁻¹) ν 3096, 2993, 2230, 1751, 1599, 1496, 1468, 1237; HRMS (ESI+) (*m/z*): [M+H]⁺ calcd for C₁₀H₁₀N₂(¹⁵N)O₂: 205.0738; found: 205.0738.

Procedure for preparation of diethyl azodicarboxylate-1,2-¹⁵N₂ (di-¹⁵N DEAD):



To a solution of hydrazine-¹⁵N₂ sulfate salt (130 mg, 1.0 mmol) and pyridine (316 mg, 4.0 mmol) in acetonitrile (4 mL), ethyl chloroformate (238 mg, 2.2 mmol) was added dropwise at 0 °C. The reaction mixture was stirred for 15 minutes at 0 °C and then for 1 h at room temperature. The reaction mixture was diluted with dichloromethane (25 mL), and the organic layer was successively washed with 1 M solution of hydrochloric acid (1M, 10 mL), a saturated solution of sodium bicarbonate (10 mL) and brine (10 mL), and dried over anhydrous sodium sulfate. Solvent evaporation afforded pure diethyl hydrazinecarboxylate-1,2-¹⁵N₂ (128 mg, 0.72 mmol, 72%) as white crystals, which was used in next step without further purification. According to the procedure for preparation of **2a**-¹⁵N and **2j**-¹⁵N, diethyl hydrazinecarboxylate-1,2-¹⁵N₂ was oxidized by treatment of NBS and pyridine in dichloromethane to give diethyl azodicarboxylate-1,2-¹⁵N₂ (78 mg, 0.445 mmol, 89%) as a yellow oil.

Diethyl hydrazine-1,2-dicarboxylate-1,2-¹⁵N₂:

mp 131–133 °C.

¹H NMR (500 MHz, CDCl₃): δ 6.44 (d, $J = 100.9$ Hz, ¹⁵NH), 4.21 (q, $J = 7.1$ Hz, 2H, CH₂), 1.28 (t, $J = 7.1$ Hz, 3H, CH₃); ¹³C NMR (126 MHz, CDCl₃): δ 156.8 (br s, CO), 62.3 (CH₂), 14.4 (CH₃); ¹⁵N NMR (51 MHz, CDCl₃): δ 97 (¹⁵NH-CO); IR (neat, cm⁻¹) ν 3228, 2990, 2917, 1745, 1693, 1516, 1481, 1233; HRMS (ESI+) (*m/z*): [M+H]⁺ calcd for C₆H₁₃(¹⁵N)₂O₄: 179.0811; found: 179.0810.

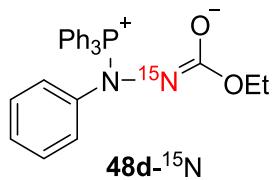
di-¹⁵N DEAD:

¹H NMR (500 MHz, CDCl₃): δ 4.51 (q, $J = 7.1$ Hz, 2H, CH₂), 1.45 (t, $J = 7.1$ Hz, 3H, CH₃); ¹³C NMR (126 MHz, CDCl₃): δ 160.3 (dd, $J = 2.3, 2.3$ Hz, CO), 65.5 (CH₂), 14.0 (CH₃); ¹⁵N NMR (51 MHz, CDCl₃): δ 150 (¹⁵N-CO); IR (neat, cm⁻¹) ν 2987, 1768, 1471, 1368; HRMS (ESI+) (*m/z*): [M+3H]⁺ calcd for C₆H₁₃(¹⁵N)₂O₄: 179.0811; found: 179.0809.

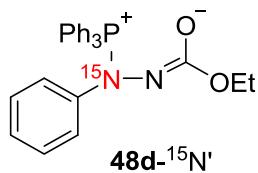
General procedure for preparation and analysis of betaine intermediates **48**:

A solution of an azo compound (0.04 mmol) in CDCl₃ (0.4 mL) was added to an NMR tube under

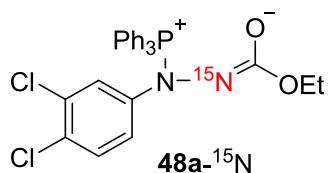
argon atmosphere, followed by the addition of a solution of triphenylphosphine (0.4 mmol) in CDCl_3 (0.4 mL). NMR spectra were recorded after 10 minutes.



^1H NMR (500 MHz, CDCl_3): δ 7.91–7.84 (m, 6H, H-2' and H-6'), 7.63–7.57 (m, 3H, H-4'), 7.51–7.40 (m, 6H, H-3' and H-5'), 7.14–7.09 (m, 2H, H-2 and H-6), 7.03–6.97 (m, 2H, H-3 and H-5), 6.91–6.86 (m, 1H, H-4), 3.82 (q, $J = 7.0$ Hz, 2H, $-\text{CH}_2\text{CH}_3$), 1.04 (t, $J = 7.0$ Hz, 3H, CH_3); ^{13}C NMR (126 MHz, CDCl_3): δ 168.1 (d, $J = 6.4$ Hz, CO), 144.1 (d, $J = 12.1$ Hz, C-1), 134.6 (d, $J = 10.3$ Hz, C-2' and C-6'), 133.3 (d, $J = 1.9$ Hz, C-4'), 128.9 (d, $J = 13.1$ Hz, C-3' and C-5'), 128.1 (C-3 and C-5), 124.8 (C-2 and C-6), 124.0 (C-4), 121.7 (d, $J = 103.0$ Hz, C-1'), 59.1 (CH_2), 15.2 (CH_3); ^{15}N NMR (51 MHz, CDCl_3): δ 182 (^{15}N -CO), 83 (Ar-N); ^{31}P NMR (202 MHz, CDCl_3): δ +33.3 (d, $J = 5.2$ Hz); HRMS (ESI+) (m/z): $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{27}\text{H}_{26}\text{N}(\text{¹⁵N})\text{O}_2\text{P}$: 442.1697; found: 442.1694.



^1H NMR (500 MHz, CDCl_3): δ 7.91–7.84 (m, 6H, H-2' and H-6'), 7.61–7.56 (m, 3H, H-4'), 7.51–7.41 (m, 6H, H-3' and H-5'), 7.15–7.10 (m, 2H, H-2 and H-6), 7.03–6.97 (m, 2H, H-3 and H-5), 6.91–6.86 (m, 1H, H-4), 3.86–3.80 (m, 2H, $-\text{CH}_2$), 1.06–1.01 (m, 3H, CH_3); ^{13}C NMR (126 MHz, CDCl_3): δ 168.1 (d, $J = 6.1$ Hz, CO), 144.0 (d, $J = 12.4$ Hz, C-1), 134.6 (d, $J = 10.3$ Hz, C-2' and C-6'), 133.3 (d, $J = 2.3$ Hz, C-4'), 128.9 (d, $J = 12.9$ Hz, C-3' and C-5'), 128.2 (d, $J = 7.0$ Hz, C-3 and C-5), 124.7 (C-2 and C-6), 124.0 (C-4), 59.1 (CH_2), 15.2 (CH_3); ^{15}N NMR (51 MHz, CDCl_3): δ 84 (Ar- ^{15}N); ^{31}P NMR (202 MHz, CDCl_3): δ +33.4 (d, $J = 6.2$ Hz); HRMS (ESI+) (m/z): $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{27}\text{H}_{26}\text{N}(\text{¹⁵N})\text{O}_2\text{P}$: 442.1697; found: 442.1691.

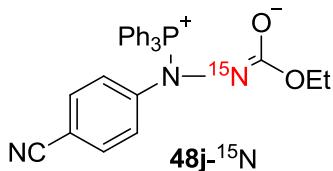


^1H NMR (500 MHz, CDCl_3): δ 7.91–7.83 (m, 6H, H-2' and H-6'), 7.67–7.62 (m, 3H, H-4'), 7.54–7.49 (m, 6H, H-3' and H-5'), 7.21–7.17 (m, 1H, H-2), 7.02 (d, $J = 8.7$ Hz, 1H, H-5), 6.86–6.82 (m, 1H, H-6), 3.82 (q, $J = 7.0$ Hz, 2H, CH_2), 1.03 (t, $J = 7.0$ Hz, 3H, CH_3); ^{13}C NMR (126 MHz, CDCl_3): δ 168.0 (dd, $J_1 = 5.5$ Hz, $J_2 = 1.8$ Hz, CO), 144.1 (d, $J = 12.8$ Hz, C-1), 134.6 (d, $J = 10.5$ Hz, C-2' and C-6'), 134.1 (d, $J = 2.5$ Hz, C-4'), 131.9 (C-3), 129.4 (C-5), 129.1 (d, $J = 12.9$ Hz, C-3')

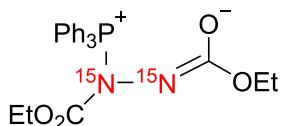
and C-5'), 127.1 (C-4), 125.6 (d, $J = 3.1$ Hz, C-2), 123.1 (d, $J = 2.5$ Hz, C-6), 122.4 (d, $J = 103.3$ Hz, C-1'), 59.3 (CH₂), 15.2 (CH₃); ¹⁵N NMR (51 MHz, CDCl₃): δ 182 (¹⁵N-CO); ³¹P NMR (202 MHz, CDCl₃): δ +34.5 (d, $J = 5.0$ Hz); HRMS (ESI+) (*m/z*): [M+H]⁺ calcd for C₂₇H₂₄Cl₂N(¹⁵N)O₂P: 510.0917; found: 510.0915.

Table S2. Chemical shifts of ³¹P NMR of betaine **48a** in different solvents

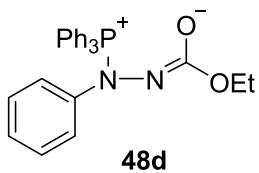
entry	solvent	³¹ P NMR (202 MHz) δ (ppm)
1	THF- <i>d</i> ₈	+21.1
2	CDCl ₃	+34.5
3	CD ₃ CN	+33.7
4	toluene- <i>d</i> ₈	Not detected



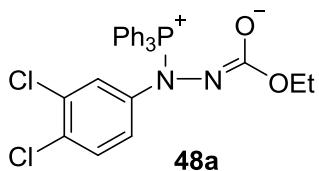
¹H NMR (500 MHz, CDCl₃): δ 7.92–7.83 (m, 6H, H-2' and H-6'), 7.71–7.65 (m, 3H, H-4'), 7.56–7.49 (m, 6H, H-3' and H-5'), 7.27 (d, $J = 8.5$ Hz, 2H, H-3 and H-5), 7.00 (d, $J = 8.5$ Hz, 2H, H-2 and H-6), 3.71 (q, $J = 7.0$ Hz, 2H, -CH₂CH₃), 0.96 (t, $J = 7.0$ Hz, 3H, CH₃); ¹³C NMR (126 MHz, CDCl₃): δ 168.4 (CO), 149.4 (d, $J = 15.7$ Hz), 134.7 (d, $J = 10.6$ Hz, C-2' and C-6'), 134.2 (d, $J = 2.6$ Hz, C-4'), 132.0 (C-3 and C-5), 129.2 (d, $J = 13.2$ Hz, C-3' and C-5'), 121.7 (d, $J = 103.1$ Hz, C-1'), 120.6 (d, $J = 3.2$ Hz, C-2 and C-6), 119.2 (CN), 104.7 (C-4), 59.3 (CH₂), 15.1 (CH₃); ¹⁵N NMR (51 MHz, CDCl₃): δ 179 (¹⁵N-CO), 90 (Ar-N); ³¹P NMR (202 MHz, CDCl₃): δ +35.4 (s); HRMS (ESI+) (*m/z*): [M+H]⁺ calcd for C₂₈H₂₅N₂(¹⁵N)O₂P: 467.1649; found: 467.1642.



¹H NMR (500 MHz, CDCl₃): δ 7.97–7.90 (m, 6H, H-2' and H-6'), 7.71–7.65 (m, 3H, H-4'), 7.58–7.52 (m, 6H, H-3' and H-5'), 4.16 (q, $J = 7.0$ Hz, 2H, Ph₃P-N-COOCH₂), 3.74 (q, $J = 7.0$ Hz, 2H, N=COOCH₂), 1.20 (t, $J = 7.0$ Hz, 3H, Ph₃P-N-COOCH₂CH₃), 0.94 (t, $J = 7.0$ Hz, 3H, N=COOCH₂CH₃); ¹³C NMR (126 MHz, CDCl₃): δ 168.2 (N=COOCH₂CH₃), 158.2 (t, $J = 23.0$ Hz, Ph₃P-N-COOCH₂CH₃), 134.6 (d, $J = 10.8$ Hz, C-2' and C-6'), 133.8 (d, $J = 2.4$ Hz, C-4'), 128.8 (d, $J = 13.2$ Hz, C-3' and C-5'), 121.7 (d, $J = 103.5$ Hz, C-1'), 63.5 (Ph₃P-N-COOCH₂CH₃), 59.5 (N=COOCH₂CH₃), 15.1 (N=COOCH₂CH₃), 14.1 (Ph₃P-N-COOCH₂CH₃); ¹⁵N NMR (51 MHz, CDCl₃): δ 171 (¹⁵N=CO), 113 (Ph₃P-¹⁵N); ³¹P NMR (202 MHz, CDCl₃): δ +44.2 (s); HRMS (ESI+) (*m/z*): [M+H]⁺ calcd for C₂₄H₂₆(¹⁵N)₂O₄P: 439.1565; found: 439.1564.



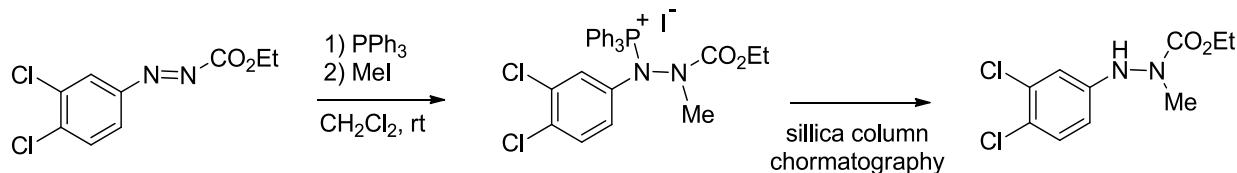
¹H NMR (500 MHz, CDCl₃): δ 7.92–7.86 (m, 6H, H-2' and H-6'), 7.61–7.56 (m, 3H, H-4'), 7.49–7.41 (m, 6H, H-3' and H-5'), 7.14–7.11 (m, 2H, H-2 and H-6), 7.03–6.98 (m, 2H, H-3 and H-5), 6.91–6.86 (m, 1H, H-4), 3.84 (q, J = 7.1 Hz, 2H, -CH₂), 1.04 (t, J_1 = 7.1 Hz, 3H, CH₃); ¹³C NMR (126 MHz, CDCl₃): δ 168.0 (br s, CO), 144.0 (d, J = 12.4 Hz, C-1), 134.6 (d, J = 10.4 Hz, C-2' and C-6'), 132.9 (C-4'), 128.9 (d, J = 13.0 Hz, C-3' and C-5'), 128.1 (C-3 and C-5), 124.8 (d, J = 3.0 Hz, C-2 and C-6), 124.0 (C-4), 59.2 (CH₂), 15.2 (CH₃), (C-1' overlaid); ¹⁵N NMR (51 MHz, CDCl₃): δ 83 (Ar-N); ³¹P NMR (202 MHz, CDCl₃): δ +33.9 (s); HRMS (ESI+) (*m/z*): [M+H]⁺ calcd for C₂₇H₂₆N₂O₂P: 441.1718; found: 441.1717.



¹H NMR (500 MHz, CDCl₃): δ 7.91–7.84 (m, 6H, H-2' and H-6'), 7.67–7.62 (m, 3H, H-4'), 7.54–7.48 (m, 6H, H-3' and H-5'), 7.22–7.18 (m, 1H, H-2), 7.02 (d, J = 8.8 Hz, 1H, H-5), 6.86–6.82 (m, 1H, H-6), 3.82 (q, J = 7.1 Hz, 2H, -CH₂), 1.03 (t, J_1 = 7.1 Hz, 3H, CH₃); ¹³C NMR (126 MHz, CDCl₃): δ 168.0 (d, J_1 = 5.5 Hz), 144.1 (d, J = 13.1 Hz, C-1), 134.6 (d, J = 10.6 Hz, C-2' and C-6'), 134.1 (d, J = 2.4 Hz, C-4'), 131.9 (C-3), 129.4 (C-5), 129.2 (d, J = 13.0 Hz, C-3' and C-5'), 127.1 (C-4), 125.6 (d, J = 3.2 Hz, C-2), 123.1 (d, J = 2.7 Hz, C-6), 122.4 (d, J = 103.3 Hz, C-1'), 59.3 (CH₂), 15.2 (CH₃); ¹⁵N NMR (51 MHz, CDCl₃): δ 85 (Ar-N); ³¹P NMR (202 MHz, CDCl₃): δ +34.5 (s); HRMS (ESI+) (*m/z*): [M+H]⁺ calcd for C₂₇H₂₄Cl₂N₂O₂P: 509.0947; found: 509.0943.

Procedure for methylation of the betaine with MeI:

a) A preparative method



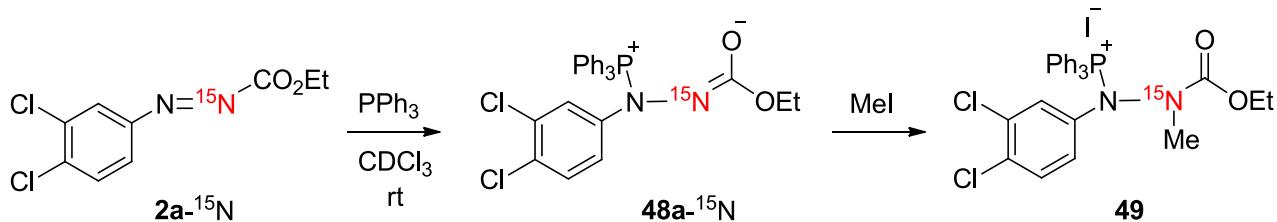
To a solution of ethyl 2-(3,4-dichlorophenyl)diazene-1-carboxylate (**2a**, 247 mg, 1.0 mmol) in dry dichloromethane (distilled over CaH₂), triphenylphosphine (1.05 g, 4.0 mmol) was added under argon atmosphere. The reaction mixture was stirred at room temperature for 5 minutes, and then methyl iodide (710 mg, 5.0 mmol) was added via syringe. After 30 minutes of stirring at room

temperature, the solvent was evaporated under reduced pressure (clean and quantitative conversion of starting diazene). The excess of PPh_3 reacts into phosphonium salt $^+\text{PPh}_3\text{Me}^- \text{I}$.

Methylated adduct: ^1H NMR (500 MHz, CDCl_3): δ 7.98–7.89 (m, 6H, H-2' and H-6'), 7.85–7.60 (H-4', H-3' and H-5' resonances overlaid with resonances for $^+\text{PPh}_3\text{Me}^- \text{I}$), 7.27 (H-5, overlaid), 6.92 (d, $J = 2.7$ Hz, H-2), 6.71 (dd, $J_1 = 8.9$, 2.7 Hz, 1H, H-6), 4.04–3.90 (m, - CH_2CH_3), 3.38 (br s, 3H, N- CH_3), 1.10–1.02 (m, 3H, - CH_2CH_3); HRMS (ESI+) (m/z): $[\text{M}-\text{I}]^+$ calcd for $\text{C}_{28}\text{H}_{26}\text{Cl}_2\text{N}_2\text{O}_2\text{P}$: 523.1103; found: 523.1099.

The crude product (methylated adduct) was subjected to a purification on silica gel column chromatography (dichloromethane/methanol/ammonium hydroxide solution 80/10/1) to obtain ethyl 2-(3,4-dichlorophenyl)-1-methylhydrazine-1-carboxylate as a brown oil. IR (neat, cm^{-1}) ν 3213, 3053, 1704, 1594, 1473, 1433, 1373, 1175; ^1H NMR (500 MHz, CDCl_3): δ 7.27 (d, $J = 8.7$ Hz, H-5), 6.81 (d, $J = 2.4$ Hz, H-2), 6.57 (dd, $J = 8.7$, 2.4 Hz, 1H, H-6), 6.01 (br s, 1H, NH), 4.17 (q, $J = 7.0$ Hz, CH_2), 3.23 (s, 3H, -N- CH_3), 1.24 (br s, 3H, - CH_2CH_3); ^{13}C NMR (126 MHz, CDCl_3): δ 157.0 (CO), 146.8 (C-1), 133.1 (C-3), 130.8 (C-5), 123.6 (C-4), 114.3 (C-2), 112.4 (C-6), 62.5 (CH_2), 37.8 (N- CH_3), 14.6 (CH_3); HRMS (ESI+) (m/z): $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{10}\text{H}_{13}\text{Cl}_2\text{N}_2\text{O}_2$: 263.0349; found: 263.0345.

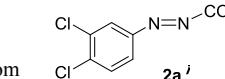
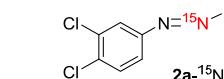
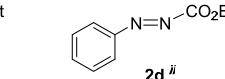
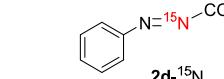
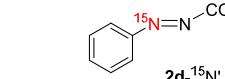
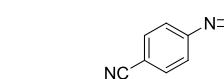
b) The reaction in NMR tube



A similar protocol as described above (preparative method) was used for the NMR experiment with $\text{2a-}^{15}\text{N}$ in CDCl_3 in an NMR tube.

49: ^1H NMR (500 MHz, CDCl_3): δ 7.90–7.40 (N- $^+\text{P}(\text{C}_6\text{H}_5)_3$ resonances overlaid with resonances for $^+\text{PPh}_3\text{Me}^- \text{I}$), 7.18 (H-5, overlaid), 6.82 (d, $J = 2.7$ Hz, 1H, H-2), 6.64 (dd, $J = 8.8$, 2.7 Hz, 1H, H-6), 3.94–3.78 (m, 2H, - CH_2CH_3), 3.27 (br s, 3H, N- CH_3), 1.02–0.90 (m, 3H, CH_3); ^{13}C NMR (126 MHz, CDCl_3): δ 155.2 (br s, CO), 137.8 (d, $J = 13.7$ Hz, C-1), 119.6 (C-2), 118.1 (C-6), 63.4 (CH_2), 38.3 (N- CH_3), 13.6 (CH_3), (C-3, C-4, C-5, N- $^+\text{P}(\text{C}_6\text{H}_5)_3$ resonances overlaid); ^{15}N NMR (51 MHz, CDCl_3): δ 109 ($^{15}\text{N-CO}$), 89 (Ar-N); ^{31}P NMR (202 MHz, CDCl_3): δ +51.1 (s); HRMS (ESI+) (m/z): $[\text{M}-\text{I}]^+$ calcd for $\text{C}_{28}\text{H}_{26}\text{Cl}_2\text{N}(^{15}\text{N})\text{O}_2\text{P}$: 524.1074; found: 524.1070.

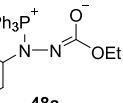
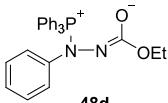
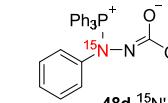
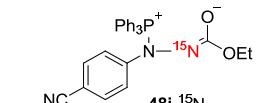
Table S3: NMR data of ethyl 2-arylazocarboxylates (2)

Atom						
CH ₃	1.48 (t, <i>J</i> = 7.2 Hz, 3H)	1.47 (t, <i>J</i> = 7.1 Hz, 3H)	1.47 (t, <i>J</i> = 7.1 Hz, 3H)	1.47 (t, <i>J</i> = 7.1 Hz, 3H)	1.48 (t, <i>J</i> = 7.1 Hz, 3H)	1.48 (t, <i>J</i> = 7.2 Hz, 3H)
CH ₂	4.53 (q, <i>J</i> = 7.2 Hz, 2H)	4.53 (q, <i>J</i> = 7.1 Hz, 2H)	4.52 (q, <i>J</i> = 7.1 Hz, 2H)	4.52 (q, <i>J</i> = 7.1 Hz, 2H)	4.53 (q, <i>J</i> = 7.1 Hz, 2H)	4.55 (q, <i>J</i> = 7.2 Hz, 2H)
H2	8.02 (d, <i>J</i> _{H2-H6} = 2.3 Hz, 1H)	8.02 (d, <i>J</i> _{H2-H6} = 2.2 Hz, 1H)	7.92–7.96 (m, 1H)	7.91–7.96 (m, 1H)	7.92–7.96 (m, 1H)	8.00 (d, <i>J</i> _{H2-H3} = 8.5 Hz, 1H)
H3	–	–	7.51–7.56 (m, 1H)	7.50–7.56 (m, 1H)	7.50–7.56 (m, 1H)	7.85 (d, <i>J</i> _{H2-H3} = 8.5 Hz, 1H)
H4	–	–	7.57–7.61 (m, 1H)	7.56–7.61 (m, 1H)	7.57–7.62 (m, 1H)	–
H5	7.64 (d, <i>J</i> _{H5-H6} = 8.5 Hz, 1H)	7.63 (d, <i>J</i> _{H5-H6} = 8.6 Hz, 1H)	7.51–7.56 (m, 1H)	7.50–7.56 (m, 1H)	7.50–7.56 (m, 1H)	7.85 (d, <i>J</i> _{H5-H6} = 8.5 Hz, 1H)
H6	7.81 (dd, <i>J</i> _{H5-H6} = 8.5 Hz, <i>J</i> _{H2-H6} = 2.3 Hz, 1H)	7.81 (dd, <i>J</i> _{H5-H6} = 8.6 Hz, <i>J</i> _{H2-H6} = 2.2 Hz, 1H)	7.92–7.96 (m, 1H)	7.91–7.96 (m, 1H)	7.92–7.96 (m, 1H)	8.00 (d, <i>J</i> _{H5-H6} = 8.5 Hz, 1H)
CO	161.7 (s)	161.6 (s)	162.2 (s)	162.2 (s)	162.2 (d, <i>J</i> _{CO-ArN} = 6.3 Hz)	161.6 (s)
C1	150.3 (s)	150.3 (d, <i>J</i> _{C1-NCO} = 5.3 Hz)	151.6 (s)	151.6 (d, <i>J</i> _{C1-NCO} = 4.9 Hz)	151.6 (d, <i>J</i> _{C1-ArN} = 2.8 Hz)	153.1 (d, <i>J</i> _{C1-NCO} = 5.3 Hz)
C2	124.6 (s)	124.6 (d, <i>J</i> _{C2-NCO} = 4.1 Hz)	123.7 (s)	123.7 (d, <i>J</i> _{C2-NCO} = 3.8 Hz)	123.7 (d, <i>J</i> _{C2-ArN} = 4.3 Hz)	124.0 (d, <i>J</i> _{C2-NCO} = 4.1 Hz)
C3	134.0 (s)	134.0 (s)	129.3 (s)	129.3 (s)	129.3 (d, <i>J</i> _{C3-ArN} = 1.8 Hz)	133.4 (s)
C4	138.1 (s)	138.1 (s)	133.8 (s)	133.8 (s)	133.8	116.7 (s)
C5	131.2 (s)	131.2 (s)	129.3 (s)	129.3 (s)	129.3 (d, <i>J</i> _{C3-ArN} = 1.8 Hz)	133.4 (s)
C6	123.6 (s)	123.6 (d, <i>J</i> _{C6-NCO} = 3.9 Hz)	123.7 (s)	123.7 (d, <i>J</i> _{C6-NCO} = 3.8 Hz)	123.7 (d, <i>J</i> _{C6-ArN} = 4.3 Hz)	124.0 (d, <i>J</i> _{C6-NCO} = 4.1 Hz)
CN	–	–	–	–	–	117.8
ArN	142	ND	149	149	149	145
NCO	ND	114	ND	107	ND	125

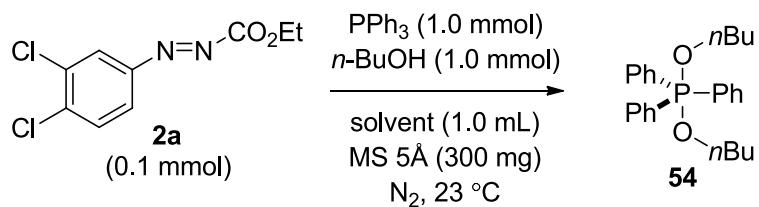
ⁱPrepared by the literature procedure.^{2a}

ⁱⁱPrepared by the literature procedure.^{2b}

Table S4: NMR data of betaine intermediates 48

Atom						
CH ₃	1.03 (t, <i>J</i> = 7.1 Hz, 3H)	1.03 (t, <i>J</i> = 7.0 Hz, 3H)	1.04 (t, <i>J</i> = 7.1 Hz, 3H)	1.04 (t, <i>J</i> = 7.0 Hz, 3H)	1.01–1.06 (m, 3H)	0.96 (t, <i>J</i> = 7.0 Hz, 3H)
CH ₂	3.82 (q, <i>J</i> = 7.1 Hz, 2H)	3.82 (q, <i>J</i> = 7.0 Hz, 2H)	3.84 (q, <i>J</i> = 7.0 Hz)	3.82 (q, <i>J</i> = 7.0 Hz)	3.80–3.86 (m, 3H)	3.71 (q, <i>J</i> = 7.0 Hz, 2H)
H2	7.18–7.22 (m, 1H)	7.17–7.21 (m, 1H)	7.11–7.14 (m, 1H)	7.09–7.14 (m, 1H)	7.10–7.15 (m, 1H)	7.00 (d, <i>J</i> _{H2–H3} = 8.5 Hz, 1H)
H3	–	–	6.98–7.03 (m, 1H)	6.97–7.03 (m, 1H)	6.97–7.03 (m, 1H)	7.27 (d, <i>J</i> _{H2–H3} = 8.5 Hz, 1H)
H4	–	–	6.86–6.91 (m, 1H)	6.86–6.91 (m, 1H)	6.86–6.91 (m, 1H)	–
H5	7.02 (d, <i>J</i> _{H5–H6} = 8.8 Hz, 1H)	7.02 (d, <i>J</i> _{H5–H6} = 8.7 Hz, 1H)	6.98–7.03 (m, 1H)	6.97–7.03 (m, 1H)	6.97–7.03 (m, 1H)	7.27 (d, <i>J</i> _{H5–H6} = 8.5 Hz, 1H)
H6	6.82–6.86 (m, 1H)	6.82–6.86 (m, 1H)	7.11–7.14 (m, 1H)	7.09–7.14 (m, 1H)	7.10–7.15 (m, 1H)	7.00 (d, <i>J</i> _{H5–H6} = 8.5 Hz, 1H)
H2',H6'	7.84–7.91 (m, 6H)	7.83–7.91 (m, 6H)	7.86–7.92 (m, 6H)	7.84–7.91 (m, 6H)	7.84–7.91 (m, 6H)	7.83–7.92 (m, 6H)
H3',H5'	7.48–7.54 (m, 6H)	7.49–7.54 (m, 6H)	7.41–7.49 (m, 6H)	7.40–7.51 (m, 6H)	7.41–7.51 (m, 6H)	7.49–7.56 (m, 6H)
H4'	7.62–7.67 (m, 3H)	7.62–7.67 (m, 3H)	7.56–7.61 (m, 3H)	7.57–7.63 (m, 3H)	7.56–7.61 (m, 3H)	7.65–7.71 (m, 3H)
CH ₃	15.2 (s)	15.2 (s)	15.2 (s)	15.2 (s)	15.2 (s)	15.1 (s)
CH ₂	59.3 (s)	59.3 (s)	59.2 (s)	59.1 (s)	59.1 (s)	59.3 (s)
CO	168.0 (d, <i>J</i> _{CO-P} = 5.5 Hz)	168.0 (dd, <i>J</i> = 5.5, 1.8 Hz)	168.0 (br s)	168.1 (d, <i>J</i> _{CO-P} = 6.4 Hz)	168.1 (d, <i>J</i> _{CO-P} = 6.1 Hz)	168.4 (br s)
C1	144.1 (d, <i>J</i> _{C1-P} = 13.1 Hz)	144.1 (d, <i>J</i> _{C1-P} = 12.8 Hz)	144.0 (d, <i>J</i> _{C1-P} = 12.4 Hz)	144.1 (d, <i>J</i> _{C1-P} = 12.1 Hz)	144.0 (dd, <i>J</i> _{C1-P} = 12.4 Hz, <i>J</i> _{C1-N} = 12.4 Hz)	149.4 (d, <i>J</i> _{C1-P} = 15.7 Hz)
C2	125.6 (d, <i>J</i> = 3.2 Hz)	125.6 (d, <i>J</i> = 3.1 Hz)	124.8 (d, <i>J</i> = 3.0 Hz)	124.8 (s)	124.7 (br s)	120.6 (d, <i>J</i> = 3.2 Hz)
C3	131.9 (s)	131.9 (s)	128.1 (s)	128.1 (s)	128.2 (d, <i>J</i> _{C3-N} = 7.0 Hz)	132.0 (s)
C4	127.1 (s)	127.1 (s)	124.0 (s)	124.0 (s)	124.0 (s)	104.7 (s)
C5	129.4 (s)	129.4 (s)	128.1 (s)	128.1 (s)	128.2 (d, <i>J</i> _{C5-N} = 7.0 Hz)	132.0 (s)
C6	123.1 (d, <i>J</i> = 2.7 Hz)	123.1 (d, <i>J</i> = 2.5 Hz)	124.8 (d, <i>J</i> = 3.0 Hz)	124.8 (s)	124.7 (br s)	120.6 (d, <i>J</i> = 3.2 Hz)
C1'	122.4 (d, <i>J</i> _{P-C} = 103.3 Hz)	122.4 (d, <i>J</i> _{P-C} = 103.3 Hz)	ND	121.7 (d, <i>J</i> _{P-C} = 103.0 Hz)	ND	121.7 (d, <i>J</i> _{P-C} = 103.1 Hz)
C2',C6'	134.6 (d, <i>J</i> _{P-C} = 10.6 Hz)	134.6 (d, <i>J</i> _{P-C} = 10.5 Hz)	134.6 (d, <i>J</i> = 10.4 Hz)	134.6 (d, <i>J</i> _{P-C} = 10.3 Hz)	134.6 (d, <i>J</i> _{P-C} = 10.3 Hz)	134.7 (d, <i>J</i> _{P-C} = 10.6 Hz)
C3',C5'	129.2 (d, <i>J</i> _{P-C} = 13.0 Hz)	129.1 (d, <i>J</i> _{P-C} = 12.9 Hz)	128.9 (d, <i>J</i> = 13.0 Hz)	128.9 (d, <i>J</i> _{P-C} = 13.1 Hz)	128.9 (d, <i>J</i> _{P-C} = 12.9 Hz)	129.2 (d, <i>J</i> _{P-C} = 13.2 Hz)
C4'	134.1 (d, <i>J</i> _{P-C} = 2.4 Hz)	134.1 (d, <i>J</i> _{P-C} = 2.5 Hz)	132.9 (br s)	133.3 (d, <i>J</i> _{P-C} = 1.9 Hz)	133.3 (d, <i>J</i> _{P-C} = 2.3 Hz)	134.2 (d, <i>J</i> _{P-C} = 2.6 Hz)
CN	–	–	–	–	–	119.2
P	34.5 (s)	34.5 (d, <i>J</i> _{P-NCO} = 5.0 Hz)	33.9 (s)	33.3 (d, <i>J</i> _{P-NCO} = 5.2 Hz)	33.4 (d, <i>J</i> _{P-NCO} = 6.2 Hz)	35.4 (s)
ArN	85	ND	83	83	84	90
NCO	ND	182	ND	182	ND	179

*Detection of di-n-butoxytriphenylphosphorane (**54**) by ^{31}P NMR analysis in several solvents:*



A solution of ethyl 2-(3,4-dichlorophenyl)azocarboxylate (**2a**) (24.7 mg, 0.1 mmol), *n*-BuOH (74.1 mg, 1.0 mmol) and powder of activated molecular sieves 5Å (300 mg) in THF-*d*₈ (1.0 mL) was stirred in a 10 mL round-bottom flask for 30 min. Triphenylphosphine (262 mg, 1.0 mmol) was added to the stirred solution at 23 °C under N₂ atmosphere. A small amount (ca. 0.5 mL) of a sample was taken from the reaction mixture, and rapidly filtered through a cotton plug. The obtained clear solution was analyzed by ^{31}P NMR.

Table S5. Chemical shifts of di-*n*-butoxytriphenylphosphorane in ^{31}P NMR spectrum

entry	solvent	^{31}P NMR (243 MHz) δ (ppm)
1	THF- <i>d</i> ₈	-56.0 (-55.0) ¹⁹
2	CDCl ₃	-55.3
3	CD ₃ CN	-55.2
4	toluene- <i>d</i> ₈	-55.8

4. Thermal analysis of azo and hydrazine compounds

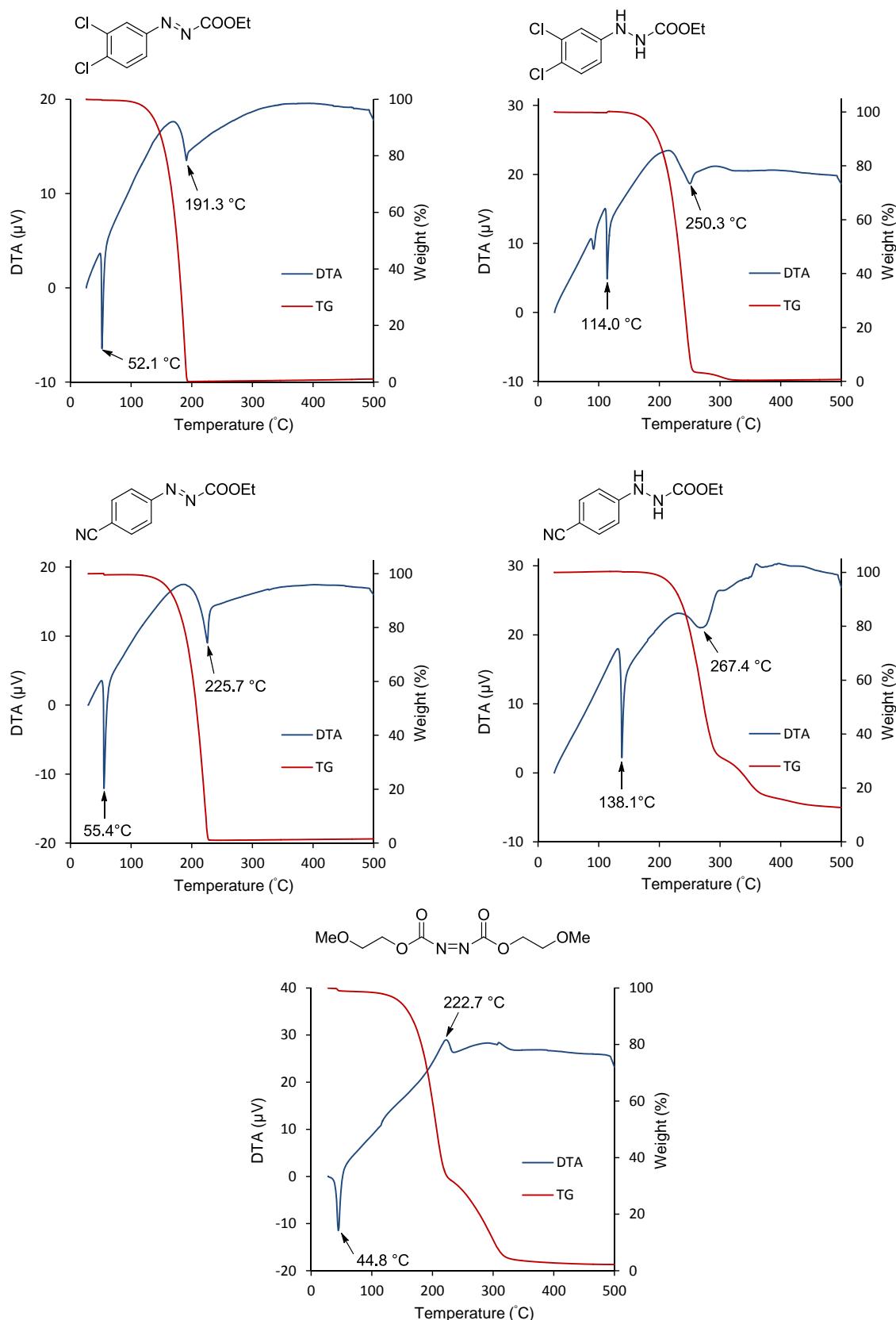


Figure S15. TG-DTA data of azo and hydrazine compounds.

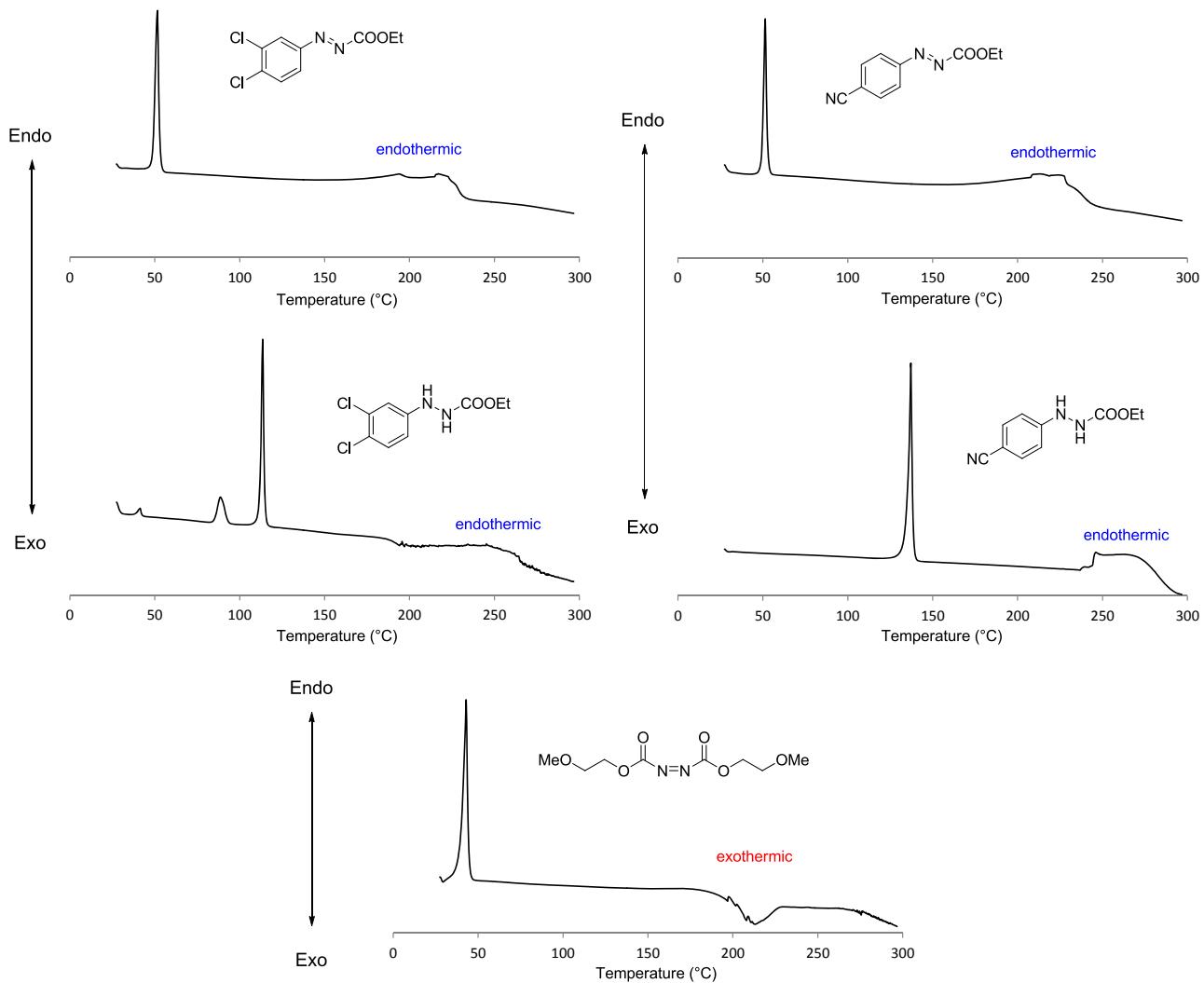
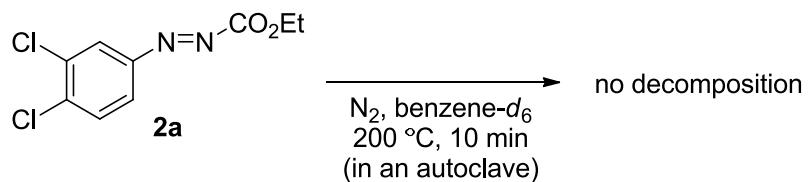


Figure S16. Preliminary DSC data of azo and hydrazine compounds.



Figure S17. Photos of Mitsunobu reagents (from left to right, **1a**, **1j**, **2a**, **2j**). No decomposition of these reagents was observed after two months under ambient conditions unlike DMEAD (See the NMR spectra data).

Procedure for thermal analysis of **2a** in a solution-phase:



A solution of ethyl 2-(3,4-dichlorophenyl)azocarboxylate (**2a**) (9.4 mg, 0.038 mmol) in benzene-*d*₆ (1 mL) was placed into an autoclave under an nitrogen atmosphere. Heating the autoclave was begun with a pre-heating oil bath (180 °C). The temperature was raised to 200 °C over 15 min, and the autoclave was further heated for 10 min at the same temperature. After the autoclave was cooled to room temperature and opened, the solution was analyzed by ¹H NMR (400 MHz) analysis. An amount of **2a** was estimated by integration values in the presence of 1,3,5-trioxane as an internal standard (full relaxation was confirmed). Neither loss of **2a** nor detection of any impurities was observed.

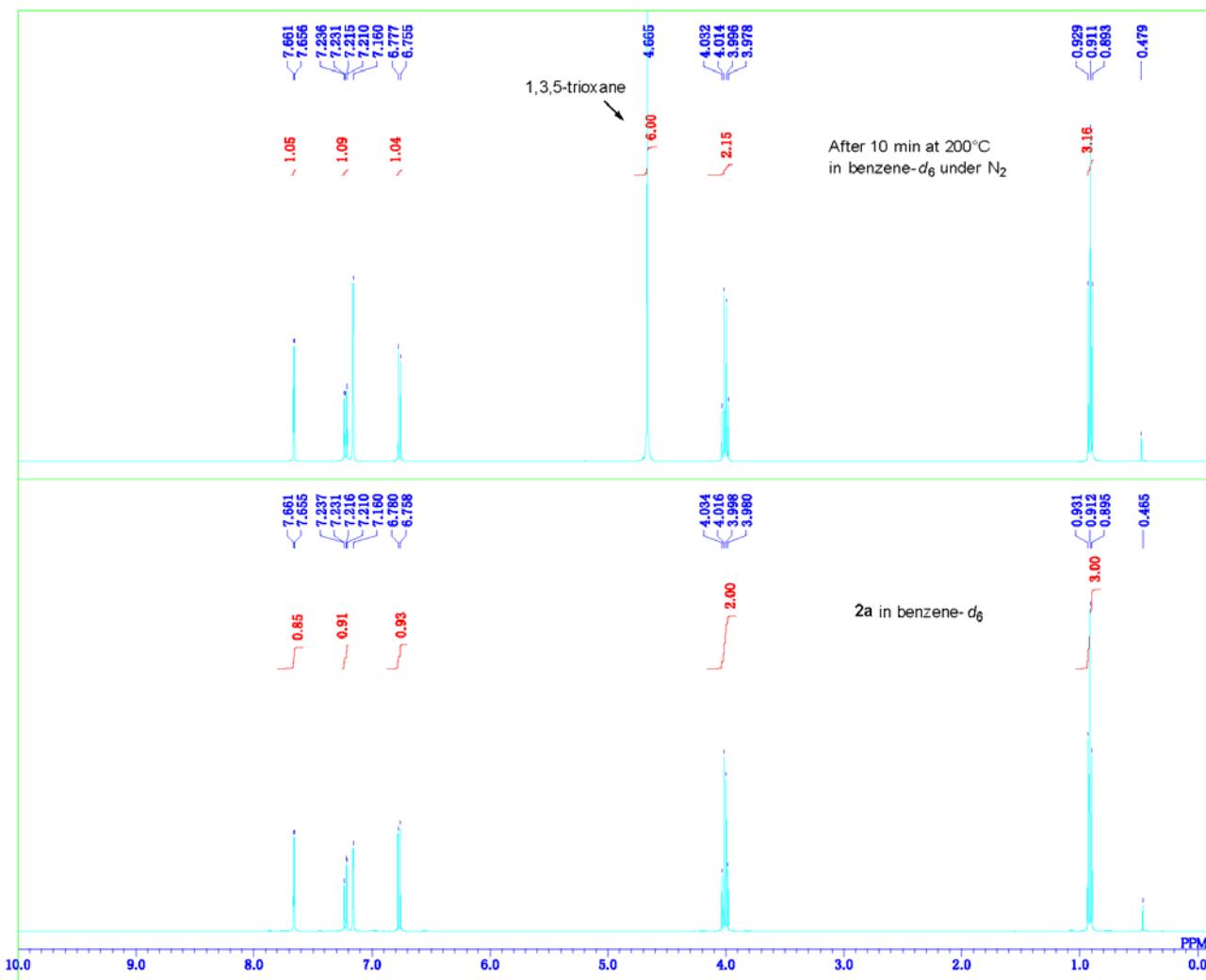


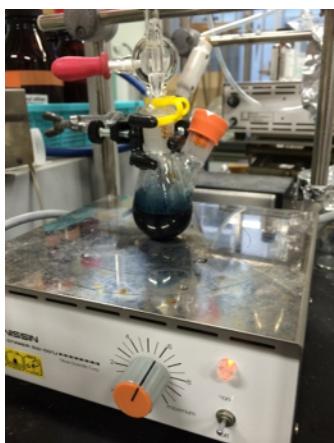
Figure S18. ¹H NMR spectra of **2a** after heating at 200 °C for 10 min (top), and before heating (bottom).

**5. Photos of the typical experiment of the catalytic Mitsunobu reaction on 10 mmol scale
(Table 3, entry 7)**

(a) Activation of MS 5Å by a heat gun in vacuo



(b) The reaction mixture



(c) The reaction mixture after filtration with Celite®



(d) Silica gel column chromatography (yellow elution is azo compound)



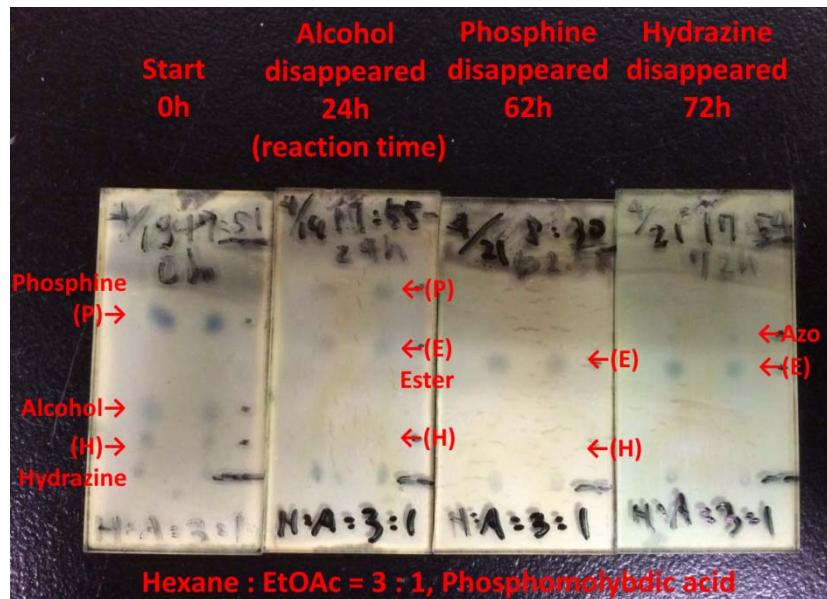
(e) After Silica gel column chromatography (left elute: **2a**, right elute: **5**)



5: 88% yield (2.35 g, 8.8 mmol, 99:1 er)

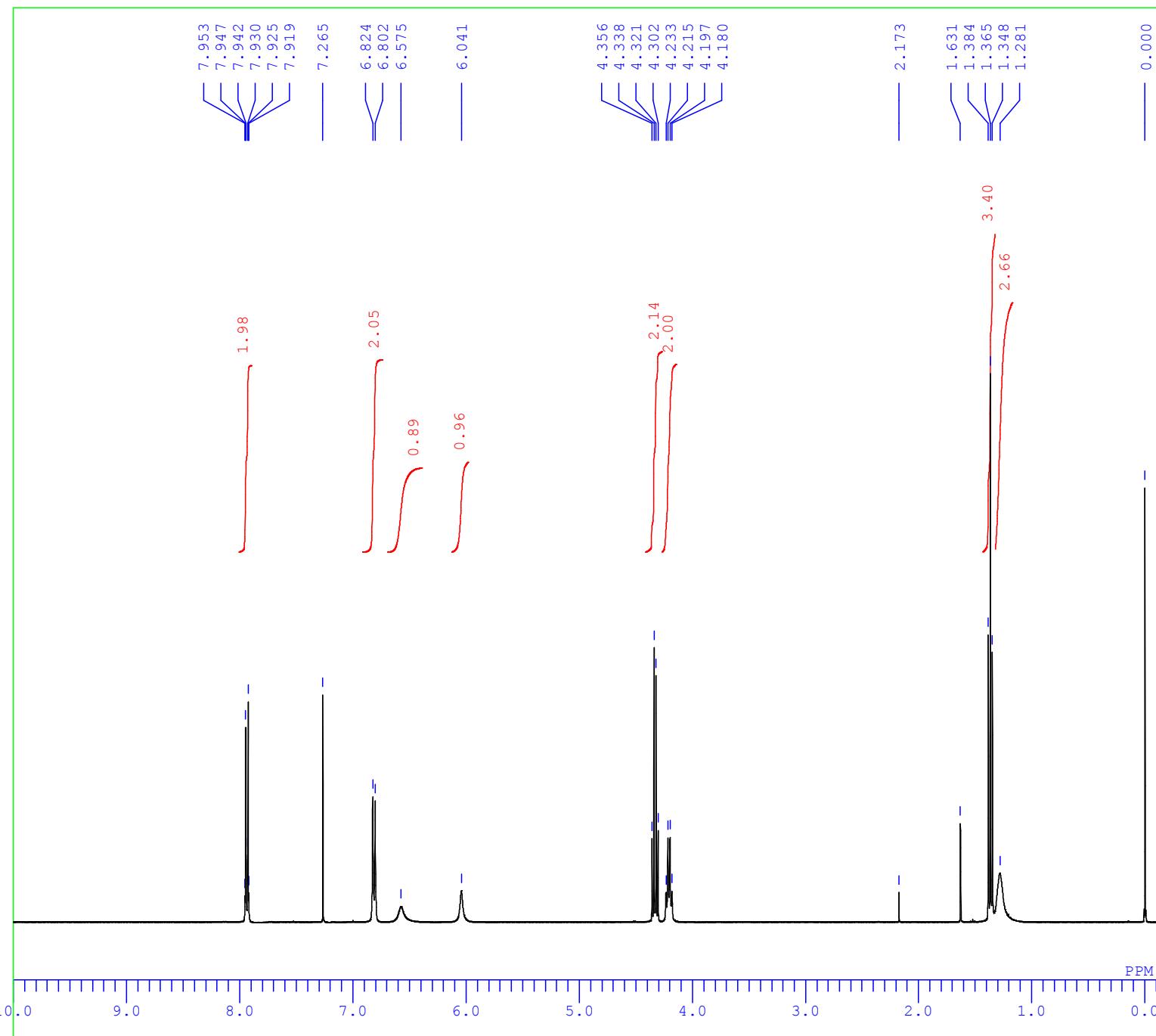
2a: 81% recovery (200 mg, 0.81 mmol)

(f) TLC at 0, 24, 62, 72 h

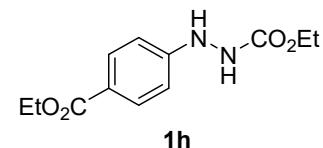


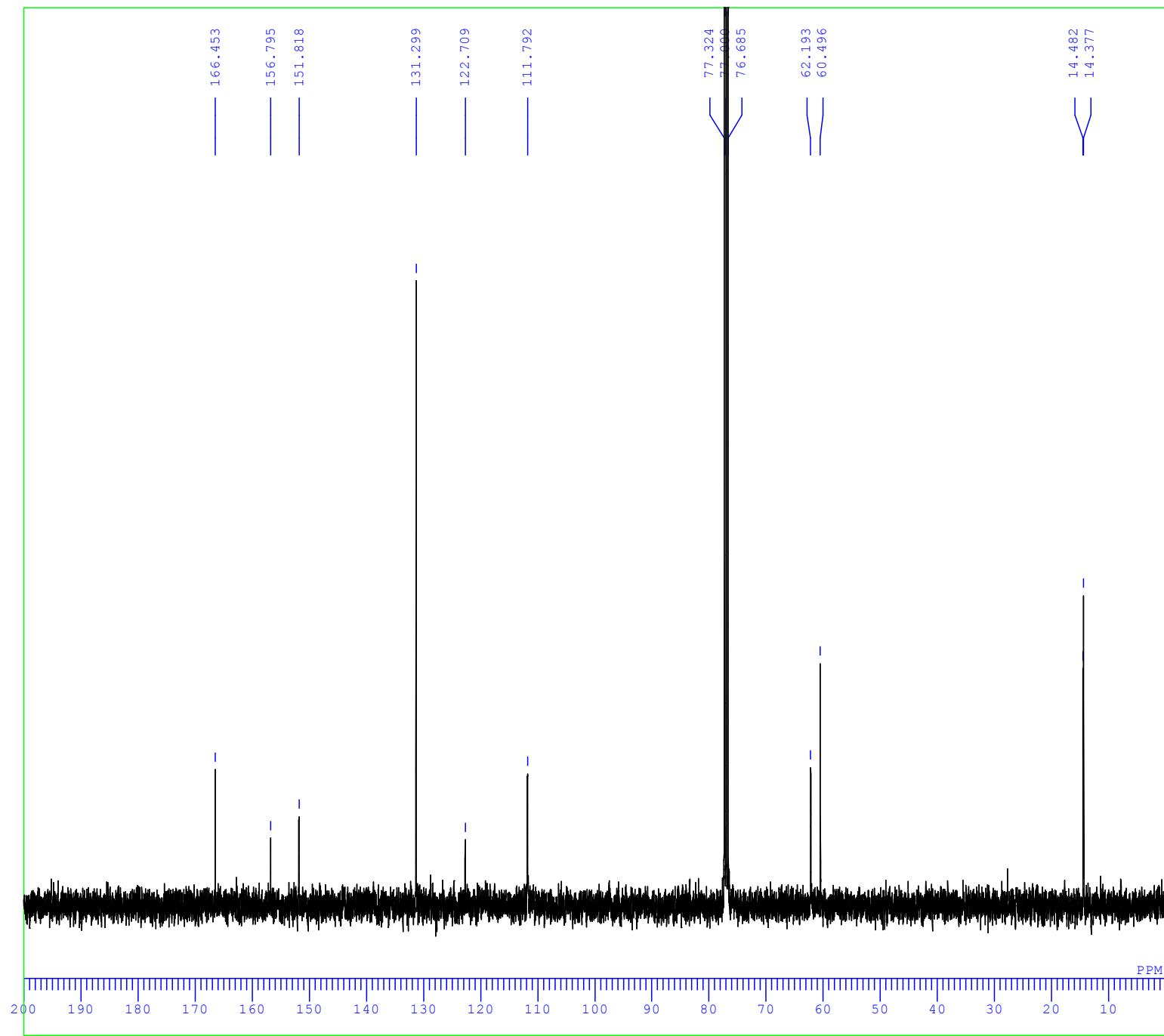
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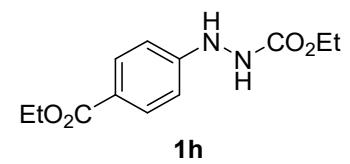


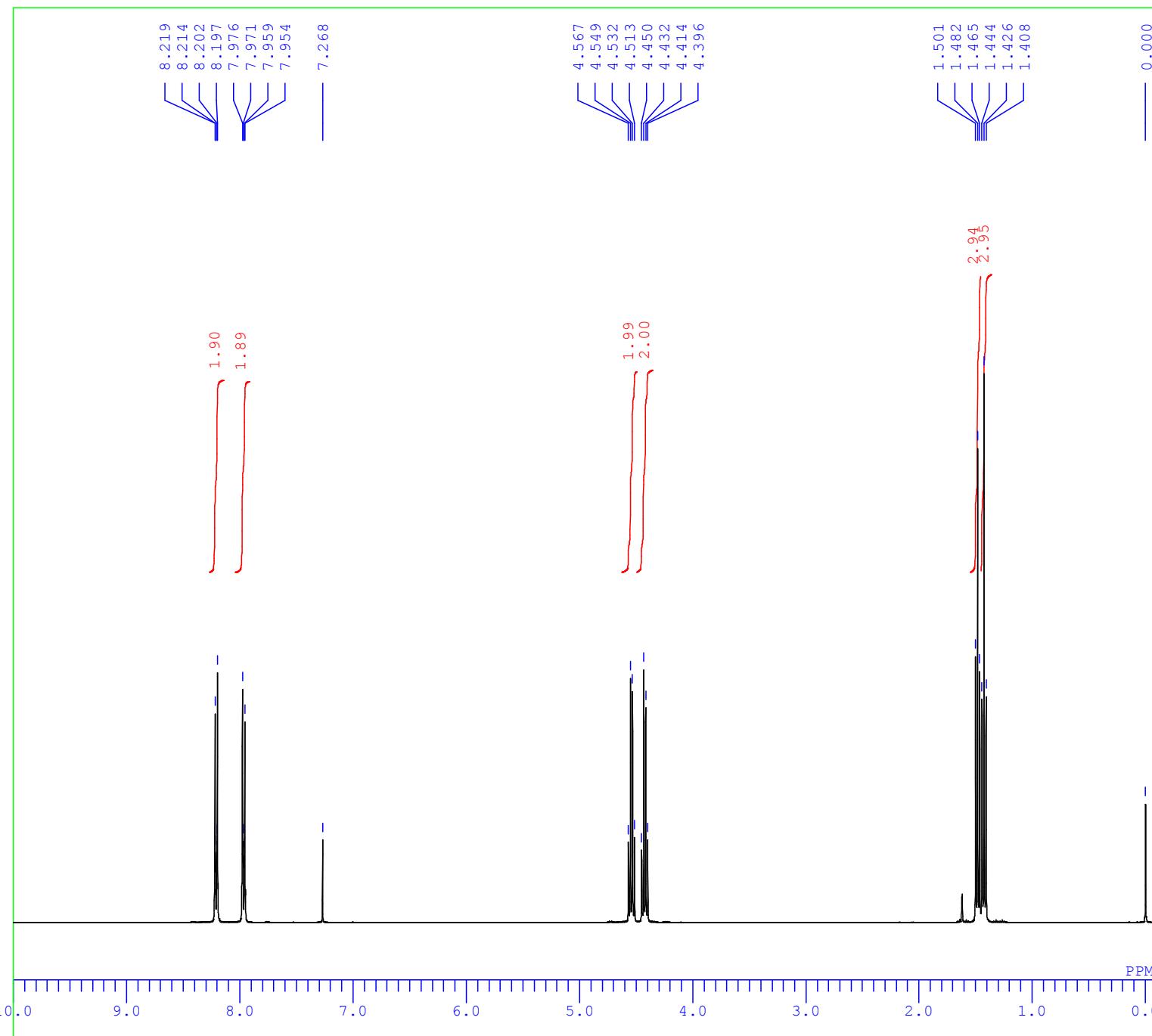
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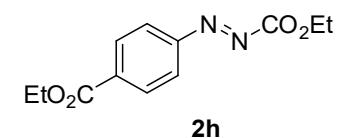


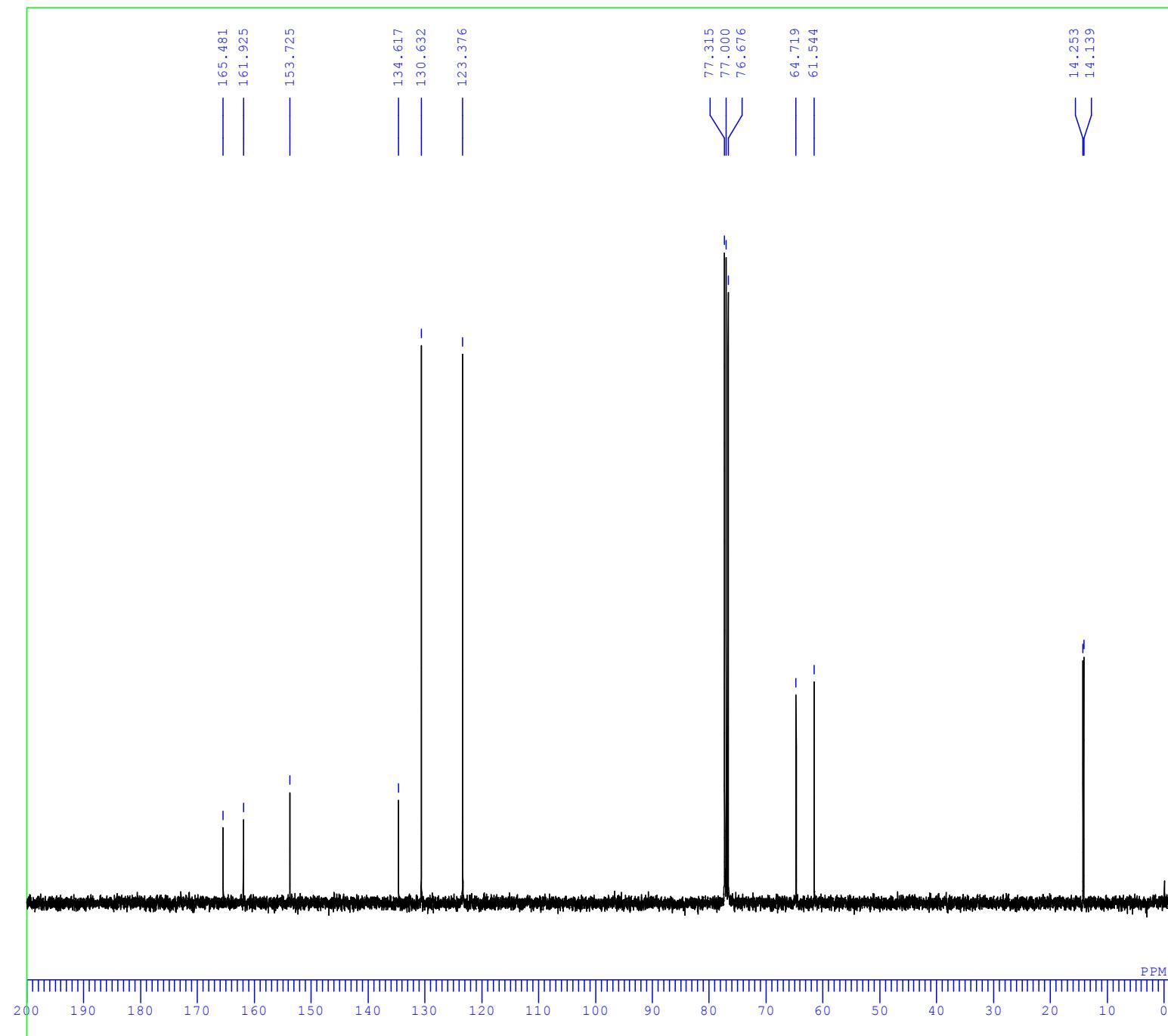
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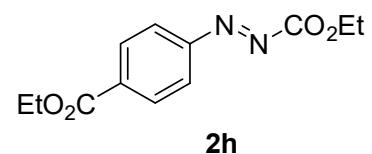


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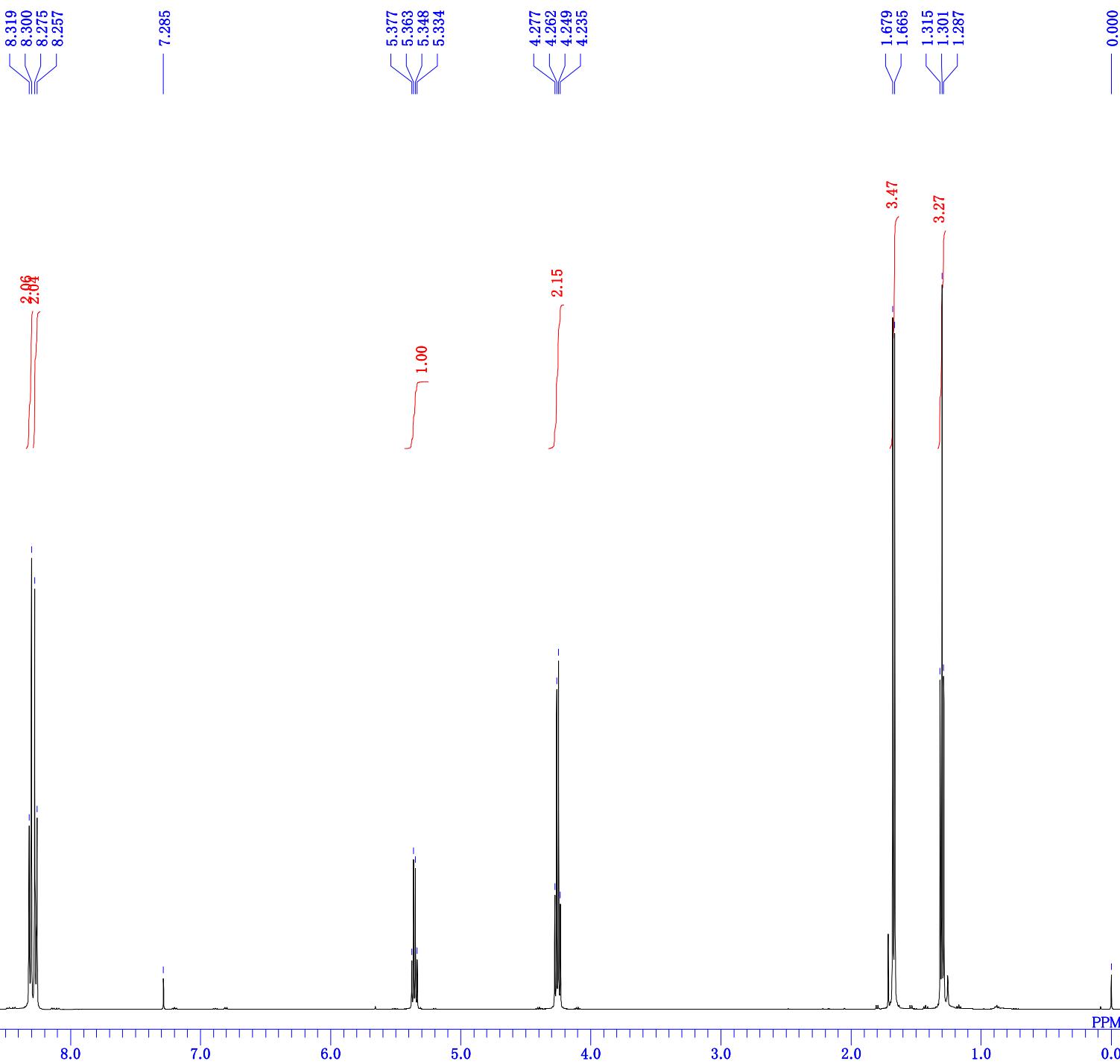




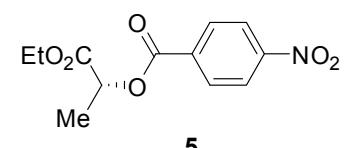
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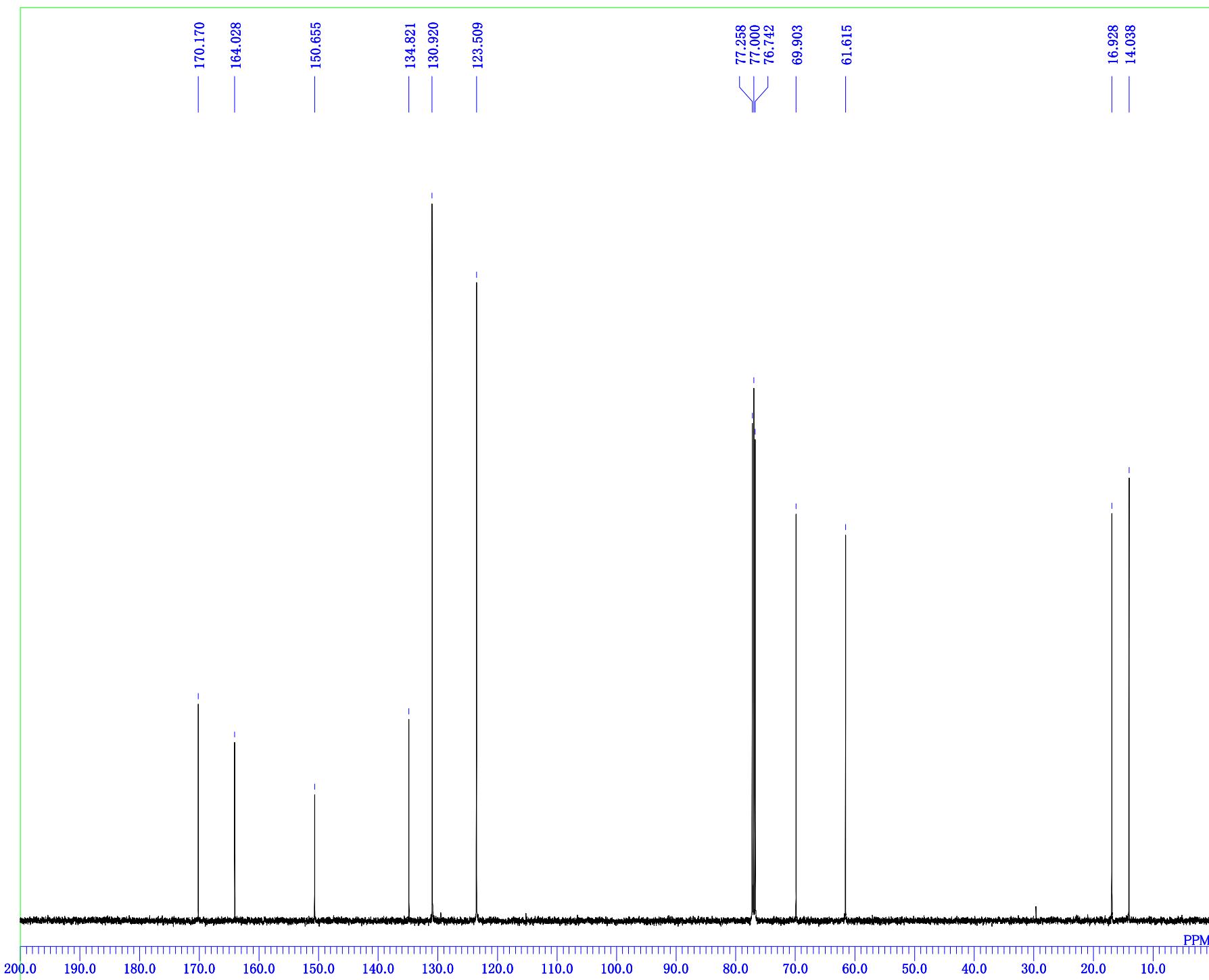


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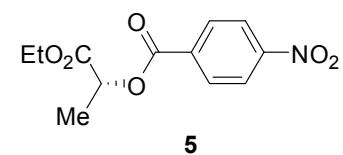


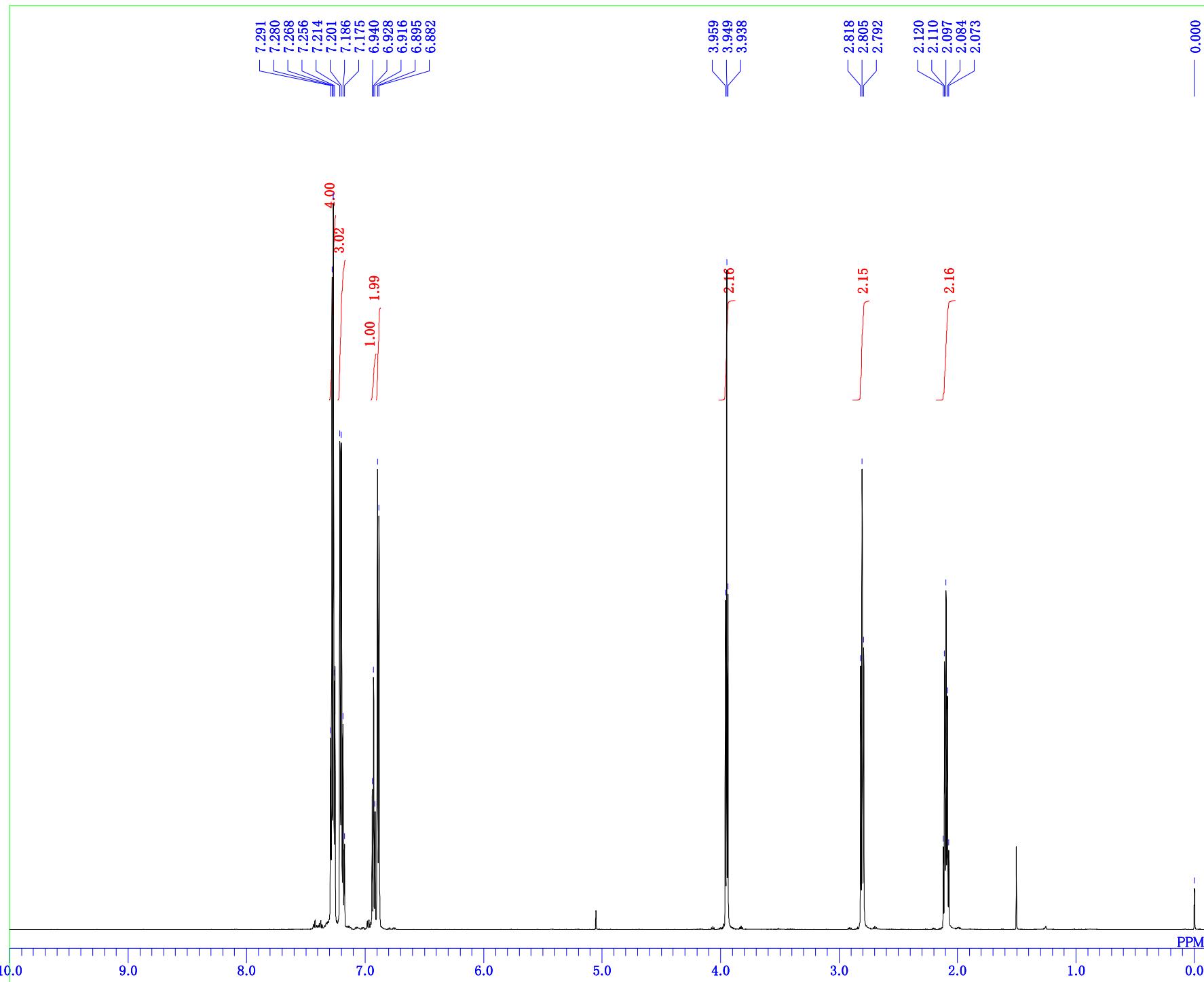
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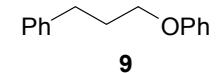


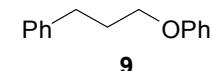
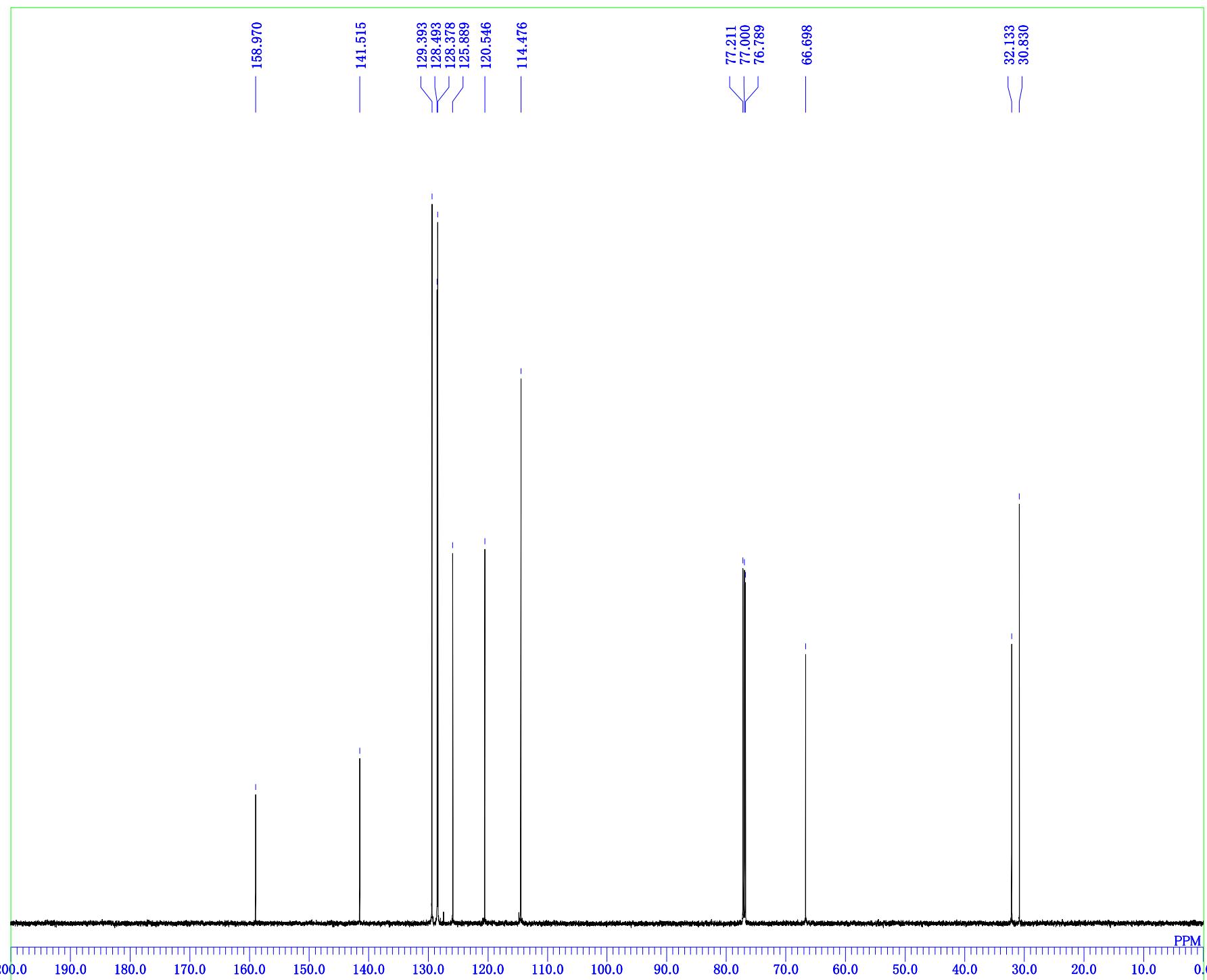
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 RGAIN 58



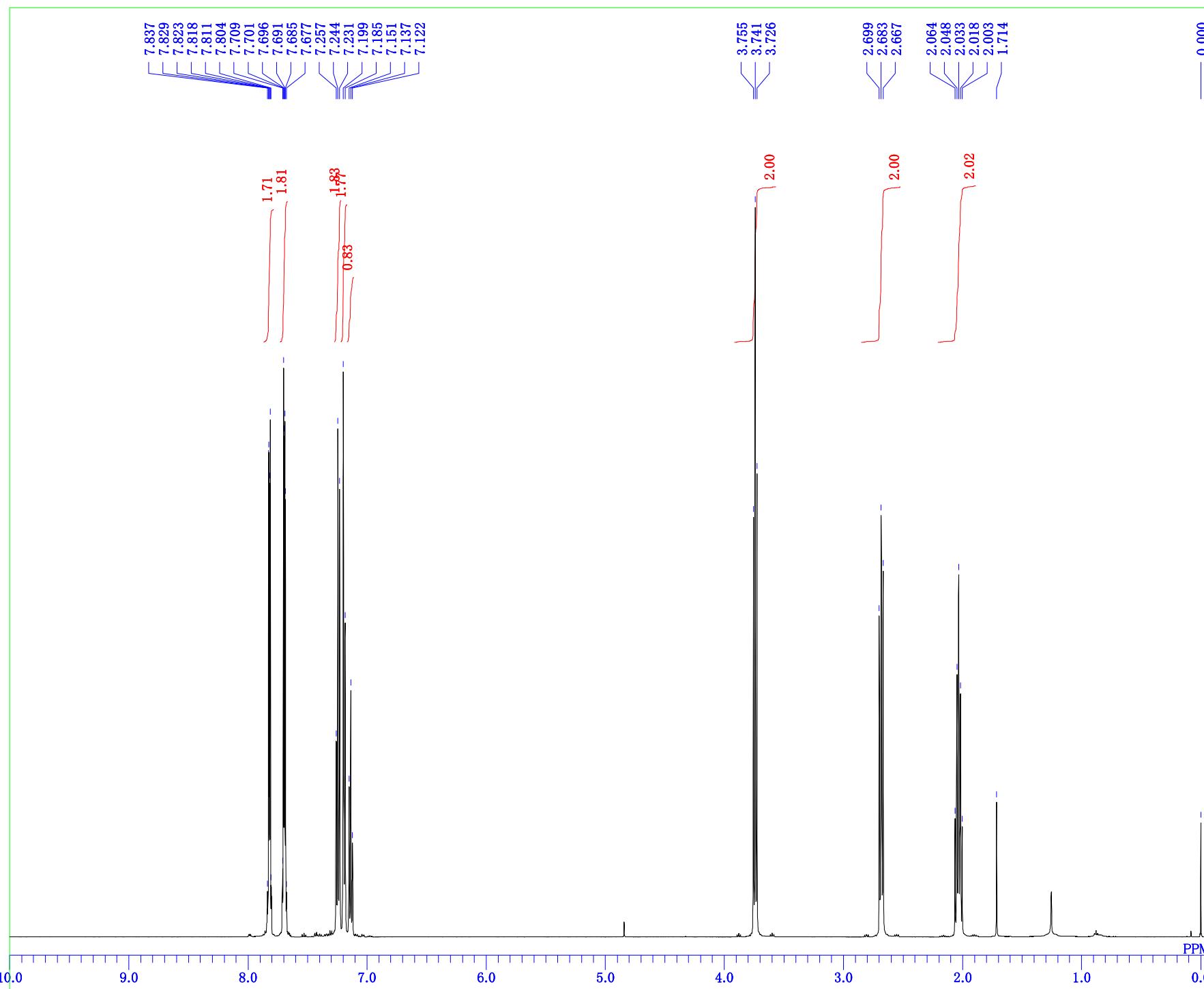


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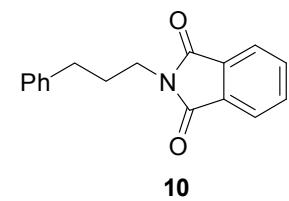


