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Electronic Supplementary Information

Active Site Structure and Absorption Spectrum of Channelrhodopsin-2 Wild-Type and C128T Mutant

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1. Modelling ChR2: MM Equilibration

As described in the main text, ChR2-WT and ChR2-C128T models were built using a homology model of the C1C2 chimera X-ray structure (PDB code: 3UG9) as template. From there, we set up the respective protein dimers surrounded by a POPC membrane and water molecules as solvent. The complete model is constituted by 492 amino acids (246 amino acids per monomer), 107 POPC molecules, and 6882 water molecules (6880 water molecules in ChR2-C128T, since no water molecule is expected in the DC gate of each monomer, see main text), in a box of 8.22 nm \times 6.74 nm \times 7.45 nm.

After setting the protonation state of the protein residues (see main text), the structure of the active site was modeled taking into account previous spectroscopic and QM/MM dynamics simulations (see refs. [19-22] and references therein). More in detail, both E123 and D253 are deprotonated, and the side chain of E123 forms a hydrogen bond with the side chain of T127, allowing a water molecule to stay next to the RSBH⁺ (Figure 1S).

The first step was the minimization of the potential energy. This was achieved by calculating 1000 steps with the steepest-descent method (tolerance of 1000 kJ mol⁻¹ nm⁻¹). The next step was the equilibration of the solvent, which was achieved by simulating a NVT-ensemble for 10 ps at a temperature of 300 K, applying the Berendsen thermostat [1S] (time step: 2 fs). The heavy atoms of the protein and lipid membrane were restrained to their initial positions by harmonic potentials (force constant: 1000 kJ mol⁻¹ nm⁻¹). Subsequently both (wt and C218T) systems (including the protein dimer and the lipid bilayer) were equilibrated for 10 ns in the NPT-ensemble, at a pressure of 1 bar (Parrinello–Rahman barostat [2S]) and a temperature of 300 K (Nosé–Hoover thermostat [3S,4S]). The time step was set to 2 fs, using the H-bond scheme for constraints as implemented in the LINCS algorithm within the GROMACS package.

As described in the main text, MM force fields show severe shortcomings in the description of strongly hydrogen bonded networks. This has been demonstrated in particular for the BR active site. Using standard force fields, the crystal water binding to the Schiff-base NH group is lost and leaves the active site. A similar finding has been reported for ChR in our previous papers (see refs. [18,19]). If this water is lost during MM equilibration, it will not be restored in QM/MM simulations due to the limited simulation time. Therefore, to avoid a bias due to the above described 10 ns MM equilibration, we harmonically restrained the positions of the oxygen atom of the water molecule and of the nitrogen and hydrogen atoms of the RSBH⁺ were with a force constant of 10000 kJ mol⁻¹ nm⁻¹ during the whole MM equilibration procedure. Moreover, the hydrogen bond between the hydrogen atom of T127 and the oxygen atom of E123 was constrained to a length of 0.18 nm. Note that we manually rotated the side chain of E123 and the water molecule for each monomer, therefore, the two monomers are not guaranteed to have exactly the same starting active site structure. This further results in two starting points for the production QM/MD simulations as shown in Figure 1S. The Charmm36 force field was applied

in the framework of the GROMACS software package.

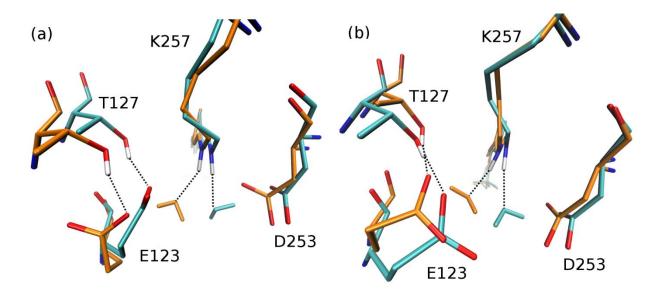


Figure 1S. Initial conditions selected for MM equilibration of the proposed ChR2 models: hydrogen bonds are set between E123 and T127 side chains, and between a water molecule (H₂O) and the protonated Schiff base, RSBH⁺ (Ret: retinal). (a) ChR2-WT, (b) ChR2-C128T. The two structures are shown in backbone colors of cyan and orange, respectively.

2. The DC/DT Gate Structures

For ChR2-WT, the MM equilibration simulations show two different DC gate structures: (i) one water molecule bridges D156 and C128 (water bridged DC gate, Figure 2S (a)); (ii) D156 and C128 form a direct hydrogen bond (direct DC gate, Figure 2S (b)), where D156 serves as a donor and C128 as an acceptor. In the starting structures, both monomers contain the water as shown in Figure 2S,a. During equilibration, however, the water in monomer 2 is lost, while it stays in monomer 1. The water bridged DC gate has been proposed by previous classical MD simulations [18] and validated by QM/MM vibrational frequency calculations [33]. To further check this assignment for the present models, we calculated the stretching vibrational frequency of D156 side chain (vCOO) for the above two DC gate structures using the normal mode analysis (NMA) with the CHARMM37b1 program at the DFTB3/MM level of theory, which has been validated in detail in ref [33]. The residues C128, D156, and the bridging water molecules (for water bridged DC gate) were included in the QM region, and the rest was treated as MM part. The calculated vCOO value is 1730 cm⁻¹ for the water bridged DC gate structure, which agrees well with the experimental data (1735 cm⁻¹) [5S], and 1693 cm⁻¹ for the direct DC gate structure, i.e. 42 cm⁻¹ red shifted compared with the FTIR value. We take this as a strong indication for the presence of one water molecule in the DC gate. The DC gate structure is strongly coupled to the active site, and the active site structures thus differ for the water bridged and direct DC gate models. Therefore, since a missing water in the DC gate may lead to wrong active site structures,

we discard the simulations of the monomer without DC water in the further analysis with QM/MM simulations.

