Substrate Selectivity of an Isolated Enoyl Reductase Catalytic Domain from an Iterative Highly Reducing Fungal Polyketide Synthase Reveals Key Components of Programming

Douglas M. Roberts,^{1,2} Christoph Bartel,² Alan Scott,¹ David Ivison,¹ Thomas J. Simpson¹ and Russell J. Cox^{*1,2}

- 1. School of Chemistry, University of Bristol, Cantock's Close, Bristol, BS8 1TS, UK.
- 2. Institute for Organic Chemistry, Leibniz Universität Hannover, Schneiderberg 1B, 30167, Hannover, Germany.

* Corresponding Author: russell.cox@oci.uni-hannover.de

Electronic Supplementary Information

1.0	Synthesis and characterisation	2
2.0	Production and purification of ER protein	55
3.0	Kinetic and Inhibition Assay procedures	58
4.0	Stereochemical assay procedures	68
5.0	Stereochemical Assignment of Proton Chemical Shifts of 2-Methyl Butyric Acid.	70
6.0	LCMS conditions	71
7.0	Multiple alignment of ER sequences	72
8.0	Modelling and Docking Procedures	73
9.0	Integration of Angelic Mandelate Spectrum	74

1.0 Synthesis of substrates

All solvents were used as purchased unless otherwise stated apart from hexane, which was distilled before use. BuLi was titrated with diphenylacetic acid before use to confirm the concentration.

E-2-Ethylbut-2-enoic acid 10¹

Chemical Formula: C₆H₁₀O₂ Exact Mass: 114.07

Potassium *tert*-butoxide (1.68 g, 15.0 mmol)was dissolved in Tetrahydrofuran (THF). The solution was cooled to 0 °C and 1 eq. triethyl 2-phosphonobutyrate was added under nitrogen atmosphere. After the solution was stirred for 10 min at 0 °C, acetaldehyde (0.9 ml, 15.00 mmol) was added and the reaction mixture was then stirred for 3 h at 22 °C. The mixture was added to H₂O, the organic phase was extracted (3 ×) with ethyl acetate, washed (1 ×) with a saturated NaCl solution and dried over MgSO₄. The solvent was removed under reduced pressure and the product was purified by column chromatography (eluent: ethyl acetate/hexane 1:10).

The ester was dissolved in a H₂O/EtOH mixture (1:5) and 10 eq. of KOH were added. The reaction mixture was stirred for 4 h at 80 °C. The solution was cooled, diethyl ether was added and the organic phase was washed (1 x) with a saturated NaHCO₃ solution. The aqueous phase was acidified with aq. HCl (2 M) and the product was extracted (3 x) with ethyl acetate. The solvent was removed *in vacuo*. The product **10** was a yellow oil (0.58 g, 5.08 mmol, 34 %). ¹H NMR (400 MHz, CDCl₃): $\delta_{\rm H} = 1.02$ (3 H, t, *J*HH = 7.5 Hz, 20-CH₃), 1.83 (3 H, d, *J*HH = 7.1 Hz, 18-CH₃), 2.32 (2 H, q, *J*HH = 7.5 Hz, 19-CH₂), 6.96 (1 H, q, *J*HH = 7.1 Hz, 17-CH) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta_{\rm C} = 13.6$ (20-CH₃), 13.9 (18-CH₃), 19.6 (19-CH₂), 134.3 (16-C), 139.6 (17-CH), 171.3 (15-CO), ppm. IR: vM_{ax} = 2969 (w), 2937 (w), 2877 (w), 2656 (w), 1680 (vs), 1640 (s), 1417 (w), 1375 (w), 1290 (m), 1255 (s), 1163 (w), 1097 (w), 1072 (w), 1037 (w), 940 (w), 861 (w), 777 (w), 751 (w), 638 (w) cm⁻¹.

E-Ethyl 2-methyl hex-2-enoate 12a²

To a stirred solution of butryaldheyde (418 ml, 5.8 mmol) in dry CH_2Cl_2 (6 ml) was added (Carbethoxycarbonylethylidene)triphenylphosphorane (2.1 g, 5.8 mmol). This was refluxed overnight then allowed to cool to room temperature. Hexane (7 ml) was added and this was stirred for 10 mins and then filtered. The filtrate was concentrate in *vacuo* to approximately 5 ml and then hexane (7 ml) was added and stirred for a further 10 mins. This was filtered and then concentrated in *vacuo*. The resulting oil was purified by column chromatography (hexane:ethyl acetate 95:5) to yield the desired product (yellow oil, 690 mg, 4.4 mmol, 76%, Rf 0.21, hexane:ethyl acetate 95:5); $\delta_{\rm H}$ (400 MHz, CDCl₃) 6.76 (1H, tq *J* 7.6 1.5, H-3), 4.18 (2H, q *J* 7.1, H2-8), 2.18-2.11 (2H, m, H2-4), 1.85 (3H, dt *J* 1.4 0.9, H3-7), 1.52-1.42 (2H, m, H2-5), 1.29 (3H, t *J* 7.1, H3-9), 0.93 (3H, t *J* 7.4, H3-6); $\delta_{\rm C}$ (100 MHz, CDCl3) 168.5 (C-1), 142.3 (C-3), 128.0 (C-2), 60.5 (C-8), 30.8 (C-4), 22.0 (C-5), 14.4 (C-7), 14.0 (C-9), 12.5 (C-6); MS (ES+): *m/z* (%) 179 (4, [M]Na+), 157 (100, [M]H+), 129 (78, [M + H - CH₂CH₃]+).

E-2-methyl hex-2-enoic acid 12²



Chemical Formula: C₇H₁₂O₂ Exact Mass: 128.08

To a stirred solution of *E*-ethyl-2-methyl hex-2-enoate (**12a**, 125 mg, 0.8 mmol) in THF (2 ml) was added an aqueous NaOH (2M, 4 ml). The reaction was refluxed overnight and on completion this was acidified with aqueous HCl (2M) to pH 4 and then extracted with diethyl ether (3×7 ml). The organic fractions were combined, dried (MgSO₄) and concentrate in *vacuo* to yield the desired product (yellow oil, 75 mg, 0.6 mmol, 78%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 6.92 (1H, dq *J* 7.5 1.5, H-3), 2.21-2.16 (2H, m, H₂-4), 1.85-1.82 (3H, m, H₃-7), 1.49 (2H, tq *J* 7.4 7.3, H₂-5), 0.94 (3H, t *J* 7.4, H₃-6).

E-4*RS*-4-methyl hex-2-enoic acid 13³

Chemical Formula: C₇H₁₂O₂ Exact Mass: 128.08

2-methyl butyraldehyde (1.6 g, 19.0 mmol), malonic acid (1.2 g, 12.0 mmol), pyridine (3.8 ml) and morpholine (18.8 μ l) were stirred at room temperature for 24 h. The temperature was then raised to 115 °C and the reactions was stirred for a further 17 h. On completion the reaction was poured into aqueous NaOH (1M, 10 ml) and extracted with diethyl ether (3 × 10 ml) the aqueous layer was then acidified with aqueous HCl (2M) and extracted with diethyl ether (5 × 10 ml). The organics from the acidified extractions were combined, washed with water (25 ml), dried (MgSO₄) and concentrated in *vacuo* to yield the desired product (yellow oil, 738 mg, 5.8 mmol, 48%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 10.95 (1H, br s, H-8), 6.97 (1H, dd *J* 15.7 7.8, H-3), 5.79 (1H, dd *J* 15.7 1.2, H-2), 2.28-2.21 (1H, m, H-4), 1.45-1.38 (2H, m, H₂-5), 1.05 (3H, d *J* 6.6, H₃- 7), 0.88 (3H, t *J* 7.7, H₃-6); $\delta_{\rm C}$ (100 MHz, CDCl₃) 172.1 (C-1), 157.1 (C-3), 119.4 (C-2), 38.3 (C-4), 28.8 (C-5), 18.9 (C-7), 11.7 (C-6); ν M_{ax} (oil)/cm⁻¹ 2965, 2931, 2871.

E-Ethyl 2,4 dimethylhex-2-enoate 14a⁴

To a stirred solution of 2-methylbutryaldehyde (500 mg, 5.8 mmol) in dry CH_2Cl_2 (6ml) was added (Carbethoxyethylidene)triphenylphosphorane (2.1g, 5.8mmol). This reaction was refluxed overnight and then allowed to cool to room temperature. Hexane (7 ml) was added and this was stirred for 10 mins and then filtered to remove the precipitate that had formed. This was concentrate in *vacuo* to approximately 5 ml and then hexane (7 ml) was added and stirred for a further 10 mins and then filtered and concentrated in *vacuo*. The resulting oil was purified by column chromatography (hexane:ethyl acetate 95:5) to yield the desired product (yellow oil, 441 mg, 2.6 mmol, 45%, Rf 0.24, hexane:ethyl acetate 95:5); δ_H (300 MHz, CDCl₃) 6.52 (1H, dq *J* 7.2 1.4, H-3), 4.18 (2H, q *J* 7.1, H2-9), 2.47-2.32 (1H, m, H-4), 1.83 (3H, d *J* 1.4, H3-8), 1.47-1.33 (2H, m, H2-5), 1.29 (3H, t *J* 7.1, H3-10), 0.99 (3H, d *J* 6.7, H3-7) 0.85 (3H, t *J* 7.4, H3-6); δ_C (75 MHz, CDCl₃) 168.6 (C-1), 148.0 (C-3), 126.6 (C-2), 60.5 (C-9), 35.0 (C-4), 29.7 (C-5)), 19.8 (C-7), 14.4 (C-10), 12.6 (C-8), 12.0 (C-6); MS (ES+): *m/z* (%) 193 (4, [M]Na+), 171 (100, [M]H+), 143 (84, [M + H - CH₂CH₃]+).

E-4RS-2,4-Dimethylhex-2-enoic acid 14⁴



Exact Mass: 142.10

To a solution of ethyl *E*-4*RS*-2,4-dimethylhex-2-enoate (**14a**, 544 mg, 3.2 mmol) in THF (4 ml) was added aqueous NaOH (2M, 5 ml) and the reaction was refluxed for 4 h. The aqueous layer acidified to pH 4 with aqueous HCl (2M) and extracted with diethyl ether (3×10 ml). The organics were combined and washed with water (10 ml) and brine solution (10 ml). The organics were then dried (MgSO₄) and concentrated in *vacuo* to yield the desired product (yellow oil, 338 mg, 2.4 mmol, 74%); $\delta_{\rm H}$ (300 MHz, CDCl₃) 7.49 (1H,br s, H-9), 6.69 (1H, dq *J* 7.2 1.4, H-3), 2.50-2.34 (1H, m, H-4), 1.84 (3H, d *J* 1.5, H₃-7), 1.37-1.15 (2H, m H₂-5), 1.01 (3H, d *J* 6.6, H₃-8), 0.86 (3H, t *J* 7.5, H₃-6); $\delta_{\rm C}$ (75 MHz, CDCl₃) 173.8 (C-1), 150.9 (C-3), 125.9 (C-2), 35.2 (C-4), 29.7 (C-5), 19.7 (C-7) 12.3 (C-8), 12.0 (C-6); MS (ES +): m/z (%) 165 (6, [M]Na+), 143 (100, [M]H+).

E-4S-2,4-Dimethylhex-2-enoic acid 15⁵

Chemical Formula: C₈H₁₄O₂ Exact Mass: 142.10

2*S*-2-methylbutanol (1.00 ml, 10 mmol), pyridinium chlorochromate (2.2 g, 10 mmol) and celite (1.31 g) were dissolved in THF (30 ml). The reaction was followed by TLC and after full consumption of the alcohol (carbethoxyethylidene)triphenylphosphorane (4.20 g, 12.00 mmol) was added and the reaction heated under reflux for 7h. Diethyl ether was added and the mixture was filtered through Florisil. The filtered solution was concentrated *in vacuo* and the residue purified by column chromatography (hexane/ ethyl acetate, 4:1) to give (2E,4S)-methyl-2,4-dimethylhex-2-enoate (0.40 g, 3.40 mmol) which was directly hydrolysed. To the ester in ethanol/water 5:1 (5 ml/ 1 ml) was added potassium hydroxide (1.22 g, 22.00 mmol). After stirring under reflux for 3 h diethyl ether was added. The mixture was washed with NaHCO₃(3 × 10 ml). Then the aqueous layer was acidified with 2M HCl until pH 1 and extracted with ethyl acetate (3 × 10 ml). The organic layer was dried over MgSO₄ and concentrated *in vacuo*. The obtained product was a red oil (0.57 g, 3.30 mmol, 98 %). ¹H-NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 0.88 (t, *J* = 7.4 Hz, 6H, CH₃); 1.17 (d, *J* = 6.9 Hz, 3H, CH₃); 1.40-1.60 (m, 2H, CH₂); 1.85 (d, *J* = 1.5 Hz, 3H, 6-CH₃); 2.32-2.49 (m, 1H, 3-CH), 6.69 (dd, *J* = 1.5 Hz, *J* = 10.0Hz, 1H, 4-CH). ¹³C-NMR (CDCl₃, 100 MHz): $\delta_{\rm C}$ 11.9 (1-CH₃), 12.6 (5-CH₃), 19.7 (6-CH₃), 29.7 (2-CH₂), 34.9 (3-CH), 126.6 (7-C), 147.9 (4-CH), 168.4 (8-CO); ESMS, *m/z* (%): 143 [M]H⁺(100 %); [a]_D = + 44.0 (CH₂Cl₂) [lit⁶ +44.4 CH₂Cl₂]; v_{max} / cm⁻¹2966 (-C-H), 2935 (-C-H), 2877 (-C-H), 1697 (C=O), 1463 (CO), 1382 (C-O), 1278 (C-O).

E-2-ethyl-2-hex-2-enoic acid 16⁷



Potassium tert-butoxide (1.63 g, 14.5 mmol) was added to dry THF (20 ml) and cooled to 0 °C. Triethylphosphonobutyrate (3.5 ml, 14.5 mmol) was added and the solution stirred for 10 min. Butyraldehyde (14 mmol) was added and the mixture warmed up to room temperature. The solution was stirred for 3 h and then water (20 ml) was added and extracted with EtOAc (3×20 ml). The combined organic layers were washed with brine and dried

over MgSO₄. The crude product was purified by column chromatography (ethyl acetate/ hexane 1:10). The mixture of the *E*- and *Z*-isomers was a colourless oil (1.50 g, 8.80 mmol, 63 %). R_f: 0.69 (ethylacetate/ hexane 1:10). ¹H-NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 0.90-1.05 (m, 6H, 1-10-CH₃); 1.28-1.32 (m, 3H, 8-CH₃); 1.38-1.52 (m, 3H, 2-CH₂); 2.16 (q, *J* = 6.9 Hz, 2H, 3-CH₂); 2.24-2.40 (m, 2H, 9-CH₂); 4.16-4.23 (m, 2H, 7-CH₂); 5.83 (t, *J* = 7.9 Hz, 0.3 H, 4-CH); 6.71 (t, *J* = 7.4 Hz, 0.7 H, 4-CH). ¹³C-NMR (CDCl₃, 100 MHz): $\delta_{\rm C}$ 13.6 (1-CH₃), 13.9 (10-CH₃), 14.3 (8-CH₃), 20.0 (9-CH_{2E}), 22.1 (4-CH_{2E}), 22.7 (9-CH_{2Z}), 27.3 (2-CH_{2Z}), 30.4 (3-CH_{2E}), 31.5 (3-CH_{2Z}), 59.8 (7-CH_{2Z}), 60.3 (7-CH_{2E}), 133.9 (5-C_Z), 134.1 (5-C_e), 140.0 (4-CH_Z), 141.4 (4-CH_E), 168.0 (6-C_Z), 168.4 (6-C_E); ESMS, *m/z* (%): 184 [M - CH₂CH₃ + CH₃CN]H⁺(18 %), 151 [M]H⁺(8 %), 143 [M - CH₂CH₃]H⁺(100 %); v_{max} = 2963 (-C-H), 2935 (-C-H), 2874 (-C-H), 1709 (C=O), 1646 (-C=C), 1462 (CO), 1379 cm⁻¹.

To a solution of the ester in ethanol/water 5:1 (5 ml/ 1 ml) was added potassium hydroxide. After stirring under reflux for 3 h diethyl ether was added. The mixture was washed with NaHCO₃ (3 × 10 ml). Then the aqueous layer was acidified with 2M HCl until pH 1 and extracted with ethyl acetate (3 x 10 ml). The organic layer was dried over MgSO₄ and concentrated *in vacuo*. The obtained product was a red oil (1.2 g, 8.40 mmol, 96 %). ¹H-NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 0.91-1.08 (m, 6H, 1-8-CH₃); 1.40-1.54 (m, 2H, 2-CH₂); 2.20 (q, *J* = 7.7 Hz, 2H, 3-CH₂); 2.26-2.34 (m, 2H, 7-CH₂); 6.03 (t, *J* = 7.6 Hz, 1H, 4-CH_Z); 6.88 (t, *J* = 7.6 Hz, 1H, 4-CH_E). ¹³C-NMR (CDCl₃, 100 MHz): $\delta_{\rm C}$ 13.8 (1-CH₃), 13.8 (8-CH₃), 19.7 (7-CH_{2Z}), 22.0 (7-CH_{2E}), 27.4 (2-CH₂), 30.5 (3-CH_{2Z}), 31.7 (3-CH_{2E}), 132.6 (5-C_Z), 133.3 (5-C_E), 144.8 (4-CH_Z), 144.9 (4-CH_E), 173.3 (6-CO). ESMS *m/z* (%) 184 [M + CH₃CN]H⁺(18 %), 143 [M]H⁺(100 %); IR ν_{max} 2963 (-CH), 2934 (-CH), 2874 (-CH), 1681 (C=O), 1637 (-C=C), 1458 (CO), 1380 (C-O), 1291 (C-O) cm⁻¹.