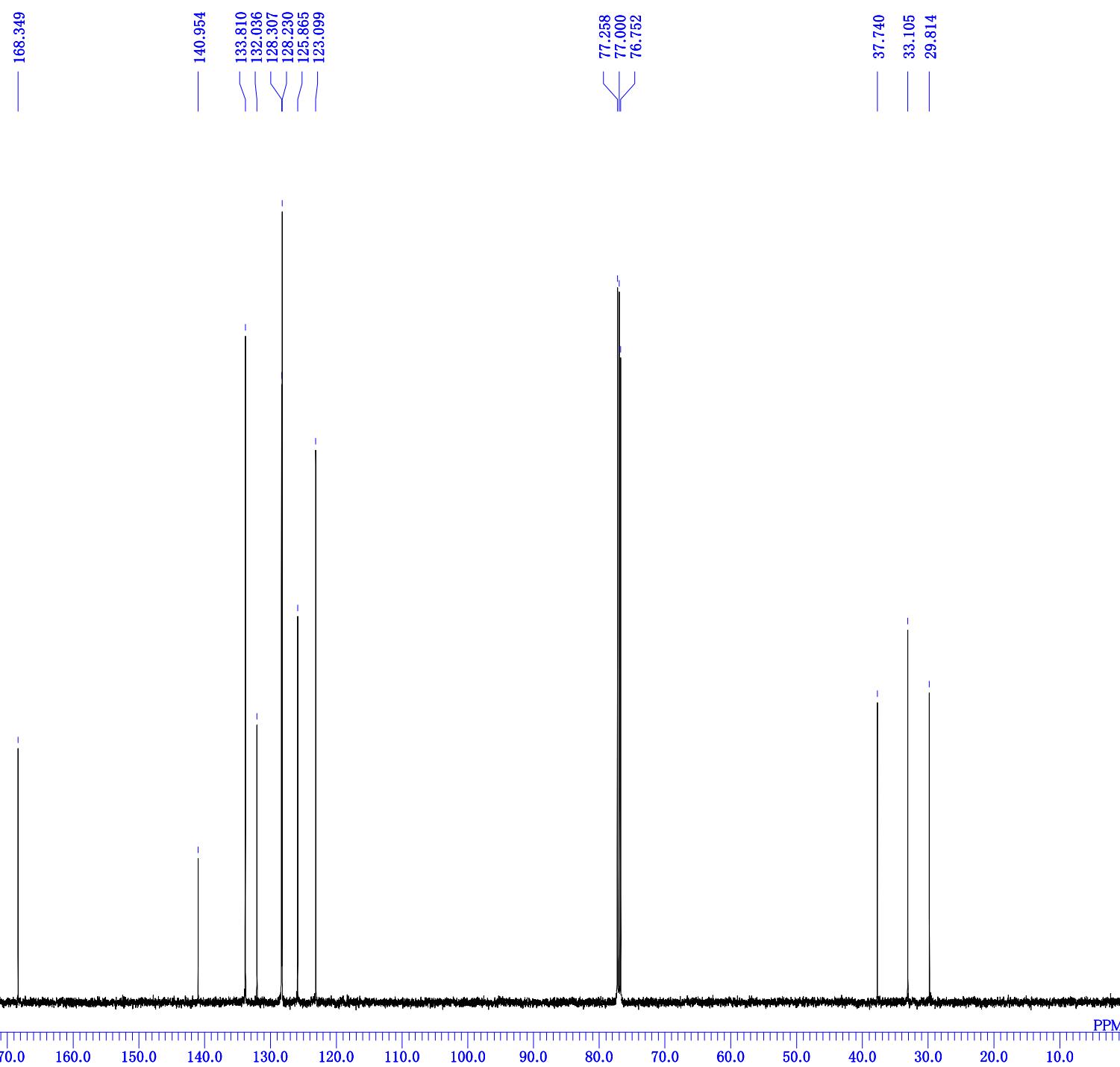


150704

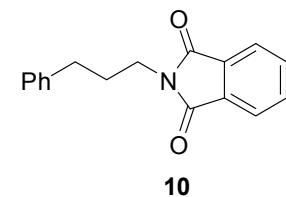


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 OBFRQ 2.41 KHz
 OBSET 6.01 Hz
 OBFIN 13107
 POINT 7507.39 Hz
 FREQU 32
 SCANS 1.7459 sec
 ACQTM 2.0000 sec
 PD 5.80 usec
 1H 20.0 c
 IRNUC CDCL3
 CTEMP 0.00 ppm
 SLVNT 0.12 Hz
 EXREF 34
 RGAIN

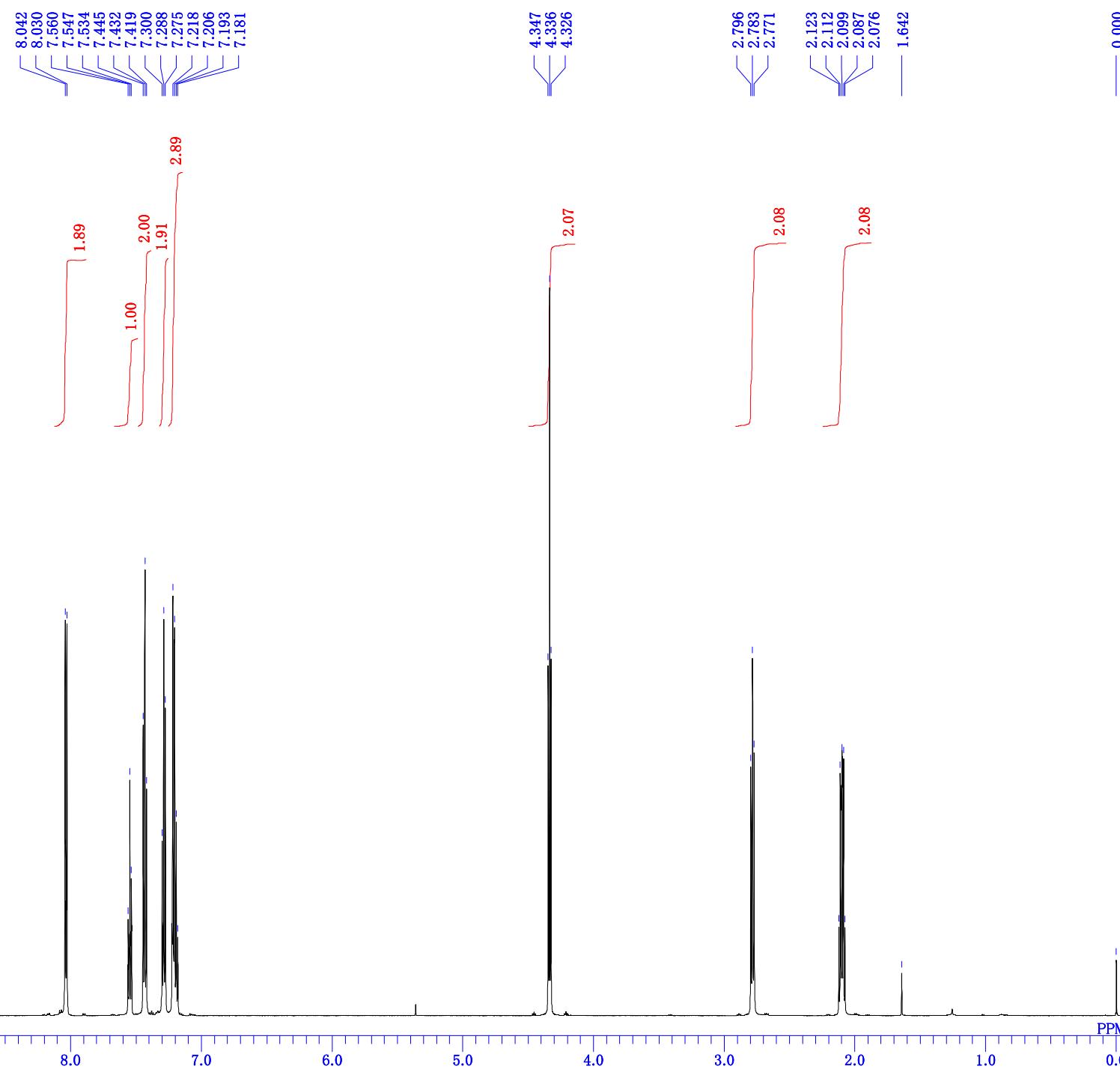




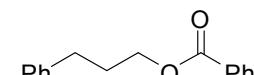
DFILE d1500-gra-13c-1.als
 COMNT 150704
 DATIM 2015-07-04 17:24:07
 OBNUC 13C
 EXMOD single_pulse_dec
 OBFRQ 125.77 MHz
 OBSET 7.87 KHz
 OBFIN 4.21 Hz
 POINT 26214
 FREQU 31446.06 Hz
 SCANS 256
 ACQTM 0.8336 sec
 PD 2.0000 sec
 PW1 3.00 usec
 IRNUC 1H
 CTEMP 20.5 c
 SLVNT CDCL₃
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 60



150701

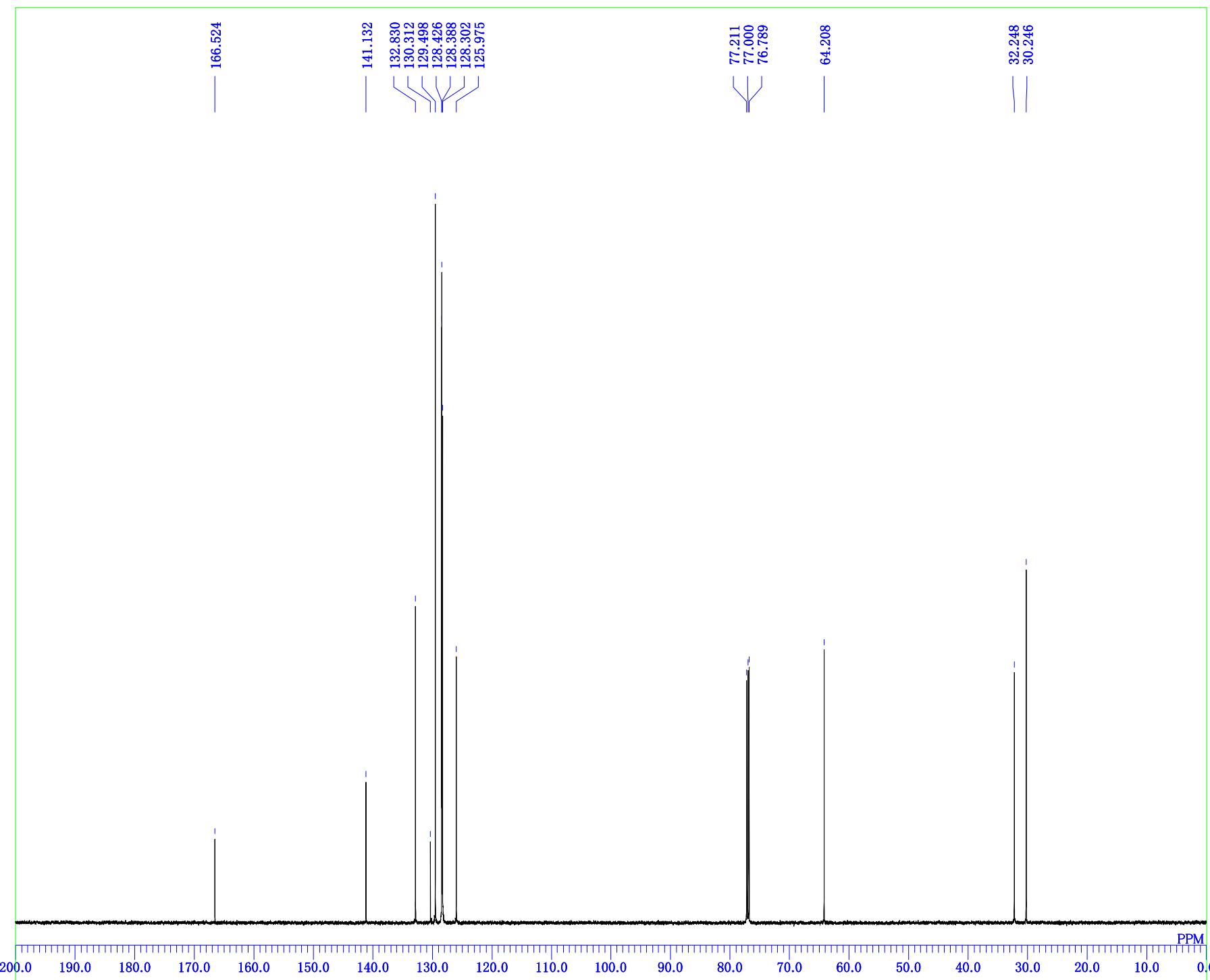


DFILE d1505-gra-1h-1.als
 COMNT 150701
 DATIM 2015-03-06 16:23:16
 1H single_pulse.ex2
 EXMOD 600.17 MHz
 OBFRQ 5.30 KHz
 OBSET 5.47 Hz
 OBFIN 26214
 POINT 26214
 FREQU 9008.87 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 5.85 usec
 IRNUC 1H
 CTEMP 20.4 c
 SLVNT CDCL₃
 EXREF 0.00 ppm
 BF 0.12 Hz
 RGAIN 30

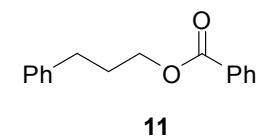


11

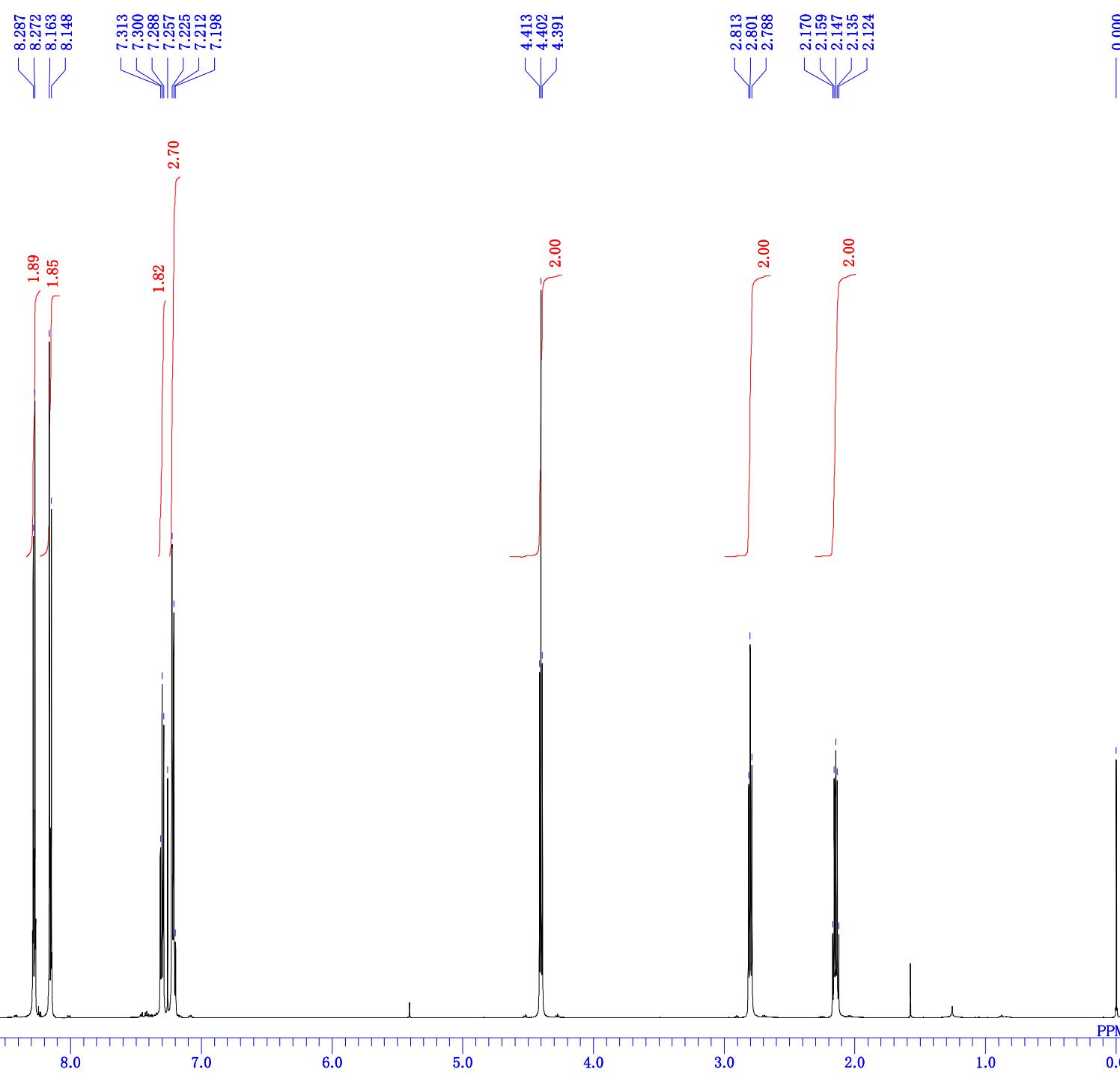
S60



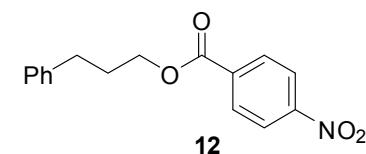
DFILE d1505-gra-13c-1.als
 COMNT 150701
 DATIM 2015-03-06 16:32:54
 13C 13C
 EXMOD single_pulse_dec
 OBFRQ 150.92 MHz
 OBSET 8.52 KHz
 OBFIN 1.74 Hz
 POINT 26214
 FREQU 37878.21 Hz
 SCANS 256
 ACQTM 0.6921 sec
 PD 1.2000 sec
 PW1 2.97 usec
 IRNUC 1H
 CTEMP 21.0 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 56



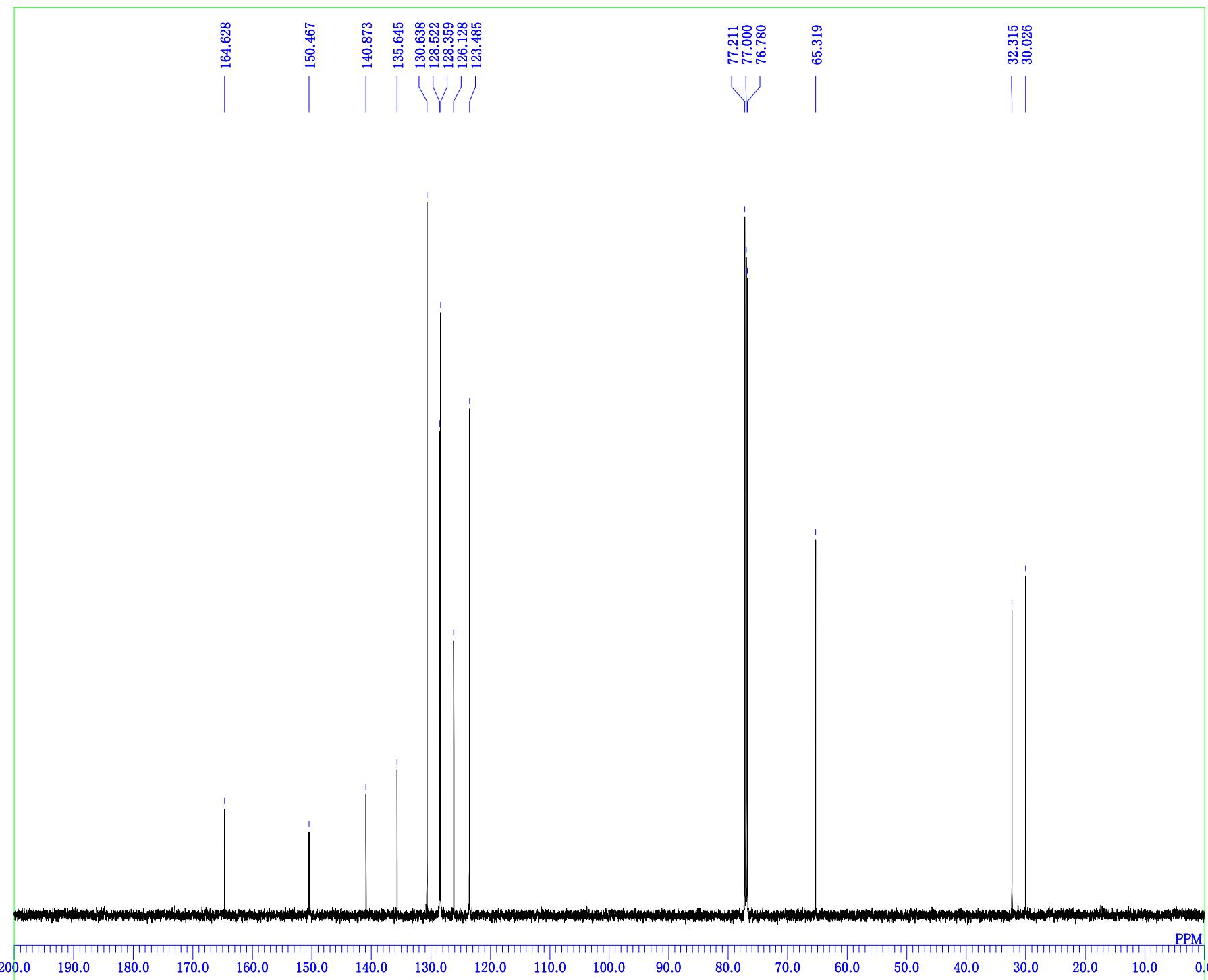
150701



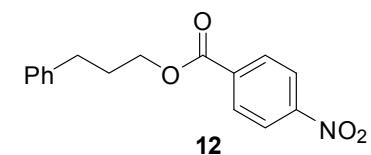
DFILE d1523-gra-1h-1.als
 COMNT 150701
 DATIM 2015-03-24 21:10:26
 1H single_pulse.ex2
 EXMOD 600.17 MHz
 OBFREQ 5.30 KHz
 OBSET 5.47 Hz
 OBFIN 5.47 Hz
 POINT 26214
 FREQU 9008.87 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 7.30 usec
 IRNUC 1H
 CTEMP 20.2 c
 SLVNT CDCL3
 EXREF 0.00 ppm
 BF 0.12 Hz
 RGAIN 40



150701

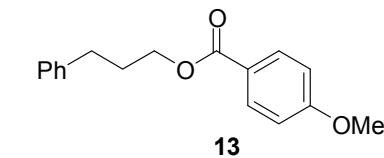
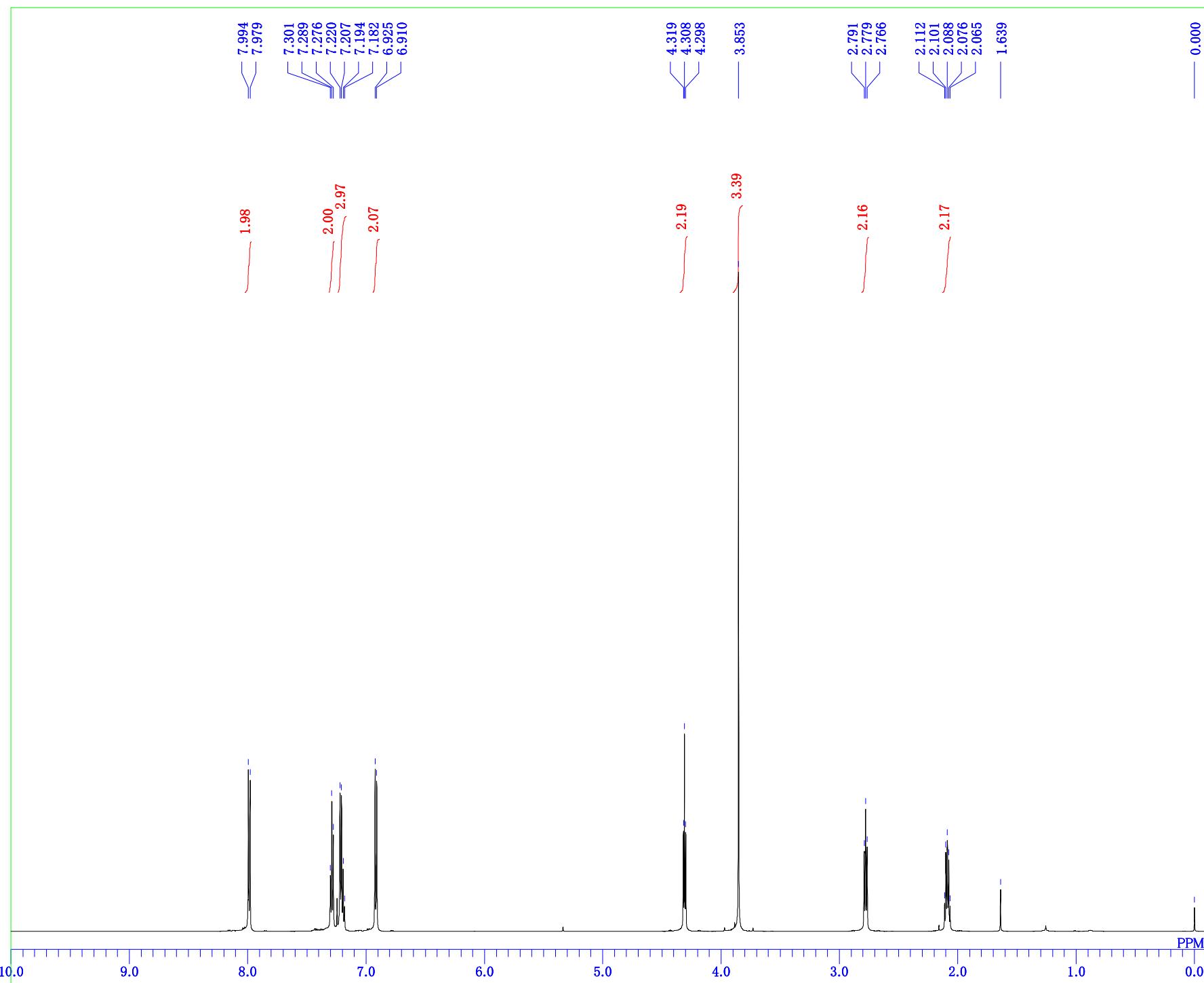


DFILE d1523-gra-13c-1.als
 COMNT 150701
 DATIM 2015-03-24 21:19:07
 13C 13C
 single_pulse_dec
 EXMOD 150.92 MHz
 OBFRQ 8.52 KHz
 OBSET 1.74 Hz
 OBFIN 26214
 POINT 37878.21 Hz
 FREQU 256
 SCANS 0.6921 sec
 ACQTM PD 1.2000 sec
 PW1 3.13 usec
 IRNUC 1H 20.6 c
 CTEMP CDCL3
 SLVNT 77.00 ppm
 EXREF 1.20 Hz
 BF 56
 RGAIN



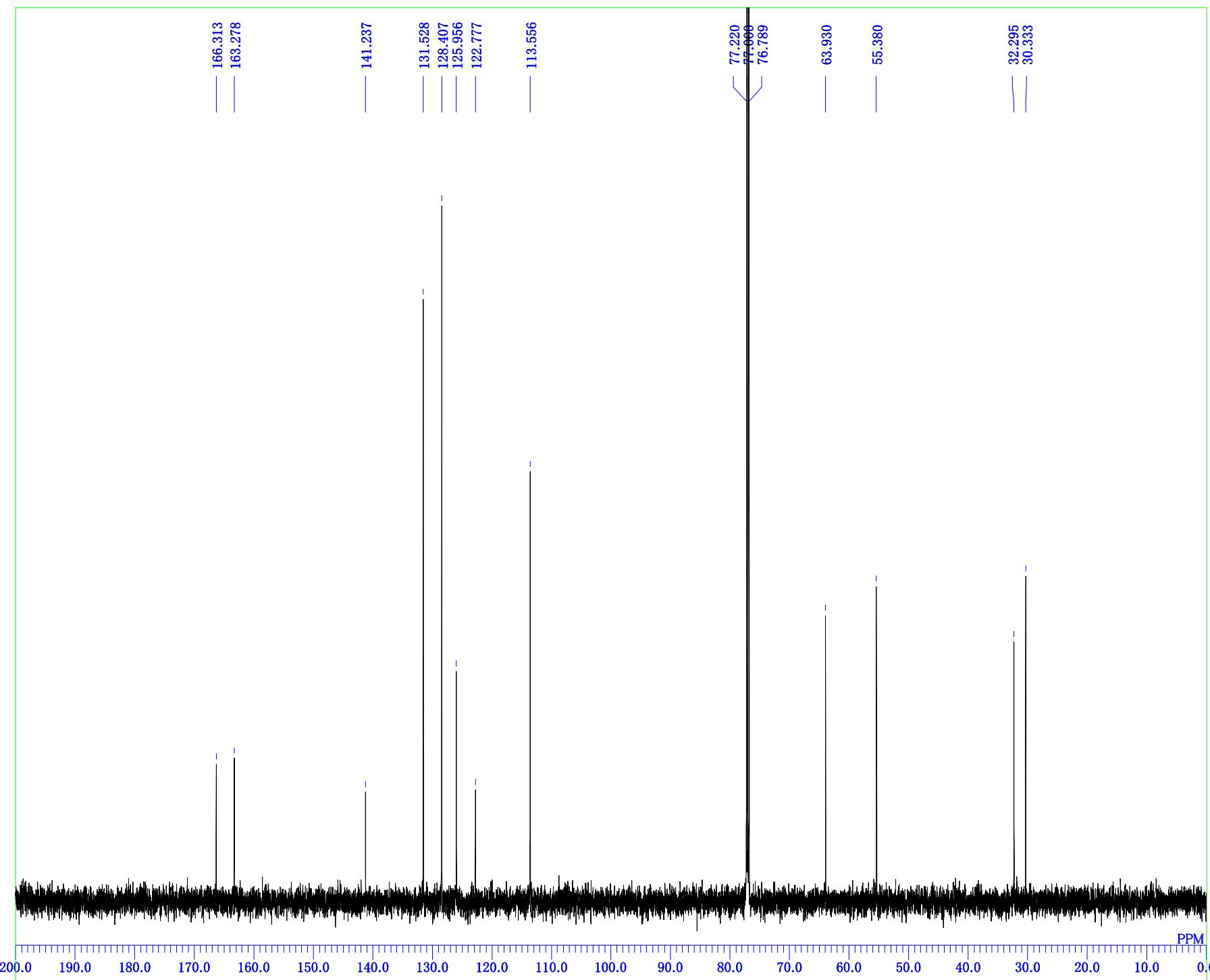
S63

150501

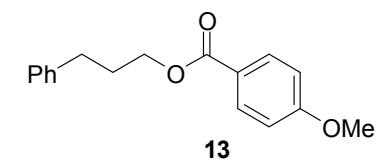


DFILE d1568-gra-1h-1.als
 COMNT 150501
 DATIM 2015-05-01 09:47:35
 1H
 single_pulse.ex2
 EXMOD 600.17 MHz
 OBFRQ 5.30 KHz
 OBSET 5.47 Hz
 OBFIN 26214
 POINT 9008.87 Hz
 FREQU 32
 SCANS 2.9098 sec
 ACQTM 2.0000 sec
 PD 7.30 usec
 PW1 1H 21.8 c
 IRNUC CDCL3
 CTEMP 0.00 ppm
 SLVNT 0.12 Hz
 EXREF 32
 BF RGAIN

150501



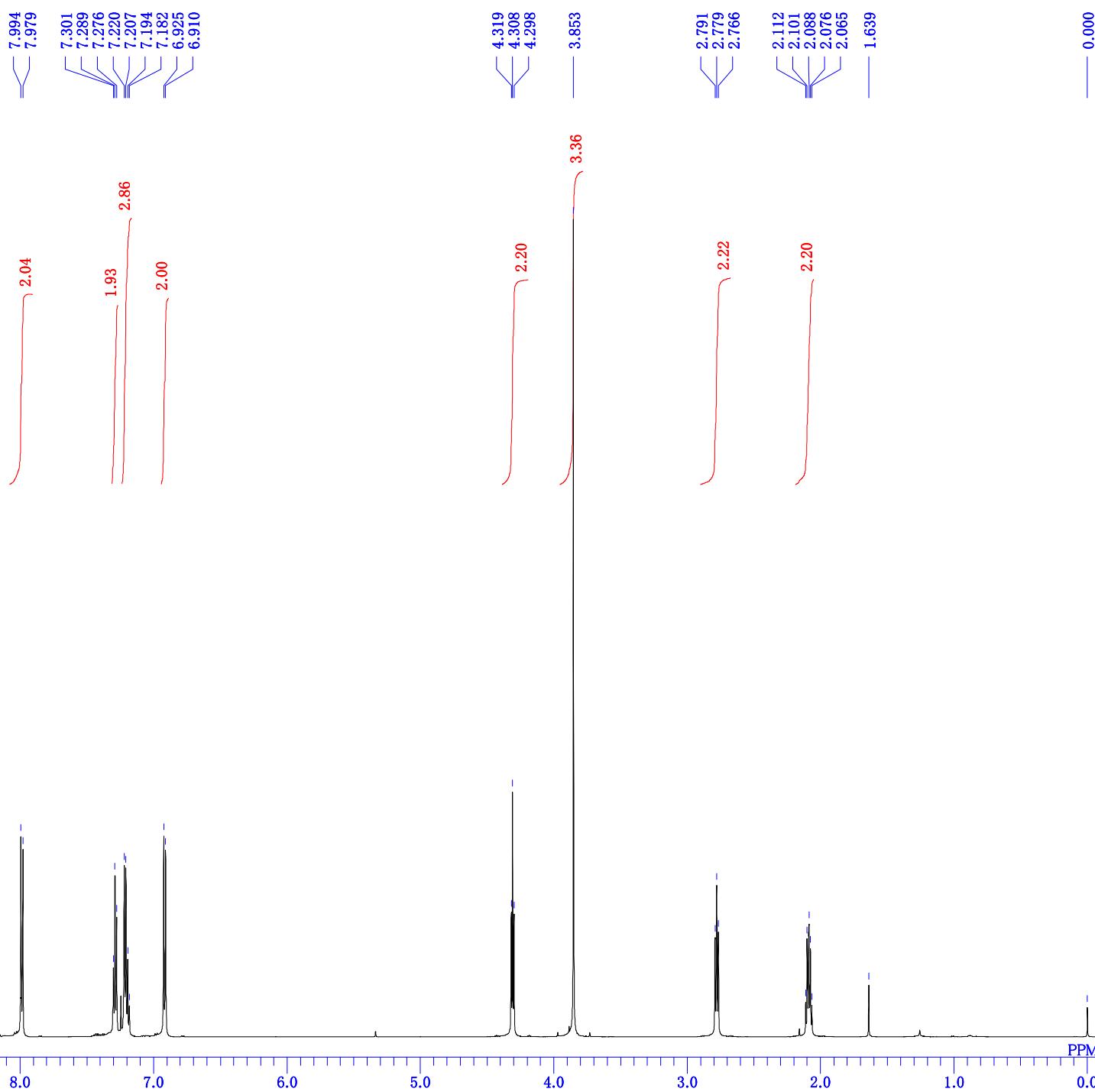
DFILE d1568-gra-13c-1.als
 COMNT 150501
 DATIM 2015-05-01 09:56:06
 13C 13C
 single_pulse_dec
 EXMOD 150.92 MHz
 OBFRQ 8.52 KHz
 OBSET 1.74 Hz
 OBFIN 26214
 POINT 37878.21 Hz
 FREQU 256
 SCANS 0.6921 sec
 ACQTM PD 1.2000 sec
 PW1 PW1 3.13 usec
 IRNUC 1H 22.4 c
 CTEMP CDCL3 77.00 ppm
 SLVNT 1.20 Hz
 EXREF 60
 BF RGAIN



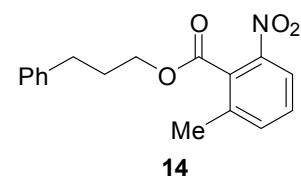
PPM

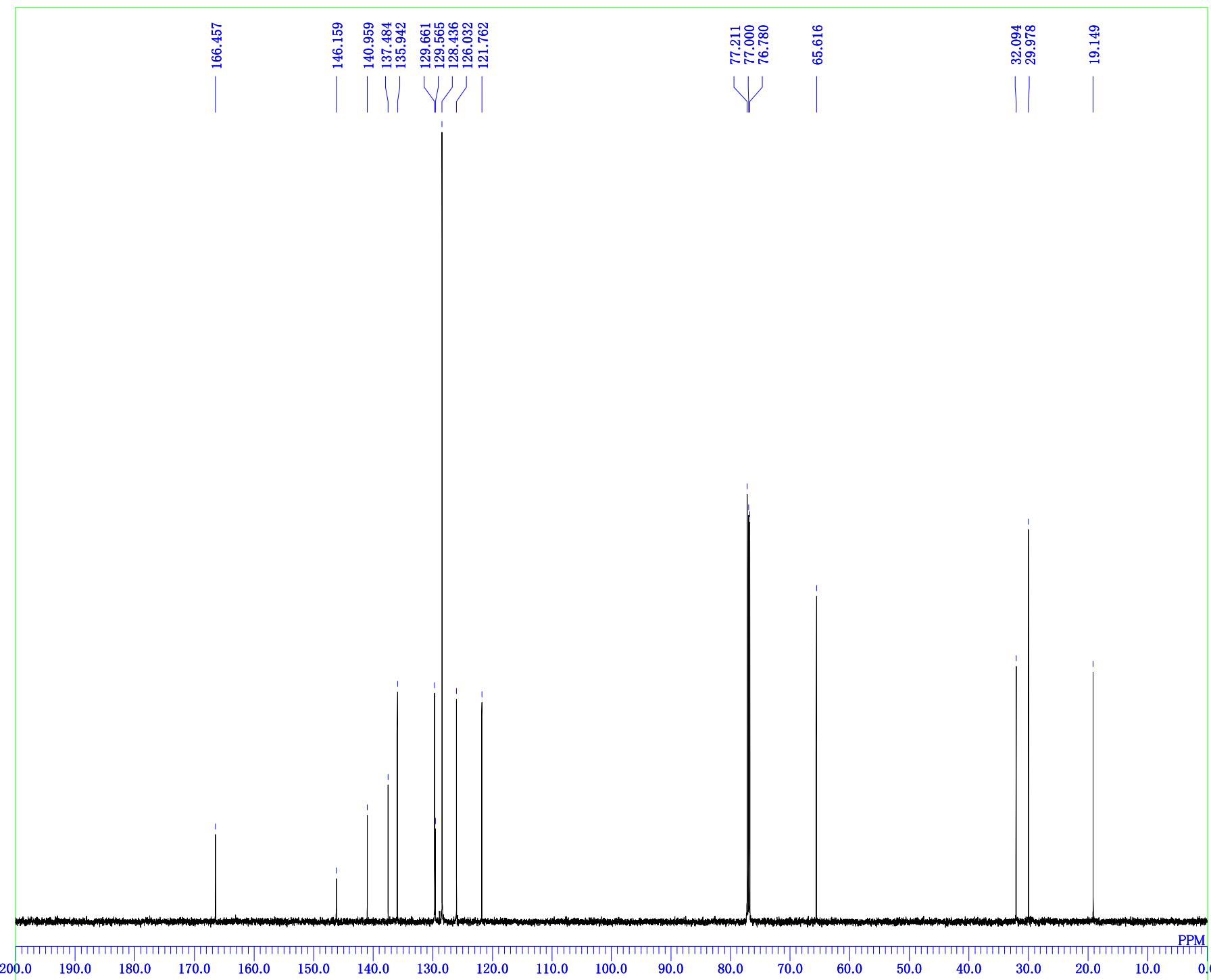
S65

150701

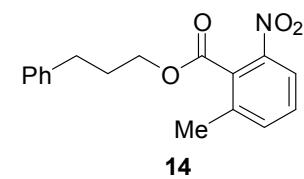


DFILE d1568-gra-1h-1.als
 COMNT 150701
 DATIM 2015-05-01 09:47:35
 1H single_pulse.ex2
 EXMOD 600.17 MHz
 OBFRQ 5.30 KHz
 OBSET 5.47 Hz
 OBFIN 26214
 POINT 26214
 FREQU 9008.87 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 7.30 usec
 IRNUC 1H
 CTEMP 21.8 c
 SLVNT CDCL₃
 EXREF 0.00 ppm
 BF 0.12 Hz
 RGAIN 32

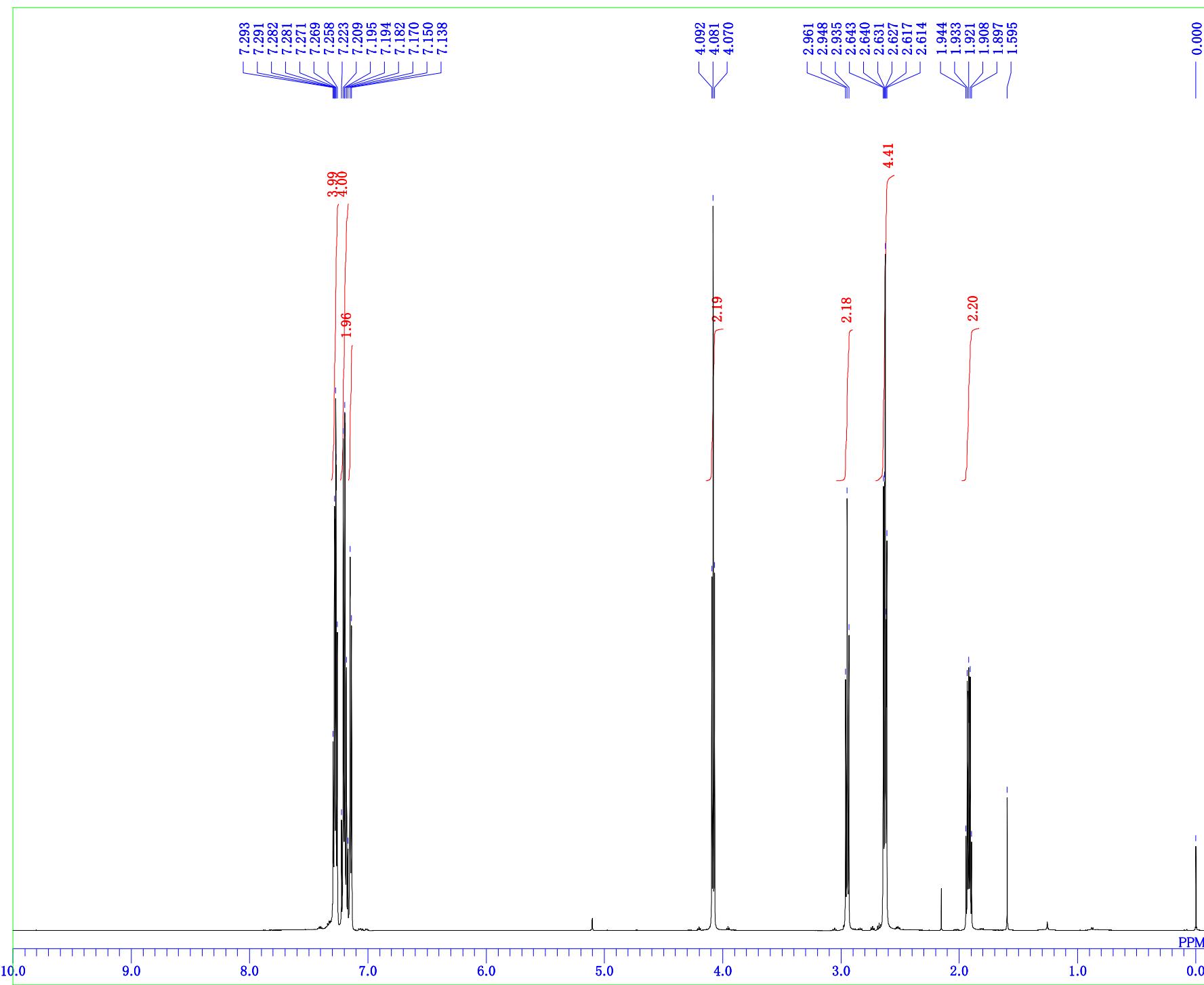




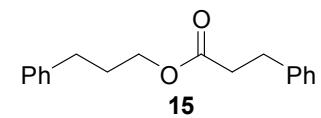
DFILE d1575-gra-13c-1.als
 COMNT 150701
 DATIM 2015-05-19 18:19:49
 13C 13C
 EXMOD single_pulse_dec
 OBFRQ 150.92 MHz
 OBSET 8.52 KHz
 OBFIN 1.74 Hz
 POINT 26214
 FREQU 37878.21 Hz
 SCANS 256
 ACQTM 0.6921 sec
 PD 1.2000 sec
 PW1 3.13 usec
 IRNUC 1H
 CTEMP 21.9 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 56



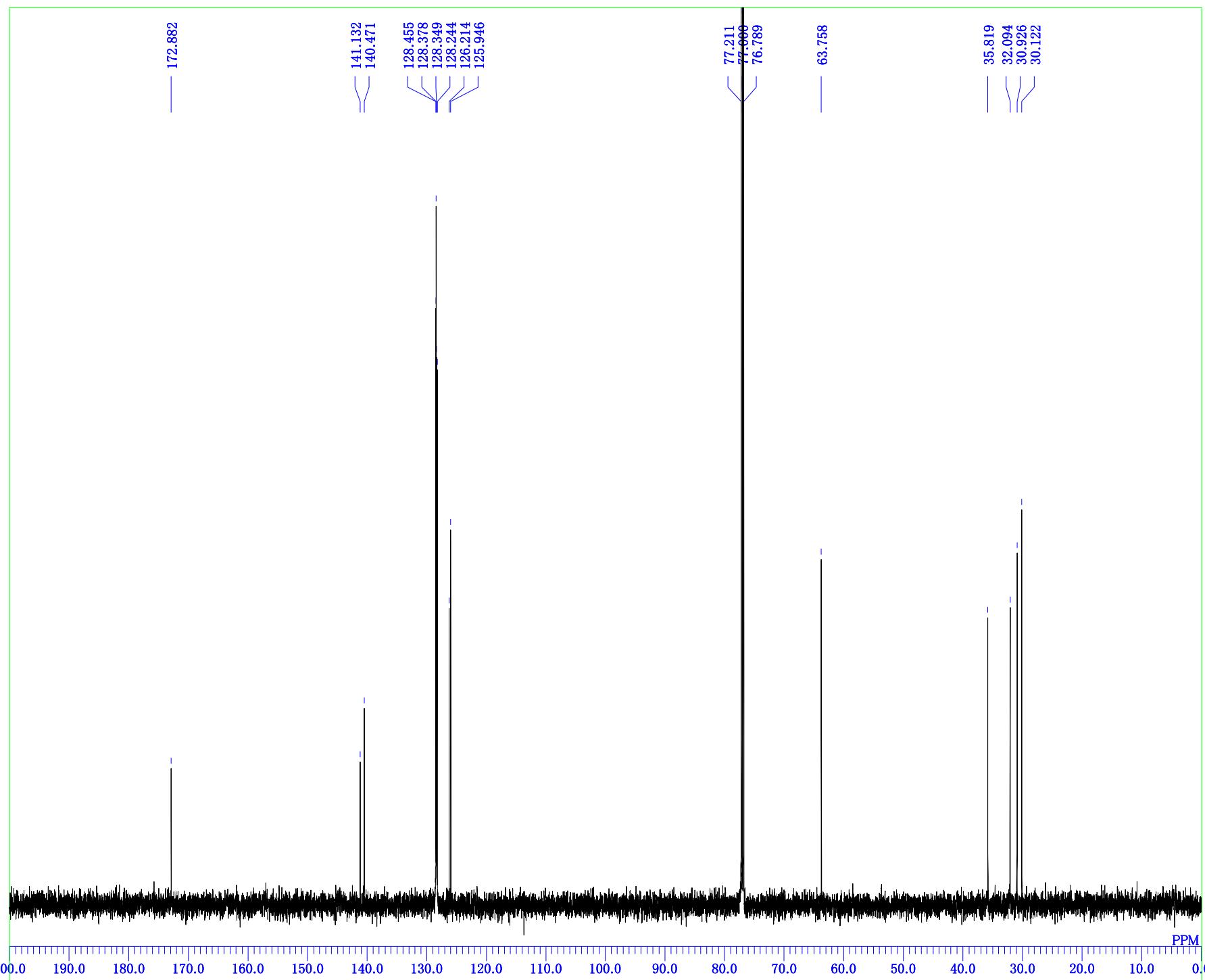
150701



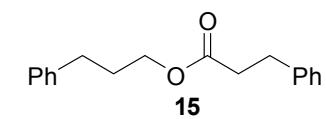
DFILE d1569-gra-1h-1.als
 COMNT 150701
 DATIM 2015-05-01 10:03:52
 1H single_pulse.ex2
 EXMOD 600.17 MHz
 OBFRQ 5.30 KHz
 OBSET 5.47 Hz
 OBFIN 26214
 POINT 26214
 FREQU 9008.87 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 7.30 usec
 IRNUC 1H
 CTEMP 21.6 c
 SLVNT CDCL3
 EXREF 0.00 ppm
 BF 0.12 Hz
 RGAIN 30



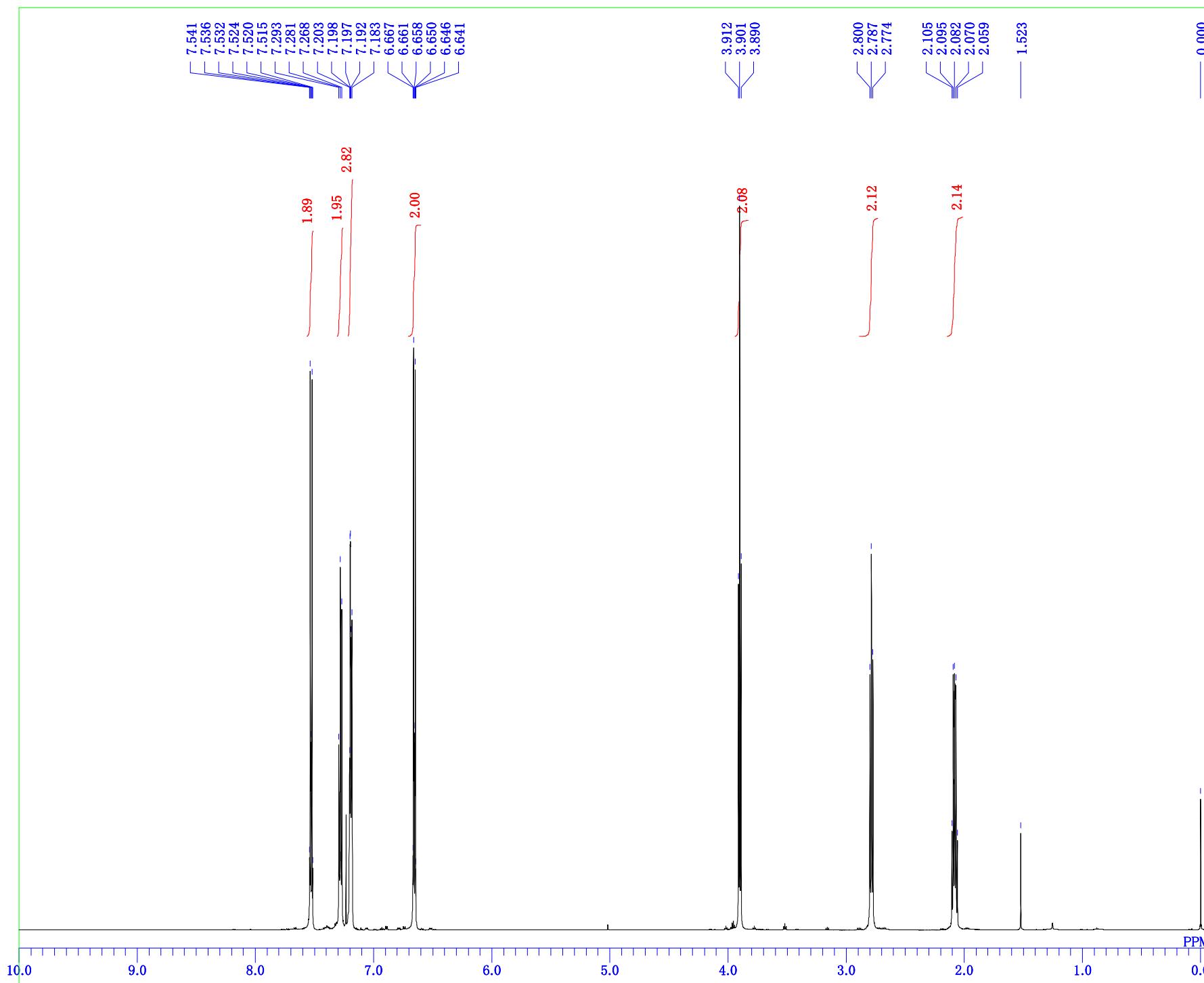
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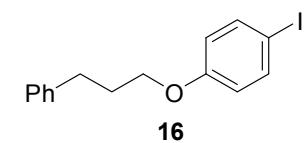
DFILE d1569-gra-13c-1.als
 COMNT 150701
 DATIM 2015-05-01 10:12:25
¹³C 13C
 EXMOD single_pulse_dec
 OBFRQ 150.92 MHz
 OBSET 8.52 KHz
 OBFIN 1.74 Hz
 POINT 26214
 FREQU 37878.21 Hz
 SCANS 256
 ACQTM 0.6921 sec
 PD 1.2000 sec
 PW1 3.13 usec
 IRNUC 1H
 CTEMP 22.6 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 60



150701



DFILE d1529-gra-1h-1.als
 COMNT 150701
 DATIM 2015-04-21 16:22:13
 1H single_pulse.ex2
 EXMOD 600.17 MHz
 OBFRQ 5.30 KHz
 OBSET 5.47 Hz
 OBFIN 26214
 POINT 9008.87 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 7.30 usec
 IRNUC 1H
 CTEMP 21.1 c
 SLVNT CDCL₃
 EXREF 0.00 ppm
 BF 0.12 Hz
 RGAIN 34



S70

158.845

141.275
138.135128.474
128.416
125.965

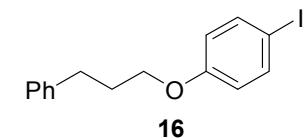
116.898

82.544
77.211
77.000
76.789

66.889

32.027
30.639

DFILE d1529-gra-13c-1.als
 COMNT 150701
 DATIM 2015-04-21 16:33:50
 13C 13C
 EXMOD single_pulse_dec
 OBFRQ 150.92 MHz
 OBSET 8.52 KHz
 OBFIN 1.74 Hz
 POINT 26214
 FREQU 37878.21 Hz
 SCANS 256
 ACQTM 0.6921 sec
 PD 1.2000 sec
 PW1 3.13 usec
 IRNUC 1H
 CTEMP 21.7 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 54

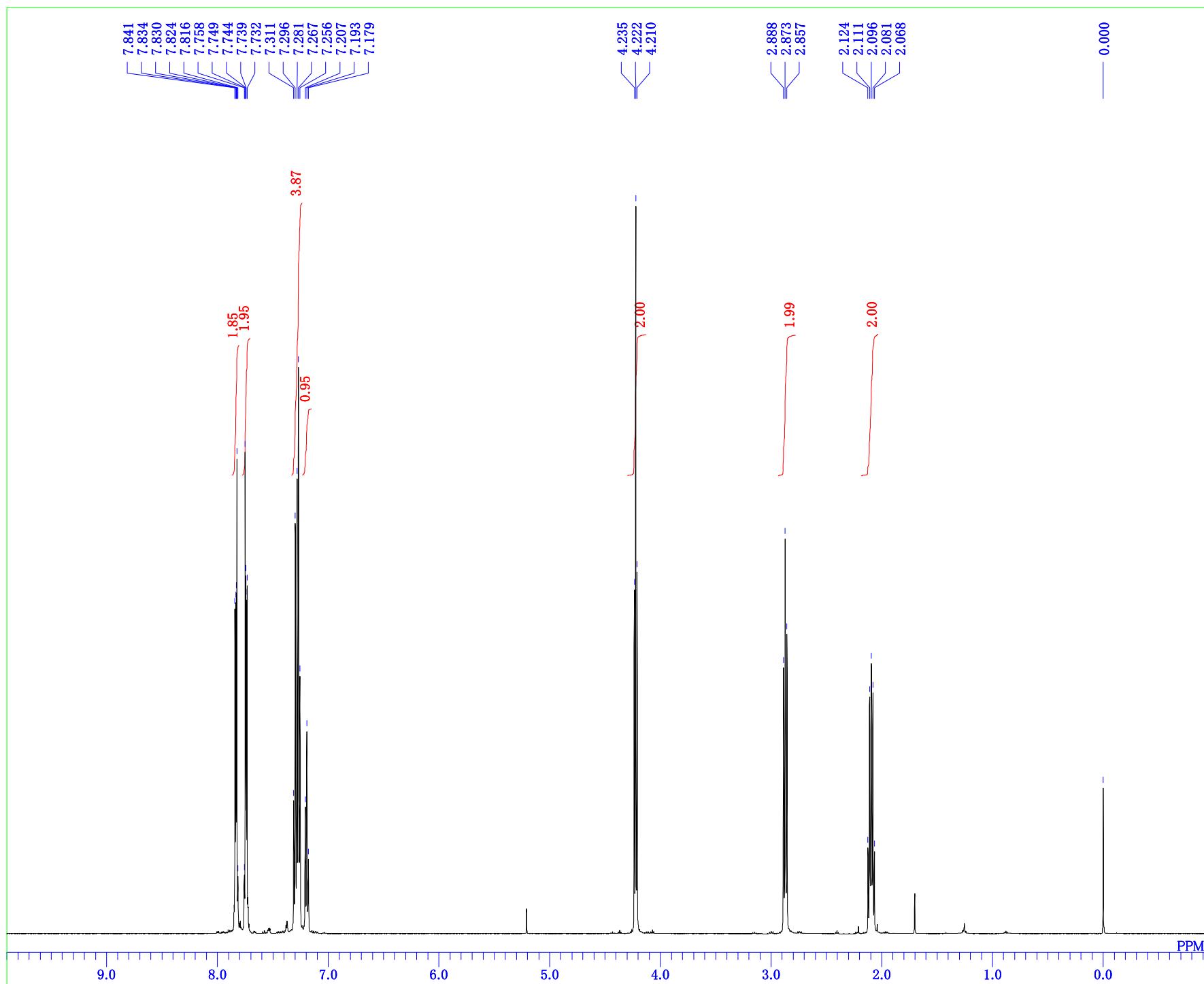


PPM

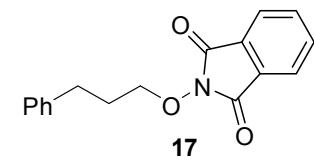
200.0 190.0 180.0 170.0 160.0 150.0 140.0 130.0 120.0 110.0 100.0 90.0 80.0 70.0 60.0 50.0 40.0 30.0 20.0 10.0 0.0

S71

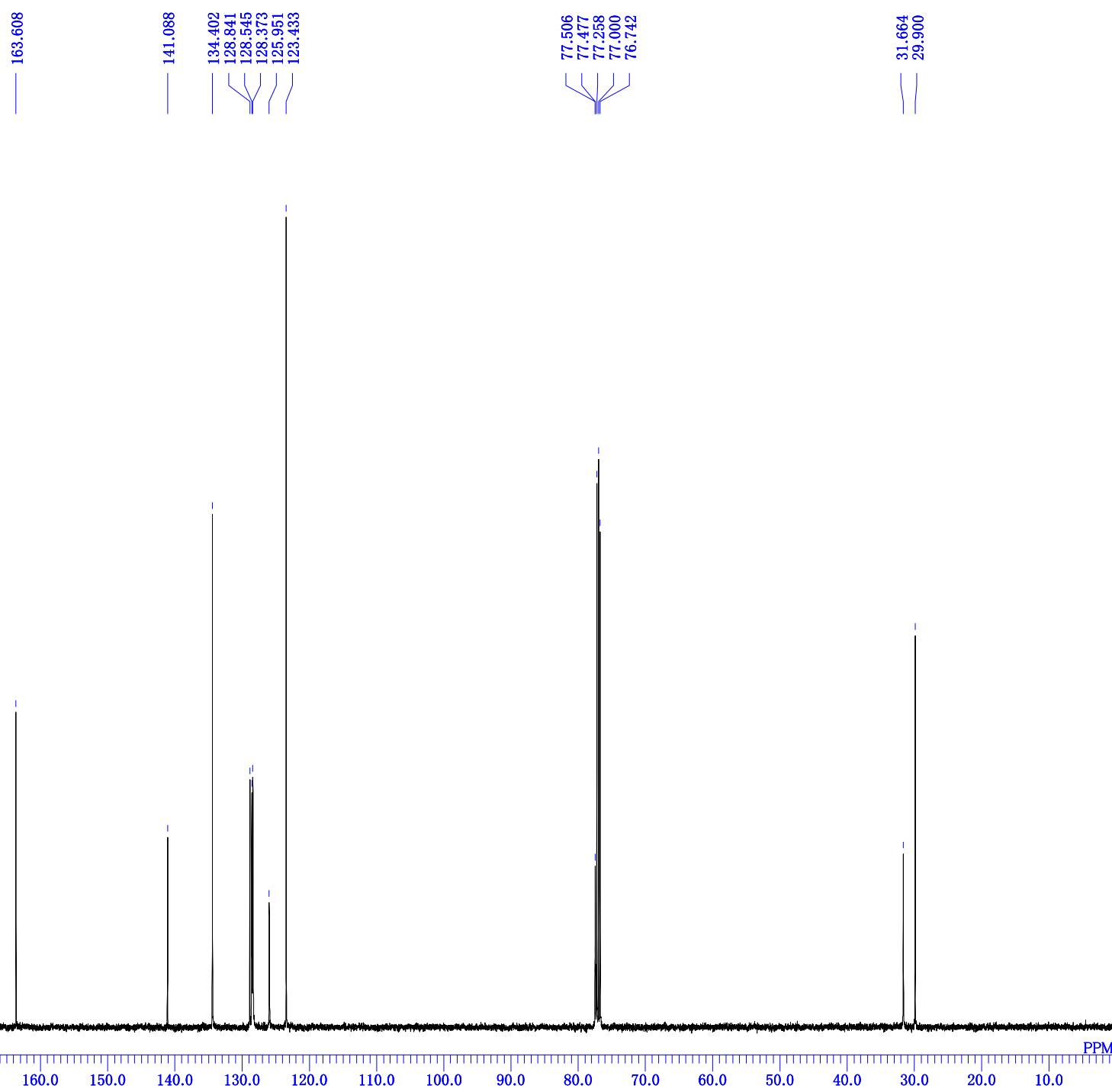
151113



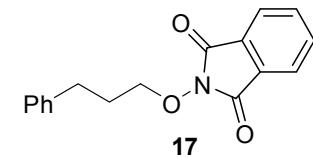
DFILE d1769-gra-1h-1.jdf
 COMNT 151113
 DATIM 2015-11-13 13:00:17
 1H single_pulse.ex2
 EXMOD 500.16 MHz
 OBFRQ 2.41 KHz
 OBSET 6.01 Hz
 OBFIN 16384
 POINT 9384.38 Hz
 FREQU 32
 SCANS 1.7459 sec
 ACQTM 2.0000 sec
 PD 5.80 usec
 PW1 1H 18.6 c
 IRNUC CDCL3
 CTEMP 0.00 ppm
 SLVNT 0.12 Hz
 EXREF 40
 RGAIN



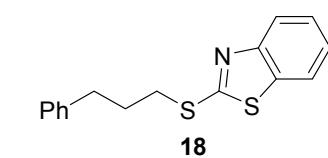
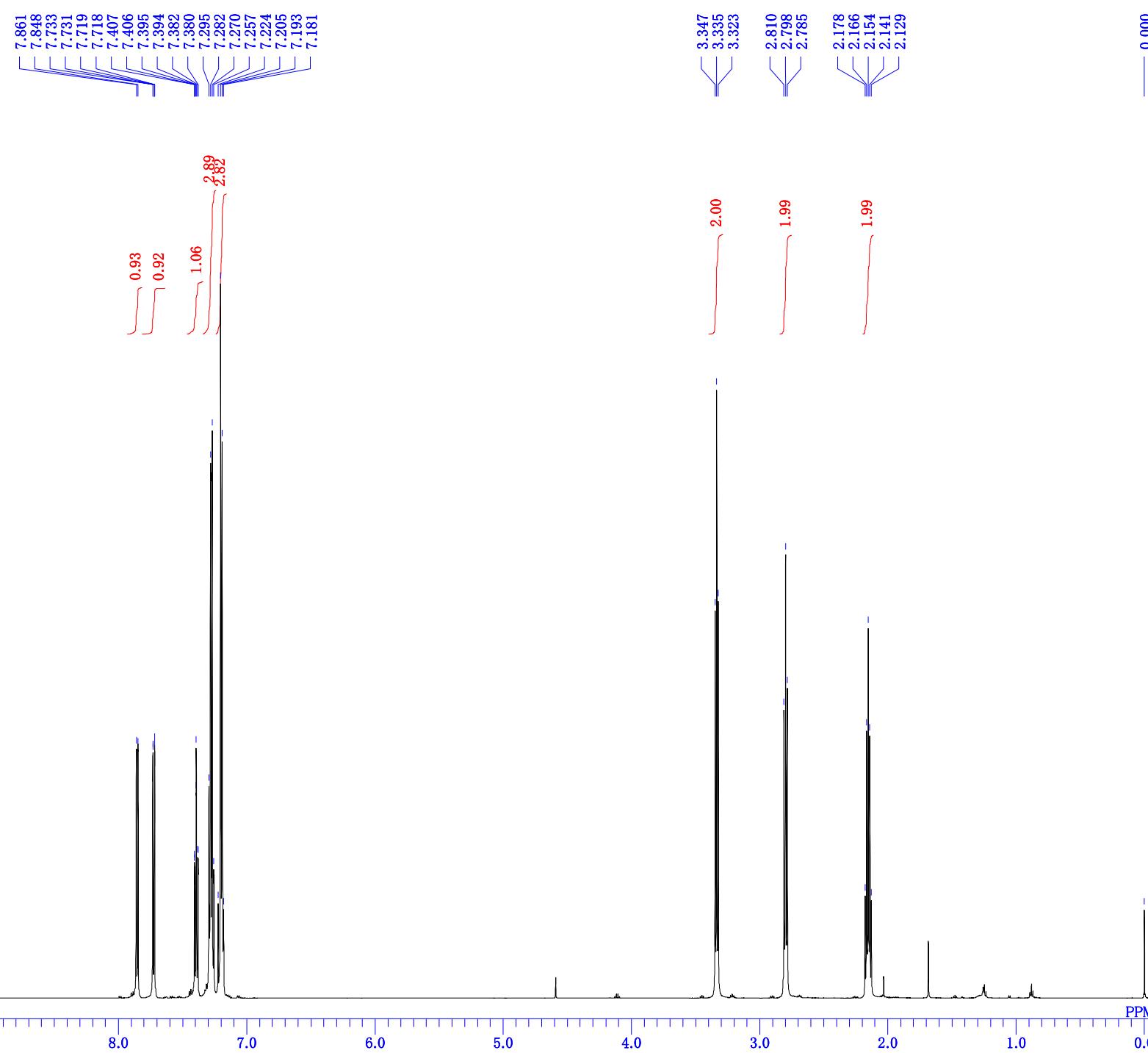
151113



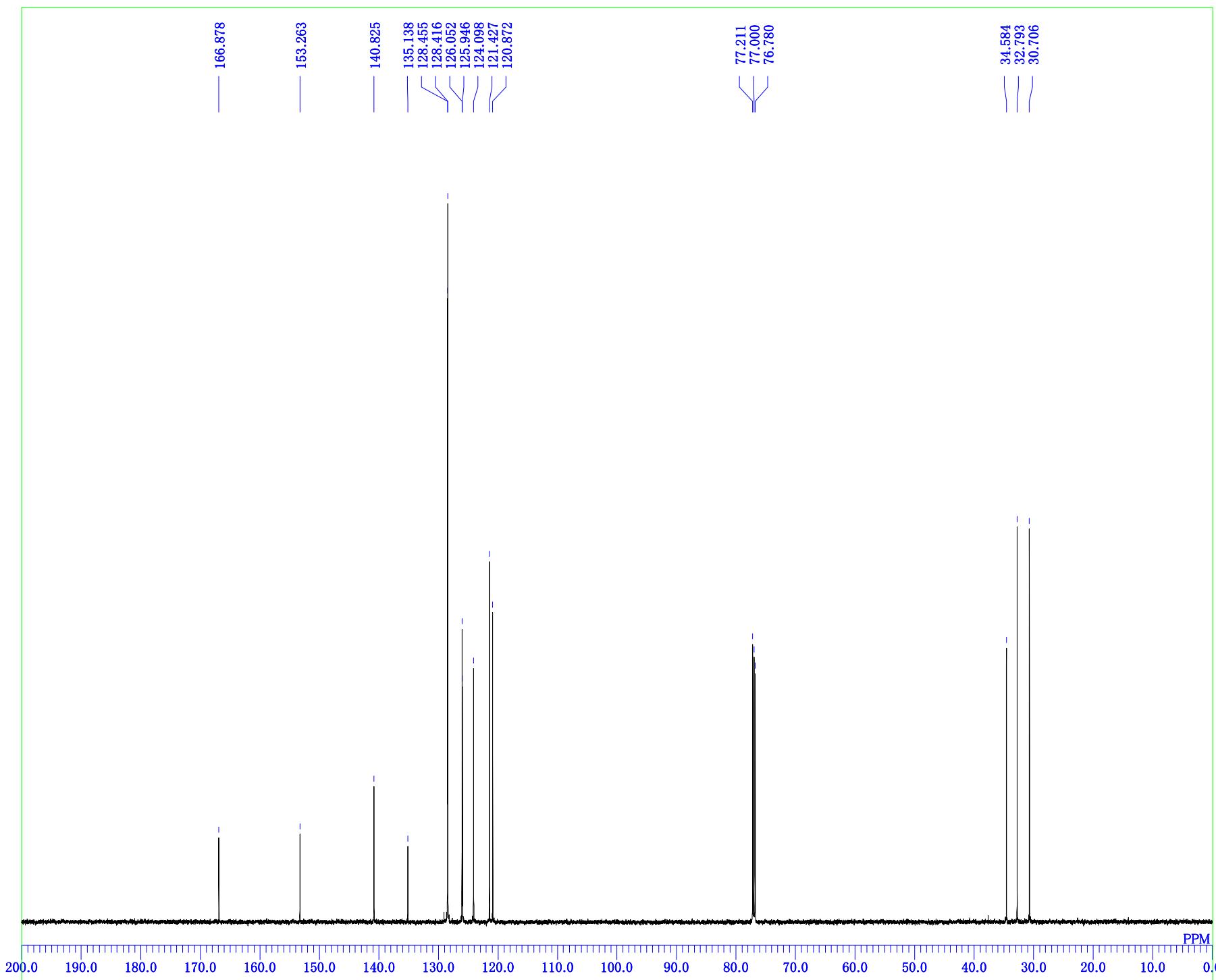
DFILE d1769-gra-13c-1.als
 COMNT 151113
 DATIM 2015-11-13 13:13:10
 13C single_pulse_dec
 EXMOD 125.77 MHz
 OBFRQ 7.87 KHz
 OBSET 4.21 Hz
 OBFIN 26214
 POINT 31446.06 Hz
 FREQU 256
 SCANS 0.8336 sec
 ACQTM PD 2.0000 sec
 PW1 PW1 3.00 usec
 IRNUC 1H 18.8 c
 CTEMP CDCL3 77.00 ppm
 SLVNT 1.20 Hz
 EXREF 54
 BF RGAIN



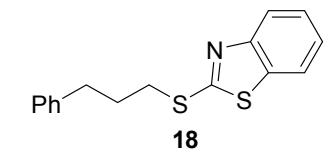
150406

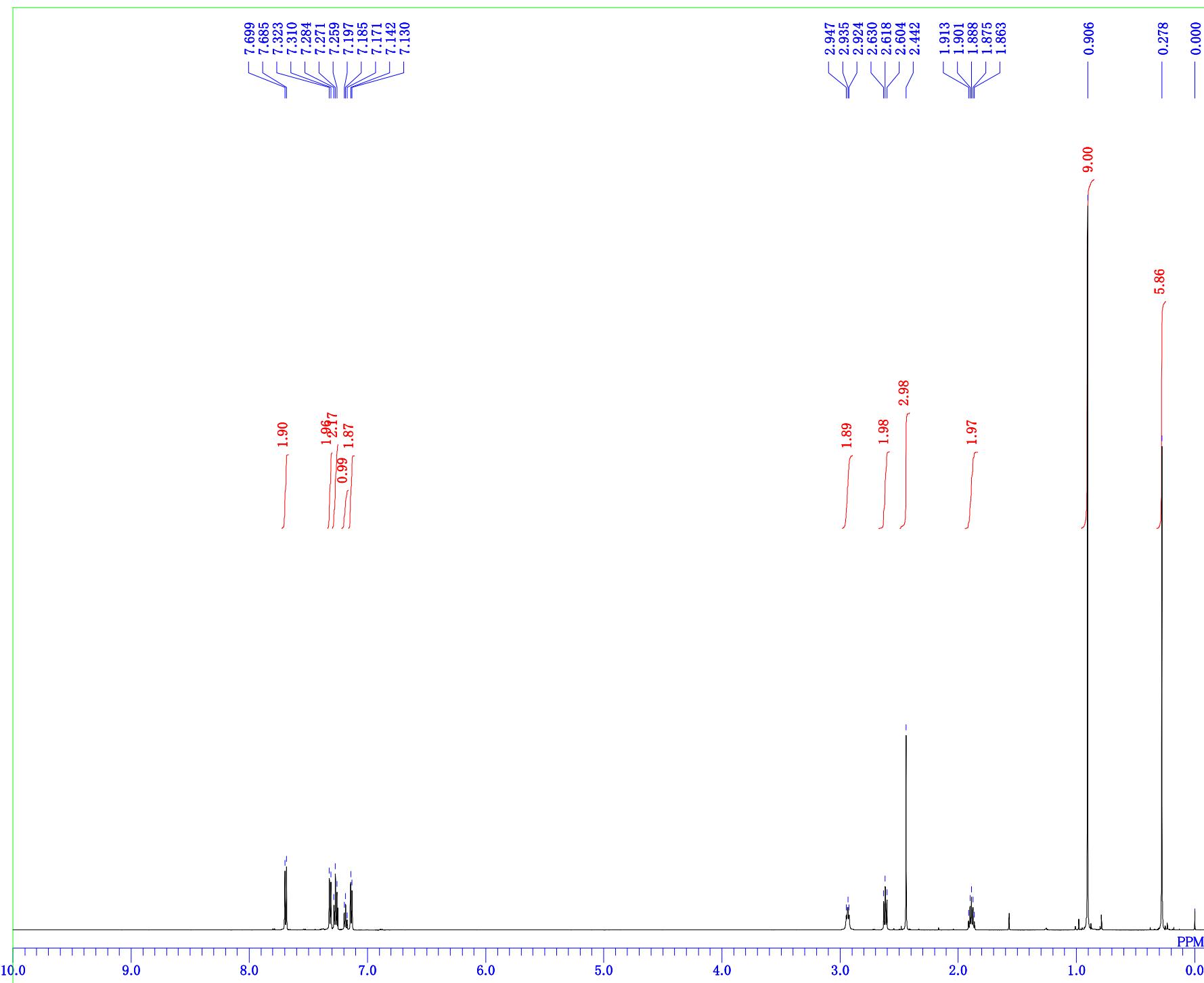


DFILE d1547-gra-1h-1.als
 COMNT 150406
 DATIM 2015-04-06 15:29:56
 1H
 single_pulse.ex2
 EXMOD 600.17 MHz
 OBFRQ 5.30 KHz
 OBSET 5.47 Hz
 OBFIN 26214
 POINT 9008.87 Hz
 FREQU 32
 SCANS 2.9098 sec
 ACQTM PD
 PW1 2.0000 sec
 IRNUC 7.30 usec
 1H 20.9 c
 CTEMP CDCL3
 SLVNT 0.00 ppm
 EXREF 0.12 Hz
 RGAIN 30

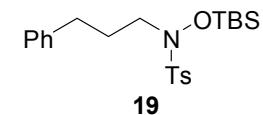


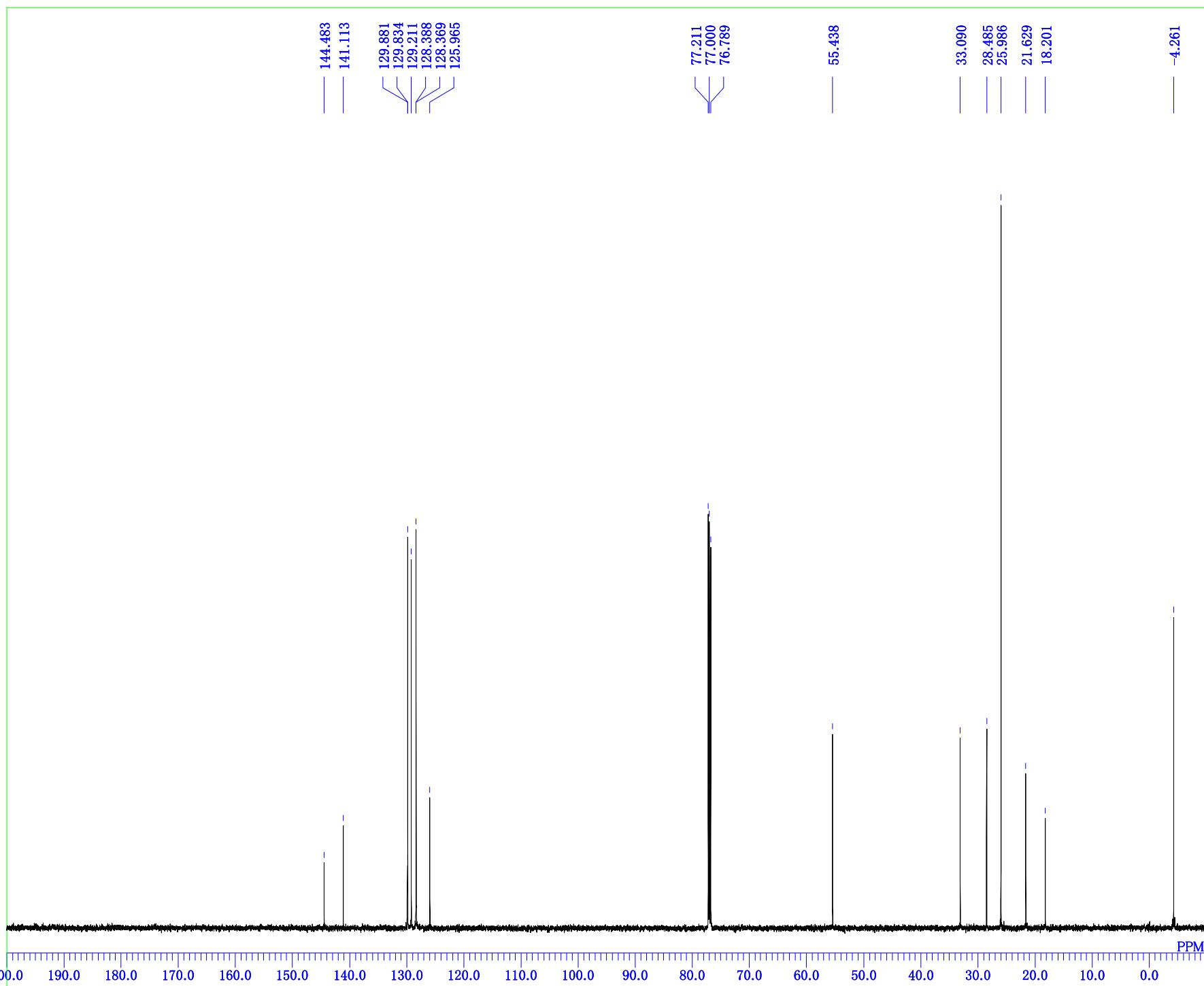
DFILE d1547-gra-13c-1.als
 COMNT 150406
 DATIM 2015-04-06 15:39:21
 13C
 single_pulse_dec
 150.92 MHz
 OBFRQ 8.52 KHz
 OBSET 1.74 Hz
 OBFIN 26214
 POINT 37878.21 Hz
 FREQU 256
 SCANS 0.6921 sec
 ACQTM 1.2000 sec
 PD 3.13 usec
 PW1
 IRNUC 1H
 CTEMP 21.7 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 56



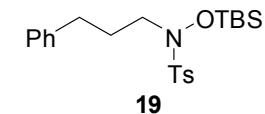


otbsnhts-1h-1.als
150704
2015-02-23 16:57:20
1H
single_pulse.ex2
600.17 MHz
5.30 KHz
5.47 Hz
26214
9008.87 Hz
16
2.9098 sec
ACQTM
PD
PW1
IRNUC
CTEMP
SLVNT
EXREF
BF
RGAIN
1H
20.4 c
CDCL3
0.00 ppm
0.12 Hz
30

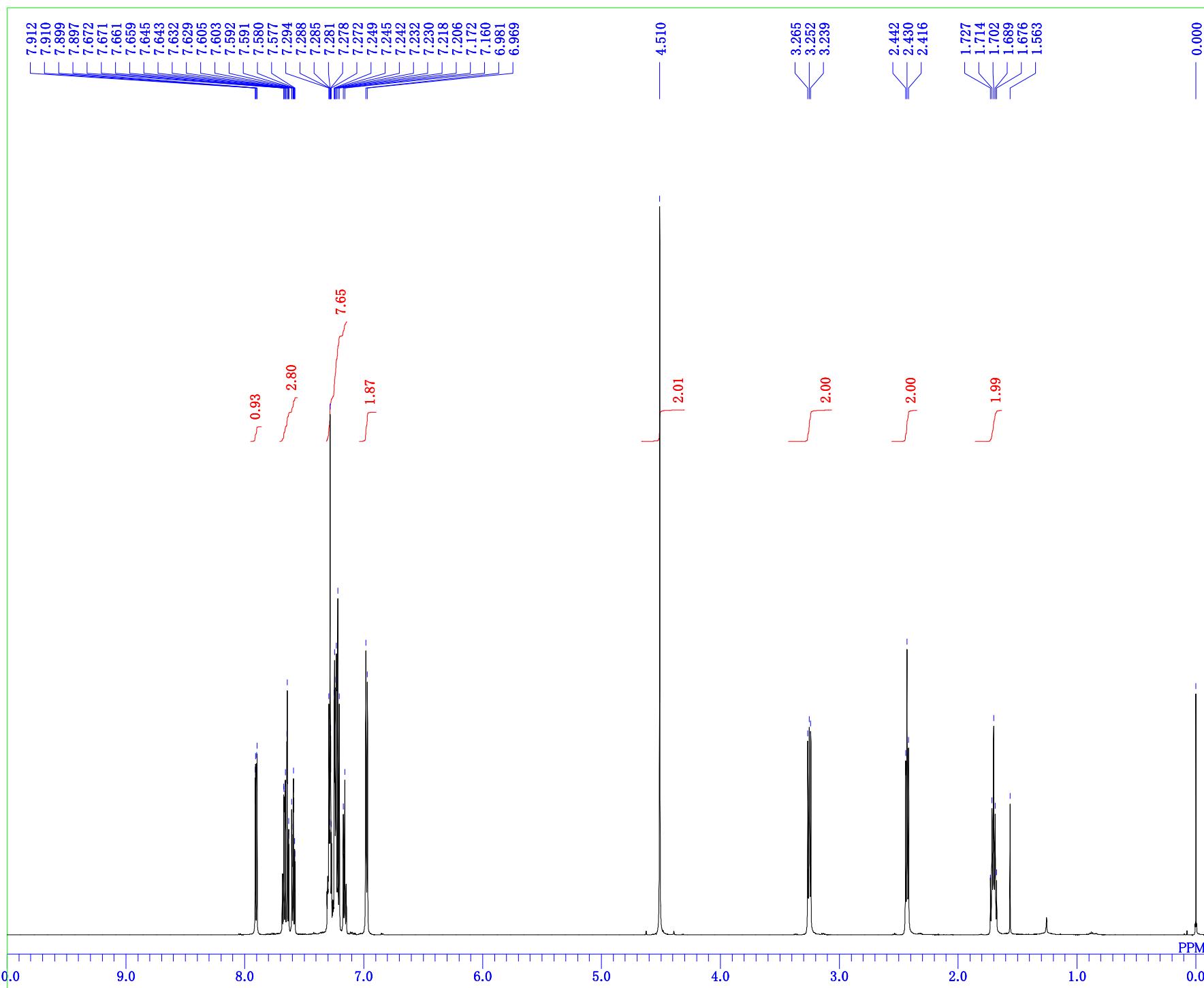




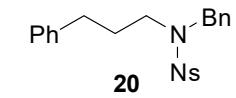
otbsnhts-13C-1.als
150704
2015-02-23 17:09:09
13C
single_pulse_dec
150.92 MHz
8.52 KHz
1.74 Hz
26214
37878.21 Hz
256
0.6921 sec
1.2000 sec
2.97 usec
1H
21.2 c
CDCL3
77.00 ppm
1.20 Hz
56

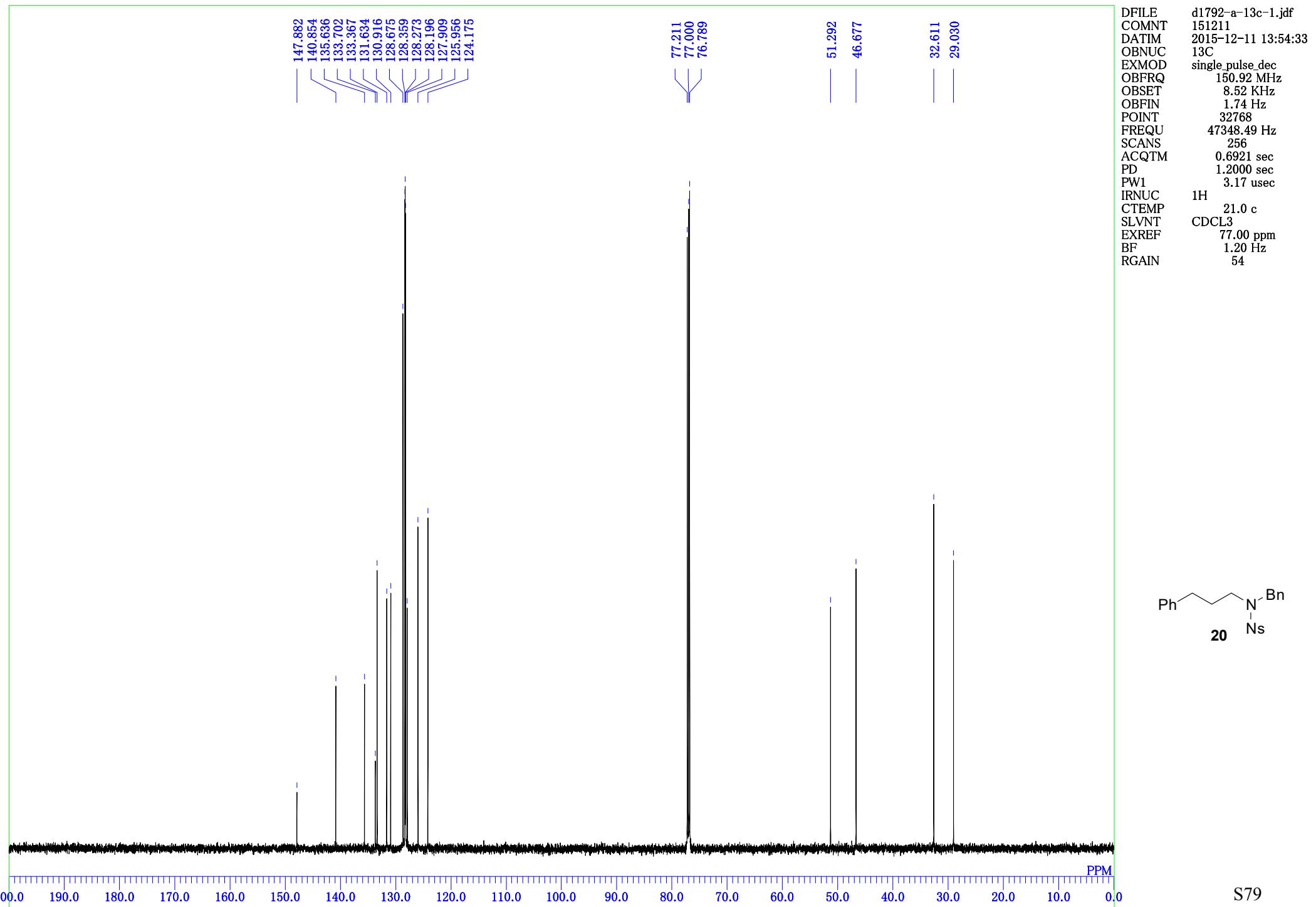


151211

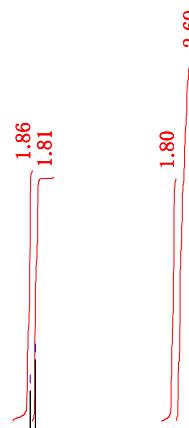
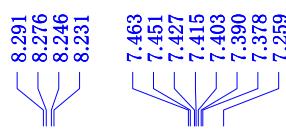


d1792-a-1h-1.jdf
151211
2015-12-11 13:43:54
1H
single_pulse.ex2
600.17 MHz
5.30 KHz
5.47 Hz
32768
11261.26 Hz
32
2.9098 sec
2.0000 sec
7.30 usec
1H
20.3 c
CDCL3
0.00 ppm
0.12 Hz
36