To summarize, our findings in principle suggest that it is crucial to include the DC gate into the QM description since, in the MM description, the DC gate motif is not stable in ChR, and the bridging water tends to leave. This, however, happens during the extended MM equilibration step, which unfortunately cannot be performed using QM/MM, due to the long simulation times. Therefore, based on MM equilibration we had to discard trajectories where the water leaves the DC gate.

Regarding the QM/MM simulations, as long as the DC gate structure is intact, the DC gate does not have to be included into the QM zone because it is far away from the counterion region. If the DC gate is broken, however, the active site structure is severely impaired. Therefore, we monitored the DC gate during the QM/MM simulations. Including the DC gate into the QM zone would increase the simulation times significantly. However, we would like to note that all calculations were performed in a QM/MM environment, therefore the Coulomb effect (which is mainly responsible for the separation of excited states) is always taken into account.

For ChR2-C128T, the simulations sample a single direct hydrogen bond between T128 and D156 (Figure 2S (c)), as discussed in a previous theoretical study [33].

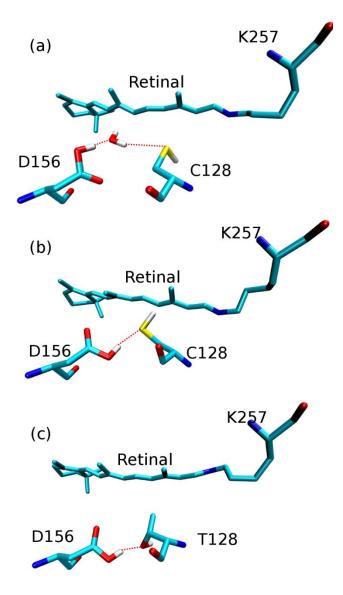


Figure 2S. The DC (D156-C128)/DT (D156-T128) gate structure sampled in the present simulations. (a) water bridged DC gate of ChR2-WT; (b) direct DC gate of ChR2-WT; (c) direct DT gate of ChR2-C128T.

3. Modelling ChR2: QM/MM Production Run

After MM equilibration, we performed QM/MM MD simulations with the GROMACS package using DFTB3 for QM region and the Charmm36 classical force field for the remainder of the system. The QM region (S-QM) includes the retinal, Lys257, the E123, D253, and K93 side chains, and four (three for ChR2-C128T) water molecules involved in the active site hydrogen bonding patterns. Four hydrogen link atoms are needed to treat the QM/MM boundary regions where a covalent bond is involved, *i.e.* between α - and β -carbon atoms of E123, D253, K257 and K93 residues.

The restraints imposed on the active site during the MM equilibration were gradually removed

when including the active site in the QM region of the DFTB3/Charmm36 model, as summarized in Table 1S. Weak restraints on the QM waters were used in order to keep the active site intact. Otherwise, QM and MM water molecules could interchange. The constraints on the water molecules are weak (500 kJ mol⁻¹ nm⁻¹). Hence, we do not expect sensible changes if a fully relaxed model is employed, as emphasized in the manuscript.

Table 1S. Gradual (step-by-step) reduction of position restraints and distance constraints while performing QM/MM simulations. The last step (step 7) corresponds to the QM/MM production run. The procedure required a total time of seven weeks on a single processor for each trajectory.

Step	Simulation time [ps]	Position restraint on non-water atoms [kJ mol ⁻¹ nm ⁻¹]	Distance constraint (E123-T127 H-bond)	Position restraint on QM water atoms (only oxygen atoms) [kJ mol ⁻¹ nm ⁻¹]
Step 1	200	5000	Yes	500
Step 2	200	3000	Yes	500
Step 3	200	2000	Yes	500
Step 4	200	1000	Yes	500
Step 5	200	500	Yes	500
Step 6	200	0	No	500
Step 7	1000	0	No	500

ChR2-WT The different DC gate structures of monomer 1 and 2 lead to different structures of the active site. This is not surprising, since a strong coupling between active site and DC gate is expected. The simulation with direct DC gate (monomer 2) shows a stable active site with RSBH⁺-E123 salt bridge during the entire 2 ns simulation time, which is different from the stable RSBH⁺-H₂O hydrogen bonding pattern sampled by the simulations with water bridged DC gate (monomer 1). It is unclear whether monomer 1 will also sample salt-bridge structures to a larger extent. Therefore, we decided to discard the monomer 2 structure and extend the sampling of monomer 1. We randomly selected 6 snapshots from the 2 ns trajectory with water bridged DC gate as the starting points for the production QM/MD simulations. Additionally, to get a more accurate description of the hydrogen bond between E123 and T127, we chose a larger QM region

(L-QM), enlarging the S-QM region by including T127. This gave rise to an additional hydrogen link atom for residue T127. New velocities were generated for each simulation.