E-Hept-2-enoic acid 17⁸

Chemical Formula: C₇H₁₂O₂ Exact Mass: 128.08

Valeraldhyde (1.6 g, 19.0 mmol), malonic acid (1.2 g, 12.0 mmol), pyridine (3.8 ml) and morpholine (18.8µl) were stirred at room temperature for 24 h. The temperature was raised to 115 °C and refluxed for 17 h. On completion the reaction was poured into aqueous NaOH (1M, 20 ml) and extracted with diethyl ether (5 × 20 ml). The aqueous phase was then acidified with aqueous HCl solution (2M, 15 ml) and extracted with diethyl ether (3 × 20 ml). The organic fractions from the extraction of the acidified aqueous phase were combined, washed with water (25 ml), dried (MgSO₄) and concentrated in *vacuo*. This crude product was purified to by Kugelrohr distillation to give the desired product (pale yellow oil, 685 mg, 5.3 mmol, 44%, 140- 150 °C, 11 mmHg); $\delta_{\rm H}$ (400 MHz, CDCl₃) 7.09 (1H, dt *J* 15.6 7.0, H-3), 5.82 (1H, dt *J* 15.6 1.6, H-2), 2.33 (2H, tdd *J* 7.0 7.0 1.6, H₂-4), 1.49-1.41 (2H, m, H₂-5), 1.38-1.31 (2H, m, H₂-6), 0.91 (3H, t *J* 7.3, H₃-7); $\delta_{\rm C}$ (100 MHz, CDCl₃) 172.3 (C-1), 152.7 (C-3), 120.7 (C-2), 32.2 (C-4), 30.1 (C-5), 22.4 (C-6), 13.9 (C-7); MS (ES-): *m/z* (%) 127 (100, [M - H]-), 83 (15, [M - COOH]-).

E-6-Methylhept-2-enoic acid 18⁹

Chemical Formula: C₈H₁₄O₂ Exact Mass: 142.10

Pyridinium chlorochromate (1.1 g, 5.0 mmol) and Celite (1.3 g) were stirred in THF (30 ml). 4-Methyl pentanol (535 μ l, 3.6 mmol) was added to the reaction, which was followed by TLC. After full consumption of the alcohol (1-2 h) methyl(triphenylphosphoranylidene)acetate (2.0g 6.1 mmol) was added and the reaction was refluxed for 7 h. On completion the reaction was cooled to RT, diethyl ether (20 ml) was added and the suspension was filtered through Florisil. The filtrate was then concentrated in *vacuo* and the resulting oil was purified by column chromatography (hexane:ethyl acetate 8:2) to yield the methyl ester (yellow oil, Rf 0.54, hexane: ethyl acetate 8:2): $\delta_{\rm H}$ (400 MHz, CDCl₃); 6.97 (1H, dt *J* 15.7 7.0, H-3), 5.87 (1H, dt *J* 15.6 1.6, H-2), 3.72 (3H, s, H₃-OMe), 2.24-2.17 (2H, m, H₂-4),

1.62-1.54 (1H, m, H-6), 1.36-1.31 (2H, m, H₂-5), 0.89 (6H, d *J* 6.6, H6-7). The resulting oil was dissolved in aqueous NaOH (6M, 3.0 ml) and THF (3.5 ml) and the solution was refluxed for 5 h. On completion the reaction mixture was acidified with HCl solution (2M) to pH 3 and extracted with diethyl ether (3 × 30 ml). The organics were dried (MgSO₄) and concentrated in *vacuo*, which gave the desired product (a pale yellow oil, 243 mg, 1.7 mmol, 46%, Rf 0.58, ethyl acetate:hexane 1:1); $\delta_{\rm H}$ (200 MHz, CDCl₃); 7.08 (1H, dt *J* 15.6 6.9, H-3), 5.83 (1H, dt *J* 15.6 1.5, H-2), 2.30-2.17 (2H, m, H₂-4), 1.68-1.48 (1H, m, H-6), 1.44-1.31 (2H, m, H₂-5), 0.90 (6H, d *J* 6.5, H6-7); $\delta_{\rm C}$ (100 MHz, CDCl₃); 171.3 (C-1), 152.8 (C-3), 120.4 (C-2), 37.0 (C-5), 30.4 (C-4), 27.7 (C-5), 22.5 (C-7); MS (ES-): *m/z* (%) 283 (73, [M₂-H]-), 141 (100, [M - H]-).

E-Ethyl 2-methyloct-2-enoate 20a¹⁰

To a stirred solution of hexanal (800 mg, 8.0 mmol) in dry CH_2Cl_2 (9 ml) was added (Carbethoxyethylidene)triphenylphosphorane (2.1 g, 8.0 mmol). The reaction mixture was refluxed overnight and then allowed to cool to RT. Then pentane (15 ml) was added and this was stirred for 15 mins. The formed suspension was filtered and concentrated in *vacuo*, which formed an orange brown solid and pentane (20 ml) was added. The suspension was stirred vigorously for 20 mins and then this was then filtered and concentrated in *vacuo*. The resulting oil was purified by column chromatography (hexane:ethyl acetate, 1:1) to yield the desired product. (yellow oil, 696 mg, 3.8 mmol, 48%, Rf 0.65, hexane:ethyl acteate 1:1); δ_H (300 MHz, CDCl₃) 6.76 (1H, tq *J* 7.5 1.5, H-3), 4.18 (2H, q *J* 7.1, H2-10), 2.19-2.12 (2H, m, H2-4), 1.82 (3H, dt *J* 1.5 1.0, H3-9), 1.47-1.40 (2H, m, H2-5), 1.32-1.25 (7H, m, H2-6 H2-7 H3-11), 0.89 (3H, t *J* 7.1, H3-8); δ_C (100 MHz, CDCl₃), 168.5 (C-1), 142.6 (C-3), 127.8 (C-2), 60.5 (C-10), 31.7 (C-6), 28.8 (C-4), 28.4 (C-5), 22.7 (C-7), 14.4 (C-11), 14.1 (C-9), 12.5 (C-8); MS (ES+): *m/z*(%) 207 (4, [M]Na+), 185 (44, [M]H+), 157 (100), 139 (6, [M + H - CO₂HCH₂CH₃]+) 87 (49).

E-2-methyloct-2-enoic acid 20¹¹

Chemical Formula: C₉H₁₆O₂ Exact Mass: 156.12

A solution of *E*-ethyl-2-methyloct-2-enoate (**20a**, 500 mg, 2.7 mmol) in a solution of THF (7.5 ml) and aqueous NaOH (6M, 6 ml) was refluxed for 4 h. On completion this was acidified with aqueous HCl (2M) to pH 4 and then extracted with diethyl ether (3×30 ml). The organic fractions were dried (MgSO₄) and concentrated in *vacuo*. The resulting oil was purified by bulb-to-bulb distillation to yield the desired product. (colourless oil, 160 mg, 1.0 mmol, 35%, bp 130-140 °C 7 mmHg); $\delta_{\rm H}$ (400 MHz, CDCl₃) 6.92 (1H, tq *J* 7.5 1.5, H-3), 2.23-2.16 (2H, m, H₂-4), 1.83 (3H, dt *J* 1.4 0.9, H₃- 9), 1.49-1.42 (2H, m, H₂-5), 1.34-1.27 (4H, m, H₂-6 and H₂-7), 0.89 (3H, t *J* 6.9, H₃-8); $\delta_{\rm C}$ (100 MHz, CDCl₃), 173.9 (C-1), 145.6 (C-3), 127.0 (C-2), 31.9 (C-6), 29.1 (C-4) 27.9 (C-5), 22.7 (C-7) 14.5 (C-9) 12.1 (C-8); MS (ES-): *m/z* (%) 155 (100, [M - H]-).

E-7-Methyloct-2-enoic acid 21¹²

Chemical Formula: C9H16O2 Exact Mass: 156.12

Pyridinium chlorochromate (1.3 g, 6.4 mmol) and Celite (1.4 g) were stirred in THF (40 ml). To this was added 5methyl hexanol (608 μ l, 4.3 mmol) and the reaction was followed by TLC. After full consumption of the alcohol (1-2 h) methyl(triphenylphosphoranylidene)acetate (2.4 g, 7.3mmol) was added and the reaction was refluxed for 7 h. On completion the reaction was cooled to RT, diethyl ether (20 ml) was added and the resulting suspension was filtered through Florisil. The filtrate was then concentrated in *vacuo* and further purified by column chromatography (hexane:ethyl acetate 8:2) to yield the methyl ester (yellow oil, Rf 0.57, hexane: ethyl acetate 8:2); $\delta_{\rm H}$ (400 MHz, CDCl₃); 6.97 (1H, dt *J* 15.6 7.0, H-3), 5.82 (1H, *J* 15.6 1.6, H-2), 3.72 (3H, s, H₃-OMe), 2.18 (2H, dtd *J* 7.2 7.2 1.6, H₂-4) 1.56-1.41 (3H, m, H₂-5 H-7), 1.23-1.16 (2H, m, H₂-6), 0.87 (6H, d *J* 6.6, H6-8). The resulting oil was dissolved in aqueous NaOH (6M, 3.0 ml) and THF (3.5 ml) and was refuxed for 5 h. On completion the reaction was acidified to pH 2 with aqueous HCl (2M) and then extracted with diethyl ether (3×30 ml). The organics were dried (MgSO₄) and concentrated in *vacuo*, this gave the desired product (yellow oil, 490 mg, 3.2 mmol, 73%, Rf 0.65, ethyl acetate:hexane, 1:1): $\delta_{\rm H}$ (400 MHz, CDCl₃); 7.09 (1H, dt *J* 15.6 6.9, H-3), 5.82 (1H, dt *J* 15.6 1.6, H-2), 2.22 (2H, dtd *J* 7.2 7.2 1.6, H₂-4), 1.58-1.44 (3H, m, H₂-5 H-7), 1.24-1.16 (2H, m, H₂-6), 0.88 (6H, d *J* 6.6, H6-8); $\delta_{\rm C}$ (100 MHz, CDCl₃); 171.7 (C-1), 152.6 (C-3), 120.6 (C-2), 38.5 (C-6), 32.7 (C-4), 30.0 (C-7), 25.9 (C-5), 22.7 (C-8); MS (ES-): *m/z* (%) 311 (100, [M₂-H]-), 155 (82, [M - H]-).

2RS,4RS-2,4-dimethyl hexanal 22a¹³



2-Butene (3.4 g, 60 mmol) was bubbled through dry THF (10 ml) at -78 °C for 5 min. Borane.THF (1M, 20 ml, 20 mmol) was added dropwise over 10 min to the reaction and after the addition was complete the reaction was warmed to -10 °C and stirred for 4 h. Methacrolein (2.1 g, 30 mmol) was then added in a dropwise manner and the reaction mixture was stirred for 30 min at RT. The reaction was quenched with water (1.5 ml) and then stirred overnight under N2 to remove excess of butene. Volatiles were removed in *vacuo* and the resultant yellow oil was purified by bulb-to-bulb distillation to yield the desired product an inseparable mixture of diastereomers (A and B 3:2, colourless oil, 1.5 g, 12 mmol, 62%, bp 45 °C/ 8 mmHg lit169 50 °C/ 13 mmHg); $\delta_{\rm H}$ (400 MHz, CDCl₃); 9.61 (0.4H, d *J* 2.0, H-1A), 9.58 (0.6H, d *J* 2.6, H-1B), 2.48-2.36 (1H, m, H-2), 1.72 (1H, m, H-3), 1.58-1.48 (1H, m, H-4), 1.45-1.33 (2.H, m, H₂-5), 1.18-1.10 (1H, m, H-3), 1.09-1.06 (3H, m, H₃-8), 0.90-0.86 (6H, m, H₃-6 H₃-7); $\delta_{\rm C}$ (100 MHz, CDCl₃); 205.7 (C-1A), 205.6 (C-1B), 44.4 (C-2B), 44.3 (C-2A), 38.0 (C-3A), 37.4 (C- 3B), 32.1 (C-4A), 31.9 (C-4B), 30.0 (C-5B), 29.3 (C-5A), 19.5 (C-7a), 18.9 (C-7B), 14.3 (C-8A), 13.5 (C-8B), 11.4 (C-6B), 11.3 (C-6A); υM_{ax} (oil)/cm⁻¹ 2961 (C-H), 2929 (C-H), 2876 (C-H), 2740 (C-H aldehyde), 1726 (C=O), 1461, 1378.

E-4RS,6*RS*-4,6-dimethyl oct-2-enoic acid 22¹⁴



Chemical Formula: C₁₀H₁₈O₂ Exact Mass: 170.13

2*RS*,4*RS*-2,4-dimethyl hexanal (**22a**, 260 mg, 2.0 mmol), malonic acid (789 mg, 1.3 mmol), pyridine (2.4 ml) and morpholine (12 μ l) were stirred at room temperature for 17 h. The reaction was refluxed at 115 °C for 17 h. On completion the solution was slowly poured into cooled aqueous NaOH (1M, 5 ml) and then extracted with diethyl ether (5 × 10 ml). After the extractions were complete the aqueous layer was acidified with aqueous HCl (2M) to pH 4, and a white suspension was formed. To this was added diethyl ether (10 ml) and the aqueous was extracted (2 × 10 ml). The organic extracts of the acidified aqueous extraction were combined, dried (MgSO₄) and concentrated in *vacuo*. This yielded the desired product as an inseperable mixture of diastereomers (A:B 3:2, colourless oil, 188 mg, 1.1 mmol, 84%); $\delta_{\rm H}$ (400 MHz, CDCl₃); 6.98 (0.6H, dd *J* 15.6 7.8, H-3A), 6.92 (0.4H, *J* 15.7 8.4, H-3B), 6.80-6.74 (1H, m, H-2), 2.47-2.38 (1H, m, H-4), 1.42-1.09 (7H, m, H₂-5 H-6 H₂-7), 1.02 (3H, t *J* 6.9, H₃-10), 0.87-0.81 (6H, m, H₃-8 H₃-9); $\delta_{\rm C}$ (100 MHz, CDCl₃); 172.4 (C-1A), 172.4 (C-1B), 158.3 (C-3A), 157.9 (C-3B), 119.1 (C-2B), 118.7 (C-2A), 43.4 (C-5B), 43.1 (C-5A), 34.5 (C-4B), 34.3 (C-4A), 32.1 (C-6B), 31.9 (C-6A), 29.9 (C-7B), 29.4 (C-7A), 20.3 (C-10B), 19.3 (C-9A), 19.2 (C-9B), 19.0 (C-10A), 11.3 (C-8B), 11.2 (C-8A); MS (ES-): *m/z* (%) 339 (14, [M₂-H]-), 169 (100, [M - H]-).

4S,6S-E-4,6-dimethyloct-2-enoic acid 1¹⁵



Chemical Formula: C₁₀H₁₈O₂ Exact Mass: 170.13

To a solution of squalestatin S1 (1.00 g, 1.40 mmol) in ethanol/water 5:1 (5 ml/ 1 ml) potassium hydroxide (1.22 g, 22.0 mmol) was added. After stirring under reflux for 3 h, diethyl ether was added to the solution and the mixture was washed ($3 \times \text{NaHCO}_3$, 10 ml). The aqueous layer with the (4S,6S)-2,4-dimethyloct-2-enoic acid was acidified with 2M HCl until pH 1 and extracted with ethyl acetate (3×10 ml). The organic layer was dried over MgSO₄ and concentrated *in vacuo*. The obtained product was not purified further and was isolated was an oil (0.21 g, 0.12 mmol, 86 %).

E-Dec-2-enoic acid 24⁸



Chemical Formula: C₁₀H₁₈O₂ Exact Mass: 170.13

Octanal (1.2g, 9.5mmol), malonic acid (0.8g, 6.0mmol), pyridine (1.9 ml) and morpholine (9.4 µl) were stirred at room temperature for 24 h. The temperature was raised to 115 °C and refluxed for 17 h. On completion the reaction mixture was poured into aqueous NaOH (1M, 20 ml) and extracted with diethyl ether (5 × 20 ml). The aqueous phase was then acidified with aqueous HCl (2M, 15 ml) and extracted with diethyl ether (3 × 20 ml). The organic fractions from the extraction of the acidified aqueous phase were combined, washed with water (25 ml), dried (MgSO₄) and concentrated in *vacuo* to give the desired product (pale yellow oil, 652 mg, 3.8 mmol, 64%); $\delta_{\rm H}$ (400 MHz, CDCl₃); 7.08 (1H, dt *J* 15.6 7.0, H-3), 5.81 (1H, dt *J* 15.6 1.6, H- 2), 2.26-2.18 (2H, m, H₂-4), 1.50-1.41 (2H, m, H₂-5), 1.35-1.22 (8H, m, H₂-6 H₂-7 H₂-8 H₂-9), 0.87 (3H, t *J* 6.9, H₃-10); $\delta_{\rm C}$ (100 MHz, CDCl₃); 172.2 (C-1), 152.7 (C-3), 120.7 (C-2), 32.5 (C-4), 31.9 (C-5), 29.2 (C-6), 29.2 (C-7), 28.0 (C-9), 14.2 (C-10); MS (ES-): *m/z* (%) 169 (100, [M - H]-).