150704



5.408

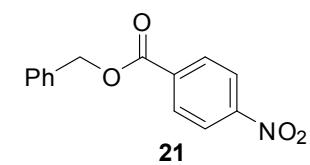
2.00

1.566

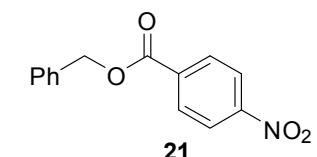
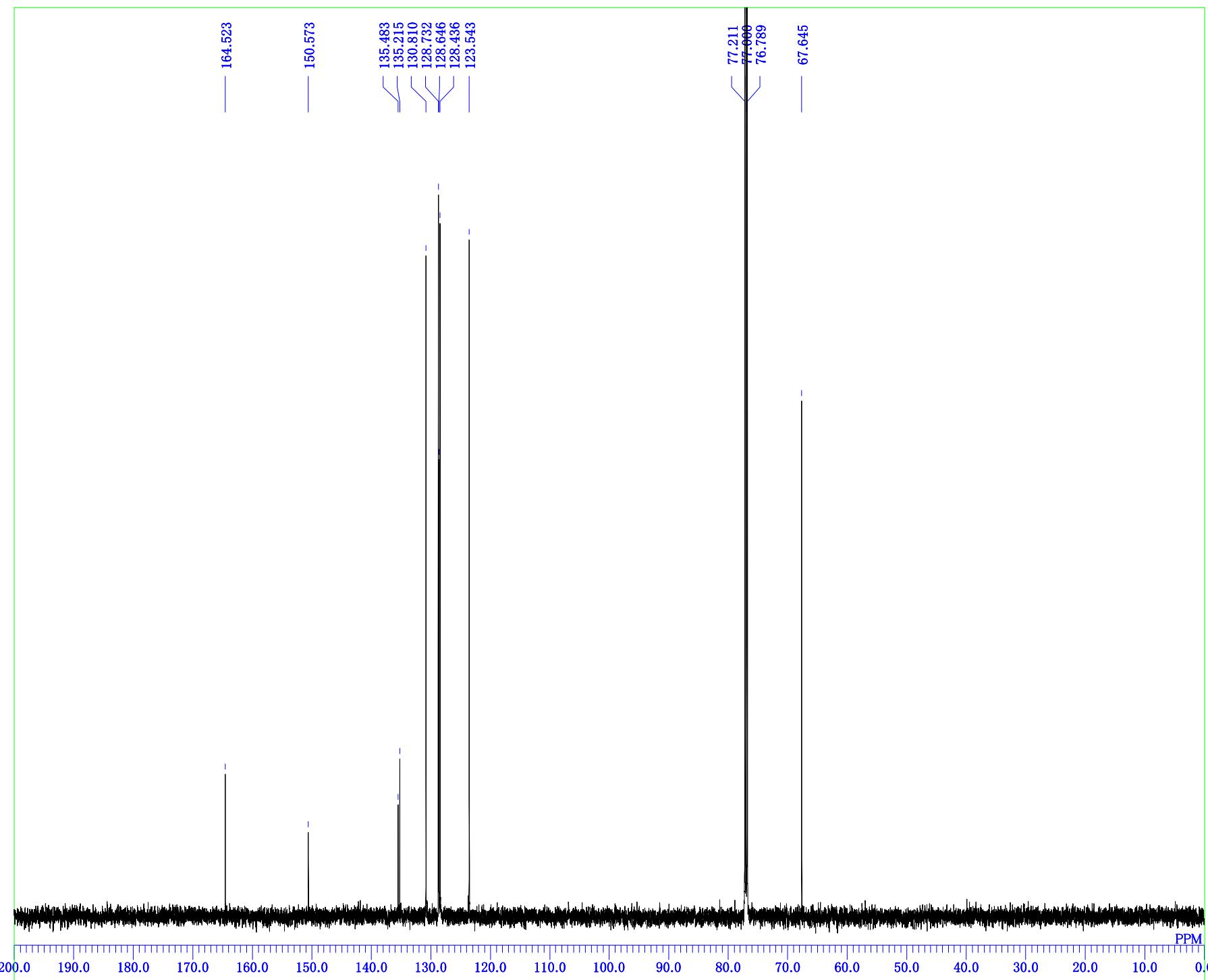
0.000

10.0 9.0 8.0 7.0 6.0 5.0 4.0 3.0 2.0 1.0 0.0 PPM

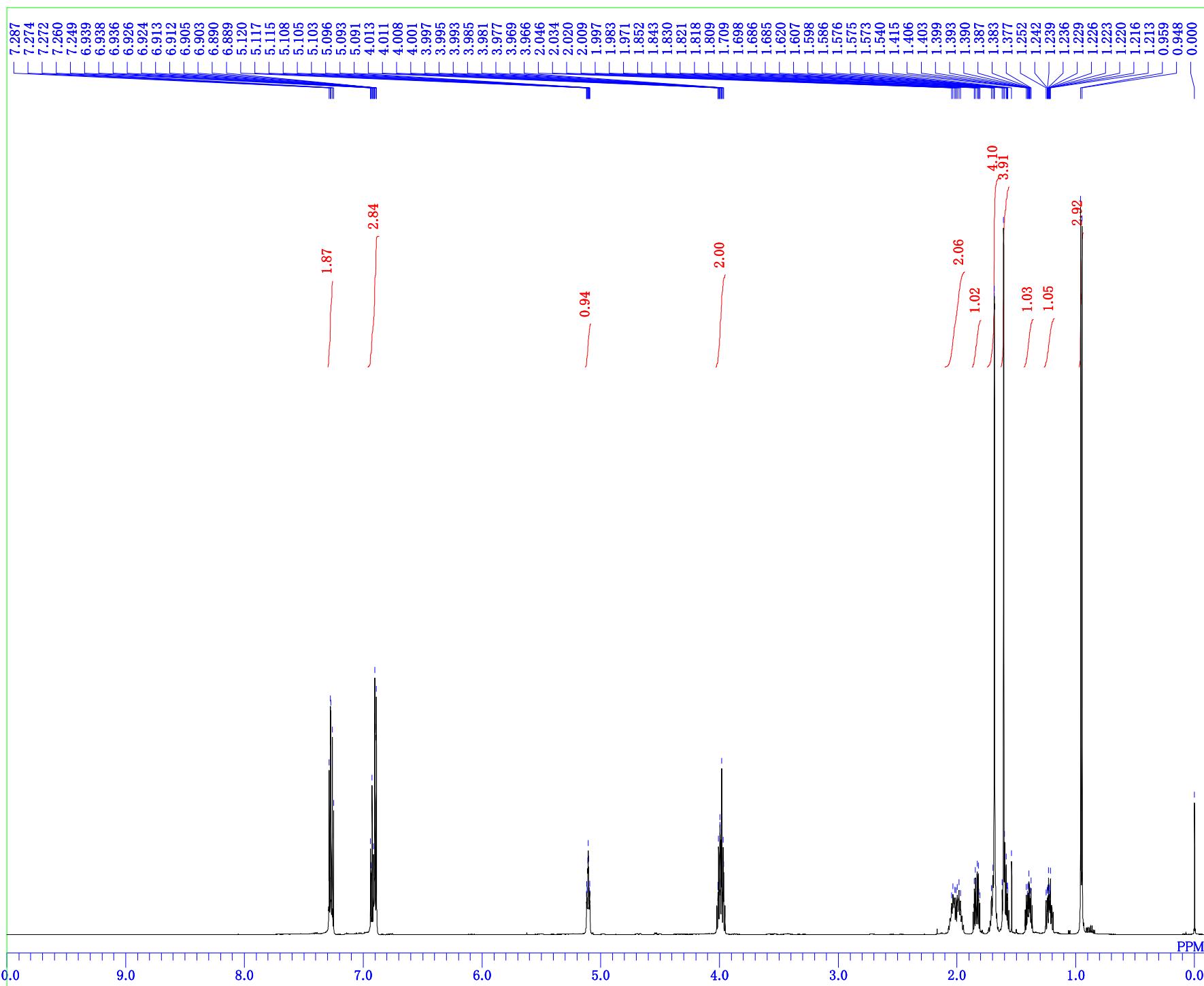
S80



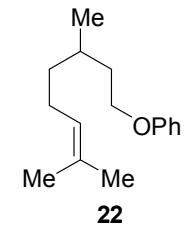
DFILE d1542-gra-1h-1.als
 COMNT 150704
 DATIM 2015-04-02 14:45:45
 1H
 single_pulse.ex2
 EXMOD 600.17 MHz
 OBFRQ 5.30 KHz
 OBSET 5.47 Hz
 OBFIN 5.47 Hz
 POINT 26214
 FREQU 9008.87 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 7.30 usec
 1H 20.9 c
 IRNUC CDCL3
 CTEMP 0.00 ppm
 SLVNT 0.12 Hz
 EXREF 44
 RGAIN



150704



DFILE d1535-gra-1h-1.als
 COMNT 150704
 DATIM 2015-03-24 21:33:37
 1H
 single_pulse.ex2
 EXMOD 600.17 MHz
 OBFRQ 5.30 KHz
 OBSET 5.47 Hz
 OBFIN 26214
 POINT 26214
 FREQU 9008.87 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 7.30 usec
 1H 20.2 c
 CDCL3 0.00 ppm
 IRNUC 0.12 Hz
 CTEMP 36
 SLVNT
 EXREF
 BF
 RGAIN



159.084
 131.289
 129.384
 124.673
 120.431
 114.476

77.211
 77.000
 76.789

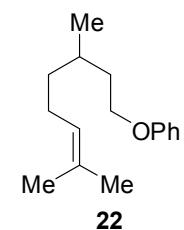
66.104

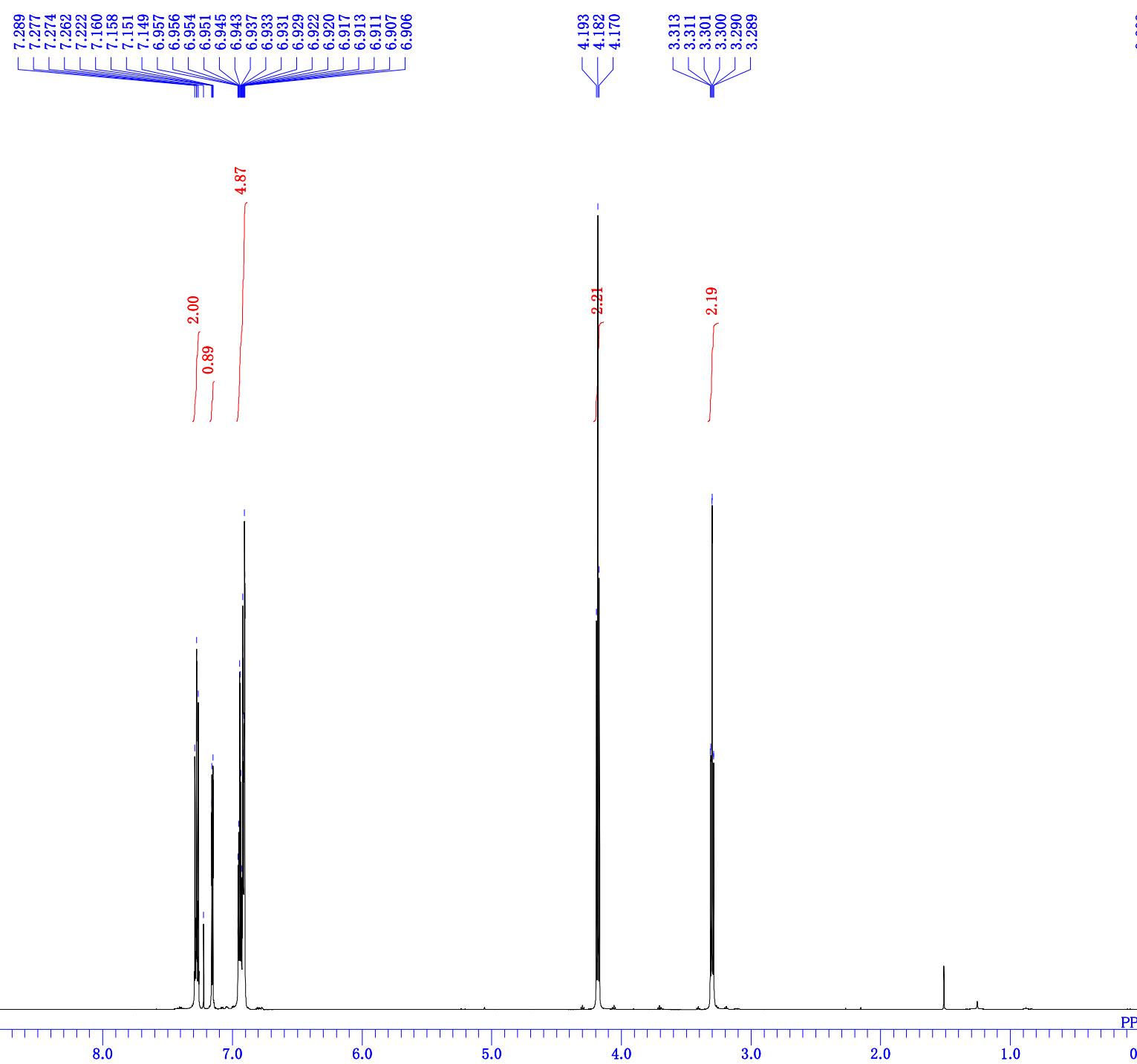
37.131
 36.144

29.519
 25.718
 25.440

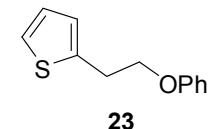
19.542
 17.656

DFILE d1535-gra-13c-1.als
 COMNT 150704
 DATIM 2015-03-24 21:42:15
 13C 13C
 single_pulse_dec
 EXMOD 150.92 MHz
 OBFRQ 8.52 KHz
 OBSET 1.74 Hz
 OBFIN 26214
 POINT 37878.21 Hz
 FREQU 256
 SCANS 0.6921 sec
 ACQTM 1.2000 sec
 PD 3.13 usec
 PW1
 IRNUC
 CTEMP 20.9 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 56





DFILE d1526-gra-1h-thio-1.als
 COMNT 150704
 DATIM 2015-03-20 10:34:22
 1H
 single_pulse.ex2
 EXMOD 600.17 MHz
 OBFRQ 5.30 KHz
 OBSET 5.47 Hz
 OBFIN 26214
 POINT 9008.87 Hz
 FREQU 32
 SCANS 2.9098 sec
 ACQTM 2.0000 sec
 PD 7.30 usec
 PW1 1H 20.9 c
 IRNUC CDCL₃
 CTEMP 0.00 ppm
 SLVNT 0.12 Hz
 EXREF 34
 RGAIN



158.587

140.385

129.451
126.770
125.458
123.916
120.862

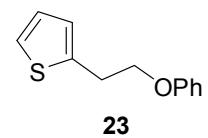
114.591

77.211
77.000
76.789

68.258

29.988

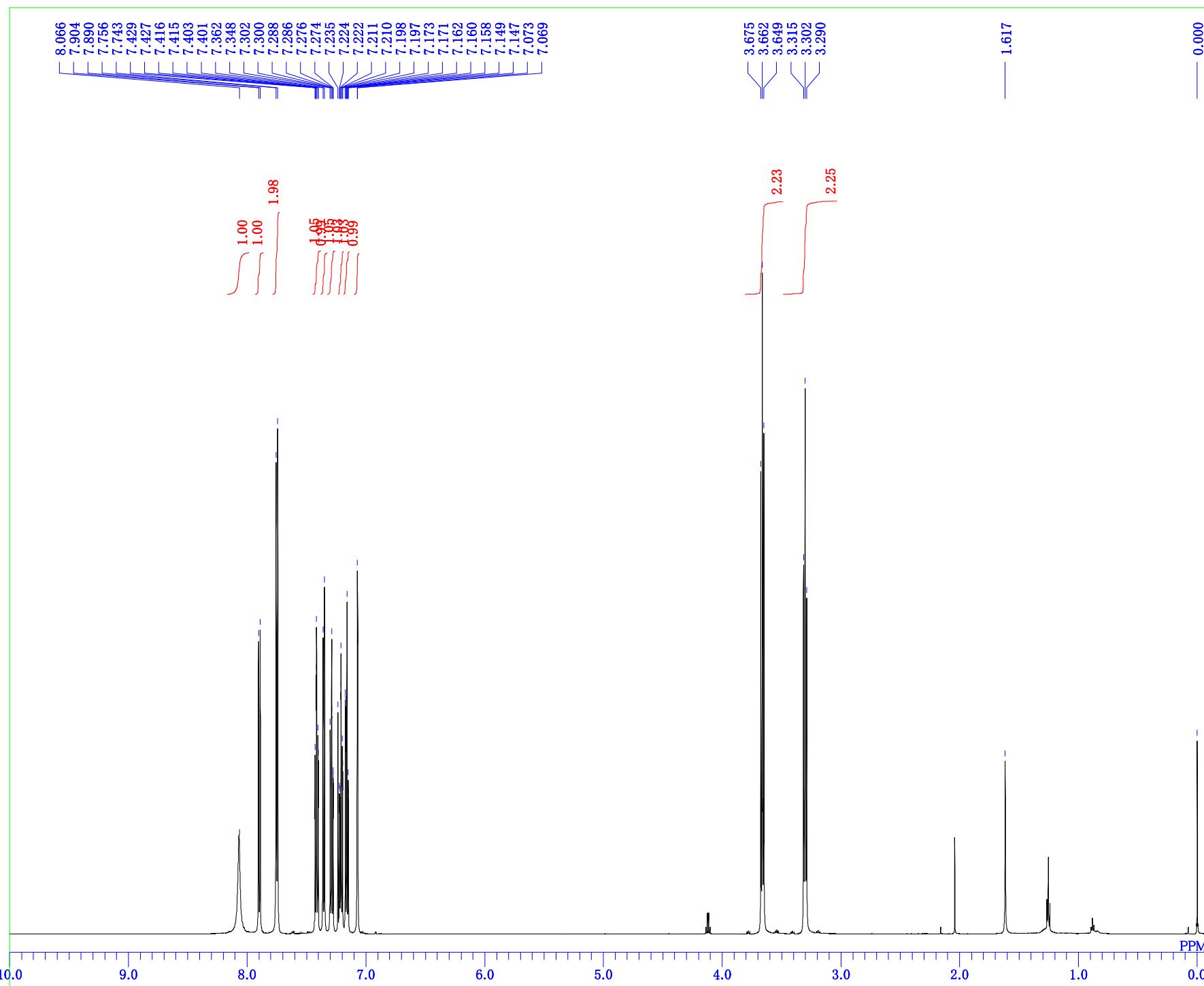
DFILE d1526-gra-13c-thio-1.als
 COMNT 150704
 DATIM 2015-03-20 10:42:58
 OBNUC 13C
 EXMOD single_pulse_dec
 OBFRQ 150.92 MHz
 OBSET 8.52 KHz
 OBFIN 1.74 Hz
 POINT 26214
 FREQU 37878.21 Hz
 SCANS 256
 ACQTM 0.6921 sec
 PD 1.2000 sec
 PW1 3.13 usec
 IRNUC 1H
 CTEMP 21.8 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 56



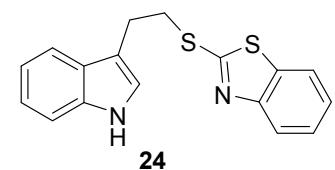
PPM

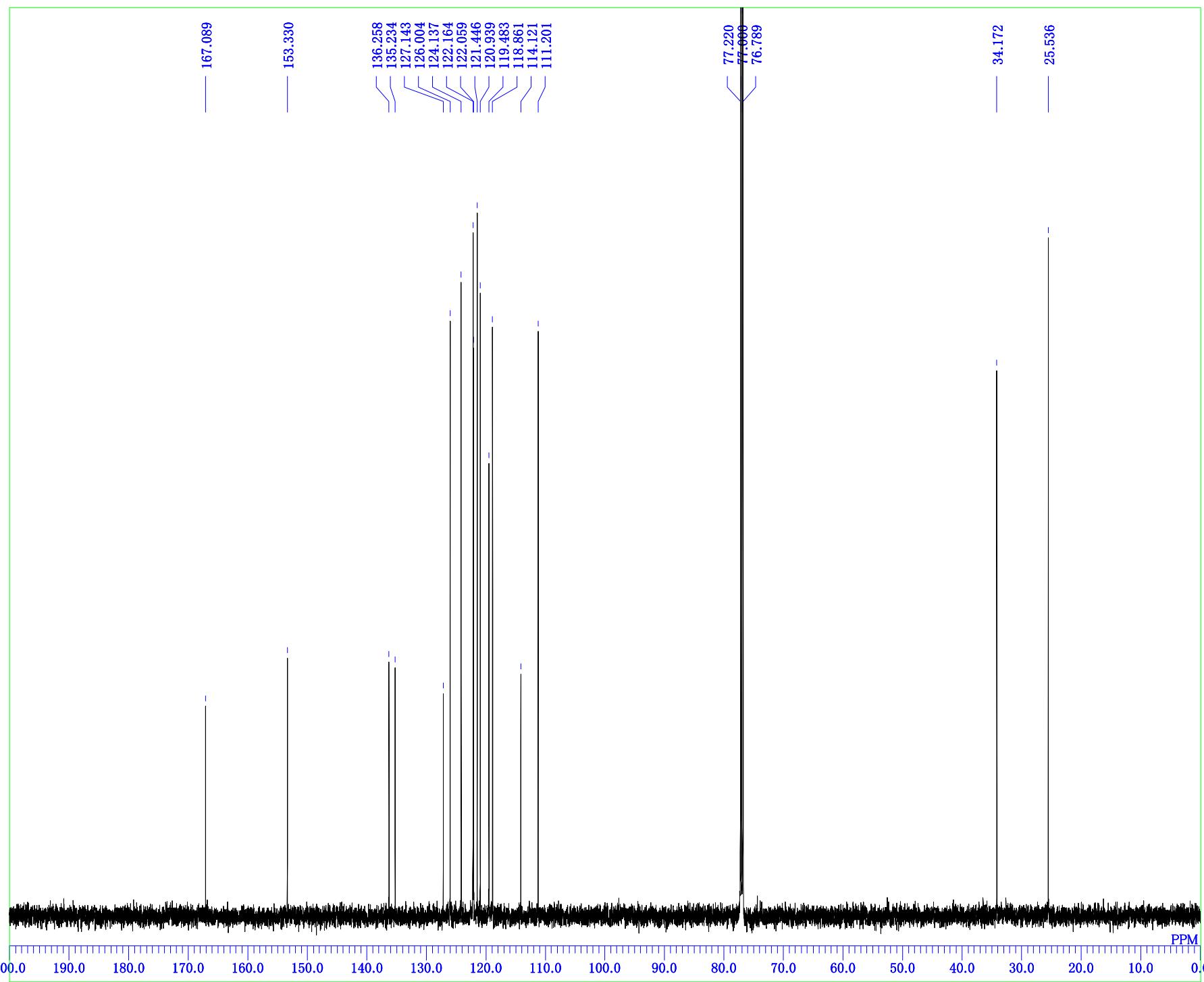
S85

200.0 190.0 180.0 170.0 160.0 150.0 140.0 130.0 120.0 110.0 100.0 90.0 80.0 70.0 60.0 50.0 40.0 30.0 20.0 10.0 0.0

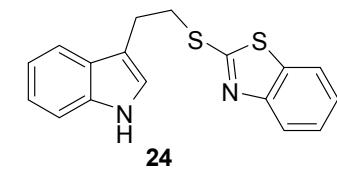


DFILE d1589-gra-1h-1.als
 COMNT 150704
 DATIM 2015-05-24 13:04:38
 1H
 single_pulse.ex2
 EXMOD 600.17 MHz
 OBFRQ 5.30 KHz
 OBSET 5.47 Hz
 OBFIN 5.47 Hz
 POINT 26214
 FREQU 9008.87 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 7.30 usec
 IRNUC 1H
 CTEMP 22.6 c
 SLVNT CDCL₃
 EXREF 0.00 ppm
 BF 0.12 Hz
 RGAIN 40





DFILE d1589-gra-13c-1.als
 COMNT 150704
 DATIM 2015-05-24 13:14:46
 OBNUC 13C
 EXMOD single_pulse_dec
 OBFRQ 150.92 MHz
 OBSET 8.52 KHz
 OBFIN 1.74 Hz
 POINT 26214
 FREQU 37878.21 Hz
 SCANS 256
 ACQTM 0.6921 sec
 PD 1.2000 sec
 PW1 3.13 usec
 IRNUC 1H
 CTEMP 23.4 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 56



150519

7.725
7.711
7.328
7.315
7.270

1.96
2.00

3.243
3.238
3.235
3.231
3.227
3.224
3.220

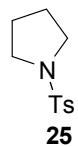
2.434

4.30
3.17

4.19

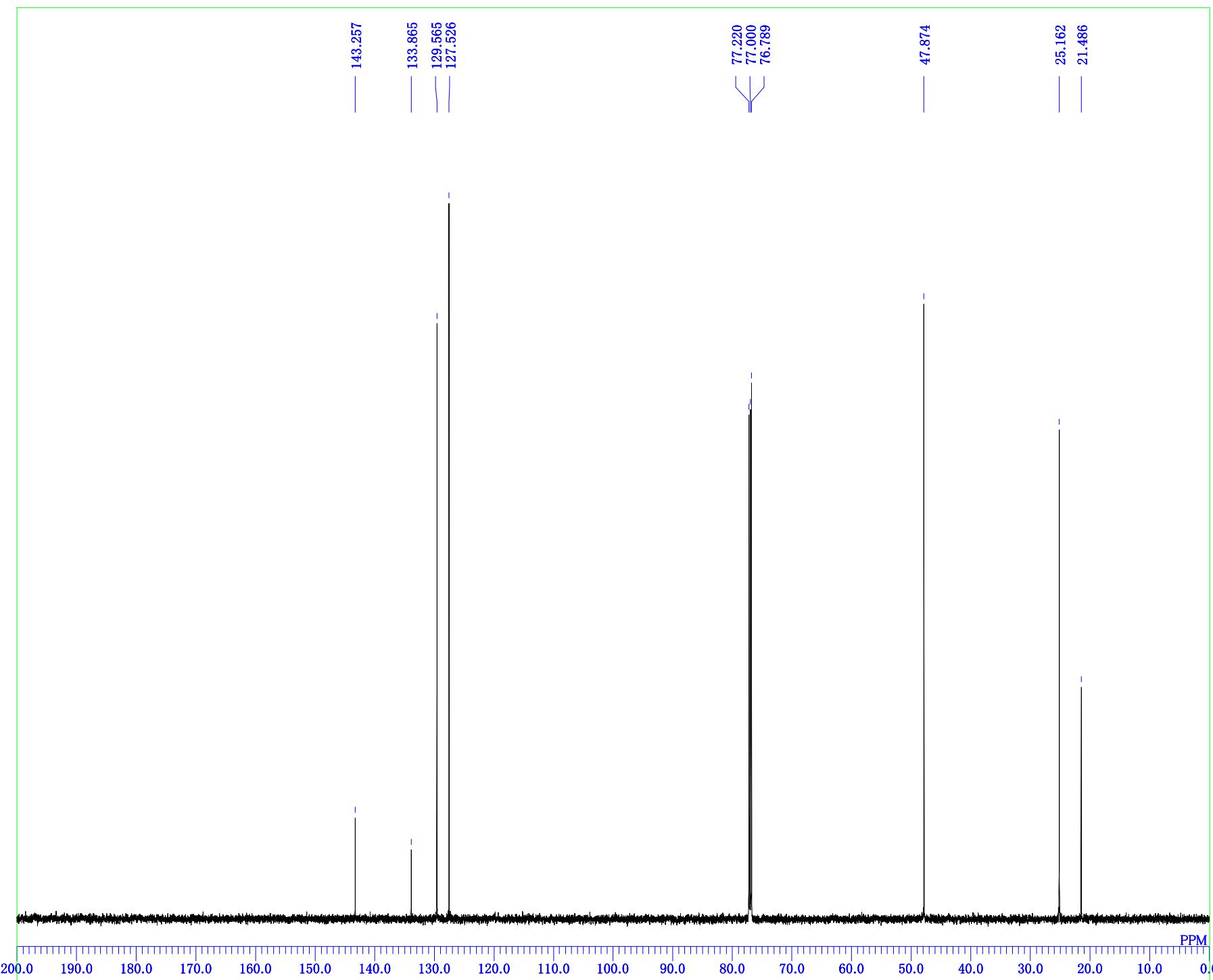
0.000

DFILE d1583-gra-1h-1.als
 COMNT 150519
 DATIM 2015-05-19 18:45:02
 1H
 single_pulse.ex2
 EXMOD 600.17 MHz
 OBFRQ 5.30 KHz
 OBSET 5.47 Hz
 OBFIN 5.47 Hz
 POINT 26214
 FREQU 9008.87 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 7.30 usec
 IRNUC 1H
 CTEMP 21.4 c
 SLVNT CDCL3
 EXREF 0.00 ppm
 BF 0.12 Hz
 RGAIN 38

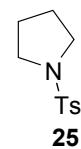


10.0 9.0 8.0 7.0 6.0 5.0 4.0 3.0 2.0 1.0 0.0 PPM

S88



DFILE d1583-gra-13c-1.als
 COMNT 150519
 DATIM 2015-05-19 18:53:36
¹³C 13C
 EXMOD single_pulse_dec
 OBFRQ 150.92 MHz
 OBSET 8.52 KHz
 OBFIN 1.74 Hz
 POINT 26214
 FREQU 37878.21 Hz
 SCANS 256
 ACQTM 0.6921 sec
 PD 1.2000 sec
 PW1 3.13 usec
 IRNUC 1H
 CTEMP 21.9 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 54



150727

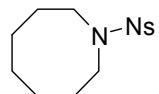
7.938
7.926
7.920
7.880
7.876
7.868
7.861
7.856
7.596
7.590
7.583
7.580
7.577
7.265

3.338
3.327
3.314

1.795
1.772
1.669
1.656
1.579

0.000

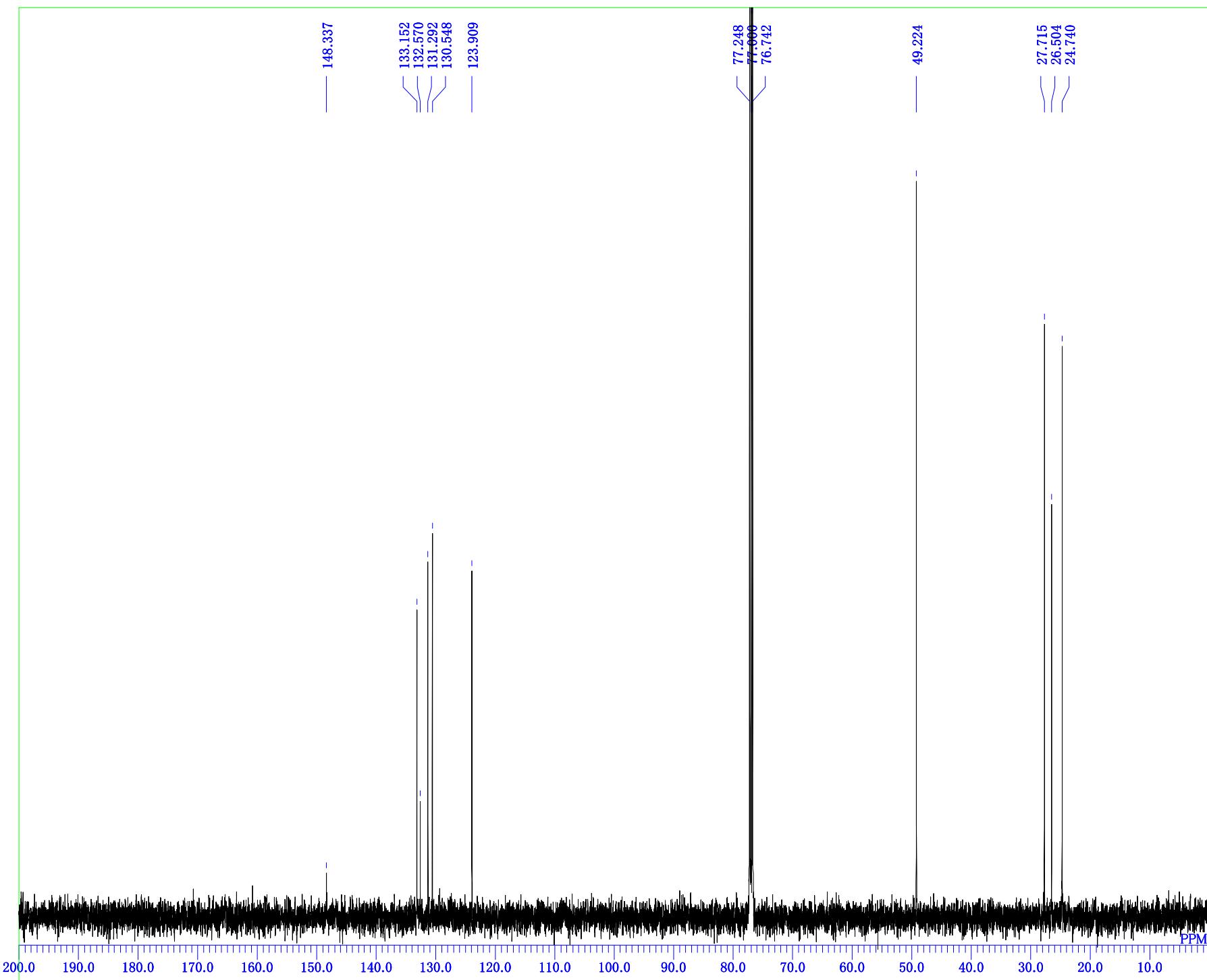
DFILE d1671-gra-1h-1.als
COMNT 150727
DATIM 2015-07-27 08:50:01
1H
EXMOD single_pulse.ex2
OBFRQ 500.16 MHz
OBSET 2.41 KHz
OBFIN 6.01 Hz
POINT 13107
FREQU 7507.39 Hz
SCANS 32
ACQTM 1.7459 sec
PD 2.0000 sec
PW1 5.80 usec
IRNUC 1H
CTEMP 20.1 c
SLVNT CDCL3
EXREF 0.00 ppm
BF 0.12 Hz
RGAIN 52



26

10.0 9.0 8.0 7.0 6.0 5.0 4.0 3.0 2.0 1.0 0.0 PPM

S90

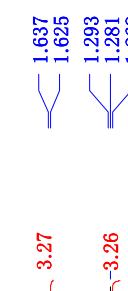
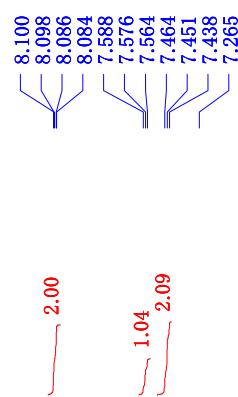


d1671-gra-13c-1.als
150727
2015-07-27 09:04:03
¹³C
single_pulse_dec
125.77 MHz
7.87 KHz
OBSET
OBFIN
POINT
FREQU
SCANS
ACQTM
PD
PW1
IRNUC
CTEMP
SLVNT
EXREF
BF
RGAIN

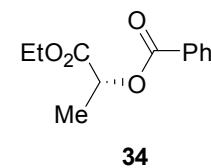
1H
CDCL₃

256
0.8336 sec
2.0000 sec
3.00 usec
21.3 c
77.00 ppm
1.20 Hz
56

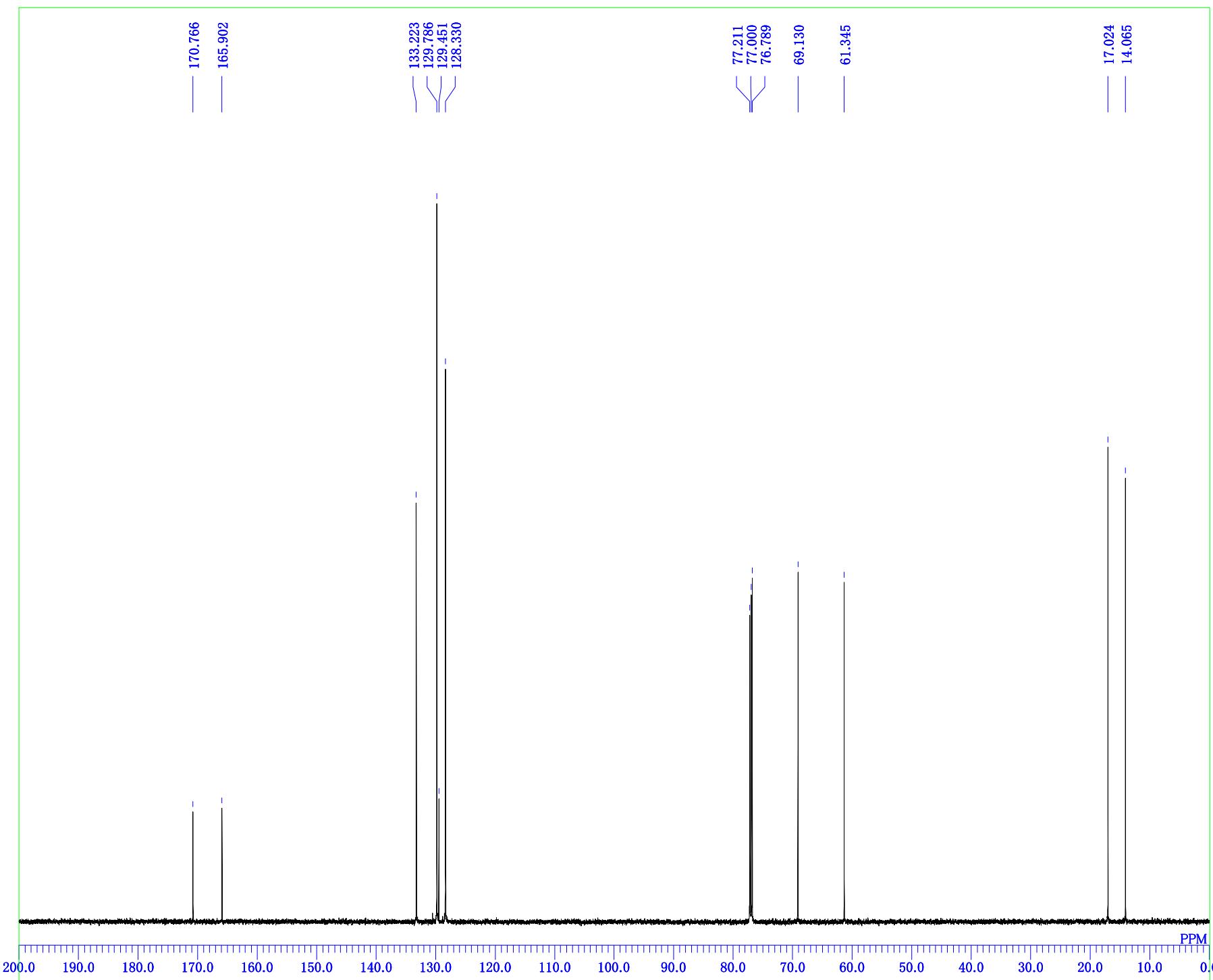
150306



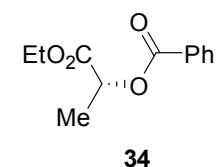
DFILE d1502-gra-1h-1.als
COMNT 150306
DATIM 2015-03-06 16:06:43
1H single_pulse.ex2
EXMOD 600.17 MHz
OBFRQ 5.30 KHz
OBSET 5.47 Hz
OBFIN 26214
POINT 9008.87 Hz
FREQU 32
SCANS 2.9098 sec
ACQTM PD 2.0000 sec
PW1 5.85 usec
IRNUC 1H 20.2 c
CTEMP CDCL3 0.00 ppm
SLVNT 0.12 Hz
EXREF 30
BF RGAIN



150306

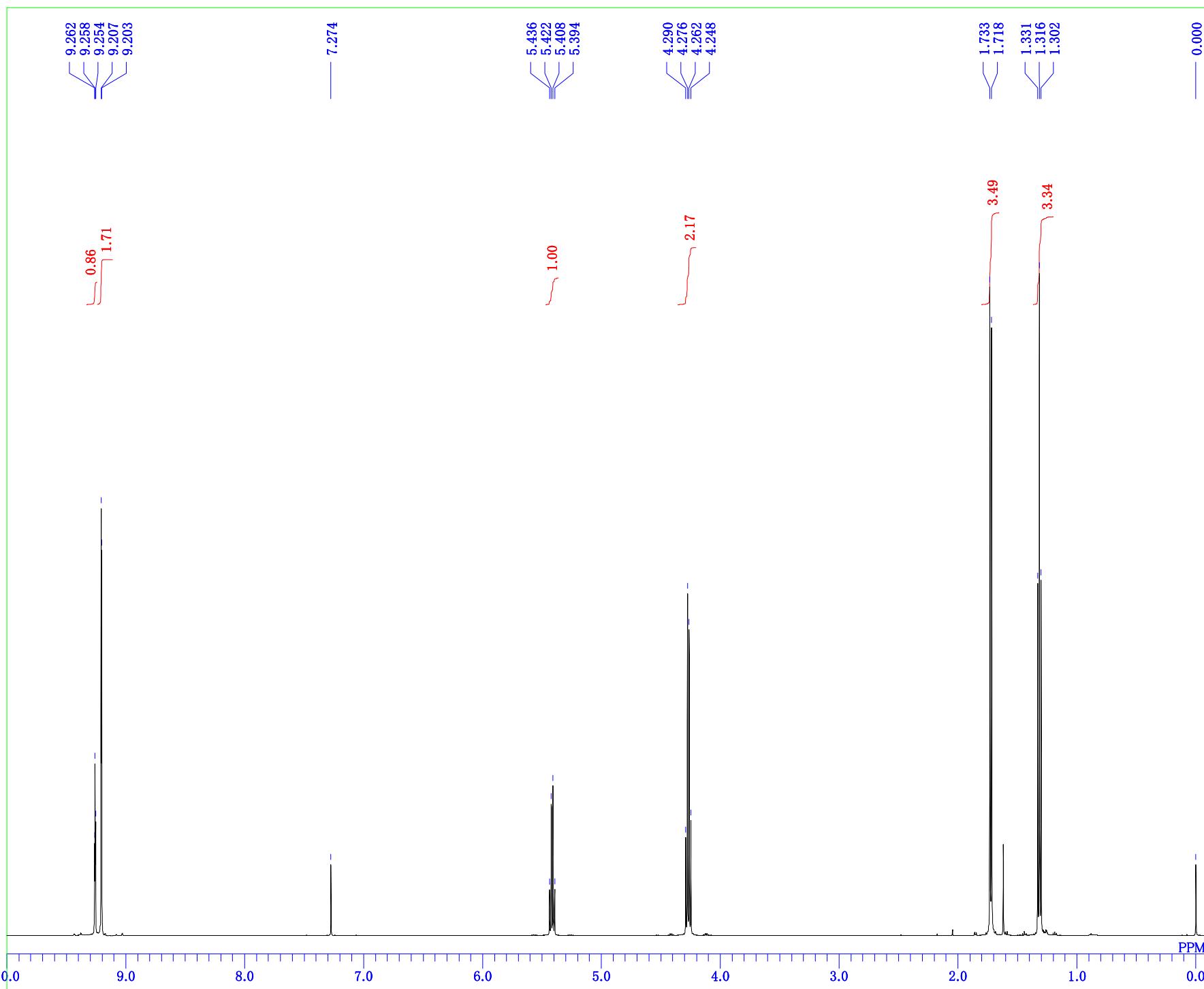


DFILE d1502-gra-13c-1.als
 COMNT 150306
 DATIM 2015-03-06 16:15:25
 13C 13C
 EXMOD single_pulse_dec
 OBFRQ 150.92 MHz
 OBSET 8.52 KHz
 OBFIN 1.74 Hz
 POINT 26214
 FREQU 37878.21 Hz
 SCANS 256
 ACQTM 0.6921 sec
 PD 1.2000 sec
 PW1 2.97 usec
 IRNUC 1H
 CTEMP 20.8 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 56

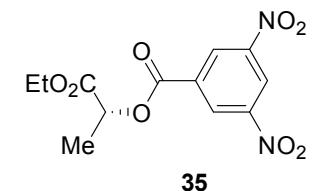


S93

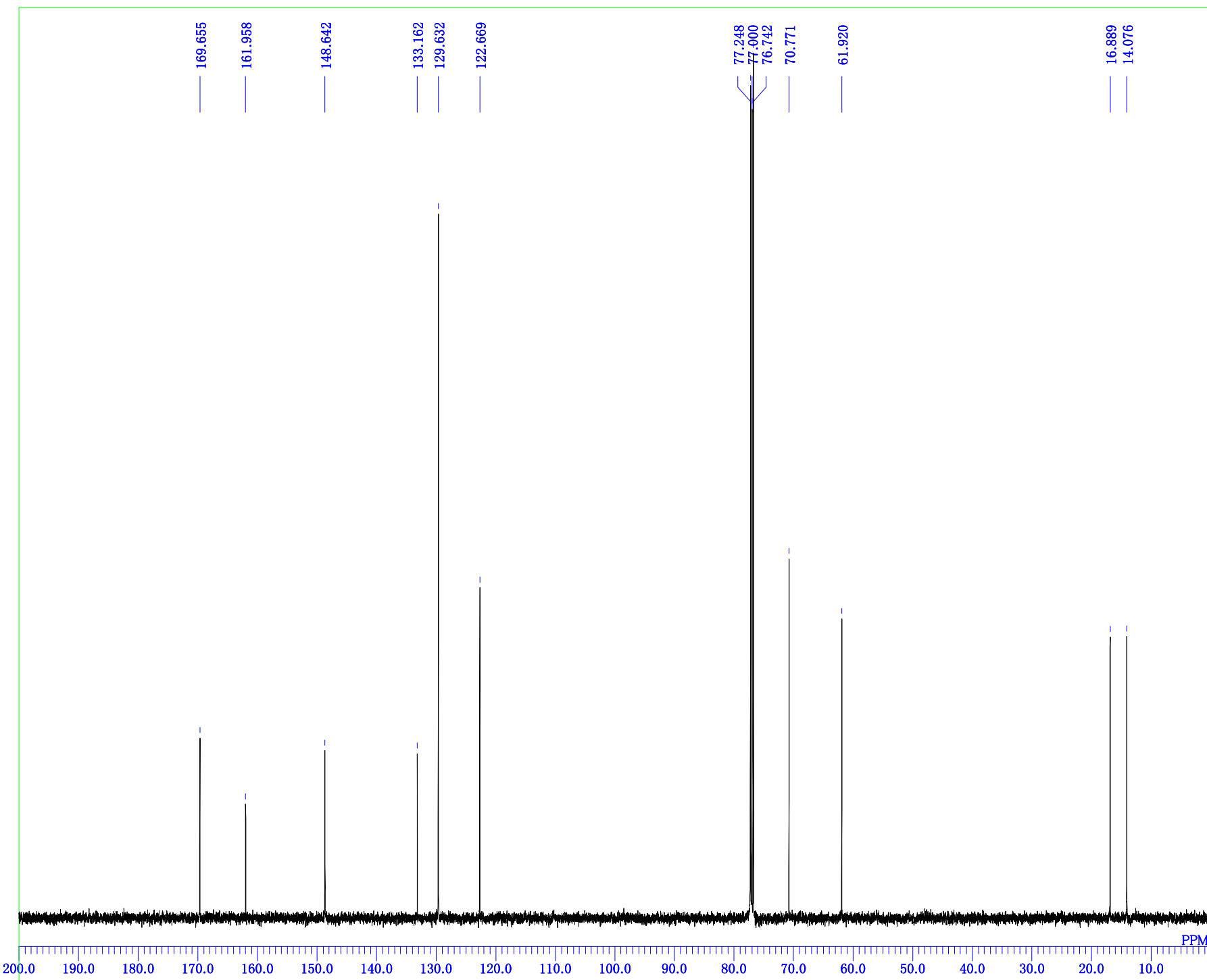
150621



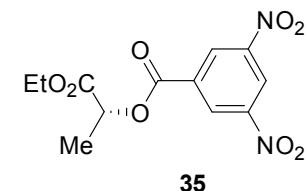
DFILE d1609-gra-1h-1.als
 COMNT 150621
 DATIM 2015-06-21 17:14:42
 1H
 single_pulse.ex2
 EXMOD 500.16 MHz
 OBFRQ 2.41 KHz
 OBSET 6.01 Hz
 OBFIN 13107
 POINT 7507.39 Hz
 FREQU 32
 SCANS 1.7459 sec
 ACQTM 2.0000 sec
 PD 5.80 usec
 PW1 1H
 IRNUC 20.5 c
 CTEMP CDCL3
 SLVNT 0.00 ppm
 EXREF 0.12 Hz
 RGAIN 40



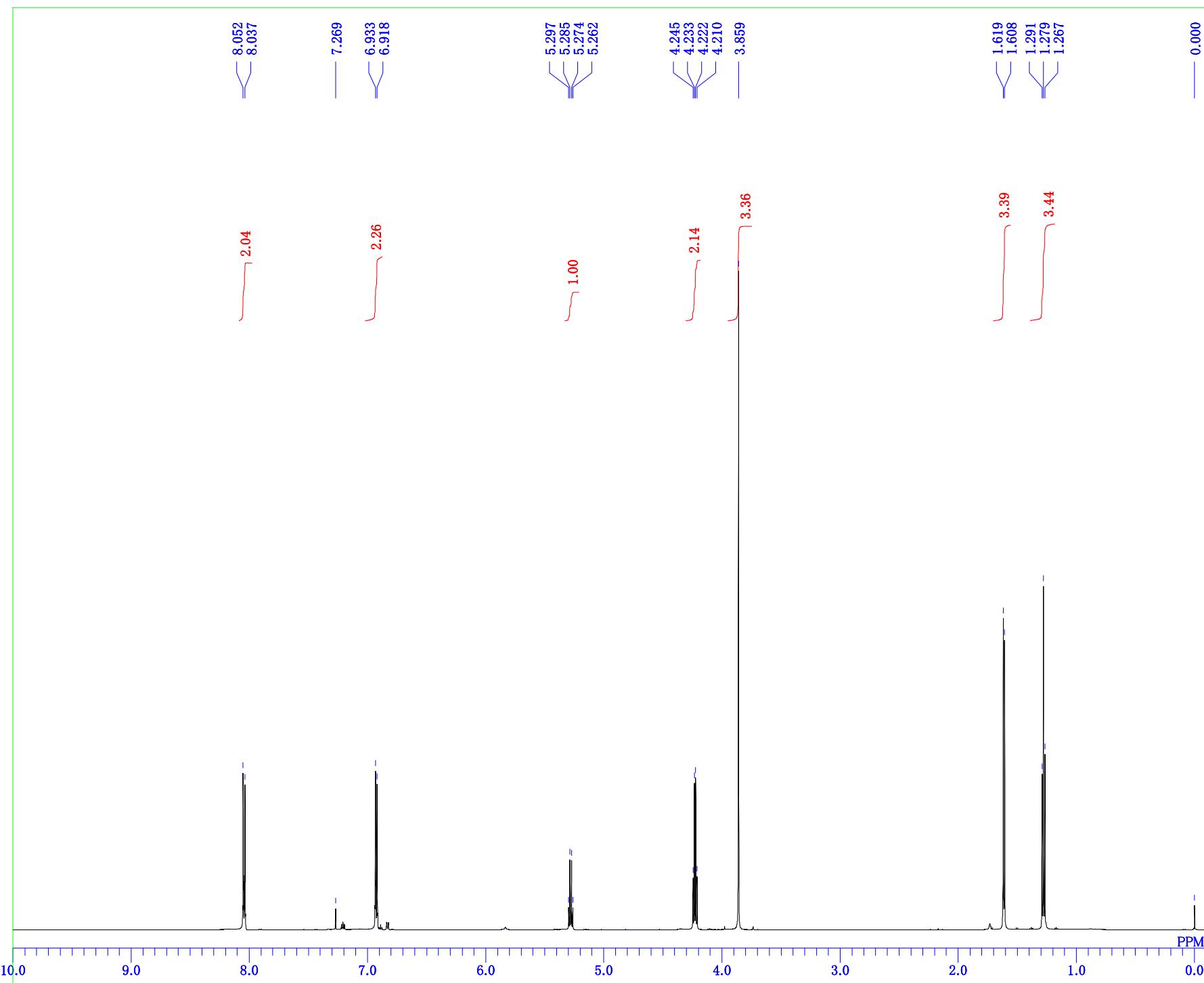
150621



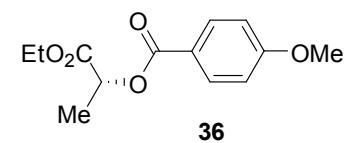
DFILE d1609-gra-13c-1.als
 COMNT 150621
 DATIM 2015-06-21 17:27:43
 13C single_pulse_dec
 EXMOD 125.77 MHz
 OBFRQ 7.87 KHz
 OBSET 4.21 Hz
 OBFIN 26214
 POINT 31446.06 Hz
 FREQU 256
 SCANS 0.8336 sec
 ACQTM PD 2.0000 sec
 PW1 3.00 usec
 IRNUC 1H 21.3 c
 CTEMP CDCL3
 SLVNT 77.00 ppm
 EXREF 1.20 Hz
 BF 58
 RGAIN

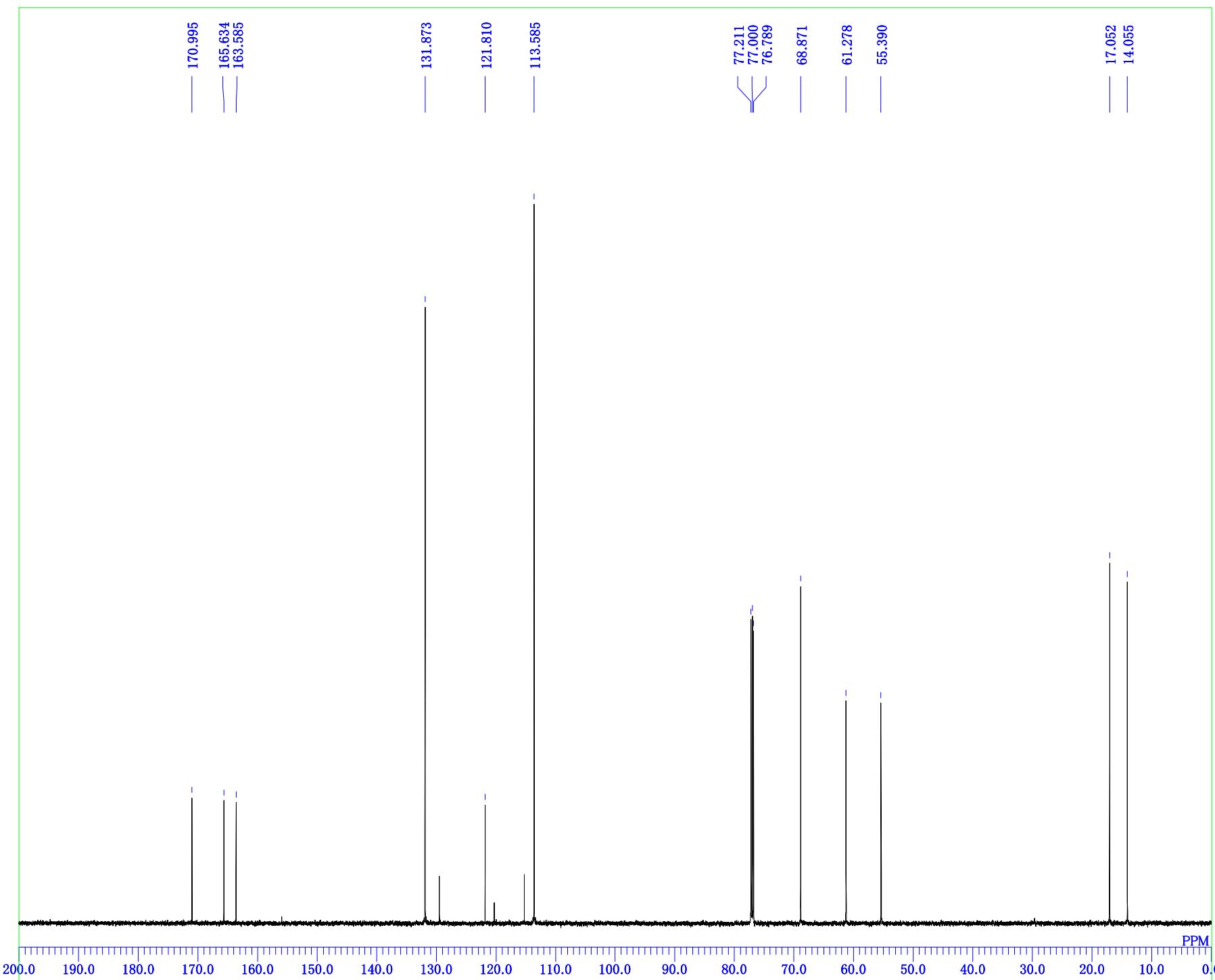


150406

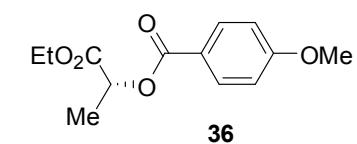


DFILE d1546-gra-1h-1.als
 COMNT 150406
 DATIM 2015-04-06 15:11:47
 1H
 single_pulse.ex2
 EXMOD 600.17 MHz
 OBFRQ 5.30 KHz
 OBSET 5.47 Hz
 OBFIN 26214
 POINT 26214
 FREQU 9008.87 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 7.30 usec
 IRNUC 1H
 CTEMP 20.9 c
 SLVNT CDCL3
 EXREF 0.00 ppm
 BF 0.12 Hz
 RGAIN 30

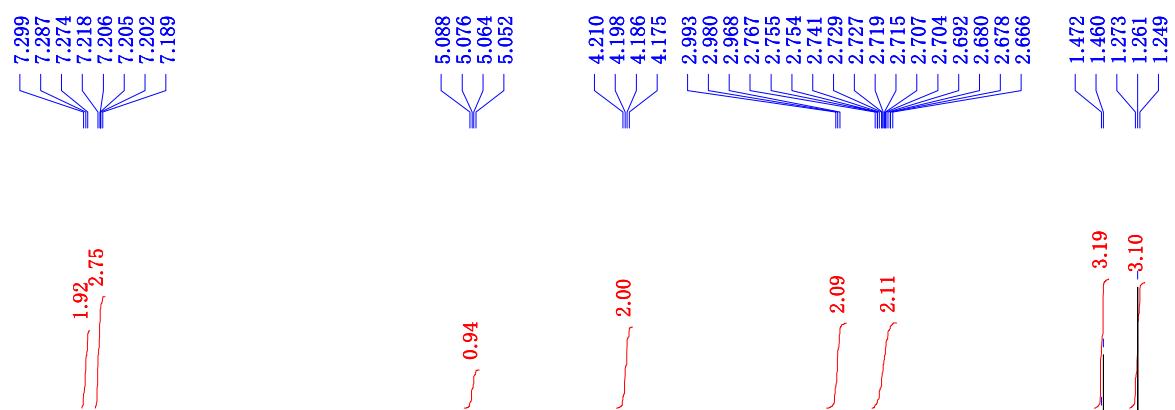




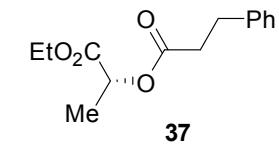
DFILE d1546-gra-13c-1.als
 COMNT 150406
 DATIM 2015-04-06 15:20:27
 13C single_pulse_dec
 EXMOD 150.92 MHz
 OBFRQ 8.52 KHz
 OBSET 1.74 Hz
 OBFIN 26214
 POINT 37878.21 Hz
 FREQU 256
 SCANS 0.6921 sec
 ACQTM PD 1.2000 sec
 PW1 3.13 usec
 IRNUC 1H 21.4 c
 CTEMP CDCL3
 SLVNT 77.00 ppm
 EXREF 1.20 Hz
 BF 56
 RGAIN



150418



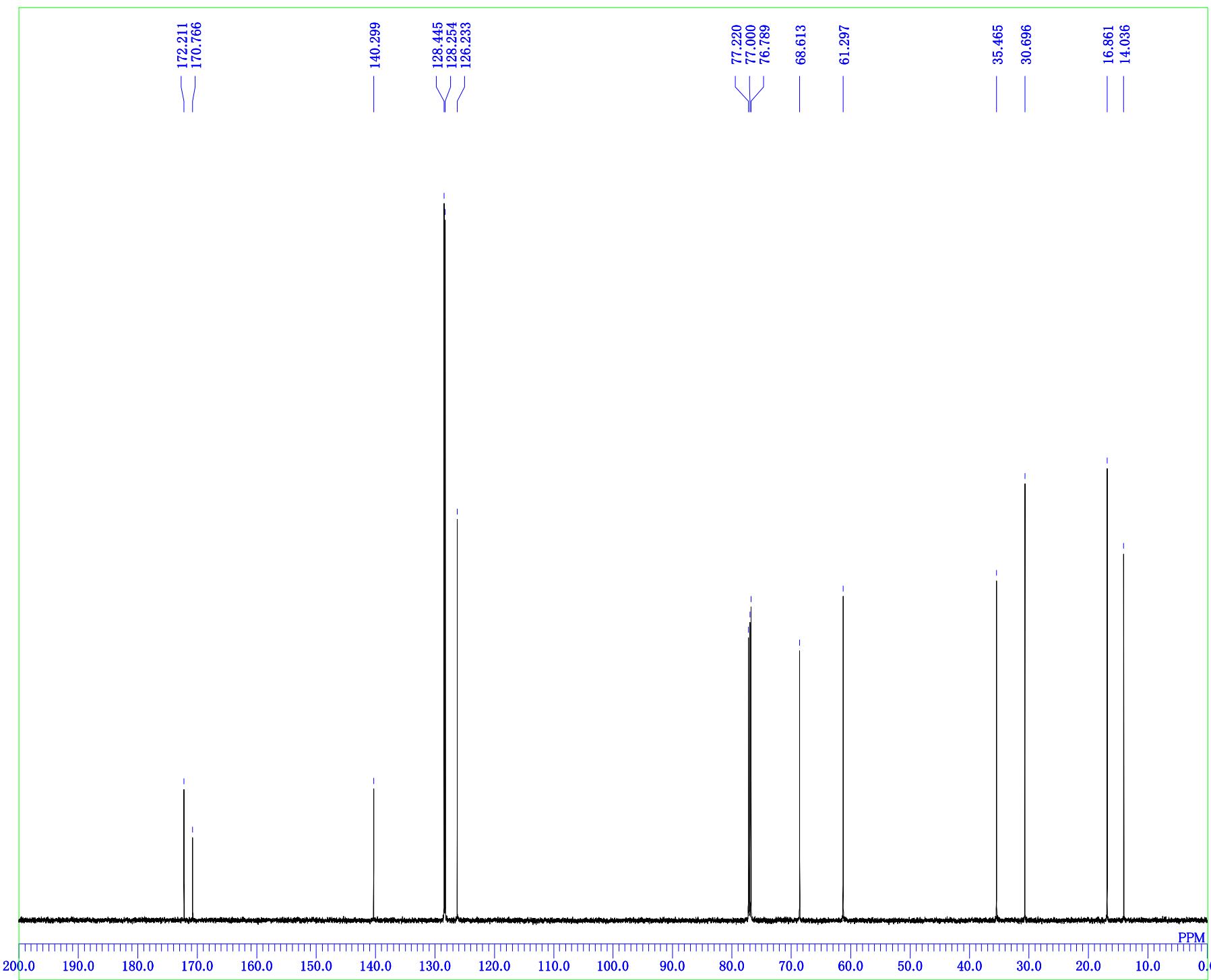
DFILE d1533-gra-1h-1.als
 COMNT 150418
 DATIM 2015-04-18 15:47:07
 1H
 single_pulse.ex2
 EXMOD 600.17 MHz
 OBFRQ 5.30 KHz
 OBSET 5.47 Hz
 OBFIN 26214
 POINT 26214
 FREQU 9008.87 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 7.30 usec
 1H 21.0 c
 CTEMP CDCL3
 SLVNT 0.00 ppm
 EXREF 0.12 Hz
 RGAIN 30



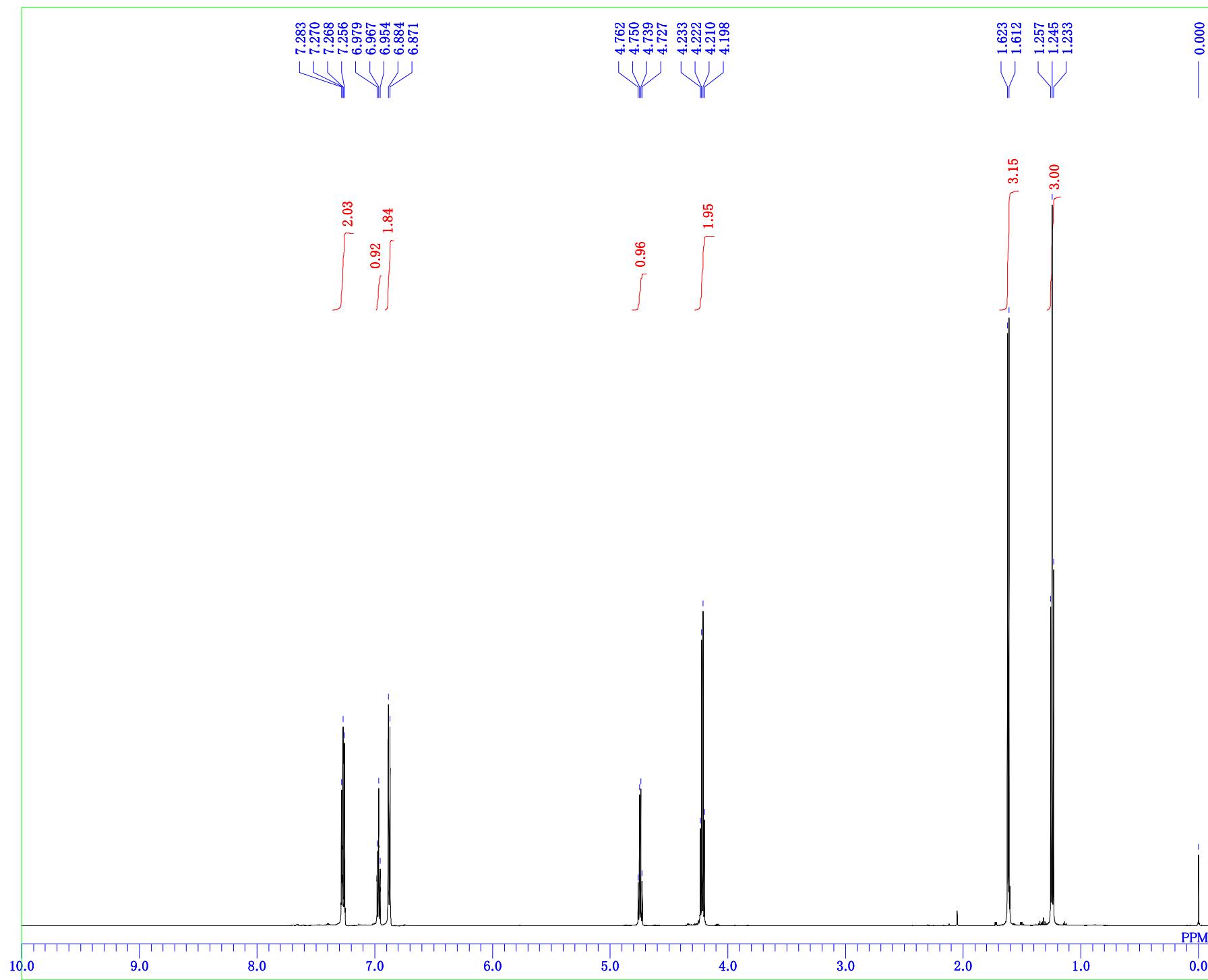
10.0 9.0 8.0 7.0 6.0 5.0 4.0 3.0 2.0 1.0 0.0 PPM

S98

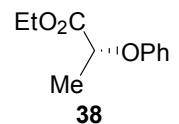
150418



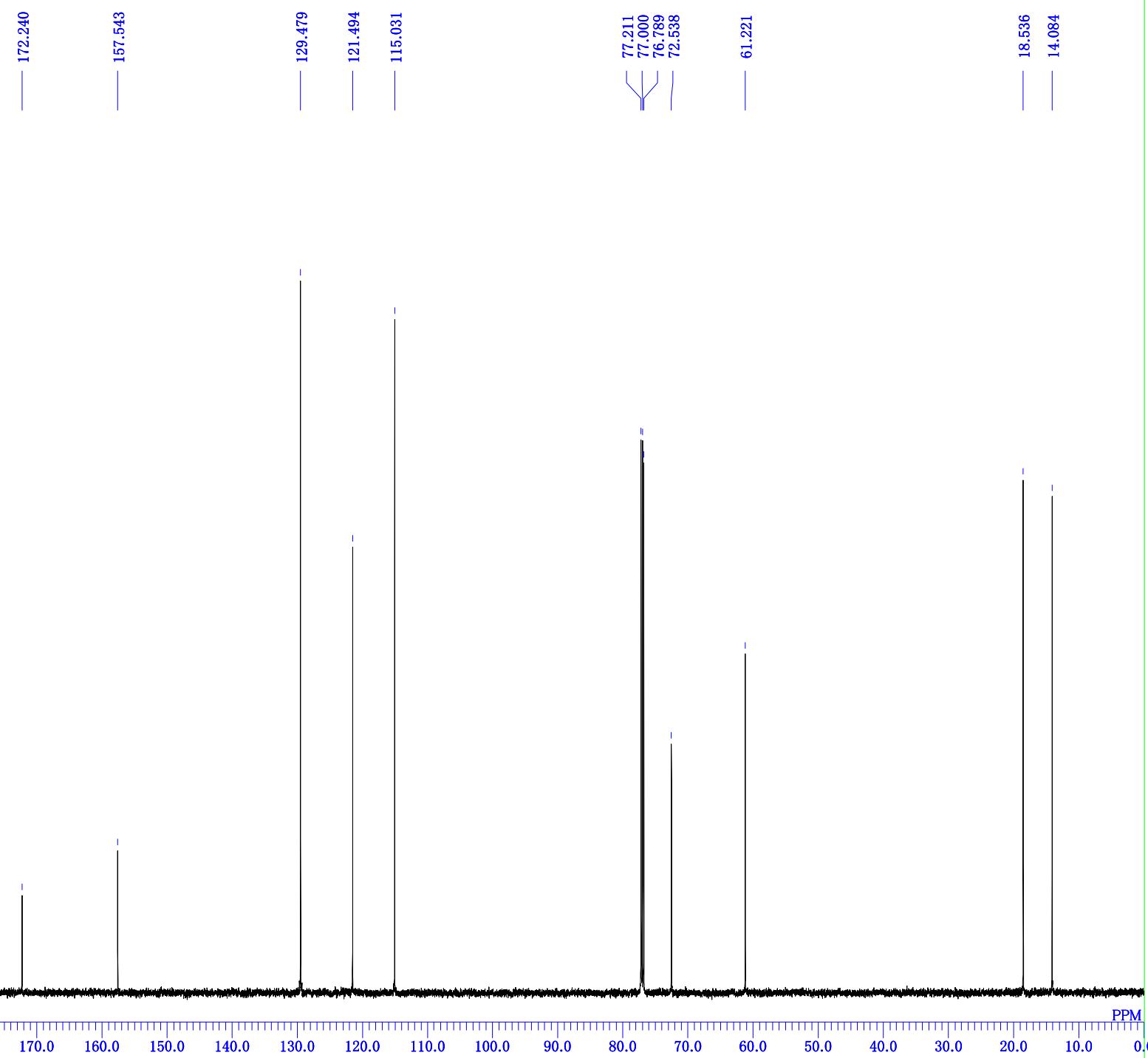
150324



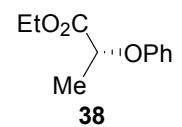
DFILE d1515-gra-1h-1.als
 COMNT 150324
 DATIM 2015-03-24 20:53:03
 1H
 single_pulse.ex2
 EXMOD 600.17 MHz
 OBFRQ 5.30 KHz
 OBSET 5.47 Hz
 OBFIN 26214
 POINT 26214
 FREQU 9008.87 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 7.30 usec
 1H 20.3 c
 CTEMP CDCL₃
 SLVNT 0.00 ppm
 EXREF 0.12 Hz
 BF 38
 RGAIN

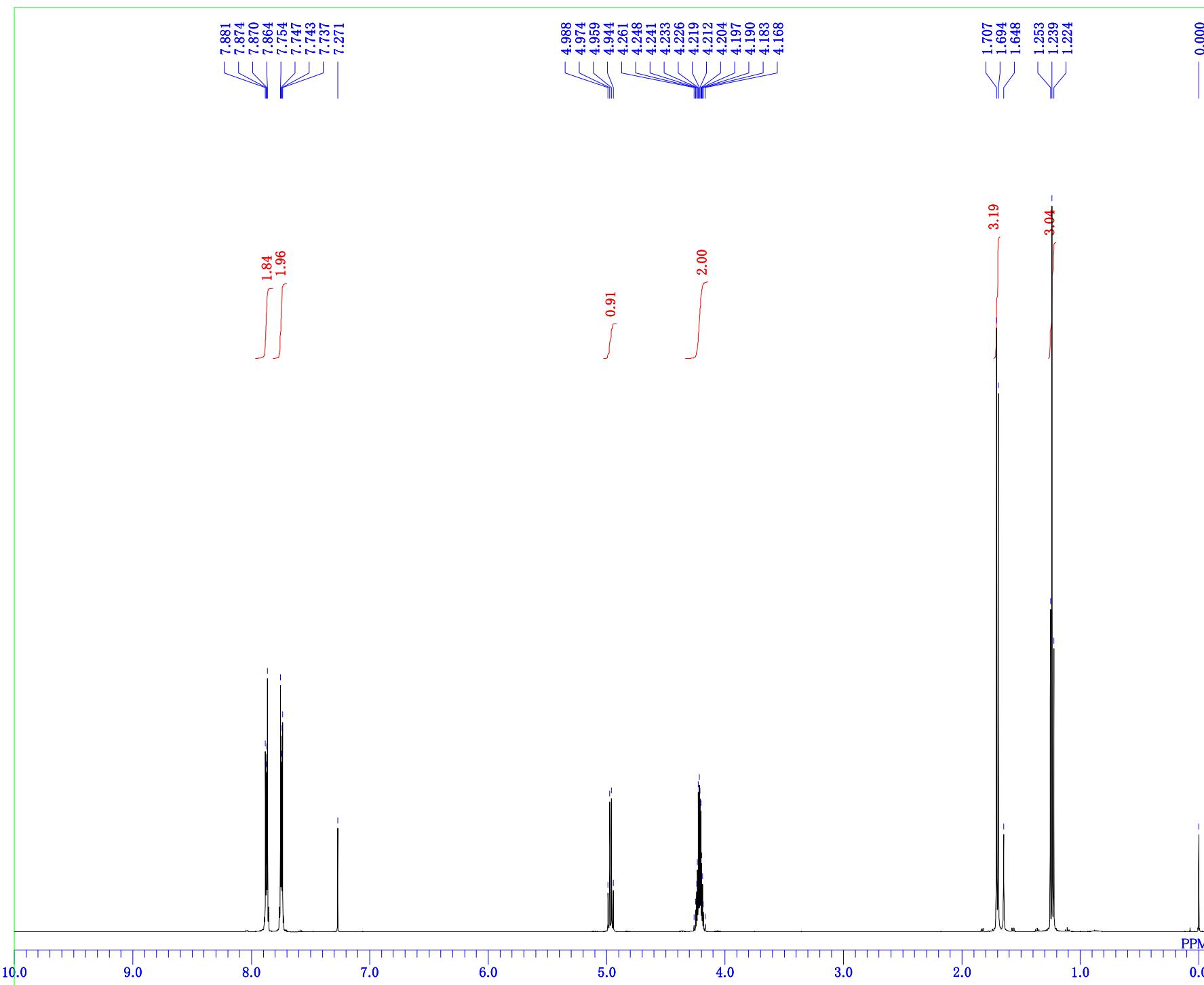


S100



DFILE d1515-gra-13c-1.als
 COMNT 150324
 DATIM 2015-03-24 21:01:43
 13C
 single_pulse_dec
 EXMOD 150.92 MHz
 OBFRQ 8.52 KHz
 OBSET 1.74 Hz
 OBFIN 26214
 POINT 37878.21 Hz
 FREQU 256
 SCANS 0.6921 sec
 ACQTM 1.2000 sec
 PD 3.13 usec
 PW1 IRNUC
 CTEMP 20.9 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 56

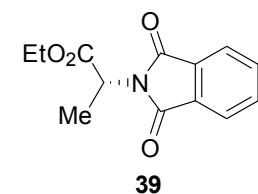


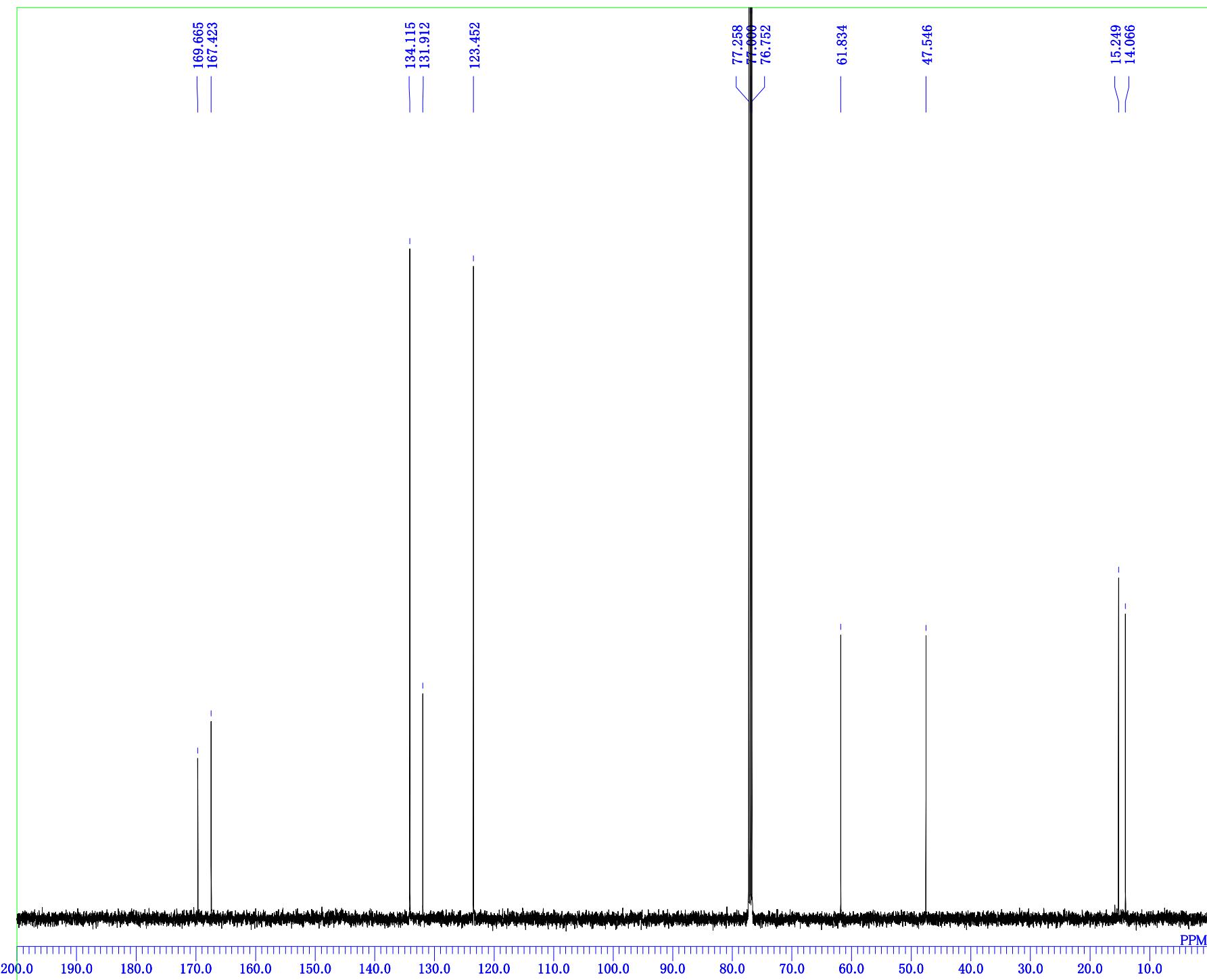


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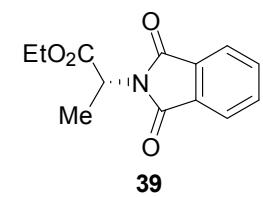
DFILE      d1503-gra-1h-1.als
COMNT     150701
DATIM   2015-07-01 21:12:31
OBNUC
1H
EXMOD
OBFRQ  500.16 MHz
OBSET  2.41 KHz
OBFIN  6.01 Hz
POINT  13107
FREQU  7507.39 Hz
SCANS   32
ACQTM  1.7459 sec
PD      2.0000 sec
PW1    5.80 usec
1H
IRNUC
CTEMP  20.2 c
SLVNT
EXREF
BF
RGAIN
CDCL3  0.00 ppm
          0.12 Hz
          42

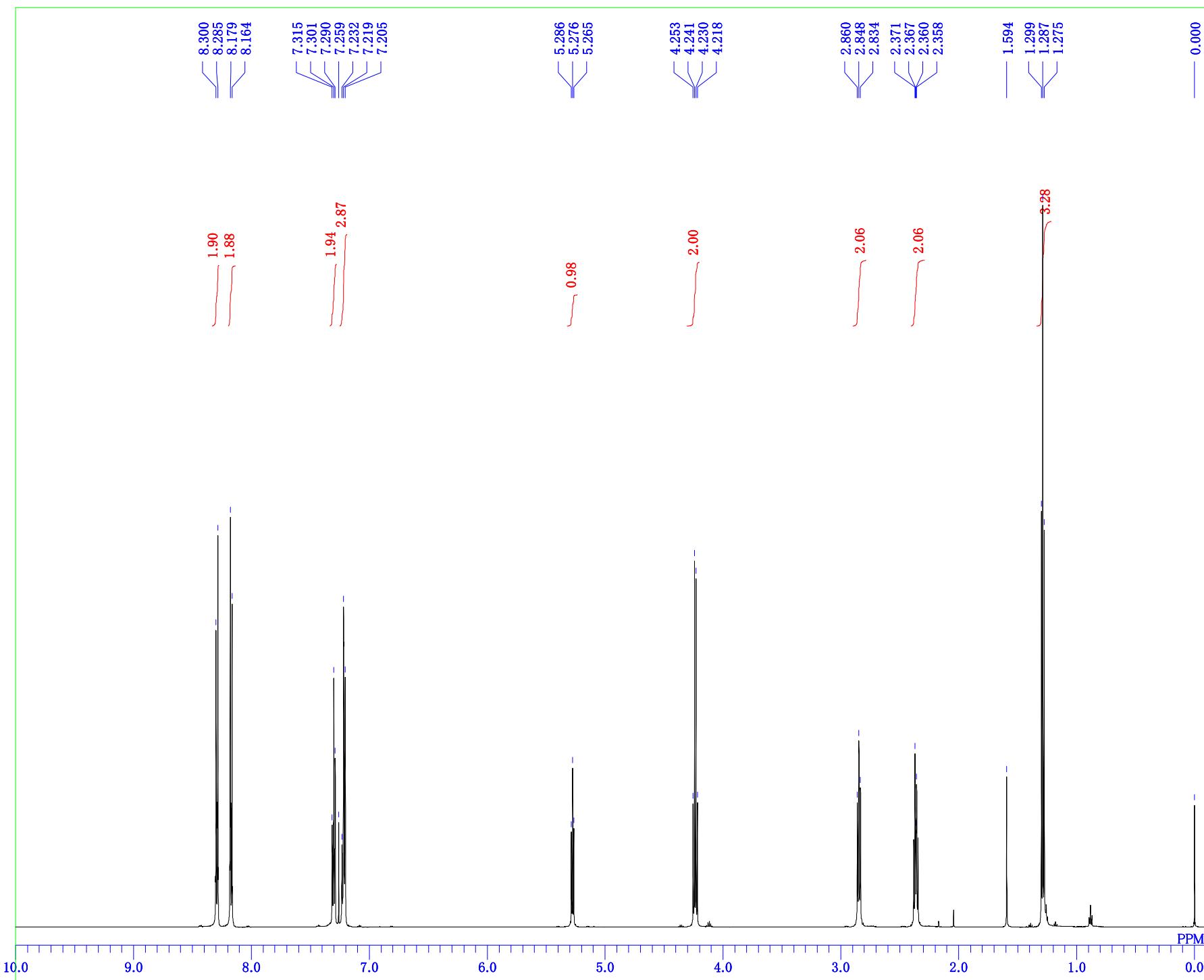
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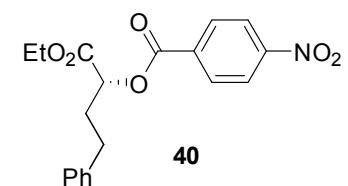


DFILE d1503-gra-13c-1.als
 COMNT 150701
 DATIM 2015-07-01 21:25:17
¹³C 13C
 EXMOD single_pulse_dec
 OBFRQ 125.77 MHz
 OBSET 7.87 KHz
 OBFIN 4.21 Hz
 POINT 26214
 FREQU 31446.06 Hz
 SCANS 256
 ACQTM 0.8336 sec
 PD 2.0000 sec
 PW1 3.00 usec
 IRNUC 1H
 CTEMP 20.7 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 56