<u>ChR2-C128T</u> The simulations sample two E123 side chain conformations as shown in Figure 4 of the main text during the 2 ns simulation time, i.e. the E123 side chain either orients towards the cytoplasmic side or moves downwards to the extracellular side. The downshift of E123 breaks the hydrogen bond between E123 and T127 side chains. Note that one MM water molecule stays stable in the active site and forms the water cluster together with the three other QM water molecules. Considering the limitations of MM force fields in describing strong hydrogen bonds in proteins, we chose the same L-QM region on ChR2-C128T as on ChR2-WT. For statistical analysis, we randomly selected 6 snapshots from the 2 ns trajectory of each simulation as the starting points for the production QM/MD simulations.

The results in the main text are based on the simulations with L-QM for both ChR2-WT and ChR2-C128T.

13-cis,**15-syn Retinal** The chemical structures of all-*trans* retinal and 13-cis, 15-syn retinal are shown in Figure 3S (a) and (b), respectively. The cis retinal from BR (PDB code: 1X0S) was aligned to the all-*trans* retinal to build the cis retinal bound ChR2-WT and ChR2-C128T structures. The RSBH⁺ has similar positions in the all-*trans* retinal bound complex and the 13-cis, 15-syn retinal bound complex as shown in Figure 3S (c). The snapshots used for aligning were randomly selected from the all-*trans* retinal trajectories with L-QM. After replacing with cis retinal, the protein and the water environment were equilibrated with position restraints (force constant: 1000 kJ mol⁻¹ nm⁻¹) on the retinal, K93, E123, T127, D253, and four water molecules involved in the active site hydrogen bonding patterns, to remove the inappropriate atomic conflicts. The equilibration was performed at the MM level of theory using the steepest descent method. The obtained structures were used as starting points for the production QM/MD simulations with the L-QM region.

BR For comparison and to compute the excitation energy shifts, we performed a 1 ns QM/MM simulation of the BR ground state, using a starting structure from our previous study [9]. BR shows a very stable and distinct active site structure, the famous pentagonal water cluster as shown in Figure 1. As we have shown in our earlier publications [58], QM/MM MD simulations only show vibrations around this stable ground states structure, no other local minima occur. Therefore, the amount of sampling to cover this phase-space is much reduced with respect to ChR, where we have shown the large structural heterogeneity. Therefore, the sufficient sampling must assure that the trajectories visit the different local minima sufficiently, in order to converge the statistics. So the BR active site structure is suitably sampled within 1 ns simulation (1000 snapshots).

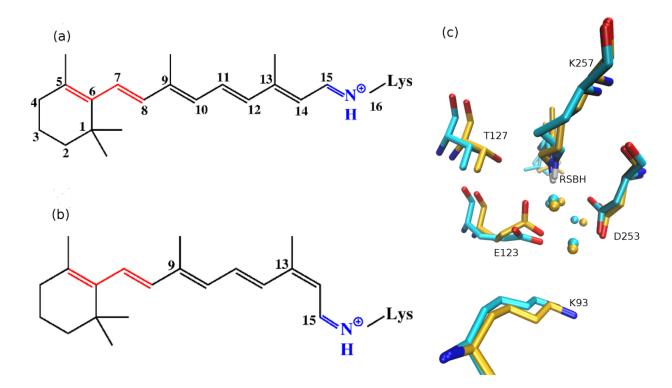


Figure 3S. The chemical structures of (a) all-*trans* retinal and (b) 13-*cis*, 15-*syn* retinal. (c) The alignment of the 13-*cis*, 15-*syn* retinal (orange backbone) to the all-*trans* retinal (cyan backbone) shows that the RSBH⁺ has similar positions in the protein. The water molecules are shown in orange and cyan balls, respectively.

<u>Simulations at 100K</u> In solid state NMR studies at 100 K [16], the authors observed a shorter distance between RSBH⁺ and counterion in ChR2 than in BR, which is in conflict with our results at 300 K: BR (3.83 Å) and ChR2 (3.82 Å) have almost the same RSBH⁺-counterion distance. Nevertheless, a previous study by our group on BR showed a different active site at low temperature with respect to that at room temperature.[38] To get an insight into the temperature dependence of our system, we therefore performed also simulations at 100 K.

We selected 14 snapshots carrying the three hydrogen bonding patterns and two E123 side chain conformations (see main text for details), from the all-*trans* retinal bound ChR2-WT production trajectories. They were used as starting points of 14 independent cooling simulations of 1 ns each. The system was cooled down from 300 K to 100 K within 10 ps, and maintained at 100 K during the remaining simulation time. The same QM/MM setups as for the 300 K production simulations in the main text were adopted.

Table 2S. Summary of structural motifs at 300K (beginning of the simulation) and after the cooling process at 100K.

