

**Asymmetric [4+2] Annulation of 5*H*-Thiazol-4-Ones with a Chiral
Dipeptide-Based Brønsted Base Catalyst**

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Supporting Information

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1. General information

General procedures and methods

Experiments involving moisture and/or air sensitive components were performed under a positive pressure of nitrogen in oven-dried glassware equipped with a rubber septum inlet. Dried solvents and liquid reagents were transferred by oven-dried syringes or hypodermic syringe cooled to ambient temperature in a desiccator. Reactions mixtures were stirred in 4 mL sample vial with Teflon-coated magnetic stirring bars unless otherwise stated. Moisture in non-volatile reagents/compounds was removed in high *vacuo* by means of an oil pump and subsequent purging with nitrogen. Solvents were removed in *vacuo* under ~30 mmHg and heated with a water bath at 30–40 °C using rotary evaporator with aspirator. The condenser was cooled with running water at 0 °C.

All experiments were monitored by analytical thin layer chromatography (TLC). TLC was performed on pre-coated plates, 60 F₂₅₄. After elution, plate was visualized under UV illumination at 254 nm for UV active material. Further visualization was achieved by staining KMnO₄, ceric molybdate, or anisaldehyde solution. For those using the aqueous stains, the TLC plates were heated on a hot plate. Columns for flash chromatography (FC) contained silica gel 200–300 mesh. Columns were packed as slurry of silica gel in petroleum ether and equilibrated solution using the appropriate solvent system. The elution was assisted by applying pressure of about 2 atm with an air pump.

Instrumentations

Proton nuclear magnetic resonance (¹H NMR), carbon NMR (¹³C NMR), and fluorous (¹⁹F NMR) spectra were recorded in CDCl₃ otherwise stated. ¹H (300 MHz) and ¹³C (75 MHz) were performed on a 300MHz spectrometer. Chemical shifts are reported in parts per million (ppm), using the residual solvent signal as an internal standard: CDCl₃ (¹H NMR: δ 7.26, singlet; ¹³C NMR: δ 77.0, triplet). Multiplicities were given as: *s* (singlet), *d* (doublet), *t* (triplet), *q* (quartet), *quintet*, *m* (multiplets), *dd* (doublet of doublets), *dt* (doublet of triplets), and *br* (broad). Coupling constants (*J*) were recorded in Hertz (Hz). The number of proton atoms (*n*) for a given resonance was indicated by *nH*. The number of carbon atoms (*n*) for a given resonance was indicated by *nC*. HRMS was reported in units of mass of charge ratio (m/z). Mass samples were dissolved in DCM and MeOH (HPLC Grade) unless otherwise stated. Optical rotations were recorded on a polarimeter with a sodium lamp of wavelength 589 nm and reported as follows; [α]_λ^{T°C} (c = g/100 mL, solvent). Melting points were determined on a melting point apparatus.

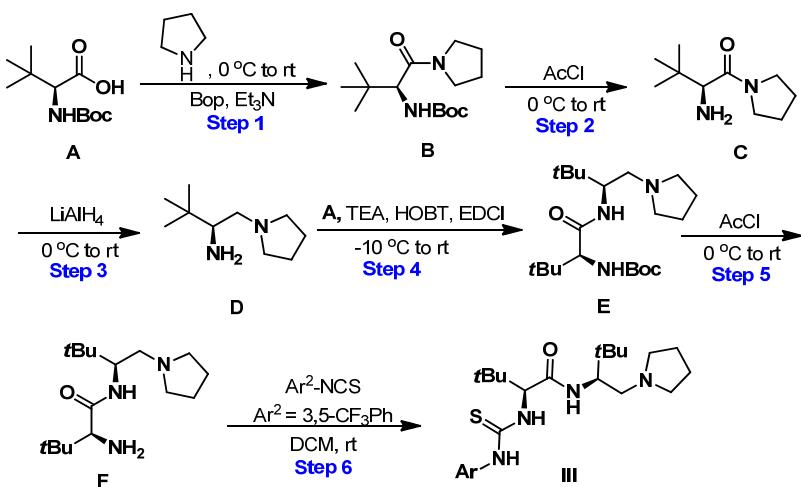
Enantiomeric excesses were determined by chiral High Performance Liquid Chromatography (HPLC) analysis. UV detection was monitored at 254 nm, 230 nm and 210 nm at the same time. HPLC samples were dissolved in HPLC grade isopropanol (IPA) unless otherwise stated.

Materials

All commercial reagents were purchased with the highest purity grade. They were used without further purification unless specified. All solvents used, mainly petroleum ether (PE) and ethyl acetate

(EtOAc), were distilled. Anhydrous CH₂Cl₂, CHCl₃ and DCE were freshly distilled from CaH₂ and stored under N₂ atmosphere. THF, Et₂O and toluene were freshly distilled from sodium/benzophenone before use. All compounds synthesized were stored in a -20 °C freezer and light-sensitive compounds were protected with aluminium foil.

2. Synthesis of dipeptide-based III



Step 1: To a solution of crude Boc-protected amino acid **A** (2.31 g, 10 mmol) in THF (15 mL), Et₃N (2.78 mL, 20 mmol) and BOP (5.41 g, 12 mmol) was added at 0 °C. Pyridine (1.16 mL, 13 mmol) was injected after the mixture was stirred for 0.5 hour at 0 °C. Then the reaction mixture was stirred at room temperature for another 8 hours (monitored by TLC). Solvent was removed under reduced pressure and **B** was obtained by flash chromatography (*silica gel*, Hexane/ethyl acetate 10:1–3:1), as a white solid, 2.32 g, 81.6% yield.

Step 2: AcCl (4.6 mL, 65.6 mmol) was added to a solution of **B** (2.32 g, 8.2 mmol) in MeOH (20 mL) 0 °C. The reaction mixture was stirred at room temperature for 8 hours (monitored by TLC). Solvent was removed under reduced pressure and solid **C** (1.51 g, 100%) was obtained for the next step without any purification.

Step 3: LiAlH₄ (1.25 g, 32.8 mmol) was added into a solution of **C** (1.51 g, 8.2 mmol) in THF (20 mL) slowly. The mixture was stirred at 0 °C for about 5.0 minutes then warmed to room temperature and stirred overnight. Subsequently, the reaction was carefully quenched by NaOH aqueous solution (4.0 N, 2 mL). The solids were filtered off and washed by THF, and the filtrate was dried over Na₂SO₄. Solvent was removed under reduced pressure and **D** was obtained as yellow oil in 86.5% yield (1.2 g).

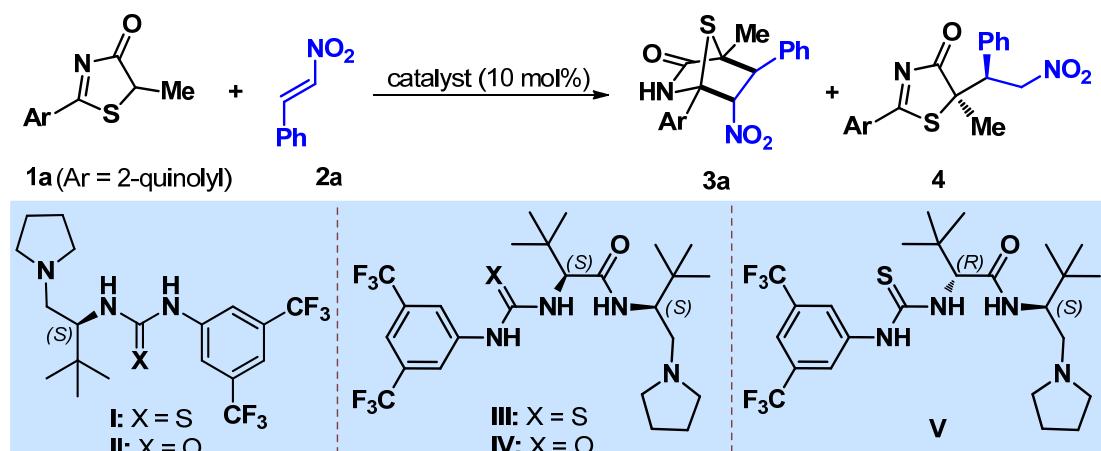
Step 4: To a solution of diamine **D** (0.75 g 4.4 mmol) in THF (25 mL) were added TEA (1.84 ml, 13.2 mmol), Boc-protected amino acid **A** (1.2 g, 5.3 mmol) and HOBT (0.89 g, 6.6 mmol) at –10 °C for 15 min. Then EDCI (1.3 g, 6.6 mmol) was added to the mixture, and the mixture was stirred overnight at room temperature. The reaction mixture was poured into water and extracted with AcOEt. The AcOEt layer was washed with water and brine and dried over Na₂SO₄. Filtration and concentration in vacuo and purification by *silica gel* flash column chromatography (AcOEt/n-hexane = 1/5) gave 1.28 g (76%) of **E** as a white solid.

Step 5 AcCl (1.89 mL, 26.7 mmol) was added to a solution of **E** (1.28 g, 3.34 mmol) in MeOH (20 mL) 0 °C. The reaction mixture was stirred at room temperature for 8 hours (monitored by TLC). Solvent was removed under reduced pressure and solid **F** (0.95 g, 100%) was obtained for the next step without any purification.

Step 6 (Final step): To a solution of **F** (308.6 mg, 1.09 mmol) in DCM (5 mL) was added 3,5-di(trifluoromethyl)phenyl isothiocyanate (295.2 mg, 1.09 mmol). The resulting solution was stirred for 5 min at 0 °C. The reaction was concentrated *in vacuo* and the residue was purified by *silica gel* chromatography (*silica gel*, DCM/MeOH 100:1–30:1) to afford product **III** as a white solid in 85% yield (513.1 mg).

3. Optimization of reaction conditions for nitroalkene **2a** as acceptor

Table 1 Asymmetric [4+2] cyclization catalyzed by amino acid-derived tertiary amine–(thio)ureas^a



Entry	Catalyst	Solvent	<i>T</i> (°C)	<i>t</i> (h)	3a : 4	3a			4		
						Yield (%) ^b	e.e. (%) ^c	d.r. ^d	Yield (%) ^b	e.e. (%) ^c	d.r. ^d
1	Et ₃ N	CH ₂ Cl ₂	25	18	trace	trace	n.a.	--	36	n.a.	n.a.
2	I	CH ₂ Cl ₂	25	12	1:1	42	0	15:1	45	64	1:1
3	II	CH ₂ Cl ₂	25	12	3:1	62	7	9:1	20	3	19:1
4	III	CH ₂ Cl ₂	25	12	4:1	73	86	19:1	13	88	12:1
5	IV	CH ₂ Cl ₂	25	12	4:1	68	87	8:1	18	87	9:1
6	III	THF	25	16	trace	trace	n.a.	n.a.	trace	n.a.	n.a.
7	III	toluene	25	16	5:1	84	94	>19:11	8	88	12:1
8	III	Et ₂ O	25	16	4:1	70	92	>19:1	11	96	>19:1
9	III	CHCl ₃	25	16	4.5:1	72	83	>19:1	9	85	>19:1
10	III	toluene	10	36	4:1	56	95	>19:1	10	31	3:1
11	III	toluene	30	18	4.5:1	78	92	19:1	12	75	19:1
12 ^e	III	toluene	25	18	5:1	83	95	>19:1	11	88	12:1
13 ^e	V	toluene	25	18	2.5:1	68	-75	>19:1	17	13	>19:1

^aThe reaction was performed using **1a** (0.05 mmol) and **2a** (0.075 mmol) in 1.0 mL of solvent.

^bIsolated yield based on **1a** after column chromatography.

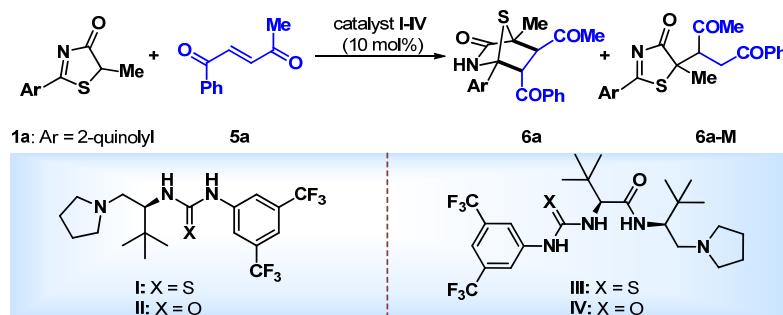
^cEnantiomeric ratio of product **3a** was determined via chiral phase HPLC analysis.

^dDetermined by crude ¹H NMR analysis.

^eThe reaction was performed using **1a** (0.2 mmol) and **2a** (0.4 mmol) in 4.0 mL of solvent.

4. Optimization of reaction conditions for 4-oxo-4-phenylbutenone **5a** as acceptor

Table 2 Condition optimization^a



Entry	Catalyst	Solvent	<i>T</i> (°C)	<i>t</i> (h)	6a		6a-M		
					Yield (%) ^b	e.e. (%) ^c	Yield (%) ^b	e.e. (%) ^c	d.r. ^d
1	Et ₃ N	CH ₂ Cl ₂	25	96	11	n.a.	72	n.a.	10:1
2	I	CH ₂ Cl ₂	25	3	27	62	57	62	>19:1
3	II	CH ₂ Cl ₂	25	3	44	54	46	37	>19:1
4	III	CH ₂ Cl ₂	25	2.5	85	87	12	32	>19:1
5	IV	CH ₂ Cl ₂	25	3	78	68	18	58	19:1
6	III	toluene	25	7	54	78	20	73	>19:1
7	III	Et ₂ O	25	7	13	71	5	72	>19:1
8	III	CHCl ₃	25	3.5	82	91	10	50	13:1
9	III	DCE	25	5	80	88	13	26/69	11:1
10	III	CHCl ₃	0	18	80	96	17	60/62	13:1
11	III	CHCl ₃	-10	24	86	98	9	69/53	19:1

^aReaction conditions: **1a** (0.05 mmol), **5a** (0.075 mmol), solvent (1.0 mL). For product **6a**, all *drs* are >20:1 determined by crude ¹H NMR.

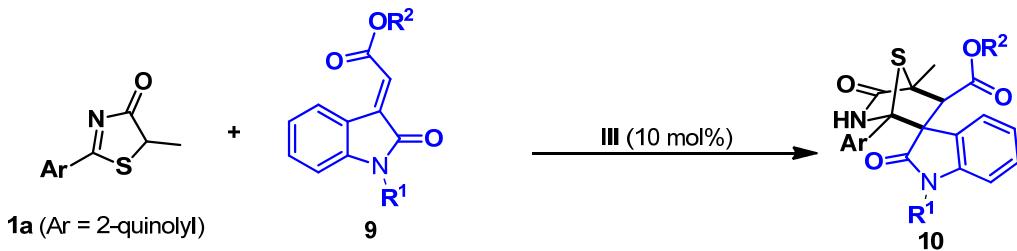
^bYield of isolated product.

^cEe was determined via chiral phase HPLC analysis.

^dDetermined by ¹H NMR analysis.

5. Optimization of reaction conditions for methyleneindolinones **9** as acceptor

Table 3 Condition optimization^a



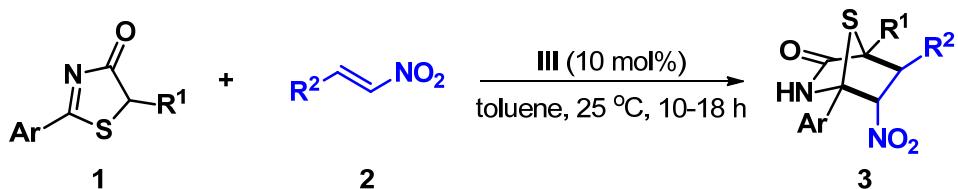
Entry	Solvent	R ¹	R ²	T (°C)	t (h)	10	
						Yield (%) ^b	ee (%) ^c
1	DCE	Me	Et	25	5	85	73
2	CHCl ₃	Me	Et	25	5	91	75
3	CHCl ₃	Me	Et	-10	10	86	85
4	CHCl ₃	Me	tBu	-10	12	84	80
5	CHCl ₃	Boc	Et	-10	12	trace	n.a.
6	CHCl ₃	Bn	Et	-10	12	90	94
7	CHCl ₃	Bn	Et	-30	24	96	95

^aReaction conditions: **1a** (0.05 mmol), **9** (0.075 mmol), solvent (1.0 mL). For product **10**, all *drs* are >20:1 determined by crude ¹H NMR.

^bYield of isolated product.

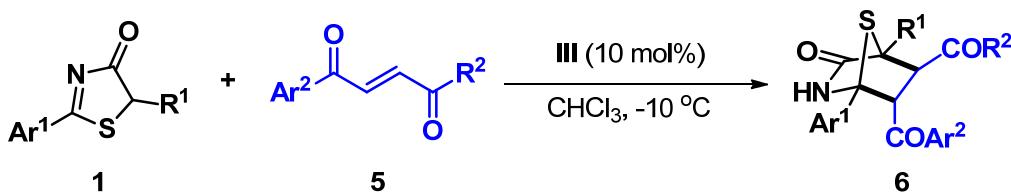
^cEe was determined via chiral phase HPLC analysis.

6. General experimental procedure for the [4+2] annulation of 5*H*-thiazol-4-ones **1 with nitroalkenes **2****



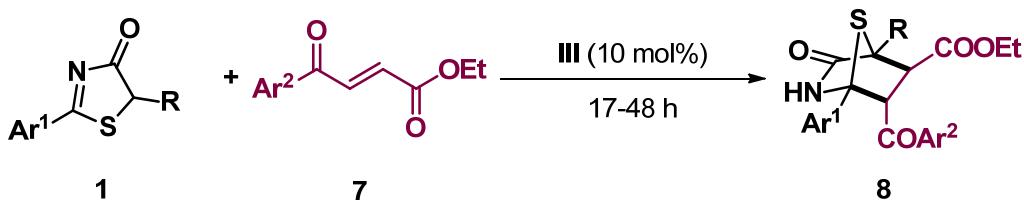
Nitroalkene **2** (0.4 mmol, 2.0 equiv) and catalyst **III** (11.5 mg, 0.02 mmol, 0.1 equiv) were dissolved in toluene (4.0 mL) and stirred at 25 °C for 15 min. Then 5*H*-thiazol-4-one **1** (0.2 mmol, 1.0 equiv) was added. The reaction mixture was stirred at 25 °C and monitored by TLC. Upon complete consumption of 5*H*-thiazol-4-one **1**, the reaction mixture was directly loaded onto a short *silica gel* column, followed by gradient elution with CH₂Cl₂/MeOH (800/1–300/1). Removing the solvent *in vacuo*, afforded products **3a-n**.

7. General experimental procedure for [4+2] annulation of 5*H*-thiazol-4-ones **1 with 4-oxo-4-arylbutenones **5****



4-Oxo-4-arylbutenone **5** (0.3 mmol, 1.5 equiv) and catalyst **III** (11.5 mg, 0.02 mmol, 0.1 equiv) were dissolved in CHCl₃ (4.0 mL) and stirred at -10 °C for 15 min. Then 5*H*-thiazol-4-one **1** (0.2 mmol, 1.0 equiv) was added. The reaction mixture was stirred at -10 °C and monitored by TLC. Upon complete consumption of 5*H*-thiazol-4-one **1**, the reaction mixture was directly loaded onto a short *silica gel* column, followed by gradient elution with CH₂Cl₂/MeOH (800/1–200/1). Removing the solvent *in vacuo*, afforded products **6a-s**.

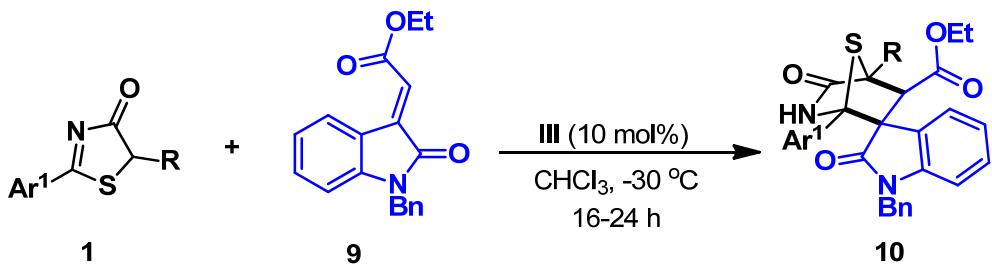
8. General experimental procedure for [4+2] annulation of 5*H*-thiazol-4-ones **1 with 4-oxo-4-arylbutenoates **7****



(R = Me): 4-Oxo-4-arylbutenoate **7** (0.3 mmol, 1.5 equiv) and catalyst **III** (11.5 mg, 0.02 mmol, 0.1 equiv) were dissolved in CHCl₃ (4.0 mL) and stirred at -30 °C for 15 min. Then 5*H*-thiazol-4-one **1** (0.2 mmol, 1.0 equiv) was added. The reaction mixture was stirred at -30 °C and monitored by TLC. Upon complete consumption of 5*H*-thiazol-4-one **1**, the reaction mixture was directly loaded onto a short *silica gel* column, followed by gradient elution with CH₂Cl₂/MeOH (800/1–200/1). Removing the solvent *in vacuo*, afforded products **8a-f** and **8h**.

(R = Ph): 4-Oxo-4-arylbutenoate **7** (0.3 mmol, 1.5 equiv), Li₃PO₄ (0.2 mmol, 1.0 equiv) and catalyst **III** (22.2 mg, 0.04 mmol, 0.2 equiv) were dissolved in CHCl₃ (4.0 mL) and stirred at 30 °C for 15 min. Then 5*H*-thiazol-4-one **1** (0.2 mmol, 1.0 equiv) was added. The reaction mixture was stirred at 30 °C and monitored by TLC. Upon complete consumption of 5*H*-thiazol-4-one **1**, the reaction mixture was directly loaded onto a short *silica gel* column, followed by gradient elution with CH₂Cl₂/MeOH (800/1–200/1). Removing the solvent *in vacuo*, afforded products **8g**.

9. General experimental procedure for [4+2] annulation of 5*H*-thiazol-4-ones **1 with methyleneindolinones **9****



Methyleneindolinones **9** (0.3 mmol, 1.5 equiv) and catalyst **III** (11.5 mg, 0.02 mmol, 0.1 equiv) were dissolved in CHCl₃ (4.0 mL) and stirred at -30 °C for 15 min. Then 5*H*-thiazol-4-one **1** (0.2 mmol, 1.0 equiv) was added. The reaction mixture was stirred at -30 °C and monitored by TLC. Upon complete consumption of 5*H*-thiazol-4-one **1**, the reaction mixture was directly loaded onto a short *silica gel* column, followed by gradient elution with CH₂Cl₂/MeOH (800/1–200/1). Removing the solvent *in vacuo*, afforded products **10a–n**.

10. Chemoselective results

Table 4 Reactions between **1** and **2** to afford **3** and **4**^a

Entry	Ar	R ¹	R ²	t (h)	3a	4
					Yield (%) ^b	Yield (%) ^b
1	2-quinolyl	Me	Ph	15	83	11
2	2-quinolyl	Me	4-FPh	12	78	16
3	2-quinolyl	Me	4-ClPh	14	89	6
4	2-quinolyl	Me	4-BrPh	13	84	8
5	2-quinolyl	Me	3-ClPh	15	85	9
6	2-quinolyl	Me	2-ClPh	18	82	12
7	2-quinolyl	Me	4-MePh	10	72	20
8	2-quinolyl	Me	3-MePh	13	76	18
9	2-quinolyl	Me	2-MePh	13	82	16
10	2-quinolyl	Me	2-naphthyl	16	88	8
11	2-quinolyl	Me	2-furyl	16	76	18
12	2-quinolyl	Me	cyclohexyl	72	41	51
13	2-quinolyl	Et	Ph	14	55	35
14	4-BrPh	Me	Ph	14	45	52
15	3-pyridyl	Me	Ph	10	76	19

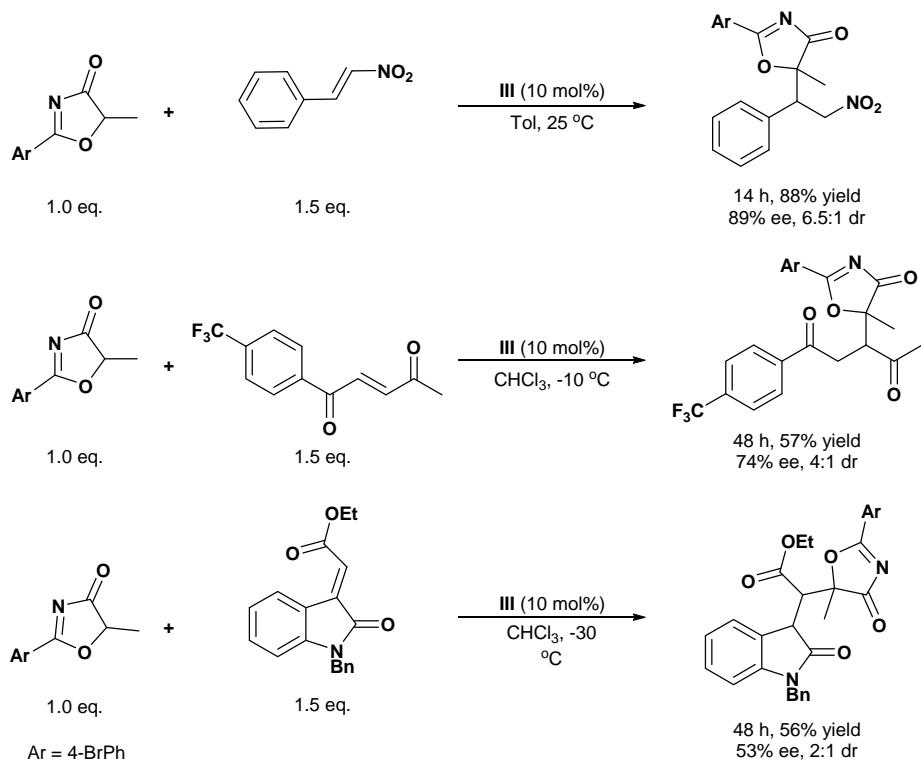
^a0.2 mmol scale. ^b Isolated yields.

Table 5 Reactions between **1** and **5** to afford **6** and **6-M**^a

entry	Ar ¹	R ¹	Ar ²	R ²	t (h)		
						6 yield (%) ^b	6-M yield (%) ^b
1	2-quinolyl	Me	Ph	Me	24	86	8
2	2-quinolyl	Me	4-CF ₃ Ph	Me	10	90	trace
3	2-quinolyl	Me	4-ClPh	Me	8	80	11
4	2-quinolyl	Me	4-BrPh	Me	24	91	trace
5	2-quinolyl	Me	3,4-Cl ₂ Ph	Me	16	95	trace
6	2-quinolyl	Me	4-MeOPh	Me	48	87	6
7	2-quinolyl	Me	3-MeOPh	Me	24	82	9
8	2-quinolyl	Me	2-MeOPh	Me	66	64	25
9	2-quinolyl	Me	2-BrPh	Me	40	60	28
10	2-quinolyl	Me	2-thienyl	Me	12	98	0
11	2-quinolyl	Me	2-furyl	Me	18	92	trace
12	2-quinolyl	Me	Ph	Et	36	90	5
13	2-pyridyl	Et	Ph	Me	24	87	7
14	3-pyridyl	Me	Ph	Me	24	86	10
15	2-thienyl	Me	Ph	Me	36	70	15
16	2-furyl	Me	Ph	Me	16	75	13
17	4-BrPh	Me	Ph	Me	24	54	42
18	2-quinolyl	Et	Ph	Me	18	72	10
19	2-quinolyl	Bn	Ph	Me	18	90	trace

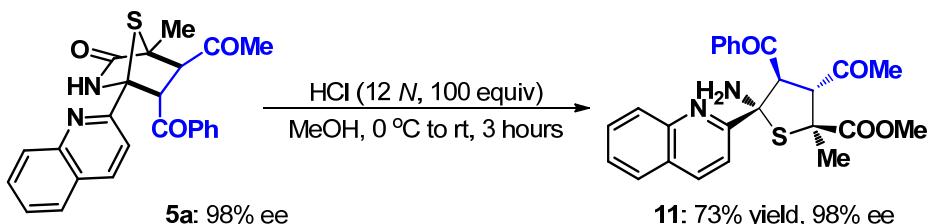
^a0.2 mmol scale. ^b Isolated yields.

11. Exploration towards reactions between 5*H*-oxazol-4-ones and alkenes



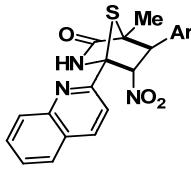
The results indicated that the reactions of 5*H*-oxazol-4-one with a series of activated alkenes under the established reaction conditions only presented conjugate addition adducts, and the corresponding annulation adducts were not observed yet. The moderate yields for 4-oxo-4-arylbutenone and methyleneindolinone as the substrates are produced by the instability of 5*H*-oxazol-4-one in the reactions.