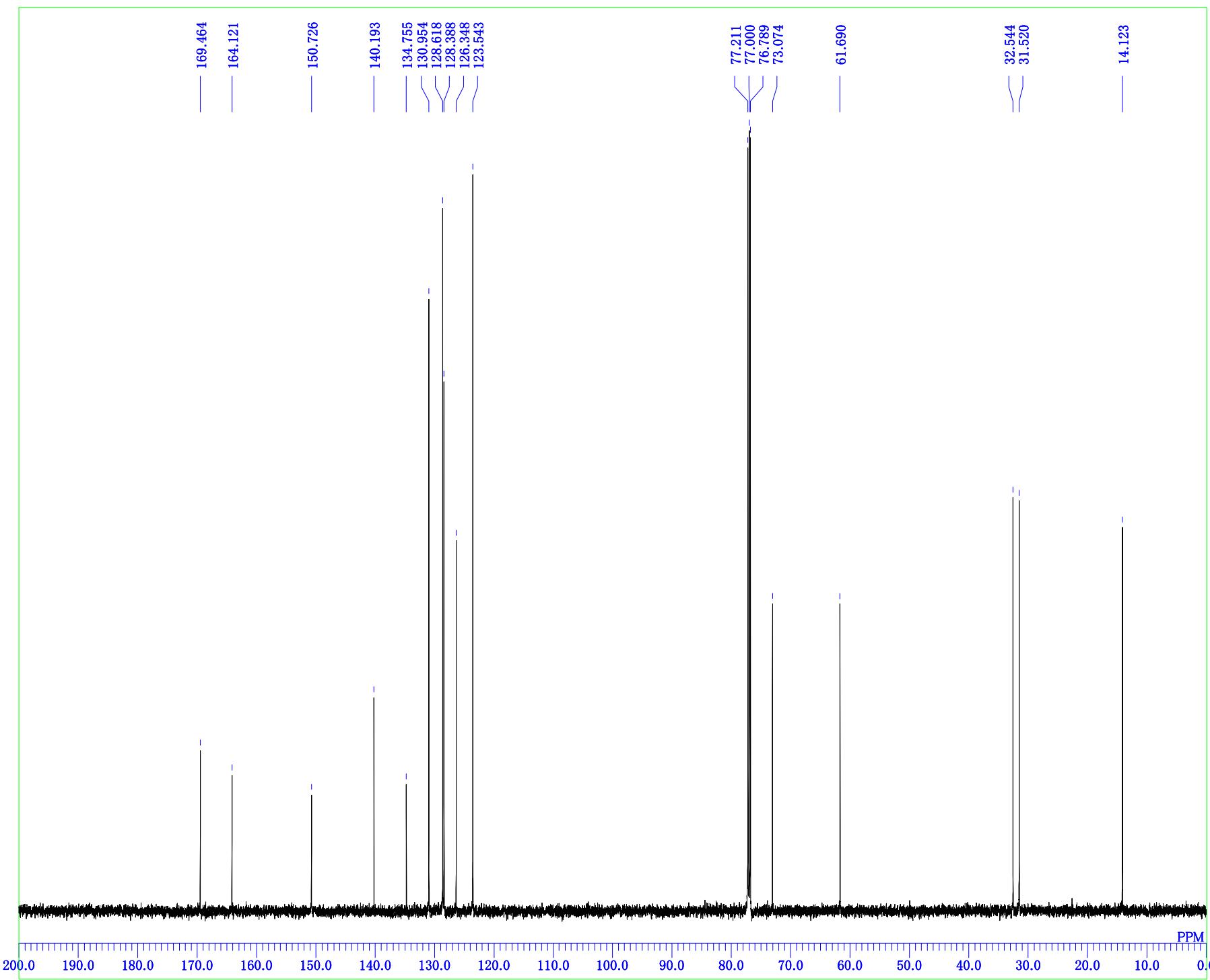




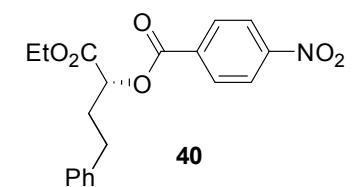
DFILE d1608-gra-1h-1.als
 COMNT 150601
 DATIM 2015-06-01 11:09:43
 1H single_pulse.ex2
 EXMOD 600.17 MHz
 OBFRQ 5.30 KHz
 OBSET 5.47 Hz
 OBFIN 26214
 POINT 9008.87 Hz
 FREQU 32
 SCANS 2.9098 sec
 ACQTM 2.0000 sec
 PD 7.30 usec
 PW1 1H 22.5 c
 IRNUC CDCL3
 CTEMP 0.00 ppm
 SLVNT 0.12 Hz
 EXREF 38
 RGAIN

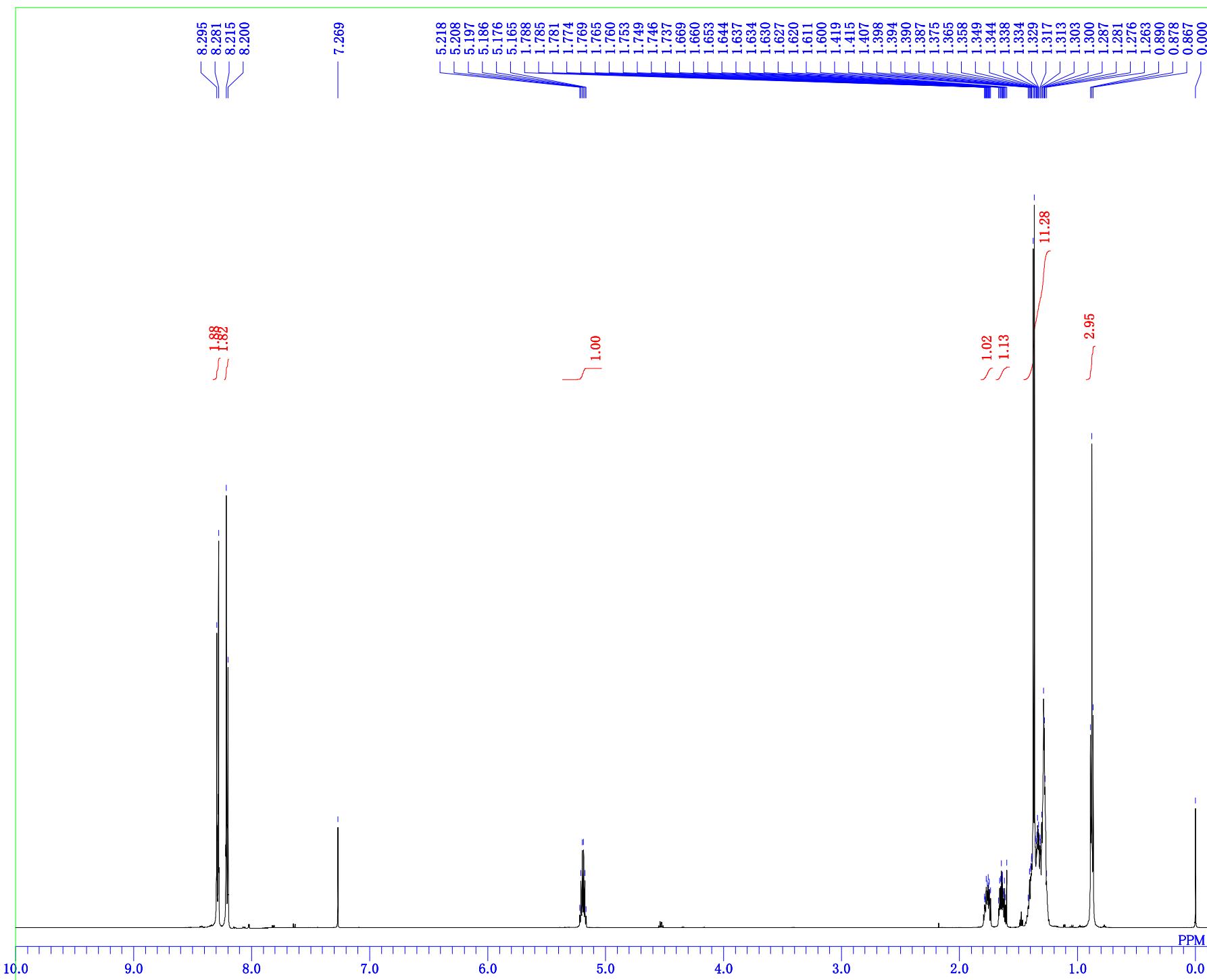


150601

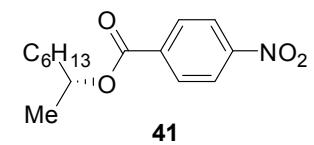


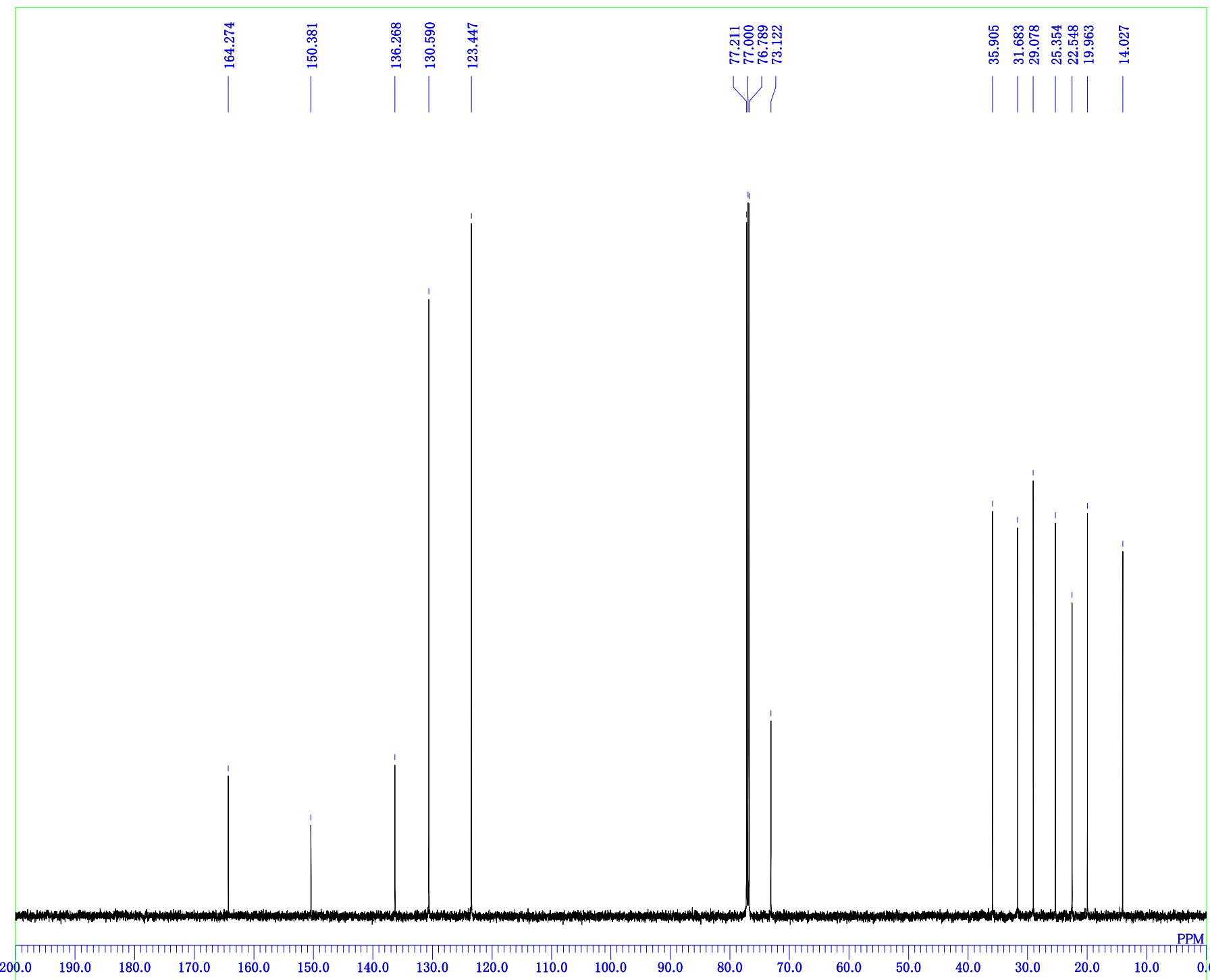
DFILE d1608-gra-13c-1.als
 COMNT 150601
 DATIM 2015-06-01 11:18:24
 13C 13C
 single_pulse_dec
 EXMOD 150.92 MHz
 OBFRQ 8.52 KHz
 OBSET 1.74 Hz
 OBFIN 26214
 POINT 37878.21 Hz
 FREQU 256
 SCANS 0.6921 sec
 ACQTM 1.2000 sec
 PD 3.13 usec
 PW1
 IRNUC 1H
 CTEMP 23.1 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 56



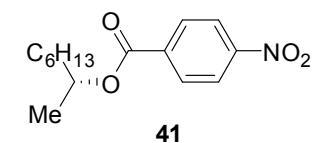


DFILE d1512-gra-1h-1.als
 COMNT 150304
 DATIM 2015-03-04 20:28:49
 1H
 single_pulse.ex2
 EXMOD 600.17 MHz
 OBFRQ 5.30 KHz
 OBSET 5.47 Hz
 OBFIN 26214
 POINT 26214
 FREQU 9008.87 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 5.85 usec
 1H 20.3 c
 CTEMP CDCL₃
 SLVNT 0.00 ppm
 EXREF 0.12 Hz
 RGAIN 36

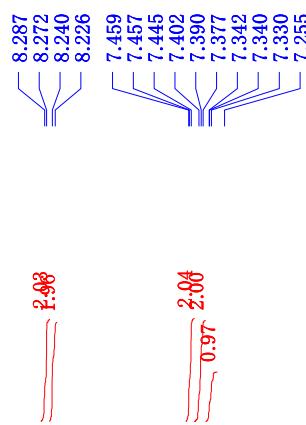




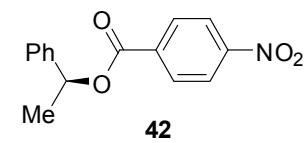
DFILE d1512-gra-13c-1.als
 COMNT 150304
 DATIM 2015-03-04 20:38:37
 13C
 single_pulse_dec
 150.92 MHz
 OBFRQ 8.52 KHz
 OBSET 1.74 Hz
 OBFIN 26214
 POINT 37878.21 Hz
 FREQU 256
 SCANS 0.6921 sec
 ACQTM 1.2000 sec
 PD 2.97 usec
 PW1
 IRNUC 1H
 CTEMP 20.6 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 56



150302

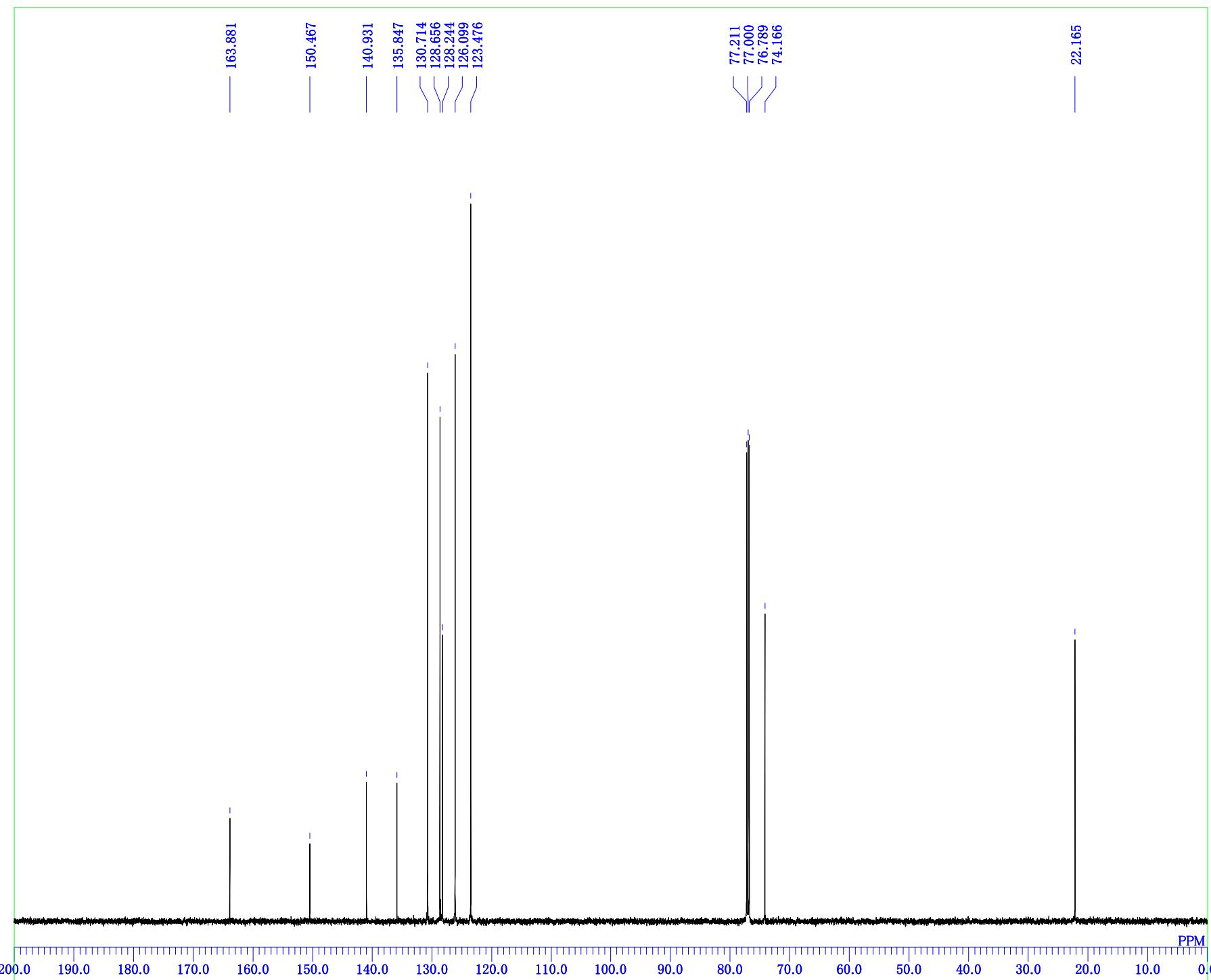


DFILE d1510-gra-1h-1.als
 COMNT 150302
 DATIM 2015-03-02 16:34:50
 OBNUC 1H
 EXMOD single_pulse.ex2
 OBFRQ 600.17 MHz
 OBSET 5.30 KHz
 OBFIN 5.47 Hz
 POINT 26214
 FREQU 9008.87 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 5.85 usec
 IRNUC 1H
 CTEMP 20.7 c
 SLVNT CDCL3
 EXREF 0.00 ppm
 BF 0.12 Hz
 RGAIN 36

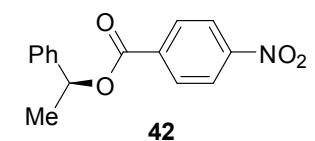


10.0 9.0 8.0 7.0 6.0 5.0 4.0 3.0 2.0 1.0 0.0 PPM

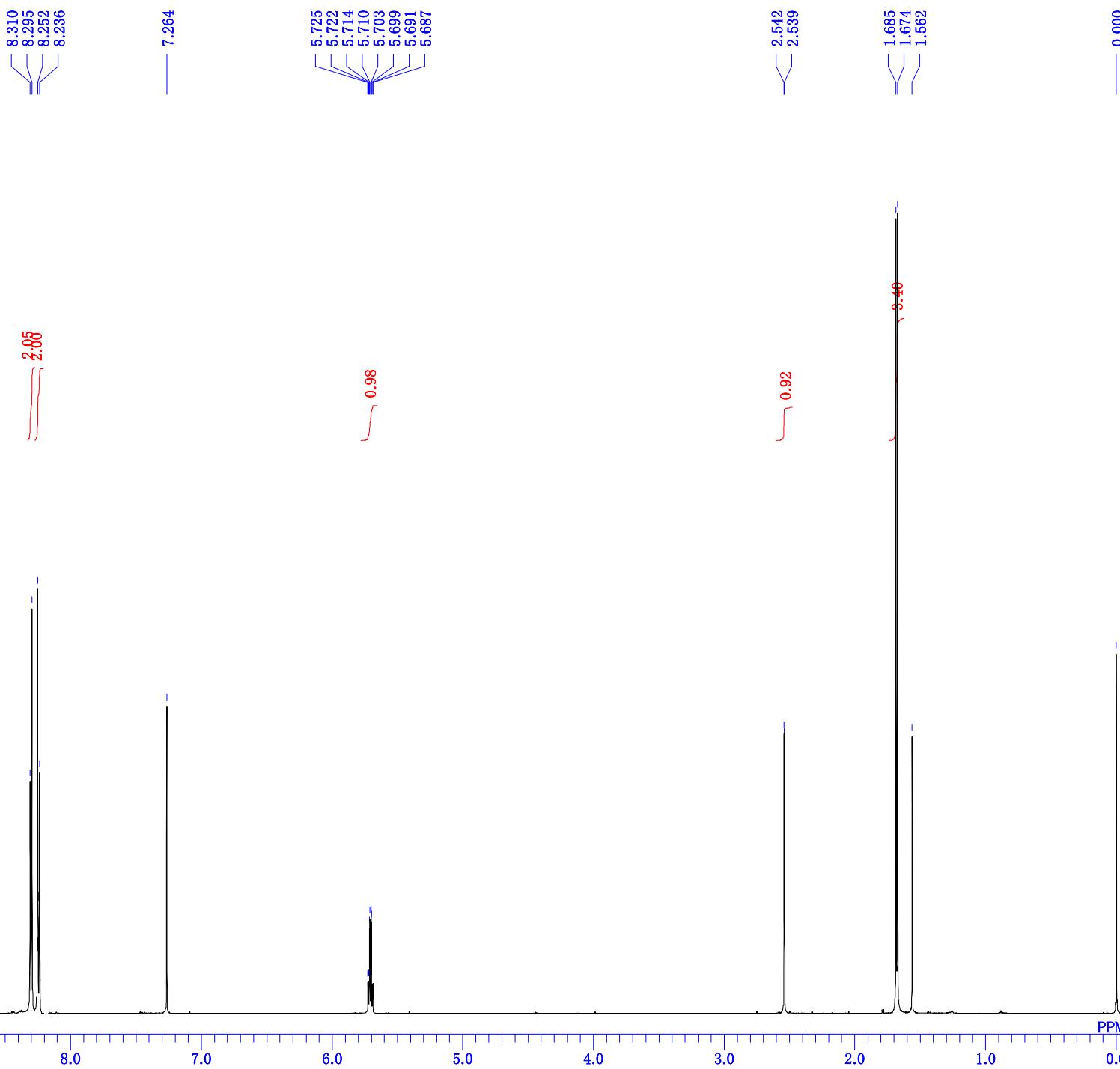
S108



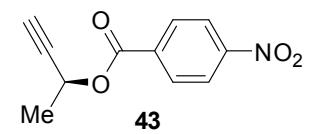
DFILE d1510-gra-13c-1.als
 COMNT 150302
 DATIM 2015-03-02 16:43:25
 13C single_pulse_dec
 EXMOD 150.92 MHz
 OBFRQ 8.52 KHz
 OBSET 1.74 Hz
 OBFIN 26214
 POINT 37878.21 Hz
 FREQU 256
 SCANS 0.6921 sec
 ACQTM 1.2000 sec
 PD 2.97 usec
 PW1 1H 21.3 c
 IRNUC CDCL3
 CTEMP 77.00 ppm
 SLVNT 1.20 Hz
 EXREF 58
 RGAIN



150421

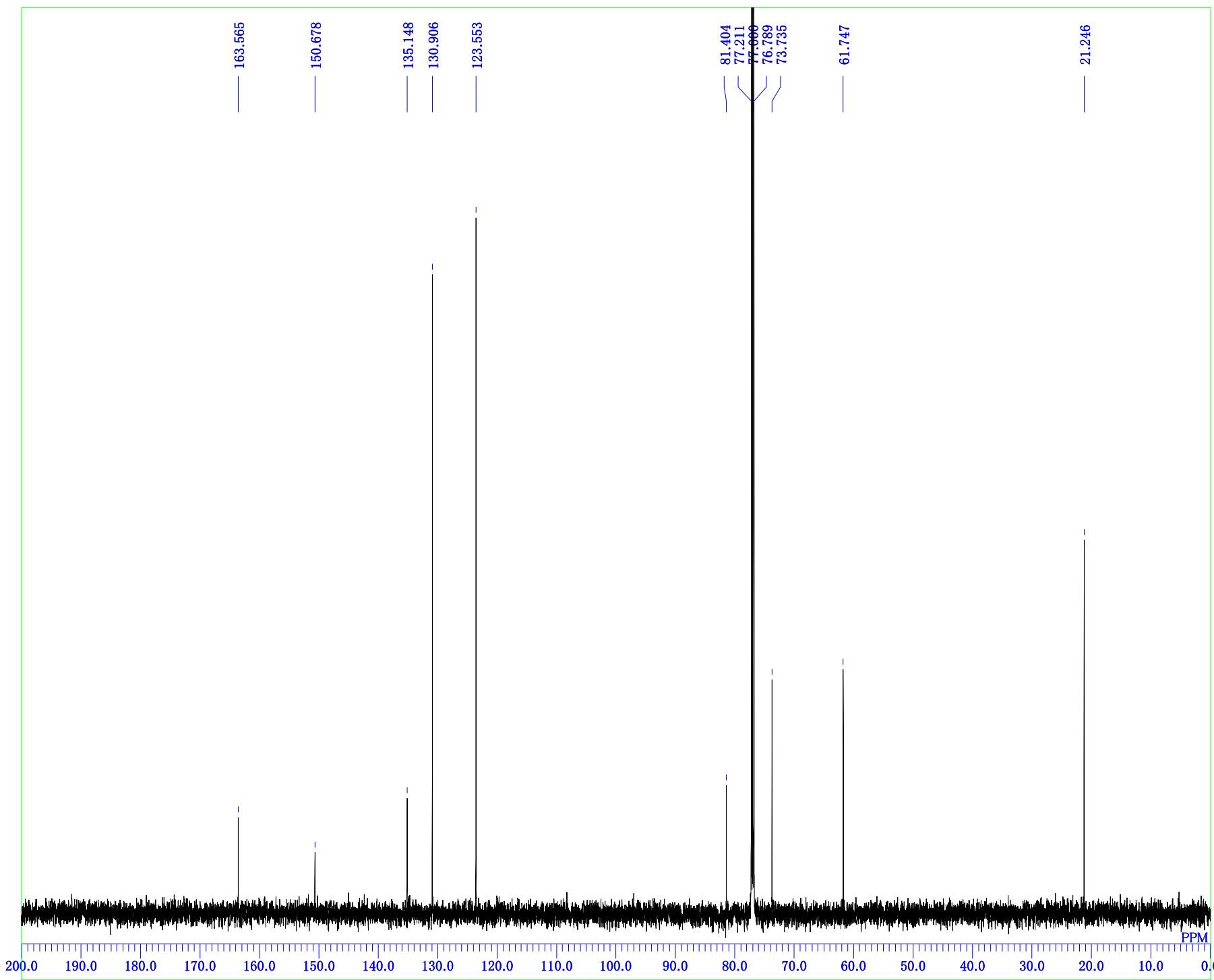


DFILE d1530-gra-1h-1.als
 COMNT 150421
 DATIM 2015-04-21 16:42:57
 1H
 single_pulse.ex2
 EXMOD 600.17 MHz
 OBFRQ 5.30 KHz
 OBSET 5.47 Hz
 OBFIN 5.47 Hz
 POINT 26214
 FREQU 9008.87 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 7.30 usec
 IRNUC 1H
 CTEMP 21.2 c
 SLVNT CDCL₃
 EXREF 0.00 ppm
 BF 0.12 Hz
 RGAIN 50

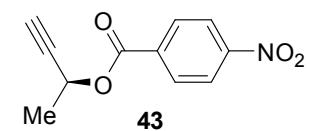


S110

150421

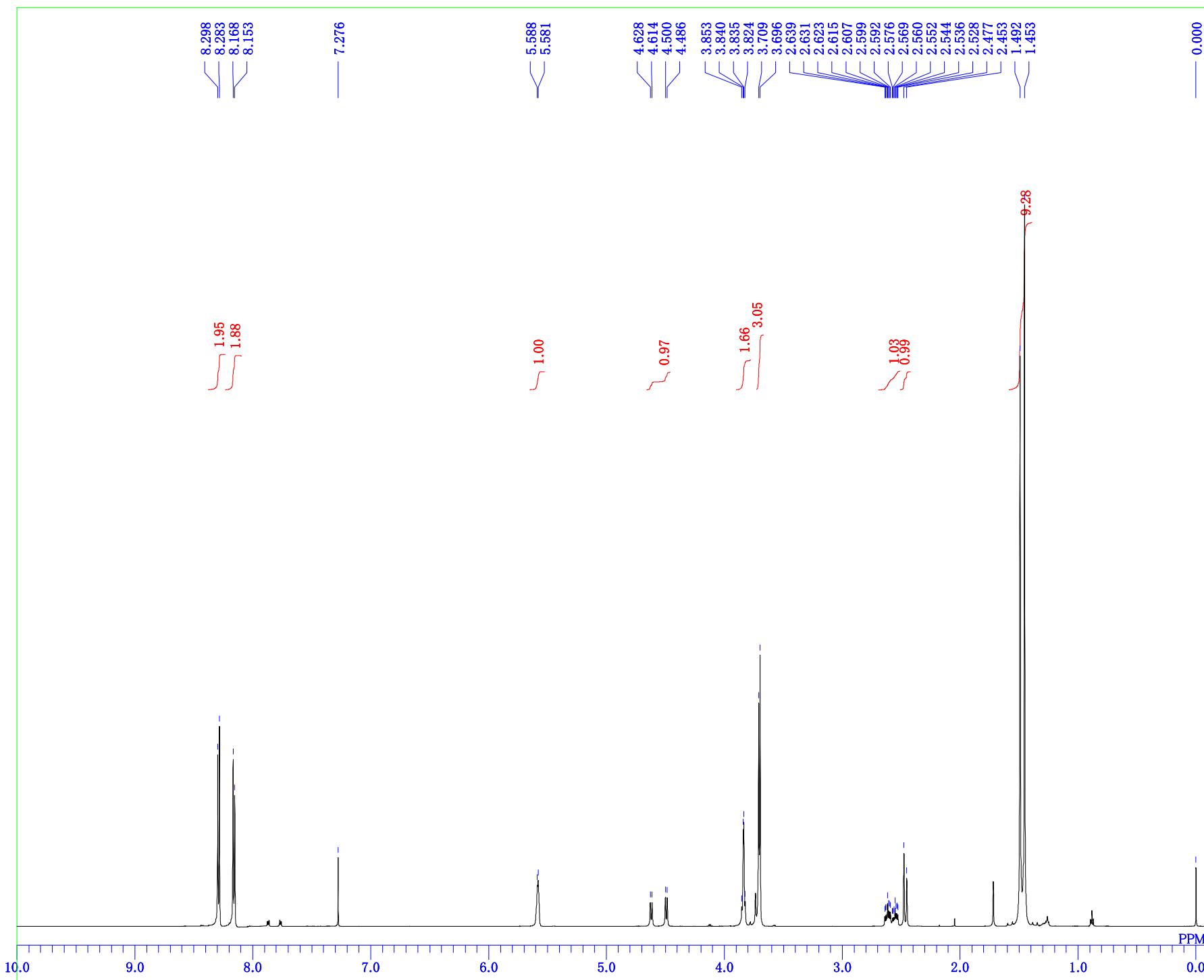


DFILE d1530-gra-13c-1.als
 COMNT 150421
 DATIM 2015-04-21 16:54:40
 OBNUC 13C
 EXMOD single_pulse_dec
 OBFRQ 150.92 MHz
 OBSET 8.52 KHz
 OBFIN 1.74 Hz
 POINT 26214
 FREQU 37878.21 Hz
 SCANS 256
 ACQTM 0.6921 sec
 PD 1.2000 sec
 PW1 3.13 usec
 IRNUC 1H
 CTEMP 21.7 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 54

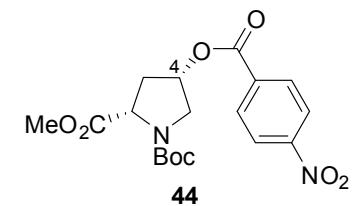


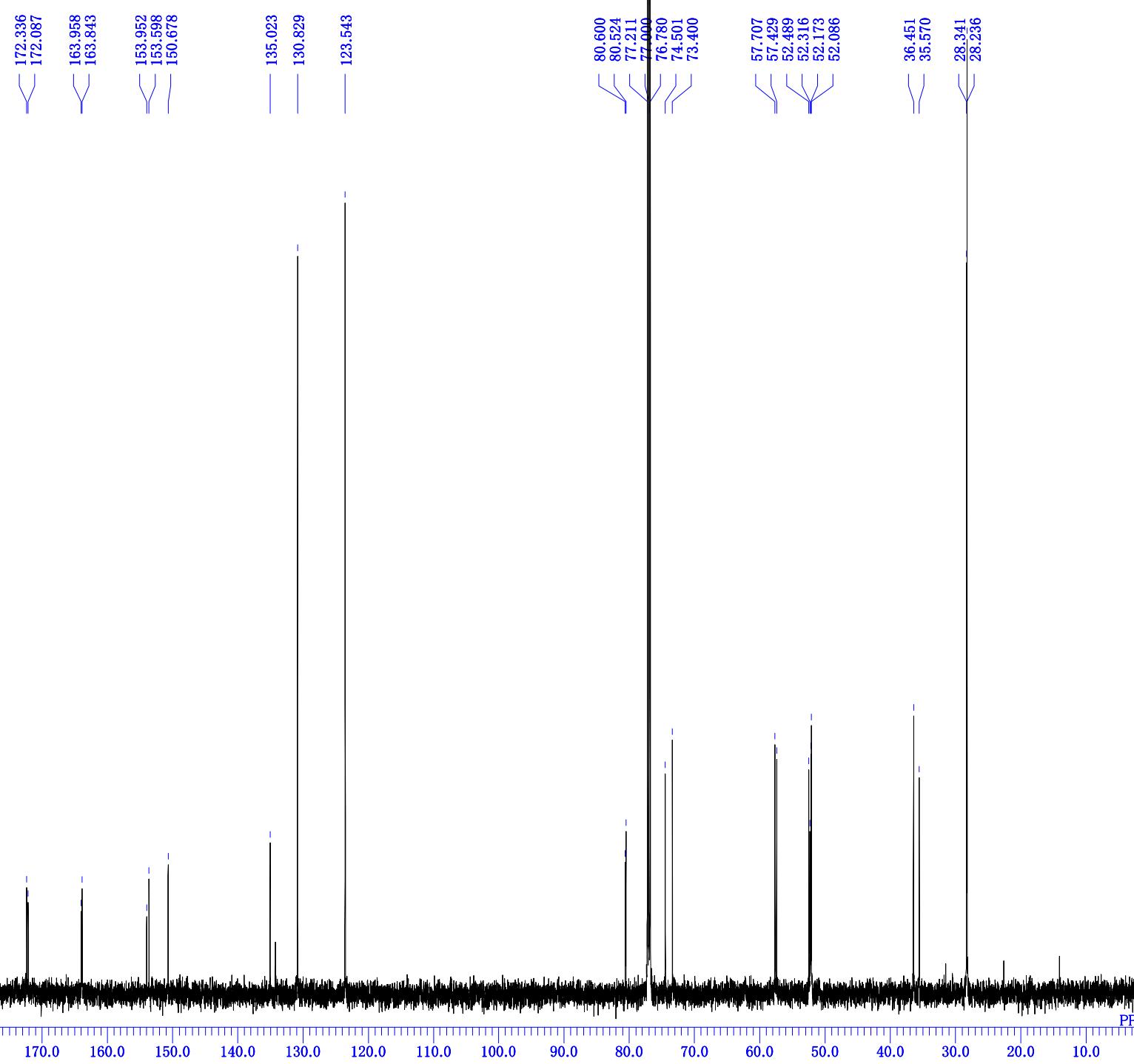
S111

150429

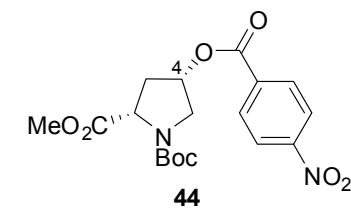


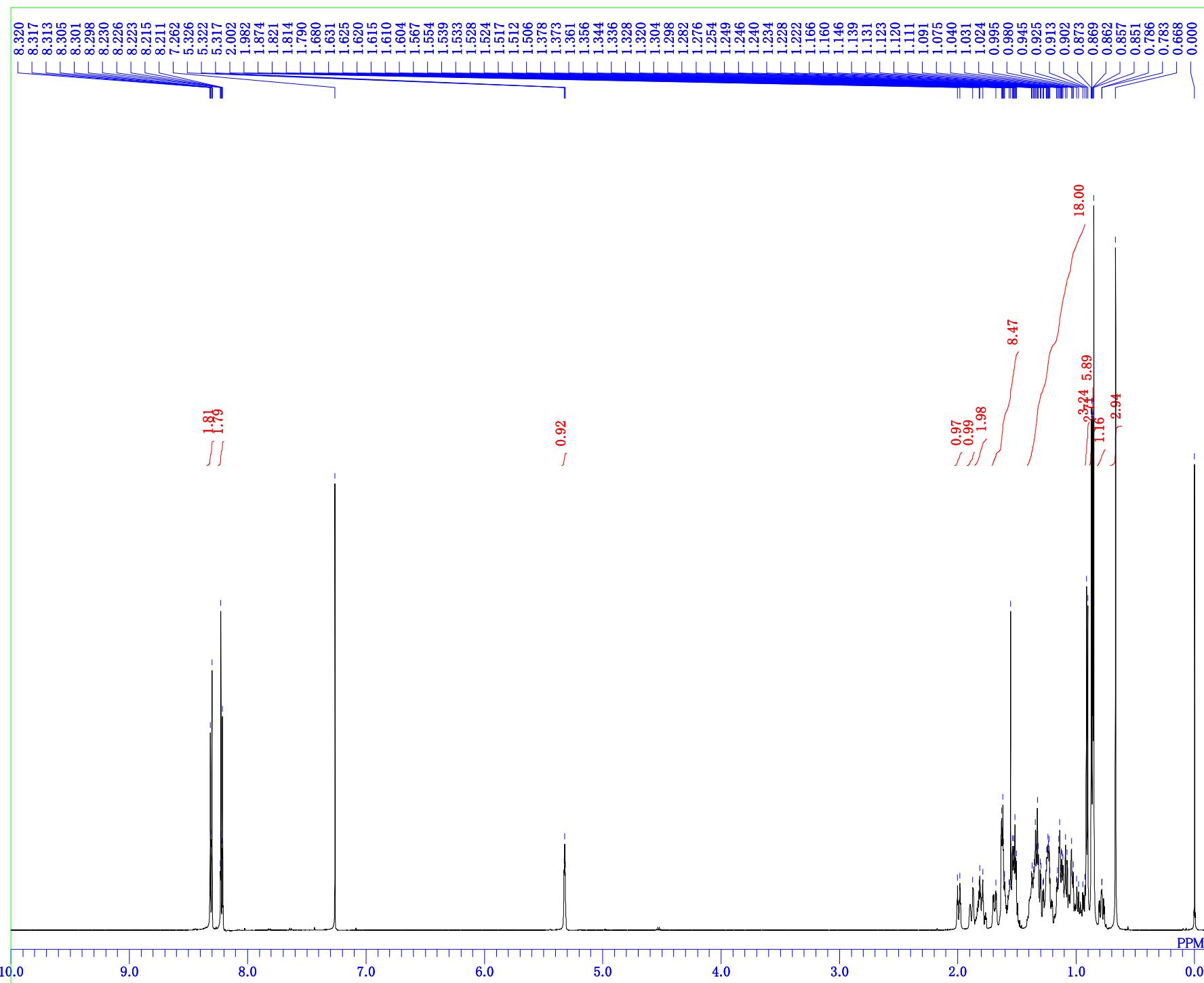
DFILE d1561-gra-1h-1.als
 COMNT 150429
 DATIM 2015-04-29 18:12:44
 1H
 single_pulse.ex2
 EXMOD 600.17 MHz
 OBFRQ 5.30 KHz
 OBSET 5.47 Hz
 OBFIN 26214
 POINT 26214
 FREQU 9008.87 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 7.30 usec
 1H 21.8 c
 CTEMP CDCL3
 SLVNT 0.00 ppm
 EXREF 0.12 Hz
 RGAIN 38



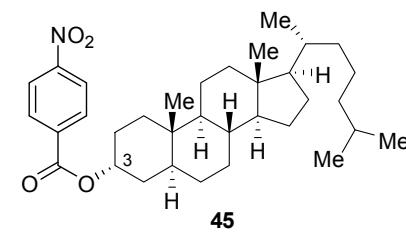


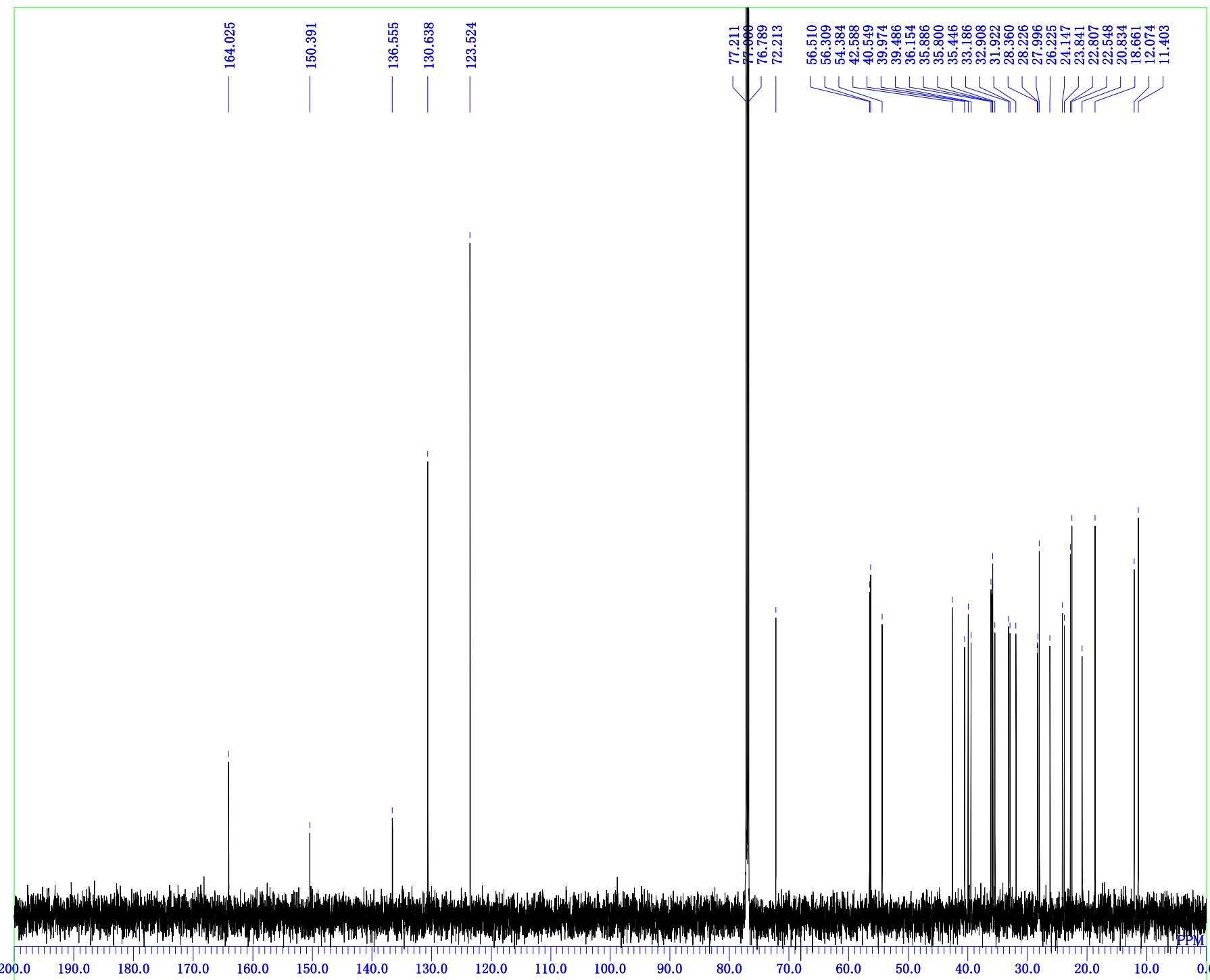
d1561-gra-13c-1.als
150429
2015-04-29 18:22:09
¹³C
single_pulse_dec
150.92 MHz
8.52 KHz
1.74 Hz
26214
37878.21 Hz
256
0.6921 sec
ACQTM
PD
1.2000 sec
PW1
3.13 usec
IRNUC
CTEMP
SLVNT
EXREF
BF
RGAIN
1H
22.4 c
CDCL3
77.00 ppm
1.20 Hz
56



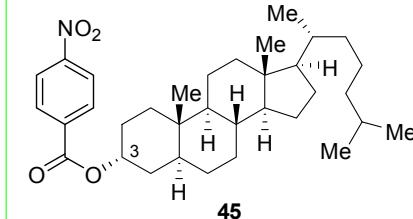


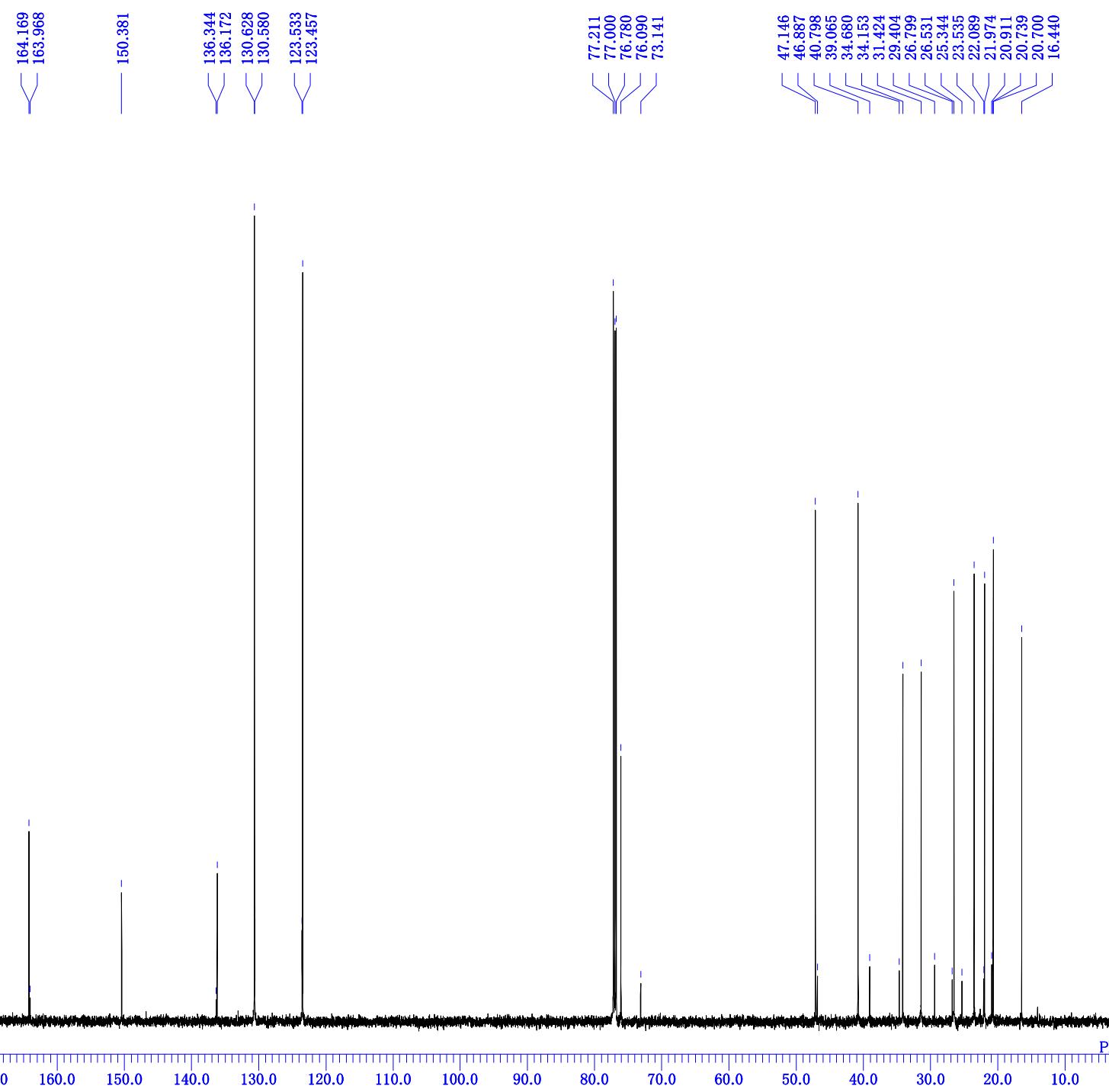
DFILE d1534-gra-1h-1.als
 COMNT 150418
 DATIM 2015-04-18 16:04:57
 1H
 single_pulse.ex2
 600.17 MHz
 5.30 KHz
 5.47 Hz
 26214
 9008.87 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 7.30 usec
 1H 21.1 c
 CDCL3 0.00 ppm
 BF 0.12 Hz
 RGAIN 40



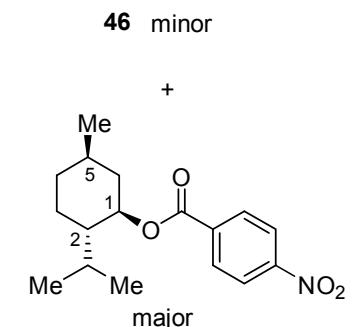
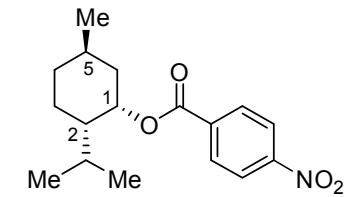


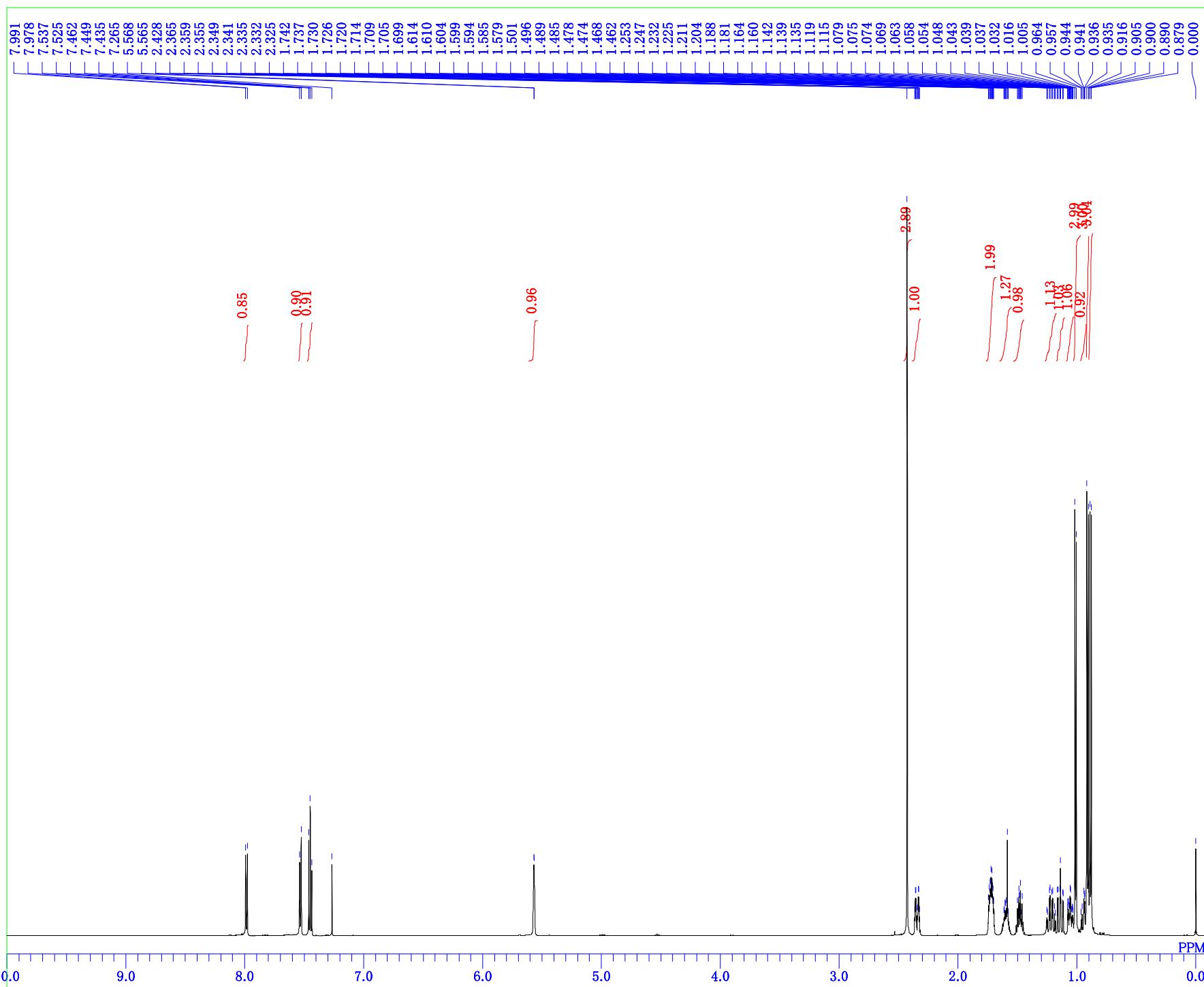
DFILE d1534-gra-13c-1.als
COMNT 150418
DATIM 2015-04-18 16:13:31
13C single_pulse_dec
EXMOD 150.92 MHz
OBFRQ 8.52 KHz
OBSET 1.74 Hz
OBFIN 26214
POINT 37878.21 Hz
FREQU 256
SCANS 0.6921 sec
ACQTM PD 1.2000 sec
PW1 3.13 usec
IRNUC 1H 21.7 c
CTEMP CDCL3 77.00 ppm
SLVNT 1.20 Hz
EXREF 54
BF RGAIN



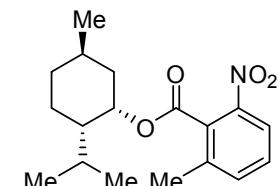


DFILE d1511-gra-13c-1.als
 COMNT 150304
 DATIM 2015-03-04 20:55:56
 OBNUC ¹³C
 EXMOD single_pulse_dec
 OBFRQ 150.92 MHz
 OBSET 8.52 KHz
 OBFIN 1.74 Hz
 POINT 26214
 FREQU 37878.21 Hz
 SCANS 256
 ACQTM 0.6921 sec
 PD 1.2000 sec
 PW1 2.97 usec
 IRNUC 1H
 CTEMP 20.5 c
 SLVNT CDCL₃
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 56

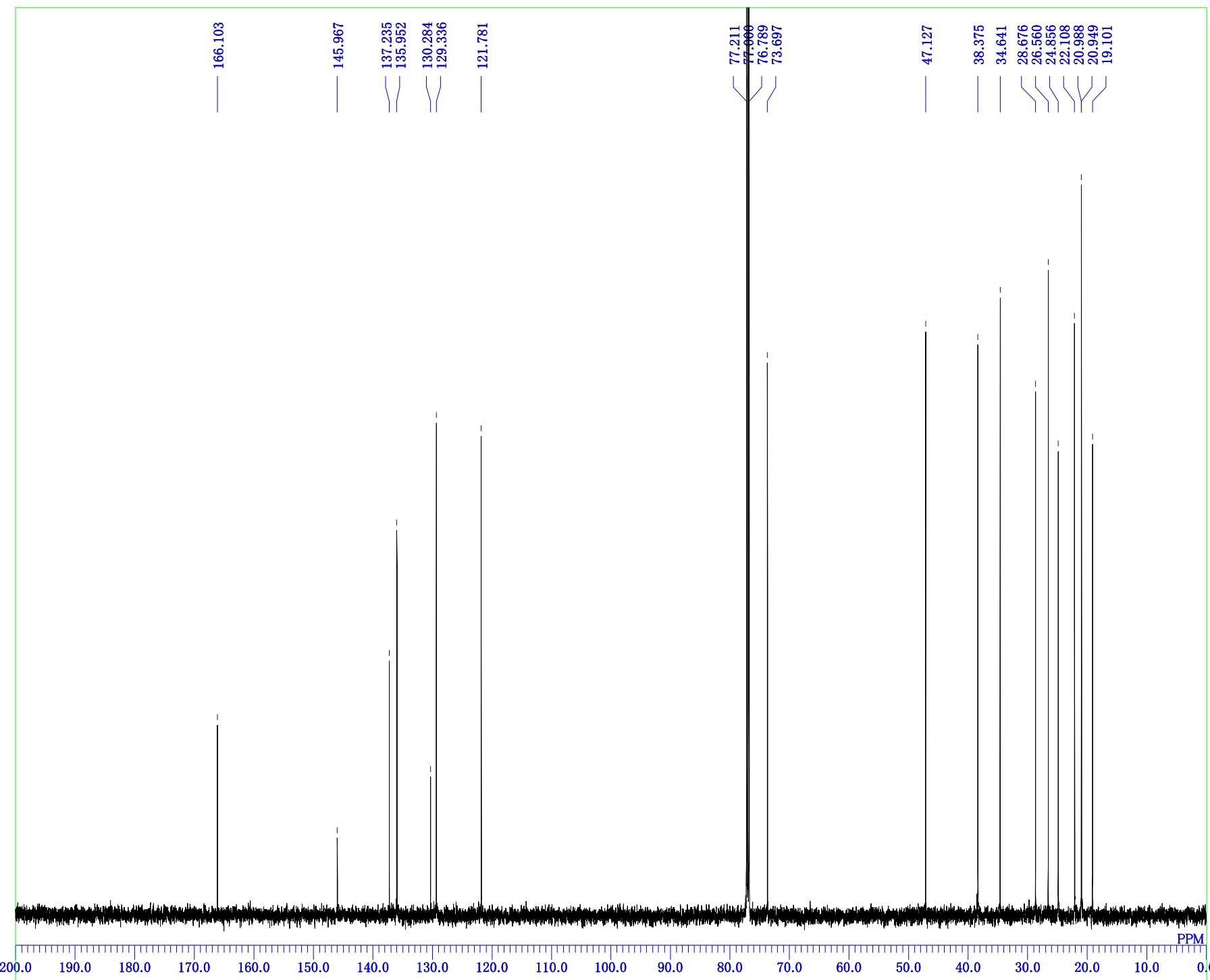




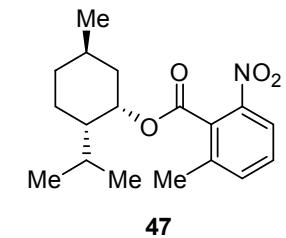
DFILE d1580-gra-1h-1.als
 COMNT 150519
 DATIM 2015-05-19 18:27:43
 1H
 single_pulse.ex2
 EXMOD 600.17 MHz
 OBFRQ 5.30 KHz
 OBSET 5.47 Hz
 OBFIN 26214
 POINT 9008.87 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 7.30 usec
 IRNUC 1H
 CTEMP 21.4 c
 SLVNT CDCL3
 EXREF 0.00 ppm
 BF 0.12 Hz
 RGAIN 34



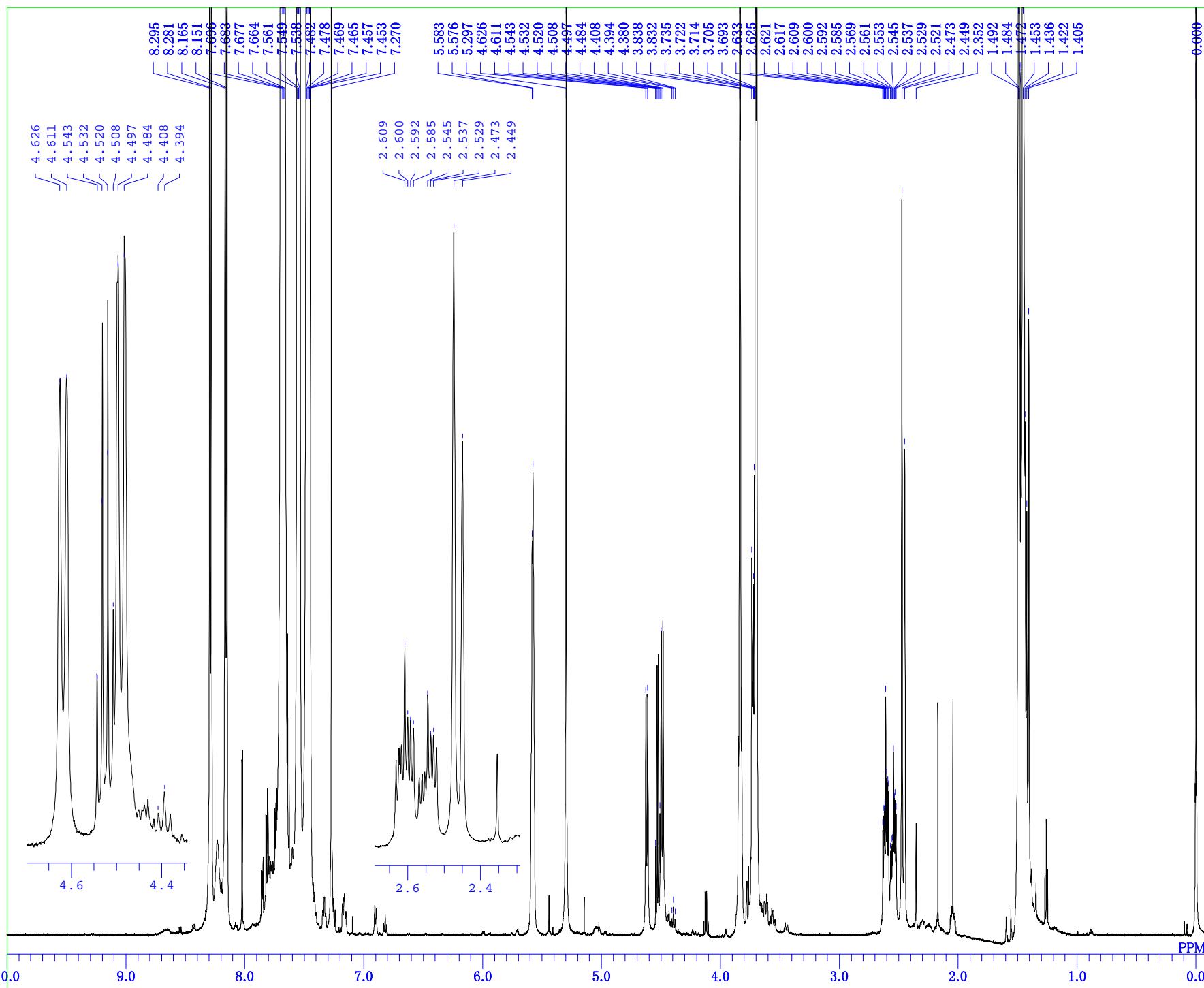
47



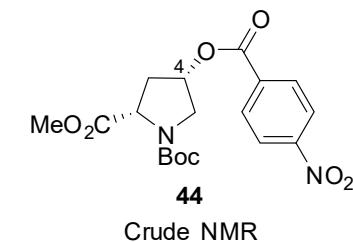
DFILE d1580-gra-13c-1.als
 COMNT 150519
 DATIM 2015-05-19 18:36:26
 13C single_pulse_dec
 EXMOD 150.92 MHz
 OBFRQ 8.52 KHz
 OBSET 1.74 Hz
 OBFIN 26214
 POINT 37878.21 Hz
 FREQU 256
 SCANS 0.6921 sec
 ACQTM 1.2000 sec
 PD 3.13 usec
 PW1
 IRNUC 1H
 CTEMP 22.0 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 54



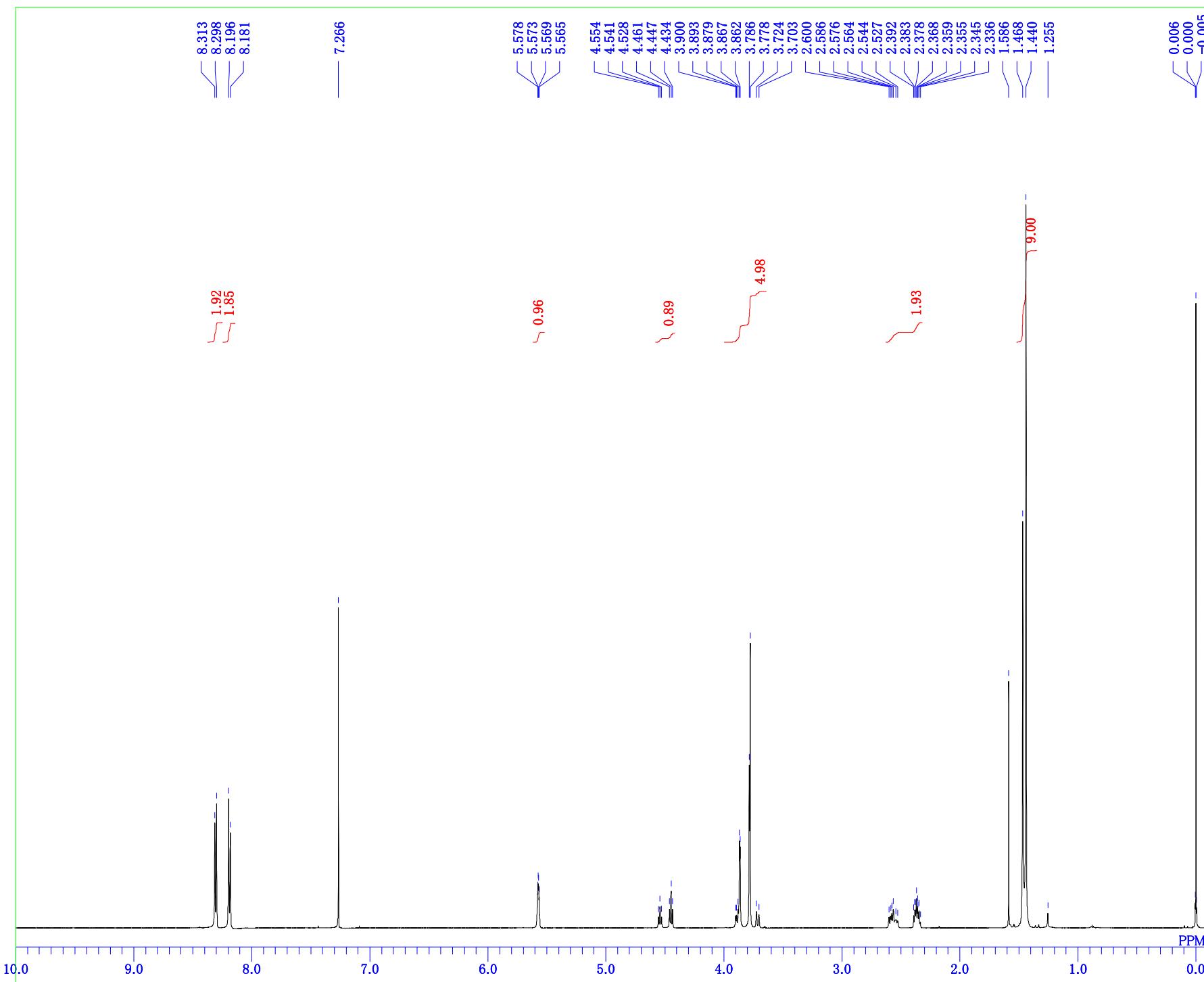
150423



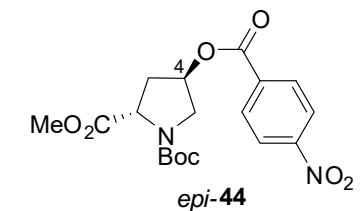
DFILE d1561-crude-diastereo-1.jdf
 COMNT 150423
 DATIM 2015-04-23 15:29:37
 1H
 single_pulse.ex2
 600.17 MHz
 5.30 KHz
 5.47 Hz
 32768
 11261.26 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 7.30 usec
 1H 21.4 c
 CDCL3 0.00 ppm
 IRNUC 0.12 Hz
 CTEMP 38
 OBSET
 OBFIN
 POINT
 FREQU
 SCANS
 ACQTM
 PD
 PW1
 IRNUC
 CTEMP
 SLVNT
 EXREF
 BF
 RGAIN



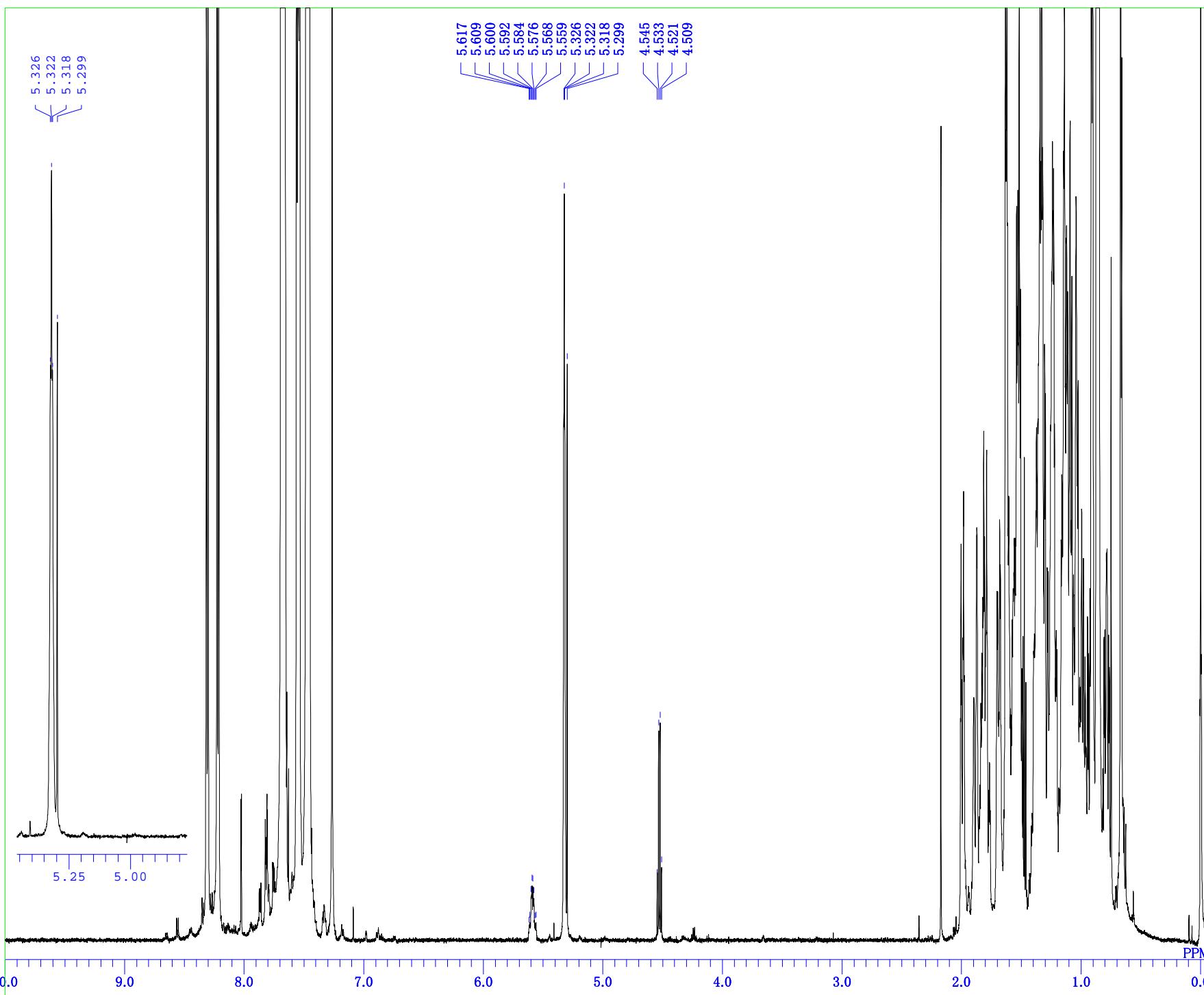
S120



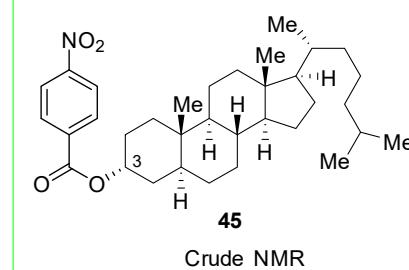
DFILE d1564-a-1.als
 COMNT 151211
 DATIM 2015-12-11 13:35:47
 1H
 single_pulse.ex2
 600.17 MHz
 5.30 KHz
 5.47 Hz
 26214
 9008.87 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 7.30 usec
 1H 20.3 c
 CDCL3 0.00 ppm
 0.12 Hz
 RGAIN 46



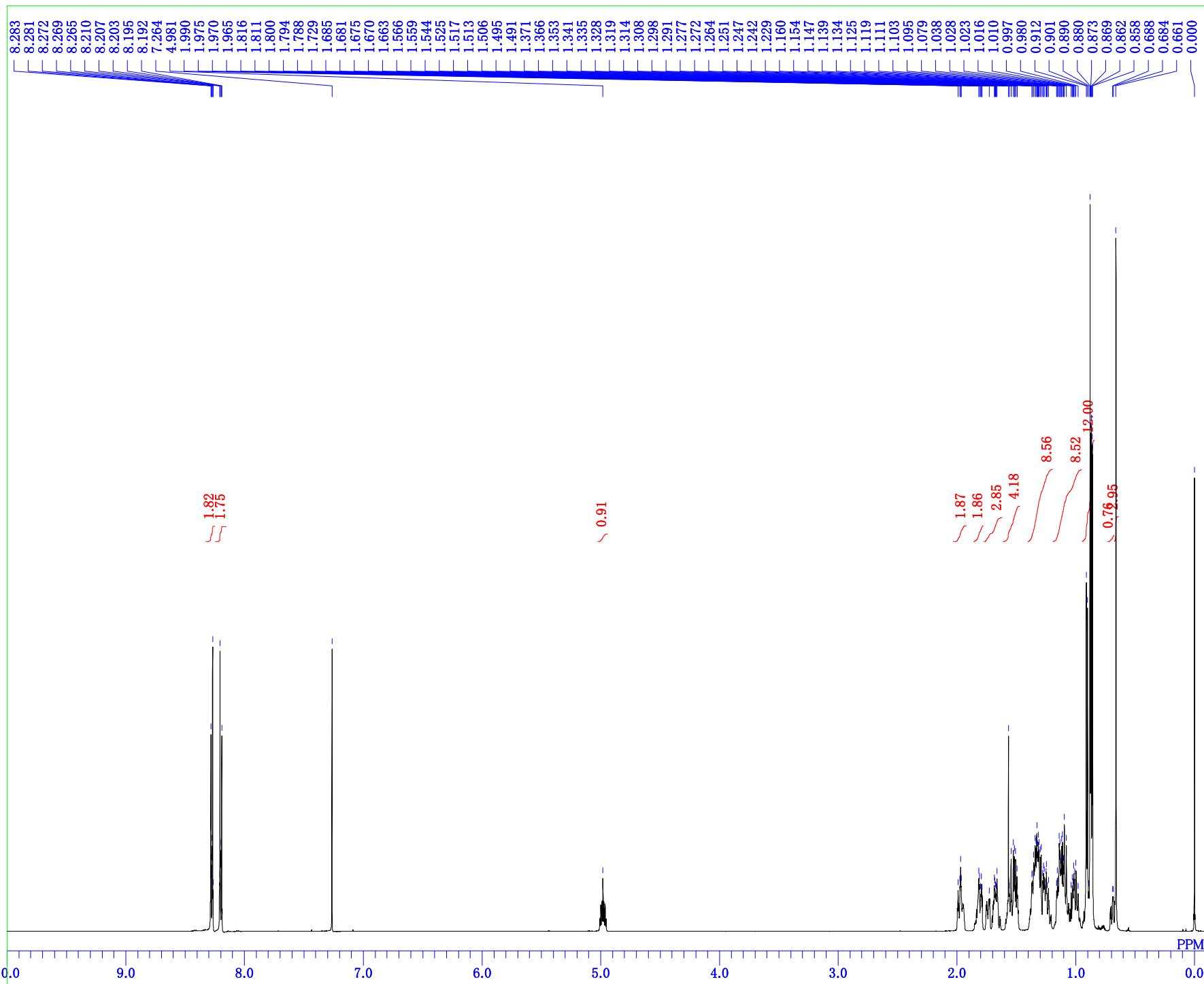
150402



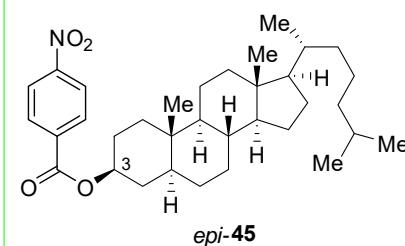
DFILE d1534-crude-diastereo-1.als
COMNT 150402
DATIM 2015-04-02 10:17:19
1H single_pulse.ex2
EXMOD 600.17 MHz
OBFRQ 5.30 KHz
OBSET 5.47 Hz
OBFIN 26214
POINT 26214
FREQU 9008.87 Hz
SCANS 32
ACQTM 2.9098 sec
PD 2.0000 sec
PW1 7.30 usec
IRNUC 1H
CTEMP 20.7 c
SLVNT CDCL₃
EXREF 0.00 ppm
BF 0.12 Hz
RGAIN 40



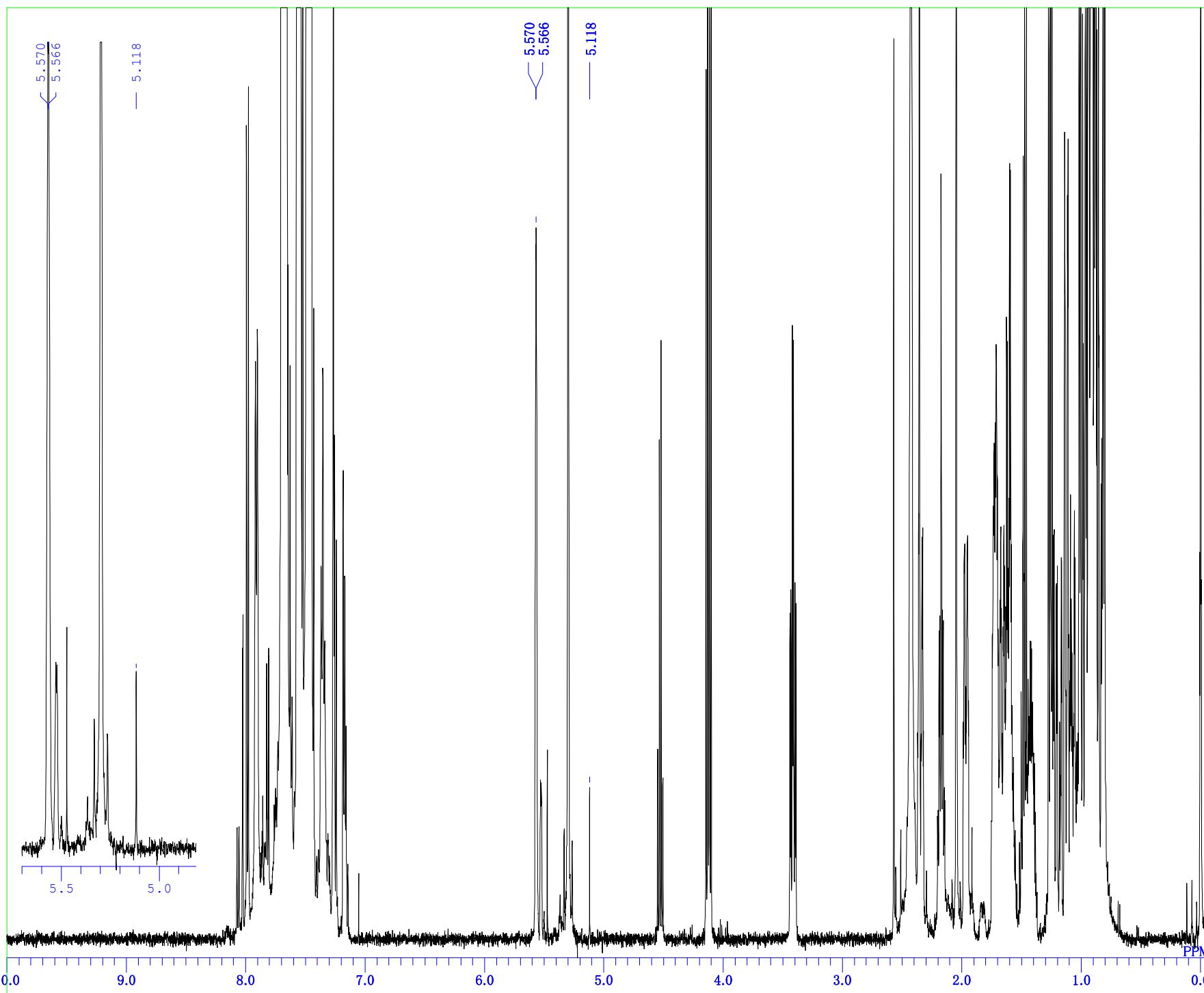
151204



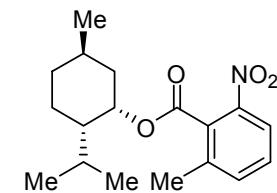
DFILE d1550-151204.jdf
 COMNT 151204
 DATIM 2015-12-04 15:45:47
 1H
 single_pulse.ex2
 600.17 MHz
 5.30 KHz
 5.47 Hz
 32768
 11261.26 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 6.90 usec
 1H 19.5 c
 CDCL3 0.00 ppm
 EXREF 0.12 Hz
 BF 38
 RGAIN



150511

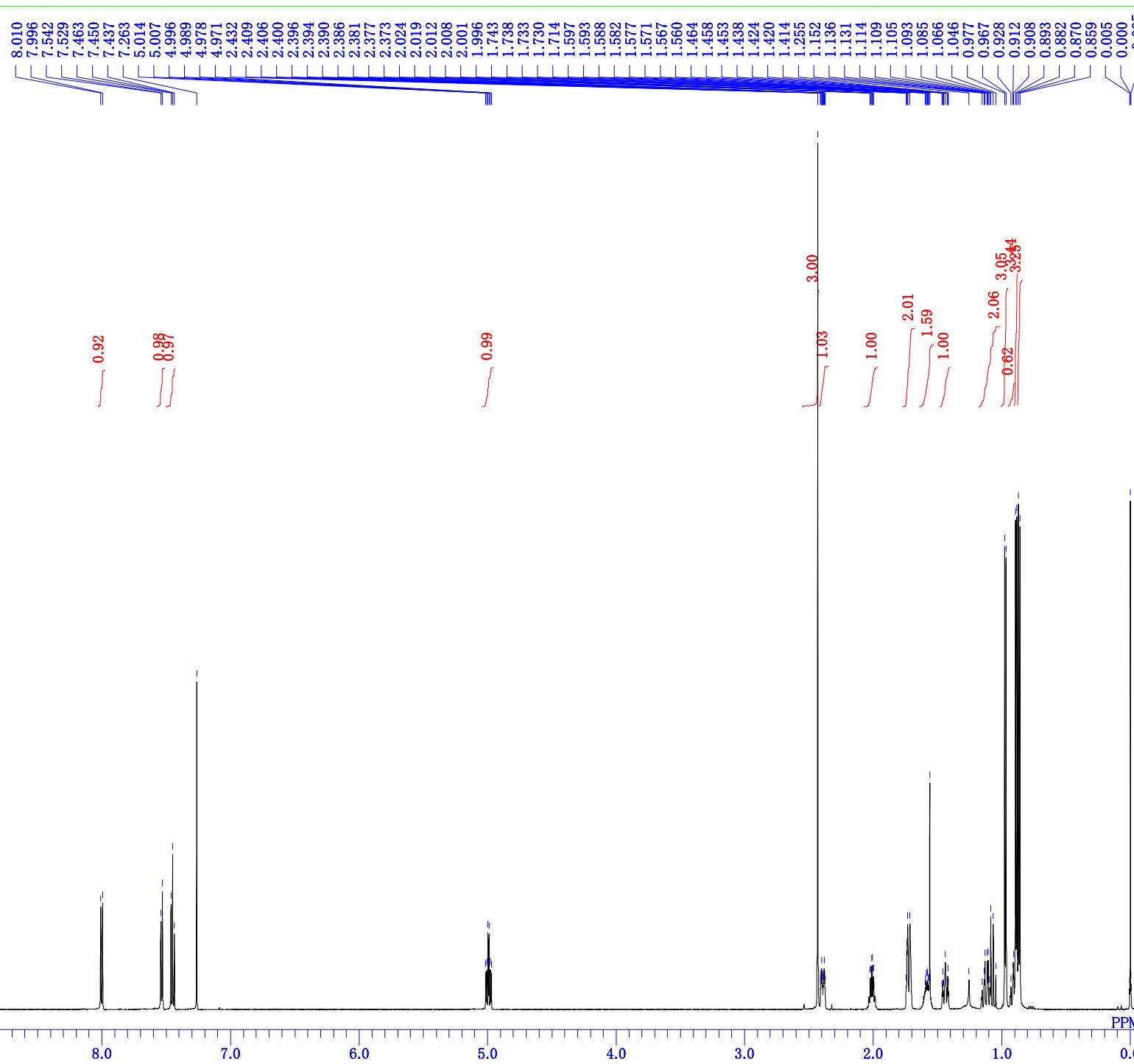


DFILE d1580-crude-1.als
 COMNT 150511
 DATIM 2015-05-11 13:53:49
 1H single_pulse.ex2
 EXMOD 500.16 MHz
 OBFREQ 2.41 KHz
 OBSET 6.01 Hz
 OBFIN 13107
 POINT 7507.39 Hz
 FREQU 1.7459 sec
 SCANS 8
 ACQTM 2.0000 sec
 PD 5.80 usec
 PW1 1H 20.4 c
 IRNUC CDCL3
 CTEMP 0.00 ppm
 SLVNT 0.12 Hz
 EXREF 42
 RGAIN

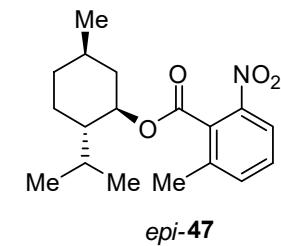


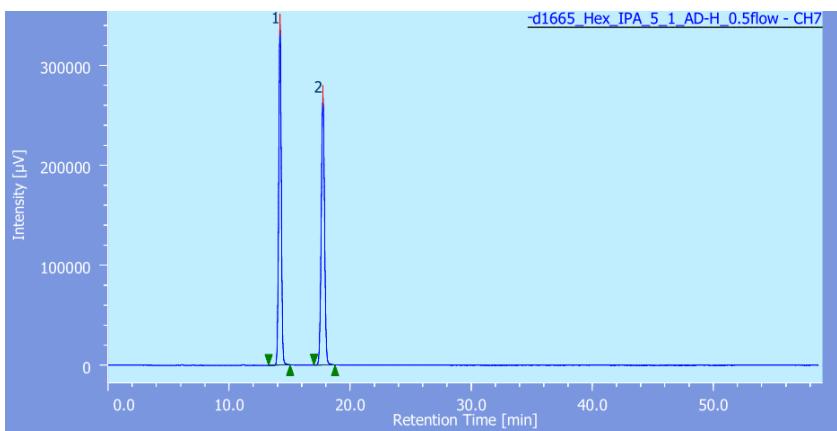
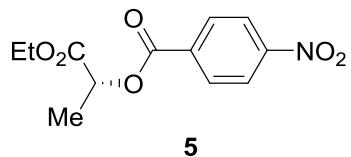
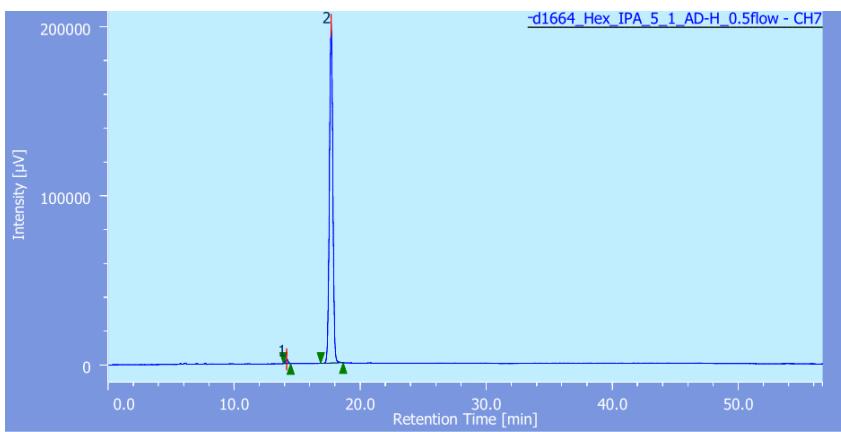
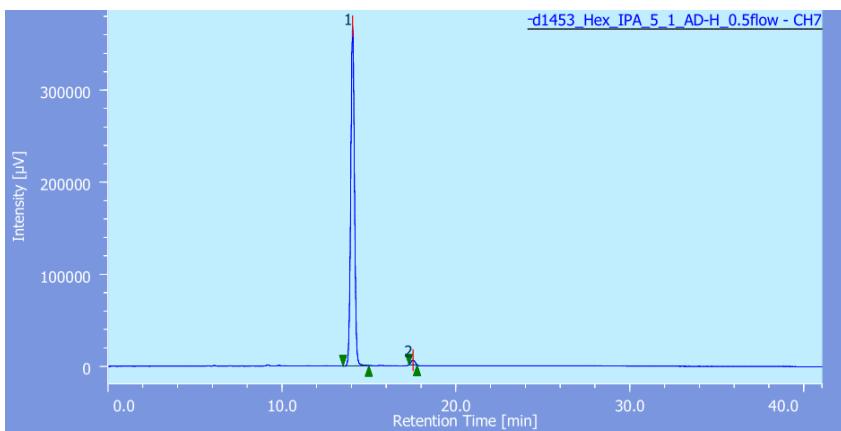
47
Crude NMR