Trajectory no.	Structural pattern at 300K (beginning of simulation)	Structural pattern at a simulation temperature of 100K
1	E123 up*	E123 up
2	E123 up	E123 up
3	E123 up	E123 up
4	E123 up	E123 up
5	H2O up**	E123 up
6	H2O up	E123 up
7	D253 up***	E123 up
8	D ₂ 53 up	E123 up
9	D253 up	E123 up
10	H2O up	H2O up
		E123 up
11	D253 down ⁺	D253 down
12	D253 down	D253 down
13	H2O down ⁺⁺	H2O down
		D253 down
14	H2O down	H2O down

^{*}E123 up: -RSBH⁺····⁻O-(**E123**) and E123-upward

^{**}H2O up: -RSBH+····OH₂; E123-upward

^{***}D253 up:-RSBH⁺····⁻O-(**D253**); E123-upward

⁺D253 down: -RSBH⁺····⁻O-(**D253**); E123-downward

⁺⁺H2O down: -RSBH⁺····O-(**H2O**); E123-downward

Table 3S. The average distance between the RSBH⁺ nitrogen atom and the center of mass of E123 side chain carboxyl oxygens for the nine trajectories sampling the -RSBH⁺····O-(E123) pattern (unit: Å). For comparison, the same distance averaged on the BR trajectory is 3.83 Å and on ChR2-WT (all-*trans* at 300K) is 3.82 Å.

Trajectories	1	2	3	4	5	6	7	8	9
RSBH ⁺ -OOC(E123) distance	2.76	2.73	2.79	3.24	3.33	2.99	2.83	3.07	2.75

4. SORCI vs OM2/MRCI: Excitation Energies

The SORCI calculations at 50 QM/MM optimized ChR2-WT geometries reveal a major difference in the electronic excited-state description with respect to BR: for both BR and ChR2 (-WT and -C128T) the optically-bright electronic transition is a $^{1}(\pi,\pi^{*})$ transition, but while in BR it is always a $S_0 \rightarrow S_1$ vertical excitation, in ChR2 it corresponds to a $S_0 \rightarrow S_1$ or to a $S_0 \rightarrow S_2$ vertical excitation. In ChR2, S₁ is described – for some of the sampled structures – by a double $^{1}(\pi,\pi^{*})$ transition (i.e. a transition involving the promotion of two electrons from bonding to antibonding π orbitals) associated with a low oscillator strength (optically dark electronic transition). The corresponding double ${}^{1}(\pi,\pi^{*})$ transition corresponds always to S_{2} in BR. This inversion between the S₁ and S₂ electronic states may be due to the different active site structure, as already been suggested to explain the hypsochromic absorption shift of ChR2 with respect to BR (as discussed in the main text). Moreover, it may lead to a different description of the transto-cis photoisomerization process. In the main text, the optically bright electronic transition was always taken into account to calculate the absorption spectra. Therefore, all SORCI calculations were performed with three roots $(S_0, S_1 \text{ and } S_2)$. In order to test the validity of this approach, we show here (Table 2S) the effect of the number of roots on the energy and oscillator strength of the lowest-lying optically bright electronic transition. Two ChR2-C128T models with different active site structures were tested. As it can be seen, the increase of the number of calculated excitation states does not have much influence on the single excitation energy, hence validating our approach.

Table 4S. Single-point excitation energy (E) and oscillator strength (f_{osc}) of the lowest-lying optically bright electronic transition, when including 3 to 6 roots (including the ground state). Two ChR2-C128T geometries were tested. Level of theory: SORCI, complete active space of 12 electrons in 12 orbitals, split-valence basis set def2-SV(P).

	Geometry 1		Geometry 2			
N _{root}	E / eV (nm)	$f_{ m osc}$	N _{root}	E / eV (nm)	$f_{ m osc}$	
3	2.61 (476)	1.83	3	2.98 (416)	1.98	
4	2.64 (469)	1.84	4	3.01 (412)	2.05	
5	2.69 (461)	1.86	5	3.00 (413)	2.03	
6	2.59 (478)	1.79	6	3.00 (412)	1.89	

For the sake of comparison, OM2/MRCI calculations were performed at the same 50 QM/MM optimized ChR2-WT geometries as in the SORCI case using the same QM region. The OM2/MRCI calculations employed closed-shell molecular orbitals (MOs) and an active space of 20 electrons in 20 MOs (i.e. the 10 highest occupied MOs and the 10 lowest unoccupied MOs). The reference configurations comprised the closed-shell ground-state configuration and four configurations generated by excitations from the two highest occupied MOs $(\pi, \pi-1)$ to the two lowest unoccupied MOs (π^*, π^*+1) , i.e. the single excitations $\pi \rightarrow \pi^*$ and $\pi-1 \rightarrow \pi^*+1$ as well as the double excitations $(\pi, \pi) \rightarrow (\pi^*, \pi^*)$ and $(\pi, \pi-1) \rightarrow (\pi^*, \pi^*+1)$. All single and double excitations from the reference configurations were included in the MRCI calculations. The same MRCI treatment was applied in the single-point calculations at the snapshot geometries extracted from the QM/MM MD simulations (see main paper).

The cross-correlation function between the SORCI and OM2/MRCI results is shown in Figure 4S, and the simulated absorption spectra are compared in Figure 5S. The relative intensity of each histogram in Figure 5S is calculated as the normalized count of geometries in a defined energy window, multiplied by the average oscillator strength within the same energy window. The energy of each single-point excitation energy can be found in Table 3S. We conclude that a shift of the OM2/MRCI spectra by ca. 0.3 eV can almost quantitatively reproduce the SORCI spectra.