12. Experimental procedure for **5a** to **11**

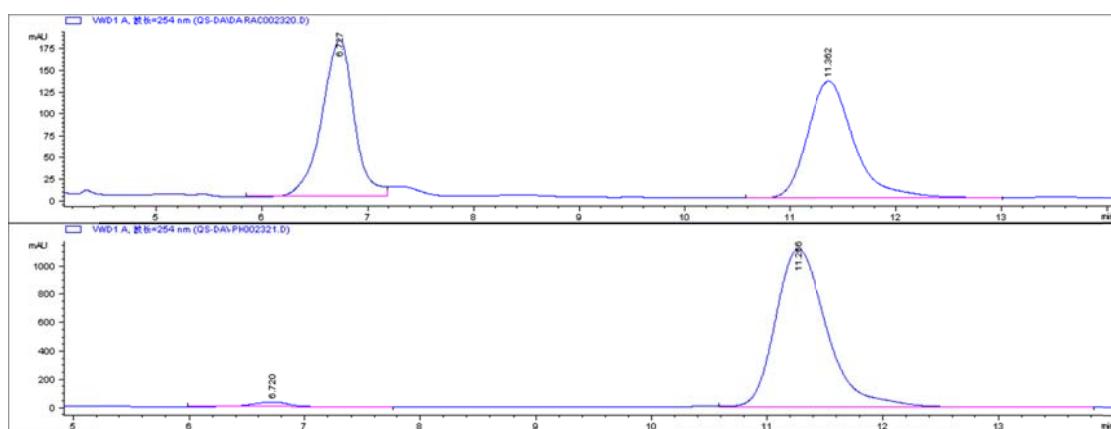


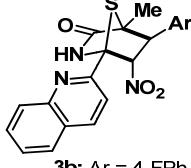
The [4+2] annulation adduct **5a** (41.6 mg, 0.1 mmol, 1.0 equiv) was dissolved in MeOH (1.0 mL) and stirred at 0 °C. Then HCl (12 N, 0.834 mL, 10.0 mmol, 100.0 equiv) was added. The reaction mixture was stirred at room temperature until the reaction completed (3.0 hours). Evaporation of methanol followed by drying under high vacuo and neutralized by NaHCO₃ (aq.) until pH > 7. The mixture was then extracted with EtOAc (3.0 mL x 3), washed with brine, and dried over Na₂SO₄. Concentration was loaded onto a short *silica gel* column, followed by gradient elution with PE/EA (20/1–2/1). Removing the solvent *in vacuo*, afforded products gave the **11** (32.7 mg, 73%).

13. Characterization of adducts

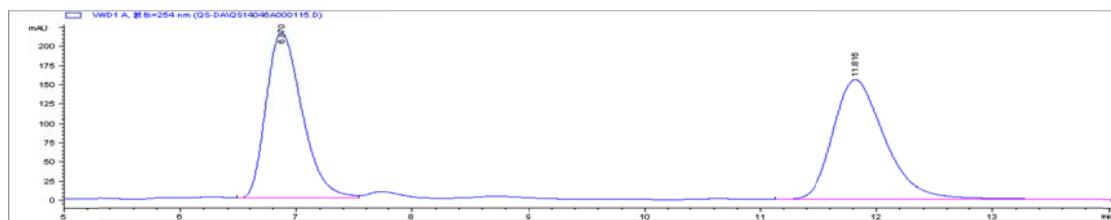
3a: Ar = Ph

 White solid, Mp 165.3–166.5 °C; 83% yield; 95% ee; $[\alpha]_D^{26} -143.0$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.34 (d, *J* = 8.5 Hz, 1H), 8.07 (d, *J* = 8.5 Hz, 1H), 7.90–7.87 (m, 2H), 7.81–7.73 (m, 2H), 7.68–7.60 (m, 3H), 7.50–7.44 (m, 3H), 6.27 (d, *J* = 4.8 Hz, 1H), 4.04 (d, *J* = 4.8 Hz, 1H), 1.29 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.8, 150.1, 147.0, 138.3, 136.0, 130.6, 129.6, 129.1, 129.0, 128.6, 128.0, 127.9, 127.6, 117.4, 98.6, 76.7, 67.1, 54.8, 13.8; HRMS (ESI) m/z 392.1074 (M+H⁺), calc. for C₂₁H₁₈N₃O₃S 392.1069.

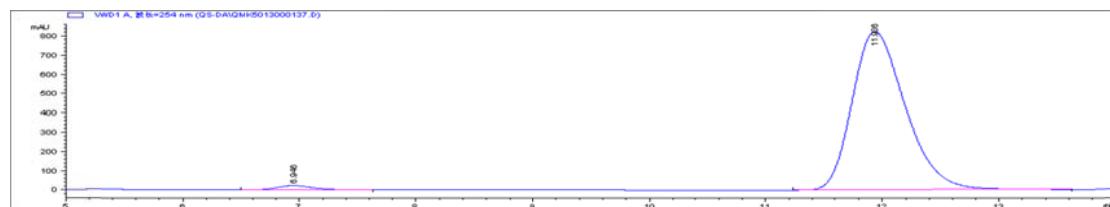
The ee was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 6.7 min (minor) and 11.3 min (major).



3b: Ar = 4-FPh

 White solid, Mp 163.0–165.0 °C; 78% yield; 97% ee; $[\alpha]_D^{26} -296.6$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.34 (d, *J* = 8.4 Hz, 1H), 8.07 (d, *J* = 8.4 Hz, 1H), 7.90–7.87 (m, 2H), 7.81–7.72 (m, 2H), 7.68–7.61 (m, 3H), 7.16 (t, *J* = 8.6 Hz, 2H), 6.23 (d, *J* = 4.8 Hz, 1H), 4.01 (d, *J* = 4.8 Hz, 1H), 1.28 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.7, 150.0, 147.1, 138.3, 131.9, 130.6, 130.3, 130.2, 129.6, 128.0, 127.7, 117.4, 116.2, 115.9, 98.6, 76.7, 66.9, 54.3, 13.8; HRMS (ESI) m/z 410.0972 (M+H⁺), calc. for C₂₁H₁₇FN₃O₃S 410.0975.

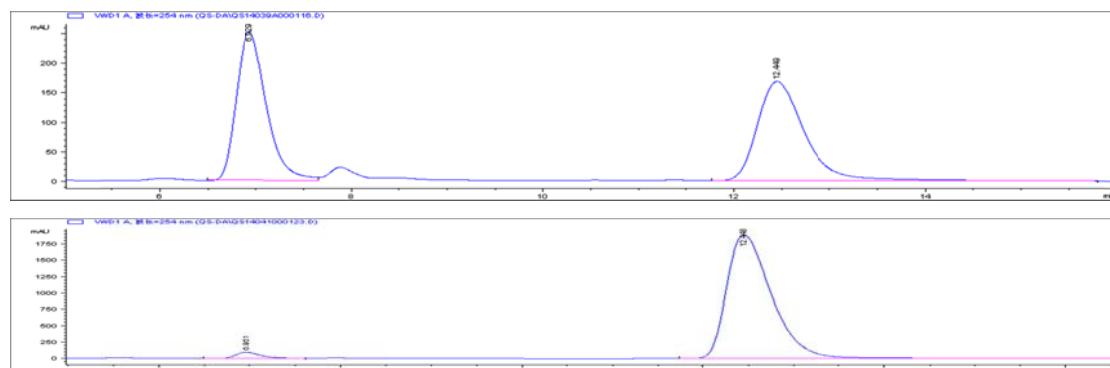
The ee was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 6.9 min (minor) and 11.9 min (major).





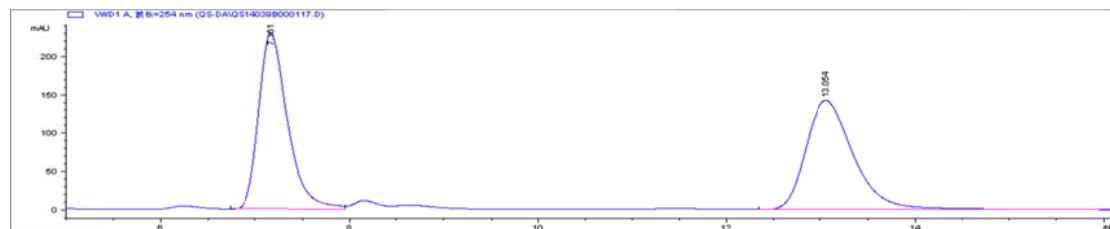
3c: Ar = 4-ClPh
 White solid, Mp 97.5–98.6 °C; 89% yield; 94% *ee*; $[\alpha]_D^{18} -298.7$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.34 (d, *J* = 8.5 Hz, 1H), 8.07 (d, *J* = 8.5 Hz, 1H), 7.90–7.85 (m, 2H), 7.81–7.72 (m, 2H), 7.66–7.61 (m, 3H), 7.44 (d, *J* = 8.5 Hz, 2H), 7.26 (s, 2H), 6.22 (d, *J* = 4.8 Hz, 1H), 4.00 (d, *J* = 4.8 Hz, 1H), 1.29 (s, 4H); ¹³C NMR (75 MHz, CDCl₃) δ 176.7, 150.0, 147.1, 138.3, 135.0, 134.6, 130.6, 129.8, 129.6, 129.3, 128.0, 127.9, 127.7, 117.4, 98.3, 76.8, 66.8, 54.4, 13.8; HRMS (ESI) m/z 426.0683 (M+H⁺), calc. for C₂₁H₁₇ClN₃O₃S 426.0679.

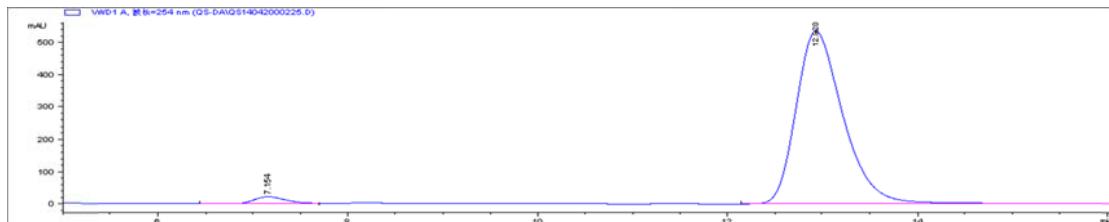
The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 7.0 min (minor) and 12.4 min (major).



3d: Ar = 4-BrPh
 White solid, Mp 95.4–97.1 °C; 84% yield; 96% *ee*; $[\alpha]_D^{21} -254.6$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.34 (d, *J* = 8.4 Hz, 1H), 8.07 (d, *J* = 8.4 Hz, 1H), 7.88 (d, *J* = 7.1 Hz, 2H), 7.81–7.72 (m, 2H), 7.66–7.54 (m, 5H), 6.22 (d, *J* = 4.8 Hz, 1H), 3.98 (d, *J* = 4.8 Hz, 1H), 1.29 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.8, 150.0, 147.1, 138.3, 135.1, 132.2, 130.6, 130.2, 129.6, 128.0, 127.9, 127.6, 123.1, 117.4, 98.1, 76.8, 66.7, 54.5, 13.8; HRMS (ESI) m/z 470.0175 (M+H⁺), calc. for C₂₁H₁₇BrN₃O₃S 470.0174.

The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 7.2 min (minor) and 12.9 min (major).

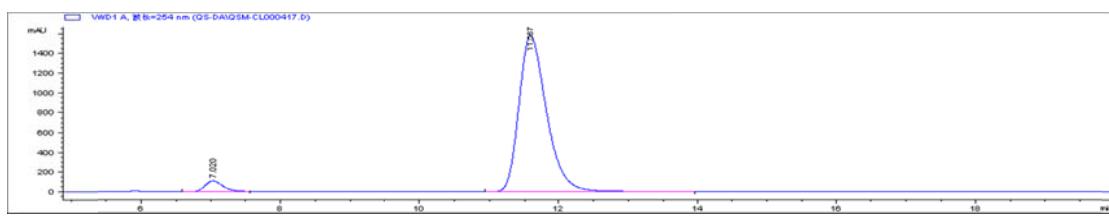
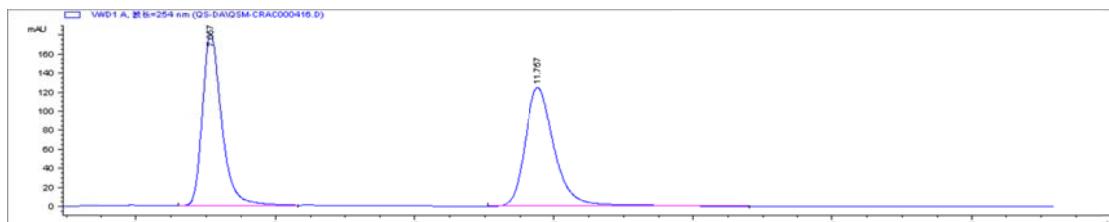




3e: Ar = 3-ClPh

White solid, Mp 163.2–165.8 °C; 85% yield; 92% ee; $[\alpha]_D^{26} -464.2$ (c 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.34 (d, J = 8.4 Hz, 1H), 8.08 (d, J = 8.4 Hz, 1H), 7.89 (d, J = 7.0 Hz, 2H), 7.81–7.73 (m, 3H), 7.66–7.61 (m, 1H), 7.53–7.51 (m, 1H), 7.45–7.37 (m, 2H), 6.23 (d, J = 4.8 Hz, 1H), 4.00 (d, J = 4.8 Hz, 1H), 1.31 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.5, 150.0, 147.1, 138.3, 138.2, 135.0, 130.6, 130.3, 129.6, 129.2, 128.2, 128.0, 127.9, 127.7, 127.2, 117.4, 98.3, 76.8, 66.8, 54.6, 13.8; HRMS (ESI) m/z 426.0684 (M+H⁺), calc. for C₂₁H₁₇ClN₃O₃S 426.0679.

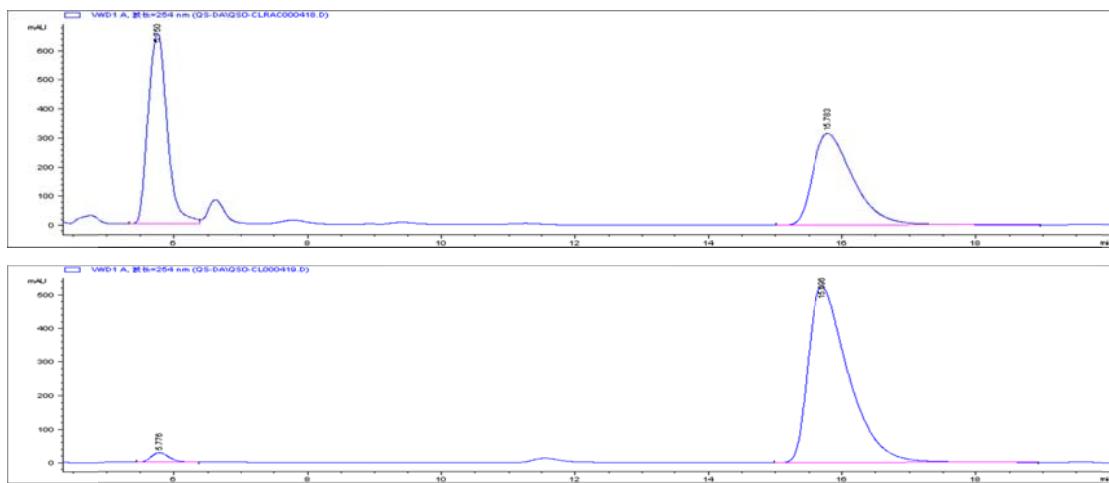
The ee was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 7.0 min (minor) and 11.6 min (major).



3f: Ar = 2-ClPh

White solid, Mp 184.0–185.6 °C; 82% yield; 95% ee; $[\alpha]_D^{26} -306.2$ (c 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.34 (d, J = 8.2 Hz, 1H), 8.10 (dd, J = 16.6, 8.2 Hz, 2H), 7.88 (d, J = 8.2 Hz, 1H), 7.77 (dd, J = 16.6, 7.6 Hz, 3H), 7.68–7.59 (m, 1H), 7.52 (d, J = 7.6 Hz, 1H), 7.41 (dt, J = 16.6, 7.6 Hz, 2H), 6.28 (d, J = 4.8 Hz, 1H), 4.89 (d, J = 4.8 Hz, 1H), 1.35 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.3, 150.1, 147.2, 138.3, 135.9, 133.9, 130.7, 130.2, 130.0, 129.7, 128.1, 128.0, 127.8, 127.7, 127.6, 117.4, 77.0, 98.1, 67.5, 49.7, 12.9; HRMS (ESI) m/z 426.0675 (M+H⁺), calc. for C₂₁H₁₇ClN₃O₃S 426.0679.

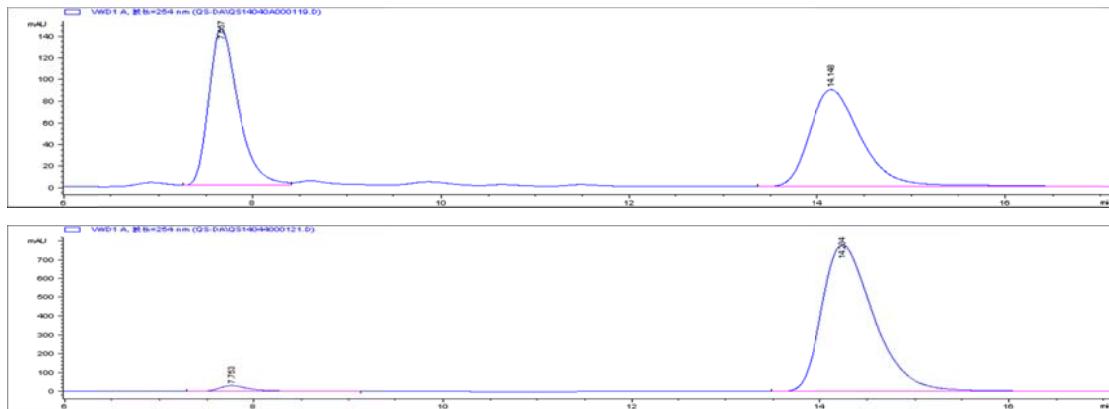
The ee was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 5.8 min (minor) and 15.7 min (major).



3g: Ar = 4-MePh

White solid, Mp 160.7–161.9 °C; 72% yield; 95% ee; $[\alpha]_D^{26} -120.4$ (c 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.33 (d, J = 8.4 Hz, 1H), 8.06 (d, J = 8.4 Hz, 1H), 7.88 (d, J = 8.4 Hz, 1H), 7.89–7.77 (m, 2H), 7.74 (d, J = 8.4 Hz, 1H), 7.65–7.60 (m, 1H), 7.54 (d, J = 8.1 Hz, 2H), 7.27 (s, 1H), 7.25 (s, 1H), 6.24 (d, J = 4.8 Hz, 1H), 4.00 (d, J = 4.8 Hz, 1H), 2.40 (s, 3H), 1.29 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.8, 150.2, 147.1, 138.8, 138.2, 133.0, 130.6, 129.7, 129.6, 128.4, 128.0, 127.9, 127.7, 117.4, 98.8, 76.7, 67.1, 54.6, 21.2, 13.8; HRMS (ESI) m/z 406.1222 (M+H⁺), calc. for C₂₂H₂₀N₃O₃S 406.1225.

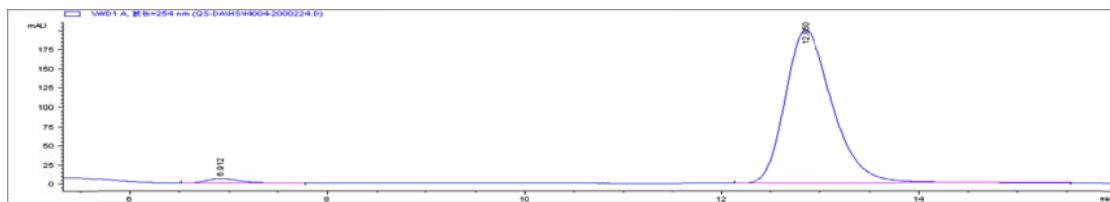
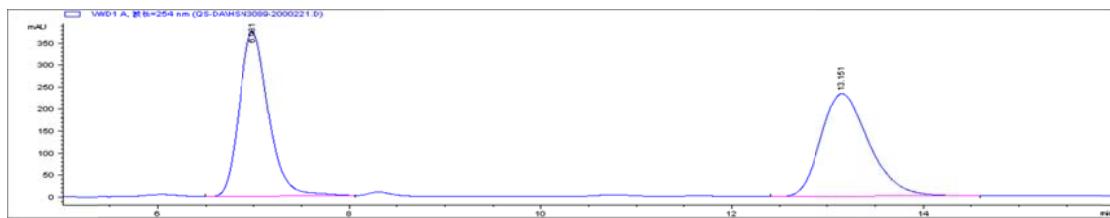
The ee was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 7.8 min (minor) and 14.2 min (major).



3h: Ar = 3-MePh

White solid, Mp 169.6–170.8 °C; 76% yield; 96% ee; $[\alpha]_D^{26} -218.1$ (c 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.34 (d, J = 8.5 Hz, 1H), 8.07 (d, J = 8.5 Hz, 1H), 7.88 (d, J = 8.2 Hz, 1H), 7.77 (dd, J = 16.3, 7.8 Hz, 3H), 7.63 (t, J = 7.5 Hz, 1H), 7.48 (d, J = 7.8 Hz, 1H), 7.43 (s, 1H), 7.34 (t, J = 7.5 Hz, 1H), 7.23 (s, 1H), 6.24 (d, J = 4.8 Hz, 1H), 3.99 (d, J = 4.8 Hz, 1H), 2.43 (s, 3H), 1.30 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.8, 150.2, 147.1, 138.8, 138.2, 135.9, 130.6, 129.7, 129.6, 129.4, 129.0, 128.0, 127.9, 127.6, 125.5, 117.5, 98.8, 76.7, 67.1, 54.9, 21.6, 13.8; HRMS (ESI) m/z 406.1223 (M+H⁺), calc. for C₂₂H₂₀N₃O₃S 406.1225.

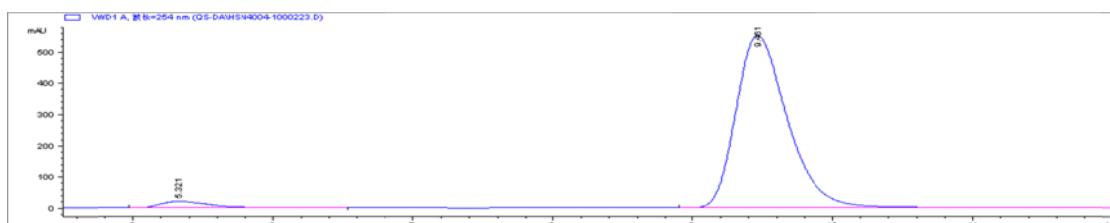
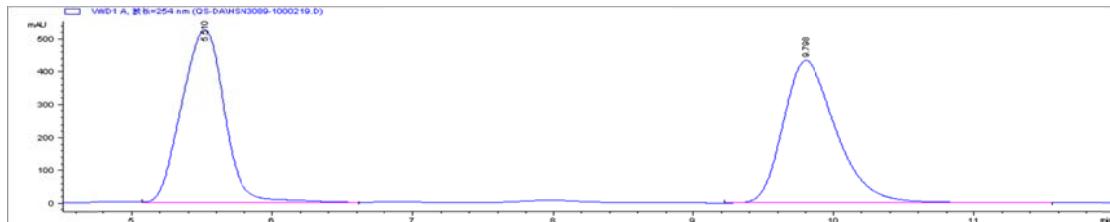
The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 6.9 min (minor) and 12.9 min (major).



3i: Ar = 2-MePh

White solid, Mp 182.2–183.5 °C; 82% yield; 93% *ee*; $[\alpha]_D^{26} -34.6$ (*c* 1.00, CHCl₃);
¹H NMR (300 MHz, CDCl₃) δ 8.33 (d, *J* = 8.3 Hz, 1H), 8.11–8.06 (m, 2H), 7.96 (s, 1H), 7.88 (d, *J* = 8.3 Hz, 1H), 7.80–7.73 (m, 2H), 7.65–7.60 (m, 1H), 7.38–7.29 (m, 3H), 6.24 (d, *J* = 4.9 Hz, 1H), 4.49 (d, *J* = 4.9 Hz, 1H), 2.40 (s, 3H), 1.29 (s, 3H);
¹³C NMR (75 MHz, CDCl₃) δ 176.9, 150.2, 147.1, 138.2, 137.8, 134.6, 131.0, 130.6, 129.6, 128.5, 128.0, 127.8, 127.6, 126.9, 125.9, 117.4, 99.6, 76.8, 67.4, 48.9, 20.4, 13.0; HRMS (ESI) m/z 406.1229 (M+H⁺), calc. for C₂₂H₂₀N₃O₃S 406.1225.

The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 5.3 min (minor) and 9.5 min (major).

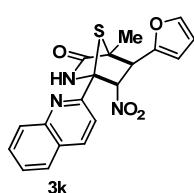
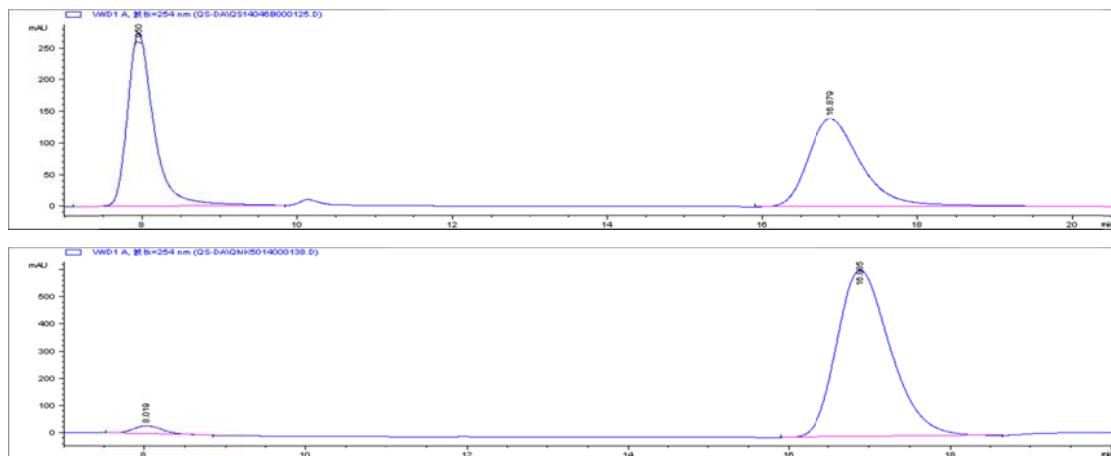


3j: Ar = 2-naphthyl

White solid, Mp 181.4–183.0 °C; 88% yield; 95% *ee*; $[\alpha]_D^{26} -384.7$ (*c* 1.00, CHCl₃);
¹H NMR (300 MHz, CDCl₃) δ 8.36 (d, *J* = 9.1 Hz, 1H), 8.09 (d, *J* = 9.7 Hz, 2H), 7.97–7.85 (m, 6H), 7.79 (t, *J* = 7.6 Hz, 2H), 7.63 (t, *J* = 7.6 Hz, 1H), 7.61–7.54 (m, 2H), 6.40 (d, *J* = 4.7 Hz, 1H), 4.20 (d, *J* = 4.7 Hz, 1H), 1.32 (s, 3H);
¹³C NMR (75 MHz, CDCl₃) δ 176.8, 150.2, 147.1, 138.3, 133.4, 133.3, 130.6, 129.6, 129.0, 128.4,

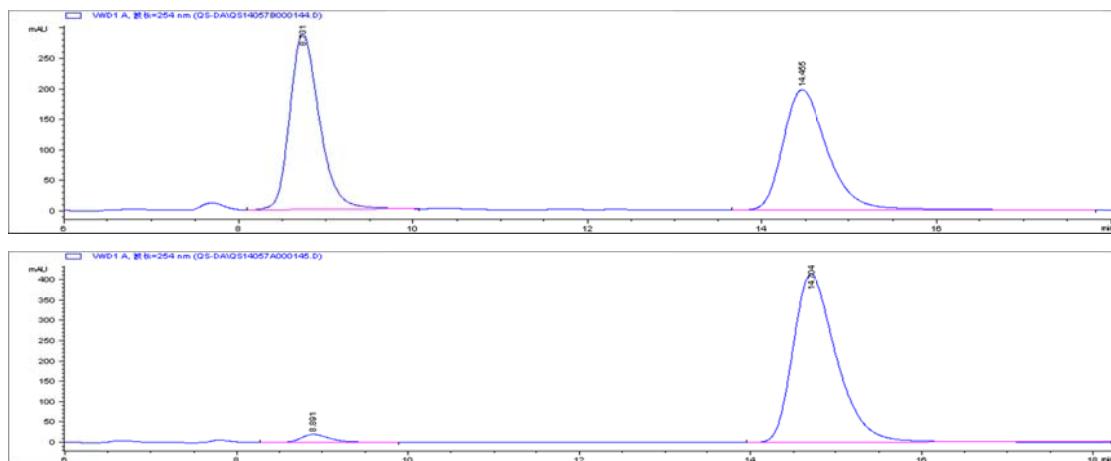
128.1, 127.9, 127.7, 126.7, 125.5, 117.5, 98.5, 76.8, 67.1, 55.2, 13.9; HRMS (ESI) m/z 442.1231 ($M+H^+$), calc. for $C_{25}H_{20}N_3O_3S$ 442.1225.