E-2-methyldec-2-enoic acid 25¹⁶



Chemical Formula: C₁₁H₂₀O₂ Exact Mass: 184.15

To a solution of ethyl *E*-2-methyldec-2-enoate (synthesised by Wittig reaction of octanal with (Carbethoxyethylidene)triphenylphosphorane, Aldrich) in ethanol/water 5:1 (5 ml/ 1 ml) was added potassium hydroxide. After stirring under reflux for 3 h diethyl ether was added. The mixture was washed with NaHCO₃ (3 × 10 ml). Then the aqueous layer was acidified with 2M HCl until pH 1 and extracted with ethyl acetate (3 × 10 ml). The organic layer was dried over MgSO₄ and concentrated *in vacuo*. The obtained product was a red oil (0.11 g, 0.60 mmol, 86 %).¹H-NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 0.87-0.90 (m, 3H, 1-CH₃); 1.26-1.30 (m, 10H, 2-3-4-5-6-CH₂); 1.83 (d, *J* = 0.93 Hz, 3H, 10-CH₃); 2.19 (q, *J* = 7.3 Hz, 2H, 7-CH₂); 6.90 (td, *J* = 1.5 Hz, *J* = 7.6 Hz, 1H, 8-CH). ¹³C-NMR (CDCl₃, 100 MHz): $\delta_{\rm C}$ 12.3 (10-CH₃), 14.1 (1-CH₃), 22.6 (2-CH₂), 28.4 (4-CH₂), 28.9 (5-CH₂), 29.1 (6-CH₂), 29.3 (7-CH₂), 31.8 (3-CH₂), 126.8 (9-C), 145.6 (8-CH), 172.8 (11-C); ESMS *m/z* (%) 369 [M₂]H⁺(10 %), 226 [M + CH₃CN]H⁺(100 %), 167 [M - H₂O] H⁺(52 %); v_{max} (oil) / cm⁻¹ 2925 (-C-H), 2363 (-O-H), 1712 (C=O), 1650 (-C=C), 1466 (CO), 1367(C-O), 1268 (C-O).

N,*S*-diacetyl cysteamine¹⁷

To a solution of cysteamine hydrochloride (3.5 g, 30.8 mmol) in water (20 ml) at 0 °C was added aqueous KOH (6M) until pH 8 was reached. Acetic anhydride (8.7 ml, 92.4 mmol) was added dropwise and the reaction was maintained at a constant pH of 8 using aqueous KOH (6M). After all the acetic anhydride was added the pH was adjusted to 7 with aqueous HCl (2M) and the reaction was stirred at 0 °C for 1.5 h. NaCl was added to the reaction mixture until saturated and the mixture was extracted with CH₂Cl₂ (5 × 25 ml). The combined organics were dried (MgSO₄) and concentrated in *vacuo* to give the desired product (colourless oil, 4.8 g, 29.6 mmol, 96%, Rf 0.18, ethyl acetate/hexane, 1:1); $\delta_{\rm H}$ (400 MHz, CDCl₃) 6.10 (1H, br s, H-3), 3.42 (2H, m, H₂-2), 3.02 (2H, t *J* 6.5, H₂-1), 2.36 (3H, s, H₃-7), 1.98 (3H, s, H₃-5); $\delta_{\rm C}$ (75 MHz, CDCl₃) 196.2 (C-6), 171.3 (C-4, 39.6 (C-2), 30.6 (C-1), 28.6 (C-7), 22.9 (C-5); ν_{max} (oil)/cm⁻¹ 3297 (NH), 3085 (CH), 2935 (CH), 1691 (CONH), 954 (COS), 734 (CS).

N-Acetyl cysteamine¹⁷

KOH (2.8 g, 49.4 mmol) was added to a solution of *N*, *S* diacetyl cysteamine (2.5 g, 15.4 mmol) in water (10 ml) at 0 °C. The reaction was warmed to RT and stirred for 50 min. The reaction was then cooled to 0 °C and the pH was adjusted to 5 with aqueous HCl (2M). NaCl was added until saturation and extracted with CH₂Cl₂ (10 × 25 ml). The combined organics were dried (MgSO₄) and concentrated in *vacuo* to give the desired product (colourless oil, 1.8 g, 14.9 mmol, 97%, Rf 0.07, ethyl acetate/hexane, 1:1); $\delta_{\rm H}$ (400 MHz, CDCl₃) 6.23 (1H, br s, H-4), 3.41 (2H, m, H₂-3), 2.66 (2H, dt *J* 8.5 6.5, H₂-2), 2.00 (3H, s, H₃-6), 1.37 (1H, t *J* 8.4, H-1); $\delta_{\rm C}$ (75 MHz, CDCl₃) 175.5 (C-5), 42.4 (C-3), 24.7 (C-2), 23.3 (C-6); vM_{ax} (oil)/cm⁻¹ 3291 (NH), 3087 (CH), 2934 (CH), 2564 (SH), 1623 (CONH), 734 (CS).

General Procedure for Synthesis of SNAC Thiolesters

N-acetylcysteamine (172 mg, 1.4 mmol) and carboxylic acid (1.3 mmol) were dissolved in CH_2Cl_2 (2 ml) and cooled to 0 °C. DMAP (35 mg, 0.3 mmol) was added followed by EDCI (275 mg, 1.4 mmol) and the reaction was warmed to RT and followed by TLC. On completion the reaction was quenched with aqueous HCl (2M, 5 ml) and extracted with CH_2Cl_2 (3 × 10 ml). The combined organics were washed with saturated NaHCO₃ (10 ml), saturated NaCl (10 ml), dried (MgSO₄) and concentrated in *vacuo*. The resulting oil was purified by silica column chromatography to yield the desired product.

Tiglic-SNAC 5S¹⁸

Chemical Formula: C9H15NO2S Exact Mass: 201.08

(yellow oil, 75%, 1.0 mmol, Rf 0.16, ethyl acetate:petroleum ether, 4:1); δ_{H} (400 MHz, CDCl₃) 6.82 (1H, qq *J* 6.9 1.3, H-3), 6.28 (1H, br s, H-8), 3.38 (2H, q *J* 6.3, H₂-7), 3.02 (2H, t, *J* 6.5, H₂-6), 1.93 (3H, s, H₃-10), 1.83 (3H, p *J* 1.3, H₃-5) 1.80 (3H, dq *J* 7.0 1.1, H₃-4); δ_{C} (100 MHz, CDCl₃) 193.6 (C-1), 170.3 (C-9), 136.7 (C-3), 136.6 (C-2), 39.6 (C-7), 28.2 (C-6), 23.0 (C-10), 14.3 (C-5), 12.0 (C-4); MS (ES +): m/z (%): 425 (16, $[M_2 + Na]$ +), 403 (5, $[M_2 + H]$ +), 224 (100, [M]Na+), 202 (78, [M]H+); v_{max} (oil)/cm⁻¹ 3283 (N-H), 3078 (C-H), 2930 (C-H), 2859 (C-H), 1650 (C=O), 1621 (C=O), 1547 (N- H); elemental analysis; found: C, 53.8; H, 7.6; N, 6.8; S, 15.8; calculated: C, 53.7; H, 7.5; N, 7.0; S, 15.9.

Chemical Formula: C₉H₁₇NO₂S Exact Mass: 203.10

(yellow oil, 71%, 0.9 mmol, Rf 0.21, ethyl acetate:petroleum ether, 4:1), δ_{H} 400 MHz, CDCl₃) 5.86 (1H, br s, H-8), 3.44 (2H, q *J* 6.2, H₂-7), 3.02 (2H, t *J* 6.8, H₂-6), 2.60 (1H, dt *J* 6.9 6.8, H-2), 1.96 (3H, s, H₃-10), 1.79-1.67 (1H, m, H-3), 1.54-1.43 (1H, m, H-3), 1.17 (3H, d *J* 6.8, H₃-5), 0.90 (3H, t *J* 7.4, H₃-4); δ_{C} (100 MHz, CDCl₃) 204.8 (C-1), 170.2 (C-9), 50.2 (C-2), 39.9 (C-7), 28.1 (C-6), 27.1 (C-3), 23.1 (C-10), 17.1 (C-5), 11.5 (C-4); MS (ES +): *m/z* (%): 429 ([M₂ + Na]+), 407 ([M₂ + H]+), 226 ([M]Na+), 204 ([M]H+); vM_{ax} (oil/cm⁻¹ 3284 (N-H), 3078 (C-H), 2967 (C-H), 2933 (C-H), 2877 (C- H), 1684 (O=C), 1651 (O=C), 1548 (N-H); elemental analysis; found: C, 53.9; H, 8.7; N, 6.7; S, 15.1; calculated: C, 53.5, H, 8.4; N, 6.9; S, 15.8.

Crotonyl-SNAC 7S²⁰

Chemical Formula: C₈H₁₃NO₂S Exact Mass: 187.07

(colourless oil, 68%, 0.9 mmol, Rf 0.23, ethyl acetate:hexane, 4:1); $\delta_{\rm H}$ (400 MHz, CDCl₃) 6.95 (1H, dq *J* 15.4 6.78, H-3), 6.17 (1H, dq *J* 15.4 1.7, H-2), 5.90 (1H, br, H-7), 3.47 (2H, q *J* 6.8, H₂-6), 3.10 (2H, t *J* 6.6, H₂-5), 1.97 (3H, s, H₃-9), 1.91 (3H, dd J 7.0 1.6, H₃-4); $\delta_{\rm C}$ (100 MHz, CDCl₃) 190.1 (C-1), 170.4 (C-8), 141.7 (C-3), 129.8 (C- 2), 39.7 (C-6), 28.1 (C-5), 20.0 (C-9), 17.9 (C-4); MS (ES +): *m/z* (%): 397 (6, [M₂ + Na]+), 375 (54, [M₂ + H]+), 188 (100, [M]H+); J max (oil/cm-1 3250 (N-H), 3070 (C=C-H), 2943 (C-H), 2852 (C-H), 2784 (C-H), 1660 (C=O), 1635 (C=O), 1563 (N-H); HRMS (ESI +): calcd for [M]Na+, C₈H₁₃NO₂SNa: 210.0565: found 210.0565.

Angelic-SNAC 8S²¹

Chemical Formula: C₉H₁₅NO₂S Exact Mass: 201.08

(yellow oil, 53%, 0.7 mmol, Rf 0.16, ethyl acetate:petroleum ether, 4:1); $\delta_{\rm H}$ (400 MHz, CDCl₃) 5.93-5.82 (2H, m, H-3 H-8), 3.48 (2H, q *J* 5.9, H₂-7), 3.09 (2H, t *J* 6.6, H₂-6), 2.00 (3H, p *J* 1.3, H₃-5), 1.98 (3H, s, H₃-10), 1.95 (3H, dq *J* 7.2 1.5, H₃-4); $\delta_{\rm C}$ (100 MHz, CDCl₃) 202.1 (C-1), 170.2 (C-9), 134.7 (C-3), 134.2 (C-2, 39.8 (C-7), 28.3 (C-6), 23.2 (C-10), 20.5 (C-4), 15.8 (C-5); MS (ES +): *m/z* (%): 425 (20, [M₂ + Na]+), 224 (100, [M]Na+), 202 (70, [M]H+); $\nu_{\rm max}$ (oil)/cm⁻¹ 3284 (N-H), 3079 (C-H), 2979 (C-H), 2934 (C-H), 1655 (C=O), 1628 (C=O), 1547 (N-H); elemental analysis; found: C, 53.5; H, 7.3; N, 6.9; S, 15.8; predicted: C, 53.7; H, 7.5; N, 7.0; S, 15.9.

E-Hex-2-enoic-SNAC 11S²⁰

Chemical Formula: C₁₀H₁₇NO₂S Exact Mass: 215.10

(clear oil, 47%, 0.6 mmol, Rf 0.37, ethyl acetate); $\delta_{\rm H}$ (400 MHz, CDCl₃) 6.96 (1H, dt, *J* 15.6 6.8, H-3), 6.17 (1H, dt, *J* 15.4 1.5, H-2), 5.91 (1H, br s, H-9), 3.51-3.47 (2H, m, H₂-8), 3.12 (2H, t, *J* 6.6, H₂-7), 2.22 (2H, m, H₂-4), 1.99 (3H, s, H₃-11), 1.58-1.49 (2H, m, H₂-5), 0.97 (3H, t, *J* 7.4, H₃-6); $\delta_{\rm C}$ (100 MHz, CDCl₃) 190.6 (C-1), 170.8 (C-10), 146.6 (C-3), 128.4 (C-2), 39.8 (C-8), 34.2 (C-9), 28.2 (C-4), 23.2 (C-11), 21.1 (C-5), 13.7 (C-6); MS (ESI +) *m/z* (%) 238 (29, [M]Na+), 216 (44, [M]H+); ν_{max} (oil)/cm⁻¹ 3317 (N-H), 3042 (C=C-H), 2930 (C-H), 2872 (C-H), 1678 (C=O), 1625 (C=O), 1540 (NH); HRMS (CI +) calcd for [M]H + , C₁₀H₁₈NO₂S: 216.1058; found 216.1062.

E-2-Methylhex-2-enoic SNAC 12S

Chemical Formula: C₁₁H₁₉NO₂S Exact Mass: 229.11

(yellow oil, 18%, 0.3 mmol, Rf 0.40, ethyl acetate); $\delta_{\rm H}$ (300 MHz, CDCl₃) 6.78 (1H, tq *J* 7.4 1.3, H-3), 5.86 (1H, br s, H-10), 3.49-3.43 (2H, m, H₂-9), 3.07 (2H, t *J* 6.52, H₂- 8), 2.23-2.16 (2H, m, H₂-4), 1.92 (3H, s, H₃-12), 1.88-1.86 (3H, m, H₃-7), 1.54-1.47 (2H, m, H₂-5), 0.95 (3H, t *J* 7.4, H₃-6); $\delta_{\rm C}$ (100 MHz, CDCl₃) 194.2 (C-1), 170.4 (C- 11), 142.2 (C-3), 136.0 (C-2), 40.0 (C-9), 30.9 (C-4), 28.5 (C-8), 23.4 (C-12), 21.9 (C- 7), 14.0 (C-5), 12.6 (C-6); MS (ES +): *m/z* (%) 252 (14, [M]Na+), 230 (8, [M]H+); $\upsilon_{\rm max}$ (oil)/cm⁻¹ 3285 (NH), 3078 (HC=C), 2960 (CH), 2931 (CH), 2872 (CH), 1651 (CO), 1547 (NH); elemental analysis, observed (%) C, 57.7; H, 8.0; N, 6.1; S, 13.6; predicted C, 57.6; H, 8.4; N 6.1, S, 14.0; HRMS (CI +): calcd for [M]H+ , C₁₁H₂₀NO₂S: 230.1215: found 230.1213.

E-4RS-4-Methylhex-2-enoic-SNAC 13S²²

Chemical Formula: C₁₁H₁₉NO₂S Exact Mass: 229.11

(yellow oil, 54%, 0.7 mmol, Rf 0.28, ethyl acetate:hexane, 4:1); δ_{H} (400 MHz, CDCl₃) 6.83 (1H, dd *J* 15.6 7.7, H-3), 6.09 (1H, dd *J* 15.6 1.1, H-2), 5.88 (1H, br s, H-10), 3.49- 3.45 (2H, m, H₂-9), 3.09 (2H, t *J* 6.1, H₂-8), 2.26-2.19 (1H, m, H-4), 1.97 (3H, s, H₃- 12), 1.46-1.39 (2H, m, H₂-5), 1.06 (3H, d *J* 6.7, H₃-7), 0.89 (3H, t *J* 7.4, H₃-6); δ_{C} (100 MHz, CDCl₃) 190.6 (C-1), 170.2 (C-11), 151.7 (C-3), 127.8 (C-2), 39.8 (C-9), 38.2 (C-4), 28.7 (C-5), 28.3 (C-8), 23.2 (C-12), 18.8 (C-7), 11.6 (C-6); MS (ES +): *m/z* (%) 252 (100, [M]Na+), 230 (13, [M]H+); v_{max} (oil)/cm⁻¹ 3283 (N-H), 3065 (C=C-H), 2963 (C-H), 2930 (C-H), 2870 (C-H), 1651 (C=O), 1629 (C=O), 1547 (N-H); elemental analysis, observed (%) C, 57.5; H, 8.3; N, 6.2; S, 14.1; predicted C, 57.6; H, 8.4; N 6.1, S, 13.8; HRMS (CI +): calcd for [M]H+ , C₁₁H₂₀NO₂S: 230.1215; found: 230.1210.

E-4RS-2,4-Dimethylhex-2-enoic-SNAC 14S



Chemical Formula: C₁₂H₂₁NO₂S Exact Mass: 243.13

(yellow oil, 51%, 0.7 mmol, Rf 0.31 ethyl acetate:hexane, 4:1); δ_{H} (400 MHz, CDCl₃) 6.53-6.49 (1H, m, H-3), 6.10 (1H, br s, H-11), 3.42 (2H, q *J* 6.1, H₂-10), 3.04 (2H, t *J* 6.4, H₂-9), 2.47-2.38 (1H, m, H-4) 1.95 (3H, s, H₃-13), 1.87-1.84 (3H, m, H₃-8), 1.47-1.38 (1H, m, H-5), 1.38-1.28 (1H, m, H-5), 1.00 (3H, d *J* 6.8, H₃-7), 0.84 (3H, t *J* 7.4, H₃-6); δ_{C} (100 MHz, CDCl₃) 194.3 (C-1), 170.5 (C-12), 147.6 (C-3), 134.7 (C-2), 39.9 (C-10), 35.2 (C-4), 29.7 (C-5), 28.5 (C-9), 23.3 (C-13), 19.6 (C-8), 12.7 (C-7), 12.0 (C- 6); MS (ES +): *m/z* (%) 266 (44, [M]Na+), 244 (4, [M]H+); ν_{max} (oil)/cm⁻¹ 3285 (N-H), 3091 (C=C-H), 2961 (C-H), 2929 (C- H), 2870 (C-H), 1670 (C=O), 1653 (C=C), 1548 (N-H); elemental analysis, observed (%) C, 59.0; H, 8.6; N, 5.7; S, 13.0; predicted C, 59.2; H, 8.7; N 5.7, S, 13.2.