S124



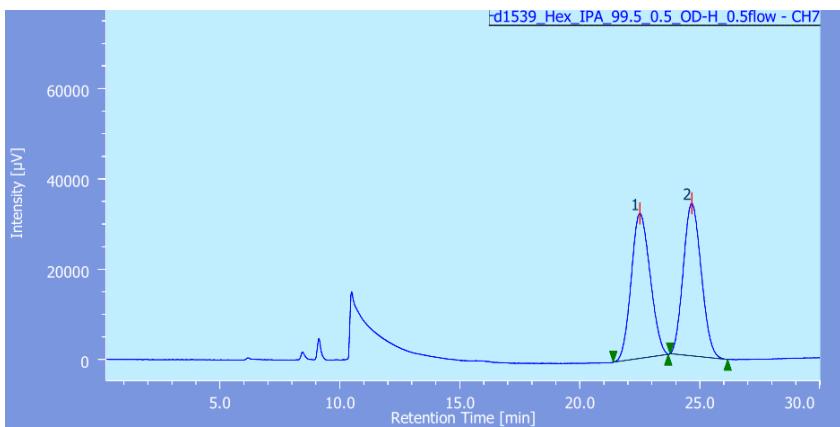
DFILE d1656-151204.jdf
 COMNT 151204
 DATIM 2015-12-04 16:02:53
 1H
 single_pulse.ex2
 600.17 MHz
 5.30 KHz
 5.47 Hz
 32768
 11261.26 Hz
 SCANS 32
 ACQTM 2.9098 sec
 PD 2.0000 sec
 PW1 6.90 usec
 1H
 IRNUC 19.7 c
 CTEMP CDCL3
 SLVNT 0.00 ppm
 EXREF 0.12 Hz
 BF 42
 RGAIN



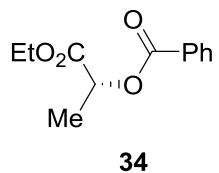
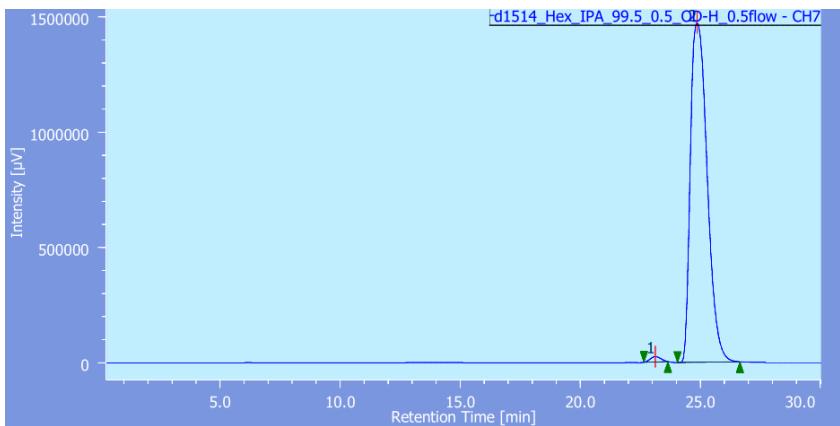
5-racemic**5-retention****5-sample**

racemic mixture	tR (min)	area	area%
peak1	14.200	5087252	49.748
peak2	17.750	5138742	50.252
retention	tR (min)	area	area%
peak1	14.167	34569	0.904
peak2	17.700	3788748	99.096
sample	tR (min)	area	area%
peak1	14.058	5475109	98.611
peak2	17.533	77132	1.389

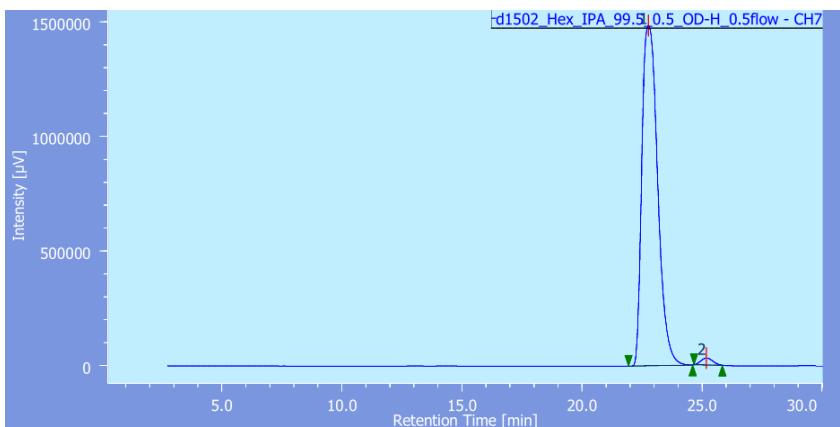
34-racemic



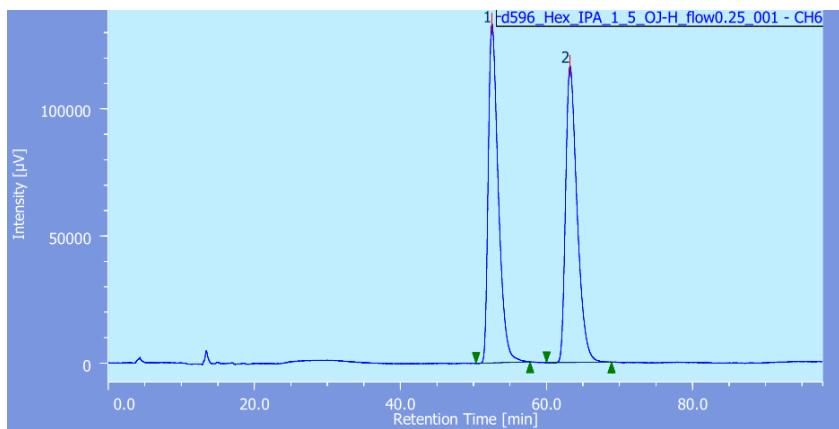
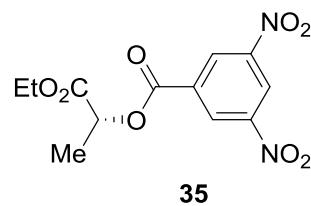
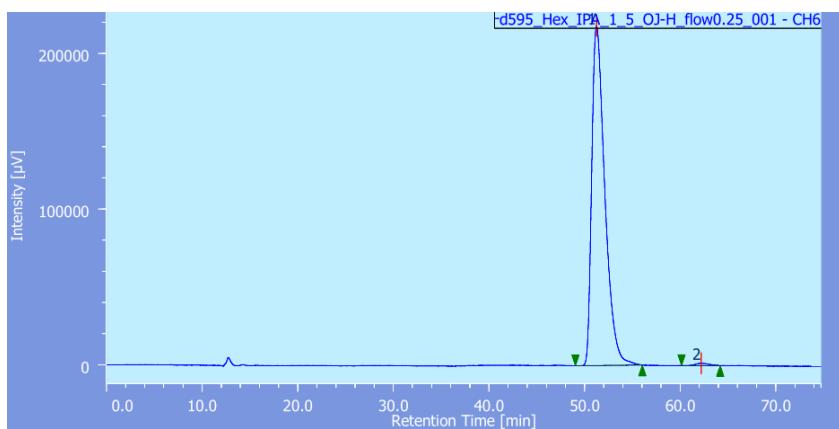
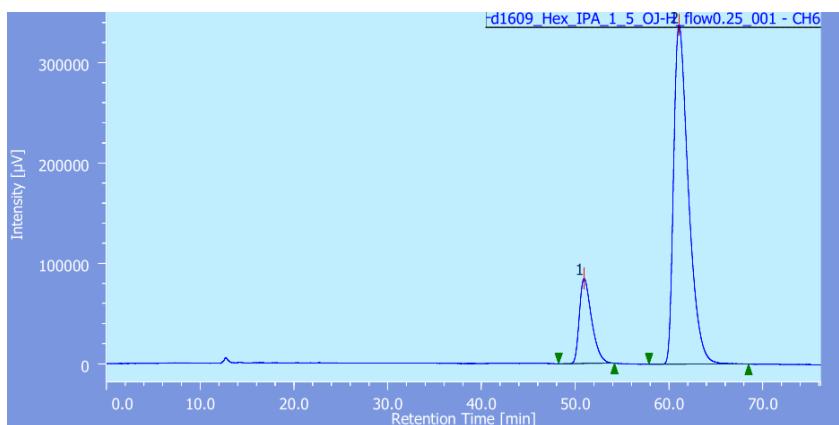
34-retention



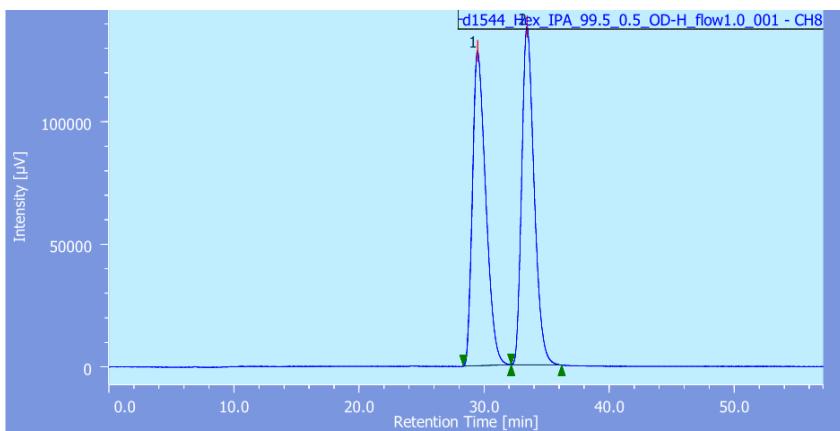
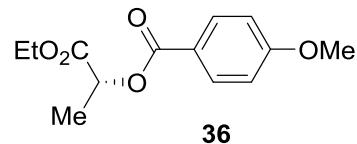
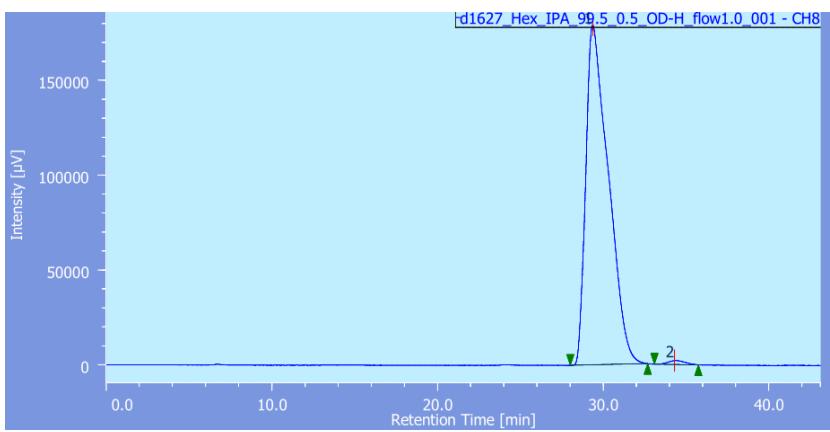
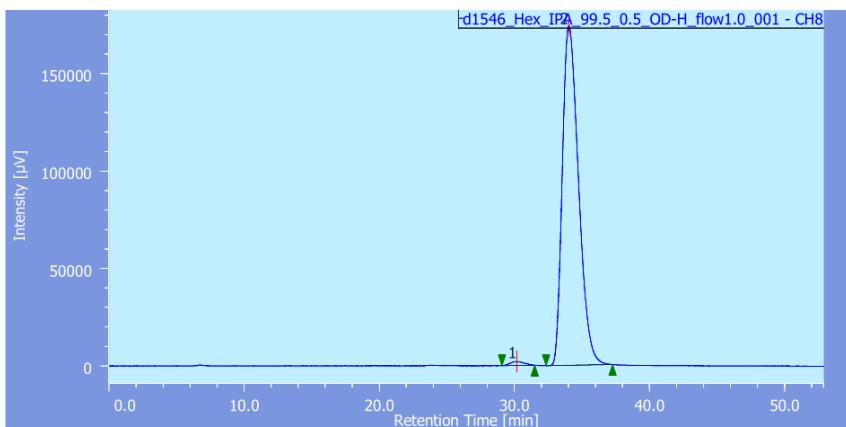
34-sample



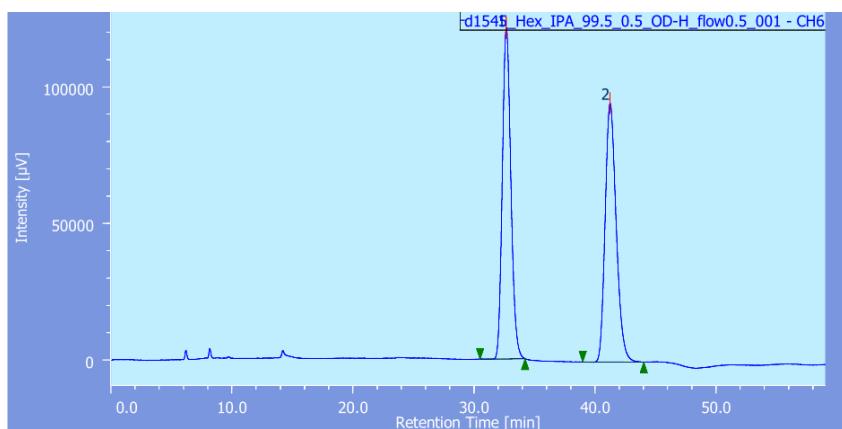
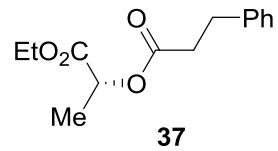
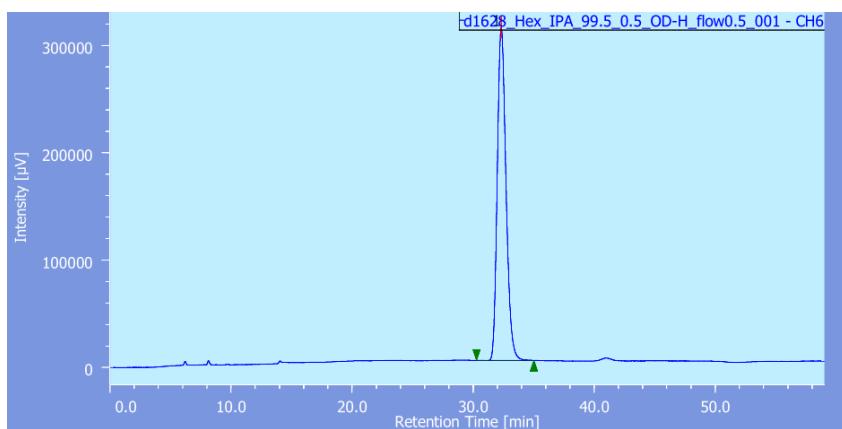
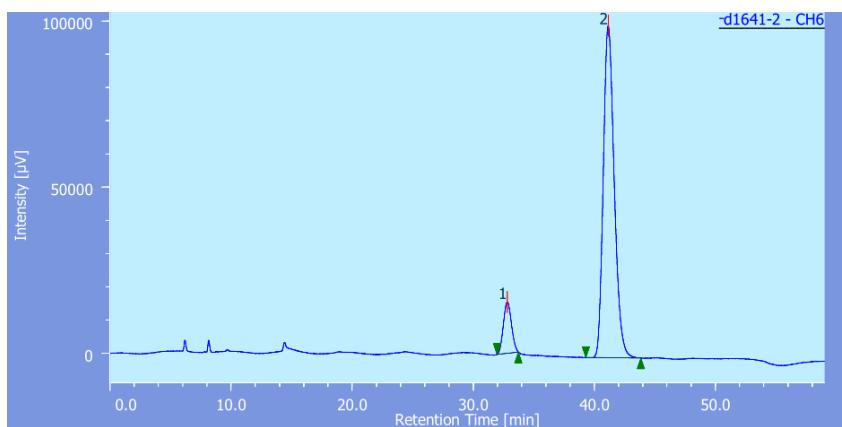
racemic mixture	tR (min)	area	area%
peak1	22.500	1762124	50.057
peak2	24.658	1758100	49.943
retention	tR (min)	area	area%
peak1	23.125	706735	0.980
peak2	24.858	71439775	99.020
sample	tR (min)	area	area%
peak1	22.758	67663135	98.502
peak2	25.175	1029277	1.498

35-racemic**35-retention****35-sample**

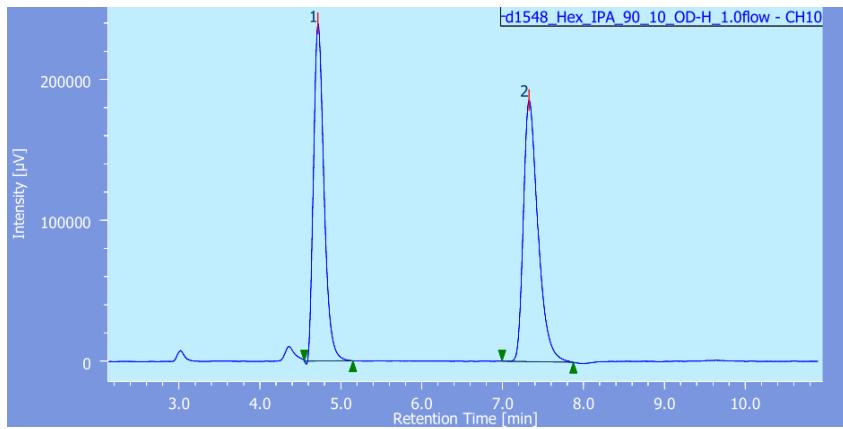
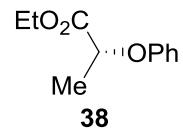
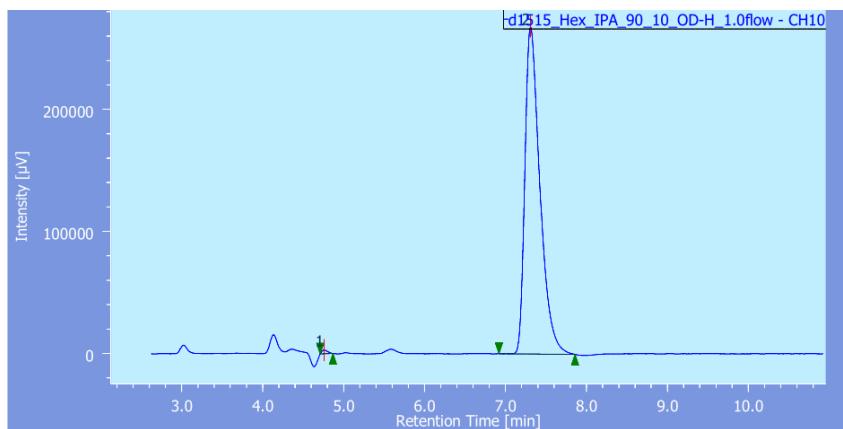
racemic mixture	tR (min)	area	area%
peak1	52.542	12747201	50.116
peak2	63.217	12688155	49.884
retention	tR (min)	area	area%
peak1	51.242	20707713	99.315
peak2	62.192	142775	0.685
sample	tR (min)	area	area%
peak1	50.950	7664177	17.167
peak2	61.067	36980193	82.833

36-racemic**36-retention****36-sample**

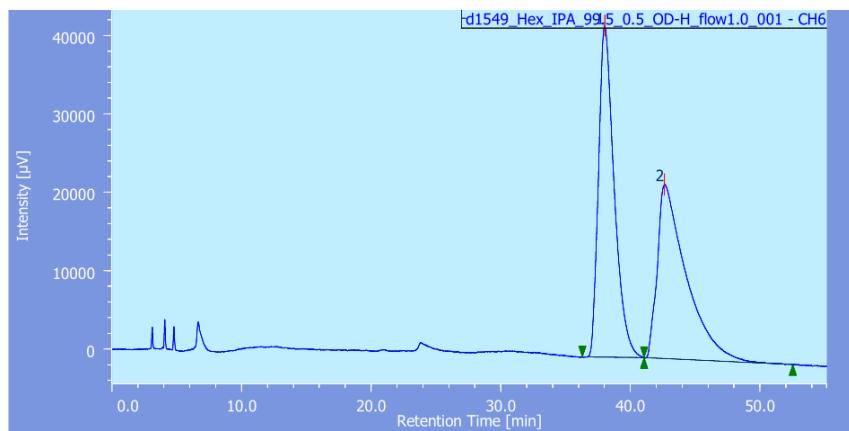
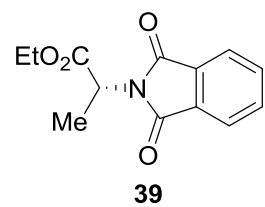
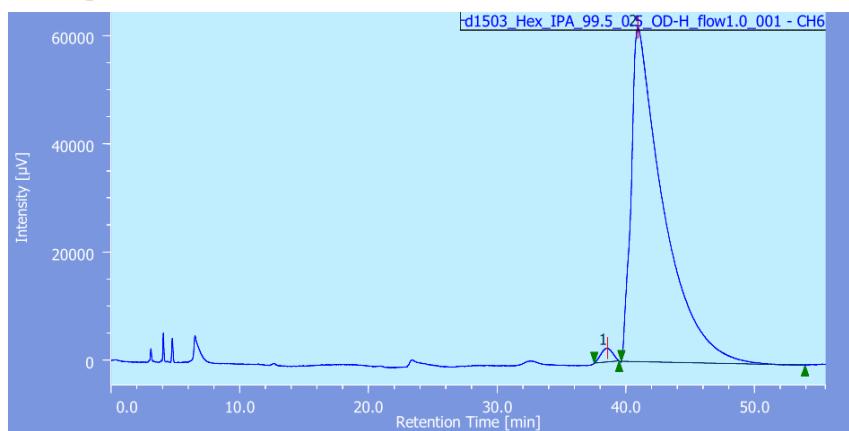
racemic mixture	tR (min)	area	area%
peak1	29.475	9510076	49.992
peak2	33.425	9513156	50.008
retention	tR (min)	area	area%
peak1	29.358	16911707	99.121
peak2	34.308	149995	0.879
sample	tR (min)	area	area%
peak1	30.183	146556	1.001
peak2	34.025	14495151	98.999

37-racemic**37-retention****37-sample**

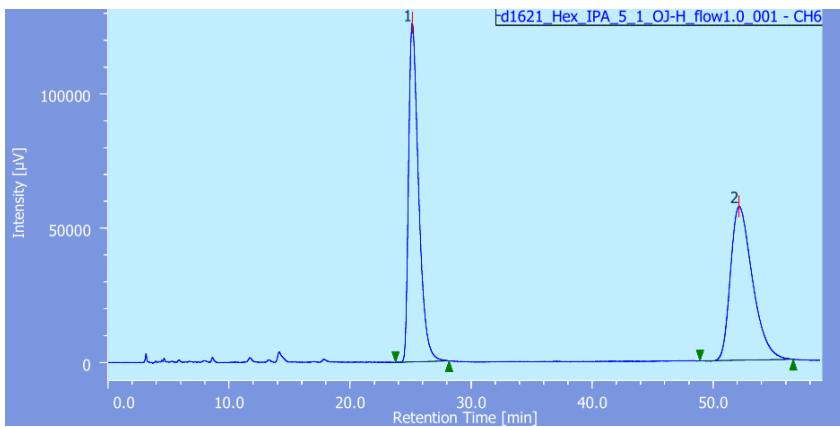
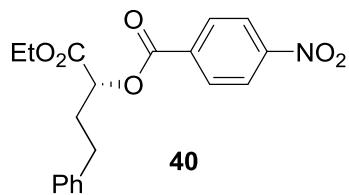
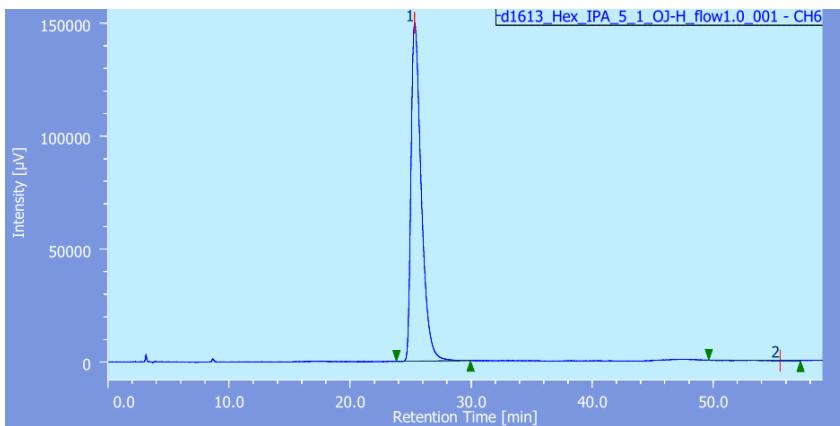
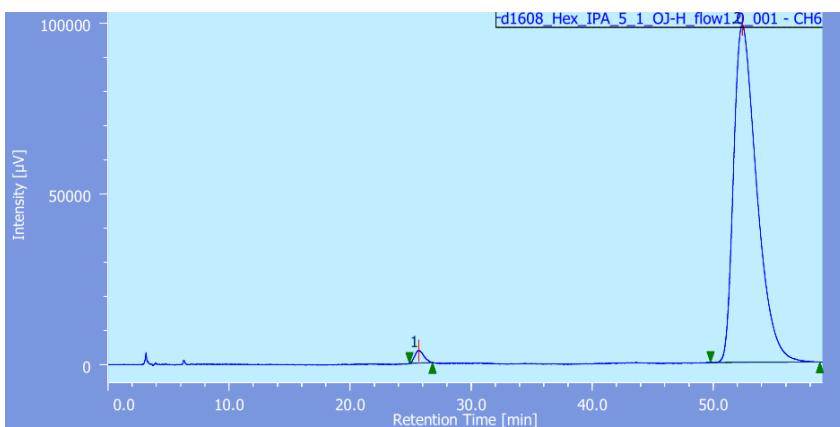
racemic mixture	tR (min)	area	area%
peak1	32.658	5946601	49.754
peak2	41.233	6005373	50.246
retention	tR (min)	area	area%
peak1	32.308	15520584	100.000
peak2			
sample	tR (min)	area	area%
peak1	32.800	713411	10.236
peak2	41.133	6255903	89.764

38-racemic**38-sample**

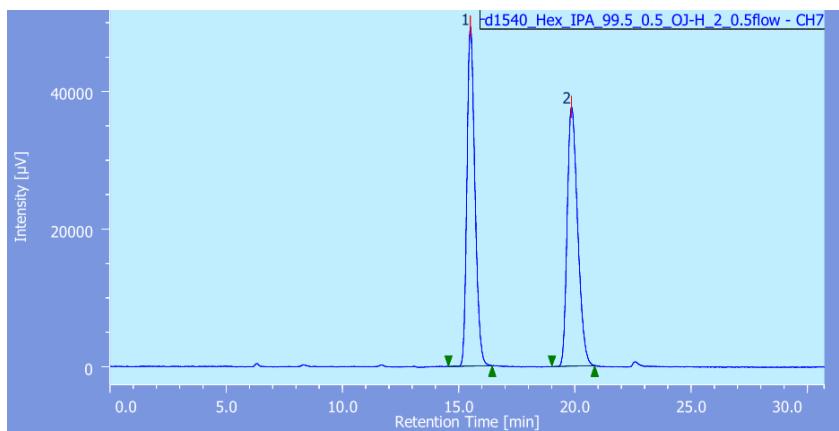
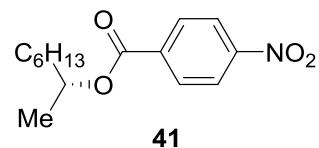
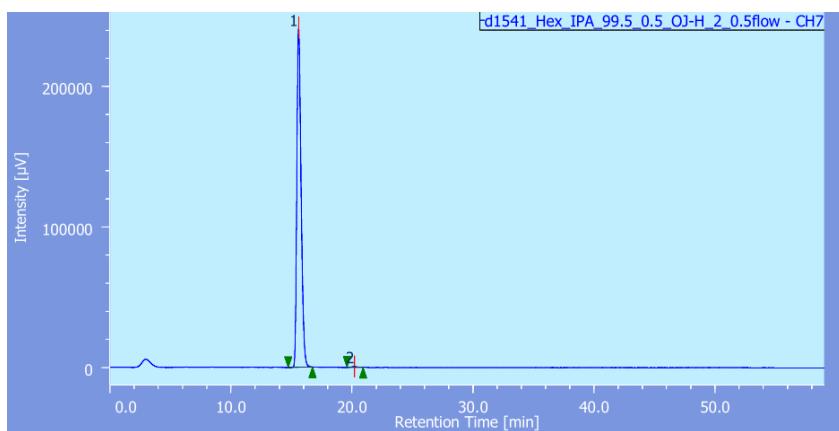
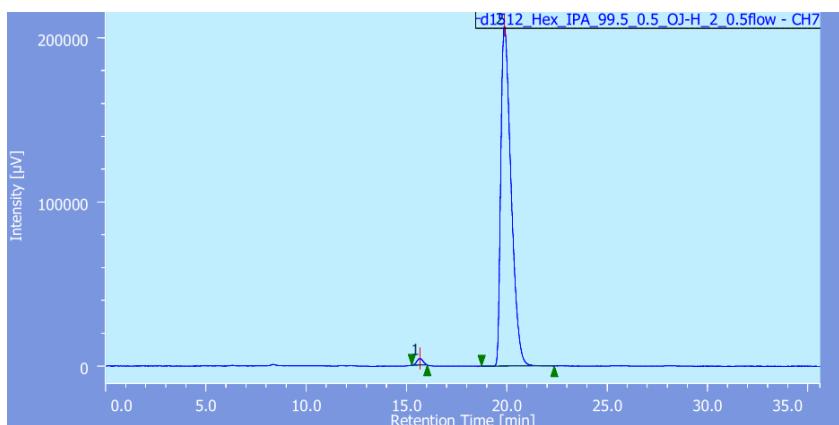
racemic mixture	tR (min)	area	area%
peak1	4.717	2147247	48.058
peak2	7.325	2320769	51.942
sample	tR (min)	area	area%
peak1	4.758	15534	0.448
peak2	7.308	3450655	99.552

39-racemic**39-sample**

racemic mixture	tR (min)	area	area%
peak1	38.008	3546595	50.756
peak2	42.633	3440934	49.244
sample	tR (min)	area	area%
peak1	38.583	161011	1.479
peak2	40.925	10727198	98.521

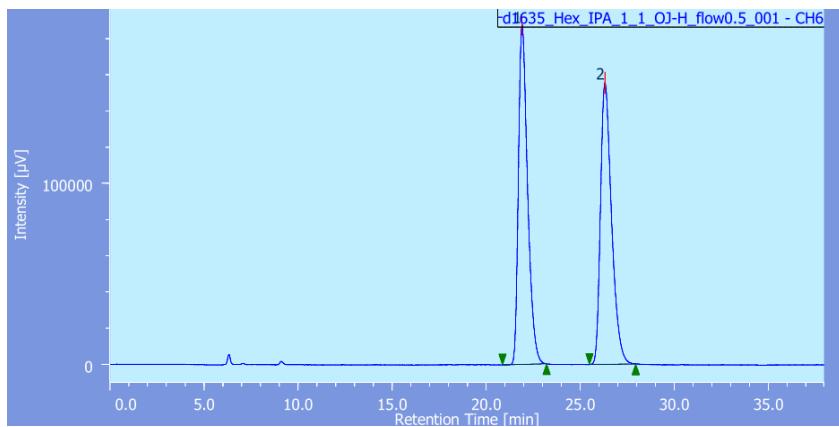
40-racemic**40-retention****40-sample**

racemic mixture	tR (min)	area	area%
peak1	25.133	7263509	50.114
peak2	52.133	7230515	49.886
retention	tR (min)	area	area%
peak1	25.333	8802532	99.688
peak2	55.517	27550	0.312
sample	tR (min)	area	area%
peak1	25.683	191068	1.438
peak2	52.392	13093028	98.562

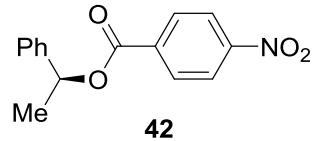
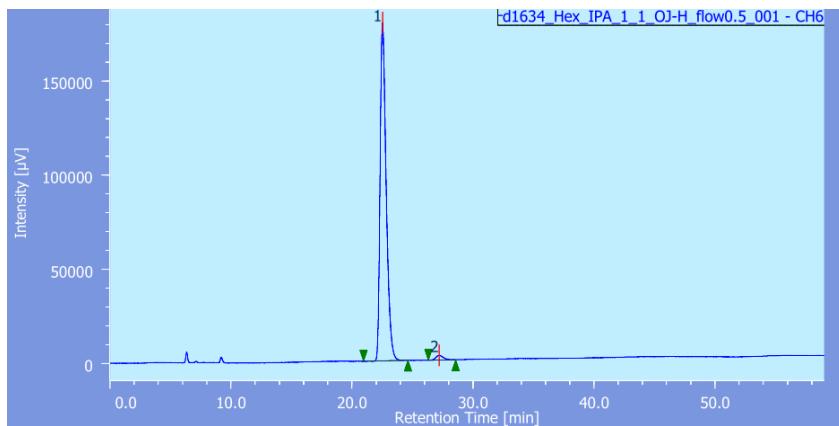
41-racemic**41-retention****41-sample**

racemic mixture	tR (min)	area	area%
peak1	15.500	1180115	49.979
peak2	19.850	1181109	50.021
retention	tR (min)	area	area%
peak1	15.583	5979359	99.684
peak2	20.200	18955	0.316
sample	tR (min)	area	area%
peak1	15.658	86560	1.158
peak2	19.875	7388866	98.842

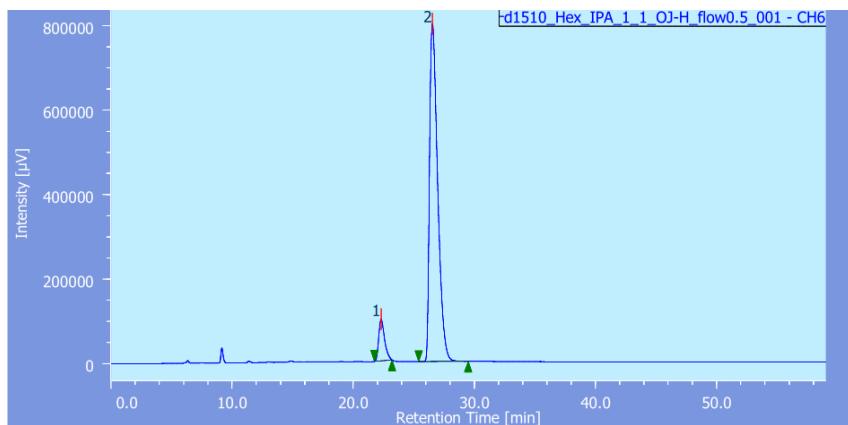
42-racemic



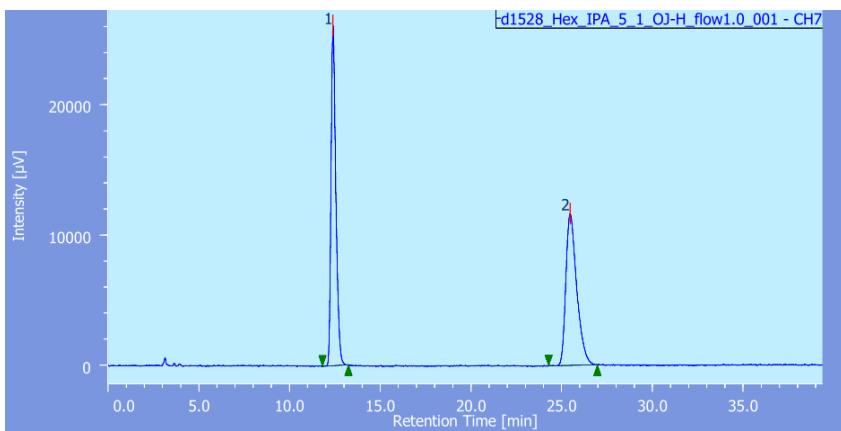
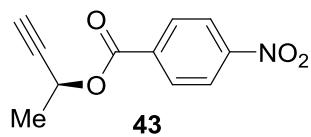
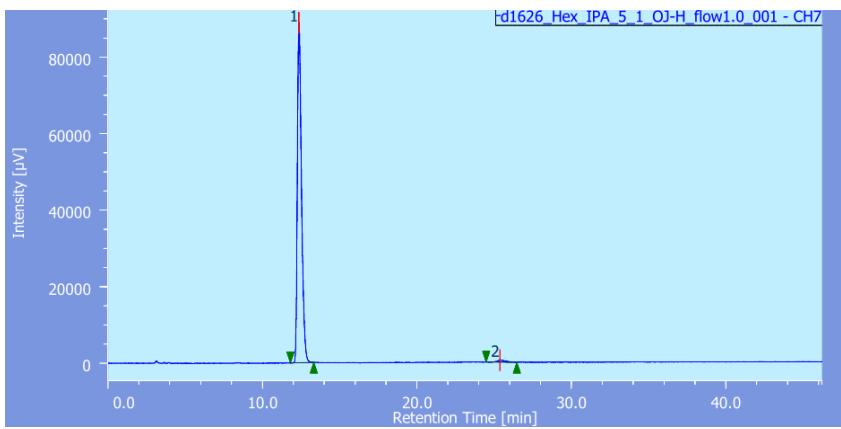
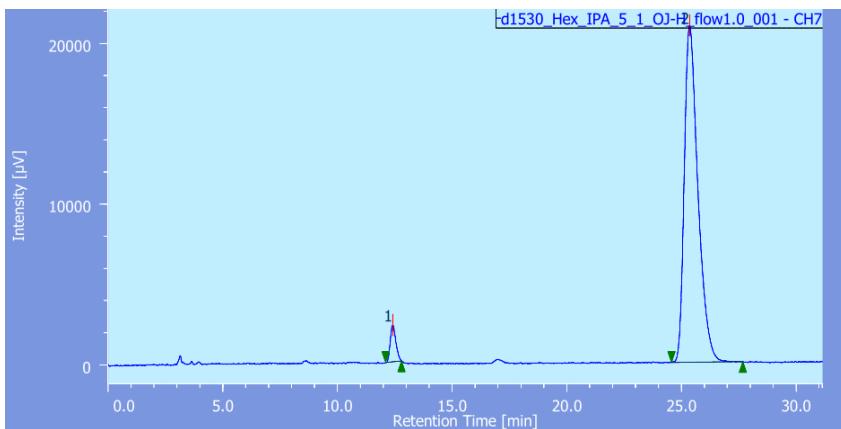
42-retention



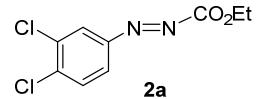
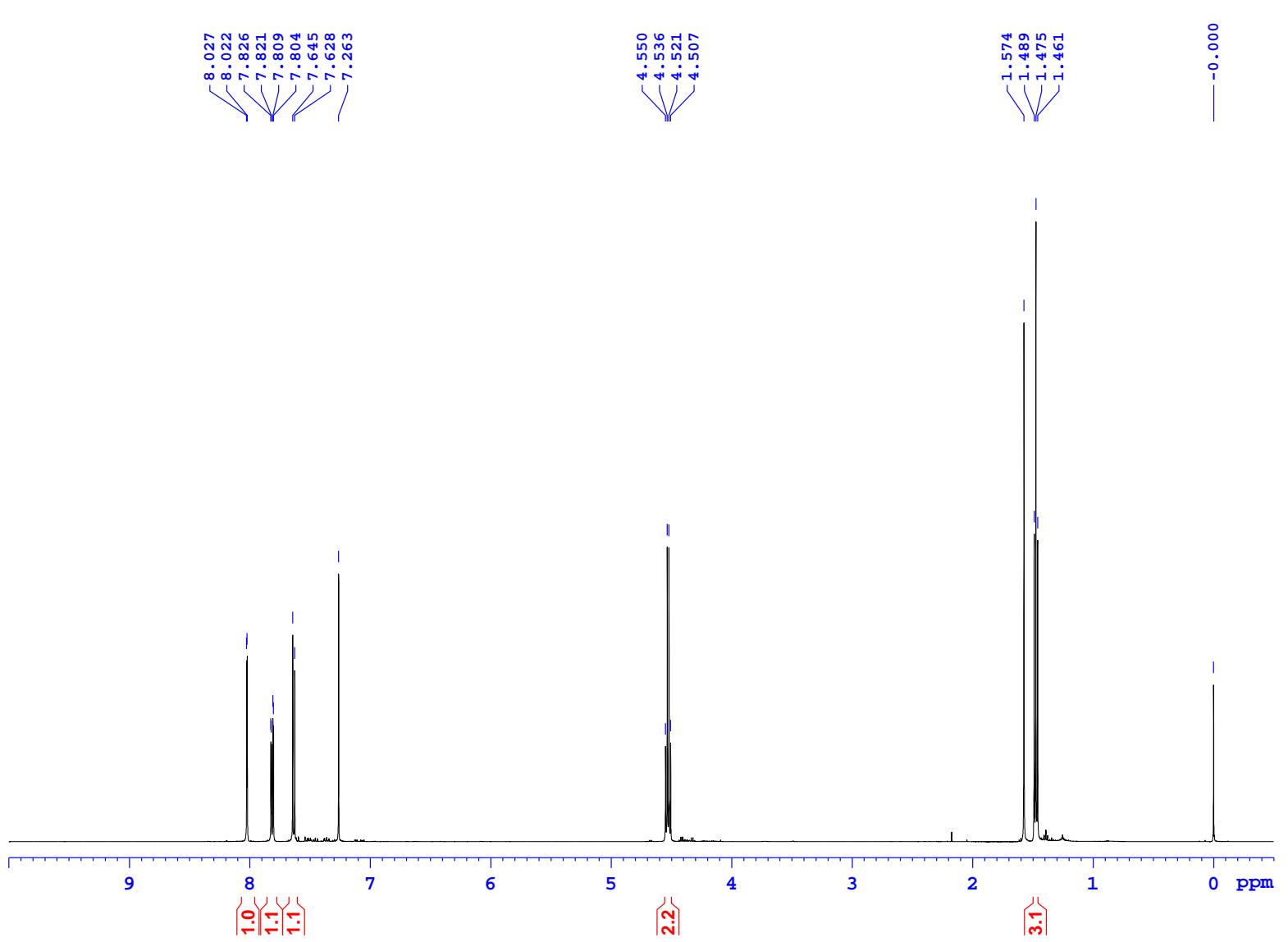
42-sample

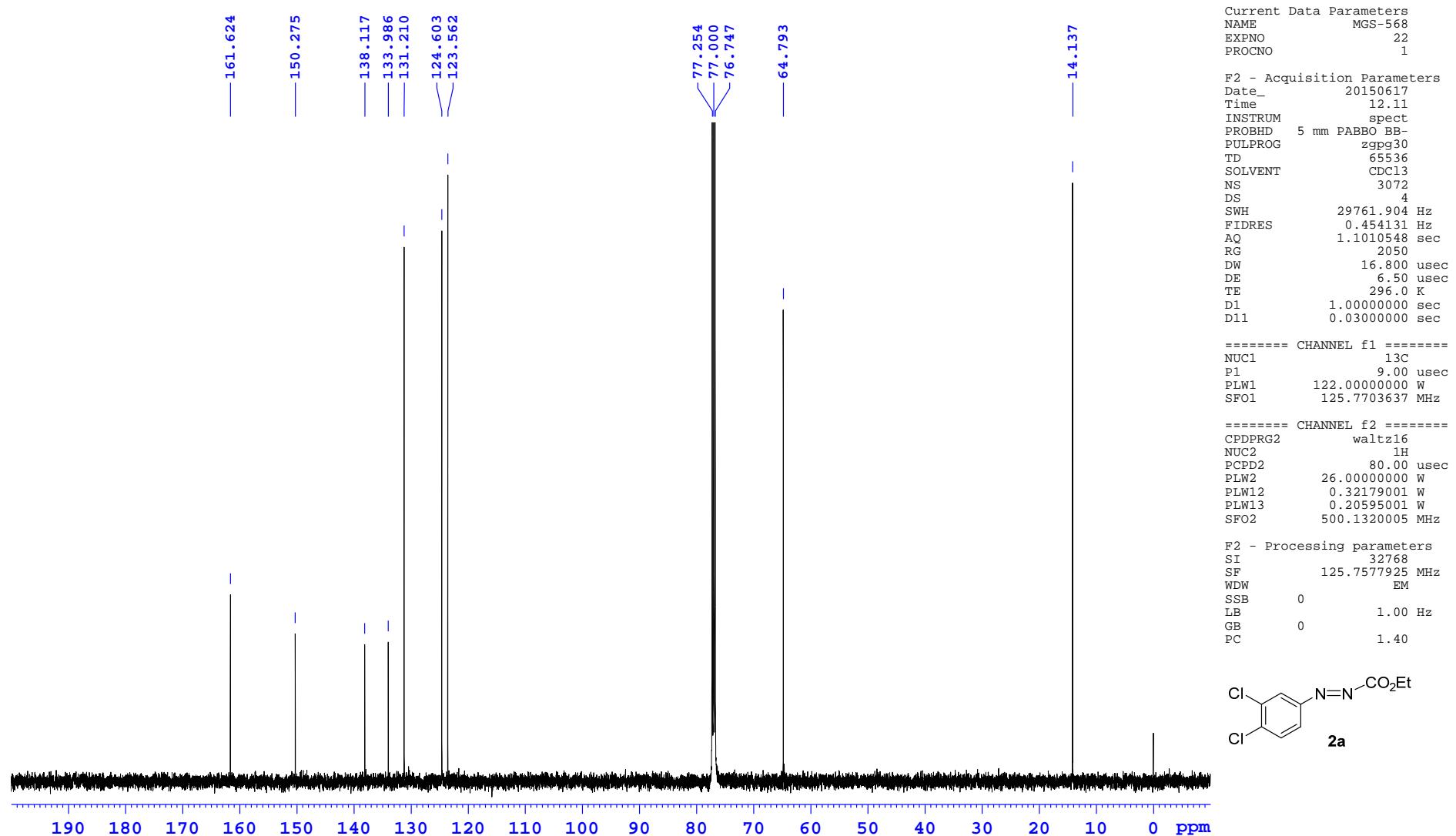


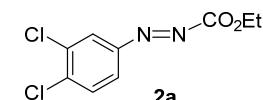
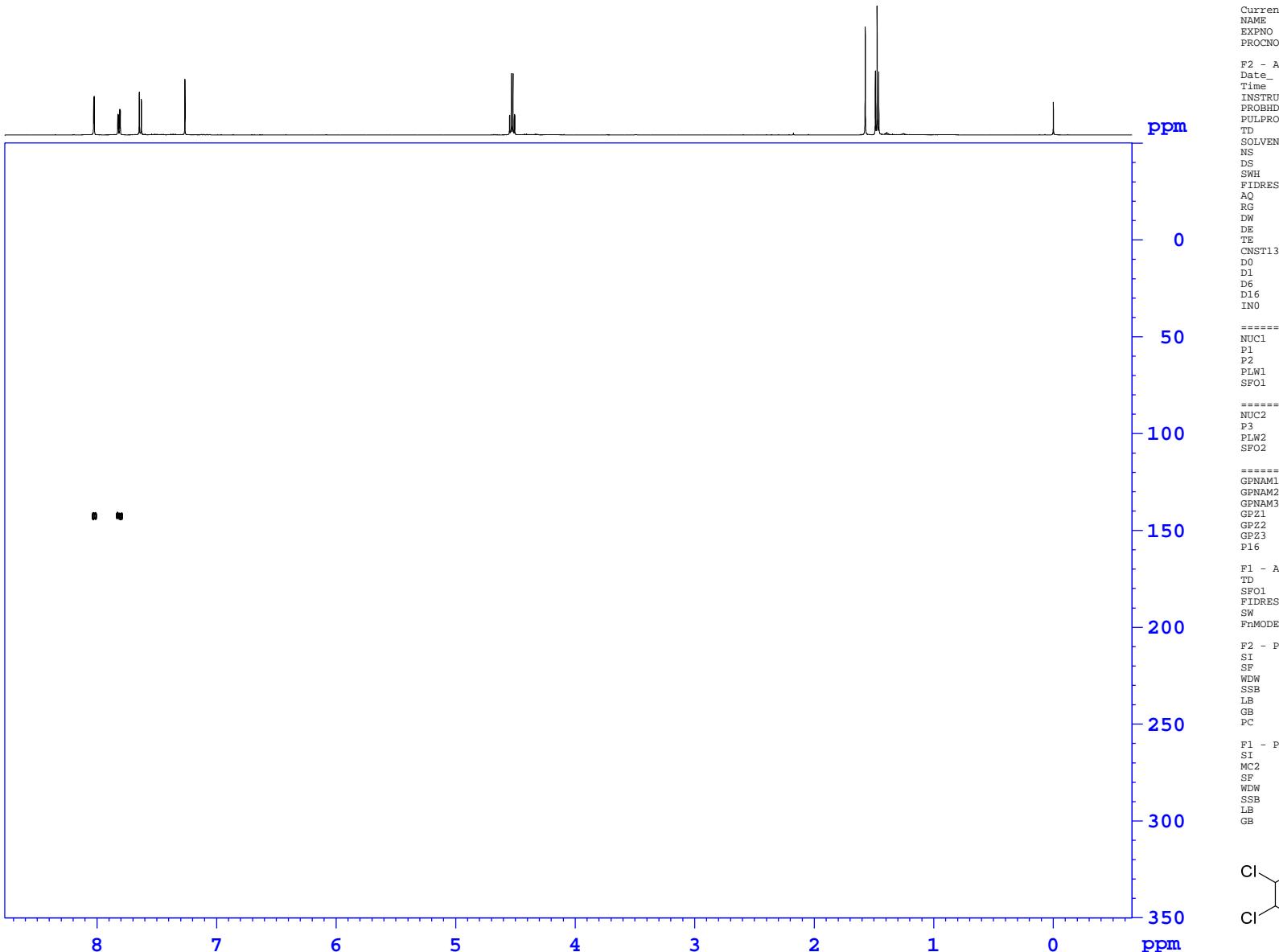
racemic mixture	tR (min)	area	area%
peak1	21.900	6358564	49.968
peak2	26.308	6366649	50.032
retention	tR (min)	area	area%
peak1	22.517	6321087	98.314
peak2	27.183	108430	1.686
sample	tR (min)	area	area%
peak1	22.292	3334306	8.216
peak2	26.533	37249530	91.784

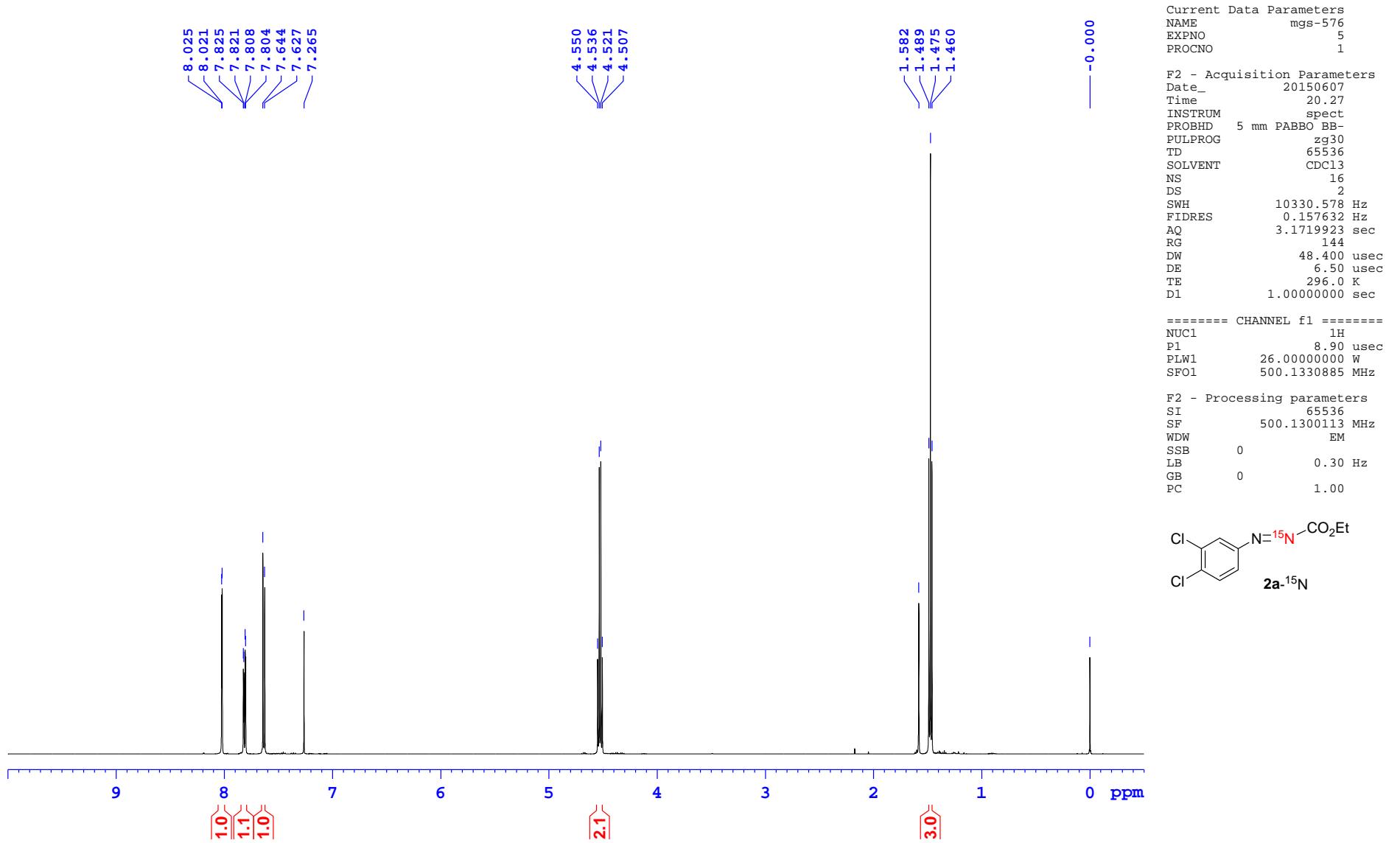
43-racemic**43-retention****43-sample**

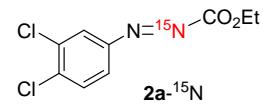
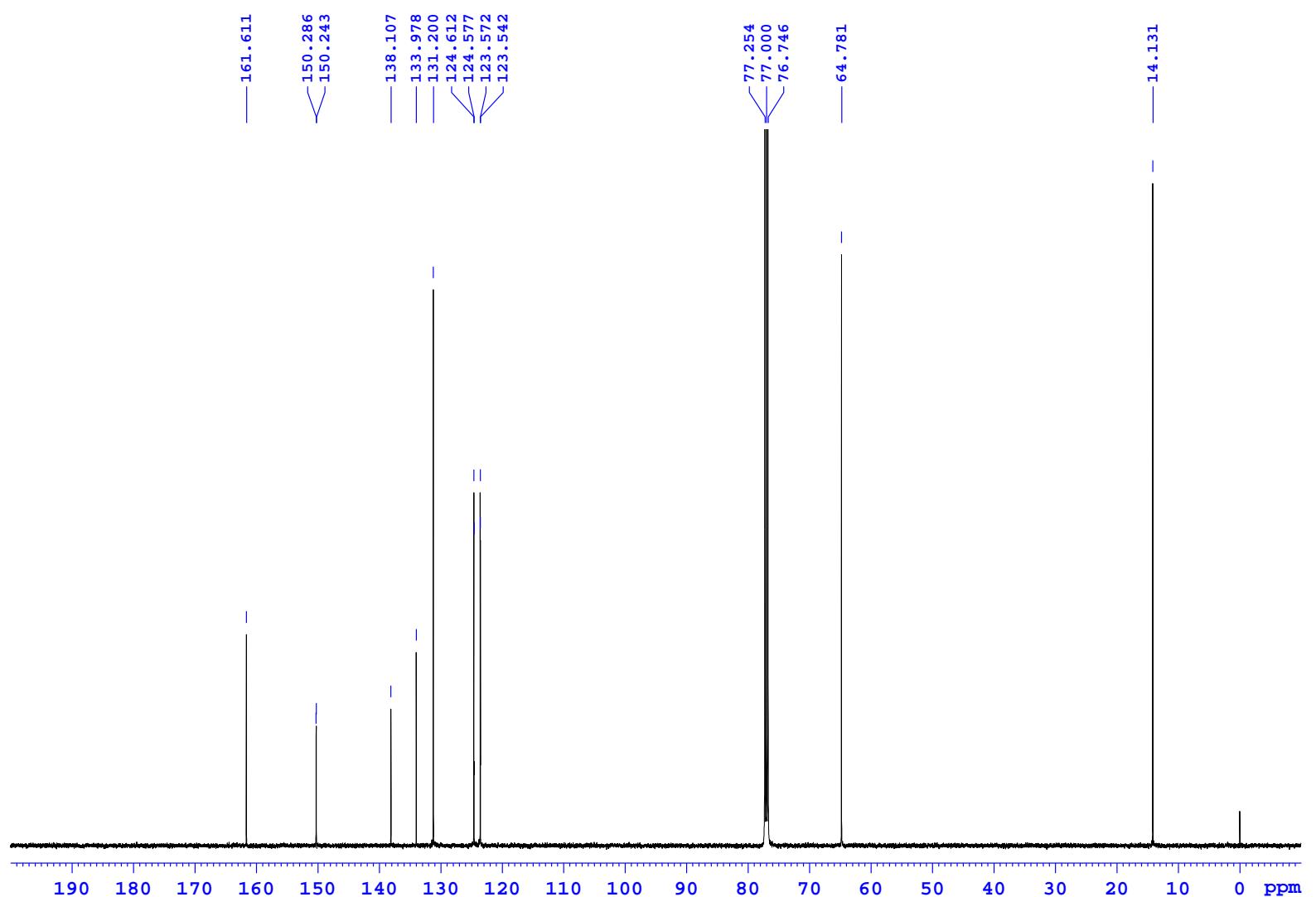
racemic mixture	tR (min)	area	area%
peak1	12.400	481719	49.988
peak2	25.458	481944	50.012
retention	tR (min)	area	area%
peak1	12.358	1659963	98.746
peak2	25.358	21073	1.254
sample	tR (min)	area	area%
peak1	12.408	38783	4.287
peak2	25.350	865888	95.713

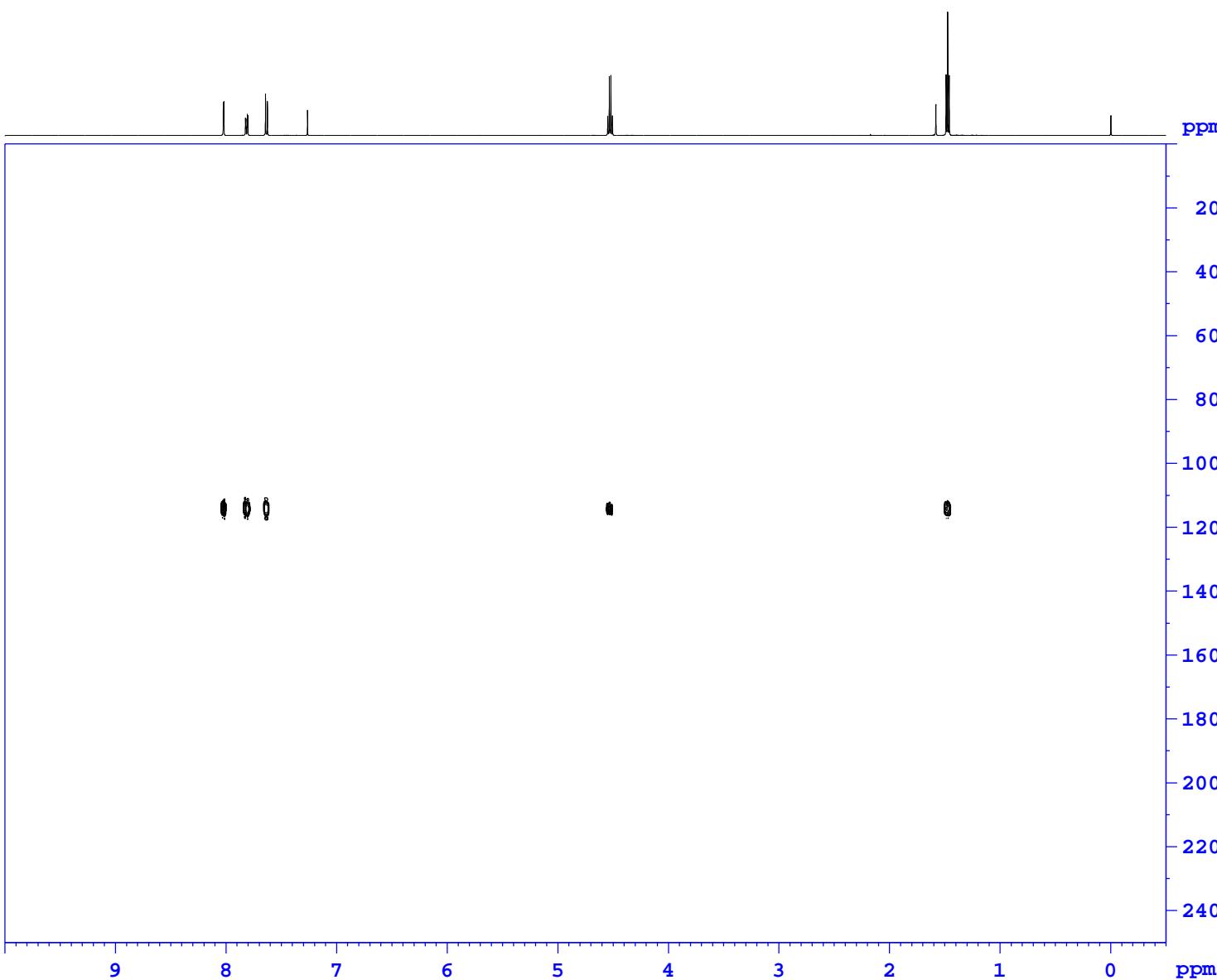












Current Data Parameters
NAME mgs-576
EXPNO 9
PROCNO 1

F2 - Acquisition Parameters
Date 20150608
Time 3.40
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG hmbcgpndqf
TD 2048
SOLVENT CDCl3
NS 8
DS 16
SWH 4672.897 Hz
FIDRES 2.281688 Hz
AQ 0.2191860 sec
RG 2050
DW 107.000 usec
DE 6.50 usec
TE 296.0 K
CNST13 5.0000000
D0 0.00000300 sec
D1 1.9381505 sec
D6 0.1000000 sec
D16 0.0002000 sec
INO 0.00002465 sec

===== CHANNEL f1 =====
NUC1 1H
P1 8.90 usec
P2 17.80 usec
PLW1 26.0000000 W
SFO1 500.1320044 MHz

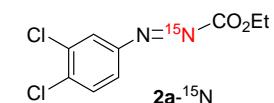
===== CHANNEL f2 =====
NUC2 15N
P3 14.40 usec
PLW2 206.0000000 W
SFO2 50.6777330 MHz

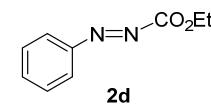
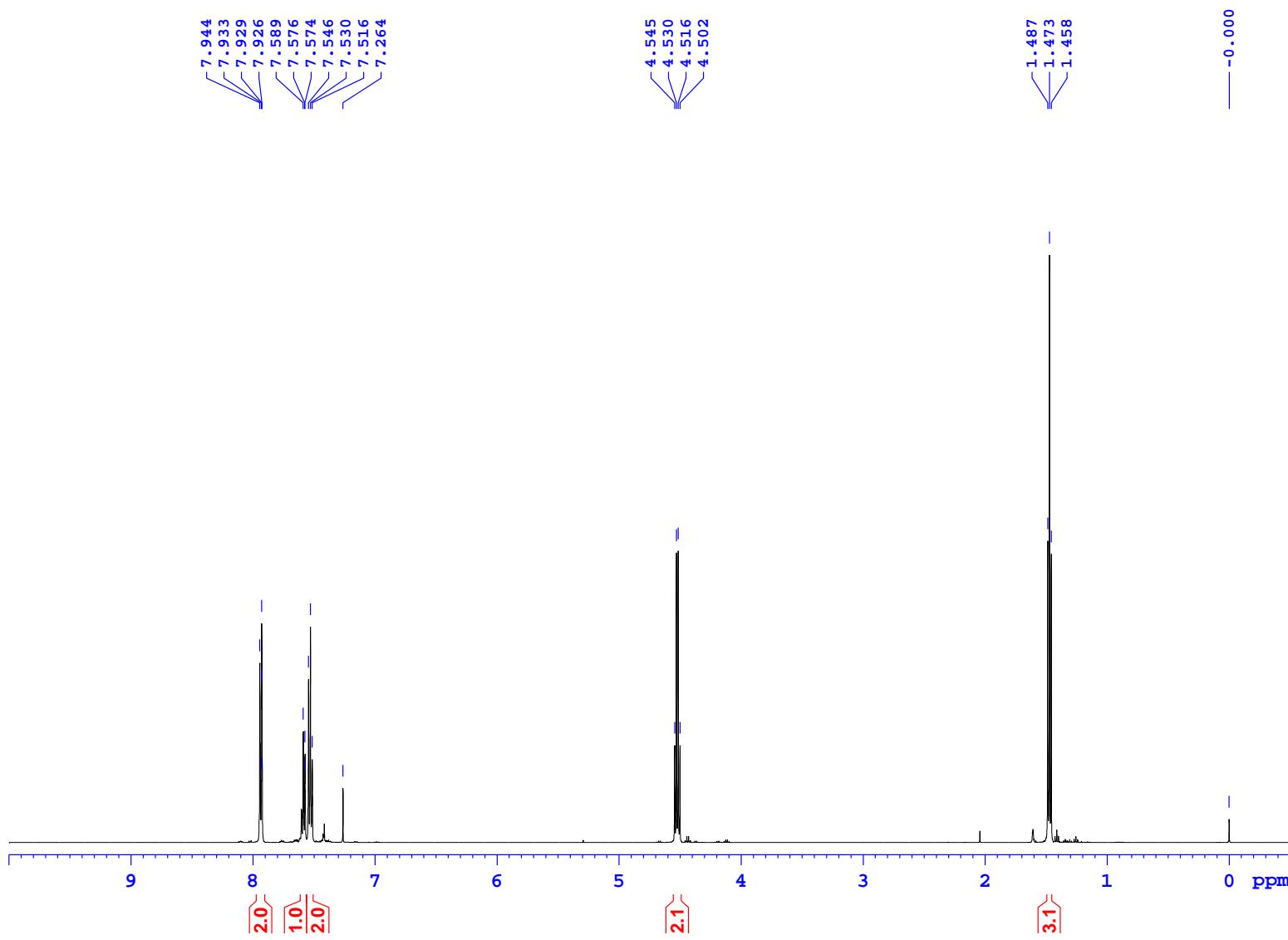
===== GRADIENT CHANNEL =====
GPNAME1 SMSQ10.100
GPNAME2 SMSQ10.100
GPNAME3 SMSQ10.100
GPZ1 70.00 %
GPZ2 30.00 %
GPZ3 50.10 %
P16 1000.00 usec

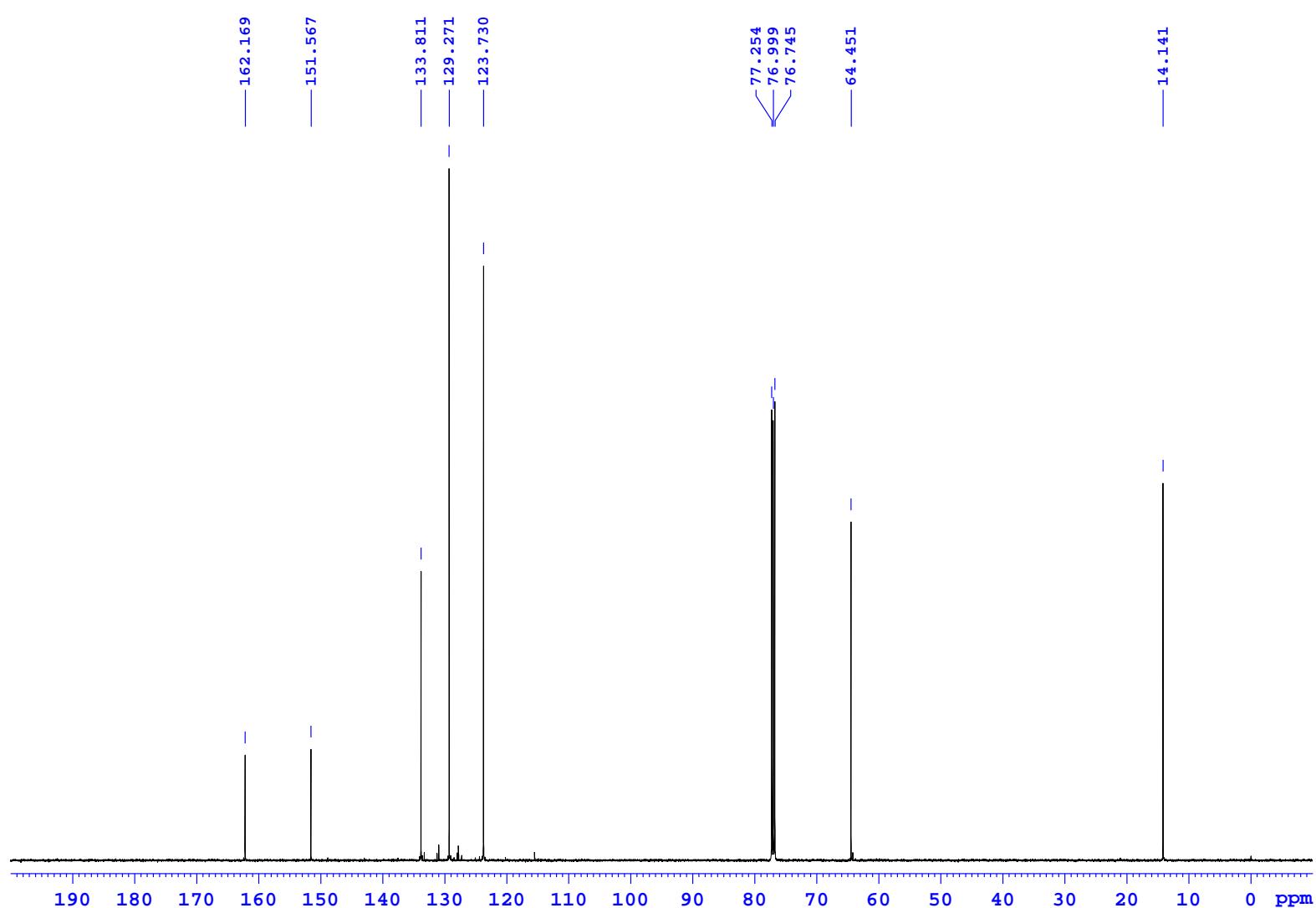
F1 - Acquisition parameters
TD 128
SFO1 50.67773 MHz
FIDRES 158.391663 Hz
SW 400.060 ppm
FnMODE QF

F2 - Processing parameters
SI 1024
SF 500.1300113 MHz
WDW SINE
SSB 0
LB 0 Hz
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
MC2 QF
SF 50.6777330 MHz
WDW States
SSB 0
LB 0 Hz
GB 0







Current Data Parameters

NAME	MGS-661
EXPNO	3
PROCNO	1

F2 - Acquisition Parameters

Date_	20151103
Time	0.58
INSTRUM	spect
PROBHD	5 mm PABBO BB-
PULPROG	zgpg30
TD	65536
SOLVENT	CDCl ₃
NS	3072
DS	4
SWH	29761.904 Hz
FIDRES	0.454131 Hz
AQ	1.1010548 sec
RG	2050
DW	16.800 usec
DE	6.50 usec
TE	296.0 K
D1	1.0000000 sec
D11	0.0300000 sec

===== CHANNEL f1 =====

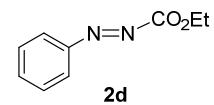
NUC1	13C
P1	9.00 usec
PLW1	122.0000000 W
SFO1	125.7703637 MHz

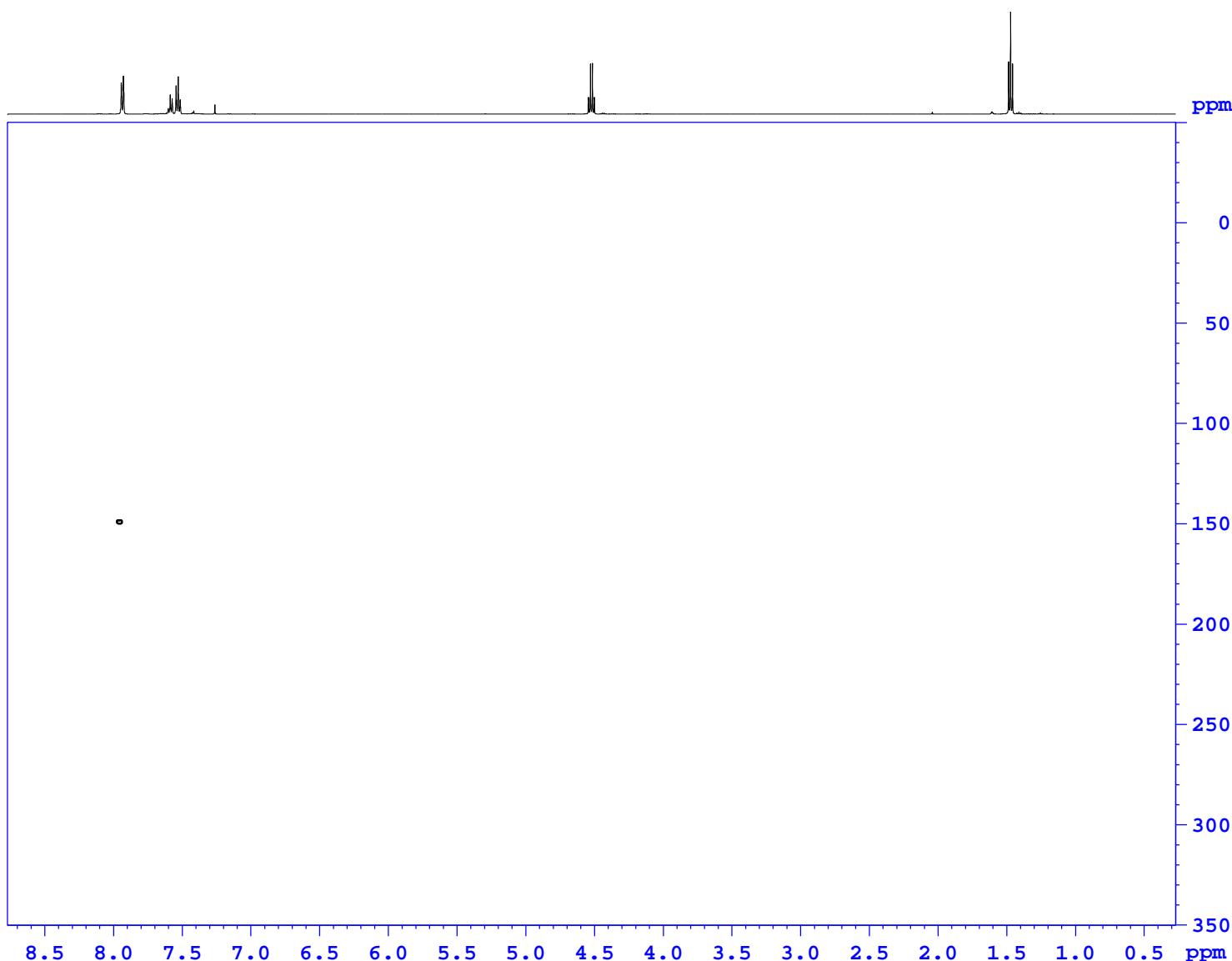
===== CHANNEL f2 =====

CPDPRG2	waltz16
NUC2	1H
PCPD2	80.00 usec
PLW2	26.0000000 W
PLW12	0.32179001 W
PLW13	0.20595001 W
SFO2	500.1320005 MHz

F2 - Processing parameters

SI	32768
SF	125.7577957 MHz
WDW	EM
SSB	0
LB	1.00 Hz
GB	0
PC	1.40



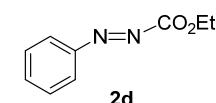


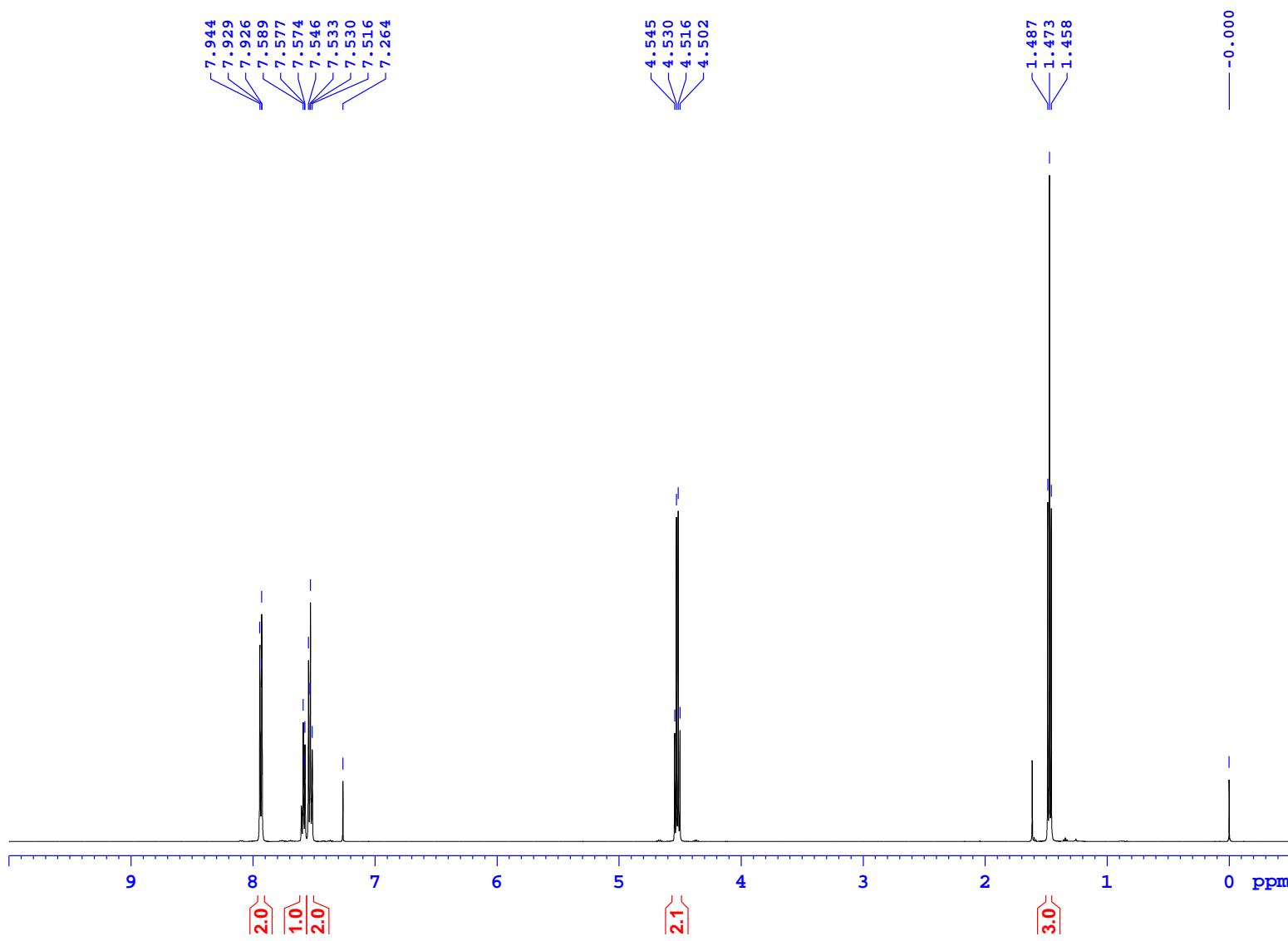
Current Data Parameters
NAME MGS-472
EXPNO 23
PROCNO 1
F2 - Acquisition Parameters
Date_ 20150222
Time 22.31
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG hmbcgpndqf
TD 2048
SOLVENT CDCl₃
NS 8
DS 16
SWH 4464.286 Hz
FIDRES 2.179827 Hz
AQ 0.2294260 sec
RG 2050
DW 112.000 usec
DE 6.50 usec
TE 296.0 K
CNST13 5.0000000
D0 0.00000300 sec
D1 1.9279097 sec
D6 0.1000000 sec
D16 0.0002000 sec
IN0 0.00002465 sec
===== CHANNEL f1 =====
NUC1 1H
P1 8.90 usec
P2 17.80 usec
PLW1 26.0000000 W
SFO1 500.1323675 MHz
===== CHANNEL f2 =====
NUC2 15N
P3 14.40 usec
PLW2 206.0000000 W
SFO2 50.6853342 MHz
===== GRADIENT CHANNEL =====
GPNAME1 SMSQ10.100
GPNAME2 SMSQ10.100
GPNAME3 SMSQ10.100
GPZ1 70.00 %
GPZ2 30.00 %
GPZ3 50.10 %
P16 1000.00 usec

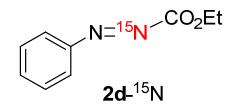
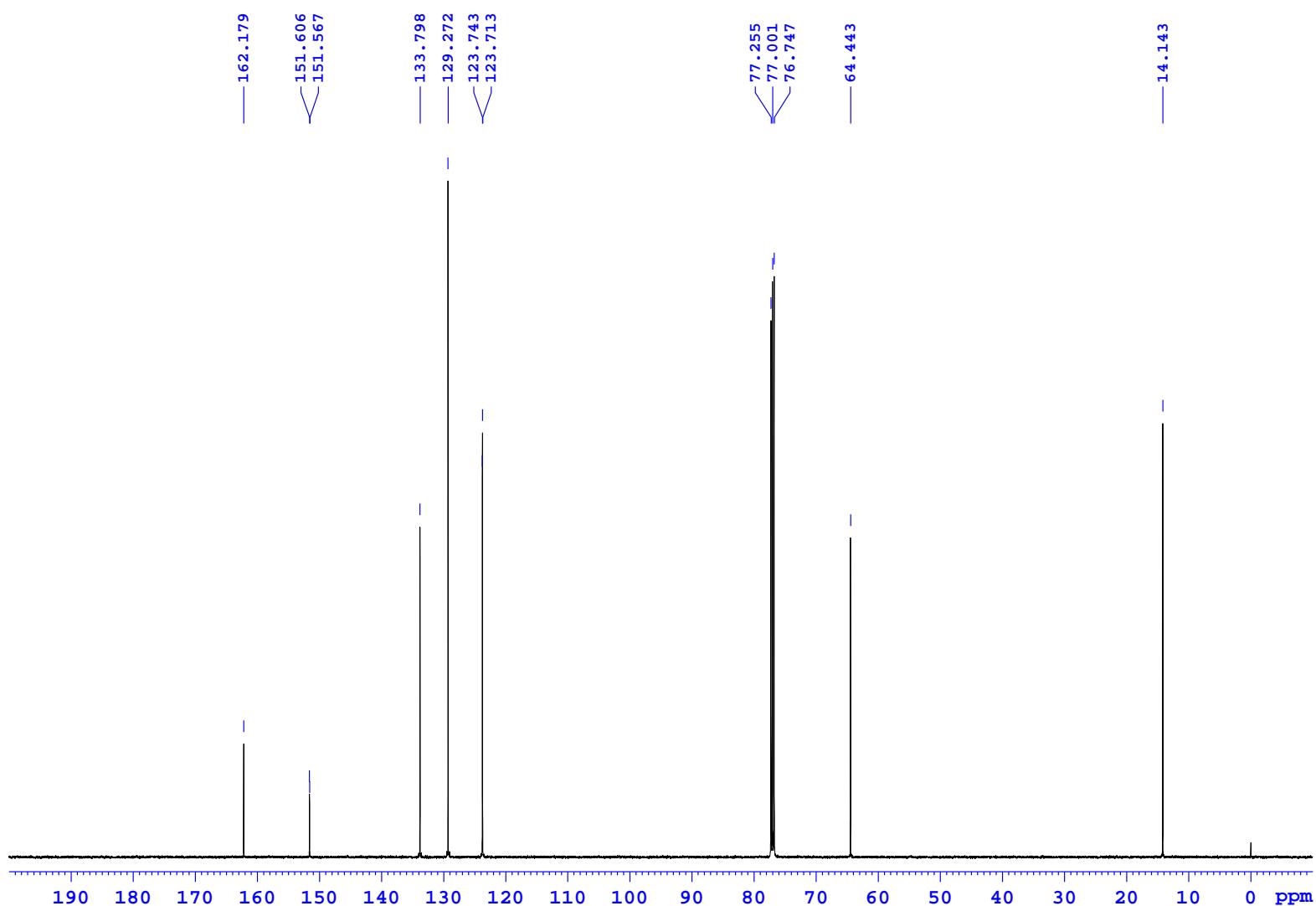
F1 - Acquisition parameters
TD 256
SFO1 50.68533 MHz
FIDRES 79.195831 Hz
SW 400.000 ppm
FnMODE QF