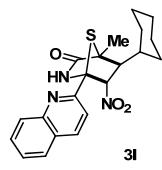
The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 8.0 min (minor) and 16.9 min (major).



White solid, Mp 156.2–157.9 °C; 76% yield; 94% *ee*; $[\alpha]_D^{26} -251.7$ (*c* 1.00, $CHCl_3$); 1H NMR (300 MHz, $CDCl_3$) δ 8.33 (d, *J* = 8.4 Hz, 1H), 8.07 (d, *J* = 8.4 Hz, 1H), 7.88 (d, *J* = 8.4 Hz, 1H), 7.83 (s, 1H), 7.78 (t, *J* = 7.6 Hz, 1H), 7.72 (d, *J* = 8.4 Hz, 1H), 7.63 (t, *J* = 7.6 Hz, 1H), 7.53 (d, *J* = 2.1 Hz, 1H), 6.56 (d, *J* = 2.1 Hz, 1H), 6.51 – 6.44 (m, 1H), 6.27 (d, *J* = 4.6 Hz, 1H), 4.27 (d, *J* = 4.6 Hz, 1H), 1.41 (s, 3H); ^{13}C NMR (75 MHz, $CDCl_3$) δ 176.1, 149.9, 149.2, 147.0, 143.4, 138.3, 130.6, 129.6, 128.0, 127.9, 127.6, 117.3, 110.8, 109.9, 96.8, 76.8, 66.4, 49.3, 13.4; HRMS (ESI) m/z 382.0864 ($M+H^+$), calc. for $C_{19}H_{16}N_3O_4S$ 382.0862.

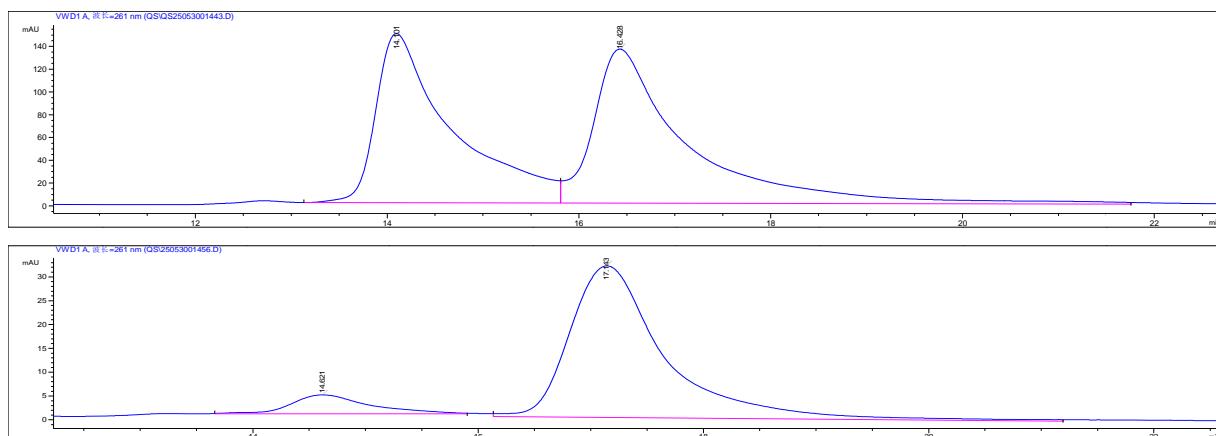
The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 8.9 min (minor) and 14.7 min (major).





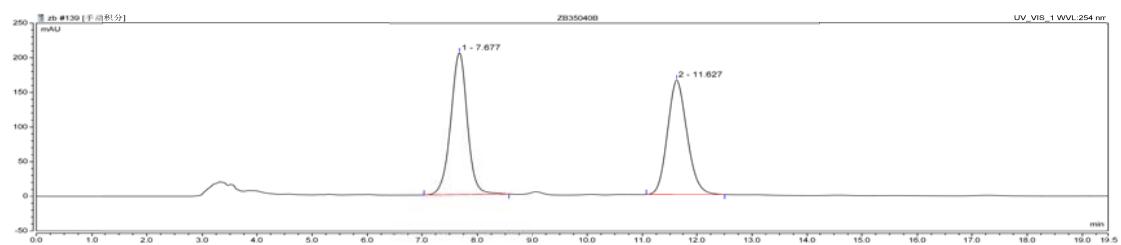
White solid, Mp 94.0–95.6 °C; 41% yield; 82% *ee*; $[\alpha]_D^{26} -136.7$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.30 (d, *J* = 8.6 Hz, 1H), 8.05 (d, *J* = 8.6 Hz, 1H), 7.86 (d, *J* = 7.6 Hz, 1H), 7.77 (d, *J* = 7.6 Hz, 2H), 7.61 (d, *J* = 7.6 Hz, 2H), 5.85 (d, *J* = 4.3 Hz, 1H), 2.97 (s, 1H), 2.09–1.78 (m, 5H), 1.70 (s, 3H), 1.37–1.19 (m, 6H); ¹³C NMR (75 MHz, CDCl₃) δ 177.1, 149.9, 147.0, 138.2, 130.6, 129.6, 128.0, 127.8, 127.6, 117.3, 95.0, 64.5, 52.9, 37.8, 32.0, 28.5, 26.5, 26.2, 26.0, 13.3; HRMS (ESI) m/z 398.1536 (M+H⁺), calc. for C₂₁H₂₄N₃O₃S 398.1538.

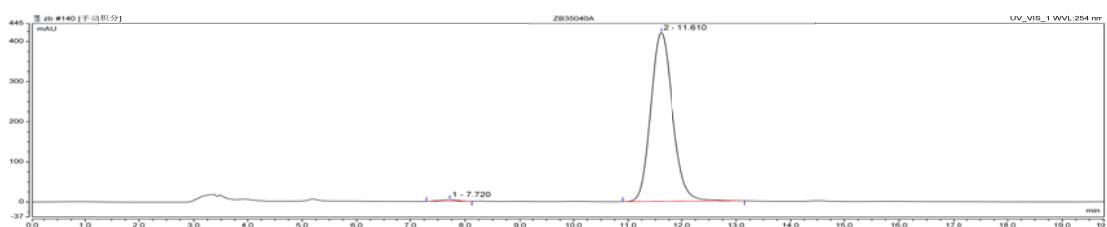
The *ee* was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 14.8 min (minor) and 16.5 min (major).



Green solid, Mp 131.8–132.9 °C; 55% yield; 98% *ee*; $[\alpha]_D^{26} -201.9$ (*c* 1.98, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.33 (d, *J* = 8.4 Hz, 1H), 8.06 (d, *J* = 8.4 Hz, 1H), 7.90–7.86 (m, 2H), 7.78–7.74 (m, 2H), 7.67–7.59 (m, 3H), 7.45–7.43 (m, 3H), 6.30 (d, *J* = 4.8 Hz, 1H), 4.03 (d, *J* = 4.8 Hz, 1H), 2.08–1.96 (m, 1H), 1.33–1.21 (m, 1H), 0.95 (t, *J* = 7.3 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.2, 150.5, 147.1, 138.2, 136.3, 130.5, 129.6, 129.0, 128.8, 128.0, 127.8, 127.6, 117.6, 98.6, 76.0, 73.6, 55.2, 21.4, 10.9; HRMS (ESI) m/z 406.1228 (M+H⁺), calc. for C₂₂H₂₀N₃O₃S 406.1225.

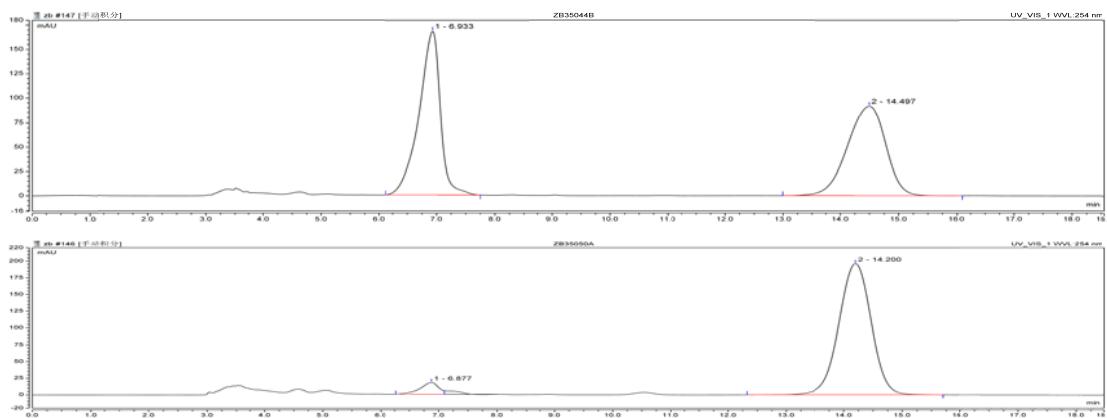
The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 7.7 min (minor) and 11.6 min (major).





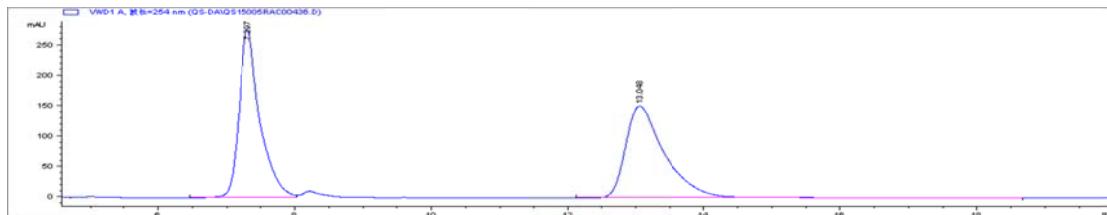
White solid, Mp 165.3–171.9 °C; 45% yield; 90% ee; $[\alpha]_D^{26} -181.8$ (*c* 1.48, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.22 (s, 1H), 7.64–7.57 (m, 4H), 7.50–7.49 (m, 2H), 7.43–7.41 (m, 3H), 5.98 (d, *J* = 4.7 Hz, 1H), 3.90 (d, *J* = 4.7 Hz, 1H), 1.23 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 178.2, 135.6, 132.6, 130.3, 129.1, 128.5, 127.4, 124.3, 98.5, 76.3, 66.1, 56.1, 13.4; HRMS (ESI) m/z 419.0064 (M+H⁺), calc. for C₁₈H₁₆BrN₂O₃S 419.0065.

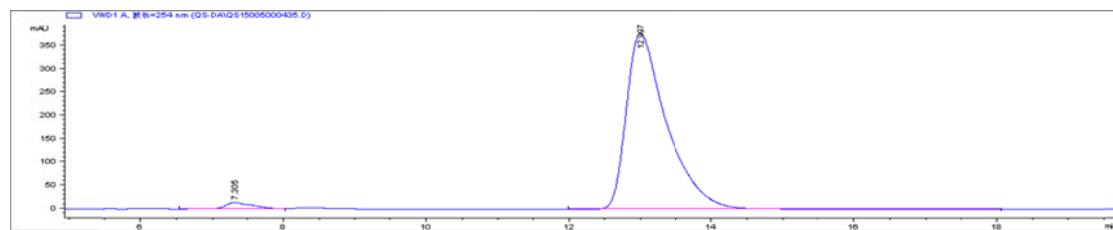
The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 6.9 min (minor) and 14.2 min (major).



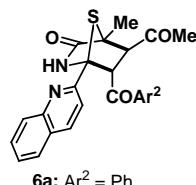
White solid, Mp 136.9–138.1 °C; 76% yield; 95% ee; $[\alpha]_D^{27} -569.3$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.62–8.61 (m, 1H), 7.89–7.83 (m, 2H), 7.70 (d, *J* = 7.9 Hz, 1H), 7.64–7.61 (m, 2H), 7.45–7.35 (m, 4H), 6.21 (d, *J* = 4.8 Hz, 1H), 3.91 (d, *J* = 4.8 Hz, 1H), 1.24 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 177.4, 150.8, 149.6, 137.7, 136.0, 129.0, 128.9, 128.6, 124.5, 120.8, 98.2, 76.6, 66.6, 55.3, 13.7; HRMS (ESI) m/z 342.0911 (M+H⁺), calc. for C₁₇H₁₆N₃O₃S 342.0912.

The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 7.3 min (minor) and 13.0 min (major).



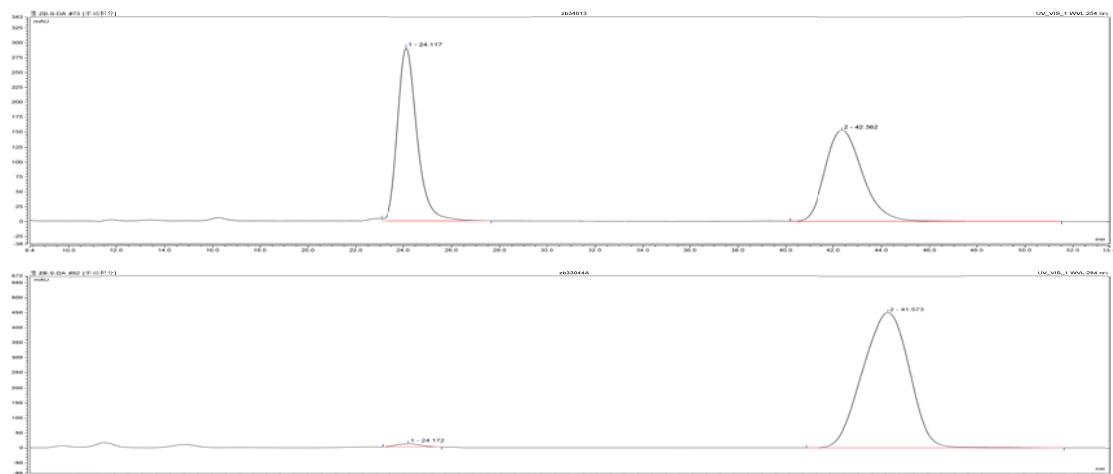


White solid, Mp 164.9–165.8 °C; 86% yield; 98% *ee*; $[\alpha]_D^{24} -169.6$ (*c* 2.37, CHCl₃);

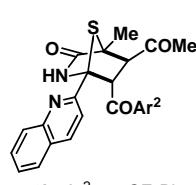


¹H NMR (300 MHz, CDCl₃) δ 8.14 (d, *J* = 8.4 Hz, 1H), 7.72 (d, *J* = 8.4 Hz, 2H), 7.68–7.63 (m, 3H), 7.61–7.56 (m, 2H), 7.48 (dd, *J* = 7.8, 7.2 Hz, 1H), 7.34 (t, *J* = 7.2 Hz, 1H), 7.22 (t, *J* = 7.8 Hz, 2H), 5.70 (d, *J* = 5.4 Hz, 1H), 3.56 (d, *J* = 5.4 Hz, 1H), 2.28 (s, 3H), 1.77 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 204.9, 198.2, 176.5, 152.2, 146.7, 137.6, 136.9, 133.1, 129.9, 129.2, 128.4, 128.0, 127.5, 127.3, 127.2, 118.2, 79.6, 66.6, 62.0, 58.4, 32.8, 14.4; HRMS (ESI) m/z 417.1277 (M+H⁺), calc. for C₂₄H₂₁N₂O₃S 417.1273.

The *ee* was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 24.2 min (minor) and 41.6 min (major).

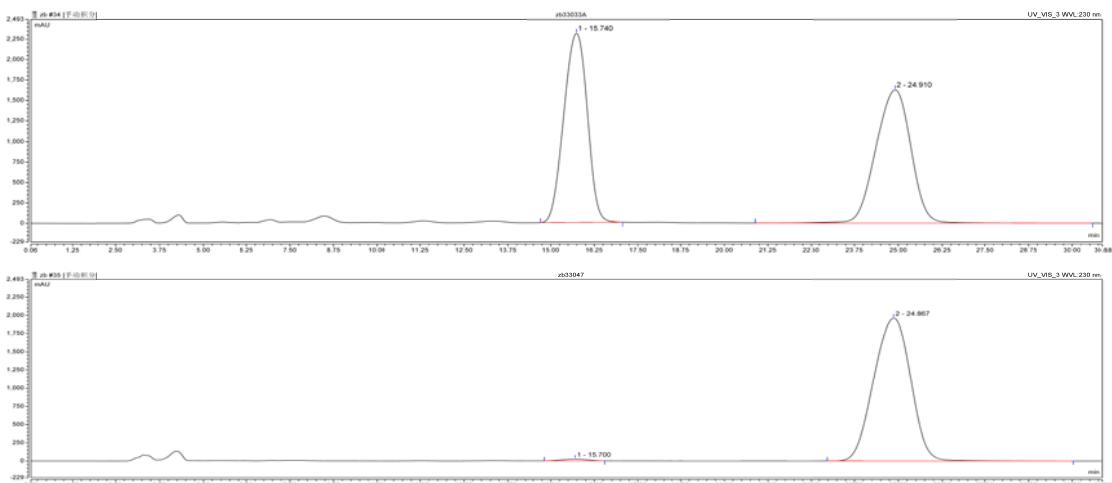


White solid, Mp 181.7–183.6 °C; 90% yield; 98% *ee*; $[\alpha]_D^{25} -144.8$ (*c* 4.22, ¹H



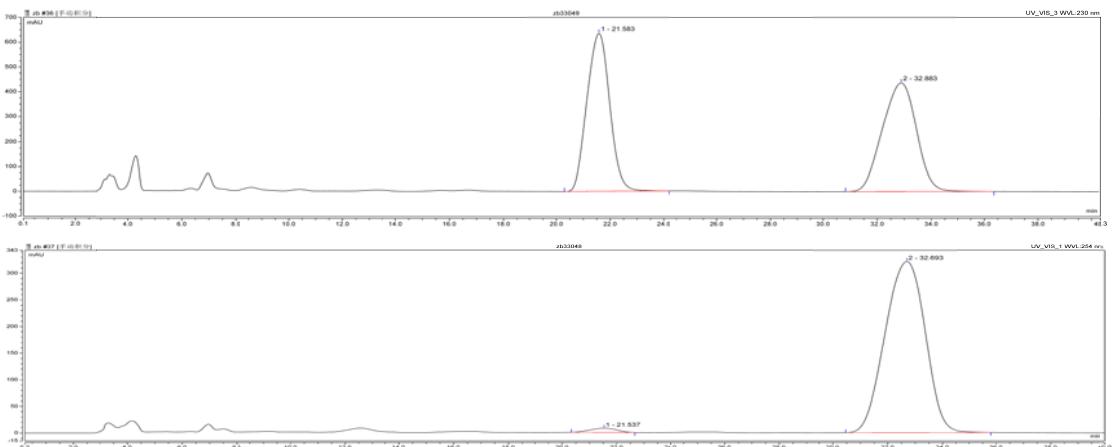
NMR (300 MHz, CDCl₃) δ 8.13 (d, *J* = 8.4 Hz, 1H), 7.71–7.67 (m, 4H), 7.60–7.53 (m, 3H), 7.51–7.46 (m, 1H), 7.38 (d, *J* = 8.4 Hz, 2H), 5.65 (d, *J* = 5.2 Hz, 1H), 3.67 (d, *J* = 5.2 Hz, 1H), 2.32 (s, 3H), 1.79 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 204.6, 197.2, 176.2, 151.7, 146.5, 139.8, 137.8, 134.2, 133.8, 130.2, 128.8, 128.2, 127.5, 127.4, 125.2, 125.1, 125.0, 121.3, 118.2, 79.7, 66.7, 62.4, 58.2, 32.6, 14.5; HRMS (ESI) m/z 485.1149 (M+H⁺), calc. for C₂₅H₂₀F₃N₂O₃S 485.1147.

The *ee* was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 230 nm; retention time: 15.7 min (minor) 24.9 min (major).



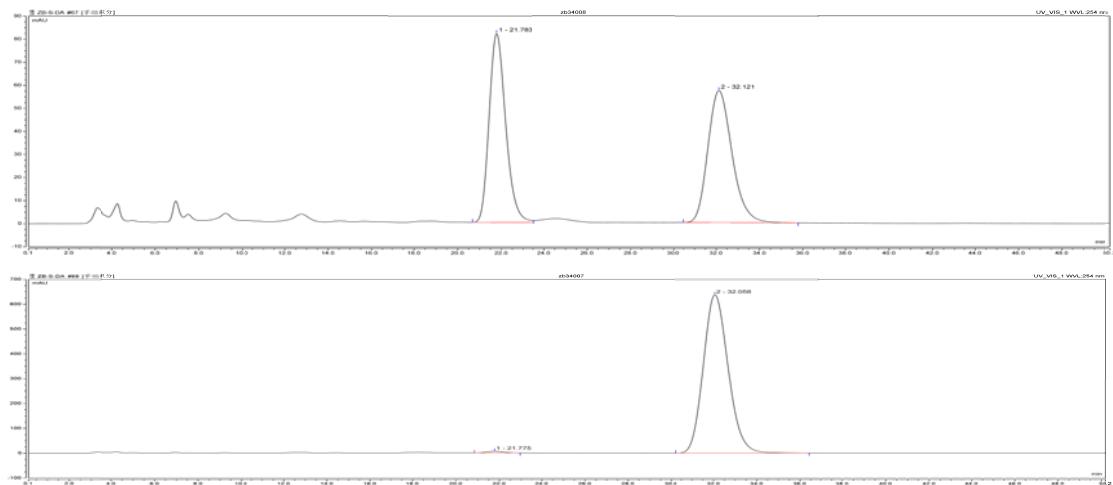
Pale yellow solid, Mp 143.2–145.0 °C; 80% yield; 97% *ee*; $[\alpha]_D^{26} -172.8$ (*c* 2.92, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.16 (d, *J* = 8.7 Hz, 1H), 7.73 (t, *J* = 8.7 Hz, 2H), 7.66–7.58 (m, 4H), 7.52–7.48 (m, 2H), 7.18 (d, *J* = 8.7 Hz, 2H), 5.64 (d, *J* = 5.3 Hz, 1H), 3.56 (d, *J* = 5.3 Hz, 1H), 2.30 (s, 3H), 1.78 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 204.7, 197.0, 176.4, 152.0, 146.7, 139.7, 137.7, 135.3, 130.1, 129.4, 129.1, 128.6, 127.6, 127.4, 118.3, 79.8, 66.7, 61.6, 58.5, 32.7, 14.5; HRMS (ESI) *m/z* 451.0881 (M+H⁺), calc. for C₂₄H₂₀ClN₂O₃S 451.0883.

The *ee* was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 21.5 min (minor) and 32.7 min (major).



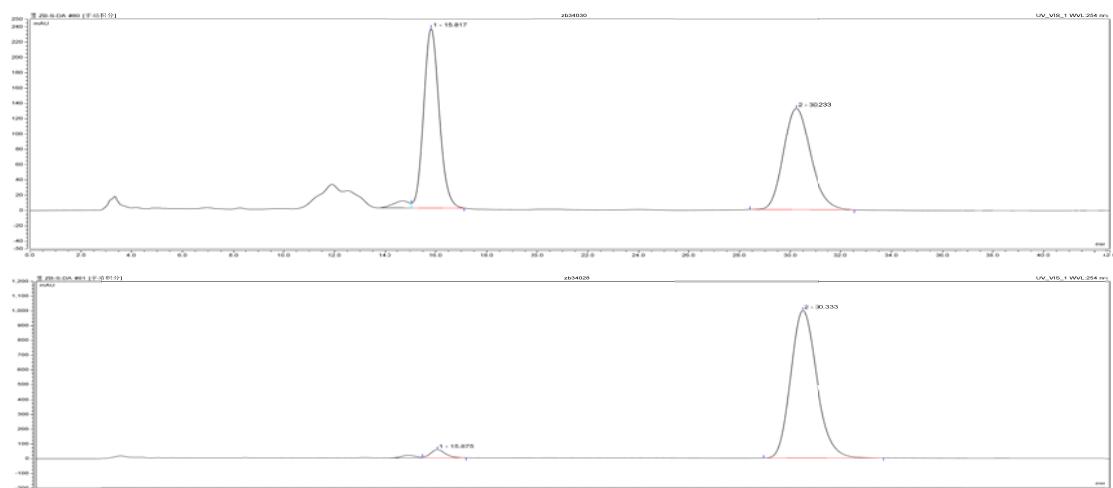
White solid, Mp 175.2–176.5 °C; 91% yield; 99% *ee*; $[\alpha]_D^{26} -167.5$ (*c* 1.26, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.18 (d, *J* = 8.4 Hz, 1H), 7.76 (d, *J* = 8.4 Hz, 1H), 7.71 (d, *J* = 8.4 Hz, 1H), 7.67–7.60 (m, 2H), 7.54–7.49 (m, 3H), 7.42 (s, 1H), 7.35 (d, *J* = 8.4 Hz, 2H), 5.62 (d, *J* = 5.3 Hz, 1H), 3.57 (d, *J* = 5.3 Hz, 1H), 2.31 (s, 3H), 1.79 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 204.7, 197.2, 176.4, 151.9, 146.7, 137.8, 135.7, 131.7, 130.2, 129.5, 129.1, 128.4, 127.6, 127.5, 127.4, 118.2, 79.7, 66.7, 61.7, 58.4, 32.8, 14.5; HRMS (ESI) *m/z* 495.0381 (M+H⁺), calc. for C₂₄H₂₀BrN₂O₃S 495.0378.

The *ee* was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 21.8min (minor) and 32.0 min (major).



White solid, Mp 176.8–178.7 °C; 95% yield; 94% *ee*; $[\alpha]_D^{25} -191.5$ (*c* 4.46, CHCl₃);
 $\text{6e: Ar}^2 = 3,4\text{-Cl}_2\text{Ph}$ ¹H NMR (300 MHz, CDCl₃) δ 8.20 (d, *J* = 8.4 Hz, 1H), 7.76–7.68 (m, 3H), 7.62–7.58 (m, 3H), 7.53–7.48 (m, 1H), 7.44 (dd, *J* = 8.4, 1.9 Hz, 1H), 7.20 (d, *J* = 8.4 Hz, 1H), 5.52 (d, *J* = 5.2 Hz, 1H), 3.69 (d, *J* = 5.2 Hz, 1H), 2.34 (s, 3H), 1.79 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 204.6, 195.6, 176.2, 151.7, 146.5, 137.8, 137.5, 136.4, 132.8, 130.3, 130.2, 129.8, 128.9, 127.6, 127.5, 127.4, 127.0, 118.2, 79.7, 66.7, 62.2, 58.1, 32.6, 14.5; HRMS (ESI) m/z 485.0494 (M+H⁺), calc. for C₂₄H₁₉Cl₂N₂O₃S 485.0493.

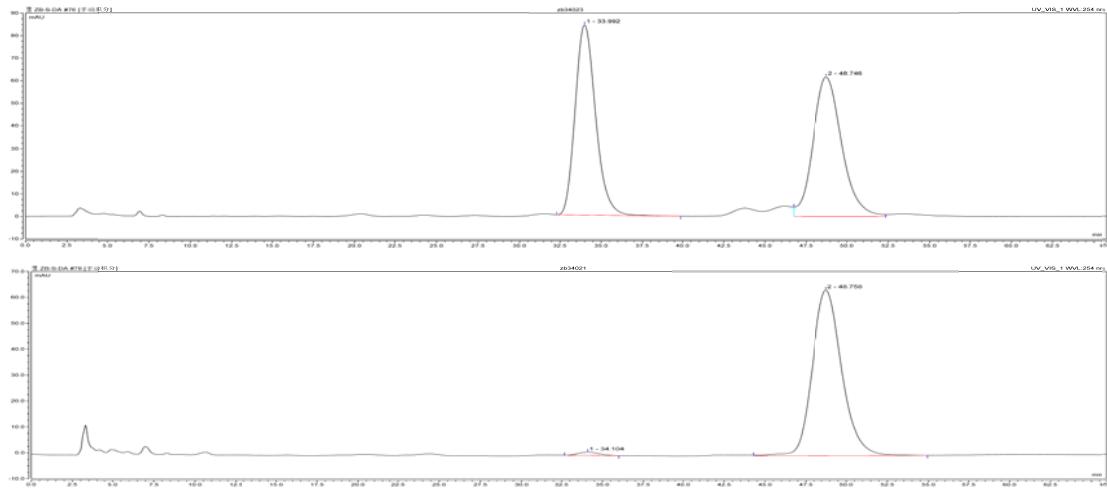
The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 15.9min (minor) and 30.3 min (major).



White solid, Mp 148.5–150.0 °C; 87% yield; 97% *ee*; $[\alpha]_{25D}^{25} -190.5$ (*c* 3.78, CHCl₃);
 $\text{6f: Ar}^2 = 4\text{-MeOPh}$ ¹H NMR (300 MHz, CDCl₃) δ 8.14 (d, *J* = 8.7 Hz, 1H), 7.73 (dd, *J* = 8.7, 5.3 Hz, 3H), 7.65 (t, *J* = 5.3 Hz, 2H), 7.67–7.55 (m, 2H), 7.51–7.46 (m, 1H), 6.69 (d, *J* = 8.7 Hz, 2H), 5.64 (d, *J* = 5.4 Hz, 1H), 3.72 (s, 3H), 3.54 (d, *J* = 5.4 Hz, 1H), 2.29 (s,

3H), 1.76 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 205.1, 196.3, 176.6, 163.6, 152.4, 146.8, 137.5, 130.4, 129.9, 129.8, 129.4, 127.6, 127.3, 127.2, , 118.2, 113.6, 79.7, 66.6, 61.5, 58.7, 55.4, 32.8, 14.4; HRMS (ESI) m/z 447.1377 ($\text{M}+\text{H}^+$), calc. for $\text{C}_{25}\text{H}_{23}\text{N}_2\text{O}_4\text{S}$ 447.1379.