E-Hept-2-enoic-SNAC 17S

~~[™]s~~[™]√

Chemical Formula: C₁₁H₁₉NO₂S Exact Mass: 229.11

(pale yellow oil, 32%, 0.4 mmol, Rf 0.45, ethyl acetate); $\delta_{\rm H}$ (400 MHz, CDCl₃) 6.89 (1H, dt *J* 15.5 6.9, H-3), 6.21 (1H, br s, H-10), 6.08 (1H, dt *J* 15.4, 1.6, H-2), 3.42 (2H, q *J* 6.1, H₂-9), 3.05 (2H, t *J* 6.4, H₂-8), 2.21-2.14 (2H, m, H₂-4), 1.93 (3H, s, H-12), 1.46-1.37 (2H, m, H₂-5), 1.34-1.27 (2H, m, H₂-6), 0.87 (3H, t *J* 7.3, H₃-7); $\delta_{\rm C}$ (100 MHz, CDCl₃) 190.4 (C-1), 170.5 (C-11), 146.8 (C-3), 128.4 (C-2), 39.8 (C-9), 31.9 (C-4), 30.5 (C-5), 28.3 (C-8), 23.2 (C-12), 22.3 (C-6), 13.9 (C-7); MS (ES +): *m/z* (%) 252 (100, [M]Na+), 230 (7, [M]H+); v_{max} (oil)/cm⁻¹ 3284 (br N-H), 3081 (C=C-H), 2958 (C-H0, 2930 (C-H), 2872 (C-H), 1653 (C=O), 1631 (C=O), 1548 (N-H); elemental analysis, observed (%) C, 58.0; H, 8.5; N, 5.7; S, 13.6; predicted C, 57.7; H, 8.4; N 6.1; S, 14.0.

E-6-Methylhept-2-enoic-SNAC 18S

~~^Hs∽^H√

Chemical Formula: C₁₂H₂₁NO₂S Exact Mass: 243.13

(colourless oil, 72% (0.9 mmol), Rf 0.34, ethyl acetate): $\delta_{\rm H}$ (400 MHz, CDCl₃); 6.94 (1H, dt *J* 15.5 6.9, H-3), 6.14 (1H, dt *J* 15.5 1.5, H-2), 5.97-5.77 (1H, br s, H-10), 3.46 (2H, td *J* 6.1, H₂-9), 3.09 (2H, t*J* 6.6, H₂-8), 2.21 (2H, tdd *J* 7.5 7.5 1.6, H₂-4), 1.96 (3H, s, H₃-12), 1.61-1.52 (1H, m, H-6), 1.38-1.32 (2H, m, H₂-5), 0.90 (6H, d*J* 6.6, H6-7); $\delta_{\rm C}$ (100 MHz, CDCl₃); 190.6 (C-1), 170.4 (C-11), 147.2 (C-3), 128.3 (C-2), 40.0 (C-9), 37.1 (C-5), 30.3 (C-4), 28.4 (C-8), 27.8 (C-6), 23.4 (C-12), 22.5 (C-7); MS (ES +): *m/z* (%) 487 (100, [M₂ + H]+), 244 (79, [M]H+); υ_{max} (oil)/cm⁻¹ 3282 (br, N-H), 3079 (C=C-H), 2956 (C-H), 2930 (C-H), 2870 (C-H), 1654 (C=O), 1631 (C=O), 1548 (N-H); HRMS (ESI +): calcd for [M]Na+, C₁₂H₂₁NO₂SNa: 266.1191: found 266.1192.

Chemical Formula: C₁₂H₂₁NO₂S Exact Mass: 243.13

(pale yellow oil, 73%, 0.8 mmol, Rf 0.37, ethyl acetate); $\delta_{\rm H}$ (300 MHz, CDCl₃) 6.93 (1H, dt *J* 15.4 7.2, H-3), 6.12 (1H, dt *J* 15.5 1.5, H-2), 5.89 (1H, br s, H-11), 3.46 (2H, dt *J* 6.6 5.7, H₂-10), 3.09 (2H, t *J* 6.5, H₂-9), 2.25-2.15 (2H, m, H₂-4), 1.96 (3H, s, H₃-13), 1.52-1.42 (2H, m, H₂-5), 1.35-1.27 (4H, m, H₂-6, H₂-7), 0.89 (3H, t, *J* 6.3, H₃-8); $\delta_{\rm C}$ (100 MHz, CDCl₃) 190.6 (C-1), 170.5 (C-12), 147.0 (C-3), 128.4 (C-2), 40.0 (C-10), 32.3 (C-4), 31.6 (C-9), 28.4 (C-5), 27.7 (C-6), 23.3 (C-13), 22.5 (C-7), 14.1 (C- 8); MS (ES +): *m/z* (%) 266 (15, [M]Na+), 244 (100, [M]H+); v_{max} (oil)/cm⁻¹ 3286 (br, N-H), 3078 (H-C=C), 2957 (C-H), 2929 (C-H), 2858 (C-H), 1654 (C=O), 1631 (C=O), 1548 (NH); elemental analysis, observed (%) C, 59.6; H, 8.5; N, 5.8; S, 13.2; predicted C, 59.2; H, 8.7; N 5.8, S, 13.1.

E-2-Methyloct-2-enoic-SNAC 20S

Ů_s~^HY

Chemical Formula: C₁₃H₂₃NO₂S Exact Mass: 257.14

(colourless oil, 22%, 0.3 mmol, Rf 0.53, ethyl acetate); $\delta_{\rm H}$ (400 MHz, CDCl₃); 6.74 (1H, tq *J* 7.3 1.3, H-3), 6.12 (1H, br s, H-12), 3.41 (2H, dt *J* 6.6 5.0, H₂-11), 3.03 (2H, t *J* 6.4, H₂-10), 2.17 (2H, dtq *J* 7.4 7.3 0.8, H₂-4), 1.94 (3H, s, H₃-14), 1.84 (3H, dt *J* 1.1 0.9, H₃-9), 1.47-1.40 (2H, m, H₂-5), 1.35-1.24 (4H, m, H₂-6 H₂-7), 0.87 (3H, t *J* 7.1, H₃- 8); $\delta_{\rm C}$ (100 MHz, CDCl₃); 190.4 (C-1), 170.4 (C-13), 142.4 (C-3), 135.8 (C-2), 39.9 (C-11), 31.4 (C-6), 28.9 (C-4), 28.5 (C-5), 28.2 (C-10), 23.3 (C-14), 22.3 (C-7), 13.9 (C-8); MS (ES +): *m/z* (%) 280 (21, [M]Na+), 258 (2, [M]H+); $\upsilon_{\rm max}$ (oil)/cm⁻¹ 3285 (br N-H), 3079 (C=C-H), 2960 (C-H), 2926 (C-H), 2858 (C-H), 1652 (C=O), 1548 (N-H); elemental analysis, observed (%) C, 60.9; H, 9.0; N, 5.4; S, 12.1; predicted C, 60.7; H, 9.0; N, 5.4; S, 12.5.

E-7-Methyl oct-2-enoic SNAC 21S²⁴

° s∽^H

Chemical Formula: C₁₃H₂₃NO₂S Exact Mass: 257.14

(colourless oil, 20%, 0.3 mmol, Rf 0.36, ethyl acetate); δ_{H} (400 MHz, CDCl₃); 6.94 (1H, dt *J* 15.5 6.9, H-3), 6.13 (1H, dt *J* 15.5 1.5, H-2), 5.89 (1H, br s, H-11), 3.47 (2H, dt *J* 6.0 6.0, H₂-10), 3.09 (2H, t *J* 6.4, H₂-9), 2.25 (2H, tdd *J* 7.0 7.0 1.5, H₂-4), 1.96 (3H, s, H₃-13), 1.57-1.42 (3H, m, H₂-5 H-7), 1.20 (2H, dt *J* 6.9 6.9, H₂-6), 0.88 (6H, d *J* 6.6, H6-8); δ_{C} (100 MHz, CDCl₃); 190.6 (C-1), 170.1 (C-12) 147.0 (C-3), 128.4 (C-2), 40.0 (C-10), 38.6 (C-6), 32.7 (C-4), 29.9 (C-5), 28.4 (C-9), 28.0 (C-7), 23.4 (C-13), 22.7 (C-8); MS (ES +): *m/z* (%) 515 (43, [M₂ + H]+), 258 (100, [M]H+); ν_{max} (oil)/cm⁻¹ 3278 (br N-H), 3079 (C=C-H), 2954 (C-H), 2930 (C-H), 2869 (C-H), 1653 (C=O), 1632 (C=O), 1547 (N-H); HRMS (ESI +): calcd for [M]Na+, C₁₃H₂₃NO₂SNa: 280.1347: found 280.1347.



Chemical Formula: C₁₄H₂₅NO₂S Exact Mass: 271.16

An inseparable mixture of diastereomers A and B 3:2 (yellow oil, 32%, 0.4 mmol, Rf 0.36, ethyl acetate); $\delta_{\rm H}$ (300 MHz, CDCl₃); 6.84 (0.5H, dd *J* 15.5 7.7, H-3A), 6.79 (0.5H, dd *J* 15.7 8.3, H-3B), 6.11-6.06 (1H, m, H-2), 5.87 (1H, br s, H-13), 3.47 (2H, q *J* 6.5. H₂-12 A), 3.09 (2H, t *J* 6.3, H₂-11), 2.45-2.34 (1H, m, H-4), 1.97 (3H, s, H₃-15), 1.42-1.23 (3H, m, H₂-5 H-6), 1.20-1.10 (2H, m, H₂-7), 1.04 (3H, dd *J* 6.7 6.7, H₃-10), 0.88-0.81 (6H, m, H₃-8 H₃-9); $\delta_{\rm C}$ (100 MHz, CDCl₃); 190 (C-1), 170.4 (C-14), 152.6 (C-3A), 152.1 (C-3B), 126.7 (C-2A), 126.4 (C-2B), 43.4 (C-5A), 43.1 (C-5B), 40.0 (C-12), 34.5 (C-4A). 34.3 (C-4B), 32.1 (C-6A), 31.9 (C-6B), 29.9 (C-7A), 29.5 (C-7B), 28.4 (C-11), 23.4 (C-15), 20.3 (C-10), 19.3 (C-9A), 19.2 (C- 9B), 11.3 (C-8A), 11.2 (C-8B); MS (ES +): *m/z* (%) 294 (27, [M]Na+), 272 (100, [M]H+); v_{max} (oil)/cm⁻¹ 3287 (br, NH), 3075 (HC=C), 2961 (C-H), 2926 (C-H), 2874 (C-H), 1654 (CO), 1629 (CO), 1548 (NH); elemental analysis, observed (%) C, 62.4; H, 9.0; N, 5.1; S, 12.2; predicted C, 62.0; H, 9.3; N, 5.2; S, 11.8.

E-Dec-2-enoic SNAC 24S²⁵



Chemical Formula: C₁₄H₂₅NO₂S Exact Mass: 271.16

(white solid, 27%, 0.4 mmol, Rf 0.33, ethyl acetate); δ_{H} (400 MHz, CDCl₃); 6.94 (1H, dt *J* 15.5 7.1, H-3), 6.13 (1H, dt *J* 15.5 1.5, H-2), 5.86 (1H, br s, H-13), 3.46 (2H, td *J* 6.2 6.2, H₂-12), 3.09 (2H, t *J* 6.3, H₂-11), 2.21 (2H, dtd *J* 7.2 7.2 1.5, H₂-4), 1.96 (3H, s, H₃-15), 1.47 (2H, tt *J* 7.3 7.2, H₂-5), 1.34-1.29 (8H, m, H₂-6 H₂-7 H₂-8 H₂-9), 0.88 (3H, t *J* 7.1, H₃-10); δ_{C} (100 MHz, CDCl₃); 190.8 (C-1), 170.4 (C-14), 147.1 (C-3), 128.4 (C- 2), 40.0 (C-12), 32.4 (C-4), 31.9 (C-8), 29.3 (C-6), 29.2 (C-7), 28.4 (C-11), 28.1 (C-5), 23.4 (C-15), 22.8 (C-9), 14.3 (C-10); MS (ES-): *m/z* (%) 543 (25, [M₂ + H]+), 272 (100, [M]H+); ν_{max} (oil)/cm⁻¹ 3284 (N-H), 3078 (C=C-H), 2954 (C-H), 2930 (C-H), 2869 (C.H), 1653 (C=O), 1632 (C=O), 1547 (N-H); HRMS (ESI +): calcd for [M]Na+ , C₁₄H₂₅NO₂SNa: 294.1504: found 294.1501.

Pantetheine dimethyl ketal²⁶



D-pantothenic acid hemi-calcium salt (2.50 g, 10.50 mmol), p-toluensulfonic acid (2.30 g, 13.00 mmol) and molecular sieves (5 g) were suspended in dry acetone (125 ml) and stirred at 25 °C for 12 h under a nitrogen atmosphere. The suspension was filtered with celite and washed with 200 ml acetone. The filtrate was concentrated to a colourless oil, redissolved in ethyl acetate (200 ml) and washed with brine (2 × 25 ml) and dried over MgSO₄. After that the ethyl acetate was removed under vacuum and hexane was added to the flask to give a white solid that was dried under high vacuum. The corresponding D-pantothenic dimethyl ketal (1.90 g, 7.00 mmol was dissolved in 40 ml dry THF with CDI (1.70 g, 11.00 mmol) and stirred for one hour at 25 °C. Then cysteamine (1.30 g, 11.00 mmol) was added to the solution and stirred for 12 h. The solution was concentrated under vacuum and CH₂Cl₂ was added. The organic layer were washed with NH₄Cl (1 × 25 ml) and brine (1 × 25 ml), dried over MgSO₄ and concentrated *in vacuo*. The colourless oil was purified by column chromatography (ethyl acetate) to give a white solid (1.90 g, 6.00 mmol, 86 %). R_f: 0.1 (ethylacetate). ¹H-NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 0.98 (s, 3H, 11-CH₃); 1.05 (s, 3H, 12-CH₃); 1.39 (t, *J* = 8.6 Hz, 1H, SH); 1.43 (s, 3H, 15-CH₃); 1.47 (s, 3H, 16-CH₃); 2.40 (t, *J* = 5.8 Hz, 2H, 5-CH₂); 2.64-2.70 (m, 2H, 1-CH₂); 3.29 (d, *J* = 12.0 Hz, 1H, 13a-CH₂); 3.37-3.63 (m, 4H, 2-6-CH₂); 3,69 (d, *J* = 12.0 Hz, 1H, 13b-CH₂); 4.09 (s,1H, 9-CH); 6.37

(bt, J = 5.2 Hz, 1H, 3-NH); 7.03 (bt, J = 5.9 Hz, 1H, 7-NH). ¹³C-NMR (CDCl₃, 100 MHz): δ_{C} 18.7 (12-CH₃); 18.9 (11-CH₂); 22.1 (15-CH₃); 24.6 (1-CH₂); 29.5 (15-CH₃); 33.0 (10-C); 34.9 (6-CH₂); 36.2 (5-CH₂); 42.4 (2-CH₂); 71.4 (13-CH₂); 77.2 (9-CH); 99.1 (14-C); 170.3 (4-CO); 171.1 (8-CO). ESMS: m/z (%): 319 [M]H⁺(65%), 261 [M-(CH₃)₂CO]H⁺(100%). IR (v_{max}/cm^{-1}): 3419 (N-H), 3324 (N-H), 2980 (CH), 2944 (CH), 2872 (CH), 2556 (C-O), 1659 (-C=O).

tigloyl pantetheine dimethyl ketal 5Pa



Tiglic acid (**5**, 0.10 g, 1.00 mmol) and pantetheine dimethyl ketal (0.32 g, 1.00 mmol,) were dissolved in CH₂Cl₂ (8 ml). The mixture was cooled to 0 °C. Then *N*,*N*-dimethylaminopyridine (0.10 g, 0.80 mmol,) and *N*-(3-Diethylaminopropyl)-*N*-ethylcarbodiimide (0.38 g, 2.00 mmol) were added. The mixture was warmed to 25 °C and stirred for 4 h. After that the mixture was quenched with 2M HCl (10 ml) and extracted with CH₂Cl₂ (3 × 25 ml). The organic layer was washed with saturated NaHCO₃ (20 ml) and brine (20 ml). The product was dried over MgSO₄ and concentrated *in vacuo*. The crude product was purified by column chromatography (ethyl acetate). The obtained product was a colourless oil (0.32 g, 0.80 mmol, 80 %). R_f: 0.46 (ethyl acetate). ¹H-NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 0.99 (s, 3H, 18-CH₃); 1.04 (s, 3H, 19-CH₃); 1.44 (s, 3H, 16-CH₃); 1.48 (s, 3H, 17-CH₃); 1.65 (s, 3H, 4-CH₃); 1.85-1.87 (m, 6H, 1-4-CH₃); 2.44 (t, *J* = 6.5 Hz, 2H, 9-CH₂); 3.08 (t, *J* = 6.3 Hz, 2H, 6-CH₂); 3.28 (d, *J* = 11.7 Hz, 1H, 14a-CH₂); 3.41-3.62 (m, 4H, 7-10-CH₂); 3.68 (d, *J* = 11.6 Hz, 1H, 14b-CH₂); 4.08 (s, 1H, 12-CH); 6.13 (bt, *J* = 5.8 Hz, 1H, 8-NH); 6.93 (qq, *J* = 1.4, 6.8 Hz, 1H, 2-CH); 7.03 (bt, *J* = 5.9 Hz, 1H, 11-NH). ¹³C-NMR (CDCl₃, 100 MHz): $\delta_{\rm C}$ 12.2 (4-CH₃); 14.4 (1-CH₃); 18.7 (18-CH₃); 18.9 (16-CH₃); 22.1 (19-CH₃); 28.3 (6-CH₂); 29.5 (17-CH₃); 32.9 (13-C); 34.8 (9-CH₂); 35.9 (10-CH₂); 39.7 (7-CH₂); 71.5 (14-CH₂); 77.2 (12-CH); 99.1 (15-C); 136.8 (3-CH); 136.9 (2-CH); 170.1 (8-CO); 171.1 (11-CO); 190.2 (5-CO). ESMS: *m/z* (%) 423.3 [M]Na⁺ (100 %). IR (v_{max} / cm⁻¹): 3306 (NH), 2940 (CH₂), 1648 (CO), 1523 (C-O), 1438 (C=C). HRMS: *m/z* (%): calculated 423.1930, found 423.1929 for [M]Na⁺.