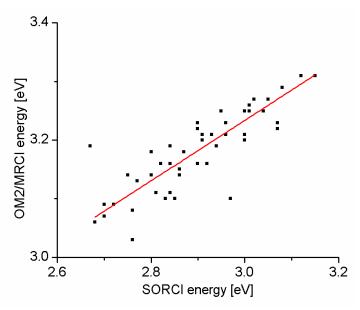


Figure 4S. Cross-correlation function between SORCI and OM2/MRCI excitation energies for the optically-bright electronic transitions. $R^2 = 0.832$.

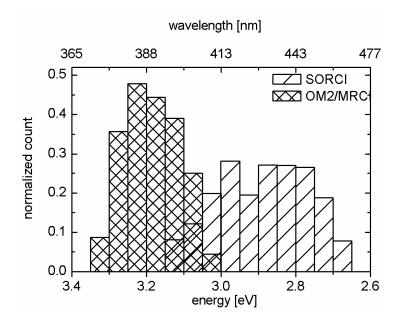


Figure 5S. Simulated ChR2-WT absorption spectra at the SORCI and OM2/MRCI levels of theory. The two histograms are centered at 2.9 and 3.2 eV, respectively (energy shift of ca. 0.3 eV).

Table 5S. Single-point excitation energies from SORCI and OM2/MRCI calculations (unit: eV).

SORCI	OM2/MRCI	ΔE _(SORCI-OM2)
3,07	3,22	-0,15
2,7	3,07	-0,37
3	3,21	-0,21
3,02	3,27	-0,25
3,01	3,25	-0,24
2,9	3,23	-0,33
3	3,25	-0,25
2,95	3,25	-0,3
2,72	3,09	-0,37
3,04	3,25	-0,21
2,81	3,11	-0,3
2,9	3,16	-0,26
2,67	3,19	-0,52
2,94	3,19	-0,25
2,76	3,08	-0,32
2,84	3,16	-0,32
2,76	3,03	-0,27
2,97	3,1	-0,13
2,85	3,1	-0,25
2,8	3,18	-0,38
2,96	3,21	-0,25
2,77	3,13	-0,36
2,9	3,22	-0,32
2,82	3,16	-0,34
2,96	3,23	-0,27
2,8	3,14	-0,34
2,93	3,21	-0,28
3,08	3,29	-0,21
2,76	3,08	-0,32
2,84	3,11	-0,27
3,12	3,31	-0,19
3,15	3,31	-0,16
2,83	3,1	-0,27
2,86	3,15	-0,29
3,05	3,27	-0,22
2,87	3,18	-0,31
2,91	3,2	-0,29
2,75	3,2 3,14	-0,39
2,86	3,14	-0,28
3,07	3,14	-0,26
2,92	3,23 3,16	-0,10 -0,24
3	3,2	-0,24
2,84	3,19	
	3,19 3,21	-0,35 -0,3
2,91		
2,68	3,06	-0,38 0.35
3,01	3,26	-0,25 0,30
2,7	3,09	-0,39

The computed excitation energies depend on the bond length alternation (BLA) of formal carbon-carbon single and double bonds within the polyene chain of the retinal chromophore, both for SORCI and OM2/MRCI (Figure 6S). As expected, the larger the BLA the higher the excitation energy.

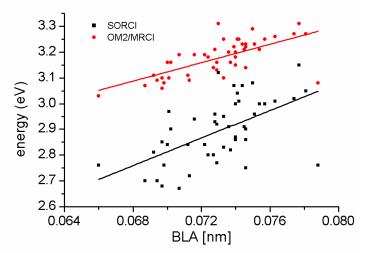


Figure 6S. Dependence of the excitation energy on the retinal bond length alternation (BLA).

Different QM regions were tested with the OM2/MRCI method, in order to establish the most convenient model to be used for the calculation of the OM2/MRCI absorption spectra (Figure 7S). As shown, realistic QM/MM results can be obtained already when only the retinal chromophore is included in the QM region; this was therefore selected as OM2/MRCI QM/MM setup.