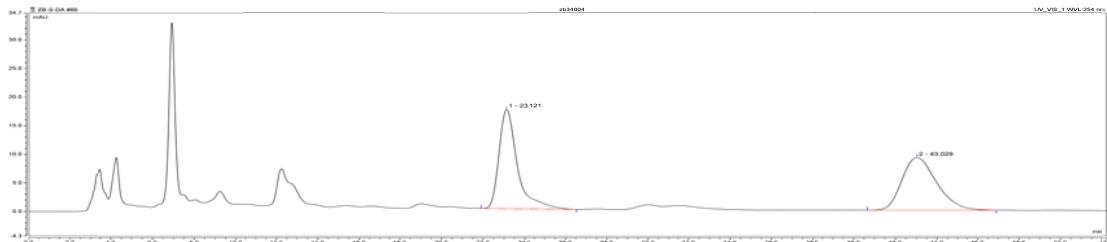
The *ee* was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 34.1 min (minor) and 48.8 min (major).

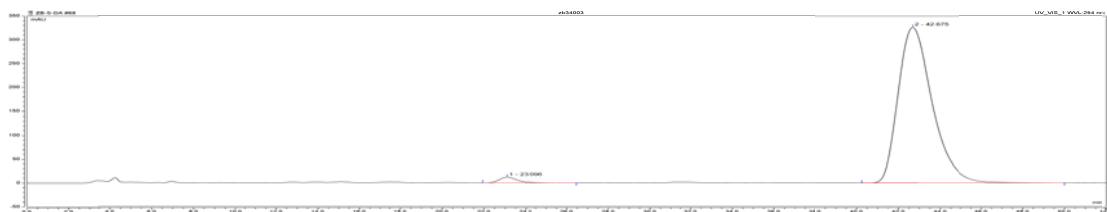


6g: $\text{Ar}^2 = 3\text{-MeOPh}$

White solid, Mp 149.1–150.8 °C; 82% yield; 96% *ee*; $[\alpha]_D^{24} -192.4$ (c 1.47, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 8.17 (d, $J = 8.3$ Hz, 1H), 7.73 (t, $J = 8.3$ Hz, 3H), 7.62 (dd, $J = 11.2, 4.6$ Hz, 1H), 7.53–7.47 (m, 2H), 7.22 (d, $J = 7.7$ Hz, 1H), 7.14 (dt, $J = 11.2, 4.6$ Hz, 2H), 6.89–6.86 (m, 1H), 5.65 (d, $J = 5.4$ Hz, 1H), 3.72 (s, 3H), 3.55 (d, $J = 5.4$ Hz, 1H), 2.30 (s, 3H), 1.78 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 204.9, 197.9, 176.4, 159.5, 152.1, 146.8, 138.2, 137.6, 130.0, 129.4, 129.3, 127.6, 127.4, 127.3, 120.6, 119.9, 118.2, 112.0, 79.5, 66.6, 62.5, 58.4, 55.34, 32.9, 14.4; HRMS (ESI) m/z 447.1381 ($\text{M}+\text{H}^+$), calc. for $\text{C}_{25}\text{H}_{23}\text{N}_2\text{O}_4\text{S}$ 447.1379.

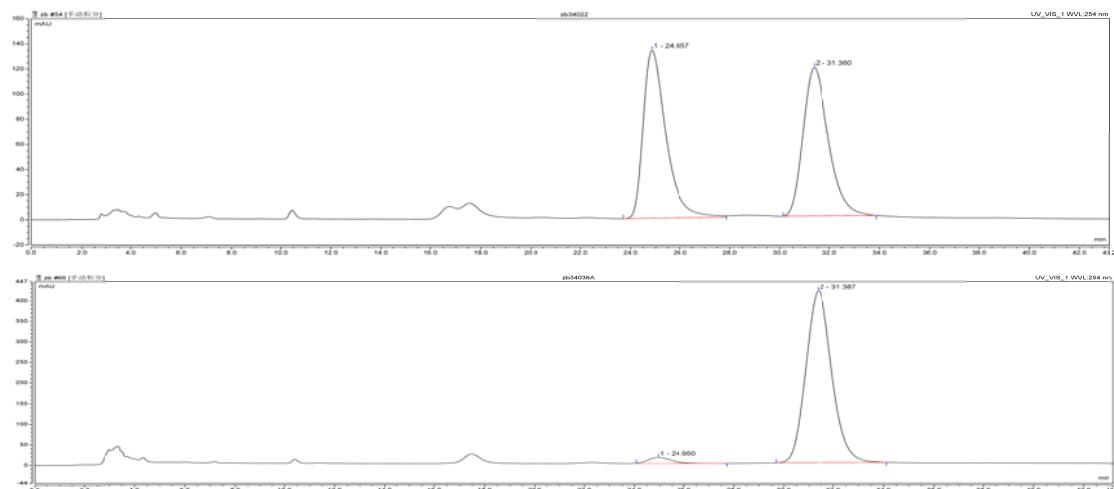
The *ee* was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 23.1 min (minor) and 42.7 min (major).





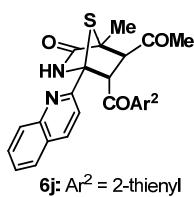
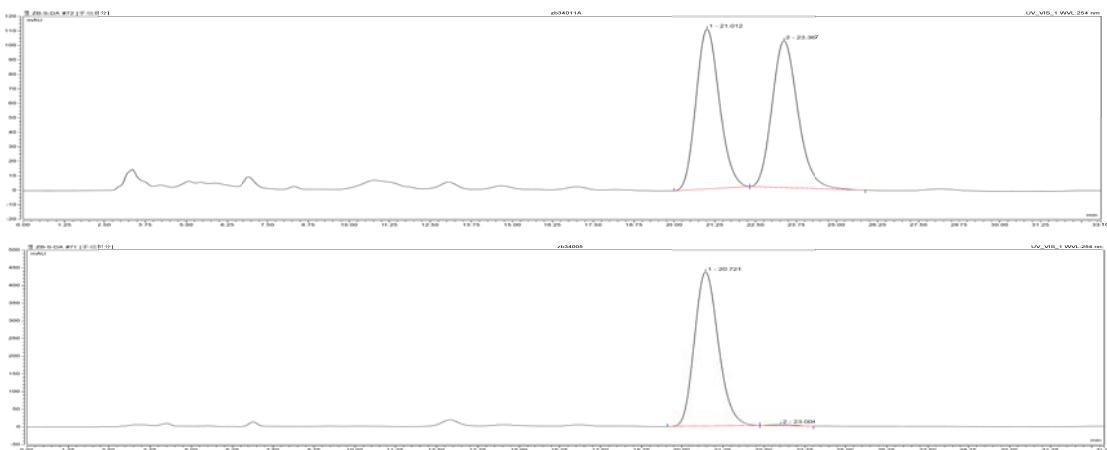
Weak yellow solid, Mp 132.1–133.8 °C; 64% yield; 94% ee; $[\alpha]_D^{26} -198.8$ (*c* 2.28, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.00 (d, *J* = 8.5 Hz, 1H), 7.79 (d, *J* = 8.5 Hz, 1H), 7.70–7.62 (m, 3H), 7.56 (d, *J* = 8.5 Hz, 1H), 7.50 (t, *J* = 7.5 Hz, 1H), 7.03–6.97 (m, 2H), 6.54–6.46 (m, 2H), 5.52 (d, *J* = 5.2 Hz, 1H), 3.80 (d, *J* = 5.2 Hz, 1H), 3.72 (s, 3H), 2.32 (s, 3H), 1.74 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 205.0, 199.3, 176.2, 156.8, 151.9, 146.6, 137.0, 132.8, 129.9, 129.7, 129.3, 128.5, 127.4, 127.3, 127.2, 120.3, 117.6, 110.4, 78.8, 68.5, 66.4, 57.1, 55.0, 32.3, 14.0; HRMS (ESI) m/z 447.1383 (M+H⁺), calc. for C₂₅H₂₃N₂O₄S 447.1379.

The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 25.0 min (minor) and 31.4 min (major).



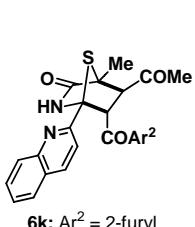
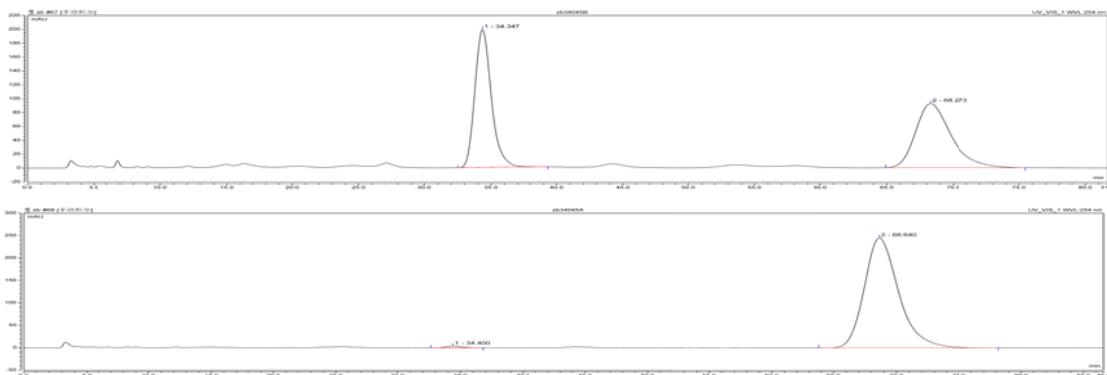
White solid, Mp 151.5–153.0 °C; 60% yield; 99% ee; $[\alpha]_D^{25} -170.0$ (*c* 0.725, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.94 (d, *J* = 8.6 Hz, 1H), 7.85 (d, *J* = 8.6 Hz, 1H), 7.72 (s, 1H), 7.67–7.62 (m, 2H), 7.54–7.45 (m, 2H), 7.11 (dd, *J* = 7.6, 1.6 Hz, 1H), 6.83 (dd, *J* = 7.6, 1.6 Hz, 1H), 6.76–6.65 (m, 2H), 5.44 (d, *J* = 5.1 Hz, 1H), 3.90 (d, *J* = 5.1 Hz, 1H), 2.29 (s, 3H), 1.71 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 204.6, 198.8, 175.9, 151.4, 146.5, 140.2, 137.5, 132.9, 131.2, 130.2, 129.3, 128.7, 127.5, 127.4, 126.6, 118.6, 117.6, 78.2, 68.2, 66.5, 56.6, 32.6, 14.3; HRMS (ESI) m/z 495.0375 (M+H⁺), calc. for C₂₄H₂₀BrN₂O₃S 495.0378.

The *ee* was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 20.7 min (major) and 23.0 min (minor).



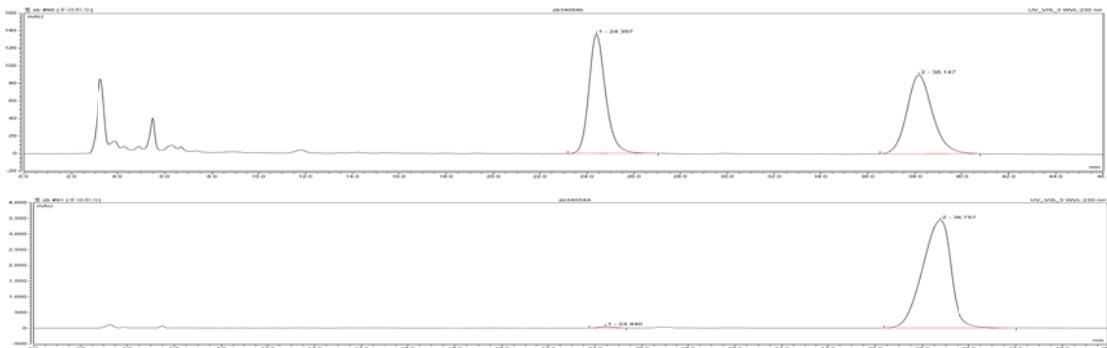
White solid, Mp 179.1–181.0 °C; 98% yield; 99% ee; $[\alpha]_D^{26} -147.2$ (*c* 4.06, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.19 (d, *J* = 8.2 Hz, 1H), 7.76–7.73 (m, 3H), 7.65 (s, 1H), 7.63–7.58 (m, 1H), 7.51 (d, *J* = 8.2 Hz, 1H), 7.46–7.44 (m, 2H), 6.89–6.86 (m, 1H), 5.52 (d, *J* = 5.4 Hz, 1H), 3.65 (d, *J* = 5.4 Hz, 1H), 2.35 (s, 3H), 1.78 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 204.8, 190.2, 176.6, 152.1, 146.8, 144.2, 137.6, 134.7, 132.5, 130.0, 129.3, 128.0, 127.6, 127.4, 127.3, 118.3, 79.8, 66.7, 62.5, 59.0, 32.8, 14.4; HRMS (ESI) m/z 423.0833 (M+H⁺), calc. for C₂₂H₁₉N₂O₃S₂ 423.0837.

The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 34.4 min (minor) and 68.6 min (major).

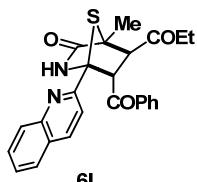


White solid, Mp 165.0–166.2 °C; 92% yield; 99% ee; $[\alpha]_D^{26} -192.1$ (*c* 1.36, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.20 (d, *J* = 8.0 Hz, 1H), 7.83–7.73 (m, 3H), 7.64 (t, *J* = 8.0 Hz, 1H), 7.55–7.50 (m, 2H), 7.27 (s, 1H), 7.03 (d, *J* = 3.6 Hz, 1H), 6.36–6.31 (m, 1H), 5.39 (d, *J* = 5.3 Hz, 1H), 3.64 (d, *J* = 5.3 Hz, 1H), 2.39 (s, 3H), 1.77 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 204.5, 185.3, 176.3, 152.3, 152.2, 146.9, 146.6, 137.5, 130.0, 129.4, 127.6, 127.4, 127.3, 118.2, 118.1, 112.5, 79.2, 66.3, 62.9, 58.3, 32.2, 14.3; HRMS (ESI) m/z 407.1069 (M+H⁺), calc. for C₂₂H₁₉N₂O₄S 407.1066.

The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 230 nm; retention time: 24.4 min (minor) and 38.8 min (major).

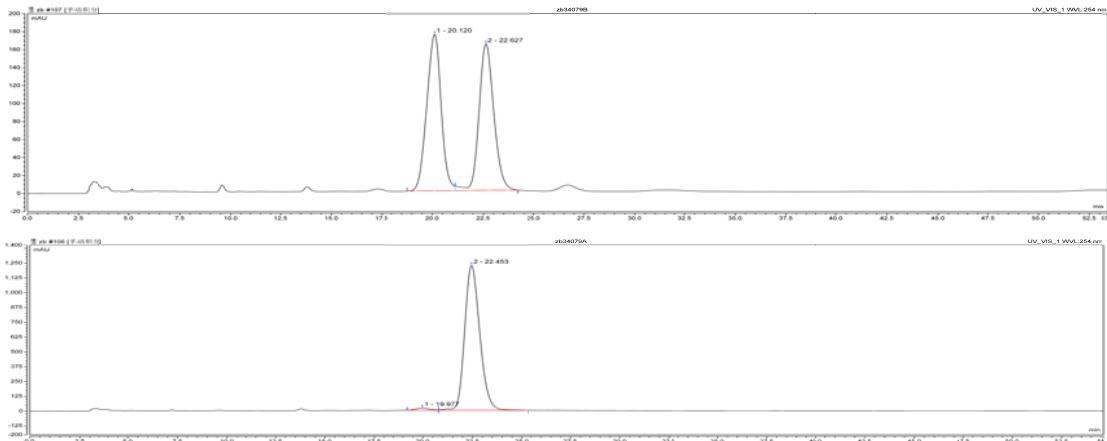


White solid, Mp 158.2–160.0 °C; 90% yield; 97% ee; $[\alpha]_D^{20} -199.8$ (*c* 2.65, CHCl₃);

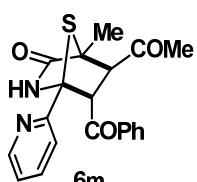


¹H NMR (300 MHz, CDCl₃) δ 8.15 (d, *J* = 8.5 Hz, 1H), 7.74–7.70 (m, 3H), 7.64–7.56 (m, 3H), 7.54–7.46 (m, 2H), 7.35 (t, *J* = 7.4 Hz, 1H), 7.22 (d, *J* = 7.4 Hz, 1H), 5.69 (d, *J* = 5.5 Hz, 1H), 3.50 (d, *J* = 5.5 Hz, 1H), 2.71–2.60 (m, 1H), 2.35–2.22 (m, 1H), 1.71 (s, 3H), 1.04 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 207.9, 198.3, 176.6, 152.2, 146.8, 137.6, 136.9, 133.2, 129.9, 129.3, 128.4, 128.0, 127.6, 127.3, 127.2, 118.2, 79.5, 66.8, 62.3, 57.4, 39.2, 14.2, 7.6; HRMS (ESI) m/z 431.1430 (M+H⁺), calc. for C₂₅H₂₃N₂O₃S 431.1429.

The ee was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 20.0 min (minor) and 22.4 min (major).

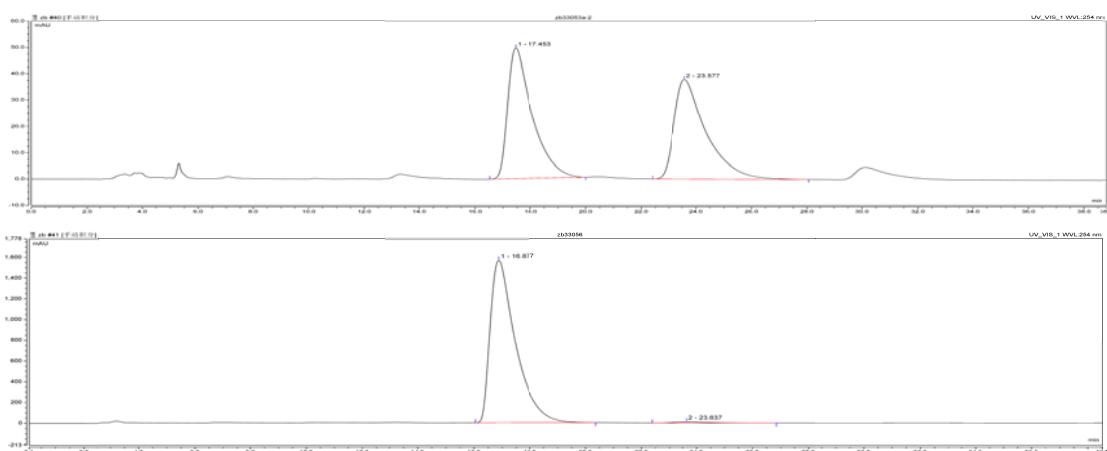


White solid, Mp 138.6–139.1 °C; 87% yield; 98% ee; $[\alpha]_D^{23} -104.5$ (*c* 1.71, CHCl₃);

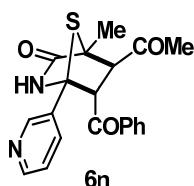


¹H NMR (300 MHz, CDCl₃) δ 8.81 (d, *J* = 1.8 Hz, 1H), 8.54 (dd, *J* = 4.8, 1.8 Hz, 1H), 7.87 – 7.83 (m, 1H), 7.64 – 7.62 (m, 3H), 7.55 (t, *J* = 7.6 Hz, 1H), 7.38 (t, *J* = 7.6 Hz, 2H), 7.29 – 7.27 (m, 1H), 5.44 (d, *J* = 5.4 Hz, 1H), 3.35 (d, *J* = 5.4 Hz, 1H), 2.24 (s, 3H), 1.71 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 204.5, 198.3, 177.1, 150.2, 147.3, 136.0, 134.2, 133.9, 130.2, 129.0, 128.0, 123.5, 76.4, 65.6, 61.6, 59.1, 33.0, 14.2; HRMS (ESI) m/z 367.1119 (M+H⁺), calc. for C₂₀H₁₉N₂O₃S 367.1116.

The ee was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 16.9 min (major) and 23.6 min (minor).

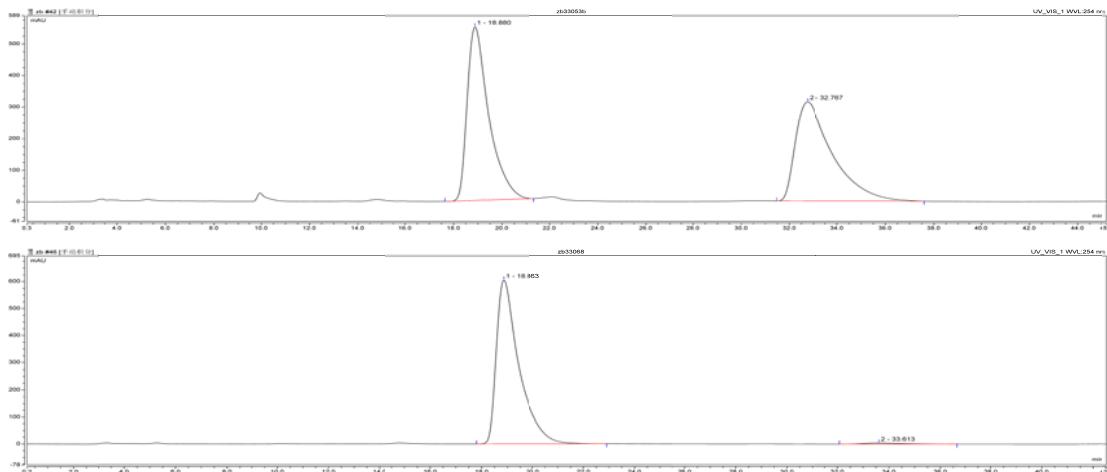


White solid, Mp 120.2–122.1 °C; 86% yield; 98% ee; $[\alpha]_D^{24} -112.9$ (c 1.81, CHCl₃);

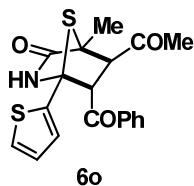


¹H NMR (300 MHz, CDCl₃) δ 8.28–8.27 (m, 1H), 7.67–7.61 (m, 1H), 7.59–7.55 (m, 3H), 7.44–7.38 (m, 1H), 7.28 – 7.23 (m, 3H), 7.10–7.06 (m, 1H), 5.50 (d, $J = 5.4$ Hz, 1H), 3.40 (d, $J = 5.4$ Hz, 1H), 2.19 (s, 3H), 1.67 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 204.8, 198.0, 176.6, 152.6, 149.1, 137.1, 136.6, 133.5, 128.6, 128.1, 123.7, 121.0, 78.9, 66.2, 62.4, 58.6, 32.8, 14.4; HRMS (ESI) m/z 367.1112 (M+H⁺), calc. for C₂₀H₁₉N₂O₃S 367.1116.

The ee was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 18.9 min (major) and 33.6 min (minor).

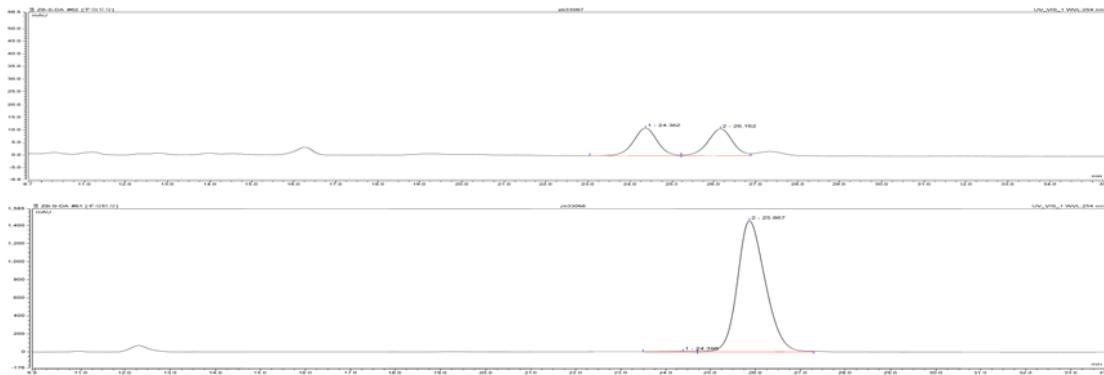


White solid, Mp 134.4–135.6 °C; 70% yield; 99% ee; $[\alpha]_D^{25} -99.7$ (c 0.48, CHCl₃);



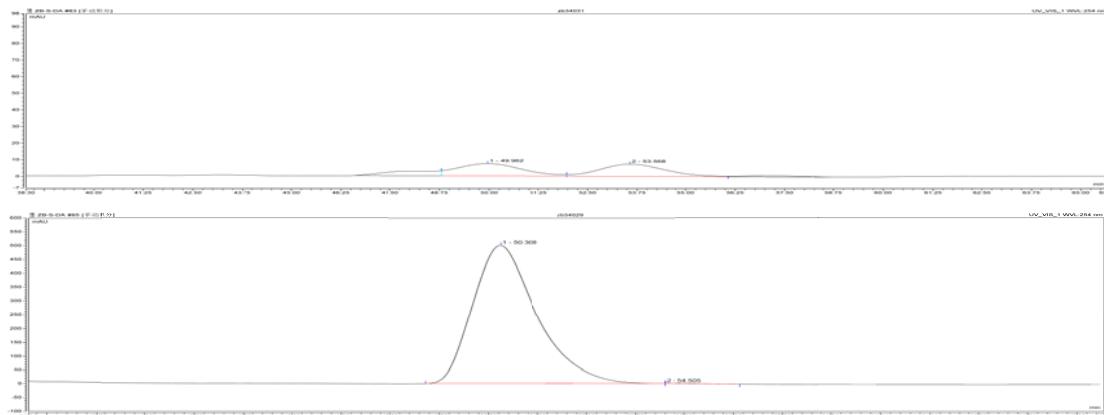
¹H NMR (300 MHz, CDCl₃) δ 7.74 (d, $J = 7.5$ Hz, 2H), 7.59 (t, $J = 7.5$ Hz, 1H), 7.44 (t, $J = 7.5$ Hz, 2H), 7.28 (dd, $J = 5.1, 2.4$ Hz, 1H), 7.11–7.10 (m, 1H), 6.98 (s, 1H), 6.90 (dd, $J = 5.1, 2.4$ Hz, 1H), 5.43 (d, $J = 5.5$ Hz, 1H), 3.26 (d, $J = 5.5$ Hz, 1H), 2.21 (s, 3H), 1.70 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 204.2, 198.6, 176.7, 136.9, 136.3, 134.2, 129.1, 128.2, 127.2, 126.6, 126.3, 75.3, 66.2, 61.8, 59.8, 32.9, 14.3; HRMS (ESI) m/z 372.0733 (M+H⁺), calc. for C₁₉H₁₈NO₃S₂ 372.0728.

The *ee* was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 24.4 min (minor) and 25.9 min (major).



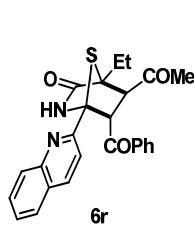
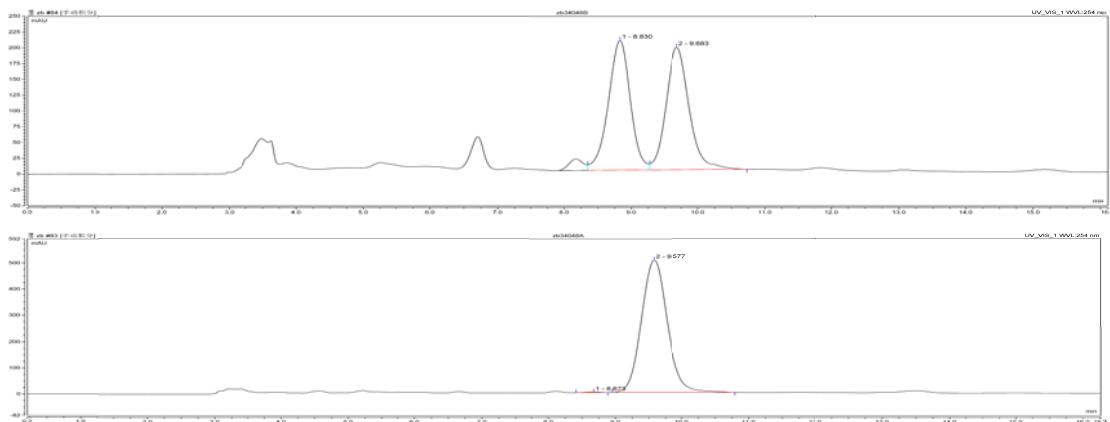
White solid, Mp 155.2–156.7 °C; 75% yield; 99% *ee*; $[\alpha]_D^{26} -94.3$ (*c* 0.74, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.72 (d, *J* = 7.6 Hz, 2H), 7.56 (t, *J* = 7.6 Hz, 1H), 7.42 (t, *J* = 7.6 Hz, 2H), 7.29 (s, 1H), 6.91 (s, 1H), 6.45 (d, *J* = 3.3 Hz, 1H), 6.32–6.30 (m, 1H), 5.42 (d, *J* = 5.5 Hz, 1H), 3.32 (d, *J* = 5.5 Hz, 1H), 2.22 (s, 3H), 1.70 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 204.4, 198.1, 176.4, 146.5, 143.0, 136.2, 134.0, 128.9, 128.2, 111.1, 109.0, 72.9, 65.6, 60.8, 58.7, 32.8, 14.3; HRMS (ESI) m/z 356.0958 (M+H⁺), calc. for C₁₉H₁₈NO₄S 356.0957.