Preparation of angeloyl pantetheine dimethyl ketal 8Pa



Angelic acid (8, 0.11 g, 1.00 mmol) and pantetheine dimethyl ketal (0.32 g, 1.00 mmol) were dissolved in CH₂Cl₂ (8 ml). Then the mixture was cooled to 0 °C. Then N,N-dimethylaminopyridine (0.1 g, 0.80 mmol) and N-(3-Diethylaminopropyl)-N-ethylcarbodiimide (0.38 g, 2.00 mmol) were added to the mixture. The mixture was warmed to 25 °C and then stirred for 4 h. After that the mixture was quenched with 2M HCl (10 ml) and extracted with CH_2Cl_2 (3 × 20 ml). The organic layer was washed with saturated NaHCO₃ (25 ml) and brine (25 ml). The product was dried over $MgSO_4$ and concentrated *in vacuo*. The crude product was purified by column chromatography (ethyl acetate). The obtained product was a yellow oil (0.29 g, 0.70 mmol, 70 %). R_f : 0.49 (ethyl acetate). ¹H-NMR (CDCl₃, 400 MHz): δ_H 0.97 (s, 3H, 19-CH₃); 1.04 (s, 3H, 18-CH₃); 1.41 (s, 3H, 16-CH₃); 1.46 (s, 3H, 17-CH₃); 1.92-1.97 (m, 3H, 1-CH₃); 1.99 $(t, J = 1.5 \text{ Hz}, 1\text{H}, 4\text{-CH}_3); 2.19 (dg, J = 1.5, 7.3 \text{ Hz}, 2\text{H}, 3\text{-CH}_2); 2.42 (t, J = 6.0 \text{ Hz}, 2\text{H}, 9\text{-CH}_2); 3.07 (t, J = 6.7 \text{ Hz}, 2\text{H}, 3\text{-CH}_2); 3.07 (t, J = 6.7 \text{ Hz}, 3\text{-CH}_2); 3.07 (t, J = 6.7$ 2H, 6-CH₂); 3.27 (d, J = 11.6 Hz, 1H, 14a-CH₂); 3.40-3.62 (m, 4H, 7-10-CH₂); 3.68 (d, J = 11.6 Hz, 1H, 14b-CH₂); 4.07 (s,1H, 12-CH); 5.89 (dt, J = 1.3, 7.3 Hz, 1H, 2-CH); 6.11 (bt, J = 5.1 Hz, 1H, 8-NH); 7.03 (bt, J = 5.9 Hz, 1H, 11-NH). ¹³C-NMR (CDCl₃, 100 MHz): δ_C 15.9 (1-CH₃); 18.7 (19-CH₃); 18.9 (16-CH₃); 20.5 (4-CH₃); 22.1 (18-CH₃); 28.4 (6-CH₂); 29.5 (17-CH₃); 32.9 (13-C); 34.8 (9-CH₂); 35.9 (10-CH₂); 39.8 (7-CH₂); 71.5 (14-CH₂); 77.3 (12-CH); 99.1 (15-C); 134.1 (3-C); 134.9 (2-CH); 170.0 (8-CO); 171.2 (11-CO); 193.9 (5-CO). ESMS: *m/z* (%) 423.4 [M]Na⁺(25 %), 401.4 [M]H⁺(28 %), 343.4 [M - C₃H₆O]H⁺(100 %). IR ($v_{max} = cm^{-1}$): 3260 (NH), 3078 (CH₂), 2924 (CH₂), 1651 (CO), 1538 (C-O), 1446 (C=C). HRMS: m/z (%): calculated 423.1930, found 423.1932 for [M]Na+.



2,3-Dimethylbut-2-enoyl panetheine dimethyl acetal 9Pa



The reaction was carried out with 0.17 g (1.50 mmol) of 2,3-dimethylbut-2-enoic acid,²⁷ 0.48 g (1.50 mmol) of pantetheine dimethyl ketal, 0.29 g (1.50 mmol) of EDCI and a catalytic amount of DMAP in 10 ml CH₂Cl₂. The solution was added to aq. HCl (10 ml, 2 M) and the product was extracted with 3×20 ml CH₂Cl₂. The organic phase was washed with 1×50 ml of a saturated NaHCO₃ solution and with 1×50 ml of a saturated NaCl solution. The product was a colorless oil (0.55 g, 1.32 mmol, 88 %). ¹H NMR (400 MHz, CDCl₃): $\delta_{\rm H} = 0.96$ (3 H, s, 10/11-CH₃), 1.03 (3 H, s, 10/11-CH₃), 1.41 (3 H, s, 13/14-CH₃), 1.45 (3 H, s, 13/14-CH₃), 1.78 (3 H, s, 19-CH₃), 1.92 (3 H, s, 18-CH₃ and 20-CH₃), 2.41 (2 H, t, JHH = 6.1 Hz, 6-CH₂), 3.04 (2 H, t, JHH = 6.5 Hz, 9-CH₂), 3.27 (1 H, d, 3JHH = 11.7 Hz, 1a/b-CH), 6.18 (1 H, br. t, NH), 7.03 (1 H, br. t, NH) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta_{\rm C} = 16.2$ (18-CH₃ and 20-CH₃), 19.1 (13/14-CH₃), 22.3 (10/11-CH₃), 23.0 (19-CH₃), 28.7 (9-CH₂), 29.6 (13/14-CH₃), 33.1 (2-C), 34.9 (5-CH₂), 36.1 (6- CH₂), 39.9 (8-CH₂), 71.6 (1-CH₂), 77.3 (3-CH), 99.2 (12-C), 129.4 (17-C), 140.6 (16-C), 170.2 (7-CO), 171.3 (4-CO), 196.2 (15-CO) ppm. IR: $\nu_{max} = 3307$ (w), 2937 (w), 2871 (w), 1737 (w), 1652 (vs), 1521 (s), 1438 (w), 1376 (m), 1288 (w), 1254 (w), 1222 (w), 1196 (m), 1159 (w), 1096 (s), 1048 (w), 1018 (w), 1000 (w), 931 (m), 900 (w), 873 (w), 784 (w), 690 (w), 642 (w) cm⁻¹. ESMS: m/z (%) = 437 (15) [M]Na+ 415 (65) [M]H+, 357 (100) [M + H - C₃H₆O]+.



E-2-Ethylbut-2-enoyl pantetheine dimethyl acetal 10Pa



The reaction was carried out with 0.58 g (5.09 mmol) of **10**, 1.62 g (5.09 mmol) of pantetheine dimethyl ketal, 0.97 g (5.09 mmol) of EDCI and a catalytic amount of DMAP in 10 ml CH₂Cl₂. The solution was dumped in 10 ml HCl (2 M) and the product was extracted with 3×20 ml CH₂Cl₂. The organic phase was washed with 1×50 ml of a saturated NaHCO3 solution and with 1×50 ml of a saturated NaCl solution. The product was a colorless oil (0.99 g, 2.38 mmol, 47 %). ¹H NMR (400 MHz, CDCl₃): $\delta_{\rm H} = 0.96$ (3 H, s, 10/11-CH₃), 0.99 (3 H, t, *J*HH = 7.5 Hz, 20-CH₃), 1.03 (3 H, s, 10/11-CH₃), 1.41 (3 H, s, 13/14-CH₃), 1.45 (3 H, s, 13/14-CH₃), 1.82-1.90 (3 H, m, 18-CH₃), 2.41 (2 H, t, *J*HH = 6.1 Hz, 6-CH₂), 2.28-2.45 (2 H, overlapped peak, 19-CH₂), 3.04 (2 H, t, *J*HH = 6.3 Hz, 9-CH₂), 3.27 (1 H, d, 3*J*HH = 11.5 Hz, 1a/b-CH), 3.38-3.64 (4 H, overlapping peaks, 5-CH₂ and 8-CH₂), 3.67 (1 H, d, 3*J*HH = 11.5 Hz, 1a/b-CH), 4.07 (1 H, s, 3-CH), 6.16 (1 H, br. t, NH), 6.81 (1 H, q, *J*HH = 7.0 Hz, 17-CH); 7.03 (1 H, br. t, NH) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta_{\rm C} = 13.5$ (20-CH₃), 14.3 (18CH₃), 18.8 (10/11-CH₃), 19.1 (13/14-CH₃), 20.2 (19-CH₂), 22.3 (10/11-CH₃), 28.5 (9-CH₂), 29.6 (13/14-CH₃), 33.1 (2-C), 35.0 (5-CH₂), 36.1 (6-CH₂), 39.9 (8-CH₂), 71.6 (1-CH₂), 77.3 (3-CH), 99.2 (12-C), 136.7 (17-CH), 143.3 (16-CH), 170.2 (7-CO), 171.3 (4-CO), 193.5 (15-CO) ppm. IR: $v_{max} = 3306$ (w), 3080 (w), 2937 (w), 2873 (w), 1738 (w), 1650 (vs), 1523 (s), 1461 (w), 1377 (m), 1288 (w), 1254 (w), 1196 (m), 1159 (w), 1096 (s), 1049 (w), 1017 (w), 946 (w), 932 (w), 900 (w), 872 (w), 844 (w), 787 (w), 755 (w), 691 (w), 653 (w) cm⁻¹. ESMS (70 eV): m/z (%) = 437 (12) [M]Na+, 415 (30) [M]H+, 357 (100) [M + H - C₃H₆O]+.



E-hex-2-enoic pantetheine dimethyl ketal 11Pa



E-Hex-2-enoic acid (11, 0.11 g, 1.00 mmol) and pantetheine dimethyl ketal (0.32 g, 1.00 mmol) were dissolved in CH₂Cl₂ (8 ml). Then the mixture was cooled to 0 °C. Then N,N-dimethylaminopyridine (0.1 g, 0.80 mmol) and N-(3-Diethyl-aminopropyl)-N-ethyl-carbodiimide (0.38 g, 2.00 mmol) were added to the mixture. The mixture was warmed to 25 °C and then stirred for 4 h. After that the mixture was quenched with 2M HCl (10 ml) and extracted with CH₂Cl₂ (3x 20 ml). The organic layer was washed with saturated NaHCO₃ (25 ml) and brine (25 ml). The product was dried over MgSO₄ and concentrated in vacuo. The crude product was purified by column chromatography (ethyl acetate). The obtained product was a yellow oil (0.32 g, 0.70 mmol, 70 %). R_f: 0.49 (ethyl acetate). ¹H-NMR (CDCl₃, 400 MHz): δ_H: 0.92-0.96 (m, 3H, 1-CH₃); 0.97 (s, 3H, 19-CH₃); 1.03 (s, 3H, 18-CH₃); 1.41 (s, 3H, 22-CH₃); 1.46 (s, 3H, 21-CH₃); 1.50 (m, 2H, 2-CH₂); 2.19 (dq, J = 1.5, 7.3 Hz, 2H, 3-CH₂); 2.42 (t, J = 6.3 Hz, 2H, 11-CH₂); 3.08 (t, J = 6.3 Hz, 2H, 7-CH₂); 3.27 (d, J = 11.6 Hz, 1H, 17a-CH₂); 3.40-3.62 (m, 4H, 8-12-CH₂); 3.68 (d, J = 11.6 Hz, 1H, 17b-CH₂); 4.07 (s,1H, 15-CH); 6.10 (bt, J = 1.6 Hz, 1H, 5-CH); 6.14 (bt, J = 1.5 Hz, 1H, 9-NH); 6.92 (dt, J = 6.9, 15.6 Hz, 1H, 4-CH); 7.03 (bt, J = 5.9 Hz, 1H, 13-NH). ¹³C-NMR (CDCl₃, 100 MHz): δ_{C} 13.7 (1-CH₃); 18.7 (19-CH₃); 18.9 (21-CH₃); 21.2 (2-CH₂); 22.1 (18-CH₃); 28.2 (7-CH₂); 29.5 (22-CH₃); 32.9 (16-C); 34.2 (3-CH₂); 34.8 (11-CH₂); 35.9 (12-CH₂); 39.7 (8-CH₂); 29.5 (22-CH₃); 29.5 CH₂); 71.5 (17-CH₂); 77.3 (15-CH); 99.1 (20-C); 128.4 (5-CH); 146.6 (4-CH); 170.0 (10-CO); 171.2 (14-CO); 190.2 (6-CO). ESMS: *m/z* (%): 415.4 [M]H⁺(100 %). IR (v_{max} / cm⁻¹): 3306 (NH), 2960 (CH₂), 2872 (CH₂), 1655 (CO), 1523 (C-O), 1461 (C=C). HRMS: m/z (%): calculated 437.2086, found 437.2088 for [M]Na+.



E-2-methyl hexenoic pantetheine dimethyl ketal 12Pa



E-2-Methylhex-2-enoic acid (12, 0.13 g, 1 mmol) and pantetheine dimethyl ketal (0.32 g, 1.00 mmol) were dissolved in CH₂Cl₂ (8 ml). The mixture was cooled to 0 °C. Then N,N-dimethylaminopyridine (0.10 g, 0.80 mmol) and N-(3-Diethylaminopropyl)-N-ethylcarbodiimide (2.00 mmol, 0.38 g) were added. The mixture was warmed to 25 °C and stirred for 8 h. After that the mixture was guenched with 2M HCl (10 ml) and extracted with CH_2Cl_2 (3 × 25 ml). The organic layer was washed with saturated NaHCO₃ (25 ml) and brine (25 ml). The product was dried over MgSO₄ and concentrated in vacuo. The crude product was purified by column chromatography (ethyl acetate). The obtained product was a yellow oil (0.38 g, 0.88 mmol, 88 %). $R_f: 0.45$ (ethyl acetate). ¹H-NMR (CDCl₃, 400 MHz): δ H 0.96 (t, J = 7.6Hz, 3H, 1-CH₃); 0.97 (s, 3H, 17-CH₃); 1.04 (s, 3H, 18-CH₃); 1.44-1.55 (m, 2H, 2-CH₂); 1.41 (s, 3H, 20-CH₃); 1.46 (s, 3H, 21-CH₃); 1.87 (s, 3H, 6-CH₃); 2.20 (q, J = 7.4 Hz, 2H, 3-CH₂); 2.42 (t, J = 6.1 Hz, 2H, 10-CH₂); 3.05 (t, J = 6.5 Hz, 2H, 8-CH₂); 3.27 (d, J = 11.2 Hz, 1H, 16a-CH₂); 3.39-3.62 (m, 4H, 9-12-CH₂); 3.68 (d, J = 11.7 Hz, 1H, 16b-CH₂); 4.07 (s,1H, 14-CH); 6.10 (bt, J = 5.7 Hz, 1H, 10-NH); 6.77 (dt, J = 1.3, 6.9 Hz, 1H, 4-CH); 7.03 (bt, J = 5.7 Hz, 1H, 13-NH). ¹³C-NMR (CDCl₃, 100 MHz): δ_C 12.5 (6-CH₃); 13.9 (1-CH₃); 18.7 (17-CH₃); 18.9 (18-CH₃); 21.8 (2-CH₂); 22.1 (20-CH₃); 28.4 (3-CH₂); 28.8 (8-CH₂); 29.5 (21-CH₃); 30.9 (7-CH₂); 32.9 (15-C); 34.8 (11-CH₂); 35.9 (12-CH₂); 39.7 (9-CH₂); 71.5 (16-CH₂); 77.2 (14-CH); 99.1 (19-C); 135.9 (5-C); 142.1 (4-CH); 170.0 (10-CO); 171.2 (13-CO); 193.7 (7-CO). ESMS: m/z (%) 429.7 [M]H⁺(100 %). IR (v_{max} / cm⁻¹): 3293 (NH), 2959 (CH₂), 2870 (CH₂), 1648 (CO), 1538 (C-O), 1449 (C=C). HRMS: m/z (%): calculated 451.2243, found 451.2244 for [M]Na+.