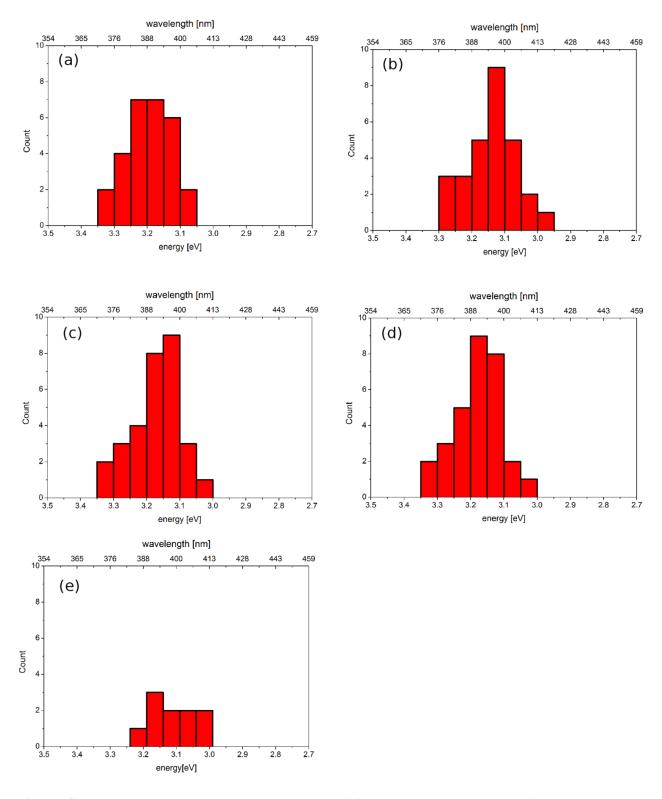


Figure 7S. OM2/MRCI absorption spectra calculated with different QM regions: (a) retinal, four water molecules, E123 and D253 side chains (*i.e.* same as SORCI); (b) only retinal; (c) retinal and D253 side chain; (d) retinal and E123 side chain; (e) retinal and water.

5. Cartesian Coordinates in Ångström of Representative BR and ChR2 Active Site Structures (L-QM region)

BR	H 26.811990 43.887170 34.569520
C 26.594540 29.322670 35.907770	H 28.482920 44.343460 34.634360
H 27.063950 30.238870 35.559420	C 28.082450 42.589410 33.423510
H 26.476490 28.680350 35.038670	H 27.749100 43.095540 32.512230
C 27.571830 28.596350 36.912000	H 29.155230 42.402940 33.308190
O 27.064690 28.037170 37.906610	C 27.329060 41.283640 33.592560
O 28.799540 28.655520 36.563400	H 27.407350 40.671490 32.689440
H 25.612640 29.521840 36.329720	H 26.266650 41.510390 33.728510
C 33.854970 30.951180 38.079040	C 27.835760 40.461490 34.809850
H 33.278240 30.108470 38.464560	C 26.746820 39.383300 35.062790
H 34.615920 30.551320 37.413490	H 26.541620 38.849350 34.137920
C 32.833830 31.795830 37.284960	H 25.820130 39.849840 35.392200
O 32.383650 32.892950 37.737940	H 27.031210 38.643080 35.802810
O 32.359450 31.220670 36.215380	C 29.183360 39.801100 34.415620
H 34.330790 31.522510 38.871780	H 29.075390 39.270610 33.471720
C 32.714810 28.653280 43.572300	H 29.527510 39.081790 35.152840
	H 29.962070 40.551050 34.289900
H 32.258810 27.738630 43.954260	O 30.813030 29.703120 37.960270
H 33.497230 28.367960 42.870390	H 31.329920 30.107460 37.232550
H 33.160930 29.233610 44.377270	H 30.086100 29.165810 37.565660
C 31.658430 29.473000 42.846540	O 31.684310 31.988830 33.543910
H 32.103370 30.390250 42.451690	
H 30.888670 29.790000 43.560780	H 30.845360 31.518240 33.713240
C 30.997700 28.710600 41.700480	H 32.147930 31.977490 34.401220
H 30.606480 27.751580 42.051310	O 29.817300 30.709430 34.987530
H 31.724980 28.493030 40.916350	H 29.440090 29.899050 35.378290
C 29.829700 29.522720 41.119920	H 30.483970 31.080730 35.589390
H 29.137090 29.780130 41.927750	ChR2, RSBH ⁺ ···E123
H 29.285070 28.940540 40.368170	N 55.21000 23.23000 31.10000
N 30.266980 30.751380 40.479510	H 55.02000 23.38000 32.06000
H 30.581800 30.639220 39.492940	C 55.23000 24.53000 30.27000
C 30.213790 31.942110 41.047460	H 54.34000 24.46000 29.67000
H 29.884030 31.968170 42.090720	C 55.45000 25.90000 30.95000
C 30.414370 33.139610 40.315670	H 54.96000 26.05000 31.90000
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H 30.152920 33.876960 42.883780	H 57.50000 27.08000 29.67000
C 29.961010 35.452630 39.832890	H 56.45000 26.62000 28.34000
H 29.999130 35.129420 38.792310	C 56.83000 28.87000 28.56000
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H 29.225490 37.080230 37.991490	H 58.17000 29.91000 27.40000
C 28.914650 38.941380 38.944520	H 59.00000 28.92000 28.52000
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	53.14000	36.63000	36.05000
	53.54000	37.10000	37.35000
	52.68000	37.10000	37.94000
	54.08000	36.43000	37.91000
	53.94000	38.04000	37.44000
	52.33000	37.44000	35.21000
	52.35000	37.44000	34.19000
	51.90000	38.70000	35.54000
	52.30000	39.11000	36.42000
	51.23000	39.11000	34.52000
	50.92000	38.83000	33.63000
1	50.78000	40.71000	34.64000
	51.01000	41.53000	35.91000
	50.15000	41.78000	36.54000
	51.69000	41.00000	36.63000