The *ee* was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 50.3 min (major) and 54.5 min (minor).



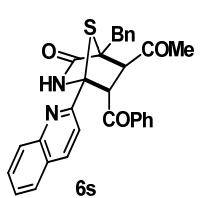
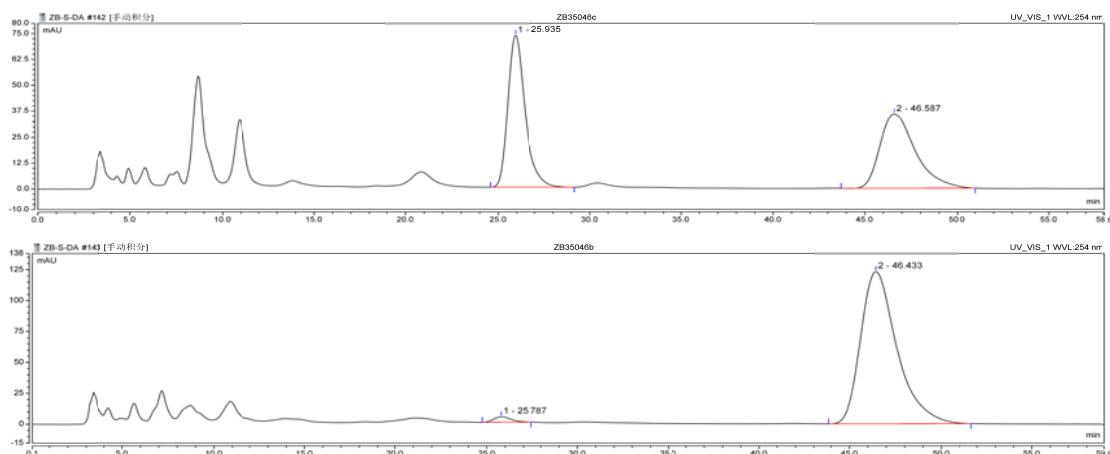
White solid, Mp 140.0–141.8 °C; 54% yield; 99% *ee*; $[\alpha]_D^{20} -111.6$ (*c* 1.61, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.68–7.63 (m, 2H), 7.59–7.54 (m, 1H), 7.48–7.45 (m, 2H), 7.41–7.38 (m, 4H), 7.29 (s, 1H), 5.40 (d, *J* = 5.4 Hz, 1H), 3.31 (d, *J* = 5.4 Hz, 1H), 2.23 (s, 3H), 1.69 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 204.6, 198.4, 177.2, 136.2, 134.2, 133.0, 132.1, 129.0, 128., 127.6, 123.2, 77.7, 65.7, 61.6, 59.4, 33.04, 14.3; HRMS (ESI) m/z 444.0273 (M+H⁺), calc. for C₂₁H₁₉BrNO₃S 444.0269.

The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 8.7 min (minor) and 9.6 min (major).



Weak yellow solid, Mp 128.1–129.6 °C; 72% yield; 97% *ee*; $[\alpha]_D^{26} -102.5$ (*c* 1.06, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 8.16 (d, *J* = 8.5 Hz, 1H), 7.77–7.66 (m, 5H), 7.62–7.56 (m, 1H), 7.52–7.46 (m, 1H), 7.43 (s, 1H), 7.40–7.35 (m, 1H), 7.28–7.23 (m, 1H), 5.70 (d, *J* = 5.5 Hz, 1H), 3.51 (d, *J* = 5.5 Hz, 1H), 2.55–2.42 (m, 1H), 2.27 (s, 3H), 1.92–1.80 (m, 1H), 1.15 (t, *J* = 7.2 Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 205.3, 198.5, 175.8, 152.4, 146.8, 137.6, 137.0, 133.2, 130.0, 129.3, 128.4, 128.1, 127.6, 127.4, 127.3, 118.3, 79.0, 74.0, 61.9, 58.6, 33.2, 22.1, 12.1; HRMS (ESI) *m/z* 431.1431 ($\text{M}+\text{H}^+$), calc. for $\text{C}_{25}\text{H}_{23}\text{N}_2\text{O}_3\text{S}$ 431.1429.

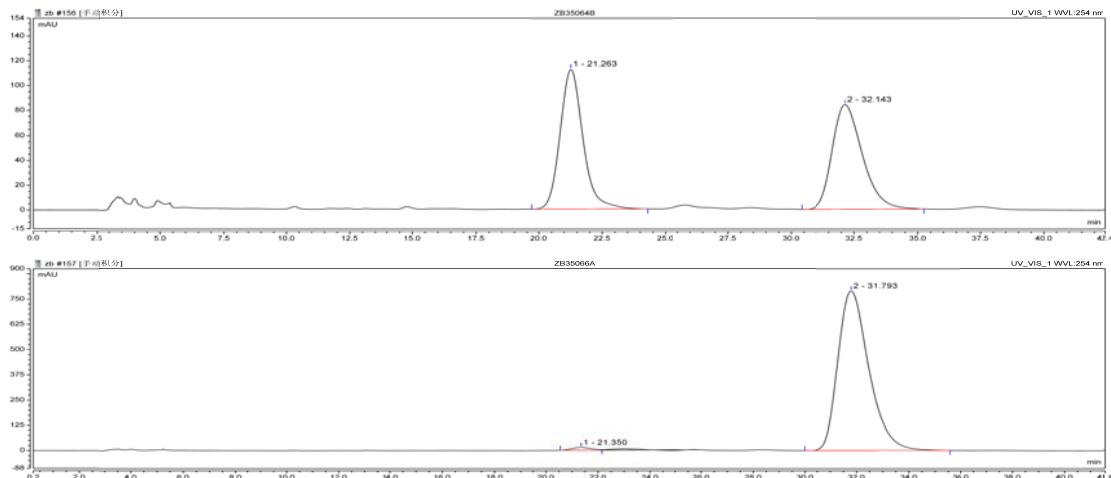
The *ee* was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 25.8 min (minor) and 46.4 min (major).



Weak yellow solid, Mp 137.3–138.8 °C; 90% yield; 98% *ee*; $[\alpha]_D^{26} -83.3$ (*c* 4.68, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 8.06 (d, *J* = 8.5 Hz, 1H), 7.69–7.62 (m, 6H), 7.58–7.53 (m, 1H), 7.48–7.42 (m, 1H), 7.39–7.34 (m, 3H), 7.30–7.22 (m, 5H), 5.66 (d, *J* = 5.6 Hz, 1H), 3.87 (d, *J* = 13.5 Hz, 1H), 3.67 (d, *J* = 5.6 Hz, 1H), 3.02 (d, *J* = 13.5 Hz, 1H), 2.29 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 205.6, 198.1, 175.6,

152.1, 146.6, 137.4, 137.2, 136.9, 133.2, 129.8, 129.1, 128.4, 128.3, 128.0, 127.5, 127.3, 127.2, 118.3, 79.1, 74.1, 62.5, 58.2, 33.8, 33.3; HRMS (ESI) m/z 493.1583 ($M+H^+$), calc. for $C_{30}H_{25}N_2O_3S$ 493.1586.

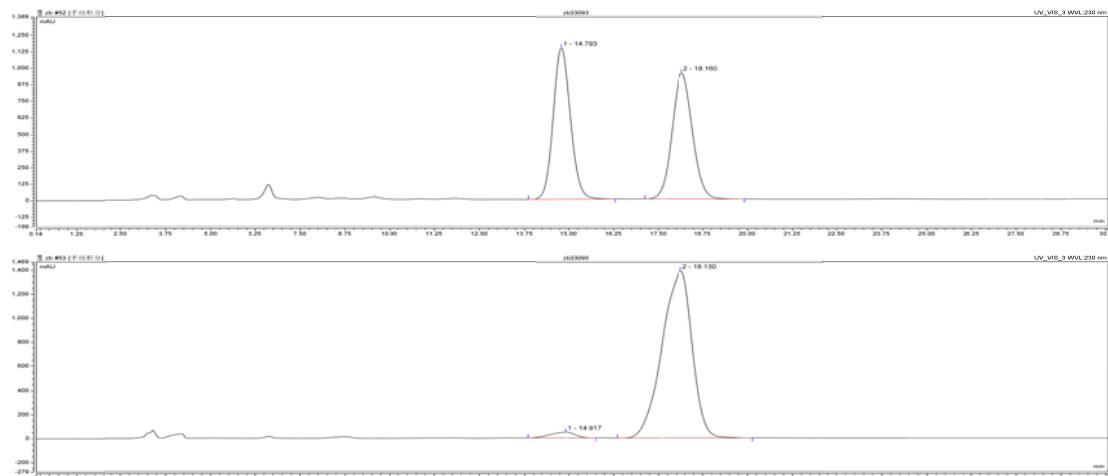
The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 21.4 min (minor) and 31.8 min (major).



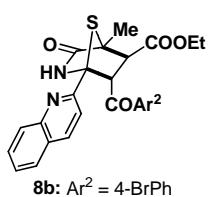
8a: $\text{Ar}^2 = \text{Ph}$,

White solid, Mp 141.8–143.2 °C; 87% yield; 95% *ee*; $[\alpha]_D^{26} -138.4$ (*c* 1.95, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 8.15 (d, *J* = 8.5 Hz, 1H), 7.74–7.66 (m, 5H), 7.62 – 7.56 (m, 1H), 7.52–7.44 (m, 2H), 7.36 (t, *J* = 7.4 Hz, 1H), 7.22 (d, *J* = 7.4 Hz, 1H), 5.76 (d, *J* = 5.4 Hz, 1H), 4.36–4.22 (m, 2H), 3.34 (d, *J* = 5.4 Hz, 1H), 1.76 (s, 3H), 1.29 (t, *J* = 7.1 Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 198.0, 176.6, 170.6, 152.2, 146.8, 137.6, 137.1, 133.1, 129.9, 129.3, 128.3, 128.1, 127.6, 127.4, 127.3, 118.3, 79.9, 66.5, 62.0, 61.8, 53.3, 14.3, 14.2; HRMS (ESI) m/z 447.1383 ($M+H^+$), calc. for $C_{25}H_{23}N_2O_4S$ 447.1379.

The *ee* was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 230 nm; retention time: 14.9 min (minor) and 18.1 min (major).

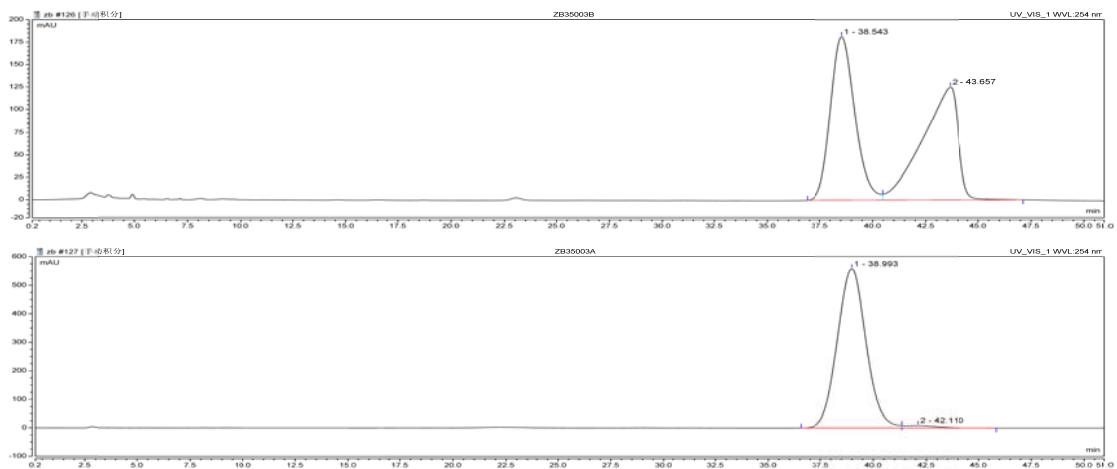


White solid, Mp 157.3–159.0 °C; 97% yield; 97% *ee*; $[\alpha]_D^{20} -170.7$ (*c* 4.98, CHCl₃);

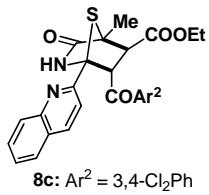


¹H NMR (300 MHz, CDCl₃) δ 8.16 (d, *J* = 8.4 Hz, 1H), 7.73 (t, *J* = 8.4 Hz, 2H), 7.65–7.57 (m, 5H), 7.53–7.47 (m, 1H), 7.35 (d, *J* = 8.4 Hz, 2H), 5.69 (d, *J* = 5.4 Hz, 1H), 4.36–4.19 (m, 2H), 3.35 (d, *J* = 5.4 Hz, 1H), 1.75 (s, 3H), 1.29 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 197.1, 176.61, 170.51, 152.1, 146.7, 137.7, 135.8, 131.5, 130.1, 129.6, 129.1, 128.4, 127.6, 127.4, 118.4, 80.1, 66.6, 61.8, 61.7, 53.2, 14.3, 14.2; HRMS (ESI) m/z 525.0486 (M+H⁺), calc. for C₂₅H₂₂BrN₂O₄S 525.0484.

The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.2 mL/min; 25 °C; 254 nm; retention time: 39.0 min (major) and 42.1 min (minor).

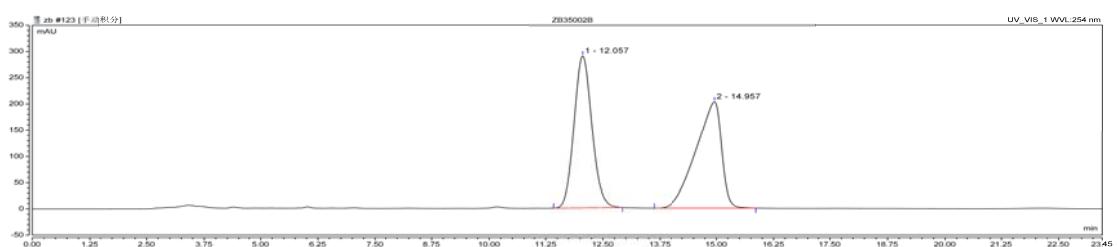


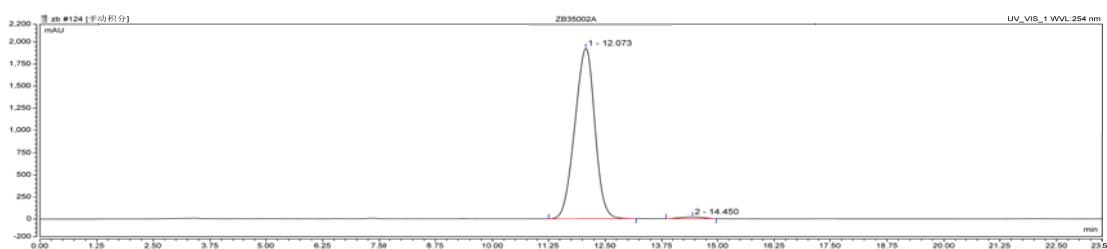
White solid, Mp 155.4–156.5 °C; 96% yield; 98% *ee*; $[\alpha]_D^{20} -124.7$ (*c* 2.76, CHCl₃);



¹H NMR (300 MHz, CDCl₃) δ 8.20 (s, 1H), 7.78–7.74 (m, 1H), 7.72–7.70 (m, 1H), 7.63 (d, *J* = 3.6 Hz, 2H), 7.58–7.49 (m, 3H), 7.27–7.24 (m, 1H), 5.59 (d, *J* = 5.2 Hz, 1H), 4.30 (qq, *J* = 10.8, 7.1 Hz, 2H), 3.43 (d, *J* = 5.2 Hz, 1H), 1.77 (s, 3H), 1.32 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 195.6, 176.5, 170.4, 151.8, 146.6, 137.8, 137.6, 136.6, 132.9, 130.3, 130.2, 130.0, 129.0, 127.6, 127.5, 127.1, 118.3, 80.0, 66.6, 62.4, 62.0, 52.9, 14.3, 14.2; HRMS (ESI) m/z 515.0602 (M+H⁺), calc. for C₂₅H₂₁Cl₂N₂O₄S 515.0599.

The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 12.1 min (major) and 14.4 min (minor).

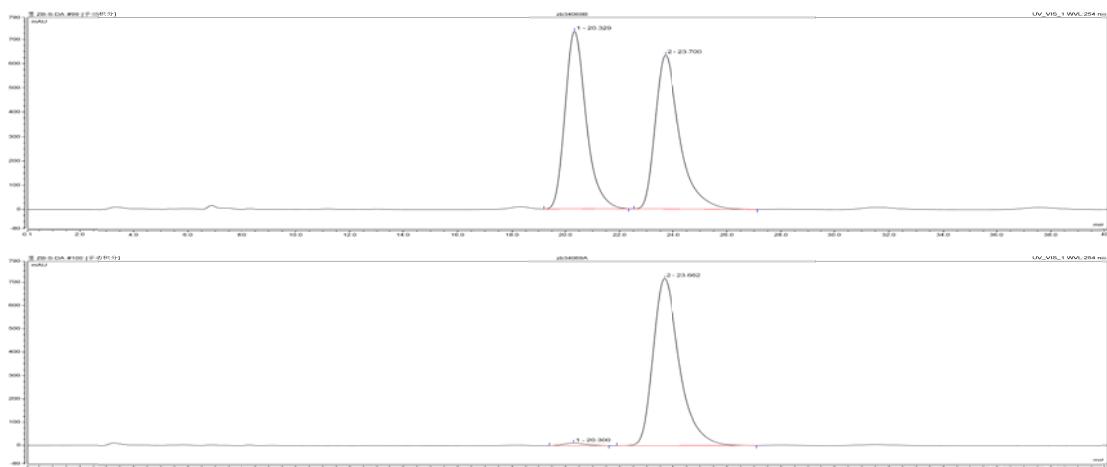




8d: $\text{Ar}^2 = 4\text{-MeOPh}$

Weak yellow solid, Mp 122.2–124.0 °C; 90% yield; 98% *ee*; $[\alpha]_D^{26} -173.9$ (*c* 4.61, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 8.16 (d, $J = 8.7$ Hz, 1H), 7.75–7.71 (m, 5H), 7.62–7.57 (m, 1H), 7.2–7.46 (m, 2H), 6.70 (d, $J = 8.7$ Hz, 2H), 5.71 (d, $J = 5.4$ Hz, 1H), 4.37–4.20 (m, 2H), 3.74 (s, 3H), 3.32 (d, $J = 5.4$ Hz, 1H), 1.75 (s, 3H), 1.30 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 196.2, 176.7, 170.7, 163.5, 152.4, 146.8, 137.5, 130.6, 130.0, 129.9, 129.4, 127.6, 127.3, 127.2, 118.3, 113.5, 80.0, 66.5, 61.7, 61.5, 55.4, 53.5, 14.3, 14.2; HRMS (ESI) m/z 477.1481 ($\text{M}+\text{H}^+$), calc. for $\text{C}_{26}\text{H}_{25}\text{N}_2\text{O}_5\text{S}$ 477.1484.

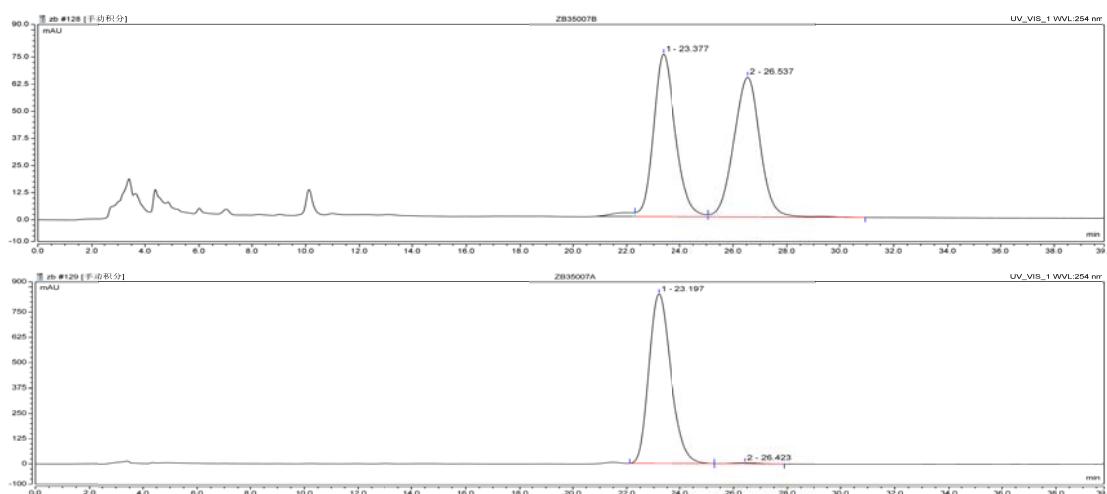
The *ee* was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 20.3 min (minor) and 23.7 min (major).



8e: $\text{Ar}^2 = 2\text{-thienyl}$

White solid, Mp 137.5–138.7 °C; 97% yield; 99% *ee*; $[\alpha]_D^{26} -132.0$ (*c* 4.67, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 8.18 (d, $J = 8.5$ Hz, 1H), 7.74–7.71 (m, 3H), 7.62–7.55 (m, 3H), 7.51–7.46 (m, 2H), 6.90–6.87 (m, 1H), 5.57 (d, $J = 5.3$ Hz, 1H), 4.37–4.20 (m, 2H), 3.41 (d, $J = 5.3$ Hz, 1H), 1.76 (s, 3H), 1.30 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 190.0, 176.6, 170.5, 152.1, 146.8, 144.3, 137.6, 134.7, 132.6, 129.9, 129.3, 127.9, 127.6, 127.4, 127.3, 118.2, 80.0, 66.5, 62.7, 61.8, 53.6, 14.2 (two peaks); HRMS (ESI) m/z 453.0940 ($\text{M}+\text{H}^+$), calc. for $\text{C}_{23}\text{H}_{21}\text{N}_2\text{O}_4\text{S}_2$ 453.0943.

The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 23.2 min (major) and 26.4 min (minor).

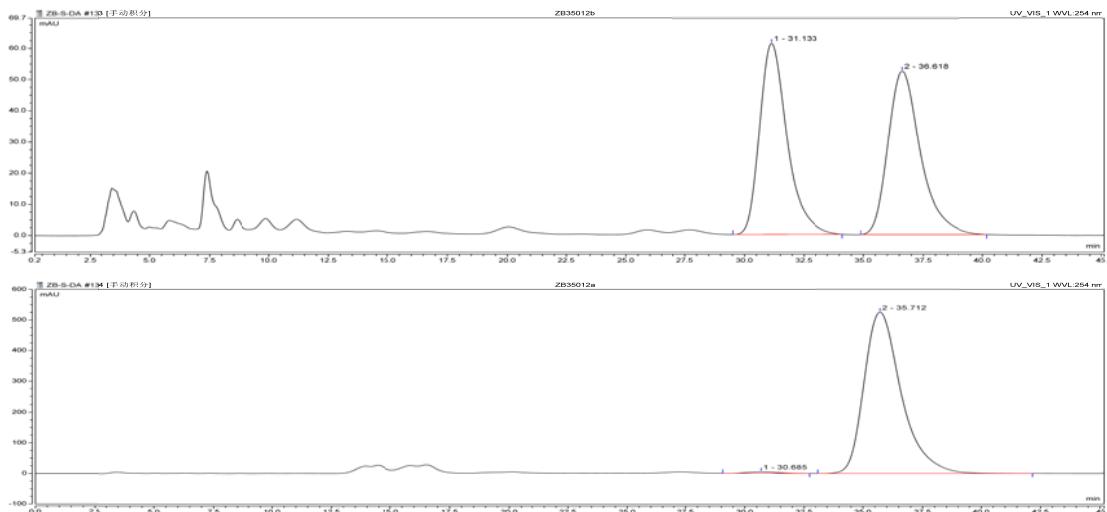


White solid, Mp 141.8–143.1 °C; 96% yield; 98% ee; $[\alpha]_D^{26} -186.1$ (*c* 4.29, CHCl₃);

8f: Ar² = 2-furyl

¹H NMR (300 MHz, CDCl₃) δ 8.19 (d, *J* = 8.5 Hz, 1H), 7.79–7.73 (m, 3H), 7.65–7.60 (m, 1H), 7.54–7.49 (m, 2H), 7.25 (s, 1H), 7.06 (d, *J* = 3.6 Hz, 1H), 6.29 (dd, *J* = 3.6, 1.7 Hz, 1H), 5.45 (d, *J* = 5.2 Hz, 1H), 4.38–4.22 (m, 2H), 3.46 (d, *J* = 5.2 Hz, 1H), 1.76 (s, 3H), 1.32 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 185.0, 176.3, 170.5, 152.3, 152.2, 146.8, 146.6, 137.4, 130.0, 129.4, 127.6, 128.4, 127.3, 118.2, 112.3, 79.5, 66.3, 63.1, 61.7, 53.0, 14.2; HRMS (ESI) m/z 437.1175 (M+H⁺), calc. for C₂₃H₂₁N₂O₅S 437.1171.

The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 30.7 min (minor) and 35.7 min (major).



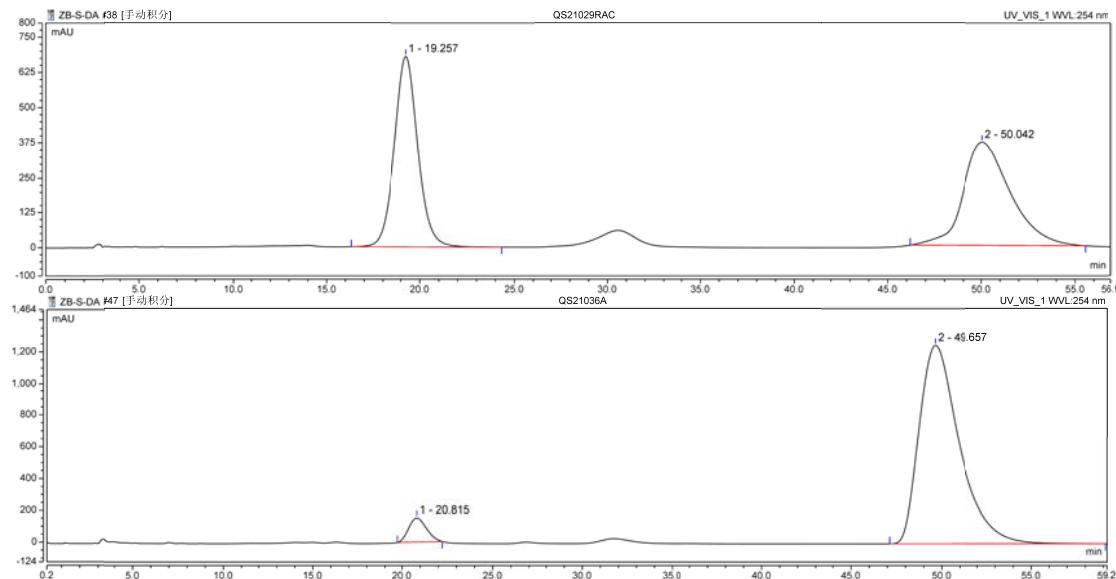
Pale yellow solid, Mp 158.4–160.1 °C; 67% yield; 90% ee; $[\alpha]_D^{28} -205.7$ (*c* 1.50, CHCl₃);

8g: Ar² = Ph

¹H NMR (300 MHz, CDCl₃) δ 8.21 (d, *J* = 8.5 Hz, 1H), 7.83 (d, *J* = 8.5 Hz, 1H), 7.78–7.70 (m, 4H), 7.64–7.49 (m, 5H), 7.43–7.35 (m, 4H), 7.28 (s, 1H), 7.23 (s, 1H), 5.86 (d, *J* = 5.4 Hz, 1H), 3.80 (d, *J* = 5.4 Hz, 1H), 3.82–3.62 (m, 2H), 0.74 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 197.7, 174.5, 170.1, 152.1,

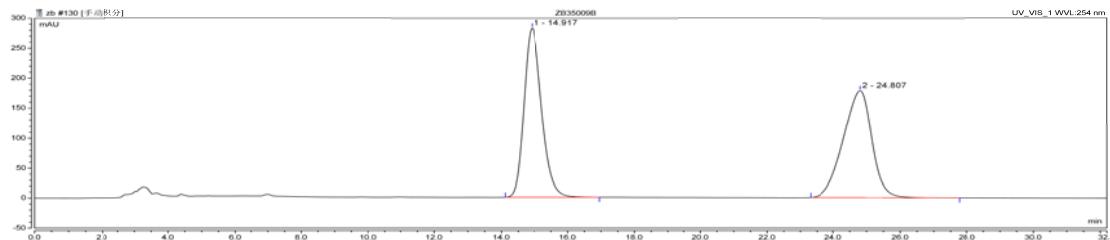
146.8, 137.7, 137.0, 133.2, 131.4, 130.0, 129.3, 128.7, 128.6, 128.4, 128.2, 128.0, 127.6, 127.4, 118.3, 78.4, 74.3, 62.5, 61.3, 54.7, 13.4; HRMS (ESI) m/z 509.1538 ($M+H^+$), calc. for $C_{30}H_{25}N_2O_4S$ 509.1535.