E-4RS-4-methylhex-2-enoic pantetheine dimethyl ketal 13Pa



4RS-4-Methylhex-2-enoic acid (13, 0.13 g, 1.00 mmol) and pantetheine dimethyl ketal (0.33 g, 1.00 mmol) were dissolved in CH₂Cl₂ (8 ml). The mixture was cooled to 0 °C. Then N,N-dimethylaminopyridine (0.10 g, 0.80 mmol) and N-(3-Diethylaminopropyl)-N-ethylcarbodiimide (0.38 g, 2.00 mmol) were added. The mixture was warmed to 25 °C and stirred for 8 h. After that the mixture was quenched with 2M HCl (10 ml) and extracted with CH₂Cl₂ (3x 20 ml). The organic layer was washed with saturated NaHCO₃ (20 ml) and brine (20 ml). The product was dried over MgSO₄ and concentrated in vacuo. The crude product was purified by column chromatography (ethyl acetate). The obtained product was a colourless oil (0.35 g, 0.82 mmol, 82 %). $R_f: 0.45$ (ethyl acetate). ¹H-NMR (CDCl₃, 400 MHz): $\delta_H 0.88$ (t, J = 7.2 Hz, 3H, 1-CH₃); 0.97 (s, 3H, 17-CH₃); 1.04 (s, 3H, 2-CH₃); 1.06 (d, J = 6.7 Hz, 1H, 4-CH₃), 1.40-1.45 (m, 2H, 2-CH₂); 1.41 (s, 3H, 20-CH₃); 1.46 (s, 3H, 21-CH₃); 2.17-2.26 (m, 1H, 3-CH); 2.42 (t, J = 6.3 Hz, 2H, 11-CH₂); 3.05 (t, J = 6.3 Hz, 2H, 8-CH₂); 3.27 (d, J = 11.9 Hz, 1H, 16a-CH₂); 3.40-3.62 (m, 4H, 9-12-CH₂); 3.68 (d, J = 11.7 Hz, 1H, 16b-CH₂); 4.07 (s,1H, 14-CH); 6.08 (dd, J = 1.2, 15.6 Hz, 1H, 6-CH); 6.12 (bt, J = 5.1 Hz, 1H, 10-NH); 6.77 (dd, J = 7.5, 15.5 Hz, 1H, 5-CH); 7.03 (bt, J = 6.0 Hz, 1H, 13-NH). ¹³C-NMR (CDCl₃, 100 MHz): δ_{C} 11.6 (1-CH₃); 18.7 (18-CH₃); 18.9 (4-CH₃); 19.1 (21-CH₃); 22.1 (17-CH₃); 28.3 (2-CH₂); 28.4 (8-CH₂); 28.7 (2-CH₂); 29.5 (21-CH₃); 32.9 (15-C); 34.8 (11-CH₂); 35.9 (12-CH₂); 38.1 (3-CH); 39.7 (9-CH₂); 71.5 (16-CH₂); 77.2 (14-CH); 99.1 (19-C); 126.7 (6-CH₂); 71.5 (16-CH₂); 77.2 (14-CH); 99.1 (19-C); 126.7 (6-CH₂); 71.5 (16-CH₂); 71.5 CH); 151.7 (5-CH); 170.0 (10-CO); 171.2 (13-CO); 190.3 (7-CO). ESMS: *m/z* (%): 429.7 [M]H⁺(100 %). IR (v_{max} / cm⁻ ¹): 3305 (NH), 2961 (CH₂), 2873 (CH₂), 1655 (CO), 1522 (C-O), 1460 (C=C). HRMS: *m/z* (%): calculated 451.2243, found 451.2243 for [M]Na+.



E-2RS-2,4-dimethylhex-2-enoic pantetheine dimethyl ketal 14Pa



2RS-2,4-dimethylhex-2-enoic acid (14, 0.14 g, 1.00 mmol) and pantetheine dimethyl ketal (0.32 g, 1.00 mmol) were dissolved in CH₂Cl₂ (8 ml). The mixture was cooled to 0 °C. Then N,N-dimethylaminopyridine (0.10 g, 0.80 mmol) and N-(3-Diethylamino-propyl)-N-ethylcarbodiimide (0.38 g, 2.00 mmol) were added. The mixture was warmed to 25 °C and stirred for 8 h. After that the mixture was quenched with 2M HCl (10 ml) and extracted with CH₂Cl₂ (3x 25 ml). The organic layer was washed with saturated NaHCO₃ (20 ml) and brine (20 ml). The product was dried over MgSO₄ and concentrated in vacuo. The crude product was purified by column chromatography (ethyl acetate). The obtained product was a colourless oil (0.38 g, 0.89 mmol, 89 %). $R_f 0.45$ (ethyl acetate). ¹H-NMR (CDCl₃, 400 MHz): $\delta_H 0.87$ (t, J = 7.9 Hz, 3H, 1-CH₃); 0.97 (s, 3H, 17-CH₃); 1.04 (s, 3H, 18-CH₃); 1.03 (d, J = 7.5 Hz, 1H, 4-CH₃), 1.24-1.49 (m, 2H, $2-CH_2$; 1.41 (s, 3H, 21-CH₃); 1.46 (s, 3H, 22-CH₃); 1.88 (s, 3H, 7-CH₃); 2.42 (t, J = 5.9 Hz, 2H, 12-CH₂); 3.05 (t, J = 5.9 Hz, 2H, 6.4 Hz, 2H, 9-CH₂); 3.27 (d, J = 11.4 Hz, 1H, 19a-CH₂); 3.39-3.64 (m, 4H, 10-13-CH₂); 3.68 (d, J = 11.7 Hz, 1H, 19b-CH₂); 4.07 (s,1H, 15-CH); 6.09 (bt, *J* = 5.4 Hz, 1H, 11-NH); 6.53 (dd, *J* = 1.4, 9.8 Hz, 1H, 5-CH); 7.03 (bt, *J* = 5.8 Hz, 1H, 11-NH); 6.53 (dd, *J* = 1.4, 9.8 Hz, 1H, 5-CH); 7.03 (bt, *J* = 5.8 Hz, 1H, 11-NH); 6.53 (dd, *J* = 1.4, 9.8 Hz, 1H, 5-CH); 7.03 (bt, *J* = 5.8 Hz, 1H, 11-NH); 6.53 (dd, *J* = 1.4, 9.8 Hz, 1H, 5-CH); 7.03 (bt, *J* = 5.8 Hz, 1H, 11-NH); 6.53 (dd, *J* = 1.4, 9.8 Hz, 1H, 5-CH); 7.03 (bt, *J* = 5.8 Hz, 1H, 11-NH); 6.53 (dd, *J* = 1.4, 9.8 Hz, 1H, 5-CH); 7.03 (bt, *J* = 5.8 Hz, 1H, 5-CH); 7.03 (bt, J = 5.8 Hz, 1H, 5-1H, 14-NH). ¹³C-NMR (CDCl₃, 100 MHz): δ_C 11.9 (1-CH₃); 12.2 (4-CH₃); 18.7 (18-CH₃); 18.9 (21-CH₃); 22.1 (17-CH₃); 28.5 (2-CH₂); 29.5 (22-CH₃); 29.6 (9-CH₂); 32.9 (16-C); 34.4 (12-CH₂); 35.0 (3-CH); 35.9 (13-CH₂); 39.7 (10-CH₂); 71.5 (19-CH₂); 77.2 (15-CH); 99.1 (20-C); 134.5 (6-C); 147.6 (5-CH); 170.0 (11-CO); 171.2 (14-CO); 193.9 (8-CH); 170.0 (11-CO); 170.0 (11-CO); 171.2 (14-CO); 193.9 (8-CH); 170.0 (11-CO); 170.0 (11-CO); 171.2 (14-CO); 193.9 (8-CH); 170.0 (11-CO); 171.2 (14-CO); 193.9 (8-CH); 170.0 (11-CO); 170.0 (11-CO CO). ESMS: m/z (%): 443.7 [M]H⁺(100 %). IR (v_{max} / cm⁻¹): 3291 (NH), 2923 (CH₂), 2872 (CH₂), 1651 (CO), 1544 (C-O), 1444 (C=C). HRMS: m/z (%): calculated 465.2399, found 465.2396 for [M]Na+.



E-4S-2,4-dimethylhex-2-enoic pantetheine dimethyl ketal 15Pa



E-4S-2,4-dimethylhex-2-enoic acid (15, 0.14 g, 1.00 mmol) and pantetheine dimethyl ketal (0.32 g, 1.00 mmol) were dissolved in CH₂Cl₂ (8 ml). The mixture was cooled to 0 °C. Then N,N-dimethylaminopyridine (0.10 g, 0.80 mmol) and N-(3-Diethylamino-propyl)-N-ethylcarbodiimide (0.38 g, 2.00 mmol) were added. The mixture was warmed to 25 °C and stirred for 8 h. After that the mixture was quenched with 2M HCl (10 ml) and extracted with CH₂Cl₂ (3x 25 ml). The organic layer was washed with saturated NaHCO₃ (20 ml) and brine (20 ml). The product was dried over MgSO₄ and concentrated in vacuo. The crude product was purified by column chromatography (ethyl acetate). The obtained product was a colourless oil (0.30 g, 0.68 mmol, 68 %). R_f 0.45 (ethyl acetate). ¹H-NMR (CDCl₃, 400 MHz): δ_H 0.91 (t, J = 7.9 Hz, 3H, 1-CH₃); 0.97 (s, 3H, 17-CH₃); 1.04 (s, 3H, 18-CH₃); 1.17 (d, J = 7.5 Hz, 1H, 4-CH₃), 1.24-1.49 (m, 2H, 2-CH₂); 1.42 (s, 3H, 21-CH₃); 1.46 (s, 3H, 22-CH₃); 1.88 (s, 3H, 7-CH₃); 2.42 (t, *J* = 6.1 Hz, 2H, 12-CH₂); 2.54-2.62 (m, 1H, 3-CH); 3.05 (t, J = 6.5 Hz, 2H, 9-CH₂); 3.27 (d, J = 11.7 Hz, 1H, 19a-CH₂); 3.38-3.62 (m, 4H, 10-13-CH₂); 3.68 (d, J = 12.1 Hz, 1H, 19b-CH₂); 4.08 (s,1H, 15-CH); 6.09 (bt, J = 5.4 Hz, 1H, 11-NH); 6.53 (dd, J = 1.5, 10.2 Hz, 1H, 5-CH); 7.03 (bt, J = 5.8 Hz, 1H, 14-NH). ¹³C-NMR (CDCl₃, 100 MHz): $\delta_{\rm C}$ 11.6(1-CH₃); 17.2 (4-CH₃); 18.7 (18-CH₃); 18.9 (21-CH₃); 22.1 (17-CH₃); 28.5 (2-CH₂); 29.5 (22-CH₃); 29.6 (9-CH₂); 32.9 (16-C); 34.8 (12-CH₂); 35.1 (3-CH); 35.9 (13-CH₂); 39.7 (10-CH₂); 71.5 (19-CH₂); 77.2 (15-CH); 99.1 (20-C); 135.6 (6-C); 147.6 (5-CH); 170.1 (11-CO); 171.1 (14-CO); 204.1 (8-CO). ESMS: m/z (%): 443.7 [M]H⁺(100 %). IR (v_{max} / cm⁻¹): 3302 (NH), 2960 (CH₂), 2873 (CH₂), 1649 (CO), 1521 (C-O), 1460 (C=C). HRMS: m/z (%): calculated 443.2580, found 443.2853 for [M]H+.



E-2-ethylhexenoic pantetheine dimethyl ketal 16Pa



E-2-Ethylhex-2-enoic acid (16, 0.13 g, 1 mmol) and pantetheine dimethyl ketal (0.32 g, 1.00 mmol) were dissolved in CH₂Cl₂ (8 ml). The mixture was cooled to 0 °C. Then N,N-dimethylaminopyridine (0.10 g, 0.80 mmol) and N-(3-Diethylaminopropyl)-N-ethylcarbodiimide (2.00 mmol, 0.38 g) were added. The mixture was warmed to 25 °C and stirred for 8 h. After that the mixture was guenched with 2M HCl (10 ml) and extracted with CH₂Cl₂ (3x 25 ml). The organic layer was washed with saturated NaHCO₃ (25 ml) and brine (25 ml). The product was dried over MgSO₄ and concentrated in vacuo. The crude product was purified by column chromatography (ethyl acetate). The obtained product was a yellow oil (0.38 g, 0.88 mmol, 88 %). R_f: 0.45 (ethyl acetate). ¹H-NMR (CDCl₃, 400 MHz): δ_H 0.86-1.08 (m, 1-7-18-19-CH₃); 1.40-1.53 (m, 2H, 2-CH₂); 1.41 (s, 3H, 21-CH₃); 1.46 (s, 3H, 22-CH₃); 2.20 (q, *J* = 7.5 Hz, 2H, 3-CH₂); 2.31-2.38 (m, 2H, 11-CH_{2E}); 2.40-2.44 (m, 2H, 6-CH_{2Z}); 3.03-3.09 (m, 2H, 9-CH₂); 3.27 (d, *J* = 11.2 Hz, 1H, 17a-CH₂); 3.39-3.64 (m, 4H, 10-13-CH₂); 3.68 (d, J = 11.7 Hz, 1H, 17b-CH₂); 4.07 (s,1H, 15-CH); 5.62 (t, J = 7.7 Hz, 1H, 4-CH_Z); 6.12 (bt, J = 5.7 Hz, 1H, 11-NH); 6.71 (t, J = 7.5 Hz, 1H, 4-CH_E); 7.04 (bt, J = 5.7 Hz, 1H, 14-NH). ¹³C-NMR (CDCl₃, 100 MHz): δ_C 13.9 (1-CH₃); 14.1 (22-CH₃); 18.7 (18-CH₃); 18.9 (22-CH₃); 20.5 (6-CH₂); 22.2 (2-CH₂); 22.1 (19-CH₃); 28.4 (9-CH₂); 28.8 (3-CH₂); 29.5 (21-CH₃); 30.9 (3-CH₂); 32.9 (15-C); 34.8 (12-CH₂); 35.9 (13-CH₂); 39.7 (10-CH₂); 71.5 (17-CH₂); 77.2 (15-CH); 99.1 (20-C); 135.9 (5-C); 142.1 (4-CH); 170.0 (11-CO); 171.2 (14-CO); 193.7 (8-CO). ESMS: m/z (%) 465.4 [M]Na⁺(51 %), 443.7 [M]H⁺(28 %), 385.4 [M - C₄H₈]H⁺(100 %). HRMS: m/z (%): calculated 465.2399, found 465.2396 for [M]Na+.



E-6RS-6-methyl hept-2-enoic pantetheine dimethyl ketal 18Pa



E-6RS-6-Methyl heptenoic acid (18, 0.16 g, 1.00 mmol) and pantetheine dimethyl ketal (0.32 g, 1.00 mmol), were dissolved in CH₂Cl₂ (8 ml). The mixture was cooled to 0 °C. Then N,N-dimethylaminopyridine (0.10 g, 0.80 mmol,) and N-(3-Diethylamino-propyl)-N-ethylcarbodiimide (0.38 g, 2.00 mmol) were added. The mixture was warmed to 25 °C and stirred for 4 h. After that the mixture was guenched with 2M HCl (10 ml) and extracted with CH₂Cl₂ (3x 25 ml). The organic layer was washed with saturated NaHCO₃ (20 ml) and brine (20 ml). The product was dried over $MgSO_4$ and concentrated in vacuo. The crude product was purified by column chromatography (ethyl acetate). The obtained product was a colourless oil (0.22 g, 0.52 mmol, 52 %). R_f: 0.53 (ethyl acetate). ¹H-NMR (CDCl₃, 400 MHz): 22-CH₃); 1.46 (s, 3H, 21-CH₃); 2.22 (dq, J = 1.9, 7.1 Hz, 2H, 5-CH₂); 2.42 (t, J = 6.1 Hz, 2H, 12-CH₂); 3.08 (t, J = 6.4Hz, 2H, 9-CH₂); 3.28 (d, J = 11.7 Hz, 1H, 17a-CH₂); 3.40-3.62 (m, 4H, 10-13-CH₂); 3.68 (d, J = 11.6 Hz, 1H, 17b-CH₂); 4.07 (s,1H, 15-CH); 6.10 (bt, *J* = 1.6 Hz, 1H, 11-NH); 6.14 (t, *J* = 1.5 Hz, 1H, 7-CH); 6.93 (dt, *J* = 7.1, 15.6 Hz, 1H, 6-CH); 7.02 (bt, J = 6.0 Hz, 1H, 14-NH). ¹³C-NMR (CDCl₃, 100 MHz): δ_{C} 18.7 (19-CH₃); 18.9 (21-CH₃); 22.1 (20-CH₃); 22.4 (1-CH₃); 22.4 (3-CH₃); 27.6 (2-CH); 28.2 (9-CH₂); 29.5 (22-CH₃); 30.1 (5-CH₂); 32.9 (16-C); 34.8 (12-CH₃); 20.1 (5-CH₂); 32.9 (16-C); 34.8 (12-CH₃); 30.1 (5-CH₂); 30 CH₂); 35.9 (13-CH₂); 36.9 (4-CH₂); 39.7 (10-CH₂); 71.5 (17-CH₂); 77.2 (15-CH); 99.1 (18-C); 128.2 (7-CH); 147.1 (6-CH); 170.1 (11-CO); 171.4 (14-CO); 190.2 (8-CO). ESMS: *m/z* (%): 465.3 [M]Na⁺(28 %), 443.3 [M]H⁺(12 %), 385 $[M + H - C_3H_6O] + (100 \%)$. IR (v_{max} / cm^{-1}): 3303 (NH), 2953 (CH₂), 2870 (CH₂), 1655 (CO), 1524 (C-O), 1465 (C=C). HRMS: *m/z* (%): calculated 465.2399, found 465.2396 for [M]Na+.