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		36.16000						
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N	31.14000	36.60000	37.96000	С		32.78000	23.70000	33.60000
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С	29.93000	36.18000	38.83000	С		33.83000	23.21000	32.72000
Н	29.82000	37.09000	39.39000	С		34.58000	22.10000	33.01000
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C	21.00000	32.23000	35.03000	Н		34.92000	25.91000	30.74000
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С	24.73000	34.87000	37.72000	Н		26.68000	33.21000	31.77000
Н	24.83000	35.67000	36.98000	0		25.77000	32.69000	29.08000
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С	25.04000	33.32000	35.71000	Н		25.68000	32.61000	27.50000
Н	23.97000	33.19000	35.49000				RSBH+ *···OH ₂	27.50000
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N	25.87000	32.22000	35.05000	N			22.66000	31.17000
				Н		55.62000	22.89000	32.13000
Н	25.66000	32.10000	34.08000	C		55.40000	23.83000	30.16000
С	26.69000	31.39000	35.60000	Н		54.54000	23.37000	29.70000
Н	26.79000	31.59000	36.69000	С		55.13000	25.24000	30.71000
C	27.39000	30.40000	34.86000	Н		54.30000	25.37000	31.36000
Н	27.44000	30.63000	33.82000	Н		55.91000	25.55000	31.49000
С	28.08000	29.36000	35.49000	С		55.11000	26.58000	29.88000
С	27.81000	28.96000	36.93000	Н		54.52000	26.59000	28.96000

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Н	55.94000	29.13000	29.09000	С	54.53000	34.35000	36.28000
N	57.79000	28.88000	28.13000	Н	54.43000	34.38000	37.35000
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Н	52.84000	29.88000	30.98000	С	51.44000	39.38000	34.81000
С	53.60000	28.84000	32.71000	Н	51.52000	39.01000	33.78000
Н	53.17000	28.31000	33.62000	C	50.91000	40.65000	34.99000
н	54.29000	28.14000	32.22000	c	50.93000	41.25000	36.33000
							36.73000
С	54.33000	30.09000	33.11000	H	49.93000	41.35000	
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С	50.82000	30.49000	33.36000	С	50.44000	41.32000	33.76000
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Ν	49.97000	29.66000	37.29000	С	49.97000	42.61000	33.75000
Н	50.12000	29.80000	36.31000	Н	50.04000	43.29000	34.59000
С	51.13000	30.06000	38.19000	С	49.34000	43.30000	32.57000
Н	51.11000	29.27000	38.93000	С	48.80000	44.50000	32.82000
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	52.79000	31.06000	35.67000	н	48.08000	46.07000	34.08000
Н							
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Н	52.58000	28.55000	36.15000	С	47.41000	44.36000	30.64000
С	50.66000	31.20000	39.22000	Н	46.59000	43.71000	31.03000
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Ν	59.92000	35.88000	34.35000	С	48.56000	43.51000	30.18000
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С	59.40000	34.42000	34.63000	Н	49.34000	44.07000	29.80000
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C	59.15000	33.46000	33.46000	c	50.51000	42.08000	30.69000
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С	58.24000	33.82000	32.19000	Н	50.41000	41.71000	29.71000
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Н	58.01000	30.19000	39.43000	H	55.76000	33.98000	30.22000
C	56.18000	31.06000	38.71000	Н	56.87000	35.12000	30.64000
H	56.15000	30.19000	38.00000	0	57.41000	30.60000	34.52000
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С	55.52000	32.29000	38.10000	Н	56.50000	30.84000	34.17000
Н	54.51000	32.38000	38.27000	0	58.16000	30.22000	31.77000

6. Metadynamics

The dihedral angle around the C_{β} - C_{γ} bond was used as the collective variable (i.e. the reaction coordinate). The parameters were set as follows: Gaussian height of 0.2 kcal/mol, Gaussian width of 0.2 rad, Gaussian depositing frequency of 2 ps (1000 time steps), bias factor of 4, and temperature of 300 K. The convergence of the metadynamics simulation was evaluated by comparing the free energy as a function of the collective variable at different times, as shown in Figure 8S. The similar profiles obtained after 85 ns indicates a good convergence of the simulation.

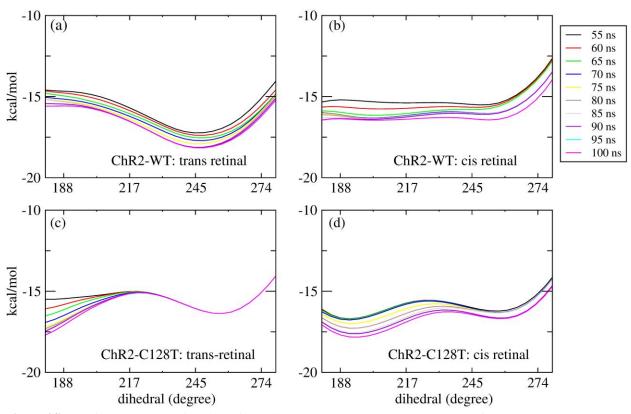


Figure 8S. The free energy as a function of the dihedral angle around the C_{β} - C_{γ} bond of E123 every 5 ns along the 100 ns well-tempered metadynamics simulation. (a) ChR2-WT bound with all-*trans* retinal; (b) ChR2-WT bound with 13-*cis*, 15-*syn* retinal; (c) ChR2-C128T bound with all-*trans* retinal; (d) ChR2-C128T bound with 13-*cis*, 15-*syn* retinal.