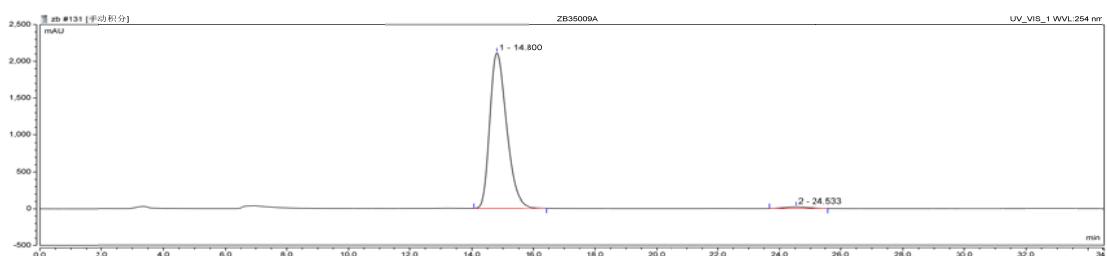
The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 60/40; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 19.3 min (minor) and 50.0 min (major).



8h White solid, Mp 125.5–126.8 °C; 95% yield; 97% *ee*; $[\alpha]_D^{26} -134.7$ (*c* 3.72, $CHCl_3$); 1H NMR (300 MHz, $CDCl_3$) δ 8.80 (d, *J* = 2.0 Hz, 1H), 8.54–8.52 (m, 1H), 7.95 (s, 1H), 7.88–7.84 (m, 1H), 7.68 (d, *J* = 7.4 Hz, 2H), 7.50 (t, *J* = 7.4 Hz, 1H), 7.35–7.24 (m, 3H), 7.24 (s, 0H), 5.43 (d, *J* = 5.4 Hz, 1H), 4.30 (q, *J* = 7.1 Hz, 2H), 3.22 (d, *J* = 5.4 Hz, 1H), 1.68 (s, 3H), 1.28 (t, *J* = 7.1 Hz, 3H); ^{13}C NMR (75 MHz, $CDCl_3$) δ 197.7, 177.3, 170.5, 150.1, 147.2, 136.1, 134.0, 133.9, 130.2, 128.8, 128.2, 123.5, 76.7, 65.4, 62.0, 54.0, 14.2; HRMS (ESI) m/z 397.1221 ($M+H^+$), calc. for $C_{21}H_{21}N_2O_4S$ 397.1222.

The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 14.8 min (major) and 24.5 min (minor).

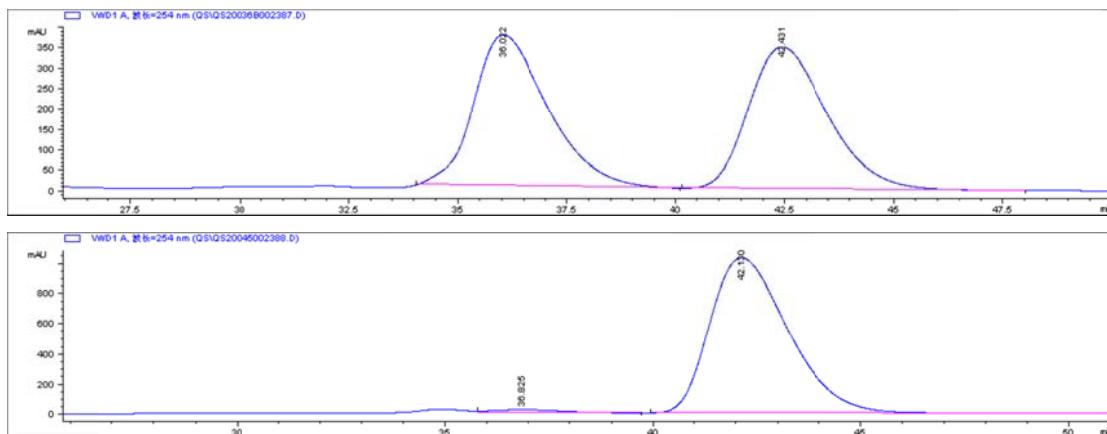




10a

Pale yellow solid, Mp 102.2–103.8 °C; 96% yield; 95% *ee*; $[\alpha]_D^{24} -348.8$ (*c* 0.05, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.95–7.91 (m, 3H), 7.79 (d, *J* = 8.5 Hz, 1H), 7.74–7.68 (m, 2H), 7.58–7.52 (m, 1H), 7.05–6.88 (m, 4H), 6.84–6.75 (m, 4H), 6.25–6.23 (m, 1H), 4.74 (s, 2H), 3.76 (s, 1H), 3.74–3.51 (m, 2H), 1.98 (s, 3H), 0.53 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.2, 174.2, 167.9, 150.4, 146.6, 143.1, 136.2, 135.0, 130.2, 129.6, 129.4, 128.4, 127.5, 127.4, 127.3, 126.5, 126.4, 126.0, 122.0, 118.9, 108.7, 82.7, 67.8, 63.0, 60.6, 60.2, 44.0, 13.3, 13.1; HRMS (ESI) m/z 550.1803 (M+H⁺), calc. for C₃₂H₂₈N₃O₄S 550.1801.

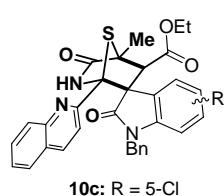
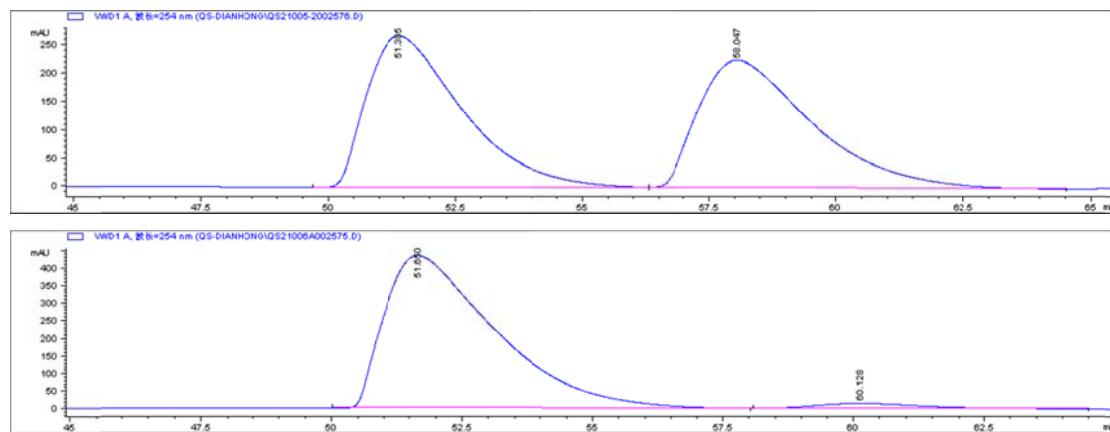
The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 36.8 min (minor) and 42.1 min (major).



10b: R = 5-F

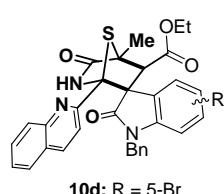
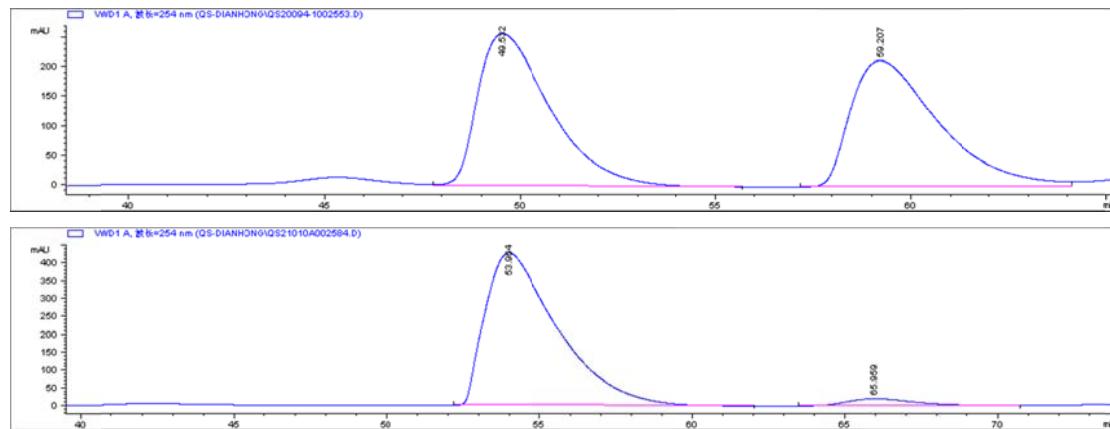
Pale yellow solid, Mp 130.4–132.2 °C; 98% yield; 94% *ee*; $[\alpha]_D^{24} -138.0$ (*c* 0.05, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.89 (t, *J* = 9.0 Hz, 3H), 7.77–7.69 (m, 3H), 7.59–7.54 (m, 1H), 7.06 (t, *J* = 7.2 Hz, 1H), 6.98 (d, *J* = 8.6 Hz, 1H), 6.88–6.78 (m, 4H), 6.69 (td, *J* = 8.6, 3.5 Hz, 1H), 6.16 (dd, *J* = 8.6, 3.5 Hz, 1H), 4.82 (d, *J* = 15.8 Hz, 1H), 4.69 (d, *J* = 15.8 Hz, 1H), 3.78–3.60 (m, 3H), 1.97 (s, 3H), 0.59 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.1, 174.1, 167.6, 160.0, 156.8, 150.2, 146.6, 139.1, 139.0, 136.6, 134.7, 130.3, 129.5, 128.4, 127.8, 127.7, 127.6, 127.5, 127.4, 126.5, 118.7, 115.7, 115.4, 114.7, 114.3, 109.2, 109.0, 82.8, 67.8, 62.9, 60.8, 60.2, 44.2, 13.2, 13.1; HRMS (ESI) m/z 590.1529 (M+Na⁺), calc. for C₃₂H₂₆FN₃O₄SnA 590.1526.

The *ee* was determined by HPLC analysis. CHIRALPAK ID-3 (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 51.7 min (major) and 60.1 min (minor).



Pale yellow solid, Mp 155.0–156.3 °C; 97% yield; 92% ee; $[\alpha]_D^{24} -140.5$ (c 0.05, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.91–7.85 (m, 4H), 7.72 (t, J = 7.4 Hz, 2H), 7.59–7.54 (m, 1H), 7.06 (t, J = 7.4 Hz, 1H), 7.00–6.94 (m, 2H), 6.88–6.78 (m, 4H), 6.17 (d, J = 8.3 Hz, 1H), 4.82 (d, J = 15.8 Hz, 1H), 4.69 (d, J = 15.8 Hz, 1H), 3.82 – 3.59 (m, 3H), 1.97 (s, 3H), 0.61 (t, J = 7.1 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.1, 173.9, 167.6, 150.2, 146.6, 141.6, 136.7, 134.6, 130.4, 129.5, 129.2, 128.5, 127.9, 127.6, 127.5, 126.5, 118.8, 109.6, 82.8, 67.6, 63.0, 60.9, 60.3, 44.1, 13.3, 13.1; HRMS (ESI) m/z 584.1415 (M+H⁺), calc. for C₃₂H₂₇ClN₃O₄S 584.1411.

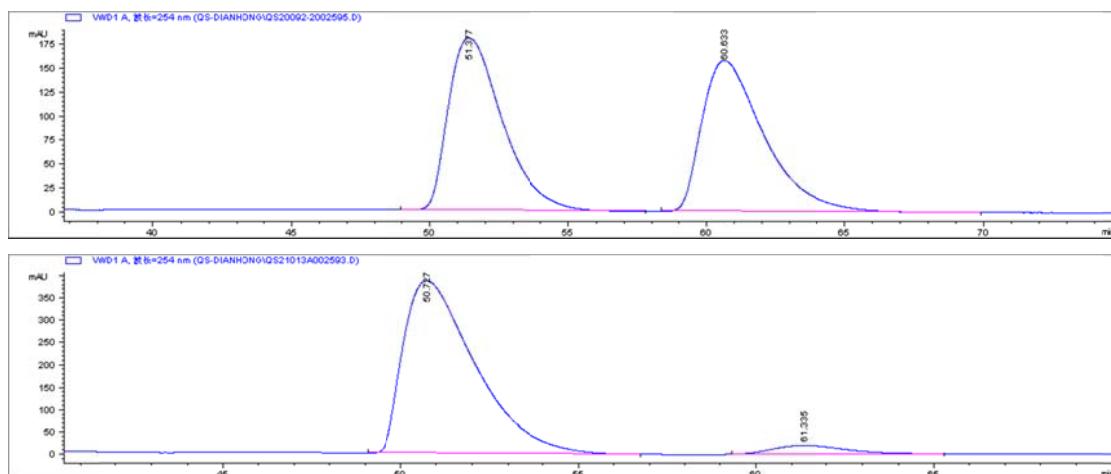
The ee was determined by HPLC analysis. CHIRALPAK ID-3 (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 54.0 min (major) and 66.0 min (minor).



Pale yellow solid, Mp 145.3–146.3 °C; 98% yield; 90% ee; $[\alpha]_D^{24} -86.1$ (c 0.05, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.03 (d, J = 2.6 Hz, 1H), 7.91 (d, J = 2.6 Hz, 1H), 7.88 (s, 2H), 7.71 (t, J = 7.4 Hz, 2H), 7.56 (t, J = 7.4 Hz, 1H), 7.12–7.03 (m, 2H), 6.98 (d, J = 8.4 Hz, 1H), 6.85 (t, J = 7.6 Hz, 2H), 6.78 (d, J = 7.6 Hz, 2H), 6.13 (d, J = 8.4 Hz, 1H), 4.81 (d, J = 15.9 Hz, 1H), 4.68 (d, J = 15.9 Hz, 1H), 3.76–3.66 (m, 3H), 1.97 (s, 3H), 0.60 (t, J = 7.1 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.1, 173.8, 167.5, 150.1, 146.6, 142.1, 136.7, 134.5, 132.1, 130.3, 129.4, 129.2, 128.4, 128.2, 127.5, 127.4, 126.5, 118.8, 114.7, 110.0,

67.5, 62.9, 60.9, 60.2, 44.1, 13.3, 13.1; HRMS (ESI) m/z 650.0720 ($M+Na^+$), calc. for $C_{32}H_{26}BrN_3O_4SNa$ 650.0725.

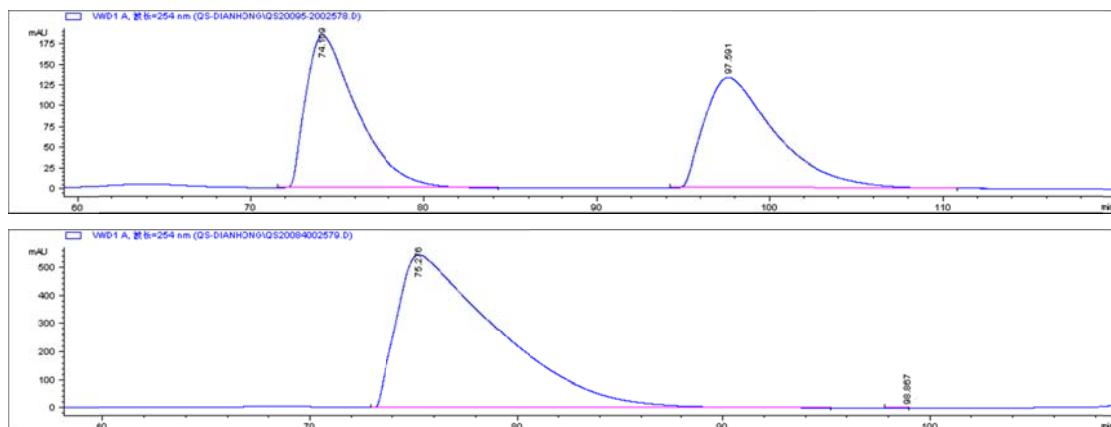
The *ee* was determined by HPLC analysis. CHIRALPAK ID-3 (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 50.7 min (minor) and 61.3 min (major).

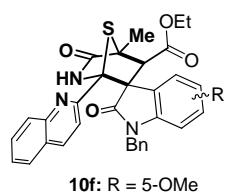


10e: R = 5-Me

 Pale yellow solid, Mp 137.5–138.5 °C; 95% yield; >99% *ee*; $[\alpha]_D^{24} -431.3$ (*c* 0.05, $CHCl_3$); 1H NMR (300 MHz, $CDCl_3$) δ 7.99 (s, 1H), 7.93 (d, J = 8.4 Hz, 1H), 7.81 (d, J = 8.4 Hz, 1H), 7.73–7.68 (m, 3H), 7.57–7.52 (m, 1H), 7.01 (t, J = 7.8 Hz, 1H), 6.89 (d, J = 7.8 Hz, 1H), 6.80–6.71 (m, 5H), 6.12 (d, J = 7.8 Hz, 1H), 4.75 (d, J = 16.0 Hz, 1H), 4.69 (d, J = 16.0 Hz, 1H), 3.75 (s, 1H), 3.73–3.54 (m, 2H), 2.29 (s, 3H), 1.98 (s, 3H), 0.53 (t, J = 7.1 Hz, 3H); ^{13}C NMR (75 MHz, $CDCl_3$) δ 176.3, 174.2, 168.0, 150.5, 146.6, 140.7, 136.2, 135.1, 131.5, 130.2, 129.6, 128.3, 127.5, 127.4, 127.2, 127.1, 126.5, 126.0, 118.9, 108.5, 82.7, 67.9, 62.9, 60.7, 60.1, 44.0, 21.2, 13.2; HRMS (ESI) m/z 586.1780 ($M+Na^+$), calc. for $C_{33}H_{29}N_3O_4SNa$ 586.1776.

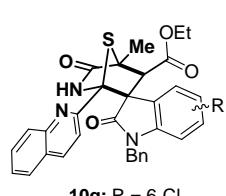
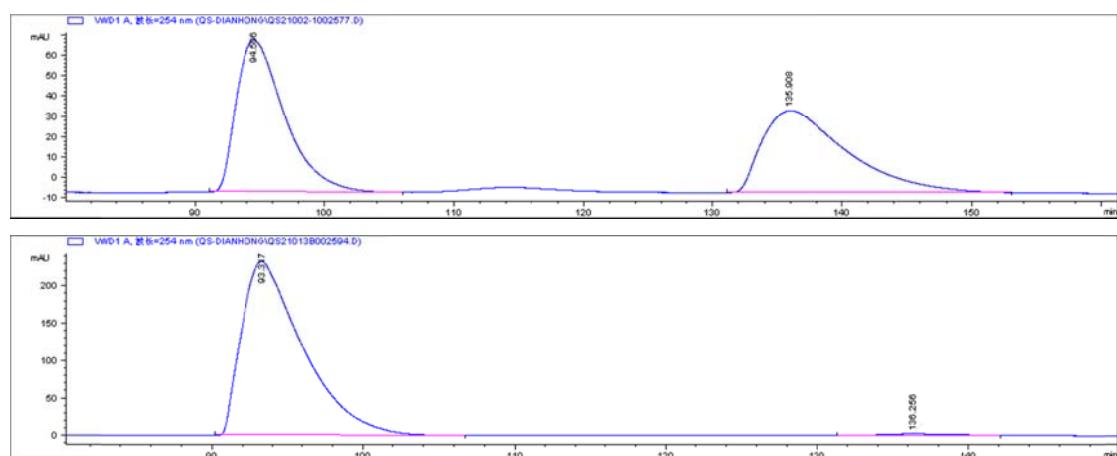
The *ee* was determined by HPLC analysis. CHIRALPAK ID-3 (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 75.3 min (major) and 98.9 min (minor).





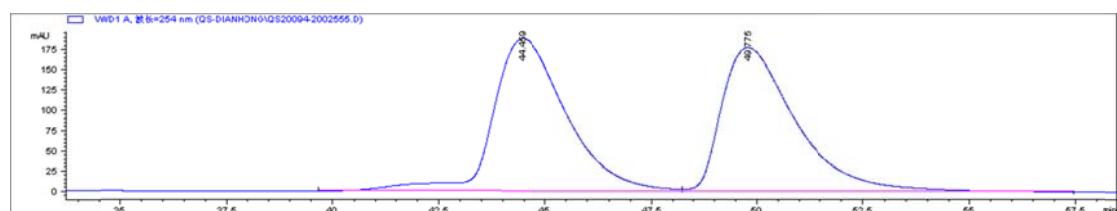
Pale yellow solid, Mp 141.1–142.3 °C; 98% yield; 98% *ee*; $[\alpha]_D^{24} -303.4$ (*c* 0.05, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.90 (s, 1H), 7.86 (d, *J* = 8.7 Hz, 1H), 7.76 (d, *J* = 8.7 Hz, 1H), 7.66–7.62 (m, 2H), 7.52–7.46 (m, 2H), 6.95 (t, *J* = 7.2 Hz, 1H), 6.87 (d, *J* = 8.5 Hz, 1H), 6.75–6.66 (m, 4H), 6.45 (dd, *J* = 8.5, 2.6 Hz, 1H), 6.06 (d, *J* = 8.5 Hz, 1H), 4.65 (s, 2H), 3.72–3.50 (m, 6H), 1.90 (s, 3H), 0.50 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.2, 173.9, 167.8, 155.2, 150.3, 146.6, 136.5, 136.3, 135.0, 130.2, 129.5, 128.3, 127.5, 127.4, 127.2, 127.1, 126.5, 118.7, 114.2, 113.5, 109.0, 82.7, 68.0, 62.9, 60.6, 60.0, 55.9, 44.0, 13.3, 13.1; HRMS (ESI) m/z 602.1724 (M+Na⁺), calc. for C₃₃H₂₉N₃O₅SNa 602.1726.

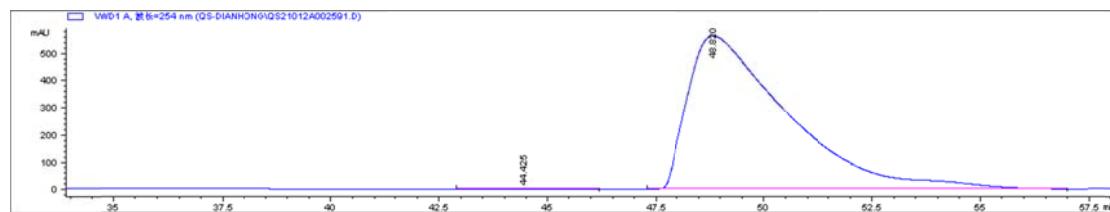
The *ee* was determined by HPLC analysis. CHIRALPAK ID-3 (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 93.3 min (major) and 136.3 min (minor).



Pale yellow solid, Mp 96.4–96.9 °C; 97% yield; >99% *ee*; $[\alpha]_D^{24} -133.7$ (*c* 0.05, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.85–7.75 (m, 4H), 7.64 (t, *J* = 7.4 Hz, 2H), 7.52–7.46 (m, 1H), 7.00 (t, *J* = 7.4 Hz, 1H), 6.88–6.71 (m, 6H), 6.19 (s, 1H), 4.73 (d, *J* = 15.9 Hz, 1H), 4.61 (d, *J* = 15.9 Hz, 1H), 3.70–3.41 (m, 3H), 1.88 (s, 3H), 0.53 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.1, 174.2, 167.6, 150.2, 146.6, 144.2, 136.6, 135.2, 134.4, 130.3, 129.4, 128.5, 127.6, 127.5, 127.4, 127.3, 126.4, 124.4, 121.8, 118.7, 109.1, 82.7, 67.2, 62.8, 60.8, 60.3, 44.1, 13.3, 13.0; HRMS (ESI) m/z 584.1414 (M+H⁺), calc. for C₃₂H₂₇ClN₃O₄S 584.1411.

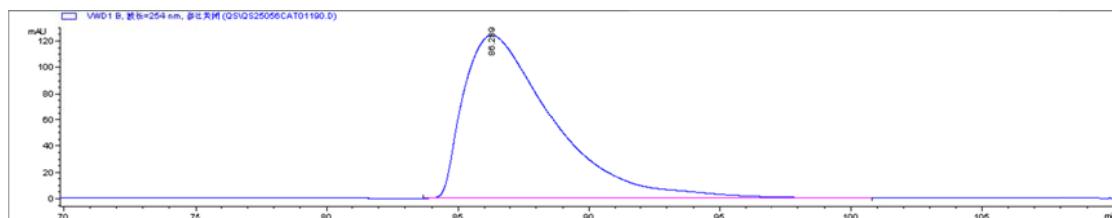
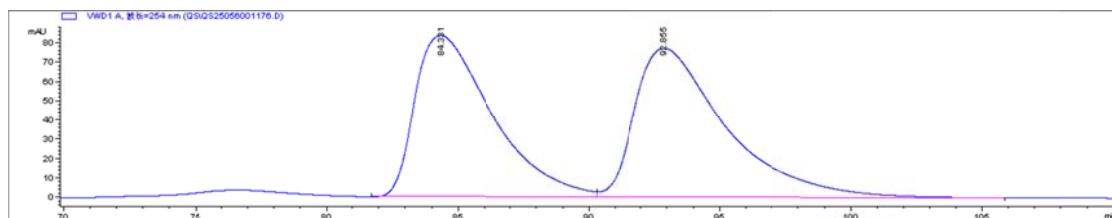
The *ee* was determined by HPLC analysis. CHIRALPAK ID-3 (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 44.4 min (minor) and 48.8 min (major).





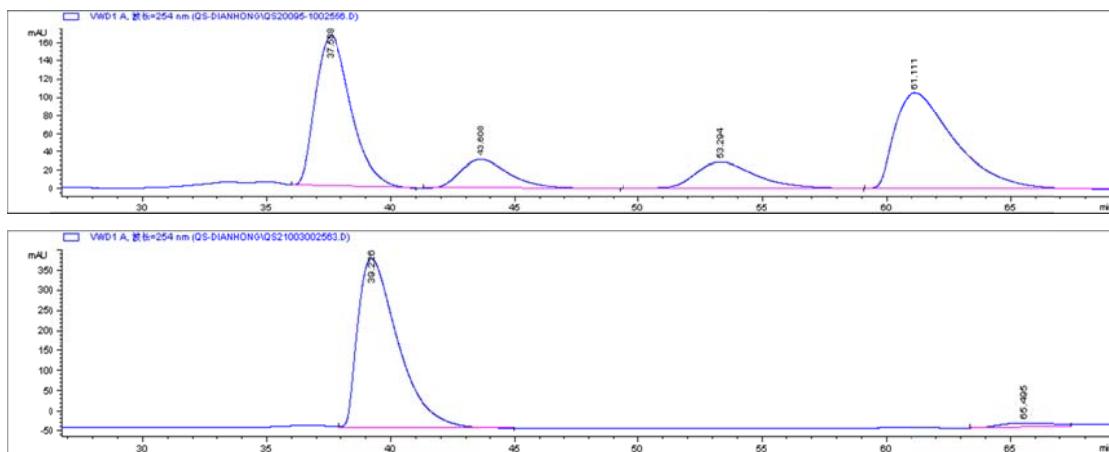
Pale yellow solid, Mp 123.0–124.8 °C; 94% yield; >99% ee; $[\alpha]_D^{24} -269.5$ (c 0.05, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.92–7.86 (m, 2H), 7.75 (dd, J = 8.5, 1.6 Hz, 2H), 7.67–7.62 (m, 2H), 7.52–7.46 (m, 1H), 6.94 (t, J = 7.1 Hz, 1H), 6.84 (d, J = 8.5 Hz, 1H), 6.73–6.63 (m, 4H), 6.38 (dd, J = 8.5, 2.3 Hz, 1H), 5.75 (d, J = 2.3 Hz, 1H), 4.68 (d, J = 15.9 Hz, 1H), 4.59 (d, J = 15.9 Hz, 1H), 3.70–3.48 (m, 6H), 1.89 (s, 3H), 0.54 (t, J = 7.2 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.2, 174.6, 168.0, 160.7, 150.5, 146.6, 144.3, 136.3, 134.9, 130.2, 129.6, 128.3, 127.5, 127.4, 127.2, 126.5, 118.9, 117.7, 105.4, 96.7, 82.6, 67.5, 62.8, 60.6, 60.1, 55.2, 44.0, 13.4, 13.1; HRMS (ESI) m/z 602.1730 (M+Na⁺), calc. for C₃₃H₂₉N₃O₅SNa 602.1726.

The ee was determined by HPLC analysis. CHIRALPAK ID-3 (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 101.9 min (minor) and 110.0 min (major).