E-2-methyloct-2-enoic pantetheine dimethyl ketal 20Pa



E-2-Methyloct-2-enoic acid (20, 0.16 g, 1.00 mmol) and pantetheine dimethyl ketal (0.32 g, 1.00 mmol) were dissolved in CH₂Cl₂ (8 ml). The mixture was cooled to 0 °C. Then N,N-dimethylaminopyridine (0.10 g, 0.80 mmol) and N-(3-Diethyl-aminopropyl)-N-ethylcarbodiimide (0.38 g, 2.00 mmol) were added. The mixture was warmed to 25 °C and stirred for 4 h. After that the mixture was guenched with 2M HCl (10 ml) and extracted with CH₂Cl₂ (3x 25 ml). The organic layer was washed with saturated NaHCO₃ (20 ml) and brine (20 ml). The product was dried over MgSO₄ and concentrated in vacuo. The crude product was purified by column chromatography (ethyl acetate). The obtained product was a colourless oil (0.27 g, 0.60 mmol, 60 %). R_f: 0.47 (ethyl acetate). ¹H-NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 0.90 (t, J = 7.4Hz, 3H, 1-CH₃); 0.97 (s, 3H, 21-CH₃); 1.04 (s, 3H, 19-CH₃); 1.31-1.51 (m, 6H, 2-3-4-CH₂); 1.41 (s, 3H, 25-CH₃); 1.46 $(s, 3H, 24-CH_3)$; 1.87 $(s, 3H, 8-CH_3)$; 2.21 $(g, J = 7.3 Hz, 2H, 5-CH_2)$; 2.42 $(t, J = 5.8 Hz, 2H, 14-CH_2)$; 3.05 $(t, J = 6.7 Hz, 2H, 14-CH_2)$; 3.05 Hz, 2H, 10-CH₂); 3.27 (d, J = 11.8 Hz, 1H, 22a-CH₂); 3.39-3.62 (m, 4H, 11-15-CH₂); 3.68 (d, J = 12.2 Hz, 1H, 22b-CH₂); 4.07 (s,1H, 18-CH); 6.08 (bt, *J* = 5.8 Hz, 1H, 12-NH); 6.92 (dt, *J* = 1.5, 7.4 Hz, 1H, 6-CH); 7.03 (bt, *J* = 5.8 Hz, 1H, 12-NH); 6.92 (dt, *J* = 1.5, 7.4 Hz, 1H, 6-CH); 7.03 (bt, *J* = 5.8 Hz, 1H, 12-NH); 6.92 (dt, *J* = 1.5, 7.4 Hz, 1H, 6-CH); 7.03 (bt, *J* = 5.8 Hz, 1H, 12-NH); 6.92 (dt, *J* = 1.5, 7.4 Hz, 1H, 6-CH); 7.03 (bt, *J* = 5.8 Hz, 1H, 12-NH); 6.92 (dt, *J* = 1.5, 7.4 Hz, 1H, 6-CH); 7.03 (bt, *J* = 5.8 Hz, 1H, 12-NH); 6.92 (dt, *J* = 1.5, 7.4 Hz, 1H, 6-CH); 7.03 (bt, *J* = 5.8 Hz, 1H, 12-NH); 6.92 (dt, *J* = 1.5, 7.4 Hz, 1H, 6-CH); 7.03 (bt, *J* = 5.8 Hz, 1H, 12-NH); 6.92 (dt, *J* = 1.5, 7.4 Hz, 1H, 6-CH); 7.03 (bt, *J* = 5.8 Hz, 1H, 12-NH); 6.92 (dt, *J* = 1.5, 7.4 Hz, 1H, 6-CH); 7.03 (bt, *J* = 5.8 Hz, 1H, 12-NH); 6.92 (dt, *J* = 1.5, 7.4 Hz, 1H, 6-CH); 7.03 (bt, *J* = 5.8 Hz, 1H, 12-NH); 6.92 (dt, *J* = 1.5, 7.4 Hz, 1H, 6-CH); 7.03 (bt, *J* = 5.8 Hz, 1H, 12-NH); 6.92 (dt, J = 1.5, 7.4 Hz, 1H, 6-CH); 7.03 (bt, J = 5.8 Hz, 1H, 12-NH); 6.92 (dt, J = 1.5, 7.4 Hz, 1H, 6-CH); 7.03 (bt, J = 5.8 Hz, 1H, 12-NH); 6.92 (dt, J = 1.5, 7.4 Hz, 1H, 6-CH); 7.03 (bt, J = 5.8 Hz, 1H, 12-NH); 6.92 (dt, J = 1.5, 7.4 Hz, 1H, 6-CH); 7.03 (bt, J = 5.8 Hz, 1H, 12-NH); 6.92 (dt, J = 1.5, 7.4 Hz, 1H, 6-CH); 7.03 (bt, J = 5.8 Hz, 1H, 12-NH); 6.92 (dt, J = 1.5, 7.4 Hz, 1H, 6-CH); 7.03 (bt, J = 5.8 Hz, 1H, 12-NH); 6.92 (dt, J = 1.5, 7.4 Hz, 1H, 6-CH); 7.03 (bt, J = 5.8 Hz, 1H, 12-NH); 6.92 (dt, J = 1.5, 7.4 Hz, 1H, 6-CH); 7.03 (bt, J = 5.8 Hz, 1H, 12-NH); 6.92 (dt, J = 1.5, 7.4 Hz, 1H, 6-CH); 7.03 (bt, J = 5.8 Hz, 1H, 12-NH); 6.92 (dt, J = 1.5, 7.4 Hz, 1H, 12-NH); 6.92 (dt, J = 1.5, 7.4 Hz, 1H, 12-NH); 6.92 (dt, J = 1.5, 7.4 Hz, 1H, 12-NH); 6.92 (dt, J = 1.5, 7.4 Hz, 1H, 12-NH); 6.92 (dt, J = 1.5, 7.4 Hz, 1H, 12-NH); 6.92 (dt, J = 1.5, 7.4 Hz, 1H, 12-NH); 7.03 (dt, J = 1.5, 7.4 Hz, 1H, 12-NH); 7.03 (dt, J = 1.5, 7.4 Hz, 1H, 12-NH); 7.03 (dt, J = 1.5, 7.4 Hz, 14-NH); 7.04 (dt, J 1H, 16-NH). ¹³C-NMR (CDCl₃, 100 MHz): δ_C 12.5 (8-CH₃); 13.9 (1-CH₃); 18.7 (19-CH₃); 18.9 (21-CH₃); 22.1 (24-CH₃); 22.5 (2-CH₃); 28.2 (4-CH₂); 28.4 (3-CH₂); 28.8 (10-CH₂); 29.5 (25-CH₃); 31.6 (5-CH₂); 32.9 (20-C); 34.8 (14-CH₂); 35.9 (15-CH₂); 39.7 (11-CH₂); 71.5 (22-CH₂); 77.2 (18-CH); 99.1 (23-C); 135.7 (7-C); 142.4 (6-CH); 170.0 (13-CO); 171.1 (17-CO); 193.7 (9-CO). ESMS: *m/z* (%): 457.7 [M]H⁺(100 %). IR (v_{max} / cm⁻¹): 3306 (NH), 2929 (CH₂), 2860 (CH₂), 1652 (CO), 1523 (C-O), 1461 (C=C). HRMS: *m/z* (%): calculated 479.2556, found 479.2552 for [M]Na+. CBHA0283/10 3 5 5 5 5 Methylogtenoic pant



E-7-methyloct-2-enoic pantetheine dimethyl ketal 21Pa



E-7-Methyloct-2-enoic acid (21, 0.16 g, 1.00 mmol) and pantetheine dimethyl ketal (0.32 g, 1.00 mmol,) were dissolved in CH₂Cl₂ (8 ml). The mixture was cooled to 0 °C. Then N.N-dimethylaminopyridine (0.10 g, 0.80 mmol,) and N-(3-Diethylamino-propyl)-N-ethylcarbodiimide (0.38 g, 2.00 mmol) were added. The mixture was warmed to 25 °C and stirred for 4 h. After that the mixture was quenched with 2M HCl (10 ml) and extracted with CH₂Cl₂ (3x 25 ml). The organic layer was washed with saturated NaHCO₃ (20 ml) and brine (20 ml). The product was dried over MgSO₄ and concentrated in vacuo. The crude product was purified by column chromatography (ethyl acetate). The obtained product was a colourless oil (0.18 g, 0.40 mmol, 40 %). R_{f} : 0.43 (ethyl acetate). ¹H-NMR (CDCl₃, 400 MHz): δ_{H} 0.89 (d, J = 6.7Hz, 6H, 1-3-CH₃); 0.97 (s, 3H, 20-CH₃); 1.04 (s, 3H, 19-CH₃); 1.42 (s, 3H, 22-CH₃); 1.46 (s, 3H, 21-CH₃); 1.43-1.62 (m, 5H, 2-CH-3-4-CH₂); 2.22 (dq, J = 1.7, 7.3 Hz, 2H, 5-CH₂); 2.42 (t, J = 6.3 Hz, 2H, 12-CH₂); 3.08 (t, J = 6.3 Hz, 2H, 9-CH₂); 3.28 (d, J = 11.7 Hz, 1H, 17a-CH₂); 3.42-3.64 (m, 4H, 10-13-CH₂); 3.68 (d, J = 11.6 Hz, 1H, 17b-CH₂); 4.08 (s,1H, 15-CH); 6.10 (bt, J = 1.6 Hz, 1H, 11-NH); 6.17 (t, J = 1.5 Hz, 1H, 7-CH); 6.93 (dt, J = 6.9, 15.4 Hz, 1H, 6-CH); 7.02 (bt, J = 6.2 Hz, 1H, 14-NH). ¹³C-NMR (CDCl₃, 100 MHz): δ_{C} 18.7 (19-CH₃); 18.9 (21-CH₃); 22.1 (20-CH₃); 22.4 (1-CH₃); 22.4 (23-CH₃); 25.8 (3-CH₂); 27.8 (2-CH); 28.2 (9-CH₂); 29.5 (22-CH₃); 32.5 (5-CH₂); 32.9 (16-C); 34.8 (12-CH₂); 35.9 (13-CH₂); 39.7 (10-CH₂); 71.5 (17-CH₂); 77.2 (15-CH); 99.1 (18-C); 128.4 (7-CH); 147.1 (6-CH); 170.1 (11-CO); 171.4 (14-CO); 190.2 (8-CO). ESMS: m/z (%): 479.3 [M]Na + (5 %), 457.4 [M]H + (18 %), 399.4 [M - $C_{3}H_{6}O$]+ (100 %). IR (v_{max} / cm⁻¹): 3307 (NH), 2952 (CH₂), 2869 (CH₂), 1655 (CO), 1524 (C-O), 1463 (C=C). HRMS: *m*/*z* (%): calculated 479.2556, found 479.2557 for [M]Na+.



E-4RS, 6RS-4,6-dimethyloct-2-en-oyl-pantetheine 22Pa



E-4RS,6RS-4,6-dimethyloct-2-enoic acid (22, 0.17 g, 1.00 mmol) and pantetheine dimethyl ketal (0.32 g, 1.00 mmol,) were dissolved in CH₂Cl₂ (8 ml). The mixture was cooled to 0 °C. Then N,N-dimethylaminopyridine (0.10 g, 0.80 mmol,) and N-(3-Diethylamino-propyl)-N-ethylcarbodiimide (0.38 g, 2.00 mmol) were added. The mixture was warmed to 25 °C and stirred for 4 h. After that the mixture was quenched with 2M HCl (10 ml) and extracted with CH₂Cl₂ (3 x 25 ml). The organic layer was washed with saturated NaHCO₃ (20 ml) and brine (20 ml). The product was dried over MgSO₄ and concentrated in vacuo. The crude product was purified by column chromatography (ethyl acetate). The obtained product was a colourless oil (0.32 g, 0.65 mmol, 65 %). R_f: 0.46 (ethyl acetate). ¹H-NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 0.83 (m, 6H, 1-3-CH₃); 0.97 (s, 3H, 20-CH₃); 1.04 (s, 3H, 21-CH₃); 1.14 (m, , 3H, 7-CH₃), 1.24-1.49 (m, 5H, 2-4-5-CH₂); 1.42 (s, 3H, 23-CH₃); 1.46 (s, 3H, 24-CH₃); 1.71-1.78 (m, 1H, 6-CH); 2.42 (t, J = 6.1 Hz, 2H,14-CH₂); 2.54-2.62 (m, 1H, 6-CH); 3.05 (t, J = 6.5 Hz, 2H, 11-CH₂); 3.27 (d, J = 11.7 Hz, 1H, 19a-CH₂); 3.38-3.62 (m, 4H, 12-15-CH₂); 3.68 (d, J = 12.1 Hz, 1H, 19b-CH₂); 4.07 (s,1H, 17-CH); 6.05 (m, 1H, 9-CH); 6.12 (bt, J = 5.4 Hz, 1H, 13-NH); 6.74-6.88 (m, 1H, 8-CH); 7.03 (bt, J = 5.8 Hz, 1H, 16-NH). ¹³C-NMR (CDCl₃, 100 MHz): δ_{C} 11.1 (1-CH₃); 11.2 (3-CH₃); 17.6 (7-CH₃); 18.7 (20-CH₃); 18.9 (23-CH₃); 22.1 (21-CH₃); 28.2 (2-CH₂); 29.5 (24-CH₃); 29.6 (11-CH₂); 32.1 (4-CH); 32.9 (18-C); 34.7 (15-CH₂) 35.9 (14-CH₂); 39.7 (12-CH₂); 40.8 (5-CH), 46-5 (6-C) 71.5 (19-CH₂); 77.2 (17-CH); 99.1 (22-C); 126.9 (9-CH); 147.6 (8-CH); 170.1 (13-CO); 171.1 (16-CO); 204.1 (10-CO). ESMS: m/z (%): 471 [M]H⁺(100%) IR (v_{max} / cm⁻¹): 2958 (CH), 2927 (CH₂), 2872 (CH₂), 1654 (CO), 1629 (CO), 1523 (C-O), 1460 (C=C). HRMS: *m/z* (%): calculated 471.2893, found 471.2891 for [M]H+.

E-4S,6S-4,6-dimethyloct-2-enoyl-pantetheine 1Pa



E-4S,6S-4,6-dimethyloct-2-enoic acid (1, 0.17 g, 1.00 mmol) and pantetheine dimethyl ketal (0.32 g, 1.00 mmol,) were dissolved in CH₂Cl₂ (8 ml). The mixture was cooled to 0 °C. Then N,N-dimethylaminopyridine (0.10 g, 0.80 mmol,) and N-(3-Diethylamino-propyl)-N-ethylcarbodiimide (0.38 g, 2.00 mmol) were added. The mixture was warmed to 25 °C and stirred for 4 h. After that the mixture was guenched with 2M HCl (10 ml) and extracted with CH₂Cl₂ (3 x 25 ml). The organic layer was washed with saturated NaHCO₃ (20 ml) and brine (20 ml). The product was dried over $MgSO_4$ and concentrated *in vacuo*. The crude product was purified by column chromatography (ethyl acetate). The obtained product was a colourless oil (0.34 g, 0.70 mmol, 70 %). R_f: 0.46 (ethyl acetate). ¹H-NMR (CDCl₃, 400 MHz): δ_H 0.87 (m, 6H, 1-3-CH₃); 0.97 (s, 3H, 20-CH₃); 1.04 (s, 3H, 21-CH₃); 1.13-1.19 (m, 3H, 7-CH₃), 1.26-1.49 (m, 5H, 2-4-5-CH₂); 1.44 (s, 3H, 23-CH₃); 1.48 (s, 3H, 24-CH₃); 2.45 (t, J = 5.9 Hz, 1H, 14-CH); 3.10 (t, J = 6.5 Hz, 2H, 11-CH₂); 3.27 (d, J = 11.7 Hz, 1H, 19a-CH₂); 3.42-3.62 (m, 4H, 12-15-CH₂); 3.70 (d, J = 11.6 Hz, 1H, 19b-CH₂); 4.09 (s, 1H, 17-CH); 6.12 (dd, J = 1.0, 16.0 Hz, 1H, 9-CH); 6.28 (bt, J = 6.3 Hz, 1H, 13-NH); 6.80 (dd, J = 8.2, 15.8 Hz, 1H, 14.2)8-CH); 7.06 (bt, J = 5.8 Hz, 1H, 16-NH). ¹³C-NMR (CDCl₃, 100 MHz): δ_{C} 11.1 (1-CH₃); 14.2 (3-CH₃); 18.7 (20-CH₃); 18.9 (23-CH₃); 22.1 (21-CH₃); 28.2 (2-CH₂); 29.5 (24-CH₃); 29.7 (3-CH₂); 29.7 (11-CH₂); 34.4 (4-CH); 32.9 (18-C); 34.7 (15-CH₂) 35.9 (14-CH₂); 39.7 (12-CH₂); 43-5 (6-C) 71.5 (19-CH₂); 77.2 (17-CH); 99.1 (22-C); 126.6 (9-CH); 152.1 (8-CH); 170.1 (13-CO); 171.1 (16-CO); 190.1 (10-CO). ESMS: *m/z* (%): 471 [M]H⁺(100%), 413 [M - C₃H₆O]H ⁺ (12%). IR (v_{max} / cm⁻¹): 2958 (CH), 2927 (CH₂), 2872 (CH₂), 1654 (CO), 1629 (CO), 1523 (C-O), 1460 (C=C). HRMS: *m/z* (%): calculated 493.2712 found 493.2619 for [M]Na+.