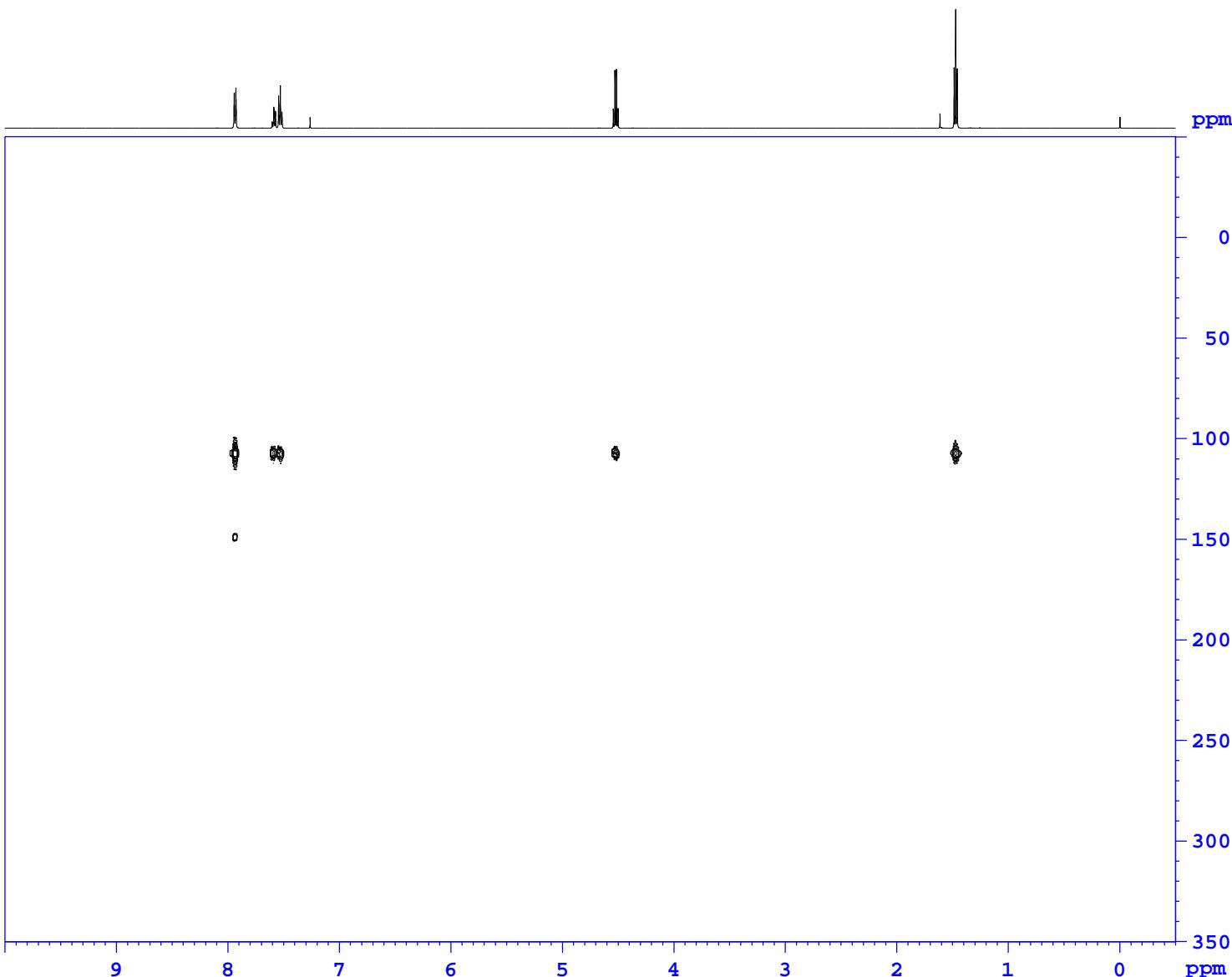
F2 - Processing parameters
SI 2048
SF 500.1300000 MHz
WDW SINE
SSB 0
LB 0 Hz
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
MC2 QF
SF 50.6777330 MHz
WDW echo-antiecho
SSB 0
LB 0 Hz
GB 0









Current Data Parameters
NAME MGS-645
EXPNO 25
PROCNO 1

F2 - Acquisition Parameters
Date_ 20151014
Time 5.54
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG hmbcgpndqf
TD 2048
SOLVENT CDCl3
NS 32
DS 16
SWH 4716.981 Hz
FIDRES 2.303213 Hz
AQ 0.2171380 sec
RG 2050
DW 106.000 usec
DE 6.50 usec
TE 296.1 K
CNST13 5.0000000
D0 0.00000300 sec
D1 1.94019794 sec
D6 0.10000000 sec
D16 0.00020000 sec
IN0 0.00002465 sec

===== CHANNEL f1 =====
NUC1 1H
P1 8.90 usec
P2 17.80 usec
PLW1 26.0000000 W
SFO1 500.1320253 MHz

===== CHANNEL f2 =====
NUC2 15N
P3 14.40 usec
PLW2 206.0000000 W
SFO2 50.6853342 MHz

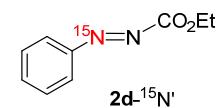
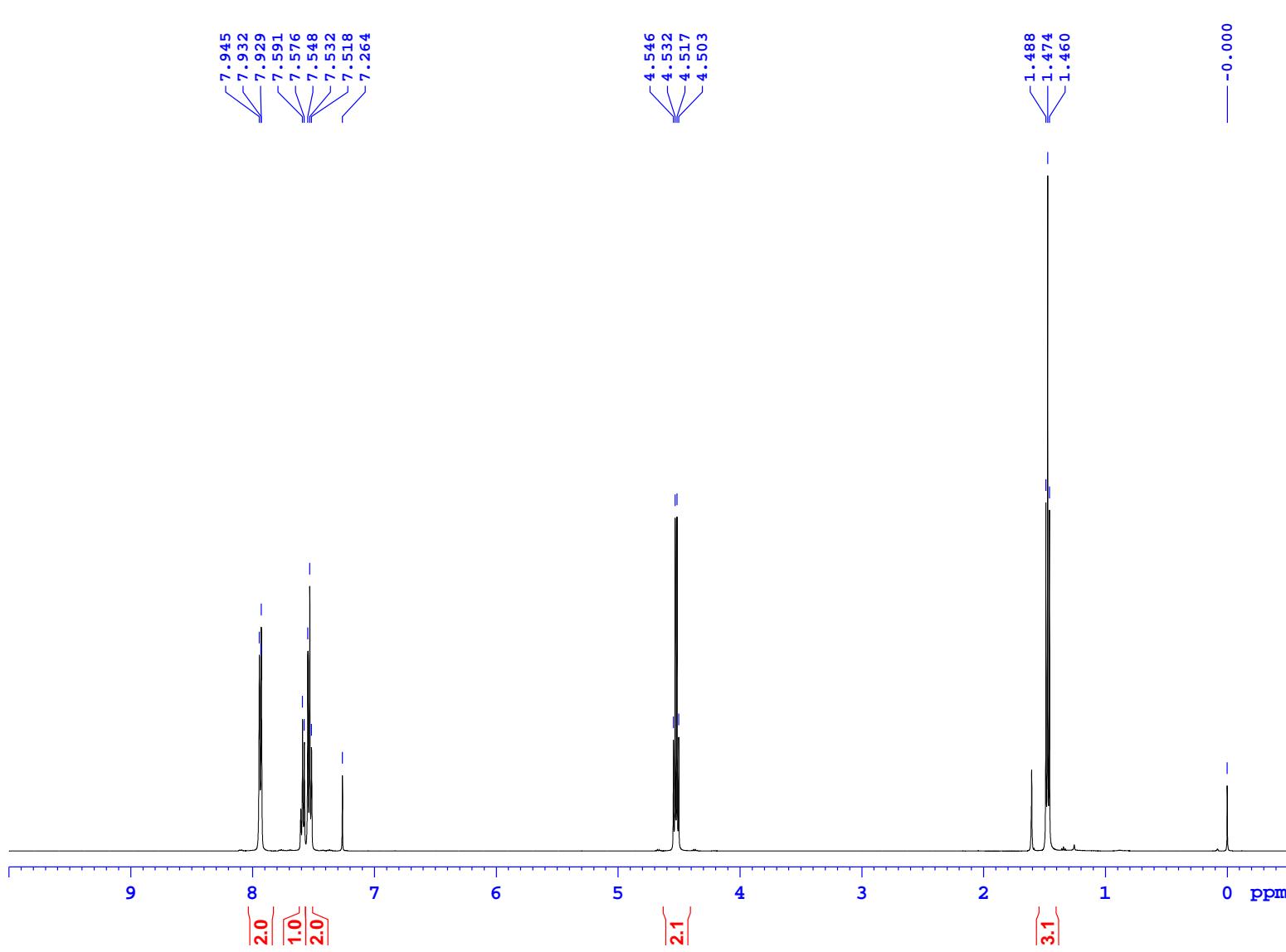
===== GRADIENT CHANNEL =====
GPNAME1 SMSQ10.100
GPNAME2 SMSQ10.100
GPNAME3 SMSQ10.100
GPZ1 70.00 %
GPZ2 30.00 %
GPZ3 50.10 %
P16 1000.00 usec

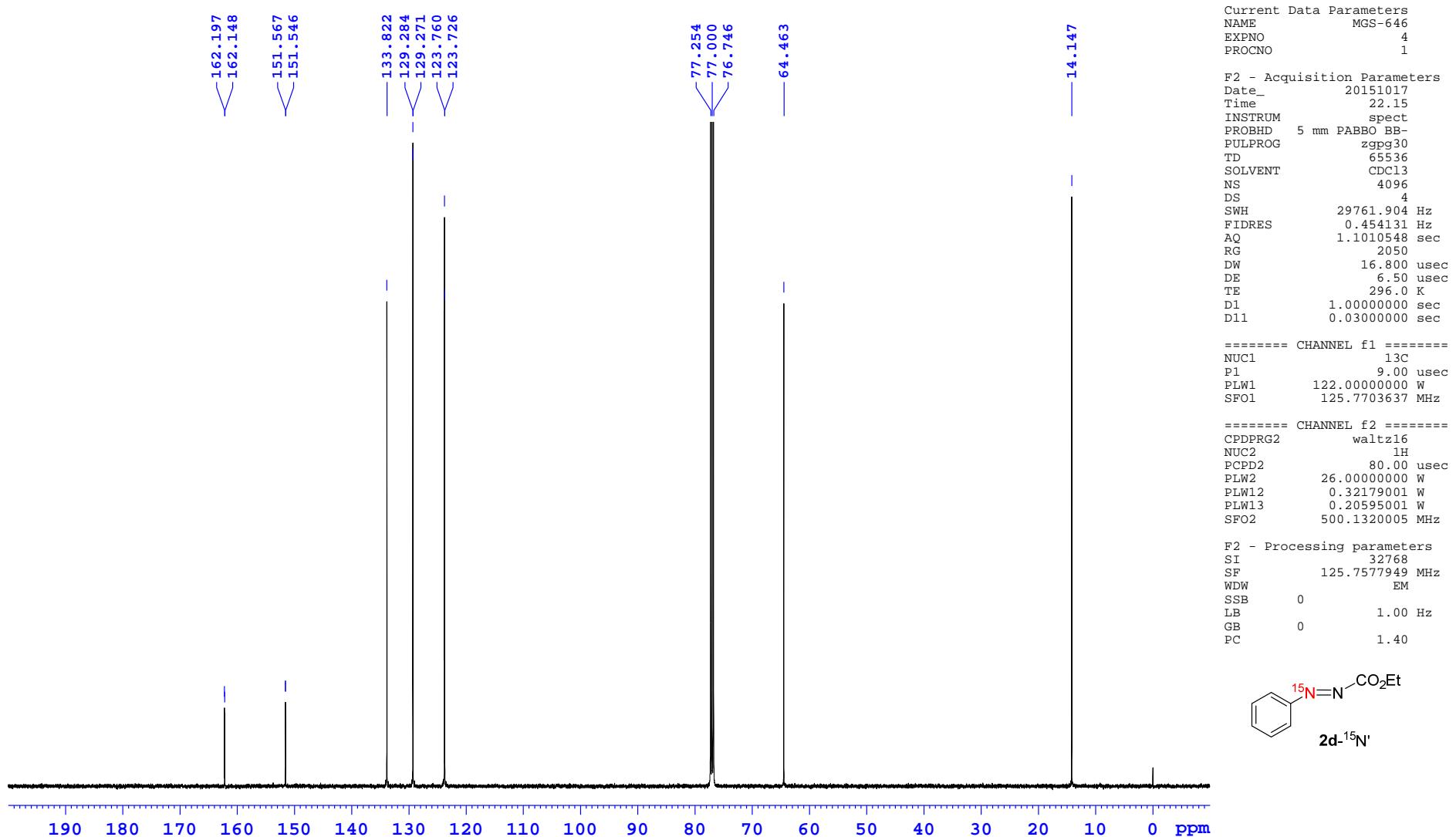
F1 - Acquisition parameters
TD 128
SFO1 50.68533 MHz
FIDRES 158.391663 Hz
SW 400.000 ppm
FnMODE QF

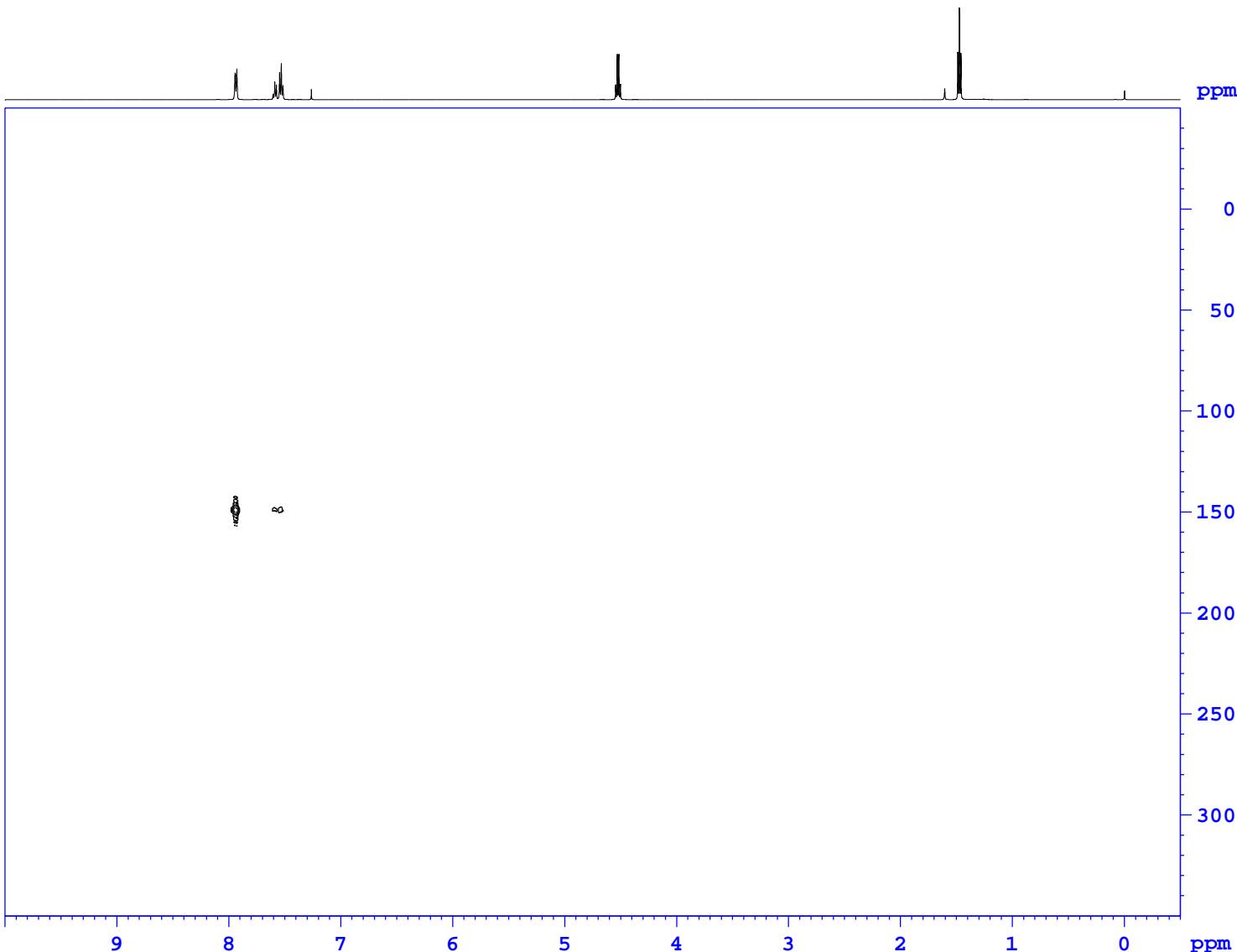
F2 - Processing parameters
SI 2048
SF 500.1300116 MHz
WDW SINE
SSB 0
LB 0 Hz
GB 0
PC 1.40

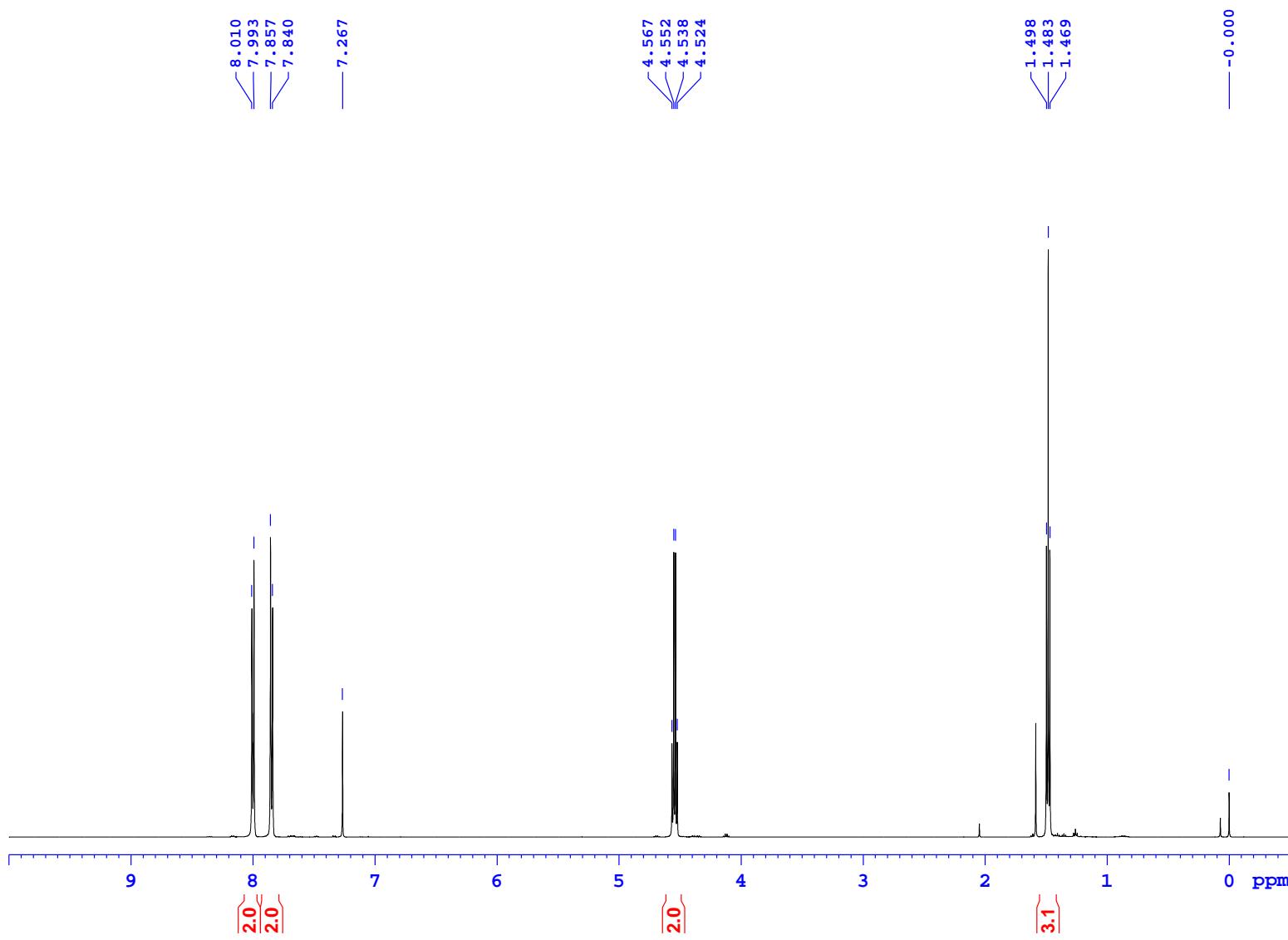
F1 - Processing parameters
SI 1024
MC2 QF
SF 50.6777330 MHz
WDW States
SSB 0
LB 0 Hz
GB 0











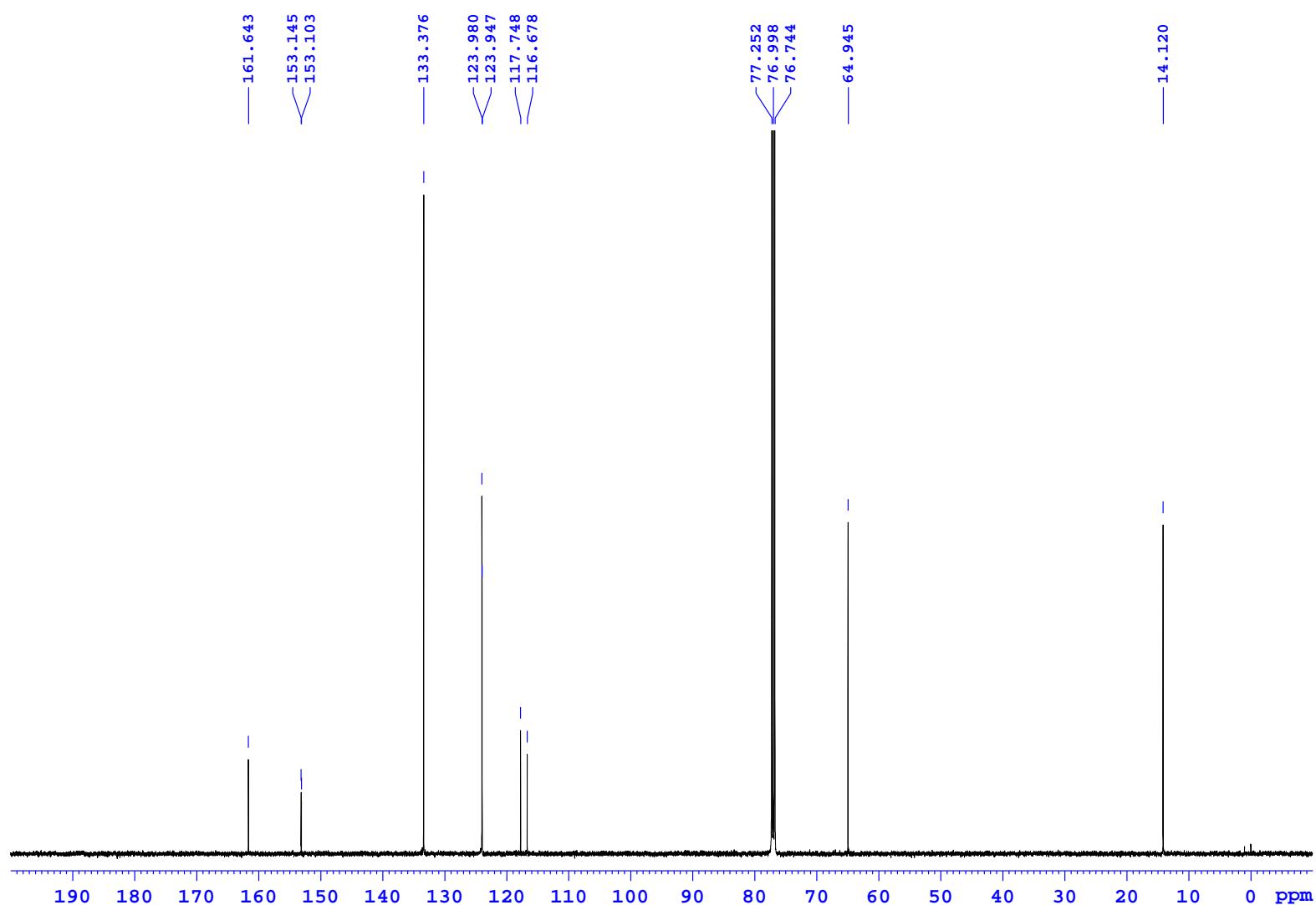
Current Data Parameters
 NAME MGS-650
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20151022
 Time 0.17
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 10330.578 Hz
 FIDRES 0.157632 Hz
 AQ 3.1719923 sec
 RG 144
 DW 48.400 usec
 DE 6.50 usec
 TE 296.0 K
 D1 1.0000000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 8.90 usec
 PLW1 26.0000000 W
 SF01 500.1330885 MHz

F2 - Processing parameters
 SI 65536
 SF 500.1300102 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00





Current Data Parameters
NAME MGS-650
EXPNO 3
PROCNO 1

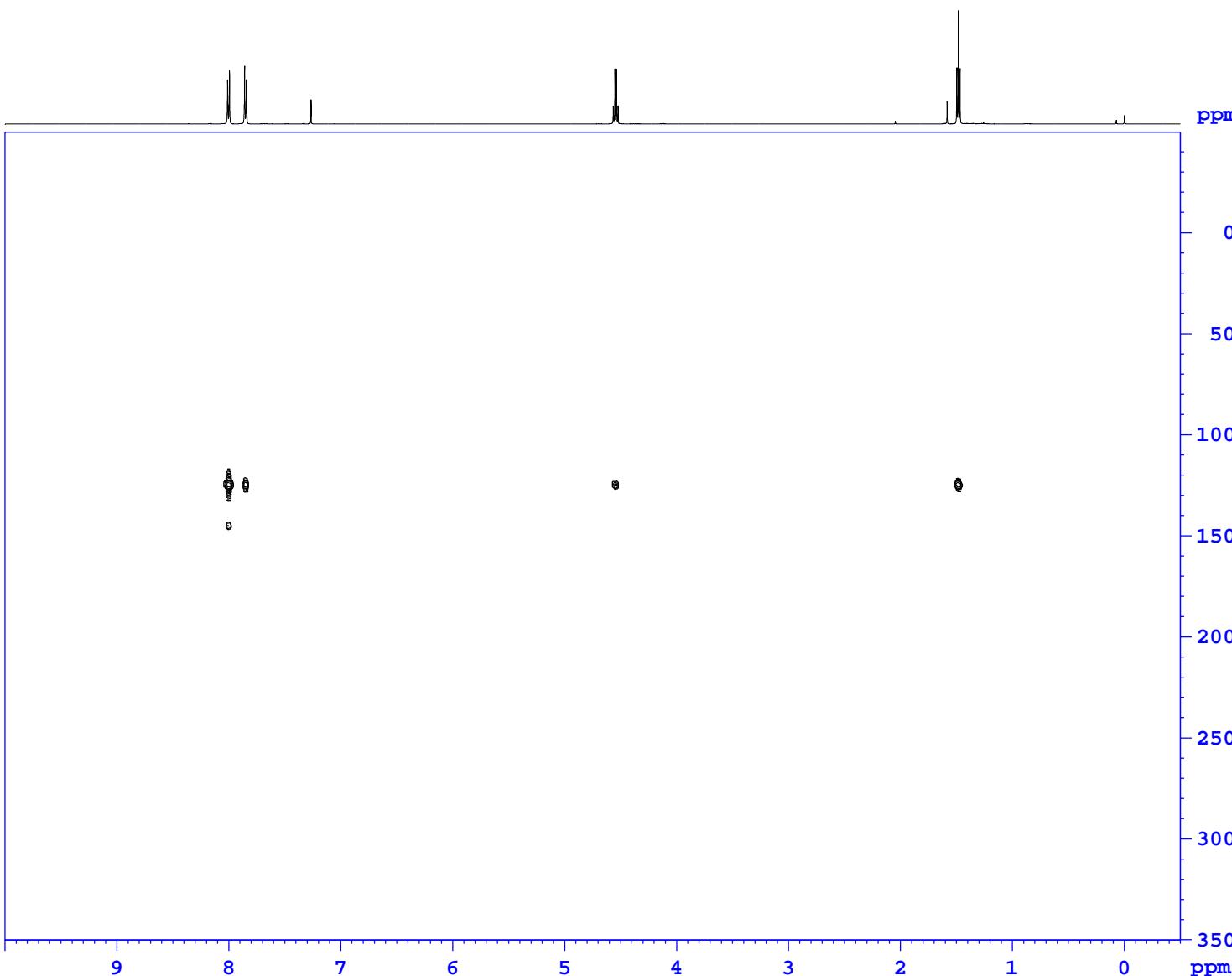
F2 - Acquisition Parameters
Date_ 20151022
Time 2.15
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 3072
DS 4
SWH 29761.904 Hz
FIDRES 0.454131 Hz
AQ 1.1010548 sec
RG 2050
DW 16.800 usec
DE 6.50 usec
TE 296.0 K
D1 1.0000000 sec
D11 0.0300000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 9.00 usec
PLW1 122.0000000 W
SFO1 125.7703637 MHz

===== CHANNEL f2 =====
CPDPG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PLW2 26.0000000 W
PLW12 0.32179001 W
PLW13 0.20595001 W
SFO2 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 125.7577945 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40





Current Data Parameters
 NAME MGS-650
 EXPNO 6
 PROCNO 1

F2 - Acquisition Parameters
 Date 20151022
 Time 3.04
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG hmbcgpndqf
 TD 2048
 SOLVENT CDCl3
 NS 16
 DS 16
 SWH 4761.905 Hz
 FIDRES 2.325149 Hz
 AQ 0.2150900 sec
 RG 2050
 DW 105.000 usec
 DE 6.50 usec
 TE 296.0 K
 CNST13 5.0000000
 D0 0.00000300 sec
 D1 1.94224596 sec
 D6 0.1000000 sec
 D16 0.00020000 sec
 IN0 0.00002465 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 8.90 usec
 P2 17.80 usec
 PLW1 26.0000000 W
 SFO1 500.1320605 MHz

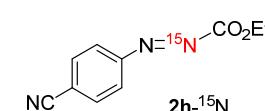
===== CHANNEL f2 =====
 NUC2 15N
 P3 14.40 usec
 PLW2 206.0000000 W
 SFO2 50.6853342 MHz

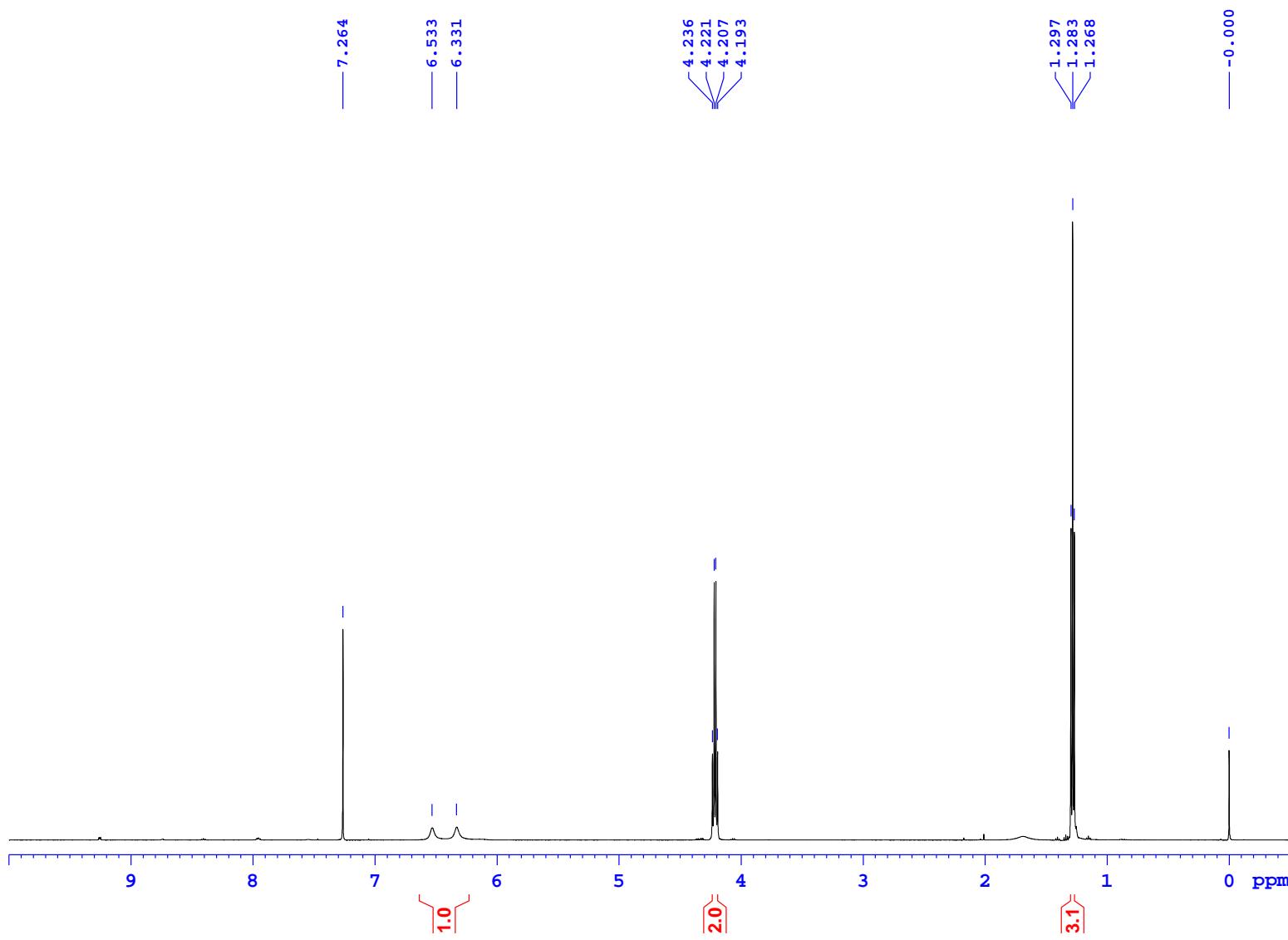
===== GRADIENT CHANNEL =====
 GPNAM1 SMSQ10.100
 GPNAM2 SMSQ10.100
 GPNAM3 SMSQ10.100
 GPZ1 70.00 %
 GPZ2 30.00 %
 GPZ3 50.10 %
 P16 1000.00 usec

F1 - Acquisition parameters
 TD 128
 SFO1 50.68533 MHz
 FIDRES 158.391663 Hz
 SW 400.000 ppm
 FnMODE QF

F2 - Processing parameters
 SI 2048
 SF 500.1300102 MHz
 WDW SINE
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.40

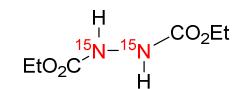
F1 - Processing parameters
 SI 1024
 MC2 QF
 SF 50.6777330 MHz
 WDW echo-antiecho
 SSB 0
 LB 0 Hz
 GB 0

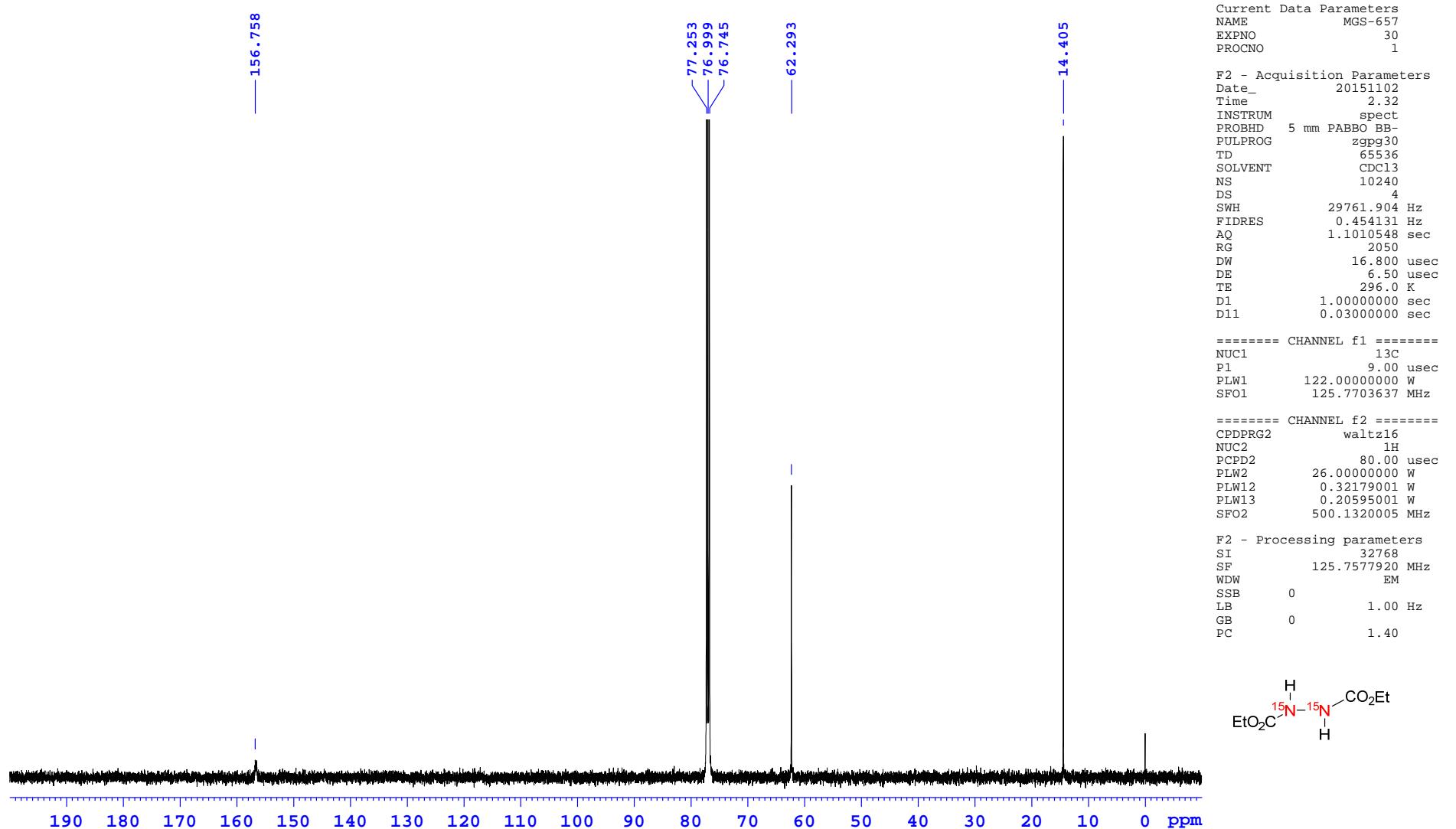


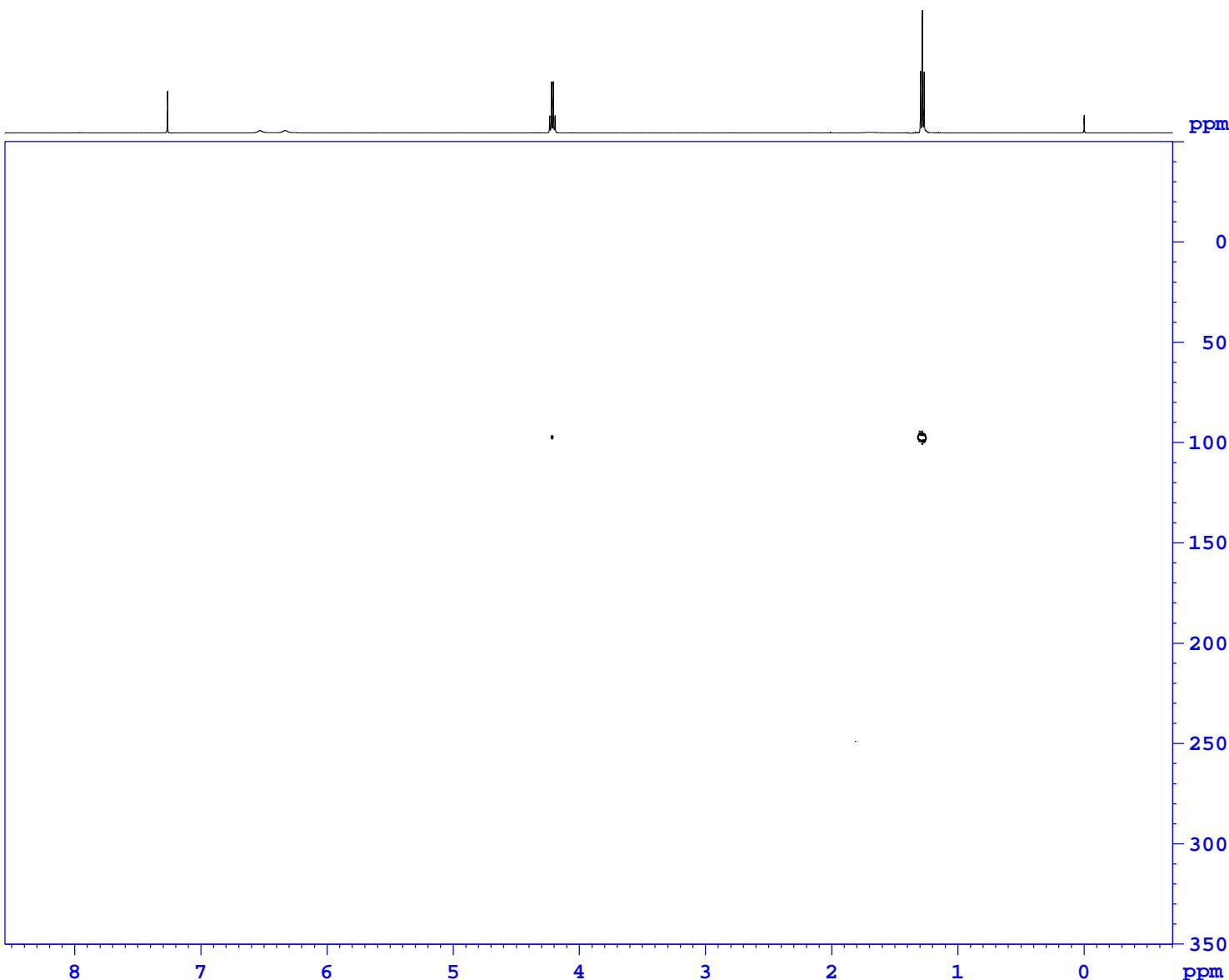


Current Data Parameters
NAME MGS-657
EXPNO 1
PROCNO 1
F2 - Acquisition Parameters
Date_ 20151028
Time 6.00
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 161
DW 48.400 usec
DE 6.50 usec
TE 296.0 K
D1 1.0000000 sec
===== CHANNEL f1 =====
NUC1 1H
P1 8.90 usec
PLW1 26.0000000 W
SF01 500.1330885 MHz

F2 - Processing parameters
SI 65536
SF 500.1300115 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00







Current Data Parameters
NAME MGS-657
EXPNO 3
PROCNO 1

F2 - Acquisition Parameters
Date 20151028
Time 7.18
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG hmbcgpndqf
TD 2048
SOLVENT CDCl₃
NS 8
DS 16
SWH 4629.629 Hz
FIDRES 2.260561 Hz
AQ 0.2212340 sec
RG 2050
DW 108.000 usec
DE 6.50 usec
TE 296.0 K
CNST13 5.0000000
D0 0.00000300 sec
D1 1.93610203 sec
D6 0.1000000 sec
D16 0.00020000 sec
INO 0.00002465 sec

===== CHANNEL f1 =====
NUC1 1H
P1 8.90 usec
P2 17.80 usec
PLW1 26.0000000 W
SFO1 500.1319747 MHz

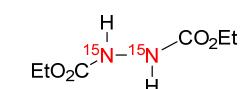
===== CHANNEL f2 =====
NUC2 15N
P3 14.40 usec
PLW2 206.0000000 W
SFO2 50.6853342 MHz

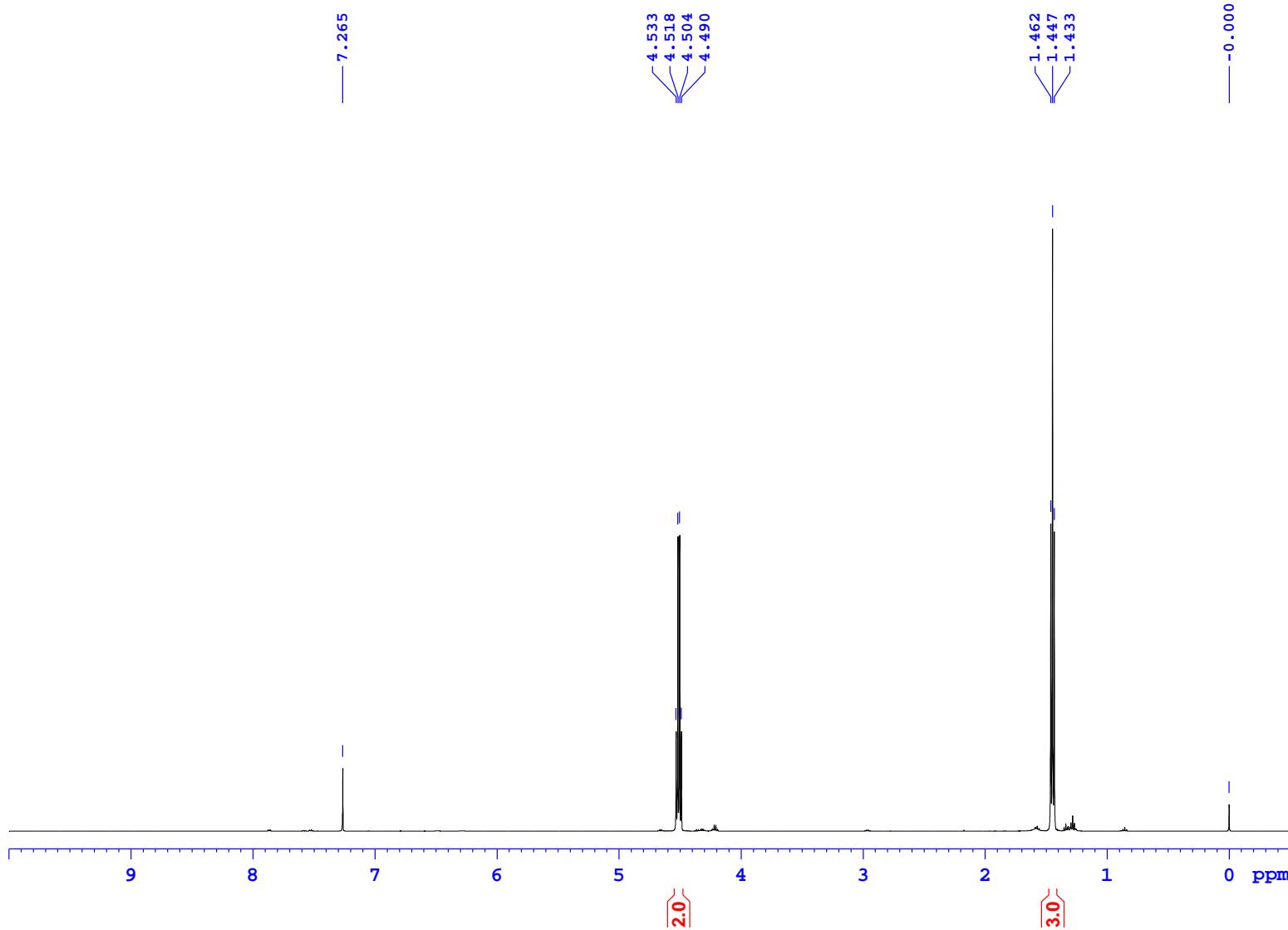
===== GRADIENT CHANNEL =====
GPNAME1 SMSQ10.100
GPNAME2 SMSQ10.100
GPNAME3 SMSQ10.100
GPZ1 70.00 %
GPZ2 30.00 %
GPZ3 50.10 %
P16 1000.00 usec

F1 - Acquisition parameters
TD 256
SFO1 50.68533 MHz
FIDRES 79.195831 Hz
SW 400.000 ppm
FnMODE QF

F2 - Processing parameters
SI 2048
SF 500.1300115 MHz
WDW SINE
SSB 0
LB 0 Hz
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
MC2 QF
SF 50.6777330 MHz
WDW echo-antiecho
SSB 0
LB 0 Hz
GB 0



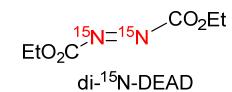


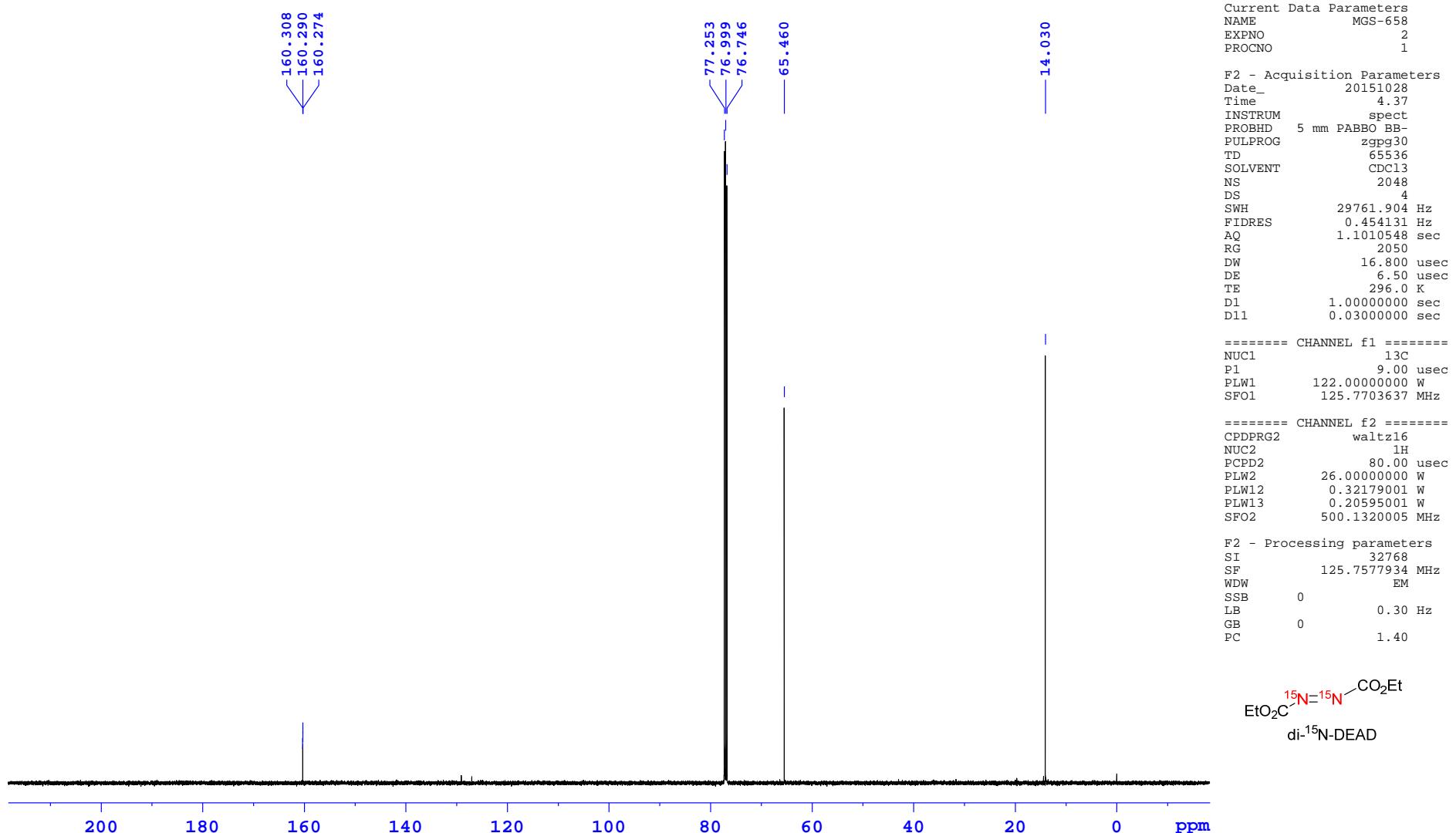
Current Data Parameters
 NAME MGS-658
 EXPNO 1
 PROCNO 1

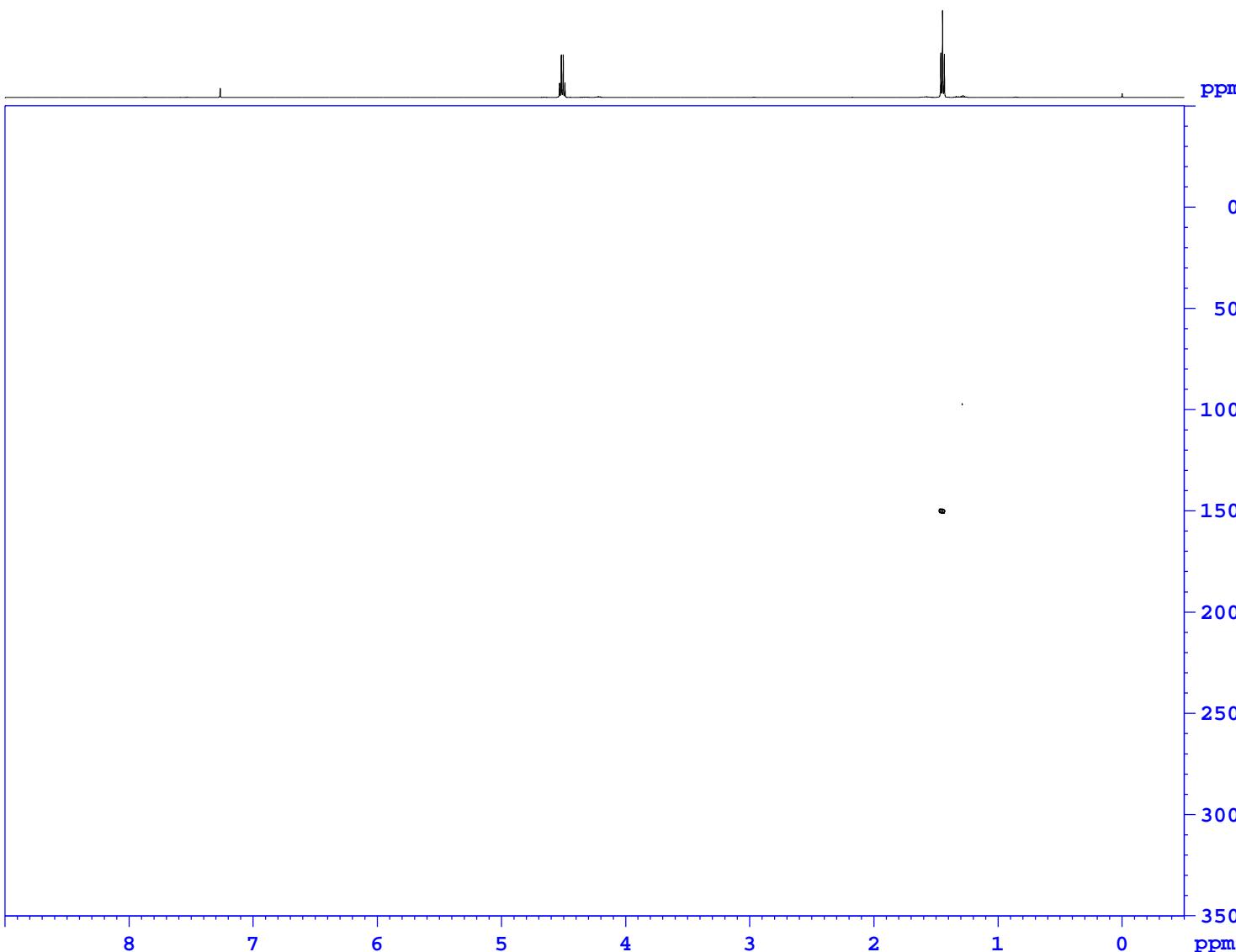
F2 - Acquisition Parameters
 Date_ 20151028
 Time 3.21
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 10330.578 Hz
 FIDRES 0.157632 Hz
 AQ 3.1719923 sec
 RG 128
 DW 48.400 usec
 DE 6.50 usec
 TE 296.0 K
 D1 1.0000000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 8.90 usec
 PLW1 26.0000000 W
 SF01 500.1330885 MHz

F2 - Processing parameters
 SI 65536
 SF 500.1300108 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00







Current Data Parameters
NAME MGS-658
EXPNO 3
PROCNO 1

F2 - Acquisition Parameters
Date 20151028
Time 4.39
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG hmbcgpndqf
TD 2048
SOLVENT CDCl₃
NS 8
DS 16
SWH 2415.459 Hz
FIDRES 1.179423 Hz
AQ 0.4239860 sec
RG 2050
DW 207.000 usec
DE 6.50 usec
TE 296.0 K
CNST13 5.0000000
D0 0.00000300 sec
D1 1.73335004 sec
D6 0.10000000 sec
D16 0.00020000 sec
INO 0.00002465 sec

===== CHANNEL f1 =====
NUC1 1H
P1 8.90 usec
P2 17.80 usec
PLW1 26.0000000 W
SFO1 500.1312404 MHz

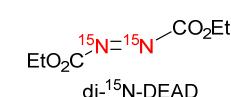
===== CHANNEL f2 =====
NUC2 15N
P3 14.40 usec
PLW2 206.0000000 W
SFO2 50.6853342 MHz

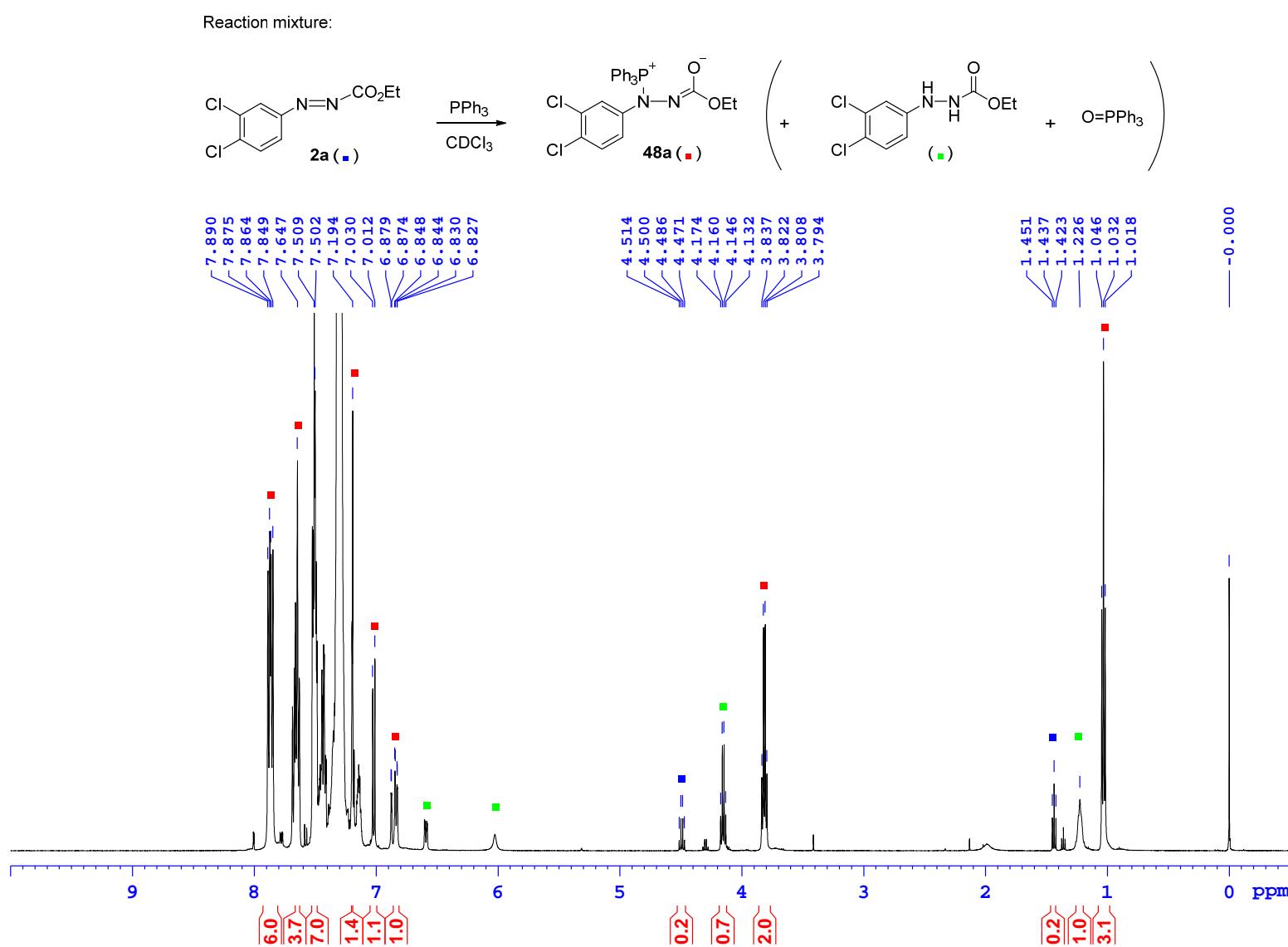
===== GRADIENT CHANNEL =====
GPNAME1 SMSQ10.100
GPNAME2 SMSQ10.100
GPNAME3 SMSQ10.100
GPZ1 70.00 %
GPZ2 30.00 %
GPZ3 50.10 %
P16 1000.00 usec

F1 - Acquisition parameters
TD 256
SFO1 50.68533 MHz
FIDRES 79.195831 Hz
SW 400.000 ppm
FnMODE QF

F2 - Processing parameters
SI 2048
SF 500.1300108 MHz
WDW SINE
SSB 0
LB 0 Hz
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
MC2 QF
SF 50.6777330 MHz
WDW echo-antiecho
SSB 0
LB 0 Hz
GB 0



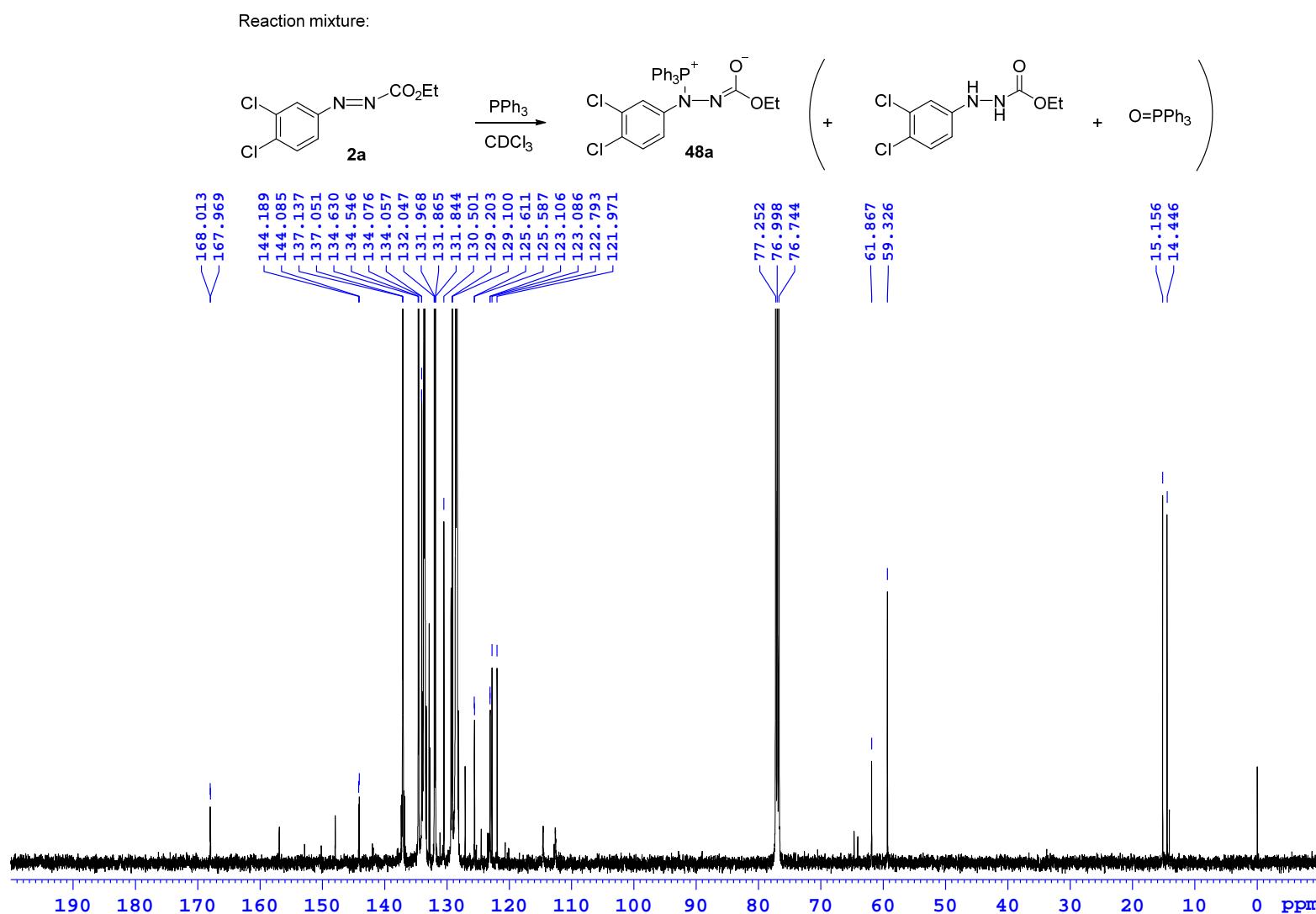


Current Data Parameters
NAME mgs-583
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150613
Time 13.07
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 32
DW 48.400 usec
DE 6.50 usec
TE 296.0 K
D1 1.0000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 8.90 usec
PLW1 26.0000000 W
SFO1 500.1330885 MHz

F2 - Processing parameters
SI 65536
SF 500.1300468 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



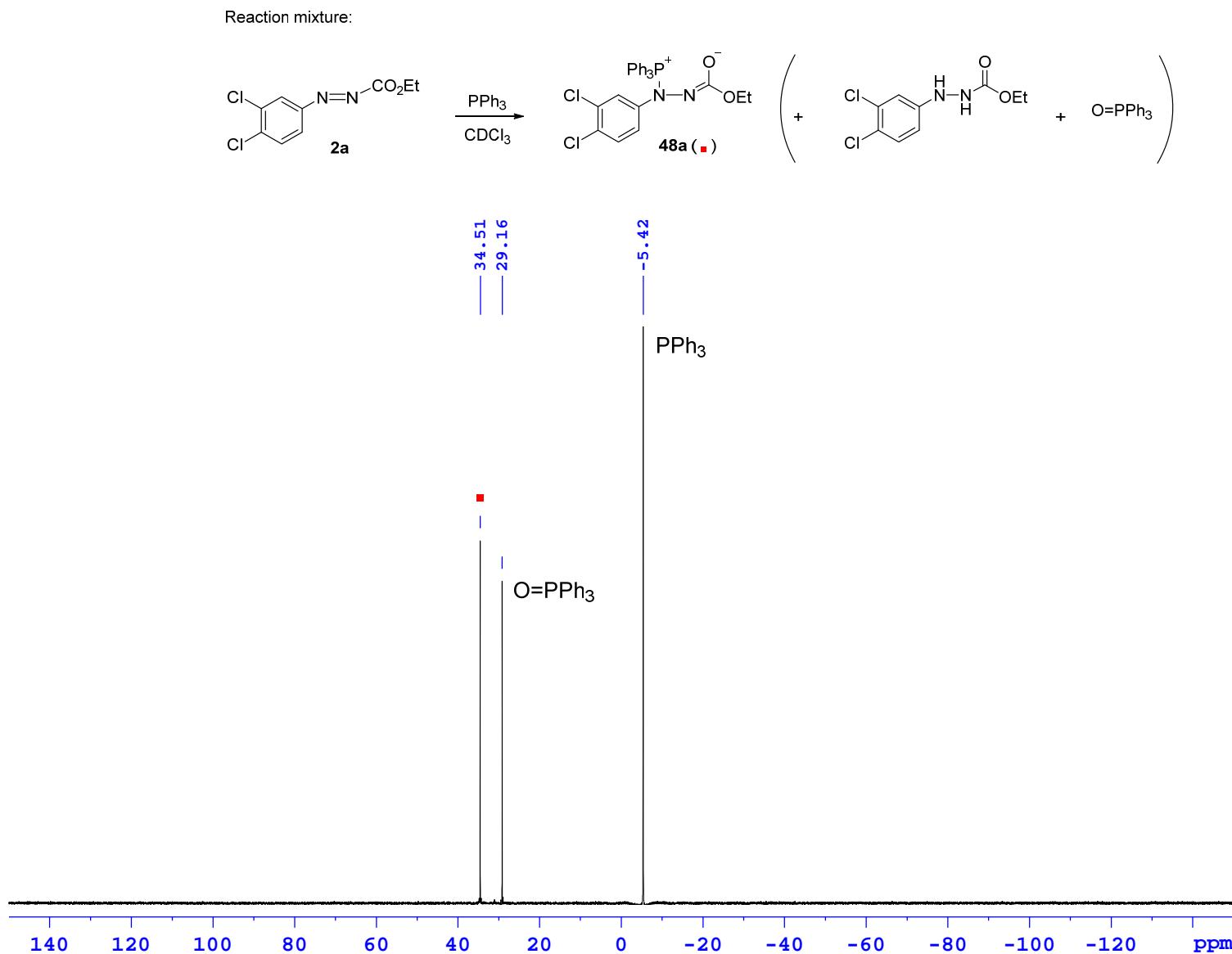
Current Data Parameters
 NAME mgs-583
 EXPNO 10
 PROCNO 1

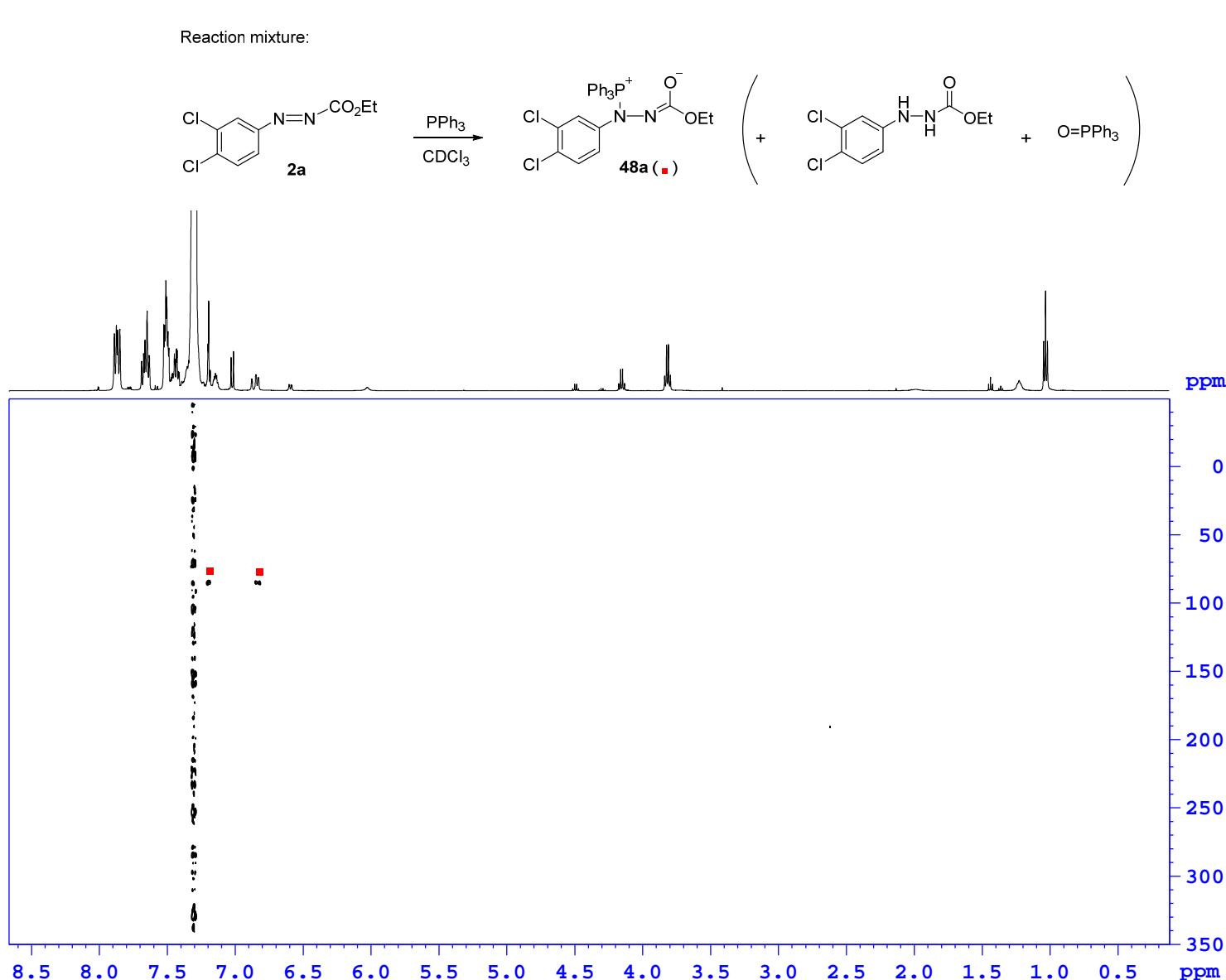
F2 - Acquisition Parameters
 Date_ 20150614
 Time 10.46
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 8192
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.454131 Hz
 AQ 1.1010548 sec
 RG 2050
 DW 16.800 usec
 DE 6.50 usec
 TE 296.0 K
 D1 1.0000000 sec
 D11 0.0300000 sec

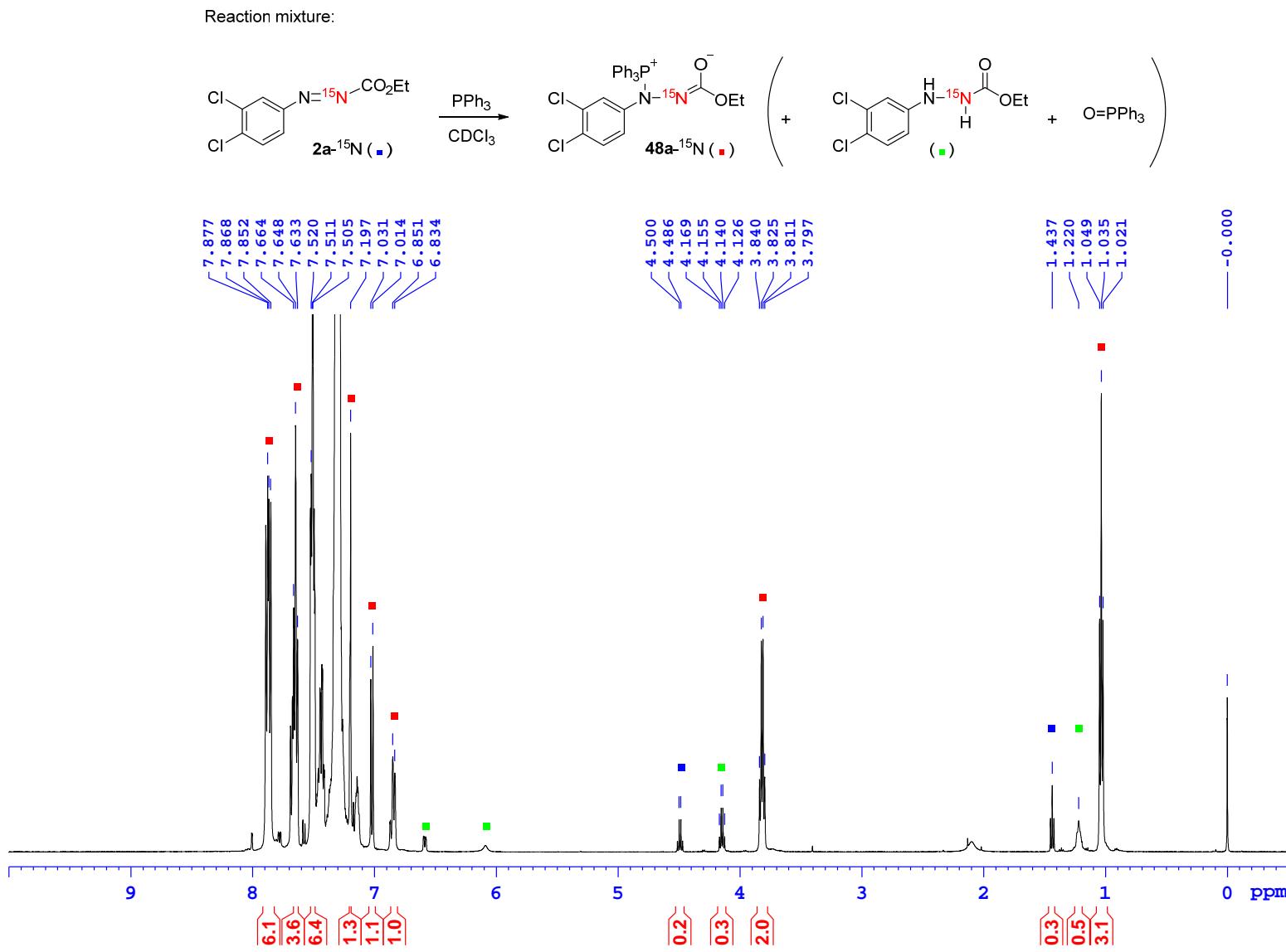
===== CHANNEL f1 =====
 NUC1 13C
 P1 9.00 usec
 PLW1 122.00000000 W
 SFO1 125.7703637 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PLW2 26.00000000 W
 PLW12 0.32179001 W
 PLW13 0.20595001 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7578100 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40







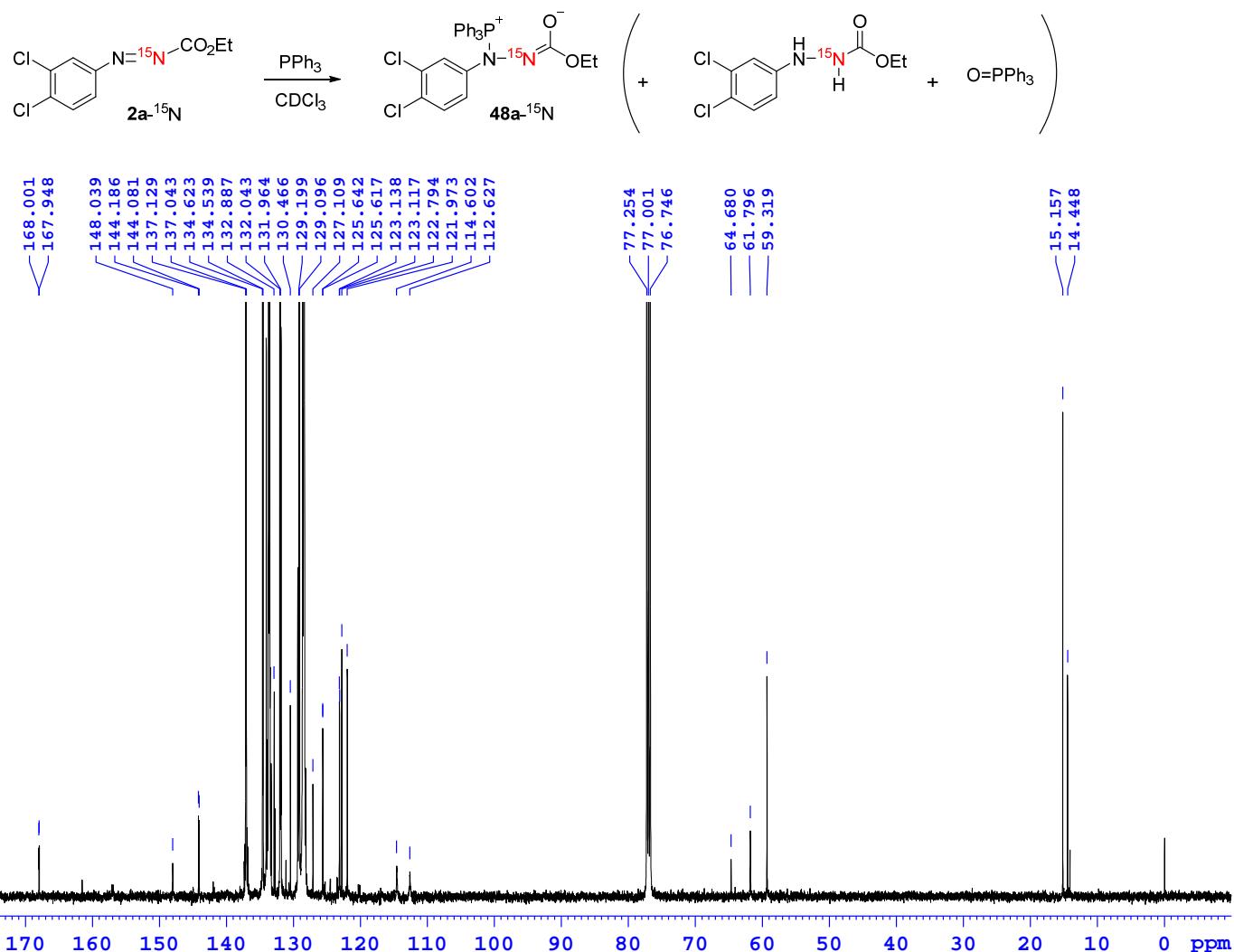
Current Data Parameters
 NAME mgs-576
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150606
 Time 13.24
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 10330.578 Hz
 FIDRES 0.157632 Hz
 AQ 3.1719923 sec
 RG 32
 DW 48.400 usec
 DE 6.50 usec
 TE 296.0 K
 D1 1.0000000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 8.90 usec
 PLW1 26.0000000 W
 SF01 500.1330885 MHz

F2 - Processing parameters
 SI 65536
 SF 500.1300453 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

Reaction mixture:



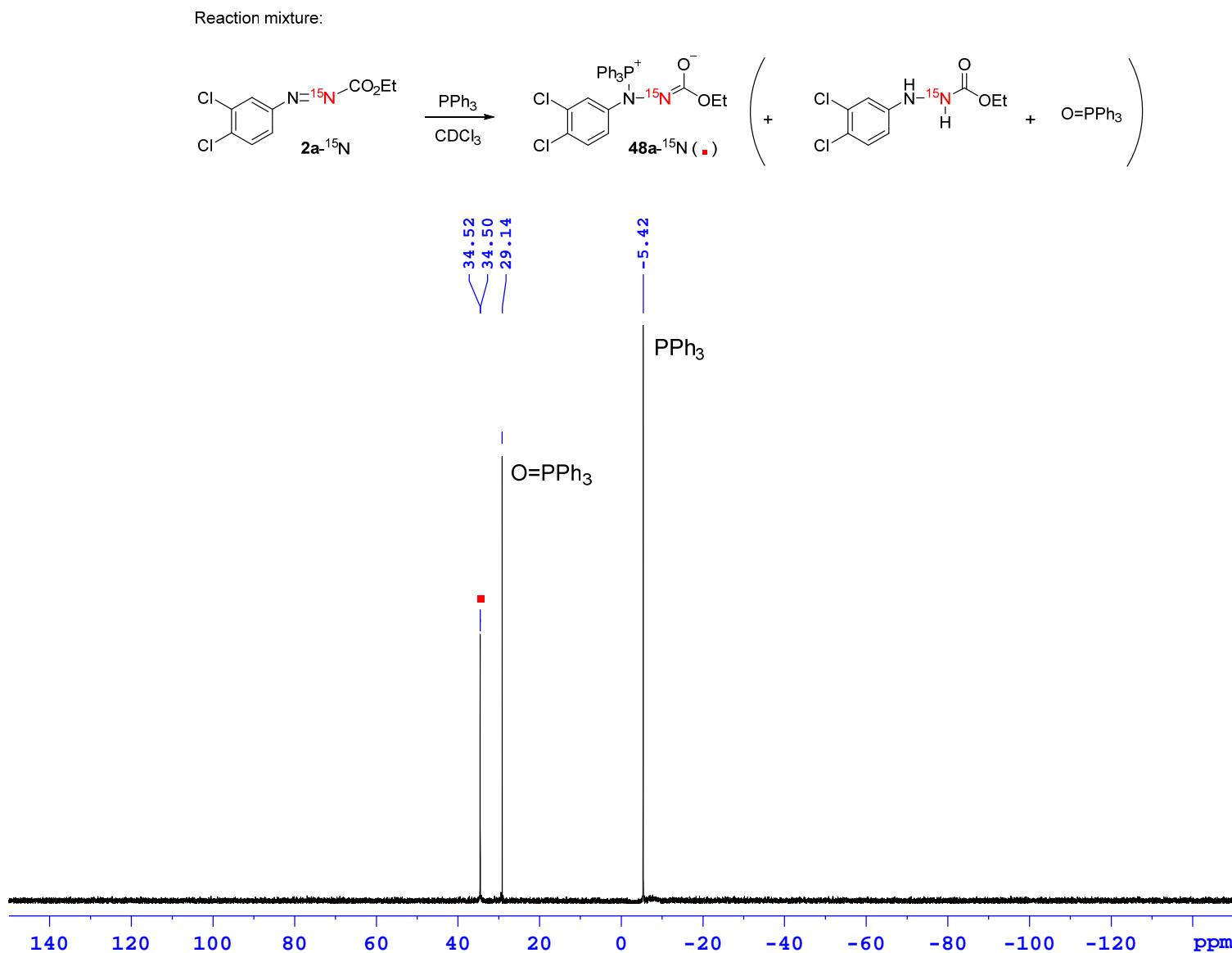
Current Data Parameters
NAME mgs-576
EXPNO 22
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150606
Time 20.16
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 11264
DS 4
SWH 29761.904 Hz
FIDRES 0.454131 Hz
AQ 1.1010548 sec
RG 2050
DW 16.800 usec
DE 6.50 usec
TE 296.0 K
D1 1.0000000 sec
D11 0.0300000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 9.00 usec
PLW1 122.0000000 W
SFO1 125.7703637 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PLW2 26.0000000 W
PLW12 0.32179001 W
PLW13 0.20595001 W
SFO2 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 125.7578104 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



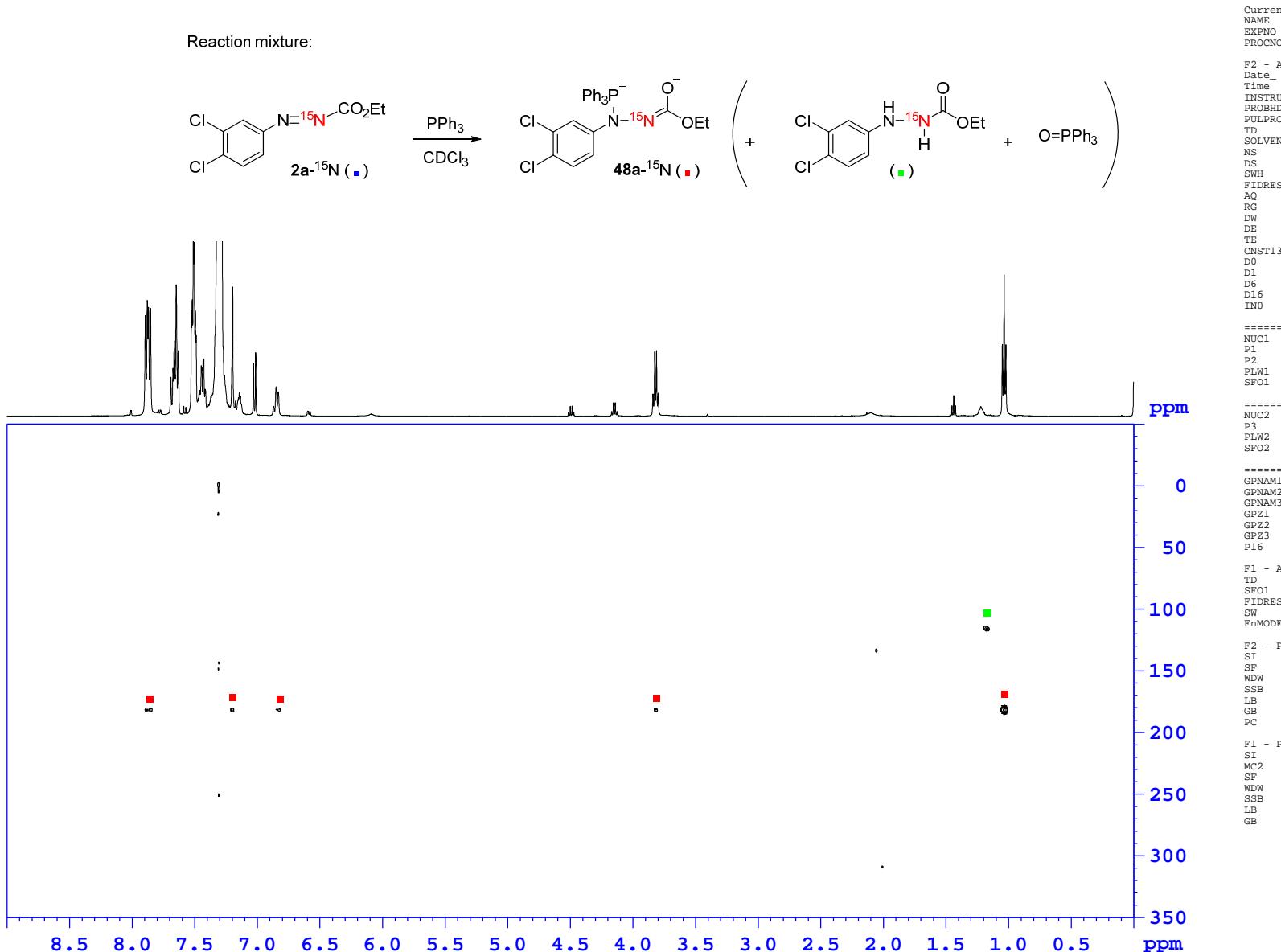
Current Data Parameters
NAME mgs-576
EXPNO 12
PROCNO 1