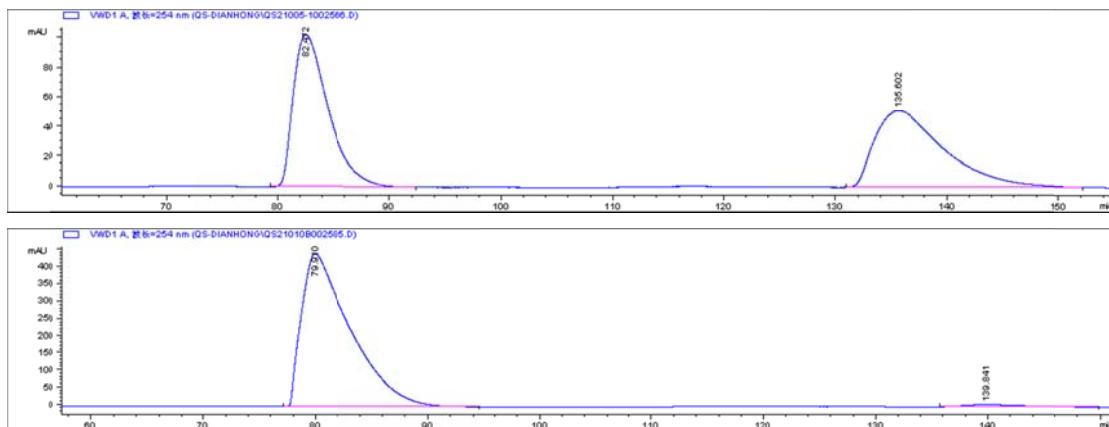
Pale yellow solid, Mp 70.4–72.1 °C; 96% yield; 93% ee; $[\alpha]_D^{24} -240.4$ (c 0.05, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.88–7.83 (m, 3H), 7.75–7.68 (m, 3H), 7.57–7.52 (m, 1H), 7.11–7.05 (m, 1H), 7.00–6.75 (m, 7H), 5.06 (d, J = 15.5 Hz, 1H), 4.78 (d, J = 15.5 Hz, 1H), 3.75 (s, 1H), 3.72–3.51 (m, 2H), 1.96 (s, 3H), 0.55 (t, J = 7.2 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.2, 174.2, 167.6, 150.1, 146.6, 136.5, 136.4, 130.3, 129.5, 128.2, 127.5, 127.4, 127.2, 126.7, 122.5, 122.4, 122.3, 122.2, 118.7, 117.6, 117.3, 82.9, 67.6, 62.8, 60.7, 60.6, 45.5, 13.2, 13.1; HRMS (ESI) m/z 606.1233 (M+Na⁺), calc. for C₃₂H₂₆ClN₃O₄SNa 606.1230.

The ee was determined by HPLC analysis. CHIRALPAK ID-3 (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 39.2 min (major) and 65.5 min (minor).



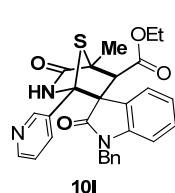
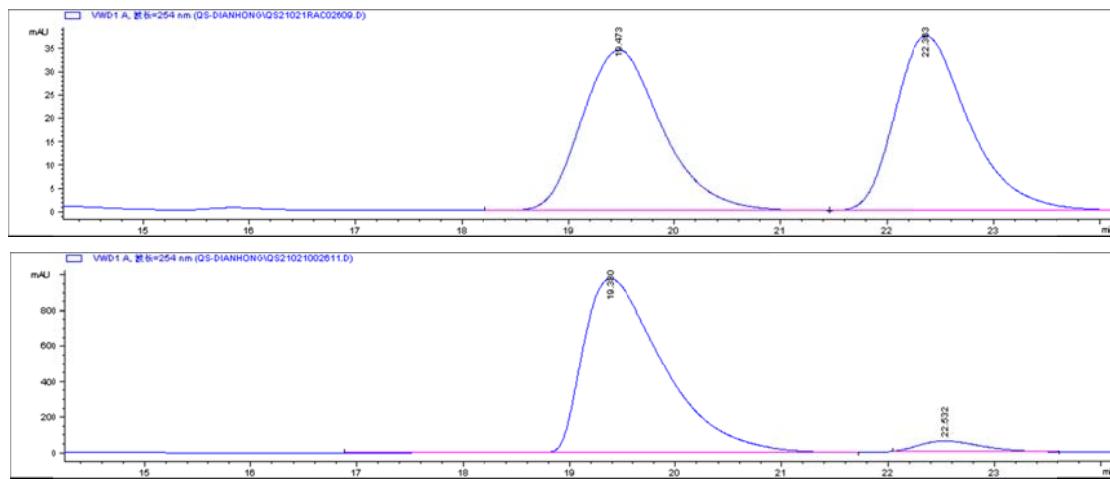
Pale yellow solid, Mp 87.0–88.6 °C; 97% yield; 96% ee; $[\alpha]_D^{24} -281.9$ (*c* 0.05, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.90–7.87 (m, 2H), 7.82–7.75 (m, 2H), 7.67–7.63 (m, 2H), 7.52–7.46 (m, 1H), 6.98 (t, *J* = 7.4 Hz, 1H), 6.86–6.77 (m, 4H), 6.71 (d, *J* = 7.4 Hz, 1H), 6.62 (d, *J* = 7.4 Hz, 2H), 5.04 (d, *J* = 17.0 Hz, 1H), 4.84 (d, *J* = 17.0 Hz, 1H), 3.74–3.52 (m, 3H), 1.91 (s, 3H), 1.79 (s, 3H), 0.59 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.3, 175.3, 168.0, 150.5, 146.6, 141.2, 137.1, 136.2, 133.4, 130.2, 129.6, 128.5, 127.5, 126.9, 126.6, 125.2, 124.4, 122.0, 119.1, 119.0, 83.0, 67.2, 63.0, 60.7, 60.6, 45.3, 18.6, 13.5, 13.2; HRMS (ESI) m/z 586.1778 (M+Na⁺), calc. for C₃₃H₂₉N₃O₄SNa 586.1776.

The ee was determined by HPLC analysis. CHIRALPAK ID-3 (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 79.9 min (major) and 139.8 min (minor).



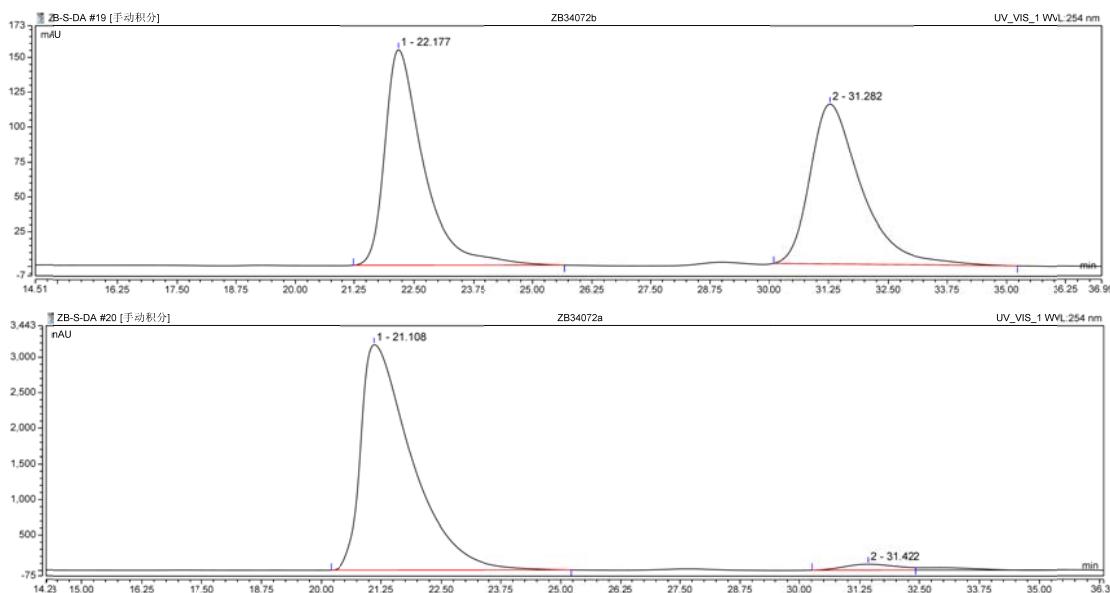
Pale yellow solid, Mp 87.8–89.8 °C; 94% yield; 90% ee; $[\alpha]_D^{24} -72.5$ (*c* 0.05, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.82 (d, *J* = 7.3 Hz, 1H), 7.61 (s, 1H), 7.28–7.25 (m, 3H), 7.08–7.04 (m, 3H), 6.99–6.88 (m, 5H), 6.46 (d, *J* = 7.3 Hz, 1H), 5.03 (d, *J* = 15.7 Hz, 1H), 4.67 (d, *J* = 15.7 Hz, 1H), 3.68–3.45 (m, 3H), 1.92 (s, 3H), 0.47 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 177.7, 174.8, 167.6, 142.5, 135.0, 134.9, 129.7, 129.4, 128.7, 128.3, 127.8, 127.3, 127.1, 126.6, 126.0, 122.4, 108.8, 82.6, 68.0, 61.3, 60.7, 60.4, 44.0, 13.2, 12.9; HRMS (ESI) m/z 555.1123 (M+Na⁺), calc. for C₂₉H₂₅ClN₂O₄SNa 555.1121.

The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 19.4 min (major) and 22.5 min (minor).



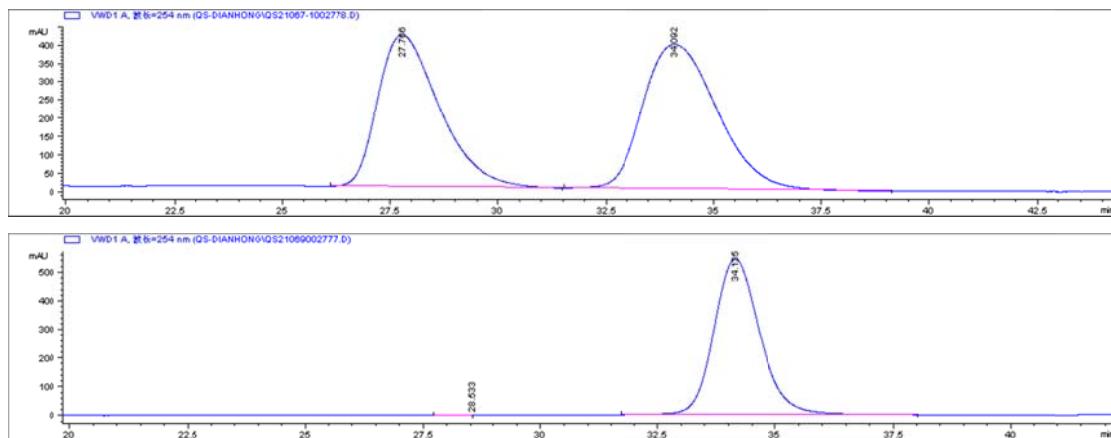
Pale yellow solid, Mp 101.3–102.6 °C; 96% yield; 95% *ee*; $[\alpha]_D^{24} -62.0$ (*c* 0.05, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.25 (d, *J* = 4.2 Hz, 1H), 7.81–7.78 (m, 1H), 7.59 (s, 1H), 7.33 (td, *J* = 7.7, 1.7 Hz, 1H), 7.26–7.22 (m, 3H), 7.12–6.98 (m, 4H), 6.92–6.82 (m, 2H), 6.39 (d, *J* = 7.7 Hz, 1H), 4.89 (d, *J* = 15.7 Hz, 1H), 4.72 (d, *J* = 15.7 Hz, 1H), 3.71–3.49 (m, 3H), 1.94 (s, 3H), 0.50 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.6, 174.4, 167.8, 150.4, 148.5, 142.9, 136.0, 135.3, 129.2, 128.5, 127.5, 127.2, 126.1, 126.0, 123.6, 122.0, 121.6, 108.4, 82.8, 67.3, 62.4, 60.6, 60.3, 44.0, 13.2, 13.1; HRMS (ESI) m/z 522.1466 (M+Na⁺), calc. for C₂₈H₂₅N₃O₄SNa 522.1463.

The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 21.1 min (major) and 31.4 min (minor).



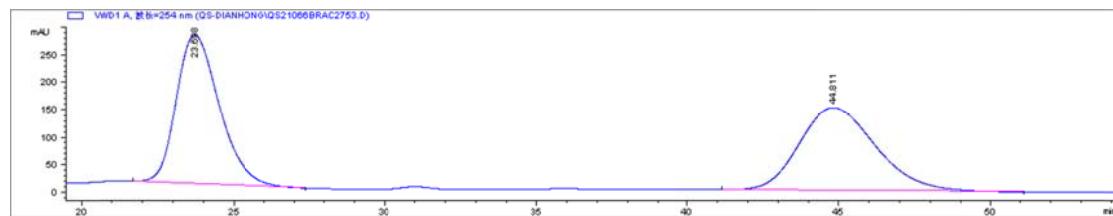
10m Pale yellow solid, Mp 92.3–94.1 °C; 87% yield; >99% ee; $[\alpha]_D^{24} -67.8$ (*c* 0.05, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.93–7.91 (m, 3H), 7.80 (d, *J* = 7.8 Hz, 1H), 7.79–7.68 (m, 2H), 7.54 (t, *J* = 7.8 Hz, 1H), 7.06–7.04 (m, 1H), 6.98–6.91 (m, 3H), 6.86–6.78 (m, 4H), 6.25–6.22 (m, 1H), 4.79 (d, *J* = 15.8 Hz, 1H), 4.71 (d, *J* = 15.8 Hz, 1H), 3.79 (s, 1H), 3.72–3.49 (m, 2H), 2.60 (dq, *J* = 14.5, 7.2 Hz, 1H), 2.40 (dq, *J* = 14.5, 7.2 Hz, 1H), 1.27 (t, *J* = 7.2 Hz, 3H), 0.52 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 175.5, 174.3, 167.9, 150.6, 146.6, 143.0, 136.2, 135.0, 130.2, 129.5, 129.3, 128.4, 127.5, 127.4, 127.3, 126.6, 125.9, 121.9, 118.9, 108.6, 81.9, 69.4, 67.5, 60.6, 59.4, 44.0, 20.4, 13.2, 12.0; HRMS (ESI) m/z 586.1775 (M+Na⁺), calc. for C₃₃H₂₉N₃O₄SNa 586.1776.

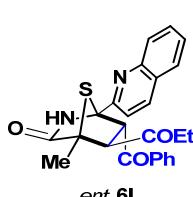
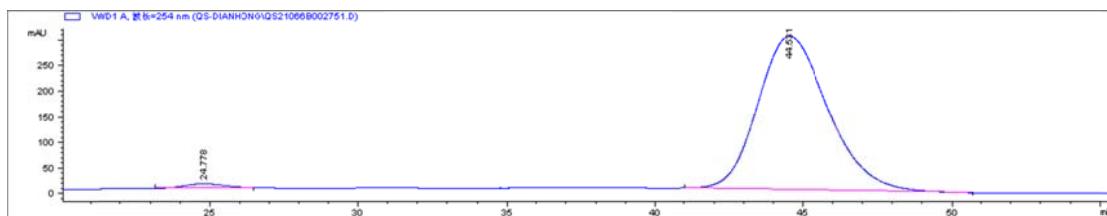
The ee was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 28.5 min (minor) and 3401 min (major).



10n Pale yellow solid, Mp 108.2–109.9 °C; 84% yield; 97% ee; $[\alpha]_D^{24} -101.0$ (*c* 0.05, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.98–7.89 (m, 3H), 7.73–7.64 (m, 3H), 7.58–7.48 (m, 3H), 7.37–7.30 (m, 3H), 7.24–7.22 (m, 1H), 7.06–6.92 (m, 3H), 6.86–6.78 (m, 5H), 6.24–6.21 (m, 1H), 4.79 (d, *J* = 15.9 Hz, 1H), 4.72 (d, *J* = 15.9 Hz, 1H), 3.85 (s, 1H), 3.81 (s, 1H), 3.76–3.53 (m, 2H), 0.54 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 175.1, 174.1, 168.1, 150.4, 146.5, 143.0, 139.1, 136.0, 135.0, 130.1, 130.0, 129.5, 129.4, 128.4, 128.3, 127.5, 127.3, 126.9, 126.6, 125.8, 122.0, 118.9, 108.7, 81.6, 70.7, 68.1, 60.7, 59.5, 44.0, 31.9, 13.3; HRMS (ESI) m/z 648.1932 (M+Na⁺), calc. for C₃₈H₃₁N₃O₄SNa 648.1933.

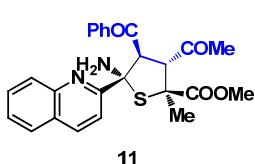
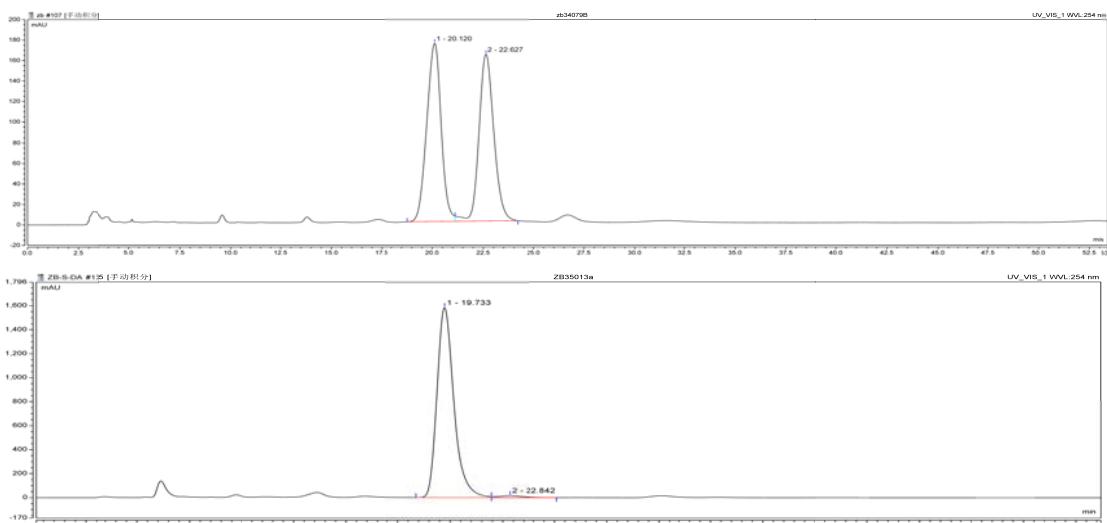
The ee was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 24.8 min (minor) and 44.5 min (major).





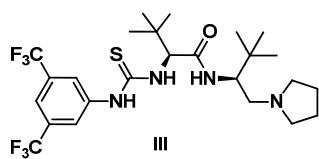
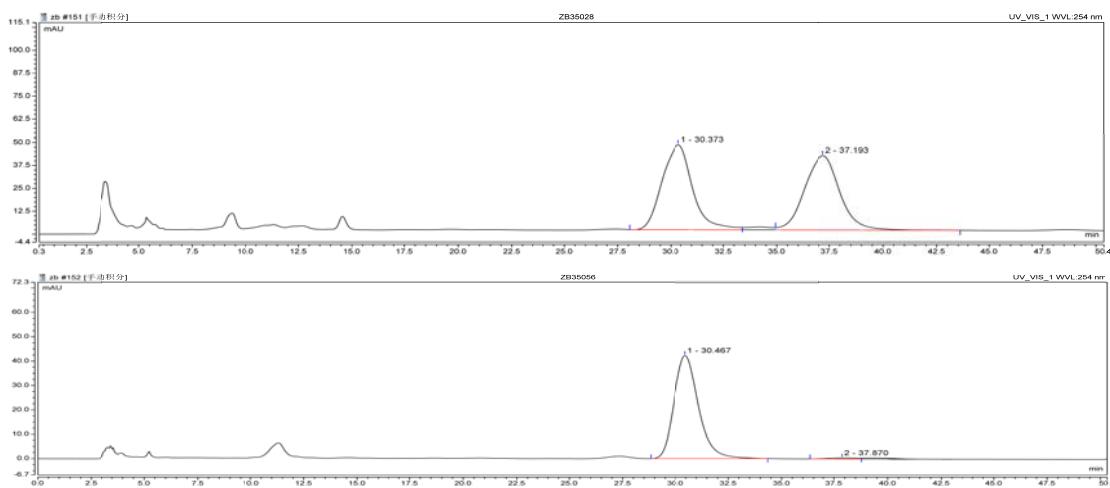
White solid, Mp 128.6–129.8 °C; 91% yield; 97% *ee*; $[\alpha]_D^{26} +183.4$ (*c* 3.44, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.13 (d, *J* = 8.5 Hz, 1H), 7.73–7.69 (m, 3H), 7.65–7.56 (m, 4H), 7.48 (t, *J* = 7.3 Hz, 1H), 7.35 (t, *J* = 7.3 Hz, 1H), 7.22 (d, *J* = 7.3 Hz, 1H), 5.70 (d, *J* = 5.5 Hz, 1H), 3.50 (d, *J* = 5.5 Hz, 1H), 2.66 (dq, *J* = 18.2, 7.2 Hz, 1H), 2.29 (dq, *J* = 18.2, 7.2 Hz, 1H), 1.72 (s, 3H), 1.04 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 207.9, 198.3, 176.6, 152.2, 146.8, 137.5, 137.0, 133.1, 129.9, 129.3, 128.4, 128.0, 127.6, 127.3, 127.2, 118.2, 79.6, 66.8, 62.3, 57.4, 39.1, 14.2, 7.6; HRMS (ESI) m/z 431.1430 (M+H⁺), calc. for C₂₅H₂₃N₂O₃S 431.1429.

The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 19.7 min (major) and 22.8 min (minor).



Green solid, Mp 121.9–123.0 °C; 73% yield; 98% *ee*; $[\alpha]_D^{26} +161.4$ (*c* 0.94, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.06 (d, *J* = 8.7 Hz, 1H), 7.92 (d, *J* = 8.7 Hz, 1H), 7.71 (d, *J* = 8.4 Hz, 2H), 7.58 (ddd, *J* = 8.4, 6.3, 1.4 Hz, 1H), 7.46 (ddd, *J* = 8.4, 6.3, 1.4 Hz, 3H), 6.99 (d, *J* = 7.5 Hz, 1H), 6.86 (t, *J* = 7.5 Hz, 2H), 5.42 (d, *J* = 12.5 Hz, 1H), 5.25 (d, *J* = 12.5 Hz, 1H), 3.91 (s, 3H), 2.28 (s, 3H), 1.75 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 205.5, 198.3, 174.0, 158.5, 146.1, 137.5, 136.6, 131.9, 129.5, 129.2, 128.1, 127.4, 127.0, 126.8, 119.3, 77.3, 62.7, 61.9, 55.0, 53.5, 30.9, 23.7; HRMS (ESI) m/z 449.1539 (M+H⁺), calc. for C₂₅H₂₅N₂O₄S 449.1535.

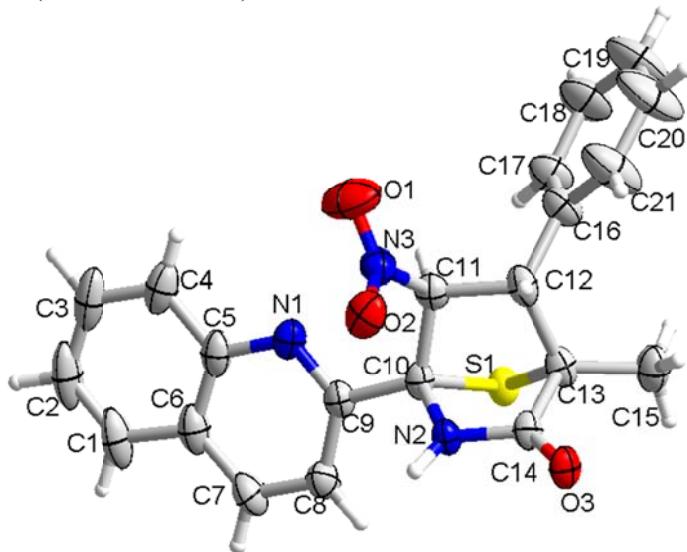
The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 30.5 min (major) and 37.9 min (minor).



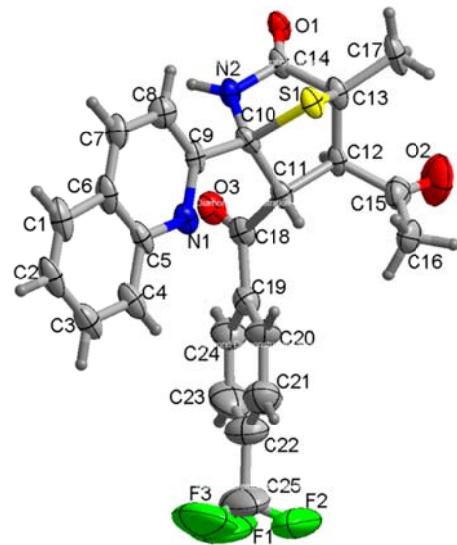
White solid, ^1H NMR (300 MHz, CDCl_3) δ 7.98 (s, 2H), 7.30 (s, 1H), 4.22 (s, 2H), 2.73 (s, 6H), 1.89 (s, 4H), 1.17 (s, 9H), 1.00 (s, 9H); ^{13}C NMR (75 MHz, CDCl_3) δ 182.0, 172.7, 140.2, 131.6, 131.2, 130.9, 130.7, 130.3, 128.8, 128.4, 124.8, 123.9, 121.2, 117.8, 67.0, 65.6, 55.7, 54.0, 34.7, 33.8, 27.3, 26.6, 23.4; HRMS (ESI) m/z 555.2593 ($\text{M}+\text{H}^+$), calc. for $\text{C}_{25}\text{H}_{37}\text{F}_6\text{N}_4\text{OS}$ 555.2592.

14. Determination of the absolute configuration

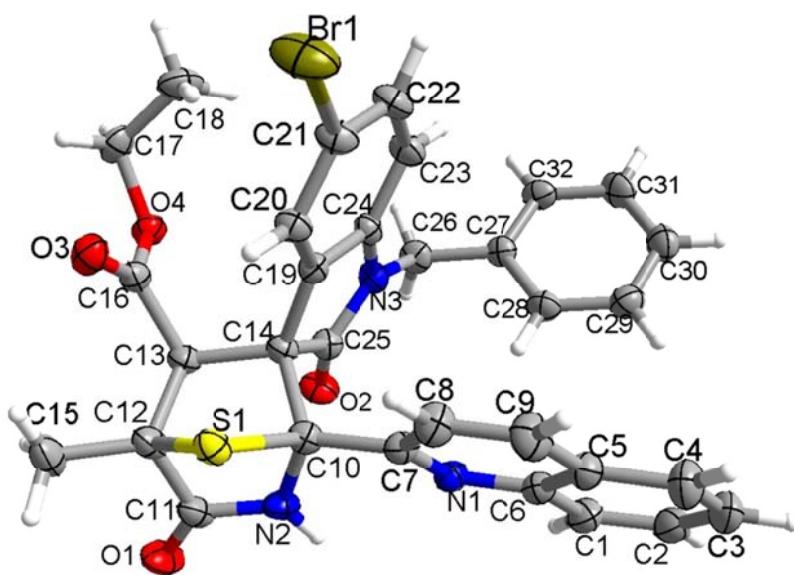
(1) Absolute configurations of the [4+2] annulation adducts **3a-n** are determined by *X*-ray structure analysis of the product **3a** (CCDC 1419583)



(2) Absolute configurations of the [4+2] annulation adducts **6a-s** and **9a-h** are determined by *X*-ray structure analysis of the product **6b** (CCDC 1419574)



(3) Absolute configurations of the [4+2] annulation adducts **11a-n** are determined by *X*-ray structure analysis of the product **11d** (CCDC 1463891)



15. Computational methods

All quantum chemical calculations were carried out with Gaussian 09¹. All ΔG values are reported relative to the individual free starting materials **1**, **2** and catalyst **III**.

Optimization of minimum and transition state (TS) structures were first carried out with Becke's three parameter² and Lee-Yang-Parr's³ B3LYP density functional using Pople's⁴ 6-31G(d,p) basis set under the SMD⁵ polarizable continuum solvent model (toluene parameters). It is important to note that Simón and Goodman⁶ showed that B3LYP/6-31G(d) is adequate to model a variety of organocatalytic reactions. Subsequently, Houk *et al*⁷ applied the dispersion-corrected meta-hybrid M06-2X(D3) approach towards modelling stereoselective intramolecular Aldol reaction and thiourea-amine catalyzed Nazarov cyclization with excellent qualitative agreement to experiment.

The B3LYP/6-31G(d,p) thermal and vibrational corrections from frequency calculations were combined to higher level single-point energies of the optimized geometries. The single-point energies were evaluated with Peverati and Truhlar's screened-exchange density functional MN12-SX under same polarizable continuum solvent model toluene parameters, herein termed MN12-SX/SMD.⁸ The newer meta-hybrid non separable gradient MN12-SX functional was assessed to perform better in chemical energetics and thus preferred over M06-2X.⁹ In conjunction with the MN12-SX/SMD, the Wiegend and Aldrich's def2-TZVPP triple zeta quality basis set was used.¹⁰ Second order derivative or Hessian of the completed calculations were checked to verify transition state structures having only one negative eigenvalue and none for minimum.