7. Gaussian Fit Parameters

The analytic fitted Gaussian function parameters used to convolute the OM2/MRCI histograms, shown in Figures 7–9 of the main text, are listed in Tables 4S.

Table 6S. Fitting parameters used to convolute the OM2/MRCI histograms into Gaussian functions of Figures 7–9 in the main text.

	Fit functi	ion: $y = -A0*exp(A2*($	A1-x)^2)	
	A0	A1	A2	Fitting coefficient: R ²
BR	-0.246117	2.62478	-24.7658	0.997614
ChR2-WT:	-0.178744	3.19532	-14.8734	0.998295
trans+cis retinals, total ChR2-WT:	-0.0781826	3.24628	-18.9074	0.996242
trans+cis retinals, RSBH-E123 ChR2-WT:	-0.0528687	3.14634	-13.1284	0.996214
trans+cis retinals, RSBH-D253 ChR2-WT: trans+cis retinals,	-0.154118	3.16907	-15.3466	0.995477
RSBH-H ₂ O ChR2-WT: trans+cis retinals,	-0.125309	3.24356	-18.7595	0.998637
E123-upward ChR2-WT: trans+cis retinals,	-0.0683909	3.10066	-15.5577	0.996431
E123-downward				
ChR2-WT: trans	-0.19488	3.20365	-16.9763	0.998440
retinal, total ChR2-WT: trans retinal, RSBH-E123	-0.0976391	3.23267	-18.5911	0.997837
ChR2-WT: trans retinal, RSBH-D253	-0.0142415	3.18572	-19.7828	0.972275
ChR2-WT: trans retinal, RSBH-H ₂ O	-0.0864315	3.17354	-16.3061	0.995963
ChR2-WT: trans retinal, E123- upward	-0.147067	3.23107	-18.8384	0.997145
ChR2-WT: trans retinal, E123- downward	-0.0551499	3.13108	-16.9933	0.994725
ChR2-WT: cis	-0.163051	3.1837	-12.7511	0.997197
retinal, total ChR2-WT: cis	-0.0976391	3.23267	-18.5911	0.997837
retinal, RSBH-E123 ChR2-WT: cis retinal, RSBH-D253	-0.0142415	3.18572	-19.7828	0.972275
ChR2-WT: cis retinal, RSBH-H ₂ O	-0.0864315	3.17354	-16.3061	0.995963
ChR2-WT: cis retinal, E123- upward	-0.147067	3.23107	-18.8384	0.997145

ChR2-WT: cis retinal, E123- downward	-0.0551499	3.13108	-16.9933	0.994725
ChR2-C128T:	-0.192937	3.05524	-15.2363	0.998059
trans+cis retinals,	0.172737	3.03324	13.2303	0.770037
total				
ChR2-C128T:	-0.00992061	3.19575	-20.4072	0.973085
trans+cis retinals,				
RSBH-E123				
ChR2-C128T:	-0.0457847	3.06655	-16.2952	0.994600
trans+cis retinals,				
RSBH-D253				
ChR2-C128T:	-0.140947	3.04233	-15.5087	0.999122
trans+cis retinals,				
RSBH-H ₂ O				
ChR2-C128T:	-0.0175266	3.17342	-18.2792	0.984749
trans+cis retinals,				
E123-upward	0.150510	2.04500	15 5011	0.007070
ChR2-C128T:	-0.179718	3.04509	-15.7911	0.997950
trans+cis retinals,				
E123-downward ChR2-C128T: trans	0.202265	3.08519	-15.7886	0.000102
retinal, total	-0.202365	3.08319	-13./880	0.998103
ChR2-C128T: trans				
retinal, RSBH-E123	-0.0196258	3.19547	-20.1408	0.970591
reuliai, KSDH-E125	***************************************			******
ChR2-C128T: trans	-0.038766	3.09045	-17.2507	0.995595
retinal, RSBH-D253	0.030700	3.07013	17.2307	0.773373
ChR2-C128T: trans	-0.149439	3.06946	-16.2166	0.995911
retinal, RSBH-H ₂ O	*******	21002		***************************************
ChR2-C128T: trans	-0.0316859	3.18153	-18.8221	0.984326
retinal, E123-				
upward				
ChR2-C128T: trans	-0.176413	3.06918	-16.4274	0.998190
retinal, E123-				
downward				
ChR2-C128T: cis	-0.189411	3.02266	-15.711	0.997310
retinal, total	0.00044			0.00.4400
ChR2-C128T: cis	-0.00044238	3.21333	-128.534	0.836400
retinal, RSBH-E123	0.0520260	2.04050	15 7042	0.005150
ChR2-C128T: cis	-0.0530368	3.04958	-15.7043	0.985150
retinal, RSBH-D253 ChR2-C128T: trans	0.12747	2.01212	-16.0913	0.006902
retinal, RSBH-H ₂ O	-0.13747	3.01213	-10.0913	0.996802
ChR2-C128T: trans	-0.0037326	3.09766	-15.5254	0.855565
retinal, E123-	-0.0037320	5.07700	-13.3434	0.033303
upward				
ChR2-C128T: trans	-0.186148	3.02121	-15.8074	0.997509
retinal, E123-				
downward				
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