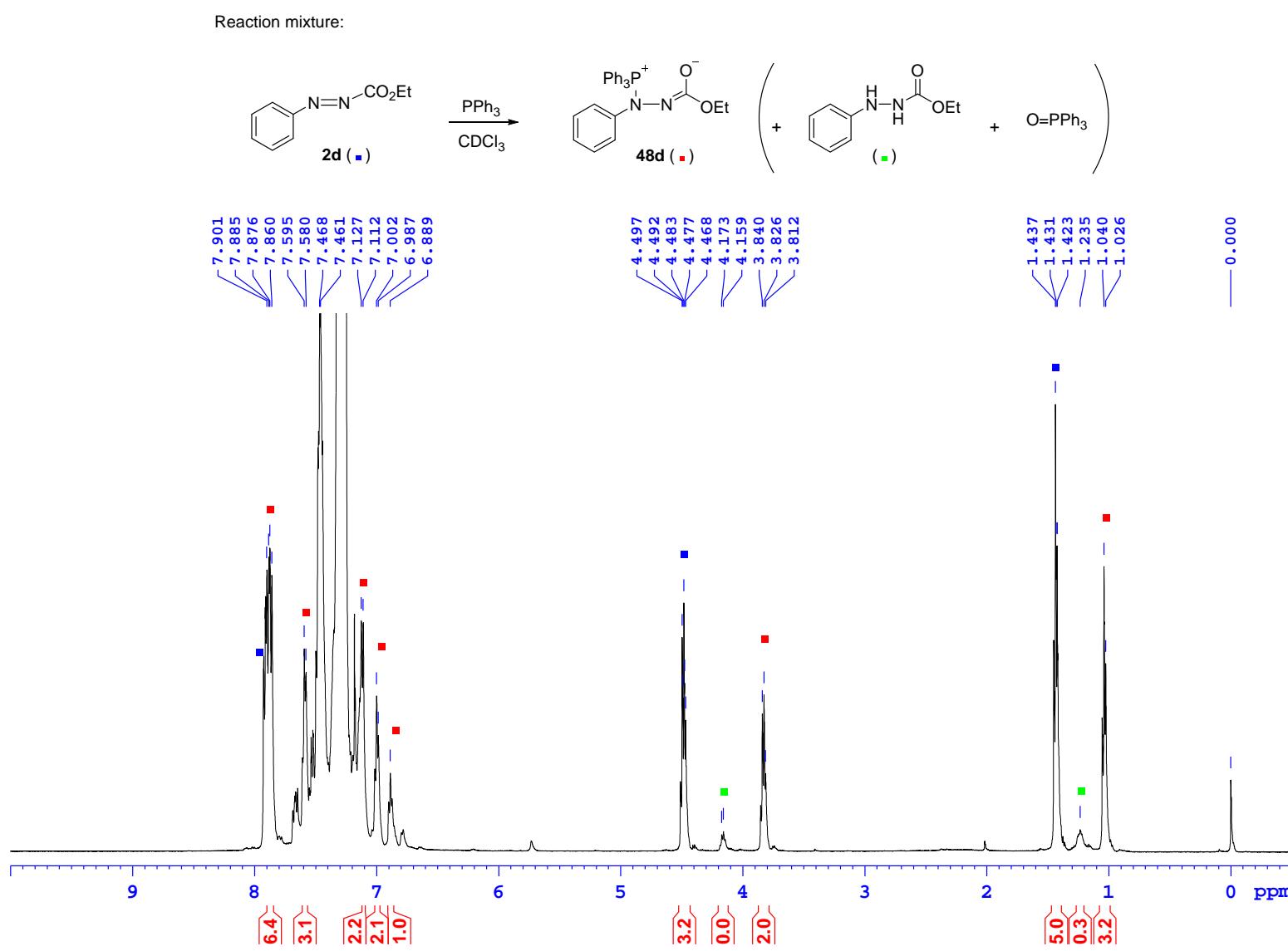
F2 - Acquisition Parameters
Date_ 20150606
Time 13.32
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDC13
NS 25
DS 4
SWH 81521.742 Hz
FIDRES 1.243923 Hz
AQ 0.4020041 sec
RG 2050
DW 6.133 usec
DE 6.50 usec
TE 296.1 K
D1 2.0000000 sec
D11 0.03000000 sec

===== CHANNEL f1 =====
NUC1 31P
P1 10.00 usec
PLW1 100.0000000 W
SFO1 202.4462121 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PLW2 26.00000000 W
PLW12 0.32179001 W
PLW13 0.20595001 W
SFO2 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 202.4563350 MHz
WDW EM
SSB 0
LB 0.25 Hz
GB 0
PC 1.40



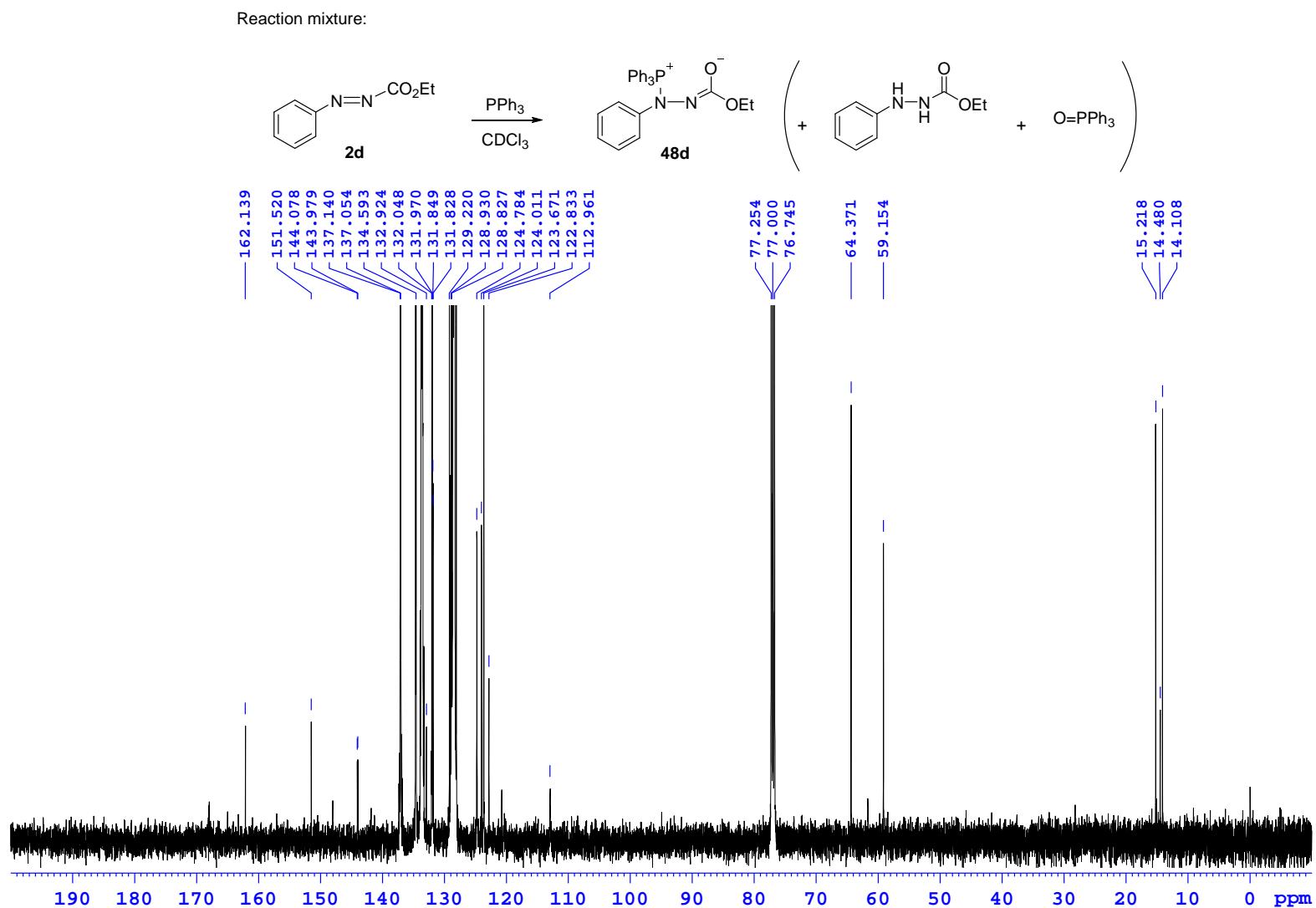


Current Data Parameters
NAME MGS-661
EXPNO 20
PROCNO 1

F2 - Acquisition Parameters
Date_ 20151103
Time 9.06
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 32
DW 48.400 usec
DE 6.50 usec
TE 296.0 K
D1 1.0000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 8.90 usec
PLW1 26.0000000 W
SF01 500.1330885 MHz

F2 - Processing parameters
SI 65536
SF 500.1300539 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



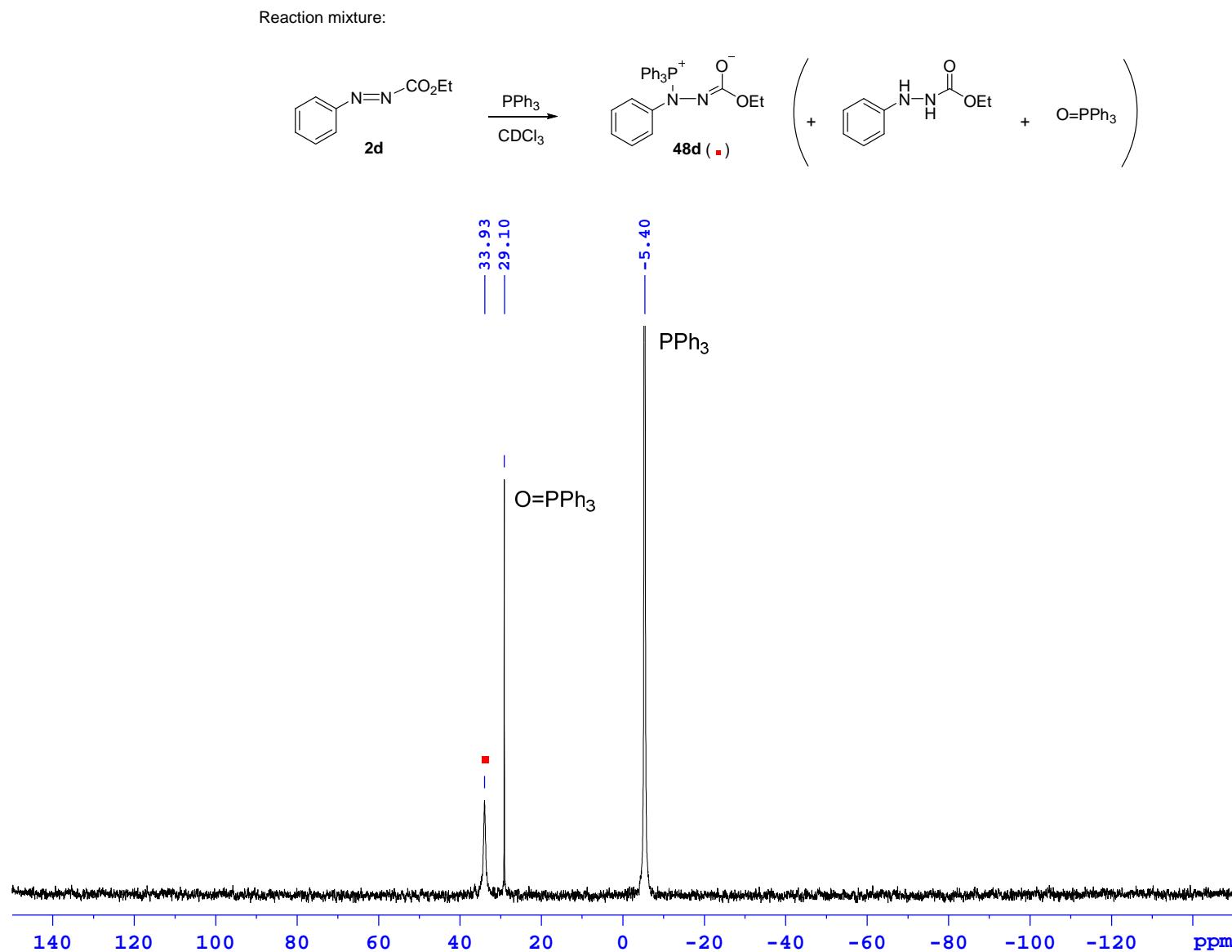
Current Data Parameters
NAME MGS-661
EXPNO 25
PROCNO 1

F2 - Acquisition Parameters
Date_ 20151103
Time 12.14
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 2048
DS 4
SWH 29761.904 Hz
FIDRES 0.454131 Hz
AQ 1.1010548 sec
RG 2050
DW 16.800 usec
DE 6.50 usec
TE 296.0 K
D1 1.0000000 sec
D11 0.0300000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 9.00 usec
PLW1 122.0000000 W
SFO1 125.7703637 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PLW2 26.0000000 W
PLW12 0.32179001 W
PLW13 0.20595001 W
SFO2 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 125.7578114 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



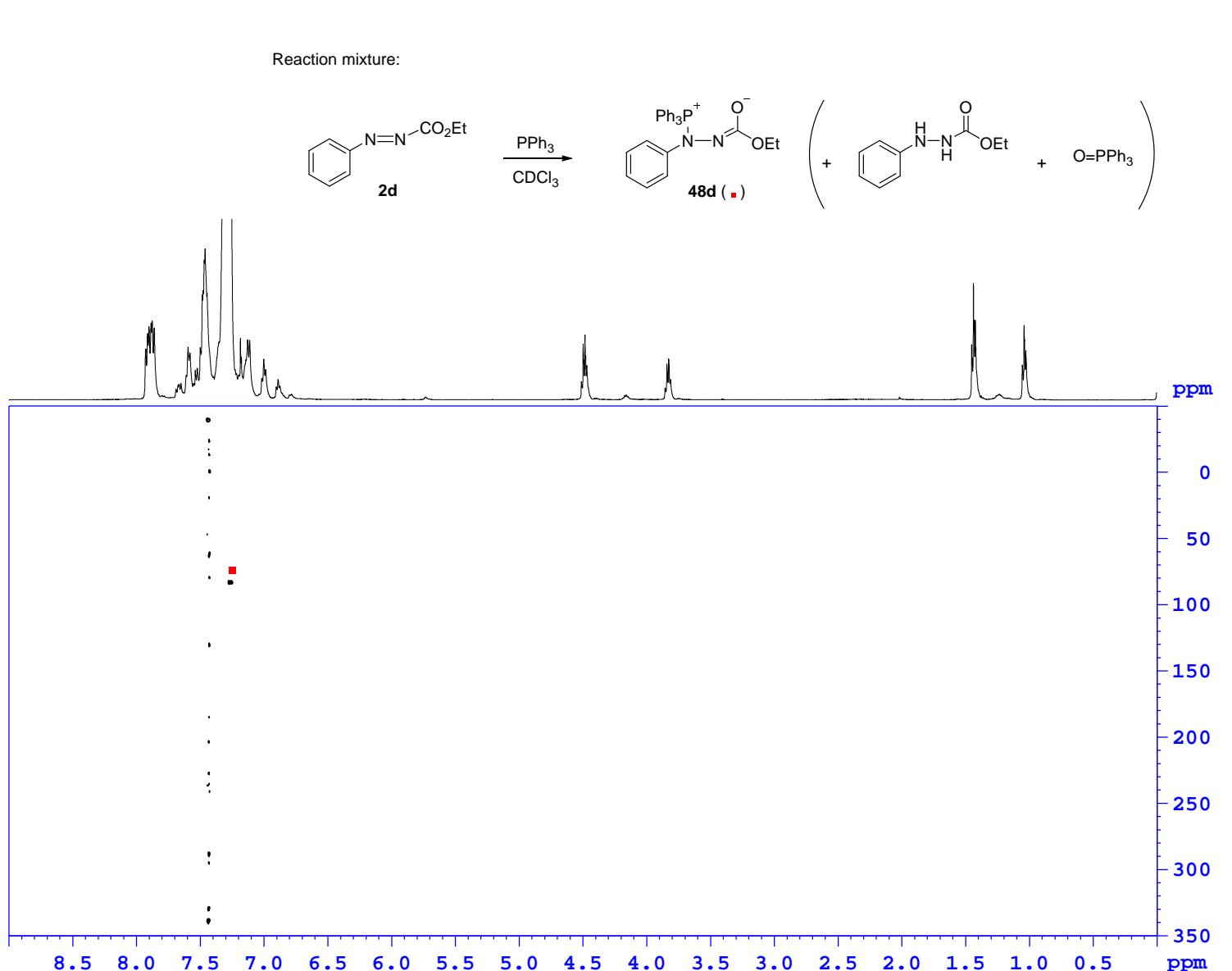
Current Data Parameters
NAME MGS-472
EXPNO 5
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150217
Time 18.38
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDC13
NS 16
DS 4
SWH 81521.742 Hz
FIDRES 1.243923 Hz
AQ 0.4020041 sec
RG 1440
DW 6.133 usec
DE 6.50 usec
TE 296.1 K
D1 2.0000000 sec
D11 0.0300000 sec

===== CHANNEL f1 =====
NUC1 31P
P1 10.00 usec
PLW1 100.0000000 W
SFO1 202.4462121 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PLW2 26.0000000 W
PLW12 0.32179001 W
PLW13 0.20595001 W
SFO2 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 202.4563350 MHz
WDW EM
SSB 0
LB 10.00 Hz
GB 0
PC 1.40



Current Data Parameters
NAME MGS-472
EXPNO 6
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150217
Time_ 18.51
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG hmbcgpndif
TD 2048
SOLVENT CDCl3
NS 128
DS 16
SWH 4273.504 Hz
FIDRES 2.086672 Hz
AQ 0.2396660 sec
RG 2050
DW 117.000 usec
DE 6.50 usec
TE 296.0 K
CNST13 5.0000000
D0 0.00000300 sec
D1 1.91767001 sec
D6 0.10000000 sec
D16 0.00020000 sec
INO 0.00002465 sec

===== CHANNEL f1 =====
NUC1 1H
P1 8.90 usec
P2 17.80 usec
PLW1 26.0000000 W
SFO1 500.1323261 MHz

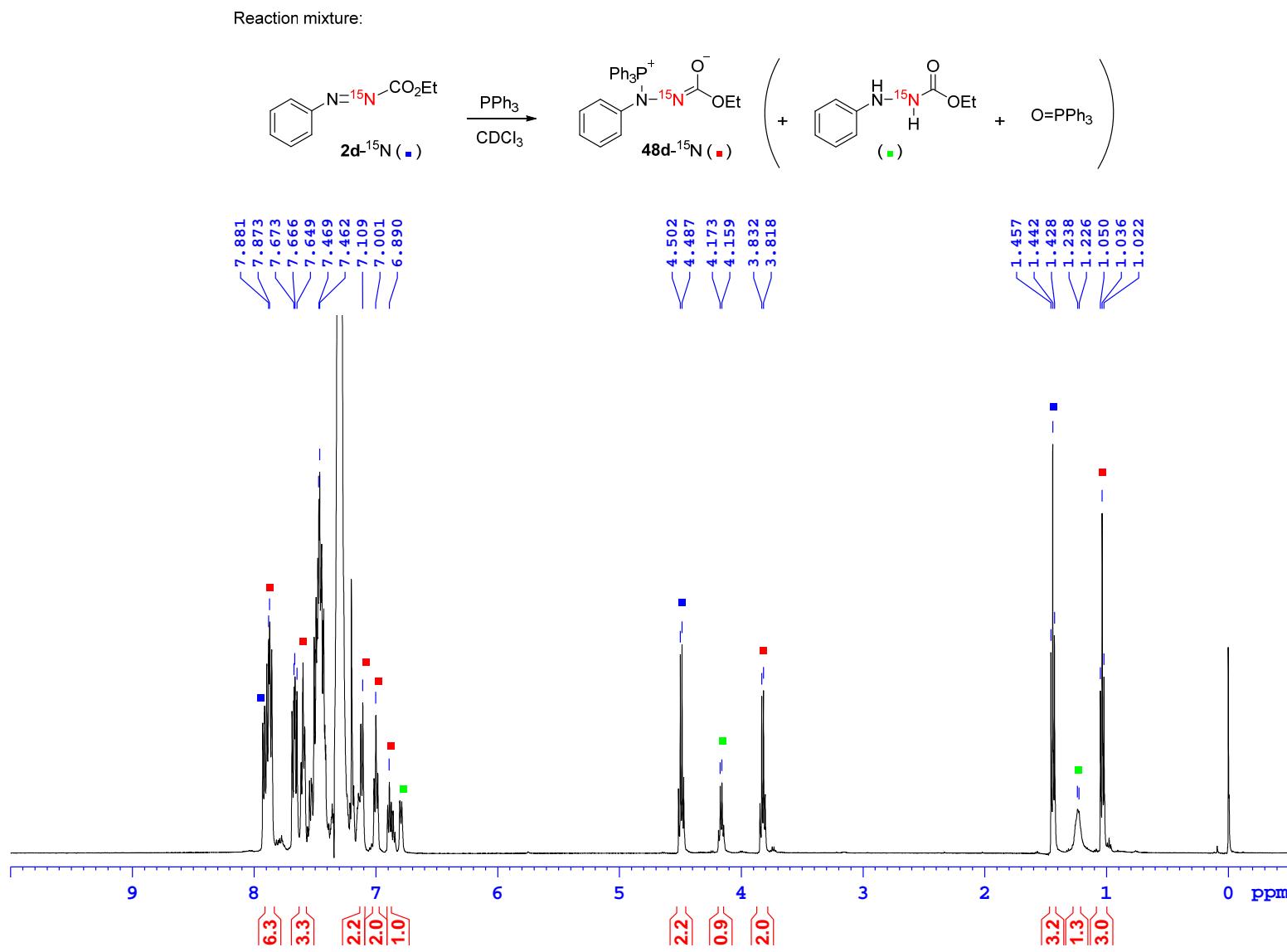
===== CHANNEL f2 =====
NUC2 15N
P3 14.40 usec
PLW2 206.0000000 W
SFO2 50.6853342 MHz

===== GRADIENT CHANNEL =====
GPNAME1 SMSQ10.100
GPNAME2 SMSQ10.100
GPNAME3 SMSQ10.100
GPZ1 70.00 %
GPZ2 30.00 %
GPZ3 50.10 %
P16 1000.00 usec

F1 - Acquisition parameters
TD 128
SFO1 50.68533 MHz
FIDRES 158.391663 Hz
SW 400.000 ppm
FnMODE QF

F2 - Processing parameters
SI 2048
SF 500.1300000 MHz
WDW SINE
SSB 0
LB 0 Hz
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
MC2 QF
SF 50.6777330 MHz
WDW echo-antiecho
SSB 0
LB 0 Hz
GB 0

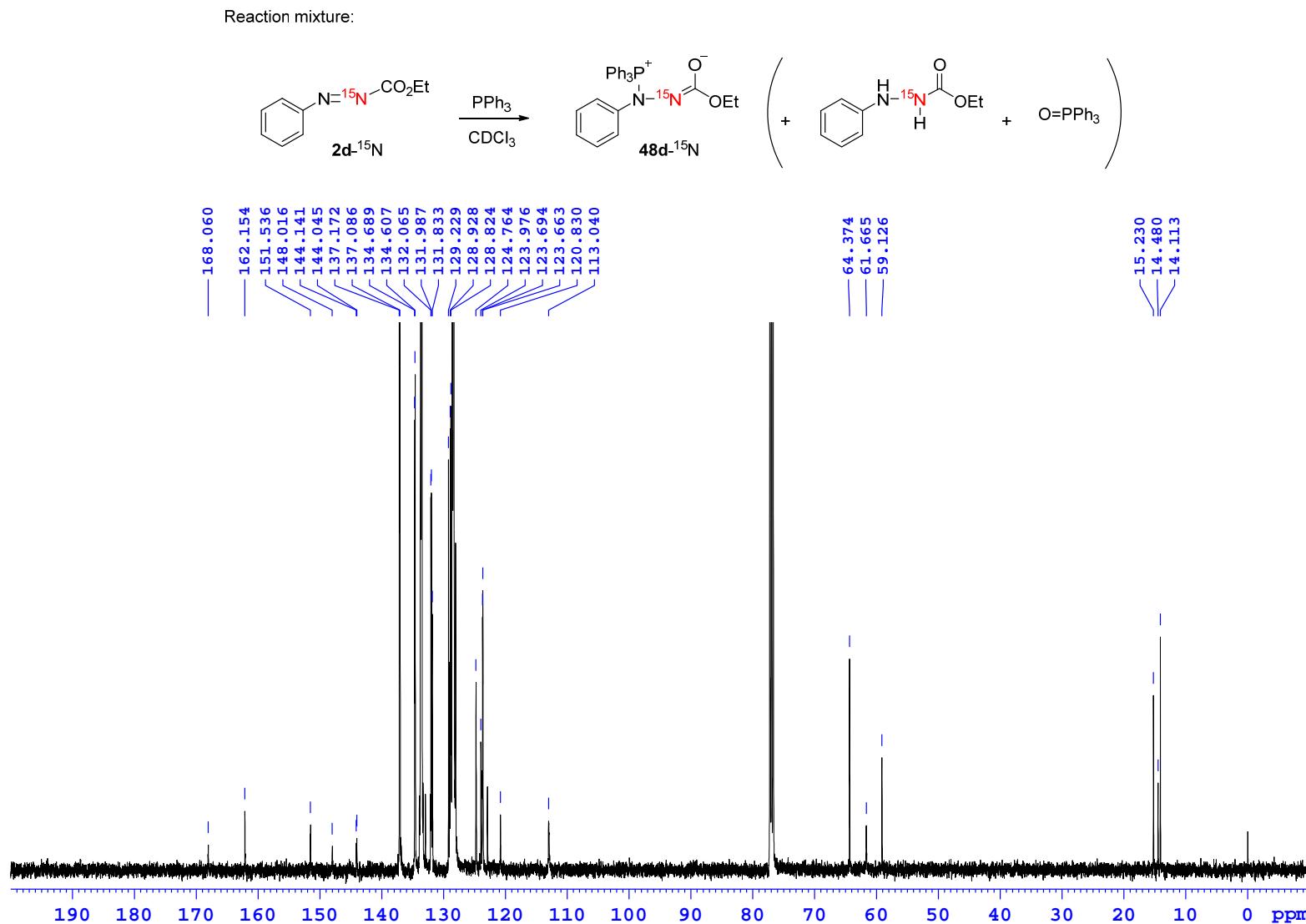


Current Data Parameters
NAME MGS-645
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20151014
Time 8.57
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 32
DW 48.400 usec
DE 6.50 usec
TE 297.2 K
D1 1.0000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 8.90 usec
PLW1 26.0000000 W
SF01 500.1330885 MHz

F2 - Processing parameters
SI 65536
SF 500.1300430 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



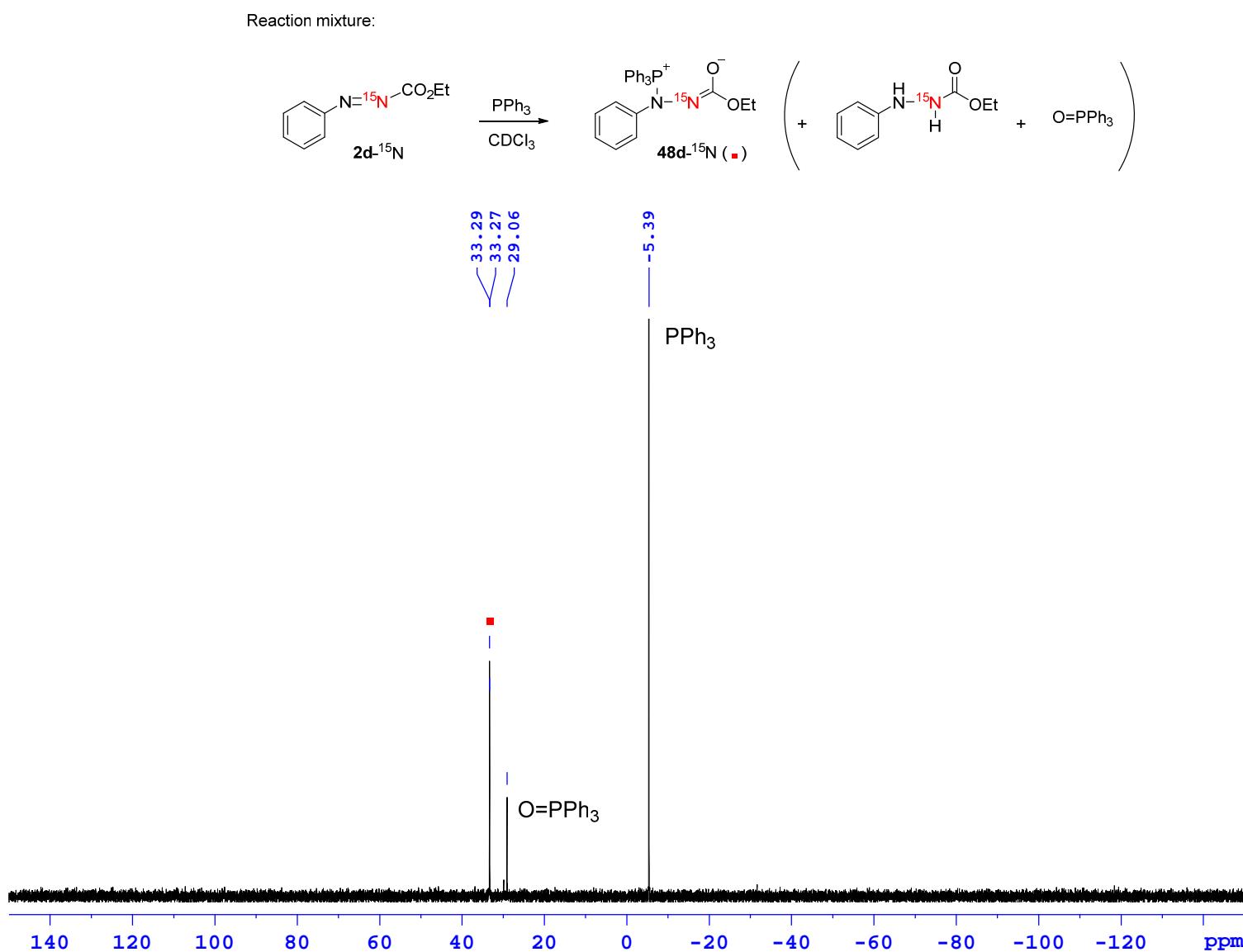
Current Data Parameters
 NAME MGS-645
 EXPNO 8
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20151013
 Time 23.39
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 5120
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.454131 Hz
 AQ 1.1010548 sec
 RG 2050
 DW 16.800 usec
 DE 6.50 usec
 TE 297.5 K
 D1 1.0000000 sec
 D11 0.0300000 sec

===== CHANNEL f1 ======
 NUC1 13C
 P1 9.00 usec
 PLW1 122.0000000 W
 SFO1 125.7703637 MHz

===== CHANNEL f2 ======
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PLW2 26.0000000 W
 PLW12 0.32179001 W
 PLW13 0.20595001 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7578067 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



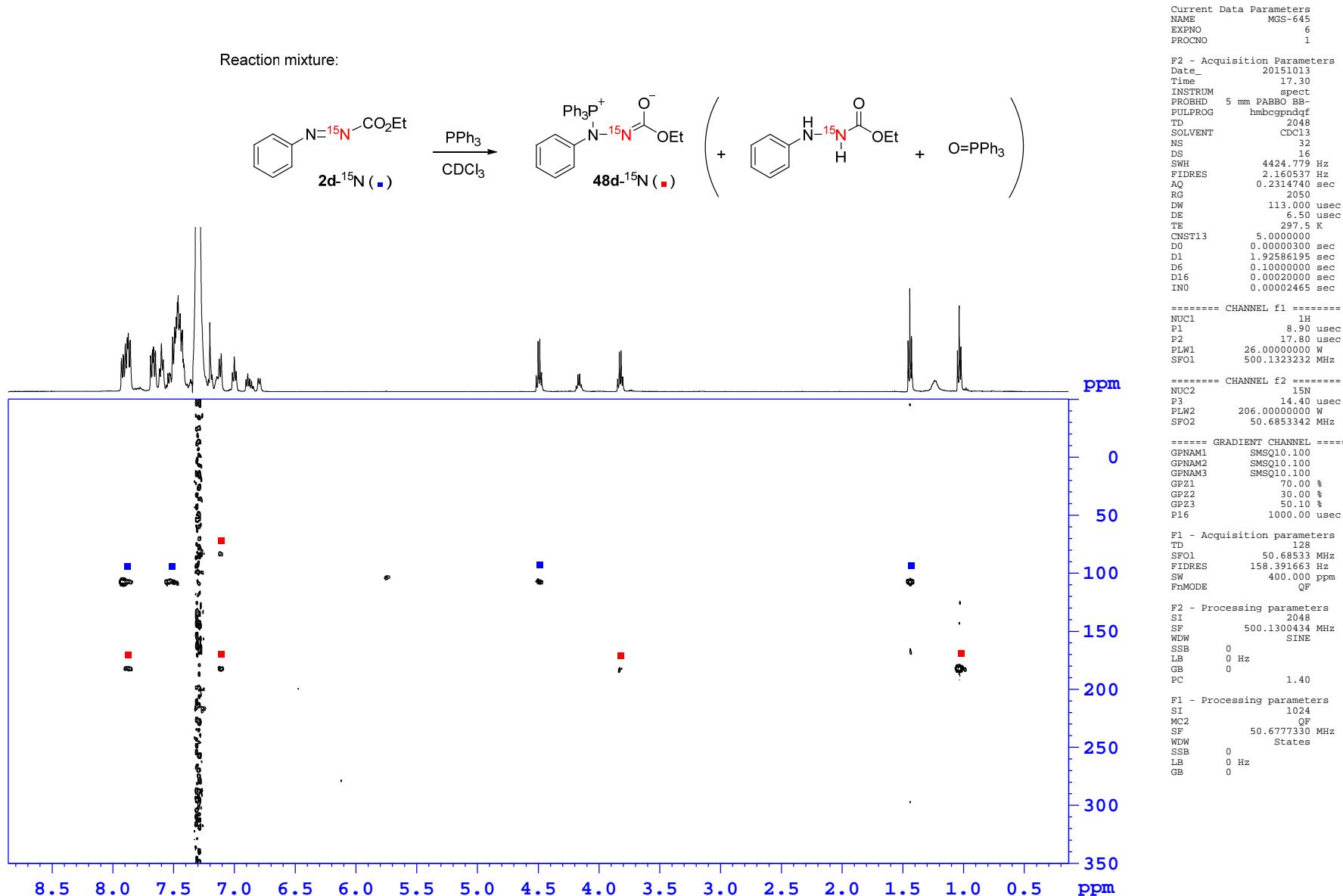
Current Data Parameters
NAME MGS-645
EXPNO 3
PROCNO 1

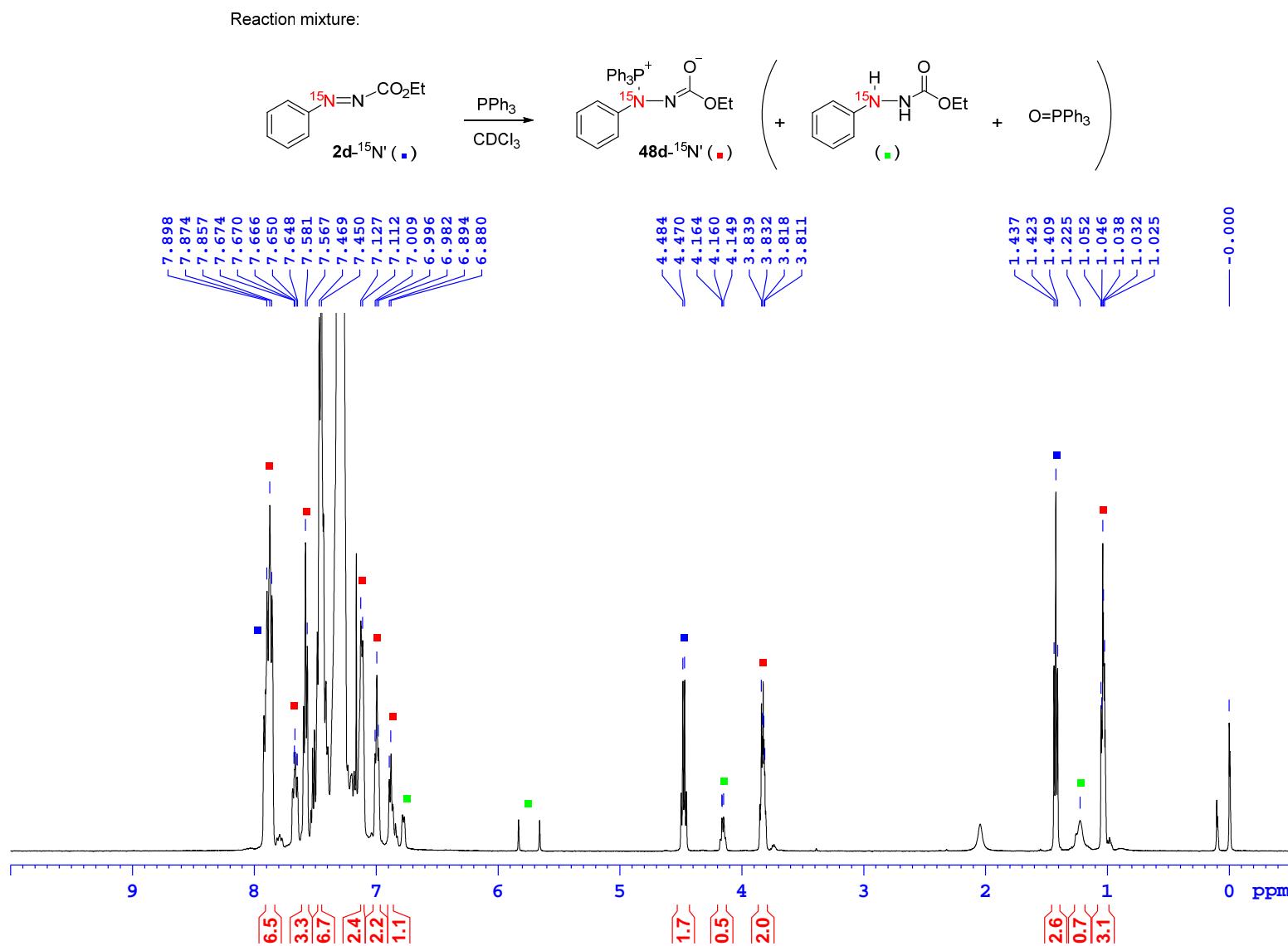
F2 - Acquisition Parameters
Date_ 20151013
Time 17.14
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDC13
NS 16
DS 4
SWH 81521.742 Hz
FIDRES 1.243923 Hz
AQ 0.4020041 sec
RG 2050
DW 6.133 usec
DE 6.50 usec
TE 297.8 K
D1 2.00000000 sec
D11 0.03000000 sec

===== CHANNEL f1 =====
NUC1 31P
P1 10.00 usec
PLW1 100.0000000 W
SFO1 202.4462121 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PLW2 26.00000000 W
PLW12 0.32179001 W
PLW13 0.20959001 W
SFO2 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 202.4563350 MHz
WDW no
SSB 0
LB 0 Hz
GB 0
PC 1.40



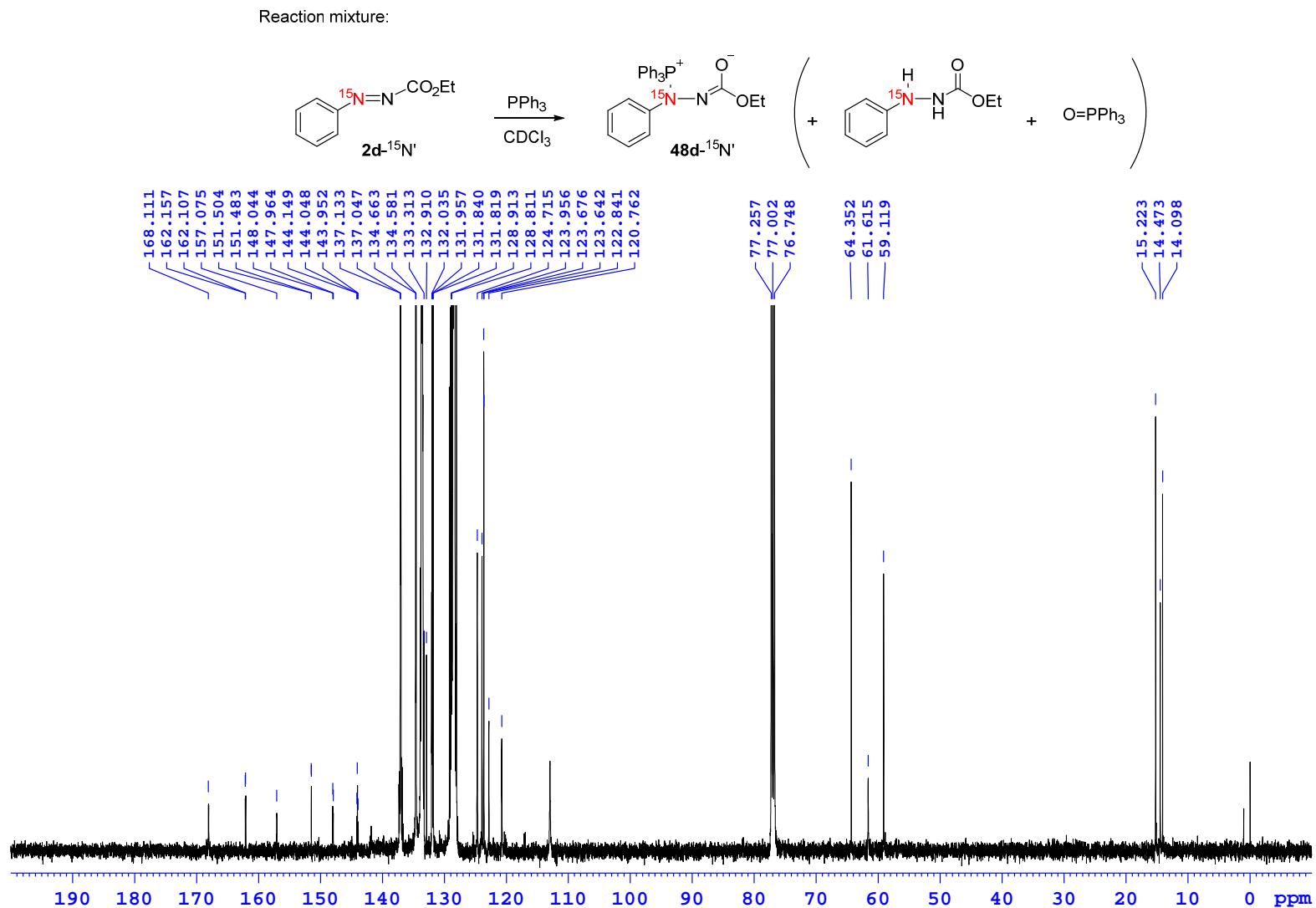


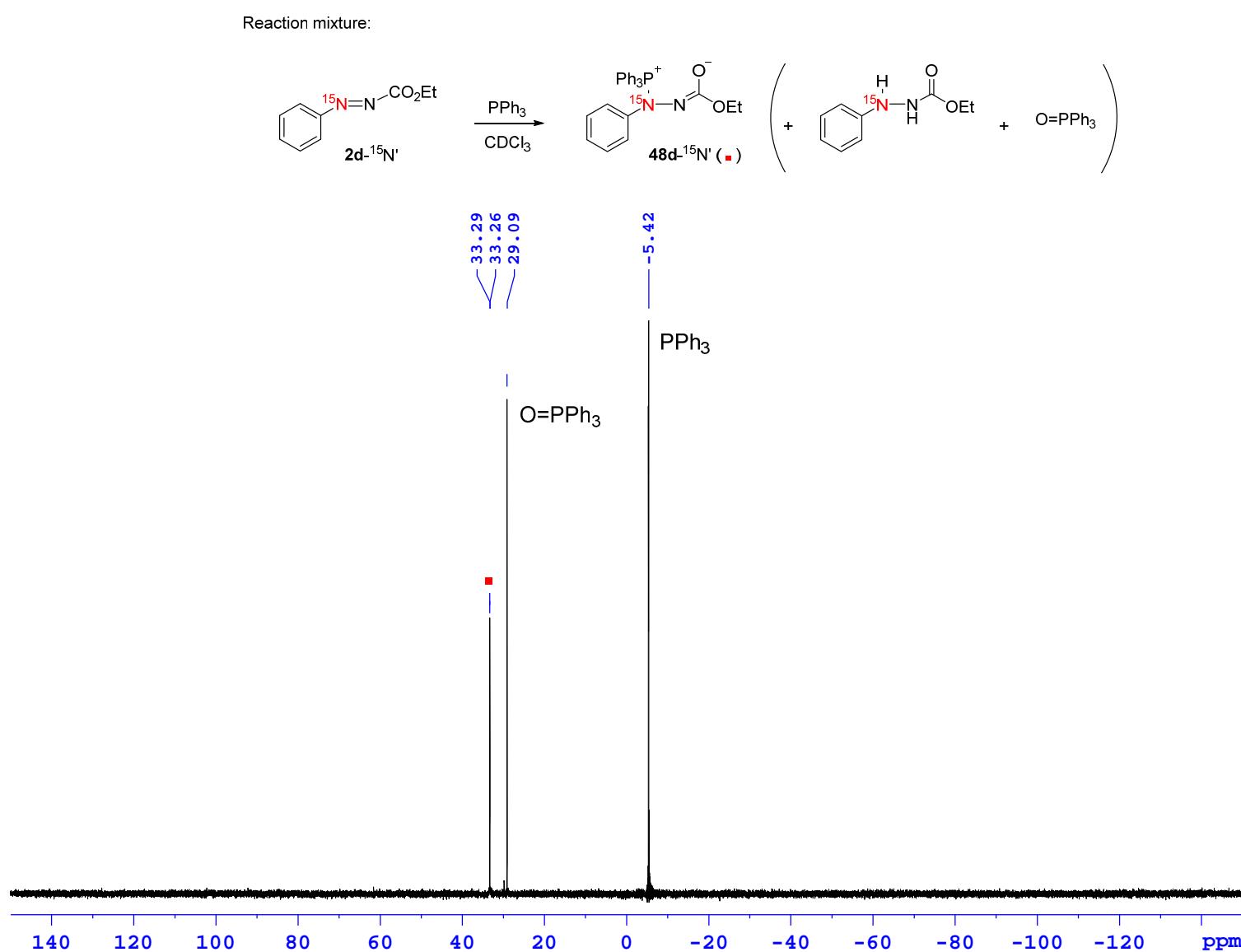
Current Data Parameters
NAME MGS-614
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150730
Time 16.24
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 20.2
DW 48.400 usec
DE 6.50 usec
TE 296.0 K
D1 1.0000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 8.90 usec
PLW1 26.0000000 W
SF01 500.1330885 MHz

F2 - Processing parameters
SI 65536
SF 500.1300618 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00





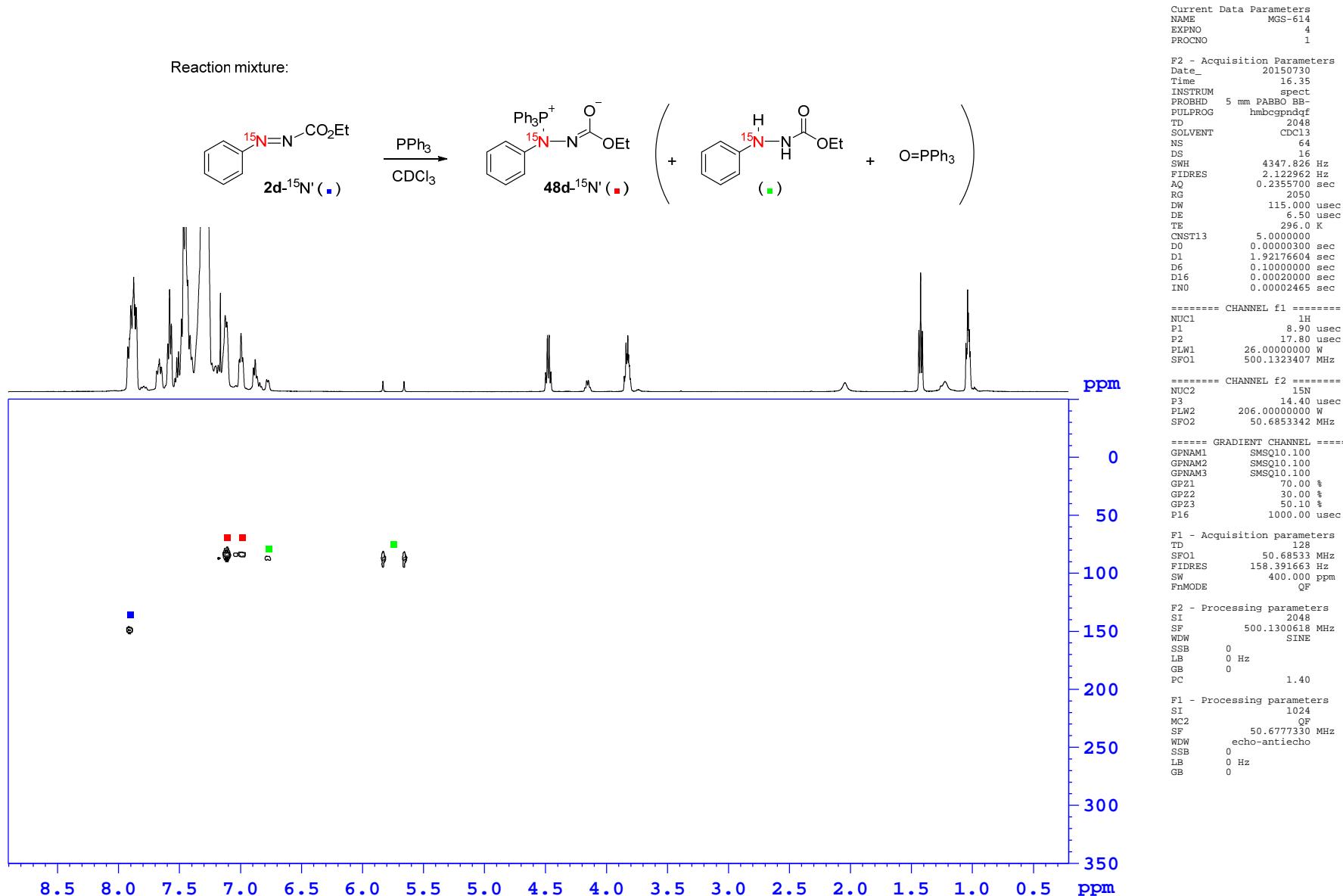
Current Data Parameters
NAME MGS-614
EXPNO 7
PROCNO 1

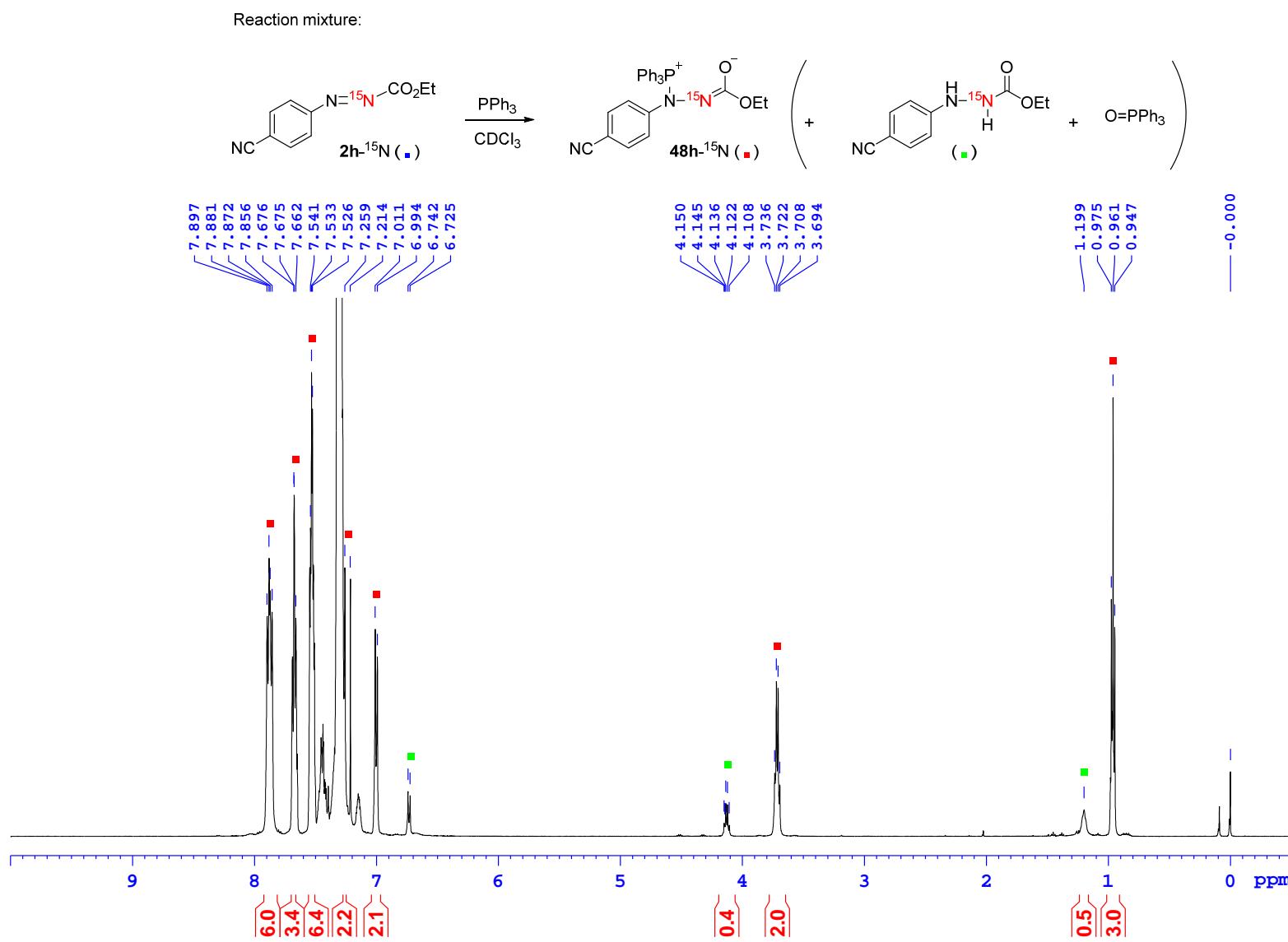
F2 - Acquisition Parameters
Date_ 20150730
Time 21.47
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 16
DS 4
SWH 81521.742 Hz
FIDRES 1.243923 Hz
AQ 0.4020041 sec
RG 1820
DW 6.133 usec
DE 6.50 usec
TE 296.1 K
D1 2.0000000 sec
D11 0.03000000 sec

===== CHANNEL f1 =====
NUC1 31P
P1 10.00 usec
PLW1 100.0000000 W
SFO1 202.4462121 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PLW2 26.00000000 W
PLW12 0.32179001 W
PLW13 0.20959001 W
SFO2 500.13200005 MHz

F2 - Processing parameters
SI 65536
SF 202.4563350 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



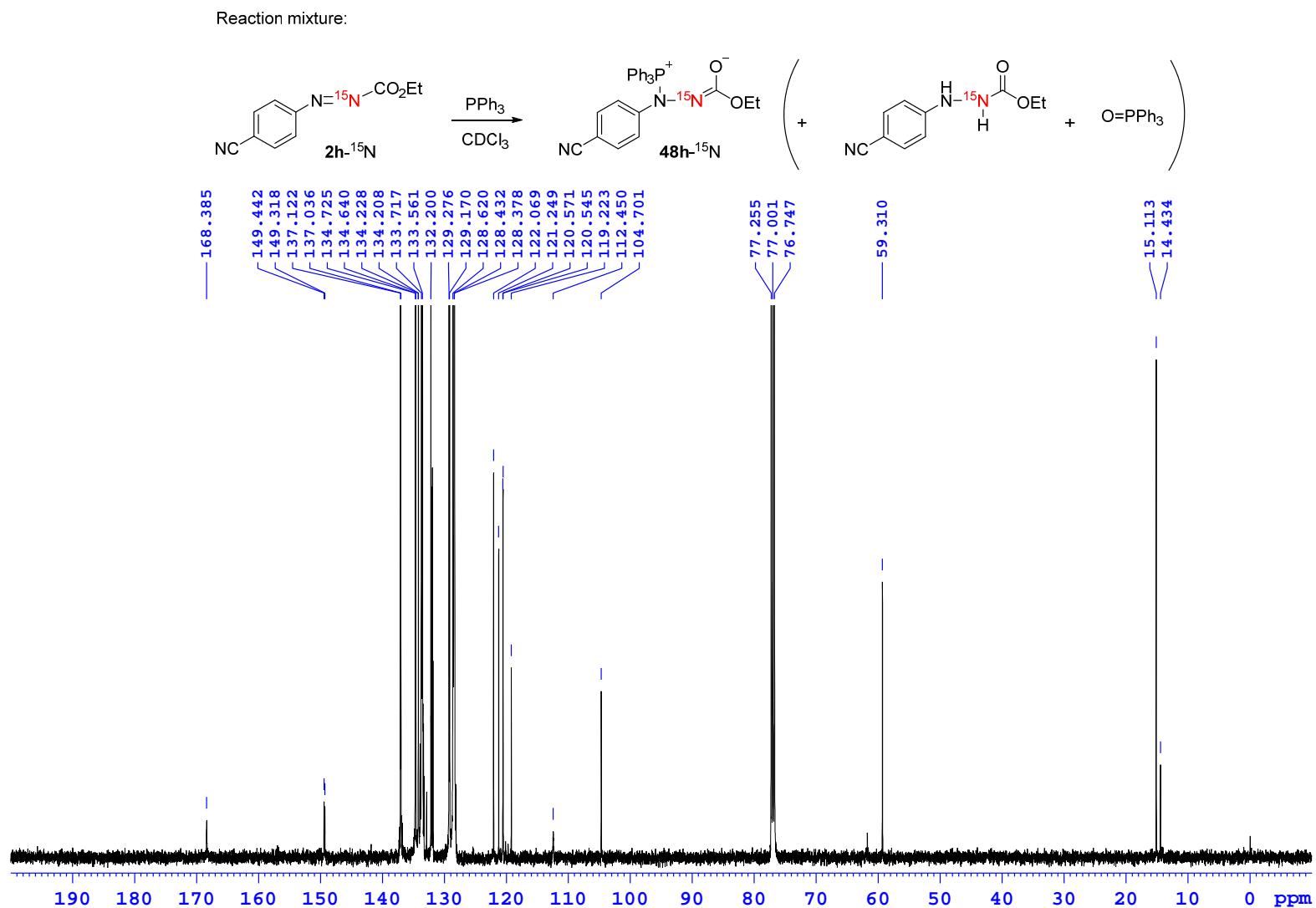


Current Data Parameters
 NAME MGS-651
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20151021
 Time 17.01
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl₃
 NS 16
 DS 2
 SWH 10330.578 Hz
 FIDRES 0.157632 Hz
 AQ 3.1719923 sec
 RG 32
 DW 48.400 usec
 DE 6.50 usec
 TE 296.0 K
 D1 1.0000000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 8.90 usec
 PLW1 26.0000000 W
 SFO1 500.1330885 MHz

F2 - Processing parameters
 SI 65536
 SF 500.1300365 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



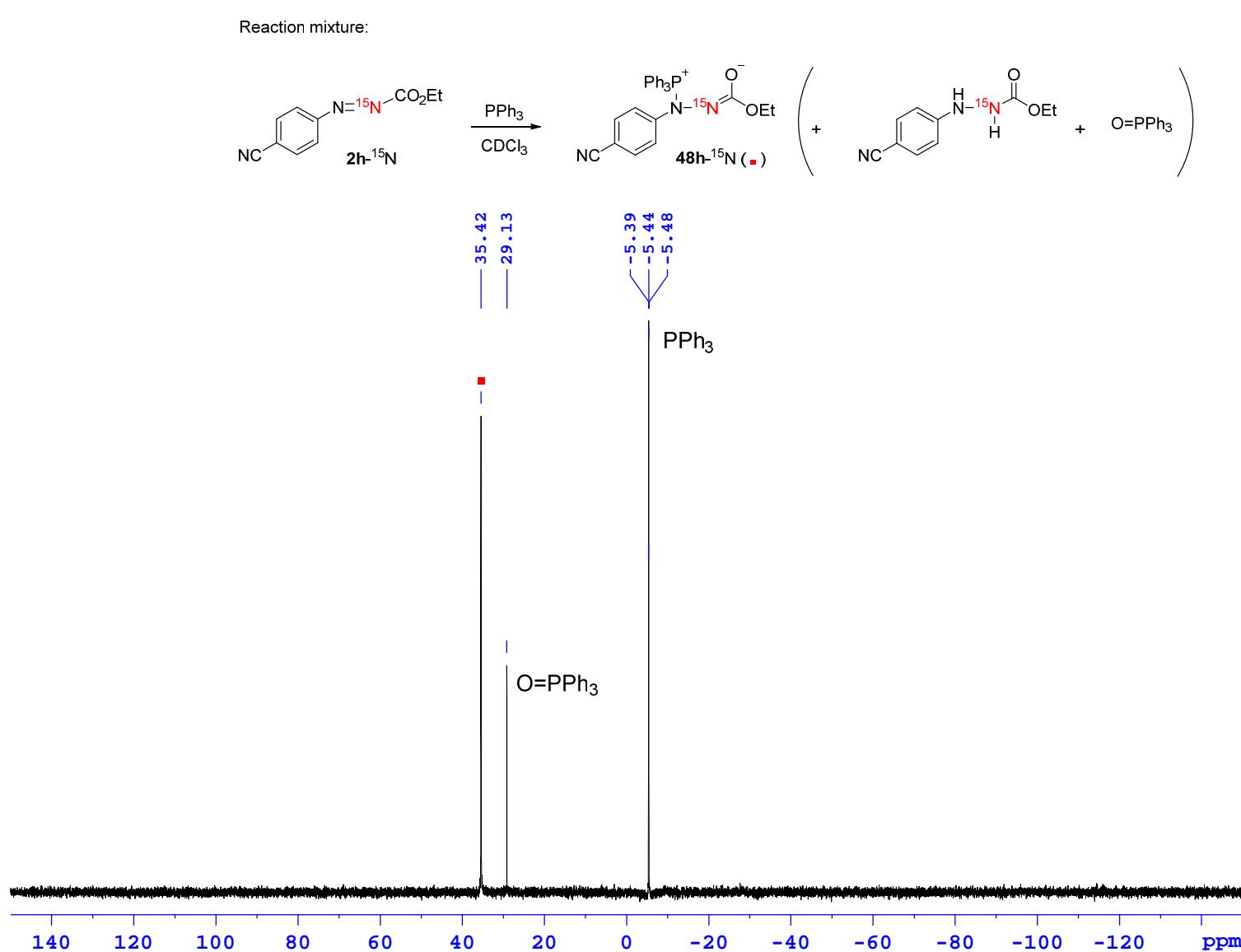
Current Data Parameters
 NAME MGS-651
 EXPNO 7
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20151021
 Time 20.38
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 4096
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.454131 Hz
 AQ 1.1010548 sec
 RG 2050
 DW 16.800 usec
 DE 6.50 usec
 TE 296.0 K
 D1 1.0000000 sec
 D11 0.0300000 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 9.00 usec
 PLW1 122.0000000 W
 SFO1 125.7703637 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PLW2 26.0000000 W
 PLW12 0.32179001 W
 PLW13 0.20595001 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7578087 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



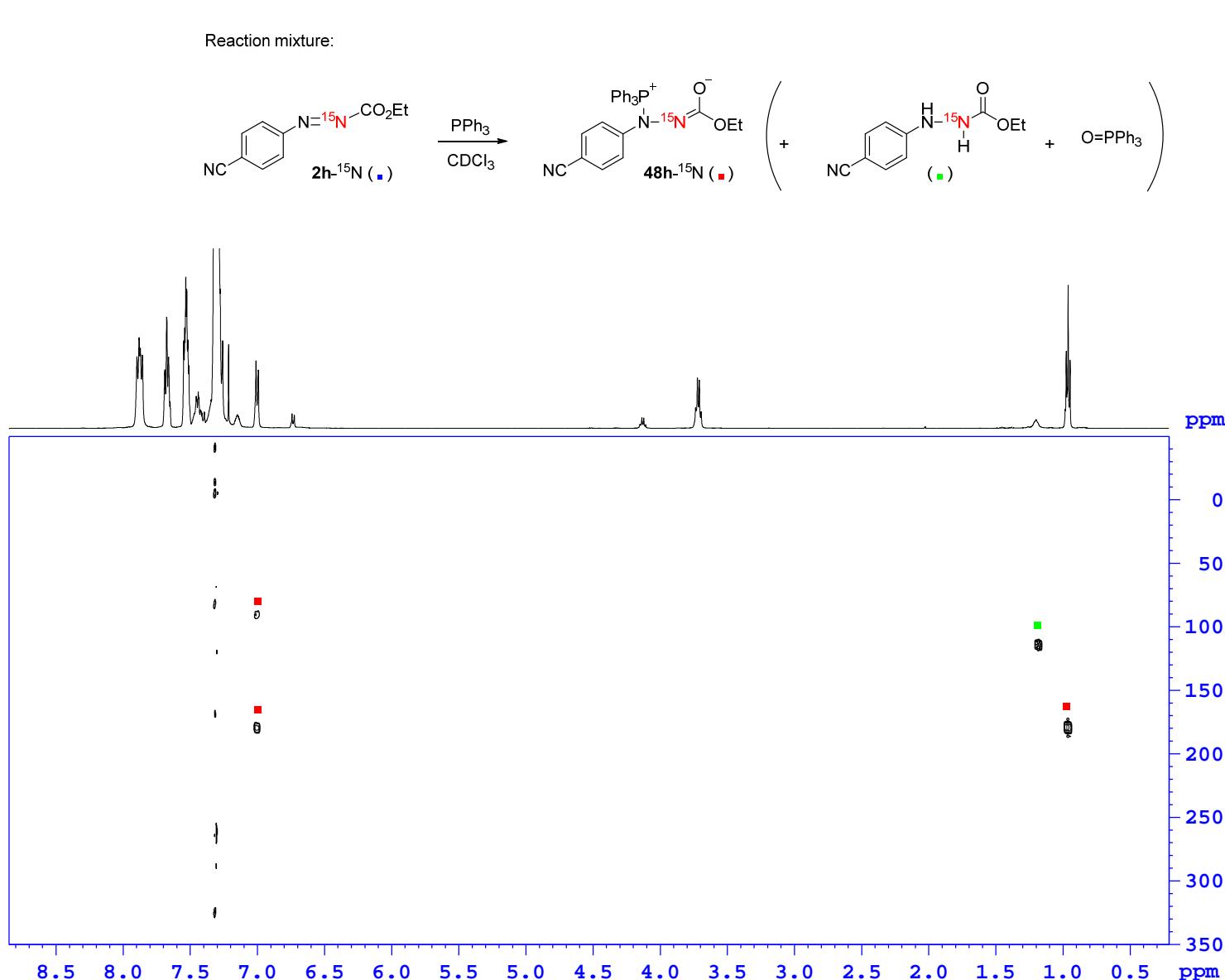
Current Data Parameters
 NAME MGS-651
 EXPNO 3
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20151021
 Time 17.06
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDC13
 NS 16
 DS 4
 SWH 81521.742 Hz
 FIDRES 1.243923 Hz
 AQ 0.4020041 sec
 RG 2050
 DW 6.133 usec
 DE 6.50 usec
 TE 296.1 K
 D1 2.0000000 sec
 D11 0.03000000 sec

===== CHANNEL f1 =====
 NUC1 31P
 P1 10.00 usec
 PLW1 100.0000000 W
 SFO1 202.4462121 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PLW2 26.00000000 W
 PLW12 0.32179001 W
 PLW13 0.20595001 W
 SFO2 500.13200005 MHz

F2 - Processing parameters
 SI 32768
 SF 202.4563350 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



Current Data Parameters
NAME MGS-651
EXPNO 4
PROCNO 1

F2 - Acquisition Parameters
Date_ 20151021
Time_ 17.09
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG hmbcgpndqf
TD 2048
SOLVENT CDCl3
NS 16
DS 16
SWH 4385.965 Hz
FIDRES 2.141584 Hz
AQ 0.2335220 sec
RG 2050
DW 114.000 usec
DE 6.50 usec
TE 296.0 K
CNST13 5.0000000
D0 0.00000300 sec
D1 1.92381406 sec
D6 0.1000000 sec
D16 0.00020000 sec
IN0 0.00002465 sec

===== CHANNEL f1 =====
NUC1 1H
P1 8.90 usec
P2 17.80 usec
PLW1 26.0000000 W
SFO1 500.1322693 MHz

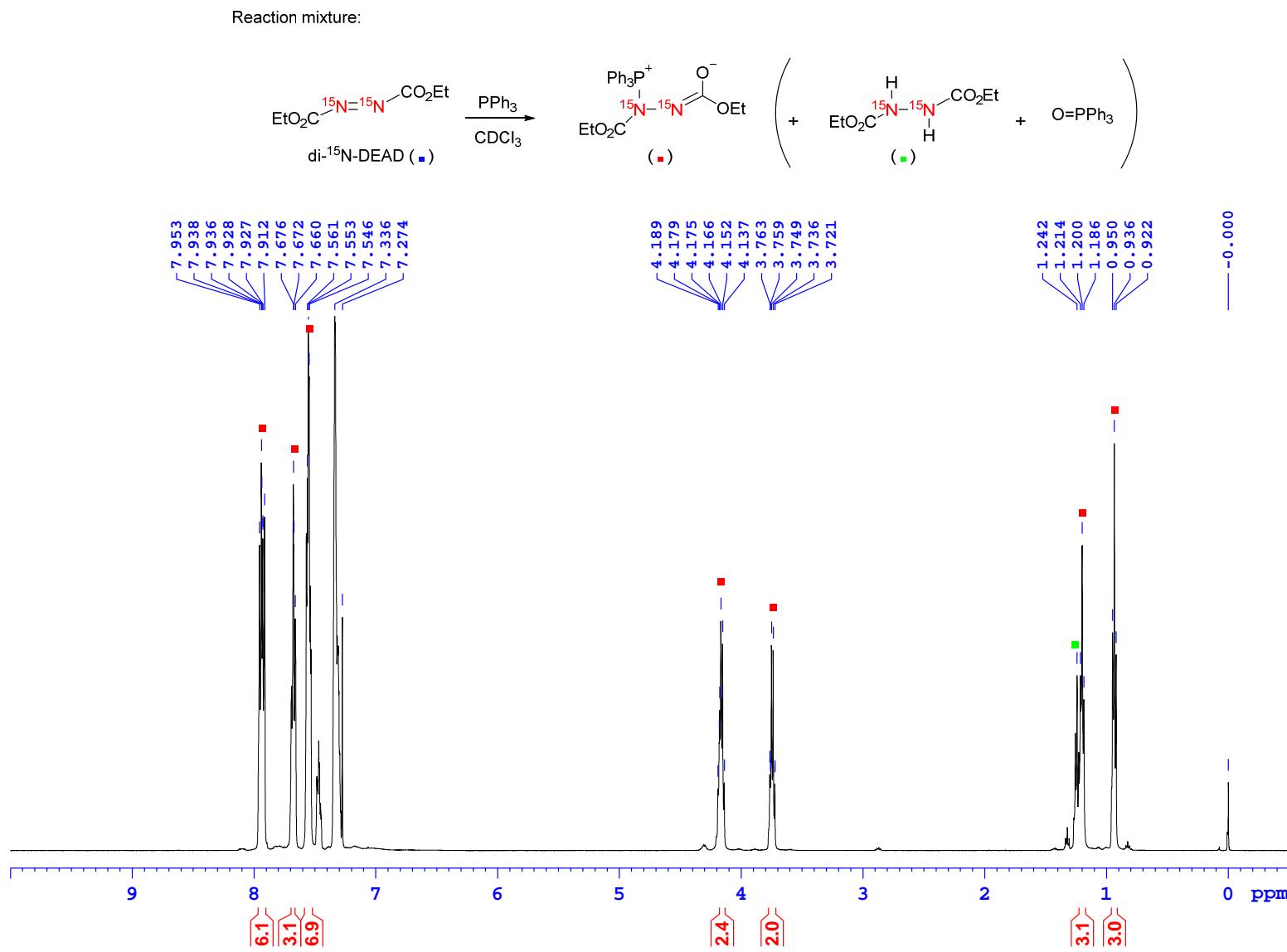
===== CHANNEL f2 =====
NUC2 15N
P3 14.40 usec
PLW2 206.0000000 W
SFO2 50.6853342 MHz

===== GRADIENT CHANNEL =====
GPNAME1 SMSQ10.100
GPNAME2 SMSQ10.100
GPNAME3 SMSQ10.100
GPZ1 70.00 %
GPZ2 30.00 %
GPZ3 50.10 %
P16 1000.00 usec

F1 - Acquisition parameters
TD 512
SFO1 50.68533 MHz
FIDRES 355.686554 Hz
SW 400.000 ppm
FnMODE QF

F2 - Processing parameters
SI 2048
SF 500.1300365 MHz
WDW SINE
SSB 0
LB 0 Hz
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
MC2 QF
SF 50.6777330 MHz
WDW States
SSB 0
LB 0 Hz
GB 0

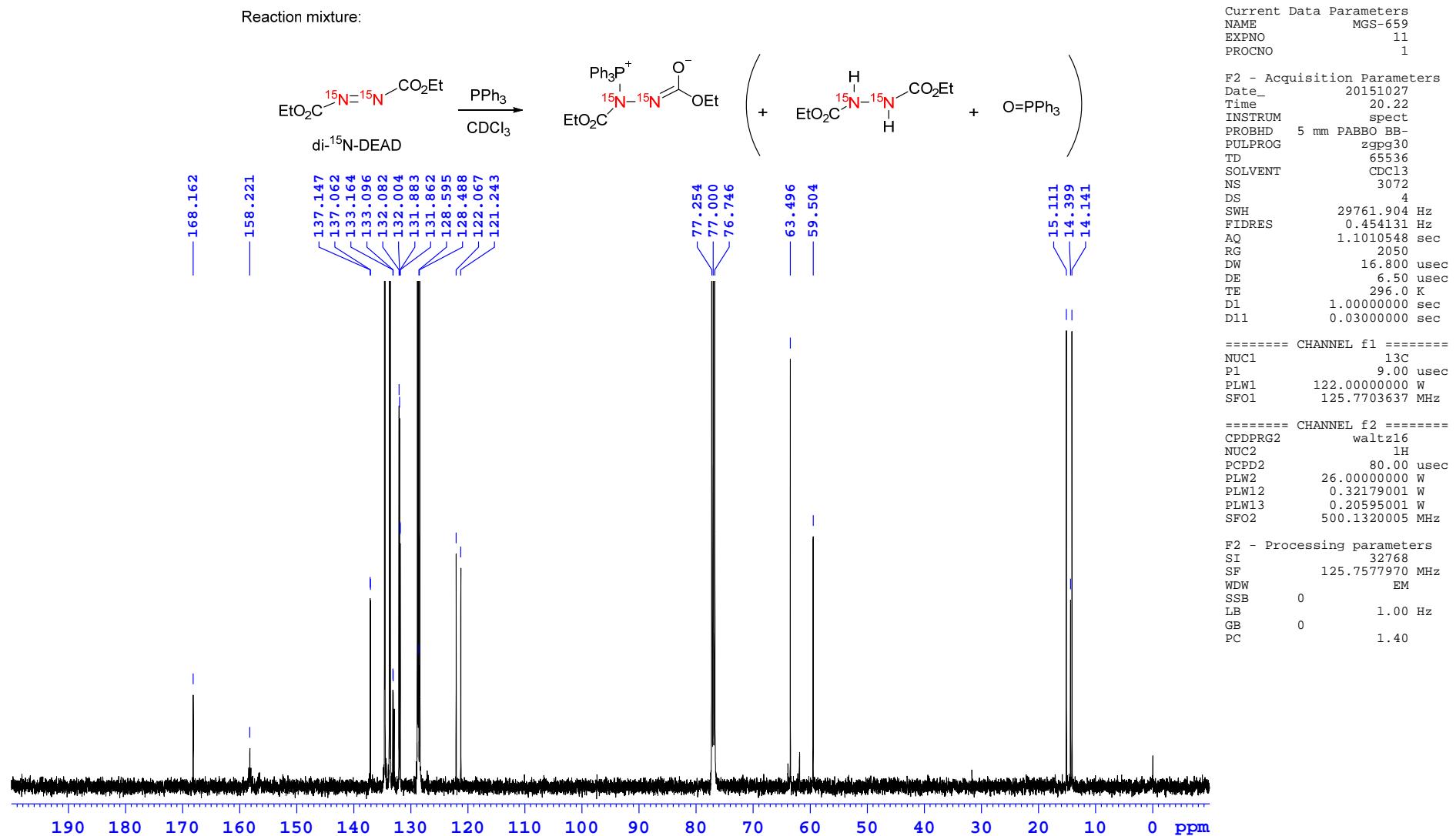


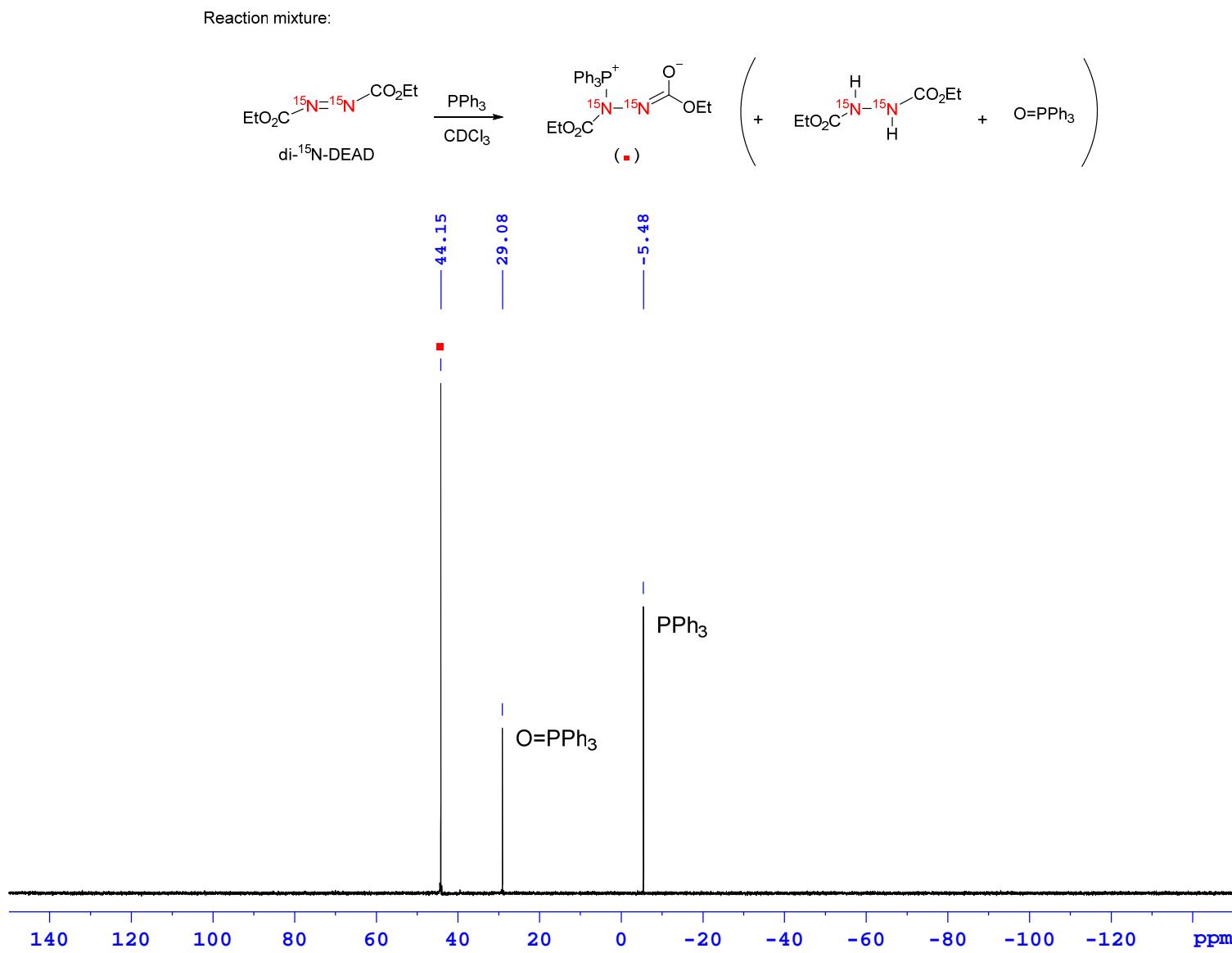
Current Data Parameters
 NAME MGS-659
 EXPNO 1
 PROCNO 1

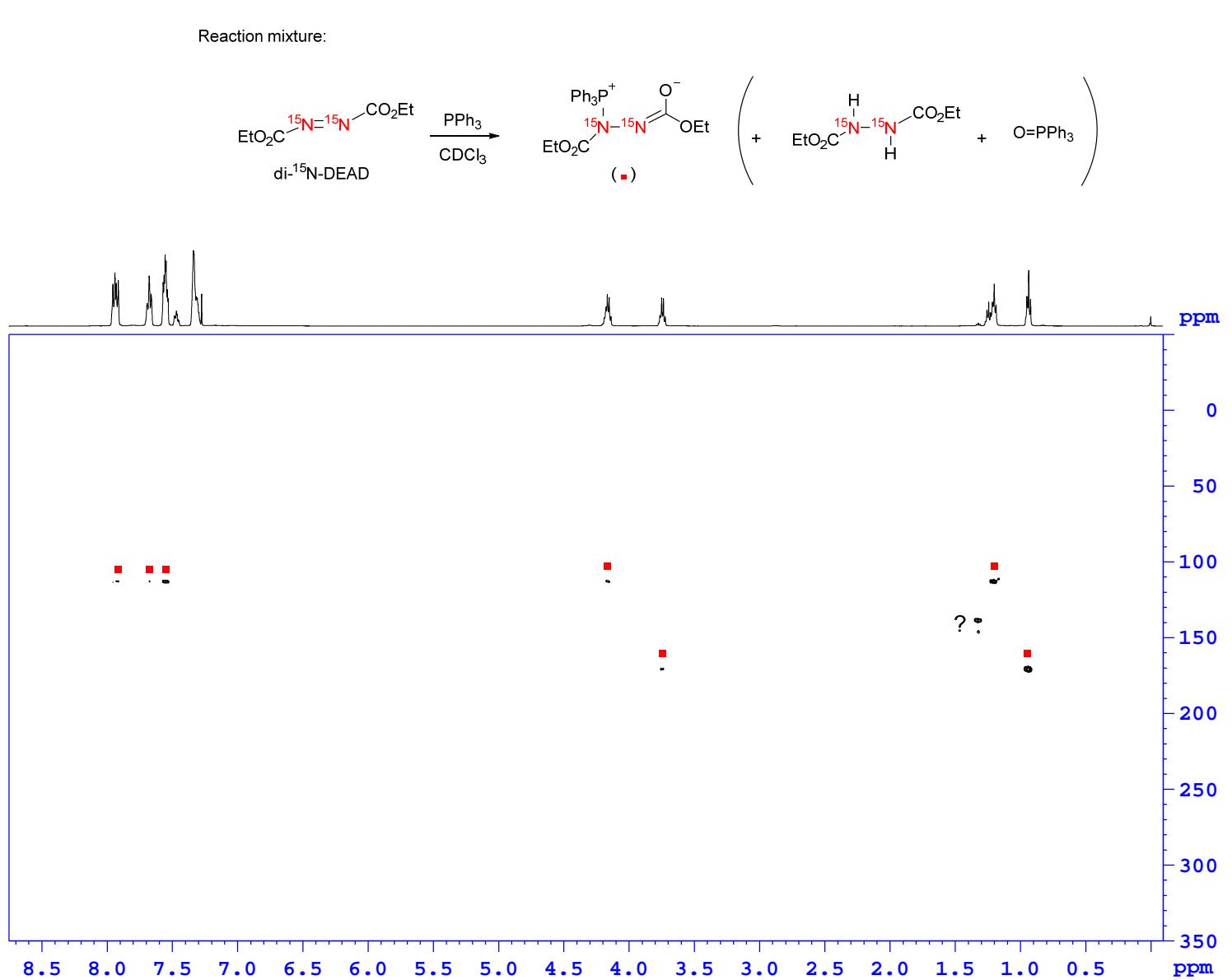
F2 - Acquisition Parameters
 Date_ 20151027
 Time 17.21
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 10330.578 Hz
 FIDRES 0.157632 Hz
 AQ 3.1719923 sec
 RG 80.6
 DW 48.400 usec
 DE 6.50 usec
 TE 296.0 K
 D1 1.0000000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 8.90 usec
 PLW1 26.0000000 W
 SF01 500.1330885 MHz

F2 - Processing parameters
 SI 65536
 SF 500.1300069 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00







Current Data Parameters
NAME MGS-659
EXPNO 15
PROCNO 1

F2 - Acquisition Parameters
Date 20151028
Time 0.43
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG hmbcpnddf
TD 2048
SOLVENT CDCl3
NS 16
DS 16
SWH 4424.779 Hz
FIDRES 2.160537 Hz
AQ 0.2314740 sec
RG 2050
DW 113.000 usec
DE 6.50 usec
TE 296.0 K
CNST13 5.0000000
D0 0.00000300 sec
D1 1.92586195 sec
D6 0.1000000 sec
D16 0.00020000 sec
INO 0.00002465 sec

===== CHANNEL f1 =====
NUC1 1H
P1 8.90 usec
P2 17.80 usec
PLW1 26.0000000 W
SFO1 500.1321725 MHz