Gibbs free energies in solution were calculated from the geometries, frequencies and improved energies using a thermocycle in which the gas-phase was treated using the standard textbook formulae for an ideal gas under the harmonic oscillator / rigid rotor approximation, and the Gibbs free energies of solvation (in toluene) were then calculated using the SMD continuum solvation model. Corrections were included to consider passage of 1 atm gas into 1M in solution, $\Delta G^{\text{1atm} \rightarrow \text{1M}}$ as follows¹¹:

$$\Delta G^{\text{1atm} \rightarrow \text{1M}} = dN * R * T * \ln(RT/P)$$

where dN is the number of moles of gas change from reactant to product and $\ln(RT/P)$ equals to 1.89 at 298 K. We have recently shown that this approach provides an excellent quantitative description of the temperature dependent behavior of solution-phase Diels-Alder reactions.¹²

Electron density topological analyses based on the reduced density gradient was carried out with NCIplot¹³ to qualify regions of non-covalent interactions especially hydrogen bonding

(H-bonding) or regions of repulsive interactions. Regions which are attractive have sign of the second density Hessian eigenvalue, $sign(\lambda_2)$, as positive and repulsive as negative. The electron density $\rho(r)$ is set at an isovalue of 0.4 au in the $sign(\lambda_2)\rho(r)$ range of -0.4 to 0.4, which corresponds to blue-green-red color scale (see Figure S1). The 3-D molecular graphics for NCIplot were rendered with VMD¹⁴ and graphics inset corresponding to the NCIplot showing key H-bond interactions in figure 3 of the manuscript were rendered with Cylview¹⁵.

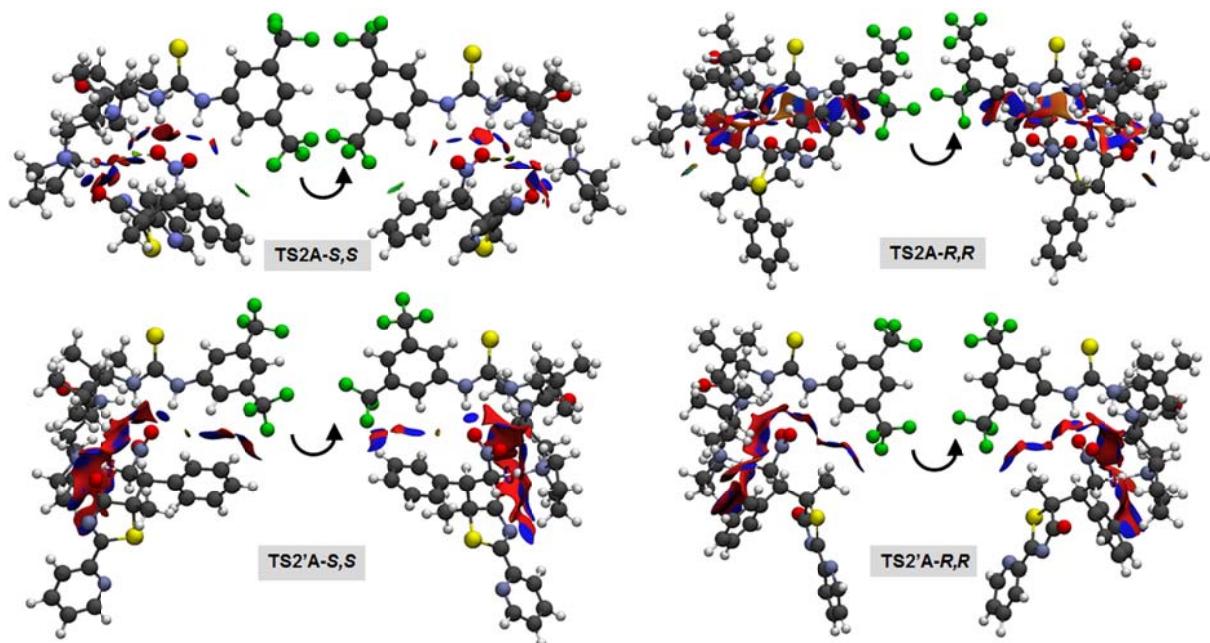


Figure S1. Non-covalent interaction isosurfaces of optimized TS structures of Mannich process (**TS2A-S,S** and **TS2A-R,R**) and the protonation process (**TS2'A-S,S** and **TS2'A-R,R**). Blue-green-red color scale from $-0.4 < sign(\lambda_2)\rho(r) > 0.4$ au, where blue (positive) is favorable and red (negative) is unfavorable interaction. Two images for each TS are flipped horizontally to allow viewing from the front and back sides.

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Molecule	MN12SX E	B3LYP E	G _{correction}	H _{correction}	ZPE (298K)	G _{sol} (MN12SX)
III	-2265.238831	-2265.933466	0.526197	0.640233	0.60057	-2264.703614
1	-930.478833	-930.724803	0.119136	0.169065	0.157119	-930.359697
2	-513.963184	-514.176050	0.101575	0.146955	0.136956	-513.861609
TS1A-R,R	-3709.692433	-3710.841985	0.796699	0.960728	0.899259	-3708.892727
TS1A-S,S	-3709.695202	-3710.844099	0.796696	0.961218	0.899614	-3708.895499
TS1B-R,R	-3709.69039	-3710.83715	0.797744	0.961166	0.899645	-3708.889639
TS1B-S,S	-3709.688422	-3710.838464	0.797171	0.961137	0.899743	-3708.888244
int1A-R,R	-3709.705159	-3710.854112	0.799168	0.96316	0.90194	-3708.902984
int1A-S,S	-3709.714753	-3710.863604	0.800902	0.963818	0.902572	-3708.910844
int1B-R,R	-3709.714366	-3710.860514	0.796381	0.961835	0.899977	-3708.914978
int1B-S,S	-3709.713598	-3710.861334	0.798688	0.962503	0.901149	-3708.911903
TS2A-R,R	-3709.699503	-3710.843566	0.804677	0.962433	0.902263	-3708.891819
TS2A-S,S	-3709.698245	-3710.843839	0.799844	0.962093	0.901234	-3708.895394
int2A-R,R	-3709.710927	-3710.853914	0.802751	0.964022	0.903051	-3708.905169
int2A-S,S	-3709.71474	-3710.857422	0.800695	0.963549	0.902543	-3708.911038
TS2'A-R,R	-3709.690718	-3710.836448	0.795499	0.957718	0.896357	-3708.892212
TS2'A-S,S	-3709.693888	-3710.837503	0.798308	0.957859	0.897082	-3708.892573
int2'A-R,R	-3709.712573	-3710.860005	0.792608	0.962243	0.899753	-3708.916958
int2'A-S,S	-3709.712922	-3710.861514	0.795597	0.962489	0.900231	-3708.914318

*All energy values in hartrees.

Gaussian Archive Files

Catalyst III

1\\1\GINC-R1102\FOpt\RB3LYP\6-31G(d, p)\C25H36F6N401S1\R00T\09-Dec-2015\\# freq=noraman rb3lyp/6-31g** opt=maxcyc=200 scf=maxcyc=200 scrf=(smd, sol vent=tol uene)\\Catalyst III\\0, 1\C, 1. 9345173079, 0. 1913285951, -1. 2675001586\\0, 2. 7222233811, -0. 0115586208, -2. 1905041561\C, 1. 0106971573, 1. 4394955675, -1. 3269008674\H, 0. 645363335, 1. 4648738374, -2. 355854527\N, 1. 8184572705, -0. 5798134111, -0. 158914149\H, 1. 0738800145, -0. 3245909597, 0. 4 718634769\C, 2. 6492454085, -1. 7284155289, 0. 2519299286\H, 2. 4689319057, -1. 8051190086, 1. 3308211293\C, 2. 1719890722, -3. 1040802263, -0. 3402051028\C, 0. 6437094855, -3. 217673919, -0. 1766577936\H, 0. 1121645481, -2. 4877299589, -0. 7921152638\H, 0. 3374482578, -3. 0747134033, 0. 8680231677\H, 0. 3043017939, -4. 2137057113, -0. 4809826574\C, 2. 5322001042, -3. 2642068952, -1. 829019438\H, 2. 1307405187, -2. 4483810422, -2. 4337829887\H, 2. 1251325727, -4. 2074545727, -2. 2112858297\H, 3. 6154797385, -3. 2896756892, -1. 9864691966\C, 2. 81628368 24, -4. 2514734696, 0. 4676217447\H, 3. 9049038954, -4. 2836959, 0. 3649586651\H, 2. 4316736996, -5. 2156645468, 0. 1178654957\H, 2. 5824557205, -4. 1737072856, 1. 5365026558\C, 4. 1490892125, -1. 4358323091, 0. 0511560015\H, 4. 7191826574, -2. 3523108328, 0. 28289469\H, 4. 3201618973, -1. 1936768557, -0. 9978704599\N, 4. 636044141, -0. 3165031866, 0. 8549308163\C, 5. 9845376224, 0. 1133510513, 0. 4 62546004\H, 5. 9653950685, 0. 5831364452, -0. 5257586169\H, 6. 6786769798, -0. 7 486247418, 0. 4079606765\C, 6. 4152538149, 1. 065952585, 1. 5896537744\H, 7. 495 9584795, 1. 0321948572, 1. 7529615673\H, 6. 15458027, 2. 098312689, 1. 340152949 1\C, 4. 7481579731, -0. 5816959913, 2. 29408829\H, 5. 2546508957, -1. 5492740071, 2. 4818874798\H, 3. 7634183122, -0. 6312522996, 2. 76944665\C, 5. 6066390903, 0. 582485208, 2. 8266273613\H, 6. 2501935688, 0. 2545677982, 3. 6478160651\H, 4. 9 736191167, 1. 3868462685, 3. 2125657352\N, -0. 1595420705, 1. 3332697755, -0. 45 22302794\H, -0. 1028967702, 1. 8337715807, 0. 4267243583\C, 1. 774772315, 2. 784 504687, -1. 0410692578\C, 2. 8993065031, 2. 9798408282, -2. 0762303819\H, 3. 382 1201824, 3. 9488390641, -1. 9080383853\H, 2. 5041851409, 2. 9749525477, -3. 0973 922727\H, 3. 657526666, 2. 1990607342, -2. 0140345912\C, 0. 7869700397, 3. 96061 87813, -1. 1910797265\H, 1. 3215996731, 4. 9099194184, -1. 081037854\H, 0. 31018 95316, 3. 9585507434, -2. 1772513457\H, -0. 0082995073, 3. 949228743, -0. 437976 7755\C, 2. 3846461119, 2. 790653866, 0. 3751985688\H, 1. 6248830384, 2. 71510565 84, 1. 1655085401\H, 2. 9159442158, 3. 7325868354, 0. 5487537325\H, 3. 094517527 2, 1. 9693082066, 0. 5168708945\C, -1. 3734823931, 0. 8308135204, -0. 8138098533 \N, -2. 3408405425, 1. 0707516051, 0. 1409349841\H, -2. 1224123162, 1. 795846689 7, 0. 8144984513\C, -3. 6722429153, 0. 6197129142, 0. 2286350453\C, -6. 34628870 77, -0. 1817627483, 0. 5763075794\C, -4. 5914077565, 1. 4627833248, 0. 868164970 7\C, -4. 0936174068, -0. 6367994761, -0. 2249863883\C, -5. 4239186959, -1. 01734 21487, -0. 0527575816\C, -5. 9129009696, 1. 0586266636, 1. 0424324835\H, -4. 277 5860816, 2. 441848976, 1. 2154116223\H, -3. 3966674364, -1. 299884231, -0. 71585 38118\H, -7. 3795857217, -0. 4857340693, 0. 6915724522\C, -5. 8582040808, -2. 39 26746795, -0. 4905093553\C, -6. 8803771074, 1. 9477472162, 1. 7772181303\F, -5. 1116366962, -2. 8626108081, -1. 5100467778\F, -5. 7534041112, -3. 2895656308, 0. 5206549455\F, -7. 1495013658, -2. 407385535, -0. 8895254796\F, -6. 9916248115 , 1. 5939605879, 3. 080272198\F, -6. 4996084107, 3. 2446310506, 1. 7564853412\F, -8. 1213692613, 1. 8803497818, 1. 2496709851\S, -1. 6294862095, 0. 0131052177, -2. 2613320743\\Versi on=ES64L-G09RevD. 01\\State=1-A\\HF=-2265. 9334657\\RMSD =3. 062e-09\\RMSF=3. 879e-06\\Di pol e=1. 5061837, 1. 3993149, 2. 5813559\\Quadrupol e=-5. 5669599, 10. 3516241, -4. 7846643, -1. 1306324, 8. 696064, -1. 3952121\\PG =C01 [X(C25H36F6N401S1)]\\@

Nucl eophi le 1

1\\1\GINC-R187\FOpt\RB3LYP\6-31G(d, p)\C9H8N201S1\R00T\08-Dec-2015\\# freq=noraman rb3lyp/6-31g** opt=(maxcyc=200) scf=maxcyc=200 scrf=(smd, sol vent=tol uene)\\thi al zone\\0, 1\C, 2. 1554781473, 0. 2865250426, -1. 087641 2741\N, 0. 7608895514, 0. 2468581964, -1. 2569431491\C, 0. 1370455655, 0. 142805 6312, -0. 1285469919\S, 1. 0717769464, 0. 0603537612, 1. 3773149285\C, 2. 614494 6534, 0. 151016418, 0. 3843888075\O, 2. 9433634364, 0. 4184260807, -2. 000445150 5\C, 3. 5631509442, 1. 2777329633, 0. 7972476518\H, 4. 4126237243, 1. 2886236346 , 0. 1081342314\H, 3. 940527843, 1. 1268027194, 1. 8118754788\H, 3. 067386483, 2. 2514982769, 0. 7525850411\C, -1. 3338590343, 0. 088377536, -0. 0324464268\C, -4 . 0405584073, -0. 0352659537, 0. 2560712795\C, -2. 128222856, 0. 1264854004, -1. 1841882541\C, -3. 1607699297, -0. 0653097693, 1. 3431224334\C, -3. 5109969468, 0. 0627724193, -1. 0293982231\H, -1. 6581958472, 0. 2038071719, -2. 1574254572\H, -3. 540957238, -0. 1406251873, 2. 3592228336\H, -4. 162067137, 0. 0893912875, -1. 8975925336\H, -5. 1115667728, -0. 0878153923, 0. 4216019107\H, 3. 123020701 8, -0. 8148681183, 0. 477055709\N, -1. 8356267876, -0. 0062094085, 1. 2097094748 \\Versi on=ES64L-G09RevD. 01\\State=1-A\\HF=-930. 7248031\\RMSD=3. 756e-09\\RM SF=1. 642e-05\\Di pol e=-1. 6832476, -0. 1296111, 0. 7592088\\Quadrupol e=11. 3769 612, -3. 1382543, -8. 2387069, 0. 2600011, 8. 9690621, 0. 7530011\\PG=C01 [X(C9H8 N201S1)]\\@

Electrophile 2

1\1\GI NC-R228\F0pt\RB3LYP\6-31G(d, p)\C8H7N102\R00T\08-Dec-2015\0\\# fr
eq=noraman rb3lyp/6-31g** opt=maxcyc=200 scf=maxcyc=200 scrf=(smd, sol v
ent=tol uene) \Ni tro-ol efi n\\0, 1\\C, -1. 2918462576, 0. 2423683973, 0. 0001013
845\\C, -0. 1927246612, -0. 5281051296, -0. 0004433292\\N, 1. 1237692805, 0. 06778
88413, 0. 0004942712\\0, 1. 2422449181, 1. 2976328932, 0. 0017773572\\0, 2. 076183
0956, -0. 7219065727, -0. 0001516785\\H, -0. 1459416234, -1. 6073583437, -0. 0015
662802\\H, -1. 1285640753, 1. 3171325925, 0. 0012663455\\C, -2. 6785138305, -0. 20
96379835, -0. 0007085052\\C, -3. 6949747019, 0. 7642788139, 0. 0000757672\\C, -3.
0475694702, -1. 5700876464, -0. 0022366964\\C, -5. 0374804338, 0. 3940614646, -0
. 0006346018\\H, -3. 4216752009, 1. 8156329434, 0. 0012512356\\C, -4. 3878067081,
-1. 9366763516, -0. 0029424809\\H, -2. 2835799191, -2. 3407096134, -0. 002869839
7\\C, -5. 386927374, -0. 9569120605, -0. 0021452073\\H, -5. 8091603691, 1. 1576023
494, -0. 0000122151\\H, -4. 6584773182, -2. 9879737308, -0. 0041181271\\H, -6. 432
6748107, -1. 2489794834, -0. 0027037997\\Versi on=ES64L-G09RevD. 01\State=1-
A\HF=-514. 1760499\RMSD=5. 485e-09\RMSF=2. 370e-05\Di pol e=-2. 5496758, -0. 5
367424, -0. 0011302\Quadrupol e=-6. 6220613, 5. 777527, 0. 8445344, -2. 3985822,
-0. 0040124, 0. 0046836\PG=C01 [X(C8H7N102)]\\@

TS1A-R, R

1\1\GI NC-R2526\FTS\RB3LYP\6-31G(d, p)\C42H51F6N7O4S2\R00T\09-Dec-2015\0
\\# freq=noraman rb3lyp/6-31g** opt=(ts, cal cfc, noeigen, maxcyc=200) scf
=maxcyc=200 scrf=(smd, sol vent=tol uene) iop(1/8=1)\\TS for 1st C-C addi
tion conf. 1: R(el ec), R(nuc) (tol opt; cat 111)\\0, 1\\C, -1. 2494256521, -
3. 3217510012, 1. 5412784212\\0, -1. 8428046469, -4. 3656027817, 1. 8276263161\\C
,-0. 4497597284, -2. 559354909, 2. 6260201828\\H, 0. 2602087816, -3. 3012465511,
3. 0030594856\\N, -1. 2451761302, -2. 7862125825, 0. 28985455\\H, -0. 7808328521,
-1. 8954887562, 0. 1743628054\\C, -1. 8576129575, -3. 3601722141, -0. 917472701\\H,
-1. 8278499219, -2. 5357806327, -1. 639196206\\C, -1. 0388791819, -4. 51870663
23, -1. 5987836517\\C, 0. 4270887284, -4. 0672780818, -1. 73737922\\H, 0. 90505582
77, -3. 9265574591, -0. 7649756148\\H, 0. 5059624251, -3. 1284151269, -2. 2995787
67\\H, 1. 0027685127, -4. 8233342217, -2. 2814790697\\C, -1. 0893396649, -5. 84796
52111, -0. 821879708\\H, -0. 6607399659, -5. 755668468, 0. 1767218786\\H, -0. 5232
31928, -6. 6112271309, -1. 3679756012\\H, -2. 1121938814, -6. 2231834928, -0. 707
5903777\\C, -1. 6028345021, -4. 7442357847, -3. 0189780114\\H, -2. 6271332884, -5
. 1329777883, -3. 0139811842\\H, -0. 9905264953, -5. 4817460953, -3. 5476638974\\H,
-1. 58864617, -3. 8229103883, -3. 6137670951\\C, -3. 3356732842, -3. 704894339
,-0. 6492774884\\H, -3. 712009638, -4. 4685146773, -1. 3305665645\\H, -3. 4422252
642, -4. 0690386299, 0. 3709633969\\N, -4. 2271605949, -2. 5055327939, -0. 812865
0749\\C, -5. 5429373154, -2. 638086815, -0. 078368513\\H, -5. 3825345194, -2. 3733
92849, 0. 9675433067\\H, -5. 8572679963, -3. 6833831058, -0. 1440380173\\C, -6. 48
38171309, -1. 7168153642, -0. 8438946751\\H, -7. 5322605662, -1. 9400666394, -0.
6338491797\\H, -6. 2844761741, -0. 6767711186, -0. 570893015\\C, -4. 5636787147,
-2. 1539368045, -2. 2613046213\\H, -4. 1991620575, -2. 9499487617, -2. 910442450
6\\H, -4. 0326332534, -1. 2325820176, -2. 4996800407\\C, -6. 0908788583, -1. 98049
98185, -2. 3063102216\\H, -6. 5659902056, -2. 8989378935, -2. 6659348678\\H, -6. 3
808054087, -1. 1705739794, -2. 9791623775\\N, 0. 3558533338, -1. 4792963388, 2. 07
16994486\\H, -0. 0934681591, -0. 5614948431, 1. 9574385303\\C, -1. 3503559441, -2
. 097985181, 3. 8385825763\\C, -1. 6645688365, -3. 3266129872, 4. 7163591637\\H, -
2. 3136754954, -3. 0354634606, 5. 5504463822\\H, -0. 7472925285, -3. 7501540105,
5. 1411523283\\H, -2. 1625751477, -4. 1100490194, 4. 1428874393\\C, -0. 573712967
1, -1. 0787483301, 4. 6963719503\\H, -1. 1349220511, -0. 8748932807, 5. 615190671
2\\H, 0. 4098327327, -1. 4626878847, 4. 9875683658\\H, -0. 4297618321, -0. 1246994
225, 4. 1820154774\\C, -2. 6714933697, -1. 4628475031, 3. 3618077972\\H, -2. 51466
1573, -0. 6375496995, 2. 6615538247\\H, -3. 225530459, -1. 0667376469, 4. 2204314
766\\H, -3. 3121451935, -2. 2088172023, 2. 8792439051\\C, 1. 6929735081, -1. 56133
09879, 1. 8353043108\\N, 2. 2268268077, -0. 3588225478, 1. 447331538\\H, 1. 628746
9459, 0. 4762828542, 1. 5451807061\\C, 3. 5112711566, -0. 0572026143, 0. 96667182
36\\C, 5. 9927017002, 0. 7744437113, -0. 0865689268\\C, 4. 0027612568, 1. 23410303
1, 1. 2250618834\\C, 4. 2722737321, -0. 9268916298, 0. 1762284079\\C, 5. 498010017
3, -0. 5020733052, -0. 3399626041\\C, 5. 2274126513, 1. 6371374954, 0. 7016425452
\\H, 3. 414015762, 1. 9196975698, 1. 8230915413\\H, 3. 9072354731, -1. 9224584426,
-0. 0342573279\\H, 6. 9427620745, 1. 0925781922, -0. 4974455989\\C, -2. 876719734
7, 0. 7725079255, -0. 7296040773\\N, -1. 6250849053, 0. 3985950223, -1. 234024007
5\\C, -1. 1329174248, 1. 3261053536, -2. 0113431405\\S, -2. 1280474027, 2. 7583278
343, -2. 2627711835\\C, -3. 2399420224, 2. 1546194918, -1. 0236388251\\O, -3. 6235
054464, -0. 0191930259, -0. 1010474722\\H, -3. 764430723, -1. 6346671385, -0. 418
089522\\C, -4. 6985294412, 2. 5164493832, -1. 1077982451\\H, -5. 2088667029, 2. 21
57359513, -0. 1892084646\\H, -5. 186176099, 1. 9939127496, -1. 941758526\\H, -4. 8
53728159, 3. 5876853737, -1. 2503322896\\C, 0. 1664670238, 1. 2454598213, -2. 697
1820778\\C, 2. 5445635579, 1. 2218132697, -4. 0443273747\\C, 1. 0876826223, 0. 228
0506119, -2. 4162892514\\C, 2. 2995974688, 0. 2225963971, -3. 1040007973\\C, 1. 56
44456269, 2. 1967129969, -4. 2518636983\\H, 0. 8586920508, -0. 5165441079, -1. 66
3879087\\H, 3. 0478925842, -0. 5366240461, -2. 9002194705\\H, 1. 7240701047, 2. 99
51738219, -4. 9732744165\\H, 3. 4760470658, 1. 2560520986, -4. 5994318758\\C, -2.
5294928753, 3. 1141587213, 0. 8472111455\\C, -1. 124120071, 3. 1270503441, 0. 886
9131861\\N, -0. 4195127932, 2. 0467121519, 1. 3263483704\\O, -1. 0295281435, 0. 98
3573429, 1. 6635278118\\O, 0. 8454863953, 2. 1233573491, 1. 4110303967\\H, -0. 508

4752287, 3. 9707213224, 0. 6163714551\H, -3. 0058722822, 2. 3298805887, 1. 42725
 10525\c, -3. 2501971741, 4. 4047915483, 0. 8056479118\c, -4. 4370828776, 4. 5504
 138315, 1. 5456047124\c, -2. 7831635948, 5. 5097256585, 0. 0692621475\c, -5. 128
 0036836, 5. 7612003981, 1. 5618536723\H, -4. 8072117657, 3. 7097503149, 2. 12585
 44666\c, -3. 4801878098, 6. 7156873746, 0. 0787523428\H, -1. 8756749689, 5. 4237
 222575, -0. 5195868372\c, -4. 6539053112, 6. 8480186877, 0. 8257004523\H, -6. 03
 6795544, 5. 8547669059, 2. 1493452473\H, -3. 1048998442, 7. 5562196481, -0. 4977
 862961\H, -5. 1918718219, 7. 7913940977, 0. 8346089122\c, 6. 2338672307, -1. 434
 5456867, -1. 2612432639\c, 5. 7720975297, 3. 0034018586, 1. 0196060122\f, 5. 597
 1646813, -1. 5460095147, -2. 4611944401\f, 6. 3189001993, -2. 6852967204, -0. 76
 13922435\f, 7. 4896759801, -1. 0174832422, -1. 5273128215\f, 6. 7265217715, 2. 9
 538694761, 1. 9800900021\f, 6. 3508011139, 3. 5738079731, -0. 0642401416\f, 4. 8
 172723195, 3. 8504130839, 1. 4577153548\s, 2. 586894857, -2. 9938714215, 2. 0216
 885915\n, 0. 4031987412, 2. 2168281812, -3. 5976451634\\Version=ES64L-G09Rev
 D. 01\State=1-A\HF=-3710. 8419853\RMSD=2. 761e-09\RMSF=2. 005e-06\Di pol e=-
 6. 6537827, -0. 2097452, -3. 366587\Quadrupole=17. 2215166, -9. 4653241, -7. 756
 1925, 17. 5997242, -2. 5011595, 16. 9565079\PG=C01 [X(C42H51F6N7O4S2)]\\@

TS1A-S, S

1\\GI NC-R2536\FTS\RB3LYP\6-31G(d, p)\C42H51F6N7O4S2\R00T\08-Dec-2015\0
 \\# freq=noraman rb3lyp/6-31g** opt=(ts, cal cfc, noeigen, maxcyc=200) scf
 =maxcyc=200 scrf=(smd, solvent=toluene)\\TS for 1st C-C addition conf.
 2: S(el ec), S(nucl) (tol opt; cat III)\\0, 1\c, 1. 0158485385, -3. 7267428083
 , 0. 3060019318\0, 1. 6919049613, -4. 7444672011, 0. 4974735651\c, -0. 087709557
 2, -3. 7157431807, -0. 7931549157\H, -0. 7778829127, -4. 5051658274, -0. 4796912
 529\n, 1. 144266676, -2. 5895557909, 1. 0305560215\H, 0. 5802611237, -1. 7901271
 804, 0. 7593720044\c, 1. 88116759, -2. 3739553205, 2. 2856050266\H, 1. 934191190
 3, -1. 2833461682, 2. 3691135934\c, 1. 0985704846, -2. 8539980721, 3. 5681699687
 \c, -0. 3559862141, -2. 3533319577, 3. 4727860153\H, -0. 9087437239, -2. 8514277
 945, 2. 6729473564\H, -0. 401056048, -1. 2728681749, 3. 2925661048\H, -0. 884025
 6361, -2. 5563869678, 4. 4104911061\c, 1. 0988316661, -4. 3857352958, 3. 7278514
 508\H, 0. 6636284677, -4. 8879510321, 2. 8614466398\H, 0. 5135381746, -4. 667466
 6049, 4. 61051337\H, 2. 1089975959, -4. 7859991239, 3. 870016884\c, 1. 738617494
 8, -2. 2096779384, 4. 8174243155\H, 2. 7448800289, -2. 5871574923, 5. 0277049442
 \H, 1. 1301433599, -2. 4299911654, 5. 7005963717\H, 1. 7989062279, -1. 119109451
 2, 4. 7237250257\c, 3. 3289579307, -2. 8860826674, 2. 2527207341\H, 3. 751193894
 , -2. 7993536314, 3. 2537440353\H, 3. 3838708192, -3. 923760604, 1. 9267960742\n
 , 4. 2275927282, -2. 0693443254, 1. 3616521661\c, 4. 1909800991, -2. 4235041906,
 -0. 1184753797\H, 3. 5676250685, -1. 6716170548, -0. 5999864055\H, 3. 735473156
 7, -3. 4074906326, -0. 2192969817\c, 5. 6551447134, -2. 3790317519, -0. 60023585
 68\H, 6. 0364617708, -3. 3948033308, -0. 7429054896\H, 5. 746208067, -1. 8516043
 098, -1. 5523438812\c, 5. 6748553376, -2. 1311313829, 1. 7795831825\H, 5. 908309
 3068, -3. 1687579417, 2. 0369742995\H, 5. 8110391804, -1. 4987015443, 2. 6585710
 228\c, 6. 4233116267, -1. 6770513103, 0. 5336290935\H, 7. 4804051956, -1. 950283
 558, 0. 569923244\H, 6. 3537160195, -0. 5894397871, 0. 433769629\n, -0. 86802681
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TS1B-R, R

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TS1B-S, S

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int1B-R, R

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int1B-S,

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TS2' A-R, R

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16. Copies of NMR Spectra