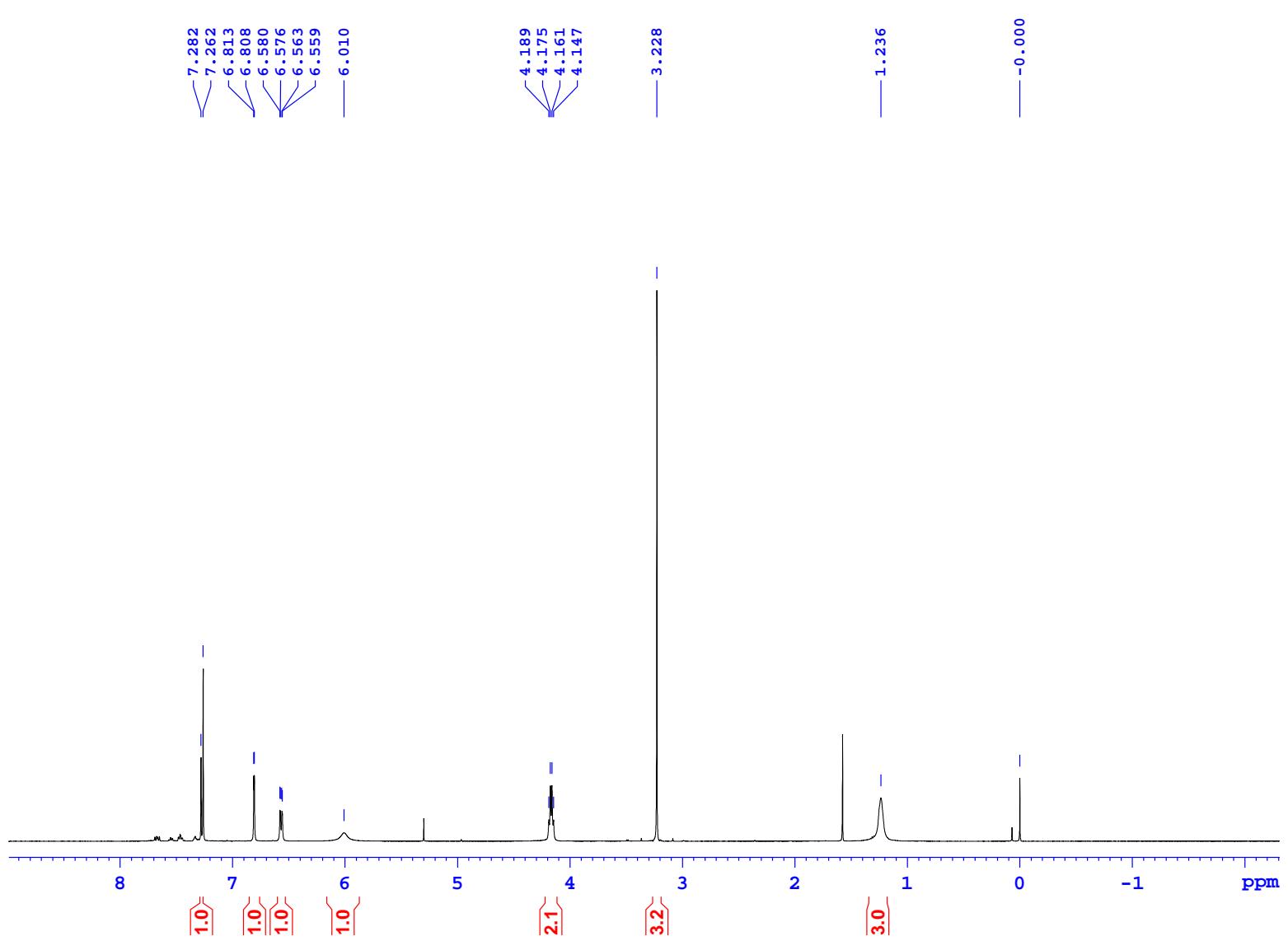
===== CHANNEL f2 =====
NUC2 15N
P3 14.40 usec
PLW2 206.0000000 W
SFO2 50.6853342 MHz

===== GRADIENT CHANNEL =====
GPNAME1 SMSQ10.100
GPNAME2 SMSQ10.100
GPNAME3 SMSQ10.100
GPZ1 70.00 %
GPZ2 30.00 %
GPZ3 50.10 %
P16 1000.00 usec

F1 - Acquisition parameters
TD 256
SFO1 50.68533 MHz
FIDRES 79.195831 Hz
SW 400.000 ppm
FnMODE QF

F2 - Processing parameters
SI 2048
SF 500.1300073 MHz
WDW SINE
SSB 0
LB 0 Hz
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
MC2 QF
SF 50.6777330 MHz
WDW echo-antiecho
SSB 0
LB 0 Hz
GB 0

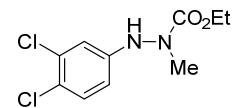


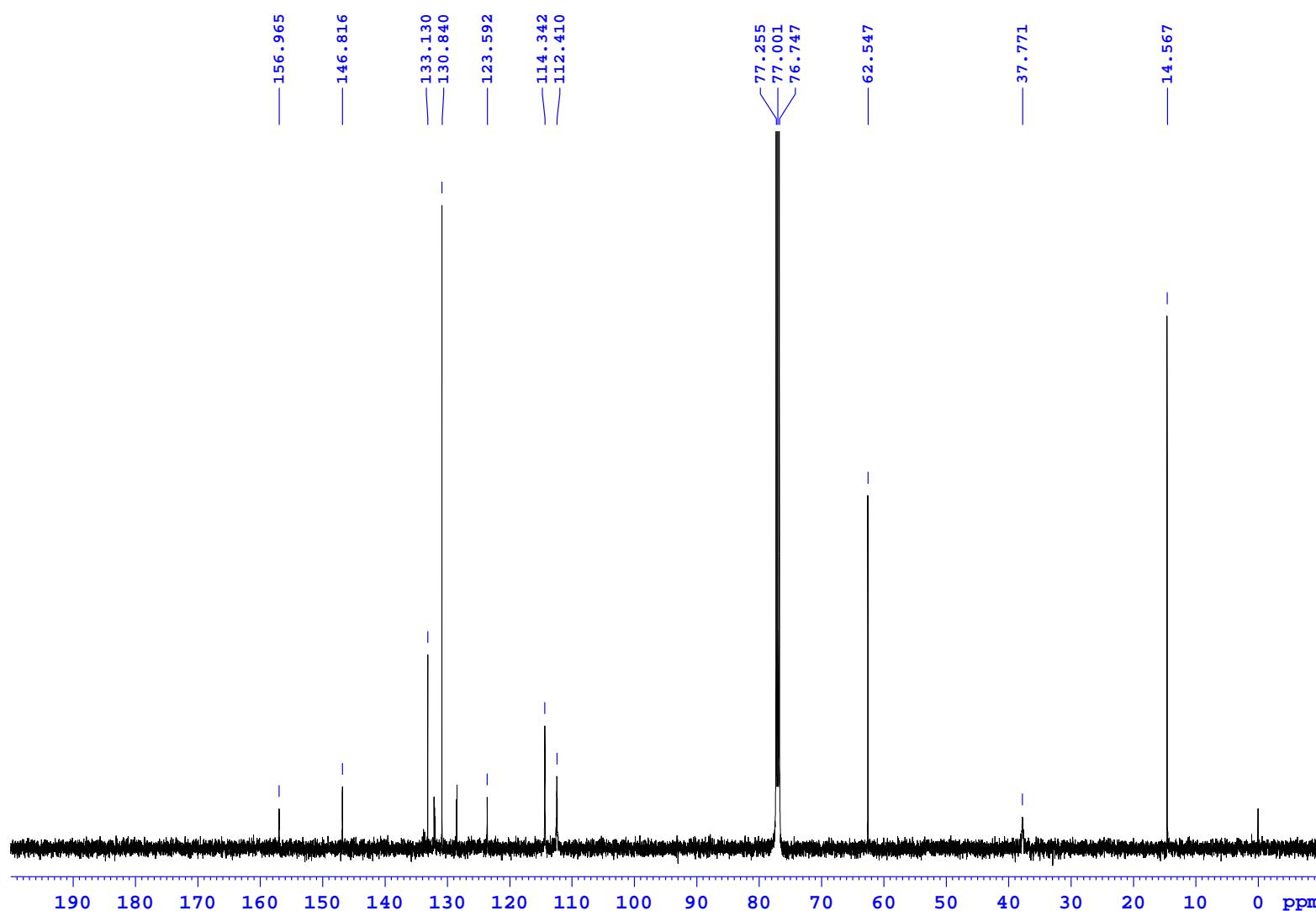
Current Data Parameters
NAME mgs-634
EXPNO 50
PROCNO 1

F2 - Acquisition Parameters
Date_ 20151028
Time 18.31
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 161
DW 48.400 usec
DE 6.50 usec
TE 296.0 K
D1 1.0000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 8.90 usec
PLW1 26.0000000 W
SF01 500.1330885 MHz

F2 - Processing parameters
SI 65536
SF 500.1300126 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00





Current Data Parameters
NAME mgs-634
EXPNO 52
PROCNO 1

F2 - Acquisition Parameters
Date_ 20151028
Time 21.20
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 4096
DS 4
SWH 29761.904 Hz
FIDRES 0.454131 Hz
AQ 1.1010548 sec
RG 2050
DW 16.800 usec
DE 6.50 usec
TE 296.0 K
D1 1.0000000 sec
D11 0.0300000 sec

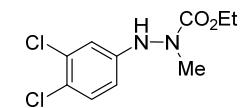
===== CHANNEL f1 ======

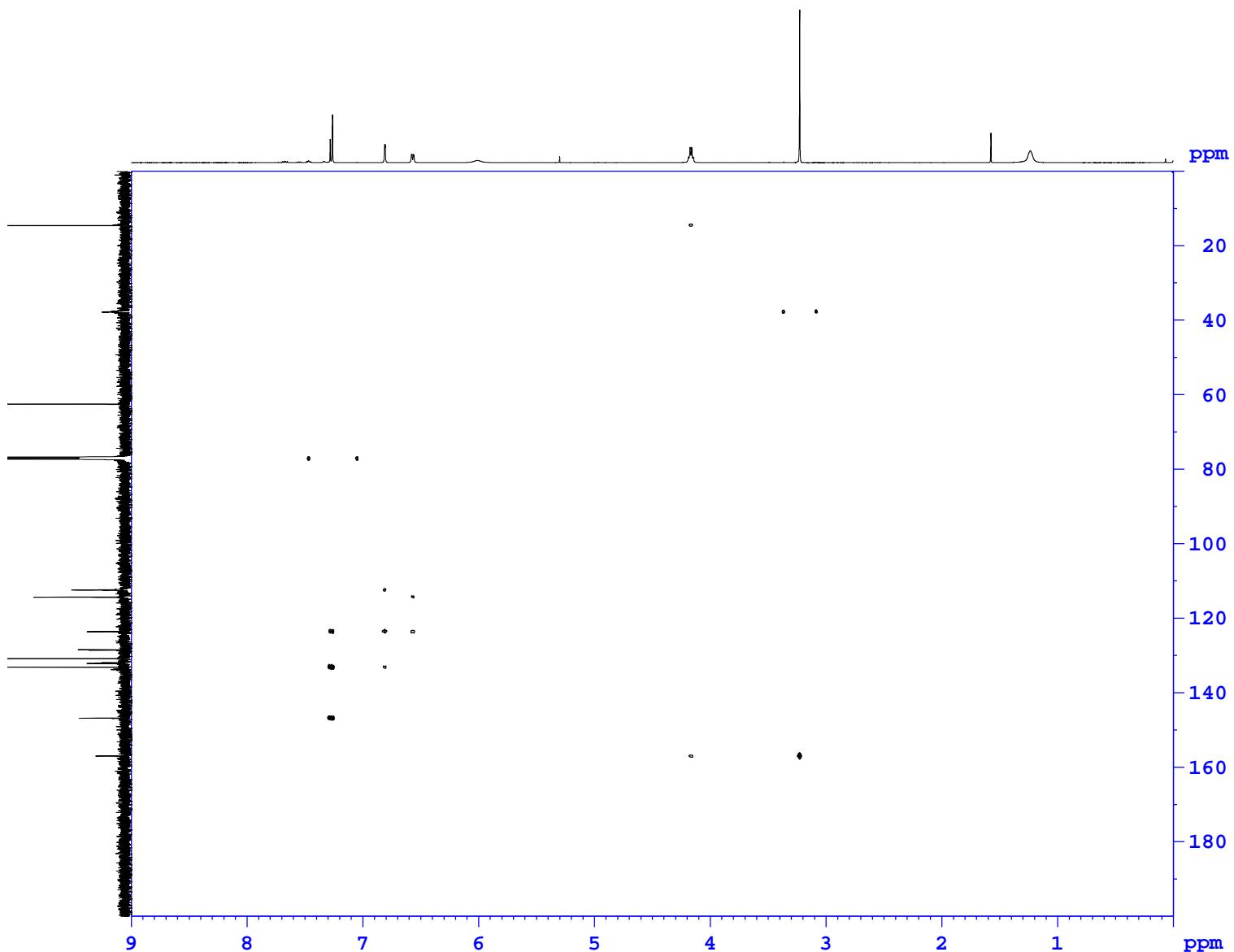
NUC1 13C
P1 9.00 usec
PLW1 122.0000000 W
SFO1 125.7703637 MHz

===== CHANNEL f2 ======

CPDPGRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PLW2 26.00000000 W
PLW12 0.32179001 W
PLW13 0.20595001 W
SFO2 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 125.7577928 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40





Current Data Parameters
 NAME mgs-634
 EXPNO 54
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20151028
 Time 21.51
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG hmbcgp1pdgf
 TD 2048
 SOLVENT CDCl3
 NS 16
 DS 16
 SWH 4424.779 Hz
 FIDRES 2.160537 Hz
 AQ 0.2314740 sec
 RG 2050
 DW 113.000 usec
 DE 6.50 usec
 TE 296.0 K
 CNST2 145.000000
 CNST13 10.000000
 D0 0.00000300 sec
 D1 1.42586195 sec
 D2 0.00344828 sec
 D6 0.05000000 sec
 D16 0.00020000 sec
 IN0 0.00001790 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 8.90 usec
 P2 17.80 usec
 PLW1 26.0000000 W
 SF01 500.1320274 MHz

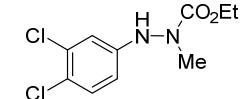
===== CHANNEL f2 =====
 NUC2 13C
 P3 9.00 usec
 PLW2 122.0000000 W
 SF02 125.7703437 MHz

===== GRADIENT CHANNEL =====
 GPNAME1 SMS10.100
 GPNAME2 SMS10.100
 GPNAME3 SMS10.100
 GPZ1 50.00 %
 GPZ2 30.00 %
 GPZ3 40.10 %
 P16 1000.00 usec

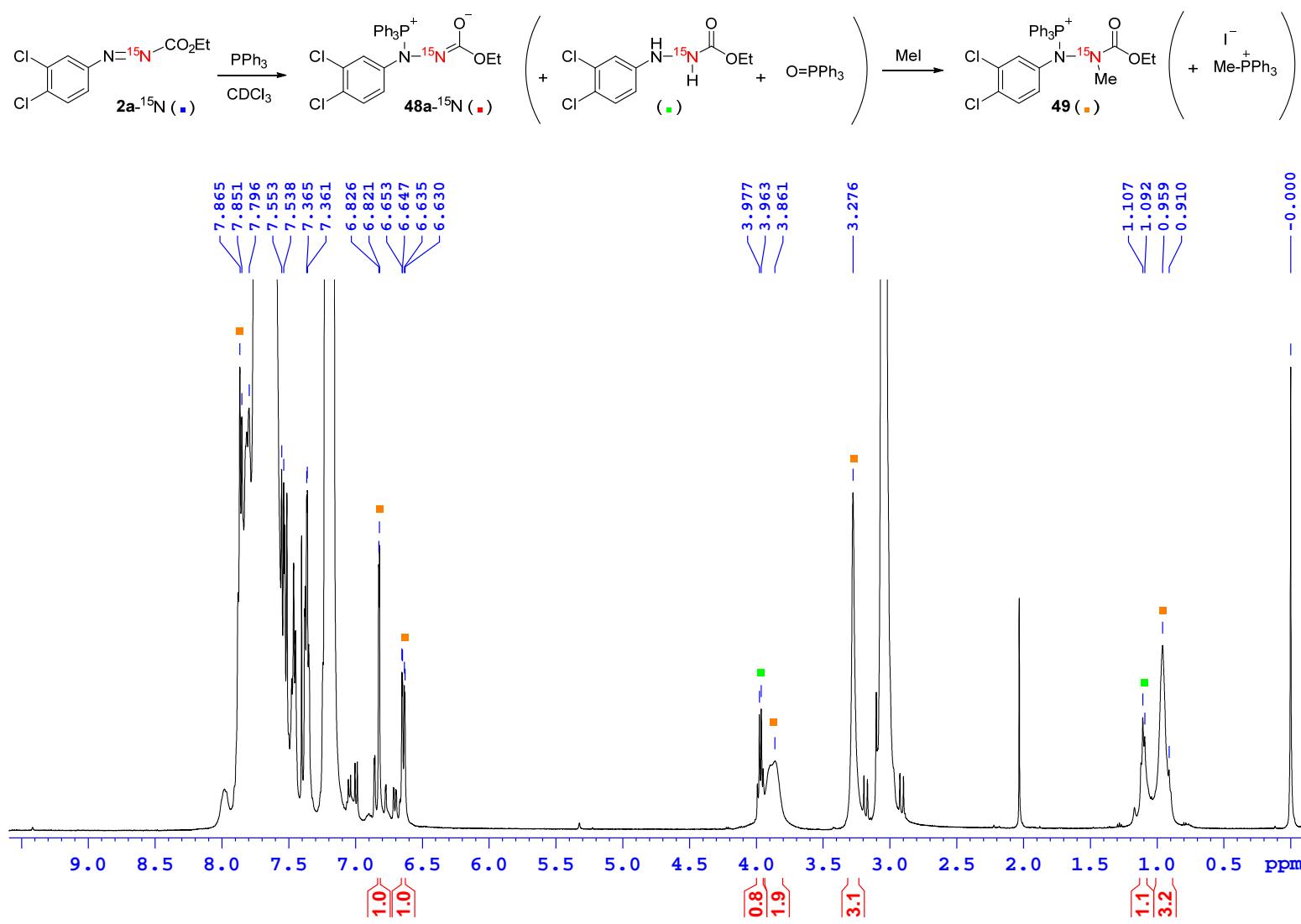
F1 - Acquisition parameters
 TD 256
 SF01 125.7703 MHz
 FIDRES 109.113174 Hz
 SW 222.095 ppm
 F1MODE QF

F2 - Processing parameters
 SI 2048
 SF 500.1300126 MHz
 WDW SINE
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.40

F1 - Processing parameters
 SI 1024
 MC2 QF
 SF 125.7577890 MHz
 WDW States
 SSB 0
 LB 0 Hz
 GB 0



Reaction mixture:



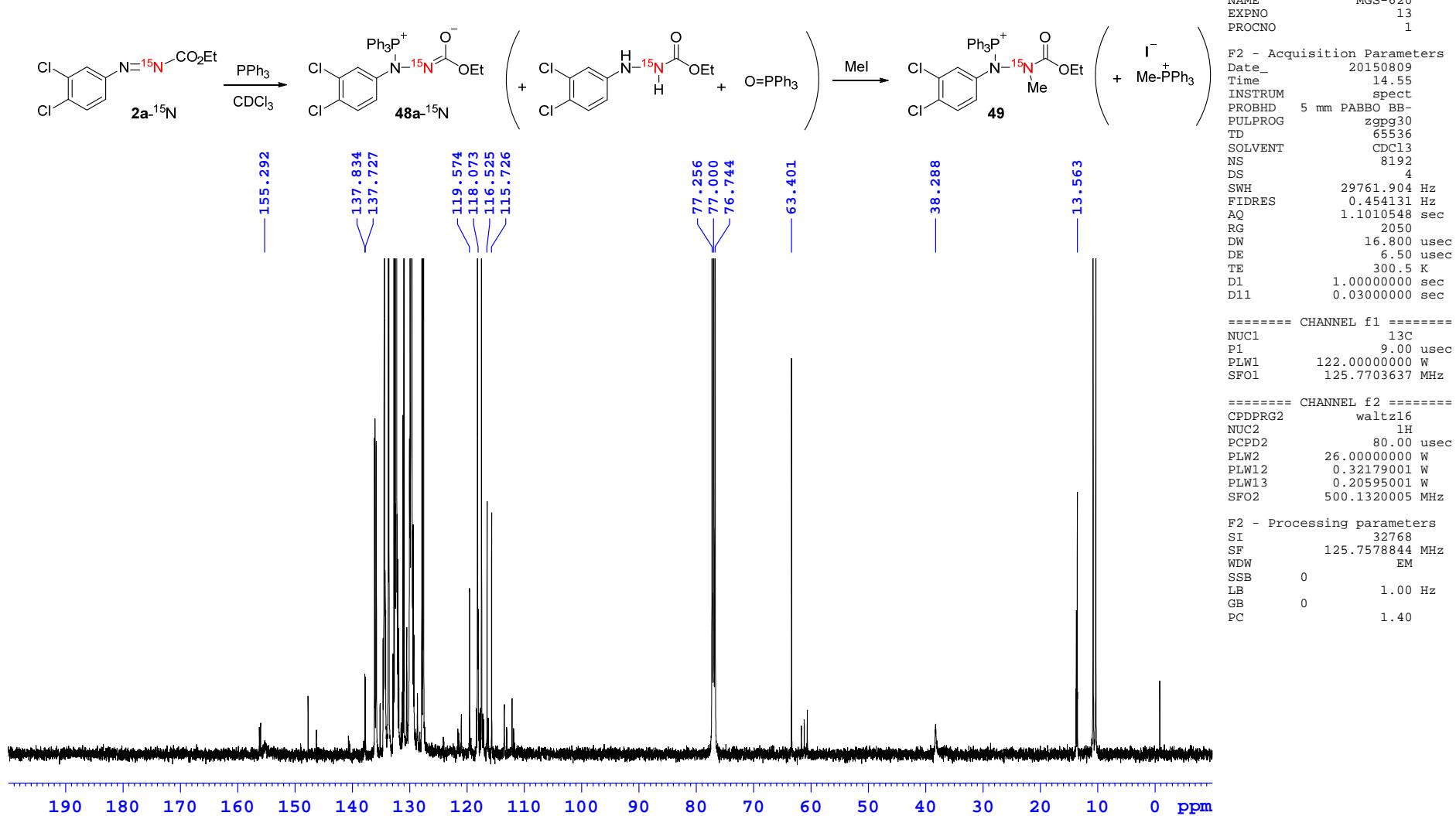
Current Data Parameters
NAME MGS-620
EXPNO 6
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150808
Time 16.09
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 32
DW 48.400 usec
DE 6.50 usec
TE 299.5 K
D1 1.0000000 sec

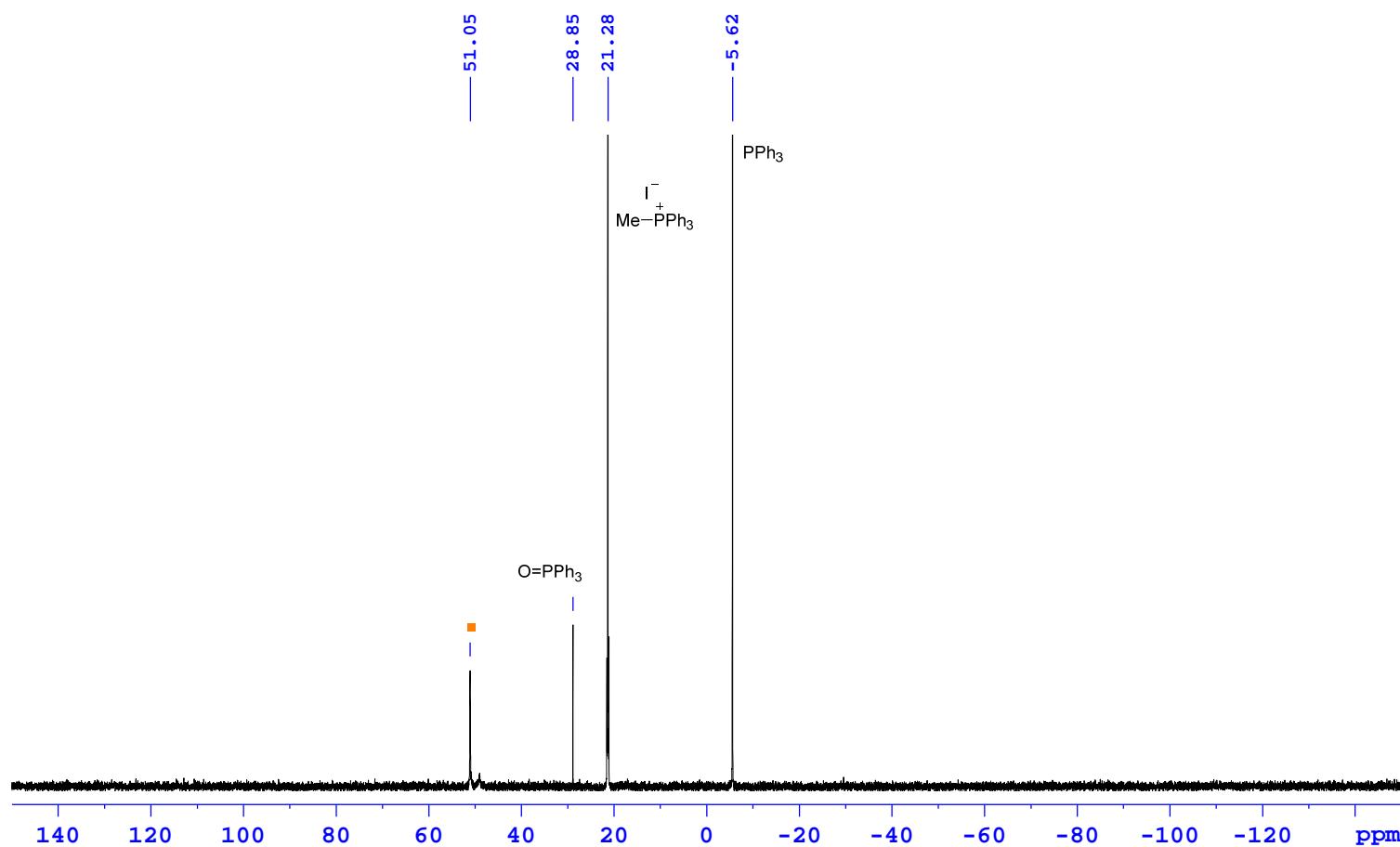
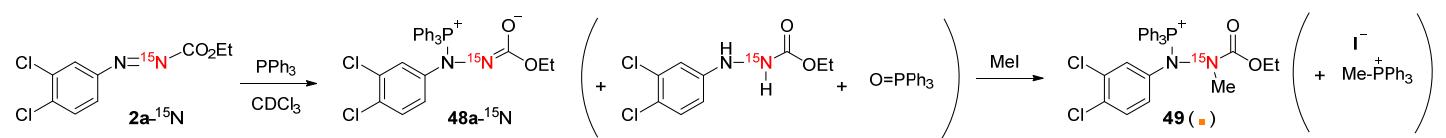
===== CHANNEL f1 =====
NUC1 1H
P1 8.90 usec
PLW1 26.0000000 W
SF01 500.1330885 MHz

F2 - Processing parameters
SI 65536
SF 500.1299483 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

Reaction mixture:



Reaction mixture:



Current Data Parameters
NAME MGS-620
EXPNO 2
PROCNO 1

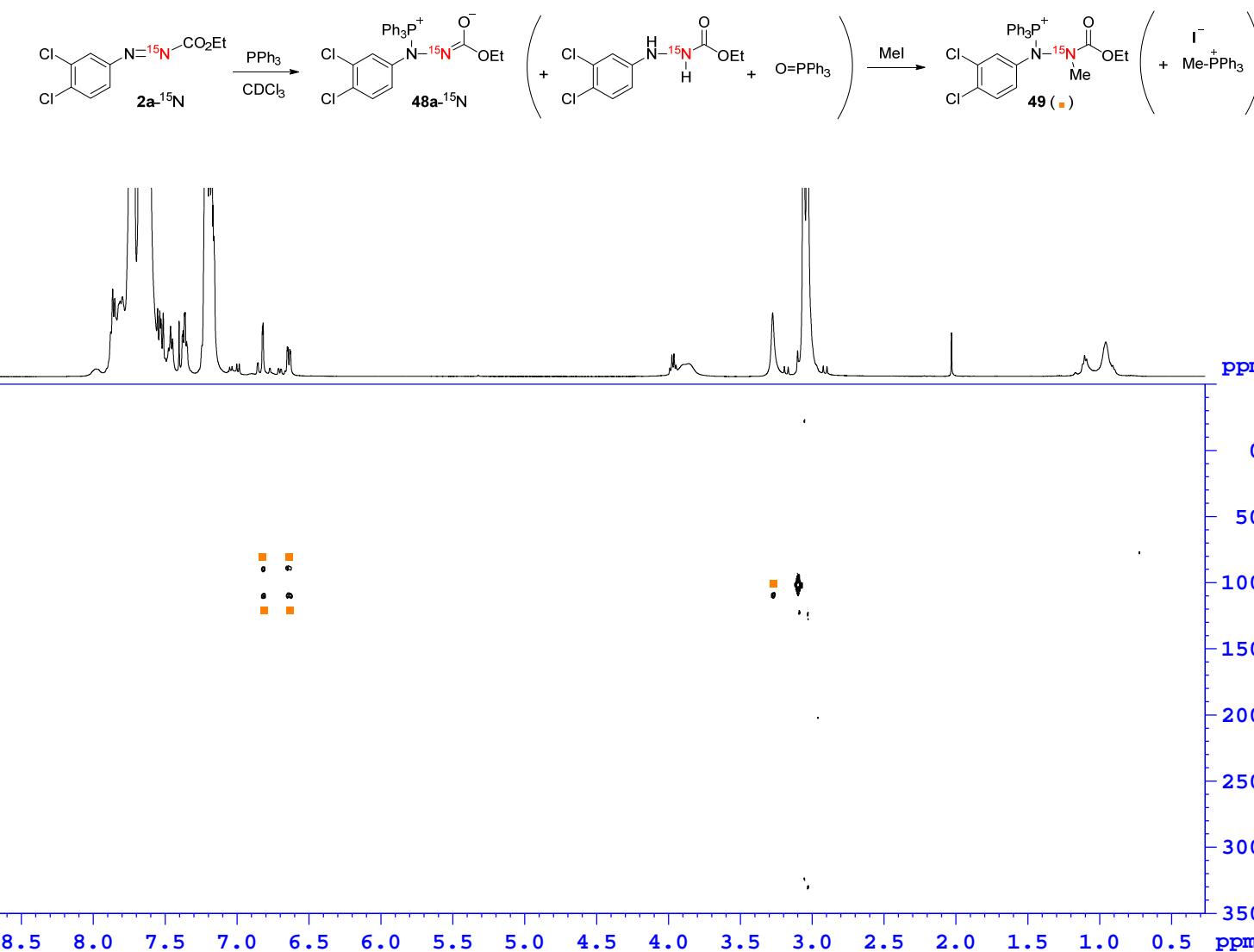
F2 - Acquisition Parameters
Date_ 20150808
Time 14.39
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl₃
NS 16
DS 4
SWH 81521.742 Hz
FIDRES 1.243923 Hz
AQ 0.4020041 sec
RG 2050
DW 6.133 usec
DE 6.50 usec
TE 299.5 K
D1 2.00000000 sec
D11 0.03000000 sec

===== CHANNEL f1 =====
NUC1 31P
P1 10.00 usec
PLW1 100.0000000 W
SFO1 202.4462121 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PLW2 26.00000000 W
PLW12 0.32179001 W
PLW13 0.20595001 W
SFO2 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 202.4563350 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Reaction mixture:



Current Data Parameters
NAME MGS-620
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date 20150808
Time 16.40
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG hmbcgpndf
TD 2048
SOLVENT CDCl₃
NS 8
DS 16
SWH 4201.681 Hz
FIDRES 2.051602 Hz
AQ 0.2437620 sec
RG 2050
DW 119.000 usec
DE 6.50 usec
TE 299.5 K
CNST13 5.0000000
D0 0.00000300 sec
D1 1.91357398 sec
D6 0.10000000 sec
D16 0.00020000 sec
IN0 0.00002465 sec

===== CHANNEL f1 =====
NUC1 1H
P1 8.90 usec
P2 17.80 usec
PLW1 26.0000000 W
SFO1 500.1321846 MHz

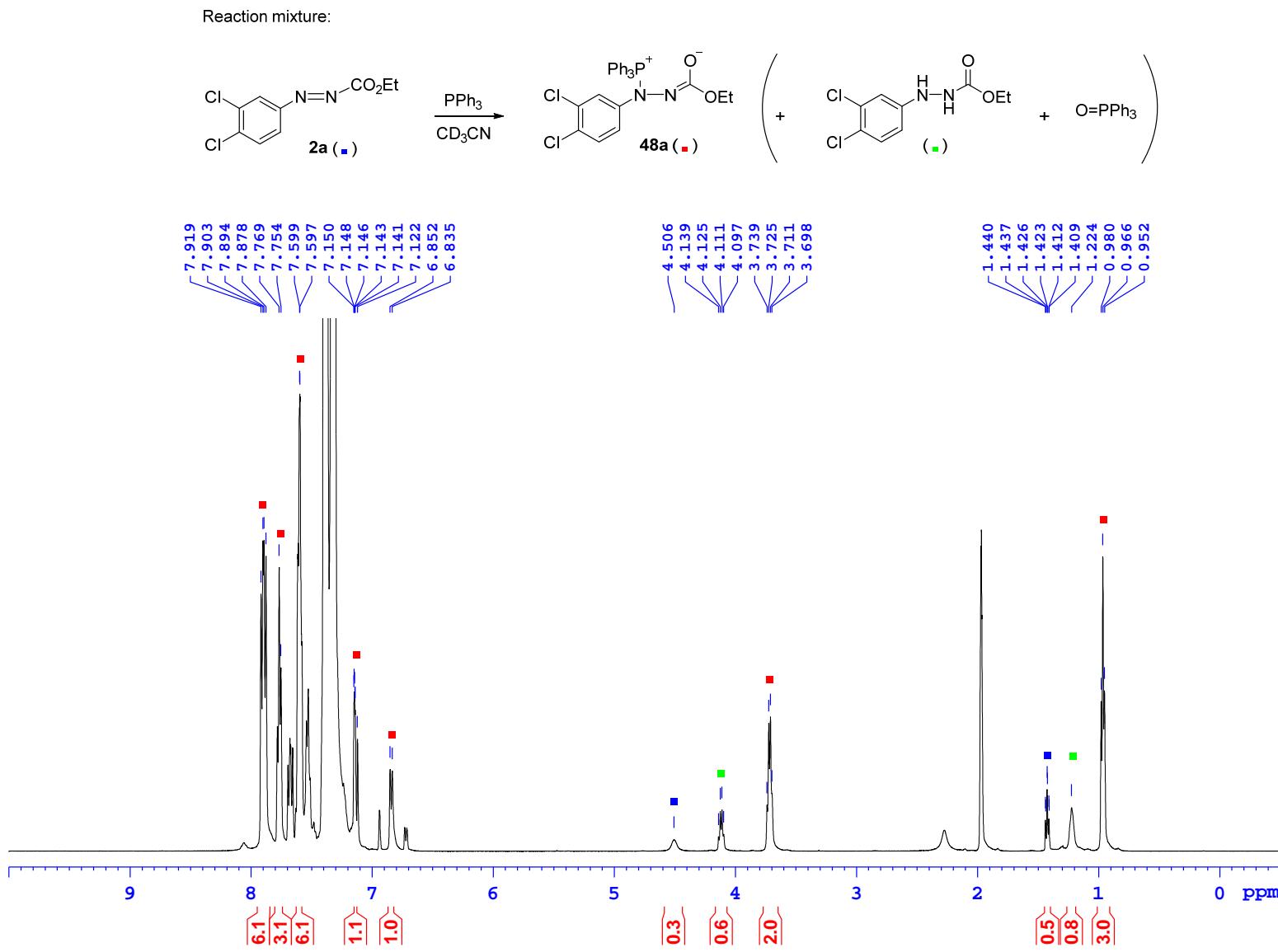
===== CHANNEL f2 =====
NUC2 15N
P3 14.40 usec
PLW2 206.0000000 W
SFO2 50.6853342 MHz

===== GRADIENT CHANNEL =====
GRNM1 SMSQ14.100
GRNM2 SMSQ10.100
GRNM3 SMSQ10.100
GPZ1 70.00 %
GPZ2 30.00 %
GPZ3 50.10 %
P16 1000.00 usec

F1 - Acquisition parameters
TD 128
SFO1 50.68533 MHz
FIDRES 158.391662 Hz
SW 400.000 ppm
FnMODE QF

F2 - Processing parameters
SI 2048
SF 500.1299502 MHz
WDW SINE
SSB 0
LB 0 Hz
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
MC2 QF
SF 50.6777330 MHz
WDW States
SSB 0
LB 0 Hz
GB 0

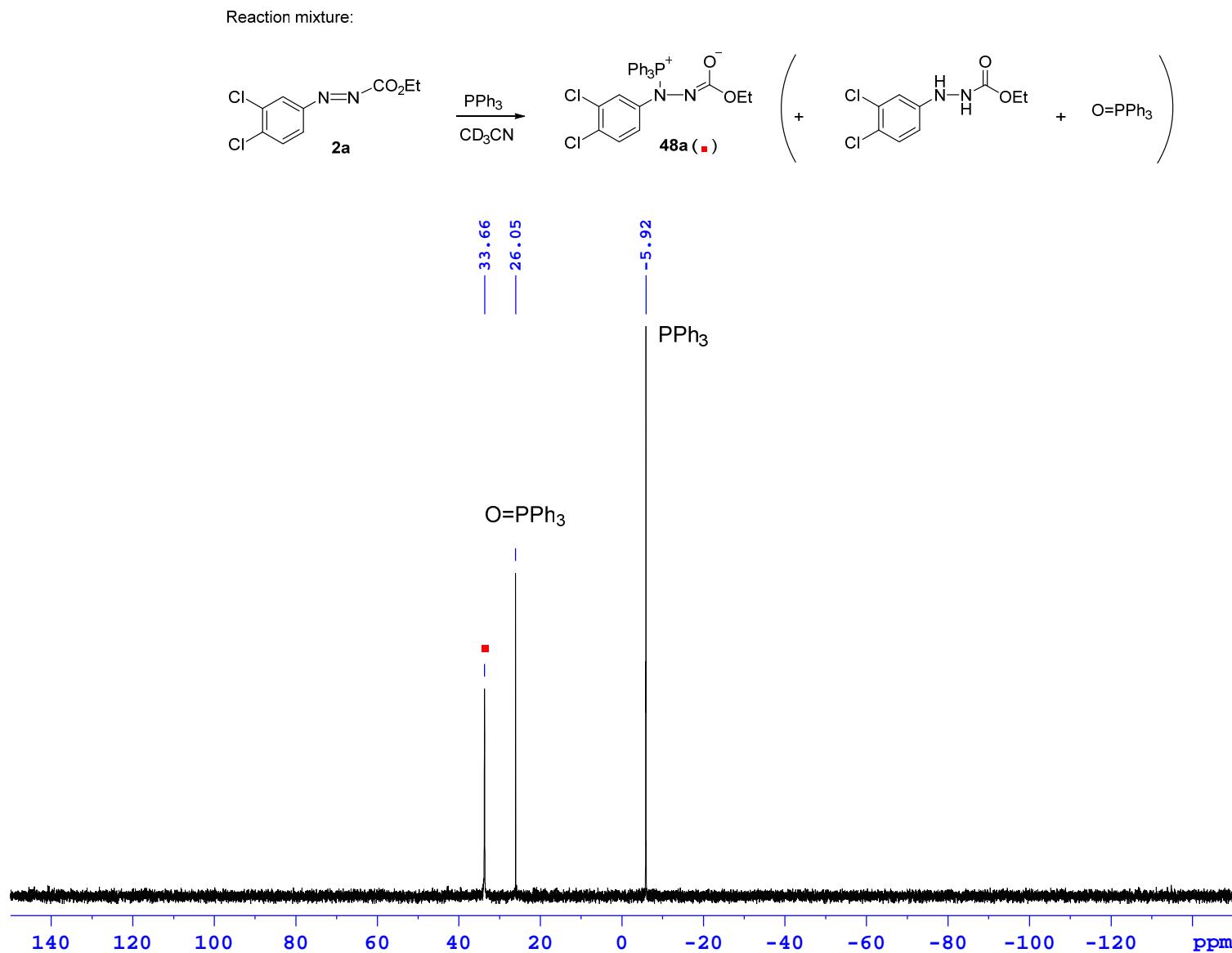


Current Data Parameters
NAME MGS-643
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20151006
Time 12.07
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CD3CN
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 32
DW 48.400 usec
DE 6.50 usec
TE 296.7 K
D1 1.0000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 8.90 usec
PLW1 26.00000000 W
SF01 500.1330885 MHz

F2 - Processing parameters
SI 65536
SF 500.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



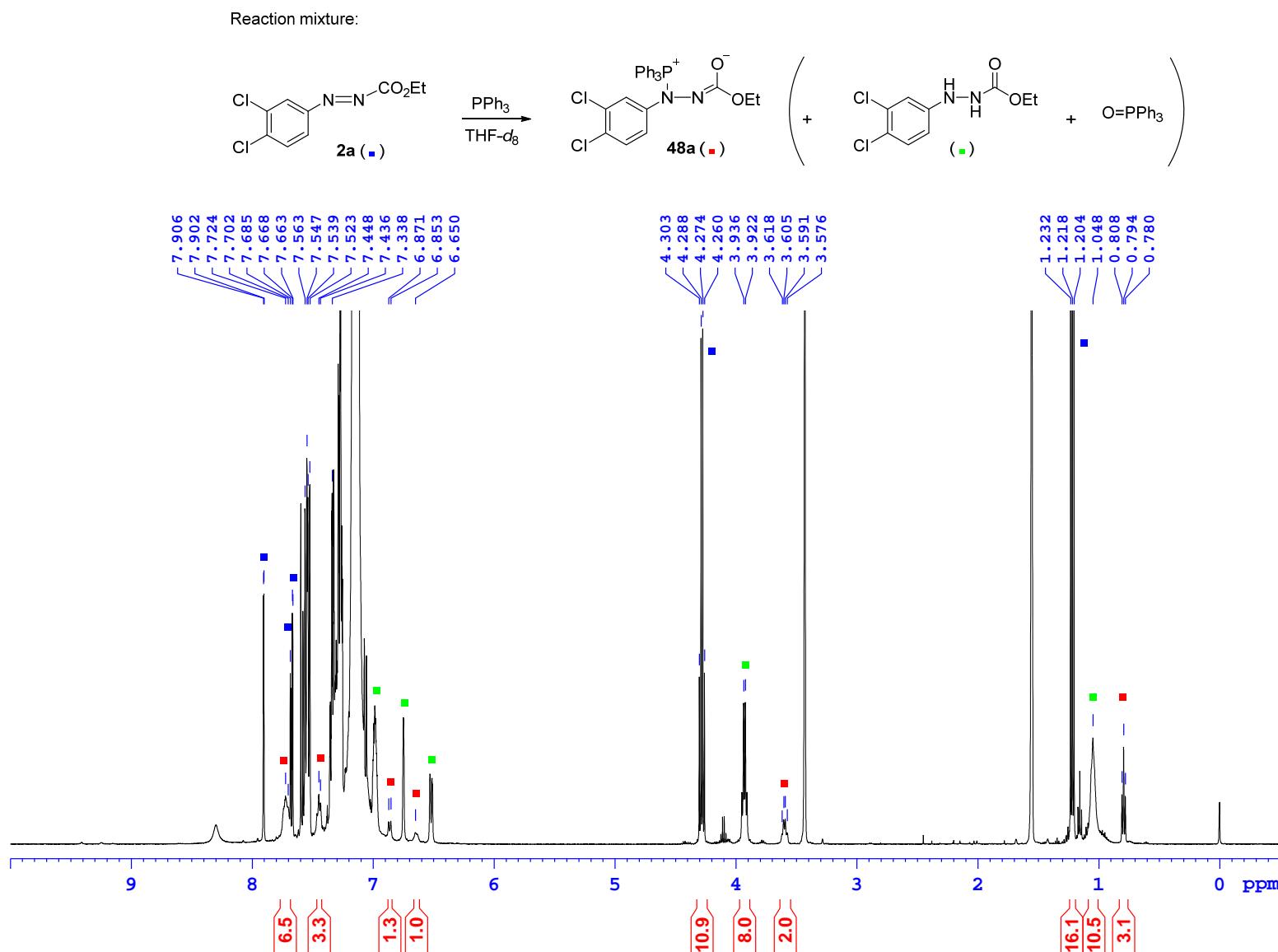
Current Data Parameters
NAME MGS-643
EXPNO 12
PROCNO 1

F2 - Acquisition Parameters
Date_ 20151006
Time 12.12
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CD3CN
NS 16
DS 4
SWH 81521.742 Hz
FIDRES 1.243923 Hz
AQ 0.4020041 sec
RG 2050
DW 6.133 usec
DE 6.50 usec
TE 296.9 K
D1 2.00000000 sec
D11 0.03000000 sec

===== CHANNEL f1 =====
NUC1 31P
P1 10.00 usec
PLW1 100.0000000 W
SFO1 202.4462121 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PLW2 26.00000000 W
PLW12 0.32179001 W
PLW13 0.20595001 W
SFO2 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 202.4563350 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

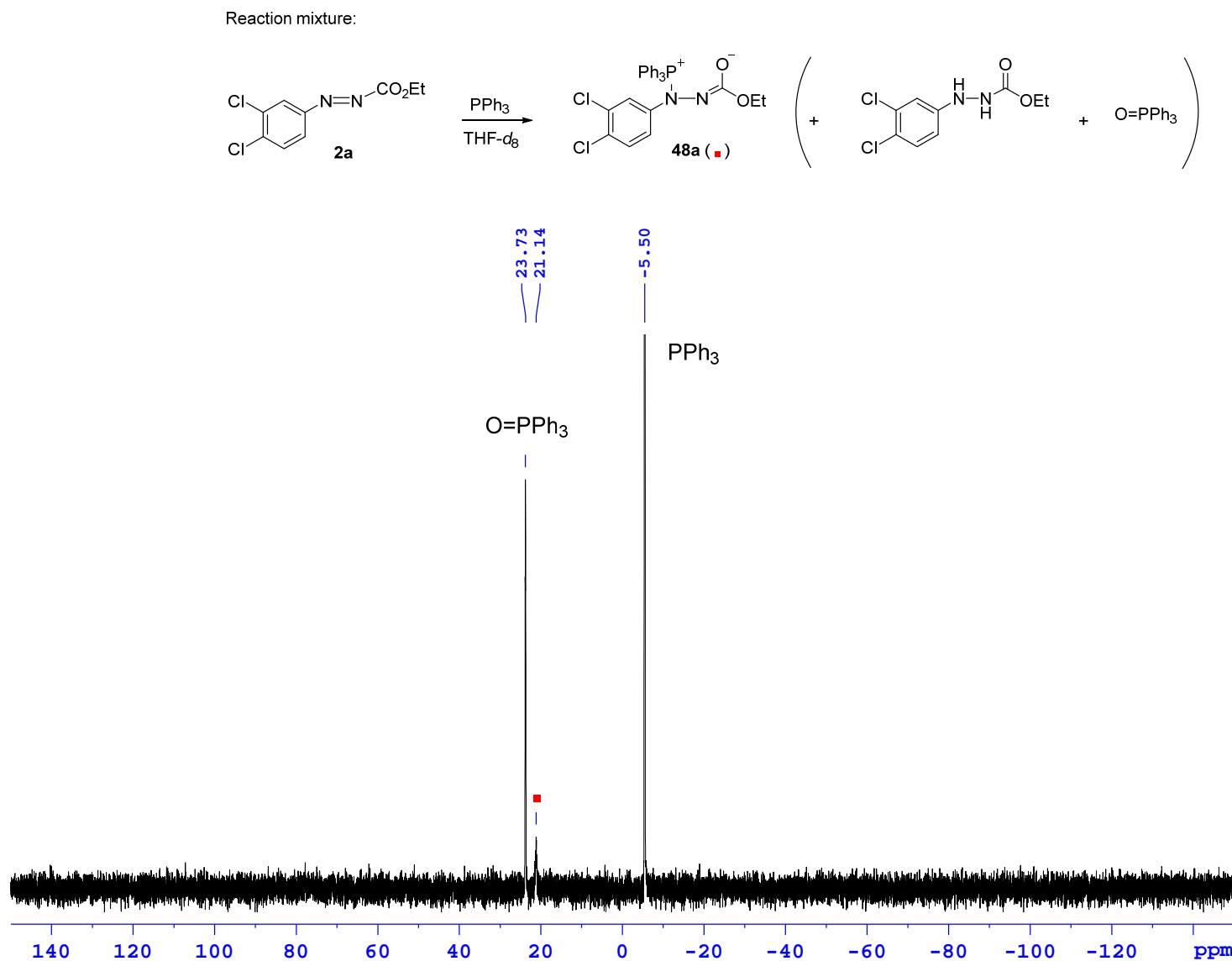


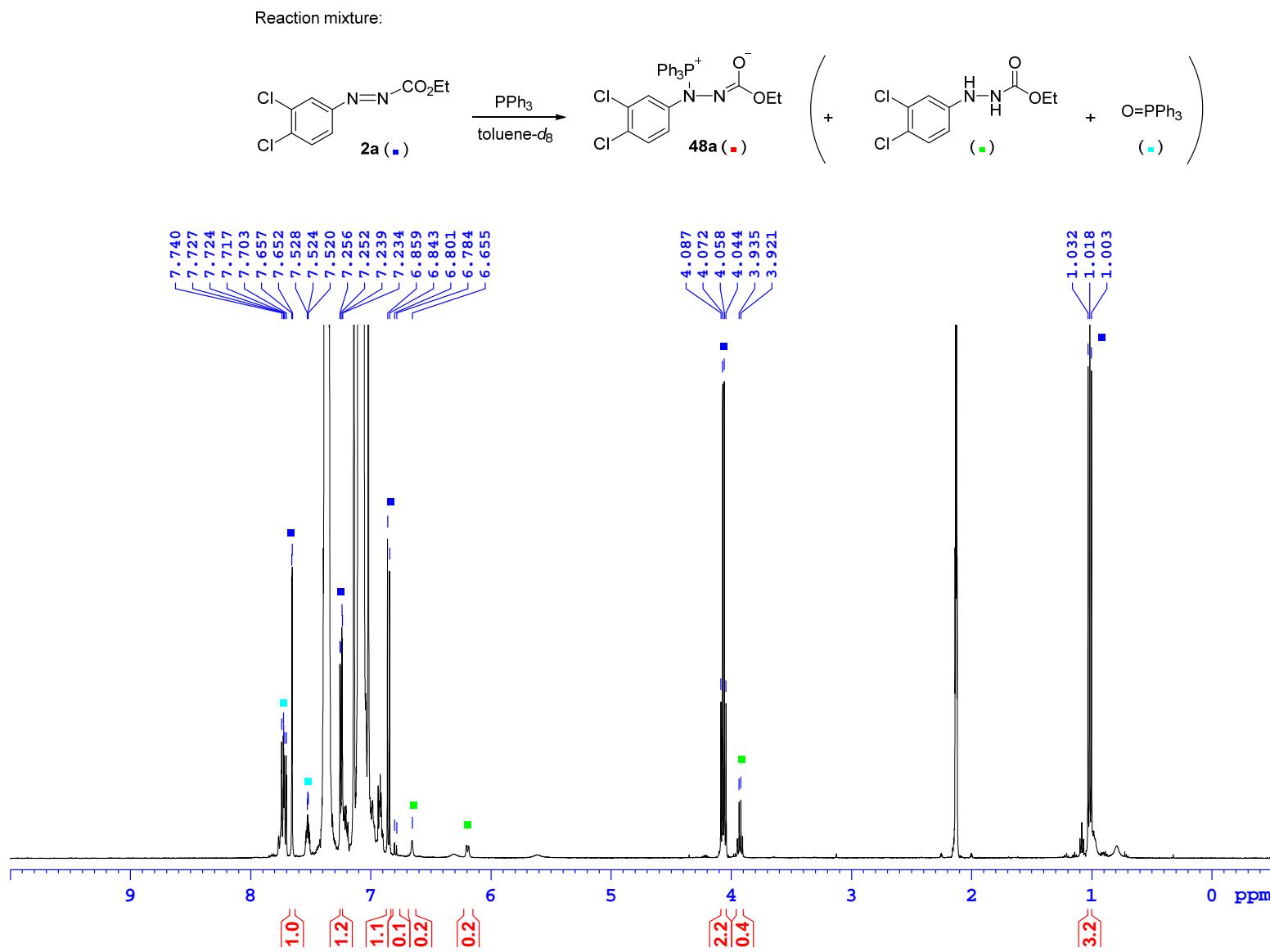
Current Data Parameters
 NAME MGS-625
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150910
 Time 17.13
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT THF
 NS 16
 DS 2
 SWH 10330.578 Hz
 FIDRES 0.157632 Hz
 AQ 3.1719923 sec
 RG 18
 DW 48.400 usec
 DE 6.50 usec
 TE 296.0 K
 D1 1.0000000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 8.90 usec
 PLW1 26.0000000 W
 SF01 500.1330885 MHz

F2 - Processing parameters
 SI 65536
 SF 500.1301070 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



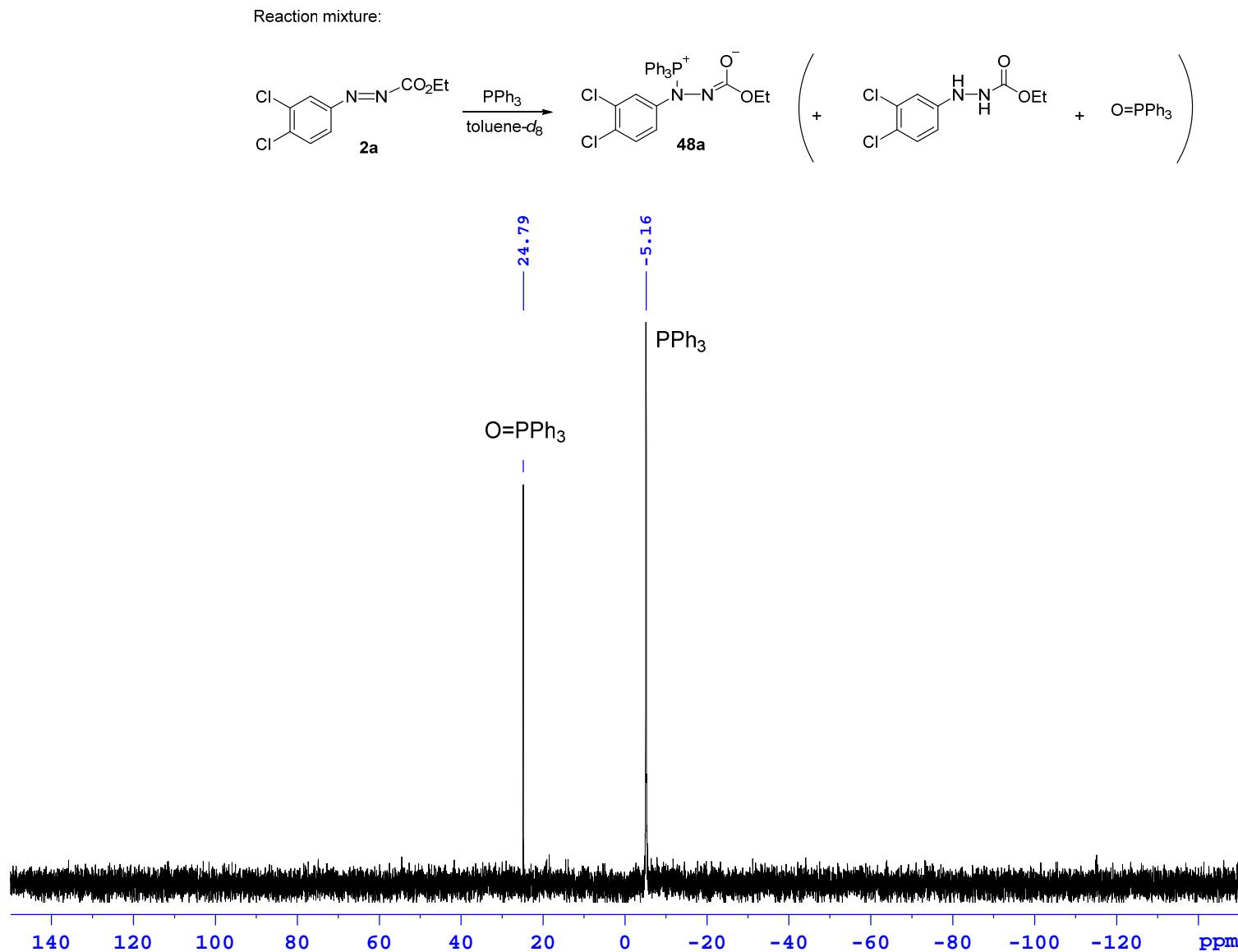


Current Data Parameters
NAME MGS-652
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20151022
Time 10.48
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT Tol
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 32
DW 48.400 usec
DE 6.50 usec
TE 296.0 K
D1 1.0000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 8.90 usec
PLW1 26.0000000 W
SF01 500.1330885 MHz

F2 - Processing parameters
SI 65536
SF 500.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Current Data Parameters
 NAME MGS-662
 EXPNO 3
 PROCNO 1

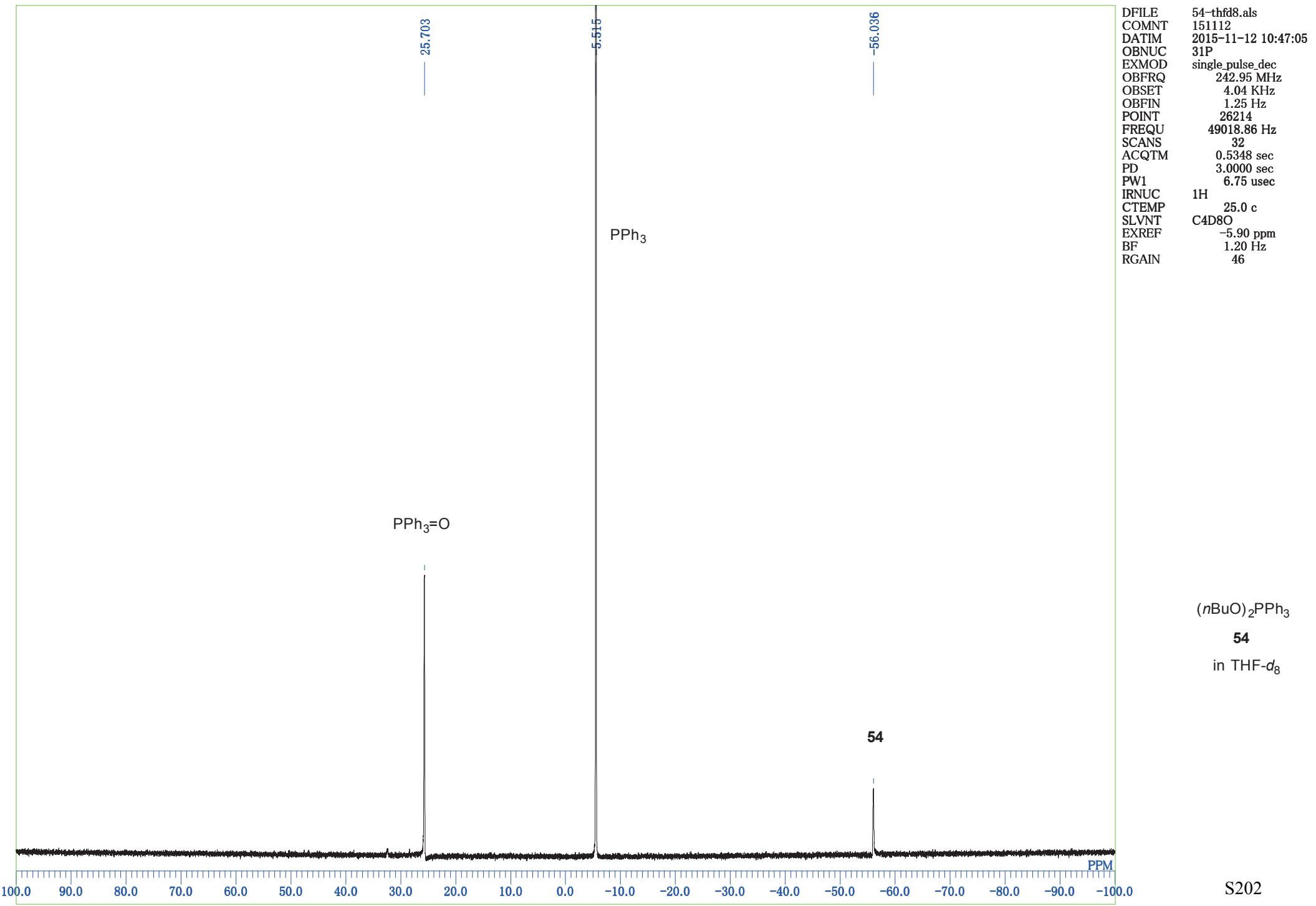
F2 - Acquisition Parameters
 Date_ 20151117
 Time 10.18
 INSTRUM spect
 PROBHD 5 mm Multinucl
 PULPROG zgpg30
 TD 65536
 SOLVENT Tol
 NS 32
 DS 4
 SWH 48661.801 Hz
 FIDRES 0.742520 Hz
 AQ 0.6734324 sec
 RG 10321.3
 DW 10.275 usec
 DE 6.00 usec
 TE 302.0 K
 D1 2.0000000 sec
 d11 0.0300000 sec
 DELTA 1.8999998 sec
 TD0 1

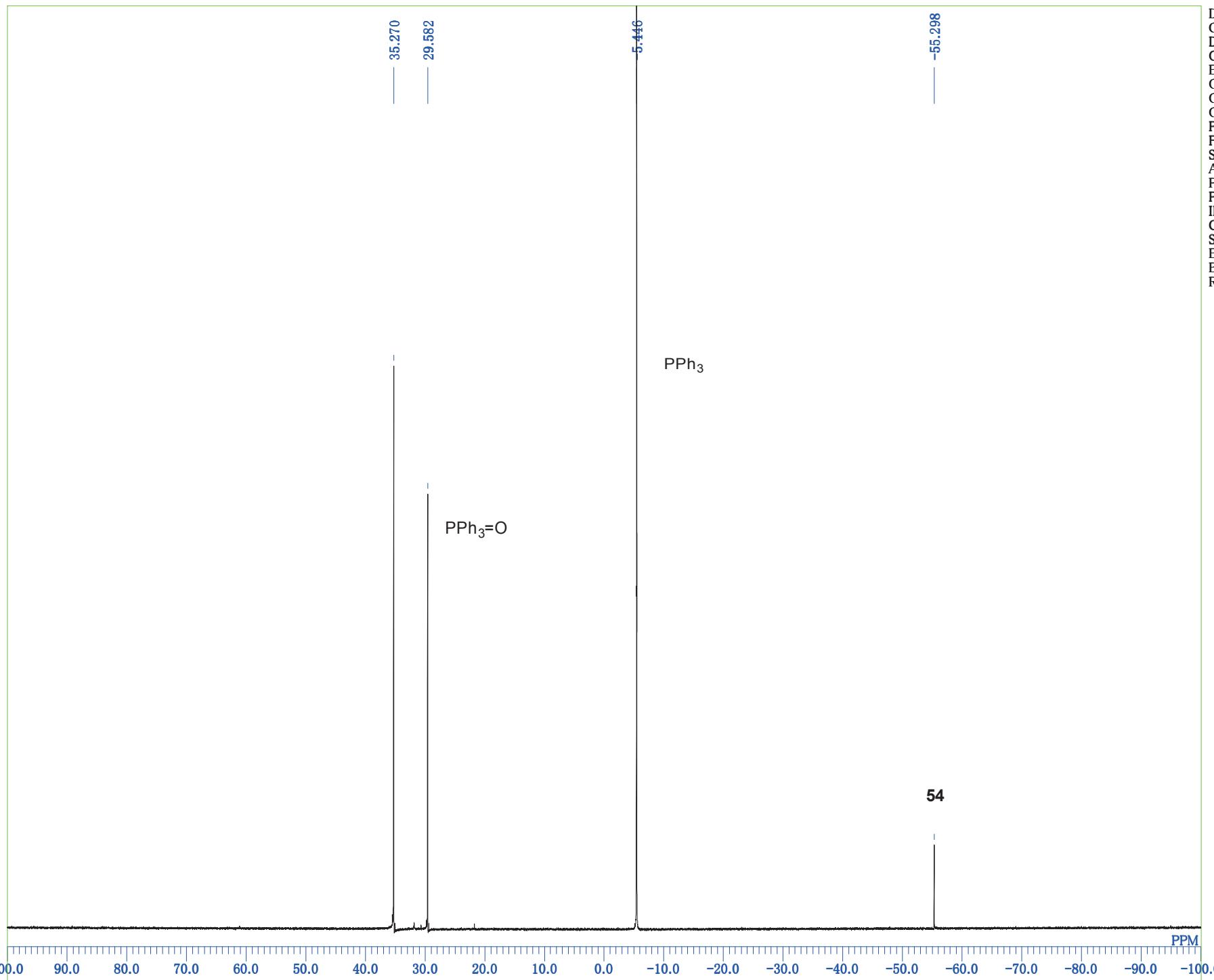
===== CHANNEL f1 ======
 NUC1 31P
 P1 10.60 usec
 PL1 -4.00 dB
 SF01 121.4887762 MHz

===== CHANNEL f2 ======
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 75.00 usec
 PL2 0 dB
 PL12 19.10 dB
 PL13 120.00 dB
 SF02 300.1312005 MHz

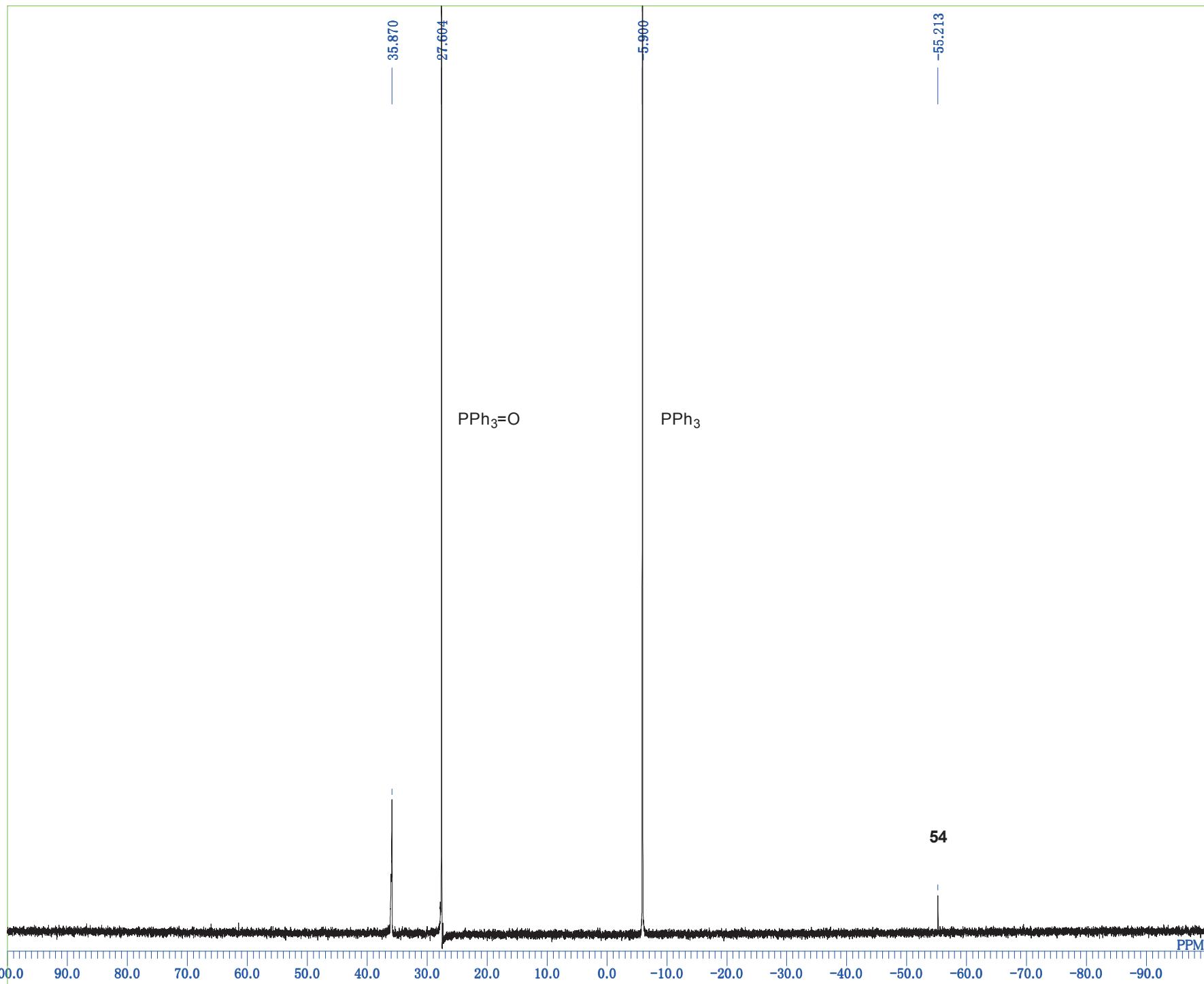
F2 - Processing parameters
 SI 32768
 SF 121.4948510 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

151112



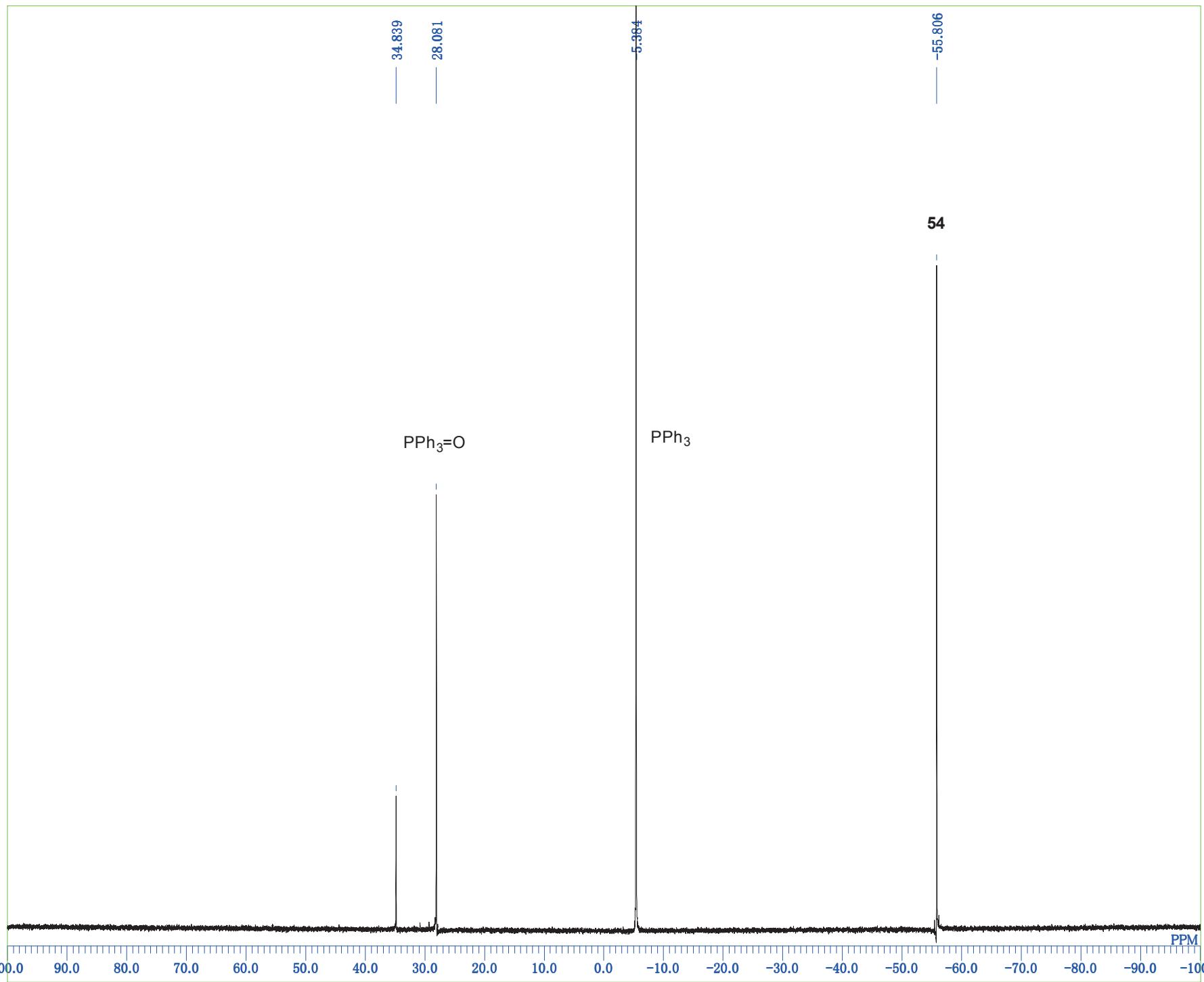


DFILE 54-cdcl3.als
 COMNT 151115
 DATIM 2015-11-15 13:08:19
 OBNUC 31P
 EXMOD single_pulse_dec
 OBFRQ 242.95 MHz
 OBSET 4.04 KHz
 OBFIN 1.25 Hz
 POINT 26214
 FREQU 49018.86 Hz
 SCANS 32
 ACQTM 0.5348 sec
 PD 3.0000 sec
 PW1 6.75 usec
 IRNUC 1H
 CTEMP 25.0 c
 SLVNT CDCL3
 EXREF -5.90 ppm
 BF 1.20 Hz
 RGAIN 46



DFILE 54-cd3cn.als
 COMNT 151115
 DATIM 2015-11-15 12:16:52
 OBNUC 31P
 EXMOD single_pulse_dec
 OBFRQ 242.95 MHz
 OBSET 4.04 KHz
 OBFIN 1.25 Hz
 POINT 26214
 FREQU 49018.86 Hz
 SCANS 32
 ACQTM 0.5348 sec
 PD 3.0000 sec
 PW1 6.75 usec
 IRNUC 1H
 CTEMP 25.0 c
 SLVNT CD3CN
 EXREF -5.90 ppm
 BF 1.20 Hz
 RGAIN 50

$(n\text{BuO})_2\text{PPh}_3$
54
 in CD_3CN



DFILE 54-toluened8.als
 COMNT 151114
 DATIM 2015-11-14 14:55:41
 OBNUC 31P
 EXMOD single_pulse_dec
 OBFRQ 242.95 MHz
 OBSET 4.04 KHz
 OBFIN 1.25 Hz
 POINT 26214
 FREQU 49018.86 Hz
 SCANS 32
 ACQTM 0.5348 sec
 PD 3.0000 sec
 PW1 6.75 usec
 IRNUC 1H
 CTEMP 25.0 c
 SLVNT C6D5CD3
 EXREF -5.90 ppm
 BF 1.20 Hz
 RGAIN 46

1a-beforeafter.als
 141222
 2014-12-22 19:46:55
 1H
 single_pulse.ex2
 399.78 MHz
 4.19 KHz
 7.29 Hz
 13107
 6002.31 Hz
 8
 2.1837 sec
 2.0000 sec
 5.00 usec
 1H
 20.4 c
 CDCL3
 0.00 ppm
 0.12 Hz
 50

DFILE
 COMNT
 DATIM
 OBNUC
 EXMOD
 OBFRQ
 OBSET
 OBFIN
 POINT
 FREQU
 SCANS
 ACQTM
 PD
 PW1
 IRNUC
 CTEMP
 SLVNT
 EXREF
 BF
 RGAIN

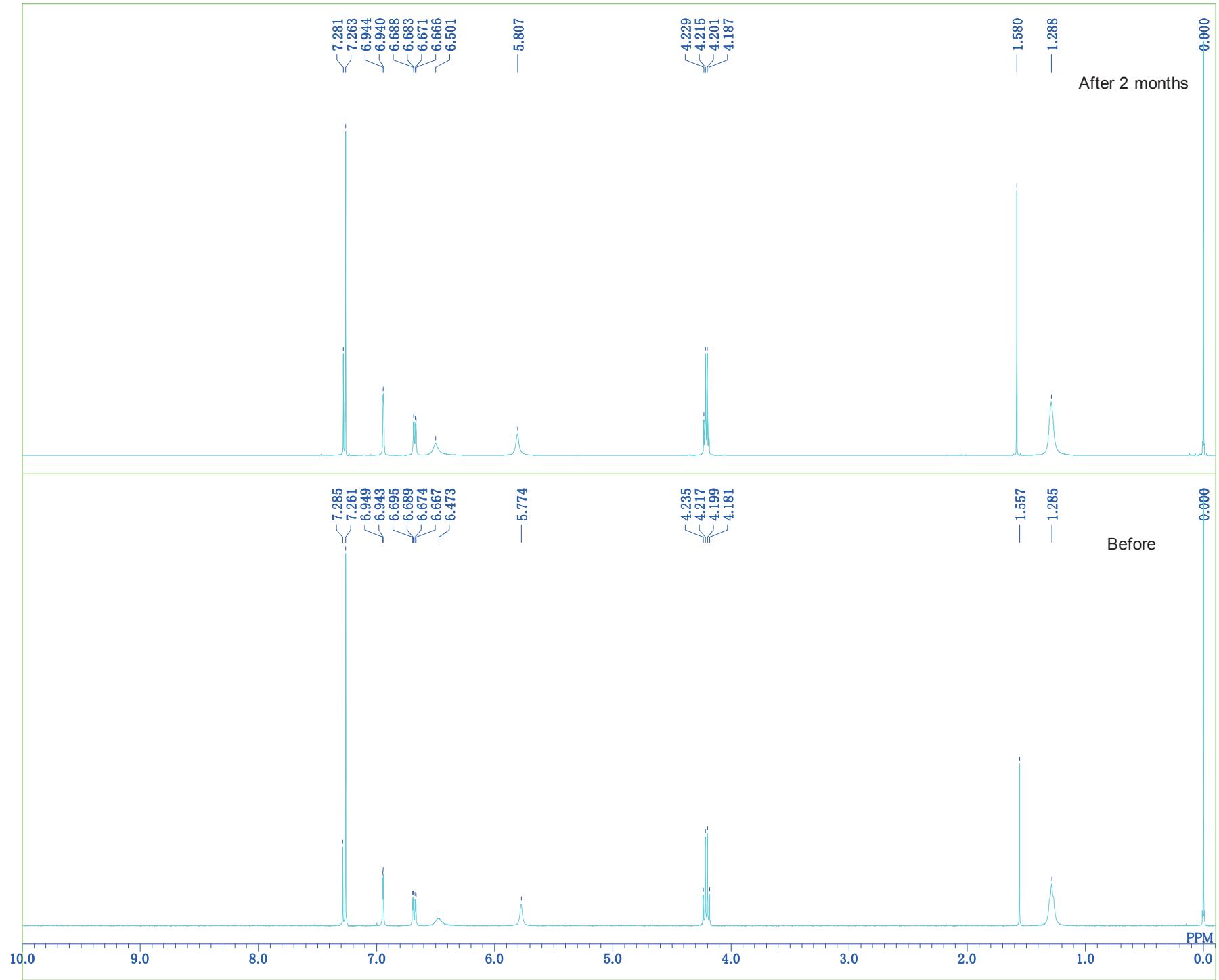
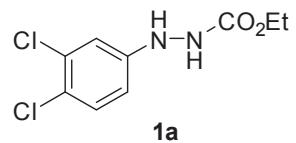
After 2 months

0.000

0.000

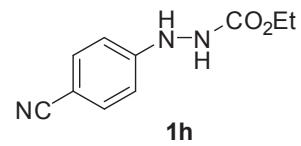
Before

PPM



1h-beforeafter.als
 150501
 2015-05-01 14:11:40
 1H
 single_pulse.ex2
 399.78 MHz
 4.19 KHz
 7.29 Hz
 13107
 6002.31 Hz
 8
 2.1837 sec
 2.0000 sec
 4.70 usec
 1H
 22.9 c
 CDCL3
 0.00 ppm
 0.12 Hz
 48

After 2 months



0.000

PPM

Before

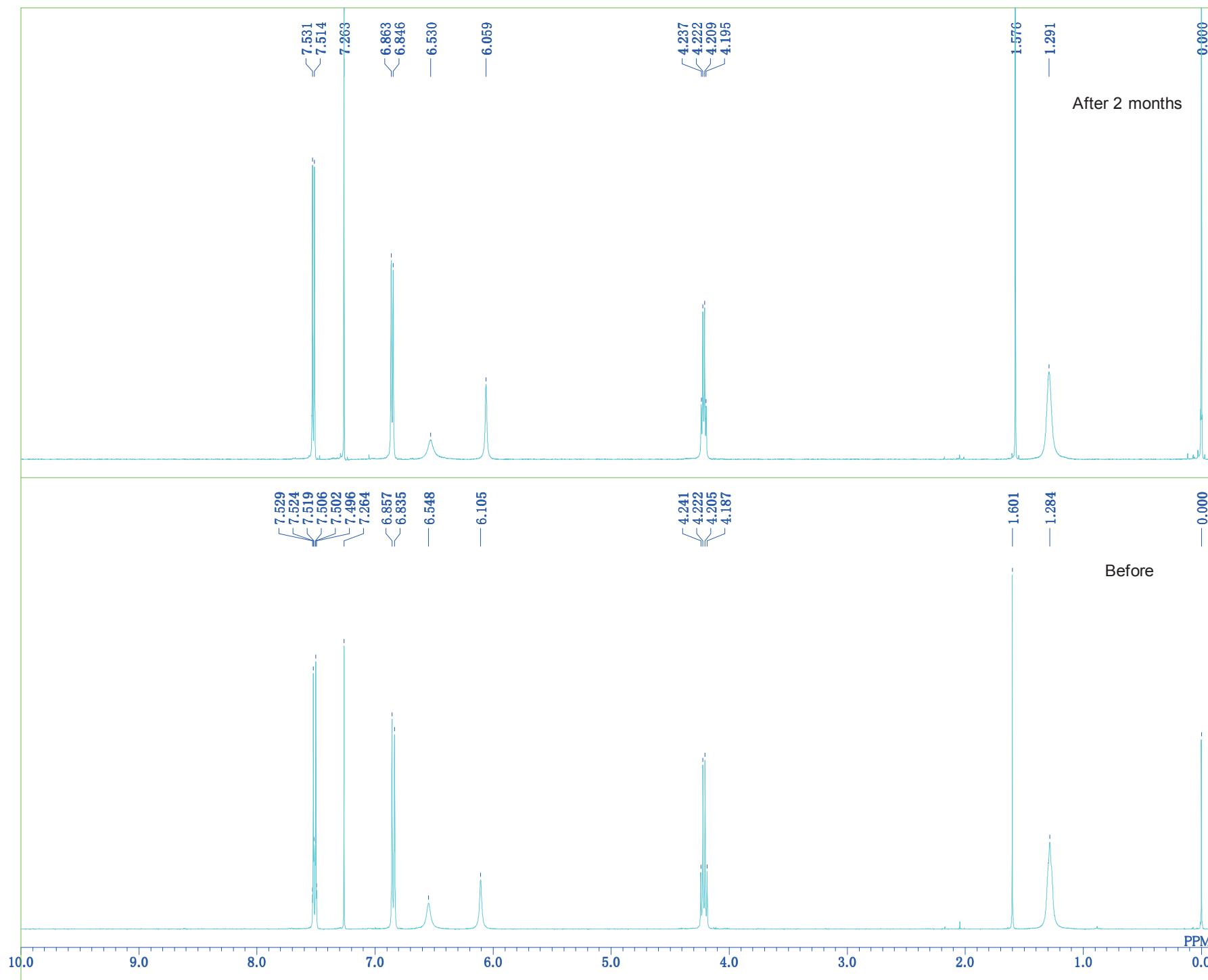
0.000

PPM



7.531
 7.514
 7.263
 6.863
 6.846
 — 6.530
 — 6.059

7.524
 7.519
 7.506
 7.502
 7.496
 7.264
 6.857
 6.835
 — 6.548
 — 6.105



2a-beforeafter.als
 150524
 2015-05-24 13:27:23
 1H
 single_pulse.ex2
 600.17 MHz
 5.30 KHz
 5.47 Hz
 26214
 9008.87 Hz
 8
 2.9098 sec
 SCANS
 ACQTM
 PD
 PW1
 IRNUC
 CTEMP
 SLVNT
 EXREF
 BF
 RGAIN

1H
 22.9 c
 CDCL3
 0.00 ppm
 0.12 Hz
 54

DFILE
 COMNT
 DATM
 OBNUC
 EXMOD
 OBFRQ
 OBSET
 OBFIN
 POINT
 FREQU
 SCANS
 ACQTM
 PD
 PW1
 IRNUC
 CTEMP
 SLVNT
 EXREF
 BF
 RGAIN

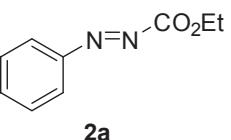
— 0.000
 PPM

After 2 months

1.569
 1.490
 1.476
 1.461

4.551
 4.537
 4.522
 4.509

8.028
 8.023
 7.827
 7.824
 7.811
 7.807
 7.646
 7.629
 — 7.264



Before

1.548
 1.486
 1.474
 1.462

4.545
 4.533
 4.521
 4.509

8.025
 8.022
 7.822
 7.818
 7.808
 7.804
 7.643
 7.628
 — 7.262

2h-beforeafter.als
 150520
 2015-05-20 15:22:12
 1H
 single_pulse.ex2
 500.16 MHz
 2.41 KHz
 6.01 Hz
 13107
 7507.39 Hz
 8
 1.7459 sec
 2.0000 sec
 5.80 usec
 1H
 20.0 c
 CDCL3
 0.00 ppm
 0.12 Hz
 48

DFILE
 COMNT
 DATIM
 OBNUC
 EXMOD
 OBFRQ
 OBSET
 OBFIN
 POINT
 FREQU
 SCANS
 ACQTM
 PD
 PW1
 IRNUC
 CTEMP
 SLVNT
 EXREF
 BF
 RGAIN

After 2 months



— 7.265



— 7.265



— 0.000

Before



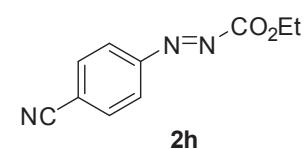
— 7.267



— 7.267



— 0.000



10.0 9.0 8.0 7.0 6.0 5.0 4.0 3.0 2.0 1.0 0.0 PPM

S209

DFILE
 COMNT
 DATIM
 OBNUC
 EXMOD
 OBFRQ
 OBSET
 OBFIN
 POINT
 FREQU
 SCANS
 ACQTM
 PD
 PW1
 IRNUC
 CTEMP
 SLVNT
 EXREF
 BF
 RGAIN

151105
 2015-11-05 16:06:46
 1H
 single_pulse.ex2
 500.16 MHz
 2.41 KHz
 6.01 Hz
 13107
 7507.39 Hz
 8
 1.7459 sec
 2.0000 sec
 5.80 usec
 1H
 19.6 c
 CDCL3
 0.00 ppm
 0.12 Hz
 30

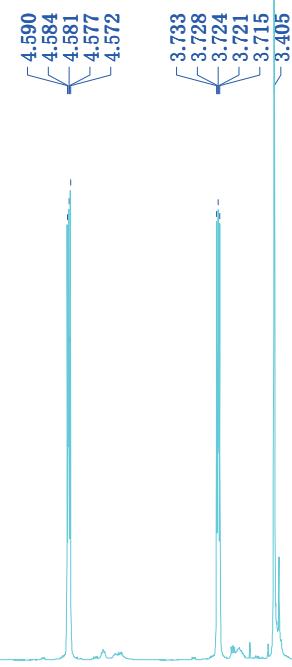
DFILE
 COMNT
 DATIM
 OBNUC
 EXMOD
 OBFRQ
 OBSET
 OBFIN
 POINT
 FREQU
 SCANS
 ACQTM
 PD
 PW1
 IRNUC
 CTEMP
 SLVNT
 EXREF
 BF
 RGAIN

DMEAD-beforeafter.xls
 151105
 2015-11-05 16:06:46

S210

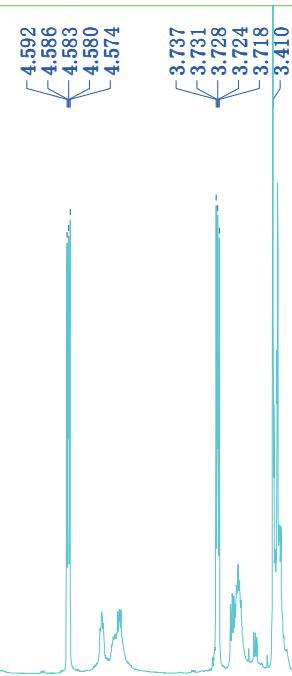
— 0.000 — PPM

Before



— 7.302 —

After 2 months



— 7.272 —