E-2-methyldec-2-enoic pantetheine dimethyl ketal 25Pa



E-2-Methyldec-2-enoic acid (25, 0.184 g, 1.00 mmol) and pantetheine dimethyl ketal (0.32 g, 1.00 mmol) were dissolved in CH₂Cl₂ (8 ml). The mixture was cooled to 0 °C. Then N,N-dimethylaminopyridine (0.10 g, 0.80 mmol) and N-(3-Diethylaminopropyl)-N-ethylcarbodiimide (0.38 g, 2.00 mmol) were added. The mixture was warmed to 25 °C and stirred for 4 h. After that the mixture was guenched with 2M HCl (10 ml) and extracted with CH₂Cl₂ (3 x 25 ml). The organic layer was washed with saturated NaHCO₃ (20 ml) and brine (20 ml). The product was dried over MgSO₄ and concentrated in vacuo. The crude product was purified by column chromatography (ethyl acetate). The obtained product was a colourless oil (0.28 g, 0.58 mmol, 58 %). R_f 0.45 (ethyl acetate). ¹H-NMR (CDCl₃, 400 MHz): δ_H 0.88 (t, $27-CH_3$; 1.46 (s, 3H, 26-CH₃); 1.87 (s, 3H, 10-CH₃); 2.21 (g, J = 7.1 Hz, 2H, 7-CH₂); 2.42 (t, J = 6.2 Hz, 2H, 13-CH₂); 3.05 (t, J = 6.2 Hz, 2H, 12-CH₂); 3.27 (d, J = 12.2 Hz, 1H, 24a-CH₂); 3.39-3.62 (m, 4H, 13-17-CH₂); 3.68 (d, J = 12.2Hz, 1H, 24b-CH₂); 4.07 (s,1H, 20-CH); 6.08 (bt, *J* = 5.0 Hz, 1H, 14-NH); 6.77 (dt, *J* = 1.9, 6.9 Hz, 1H, 8-CH); 7.03 (bt, J = 5.5 Hz, 1H, 18-NH). ¹³C-NMR (CDCl₃, 100 MHz): δ_{C} 12.5 (10-CH₃); 14.0 (1-CH₃); 18.7 (23-CH₃); 18.9 (21-CH₃); 22.1 (26-CH₃); 22.6 (2-CH₂); 28.4 (4-CH₂); 28.5 (5-CH₂); 28.8 (12-CH₂); 29.1 (3-CH₂); 29.5 (27-CH₃); 31.8 (6-CH₂); 32.9 (22-C); 34.8 (16-CH₂); 35.9 (17-CH₂); 39.7 (13-CH₂); 71.5 (24-CH₂); 77.2 (20-CH); 99.1 (25-C); 135.7 (9-C); 142.4 (8-CH); 170.0 (15-CO); 171.2 (19-CO); 193.7 (11-CO). ESMS: m/z (%): 485.7 [M]H⁺(100 %). IR (v_{max} / cm⁻¹): 3307 (NH), 2926 (CH₂), 2856 (CH₂), 1652 (CO), 1523 (C-O), 1461 (C=C). HRMS: m/z (%): calculated 507.2869, found 507.2870 for [M]Na+.





A solution of pantetheine (0.45 g, 0.80 mmol) and DTT (0.12 g, 0.80 mmol) in THF (4 ml) was stirred in a nitrogen atmosphäre at 30 °C for 3 h. After that triethylamine (0.3 ml) and DMAP (0.09 mmol, 12 mg) were added to the reaction and stirred for 15 min. Then the reaction was cooled down to 0 °C and tigloyl chloride (0.19 g, 1.60 mmol) was added to the reaction. The mixture was stirred for one hour at this temperature. The solvents were removed under a nitrogen flow. The product was purified by HPLC (acetonitrile) and concentrated under a nitrogen flow. The obtained product was a colourless oil (0.14 g, 0.40 mmol, 50 %). ¹H-NMR (CDCl₃, 400 MHz) $\delta_{\rm H}$ 0.92 (s, 3H, 13-CH₃); 0.92 (s, 3H, 12-CH₃); 1.82-1.87 (m, 6H, 1-CH₃, 4-CH₃); 2.41 (t, *J* = 5.9 Hz, 3H, 8-CH₂); 3.01-3.10 (m, 2H, 5-CH₂); 3.32-3.60 (m, 6H, 6-9-14-CH₂); 3.99 (s, ¹H, 11-CH); 6.33 (bt, *J* = 5.8 Hz, ¹H, 7-NH); 6.86 (qq, *J* = 1.3, 6.8 Hz, ¹H, 2-CH); 7.40 (bt, *J* = 5.6 Hz, ¹H, 10-NH). ¹³C-NMR (CDCl₃, 100 MHz): $\delta_{\rm C}$ 12.2 (4-CH₃); 14.5 (1-CH₃), 20.4 (13-CH₃); 21.5 (12-CH₃); 28.2 (5-CH₂); 35.2 (9-CH₂); 35.6 (8-CH₂); 39.3 (C); 39.9 (6-CH₂); 70.9 (14-CH₂); 77.9 (11-CH); 136.8 (3-C); 137.3 (2-CH); 171.8 (CO); 173.5 (CO); 194.1 (COS). ESMS *m/z* (%): 383 [M]Na⁺(8 %), 361 [M]H⁺(100%), 343 [M - H₂O]H⁺(2 %), 231 [M - C₆H₁₁O₃]H⁺(5 %). HRMS *m/z* (%): calculated 383.1617, found 383.1621 for [M]Na⁺.



2RS-2-methylbutyryl pantetheine 6P



A solution of pantetheine (0.15 g, 27 µmol) and DTT (0.04 g, 27 µmol) in THF (1 ml) was stirred in a nitrogen atmosphäre at 30 °C for 3 h. After that pyridine (0.1 ml) and DMAP (4 mg, 0.03 mmol) were added to the reaction and stirred for 15 min. Then the reaction was cooled down to 0 °C and (±) 2-methylbutyryl chloride (0.06 g, 54 µmol) was added to the reaction. The mixture was stirred for one hour at this temperature. The solvents were removed under a nitrogen flow. The product was purified by HPLC (acetonitrile) and concentrated under a nitrogen flow. The obtained product was a colourless oil (0.035 g, 0.10 mmol, 36 %) ¹H-NMR (CDCl₃, 400 MHz) $\delta_{\rm H}$ 0.90 (t, *J* = 7.6 Hz, 3H, 1-CH₃); 0.92 (s, 3H, 13-CH₃); 1.01 (s, 3H, 12-CH₃); 1.15 (d, *J* = 7.5 Hz, 3H, 4-CH₃); 1.36-1.78 (m, 2H, 2-CH₂); 2.40 (t, *J* = 6.1 Hz, 2H, 8-CH₂); 2.53-2.63 (m, ¹H, 3-CH); 2.96-3.05 (m, 2H, 5-CH₂); 3.37-3.60 (m, 6H, 6-9-14-CH₂); 3.99 (s, ¹H, 15-CH₂); 6.32 (t, *J* = 5.4 Hz, ¹H, 7-NH); 7.44 (bt, *J* = 5.9 Hz, ¹H, 10-NH). ¹³C-NMR (CDCl₃, 100 MHz) $\delta_{\rm C}$ 17.1 (4-CH₃), 20.4 (13-CH₃); 21.5 (12-CH₃); 27.1 (2-CH₂); 27.4 (1-CH₂); 27.9 (5-CH₂); 35.1 (9-CH₂); 35.6 (8-CH₂); 39.3 (C); 39.7 (6-CH₂); 50.2 (14-CH₂); 70.8 (11-CH); 172.5 (CO); 174.5 (CO); 205.1 (COS). ESMS *m/z* (%): 385 [M]Na⁺(3 %), 363 [M]H⁺(100 %), 345 [M - H₂O]H⁺(3 %), 233 [M - C₆H₁₁O₃]H⁺(8 %), 100 [C₅H₁₁O₂] (35 %) HRMS *m/z* (%): calculated 361.1797, found 361.1796 for [M - H]-.





A solution of pantetheine (0.45 g, 0.80 mmol) and DTT (0.13 g, 0.80 mmol) in THF (4 ml) was stirred in a nitrogen atmosphäre at 30 °C for 3 h. After that triethylamine (0.3 ml) and DMAP (0.01 g, 0.09 mmol) were added to the reaction and stirred for 15 min. Then the reaction was cooled down to 0 °C and crotonyl chloride (0.16 g, 1.66 mmol) was added to the reaction. The mixture was stirred for one hour at this temperature. The solvents were removed under a nitrogen flow. The product was purified by HPLC (acetonitrile) and concentrated under a nitrogen flow. The obtained product was a colourless oil (0.18 g, 0.50 mmol, 34%) ¹H-NMR (CDCl₃, 400 MHz) $\delta_{\rm H}$ 0.92 (s, 3H, 13-CH₃); 1.03 (s, 3H, 12-CH₃); 1.90 (dd, *J* = 1.3, 6.8 Hz, 3H, 1-CH₃); 2.41 (t, *J* = 5.9 Hz, 3H, 8-CH₂); 3.02-3.16 (m, 2H, 5-CH₂); 3.36-3.60 (m, 6H, 6-9-14-CH₂); 3.99 (s, ¹H, 11-CH); 6.13 (dd, *J* = 5.8 Hz, ¹H, 3-CH); 6.23 (bt, *J* = 5.8 Hz, ¹H, 7-NH); 6.95 (dq, *J* = 1.3, 6.8 Hz, ¹H, 2-CH); 7.36 (bt, *J* = 5.6 Hz, ¹H, 10-NH). ¹³C-NMR (CDCl₃, 100 MHz) $\delta_{\rm C}$ 18.1 (1-CH₃), 20.4 (13-CH₃); 21.7 (12-CH₃); 28.1 (5-CH₂); 35.2 (9-CH₂); 35.6 (8-CH₂); 39.3 (C); 39.9 (6-CH₂); 70.9 (14-CH₂); 77.9 (11-CH); 129.7 (3-CH); 142.3 (2-CH); 171.8 (CO); 173.5 (CO); 190.5 (COS). ESMS *m/z* (%): 715.9 [M₂]Na⁺(5 %), 693 [M₂]H⁺(48 %), 369 [M]Na⁺(2 %), 347 [M]H⁺(100 %), 329 [M - H₂O]H⁺(2 %). HRMS *m/z* (%): calculated 369.1460, found 369.1461 for [M]Na+.

angeloyl-pantetheine 8P



Angeloyl pantetheine dimethyl ketal (**8Pa**, 0.02 g, 0.002 mmol) stirred in a mixture of acetonitrile and water (1:1) and 10 % TFA for 20 min. The reaction was followed by TLC and LCMS. After that the solvents were liphophilized. 0.03 g of the product were purified by HPLC. ¹H-NMR (CDCl₃, 400 MHz) $\delta_{\rm H}$ 0.92 (s, 3H, 13-CH₃); 1.01 (s, 3H, 12-CH₃); 1.83-1.90 (m, 6H, 1-CH₃, 4-CH₃); 2.40-2.44 (m, 2H, 8-CH₂); 2.99-3.14 (m, 2H, 5-CH₂); 3.34-3.59 (m, 6H, 6-9-14-CH₂); 3.99 (s, ¹H, 11-CH); 5.90 (dq, *J* = 1.2, 7.3 Hz, ¹H, 2-CH); 6.35 (bt, *J* = 5.5 Hz, ¹H, 7-NH); 7.40 (bt, *J* = 6.1 Hz, ¹H, 10-NH). ¹³C-NMR (CDCl₃, 100 MHz) $\delta_{\rm C}$ 15.9 (1-CH₃), 20.4 (13-CH₃); 20.6 (4-CH₃); 21.5 (12-CH₃); 28.2 (5-CH₂); 35.2 (9-CH₂); 35.6 (8-CH₂); 39.4 (C); 39.8 (6-CH₂); 70.9 (14-CH₂); 77.9 (11-CH); 134.0 (3-C); 135.3 (2-CH); 171.8 (CO); 173.5 (CO); 194.1 (COS). ESMS *m/z* (%): 383 [M]Na⁺(8 %), 361 [M]H⁺(58 %), 343 [M - H₂O]H⁺(88 %), 231 [M - C₆H₁₁O₃]H⁺(100 %). HRMS *m/z* (%): calculated 383.1617, found 383.1616 for [M]Na⁺.



E-2,3-Dimethylbut-2-enoyl pantetheine 9P



The acyl pantetheine dimethyl acetal (**9Pa**, 0.55 g ,1.32 mmol) was dissolved in $CH_2Cl_2(2 \text{ ml})$. TFA (0.2 ml) was then added and the solution was stirred for 15 min at 22 °C. Thereafter, H_2O was added and the solution was stirred for an additional 10 min at 22 °C. The solvent was then removed by blowing of N_2 flow and the product was purified by HPLC. After purification, 202.0 mg (0.54 mmol) of the colorless oily product 3c were collected. ¹H NMR (400 MHz, CDCl₃): $\delta_H = 0.91$ (3 H, s, 10/11-CH₃), 1.01 (3 H, s, 10/11-CH₃), 1.79 (3 H, s, 19-CH₃), 1.92 (6 H, s, 18-CH₃ and 20-CH₃), 2.38- 2.45 (2 H, m, 6-CH₂), 2.93-3.14 (2 H, m, 9-CH₂), 3.29-3.61 (7 H, overlapping peaks, OH, 1-CH₂, 5-CH₂ and 8-CH₂), 3.98 (1 H, s, 3-CH), 6.32 (1 H, br. t, NH), 7.40 (1 H, br. t, NH) pm. ¹³C NMR (100 MHz, CDCl₃): $\delta_C = 16.2$ (18-CH₃ and 20-CH₃), 20.5 (10/11-CH₃), 21.8 (10/11-CH₃), 23.0 (19-CH₃), 28.6 (9-CH₂), 35.3 (5-CH₂), 35.7 (6-CH₂), 39.5 (2-C), 40.1 (8-CH₂), 71.1 (1- CH₂), 77.8 (3-CH), 129.3 (17-C), 141.0 (16-C), 171.8 (7-CO), 173.6 (4-CO), 196.7 (15-CO) ppm. IR: $v_{max} = 3291$ (w), 2933 (w), 2873 (w), 1642 (vs), 1526 (s), 1438 (w), 1374 (m), 1292 (w), 1262 (w), 1198 (w), 1167 (w), 1136 (w), 1077 (w), 1041 (m), 950 (m), 784 (w), 689 (w), 643 (w) cm⁻¹. ESMS: *m/z* (%) = 375 (82) [M]H+, 357 (76) [M + H - H₂O]+, 261 (43) [M + H - C₆H₉O - H₂O]+, 245 (100) [M + H - C₆H₁O₃]+.


E-2-Ethylbut-2-enoyl pantetheine 10P



The acyl pantetheine dimethyl acetal (**10Pa**, 0.99 g, 2.38 mmol) of was dissolved in CH_2Cl_2 (2 ml). TFA (0.2 ml) was then added and the solution was stirred for 15 min at 22 °C. Thereafter, H₂O was added and the solution was stirred for an additional 10 min at 22 °C. The solvent was then removed by blowing of N₂ flow and the product was purified by HPLC. After purification, 261.0 mg (0.70 mmol) of the colorless oily product were collected. ¹H NMR (400 MHz, CDCl₃): $\delta_H = 0.91$ (3 H, s, 10/11-CH₃), 0.99 (3 H, t, JHH = 7.4 Hz, 20-CH₃), 1.01 (3 H, s, 10/11-CH₃), 1.85 (3 H, d, JHH = 7.0 Hz, 18-CH₃), 2.35 (2H, q, JHH = 7.5 Hz, 19-CH₂), 2.40 (2 H, t, JHH = 5.8 Hz, 6-CH₂), 2.98-3.16 (2 H, m, 9-CH₂), 3.33-3.61 (7 H, overlapping peaks, OH, 1-CH₂, 5-CH₂ and 8-CH₂), 3.99 (1 H, s, 3-CH), 6.33 (1 H, br. t, NH), 6.82 (1 H, q, JHH = 7.1 Hz, 17-CH), 7.40 (1 H, br. t, NH) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta_C = 13.5$ (20-CH₃), 14.3 (18-CH₃), 20.2 (19-CH₃), 20.5 (10/11-CH₃), 21.8 (10/11-CH₃), 28.3 (9-CH₂), 35.3 (5-CH₂), 35.7 (6-CH₂), 39.5 (2-C), 40.0 (8-CH₂), 71.1 (1-CH₂), 77.8 (3-CH), 137.0 (17-CH), 143.2 (16-C), 171.8 (7-CO), 173.6 (4-CO), 193.9 (15-CO) ppm. IR: vM_{ax} = 3294 (w), 2965 (w), 2934 (w), 2874 (w), 1641 (vs), 1526 (s), 1438 (w), 1380 (m), 1292 (w), 1206 (w), 1073 (w), 1042 (m), 1011 (w), 958 (w), 844 (w), 735 (w), 690 (w), 652 (w) cm⁻¹. ESMS: *m/z* (%) = 397 (51) [M]Na + , 375 (67) [M]H + , 357 (88) [M + H - H₂O] + , 261 (73) [M + H - C₆H₄O - H₂O] + , 245 (100) [M + H - C₆H₁O₃] + .





E-Hex-2-enoyl pantetheine dimethyl ketal (**11Pa**, 0.02 g, 0.002 mmol) stirred in a mixture of acetonitrile and water (1:1) and 10 % TFA for 20 min. The reaction was followed by TLC and LCMS. After that the solvents were liphophilized. 0.003 g of the product were purified by HPLC. ¹H-NMR (CDCl₃, 400 MHz) $\delta_{\rm H}$ 0.93 (s, 3H, 21-CH₃); 0.95 (t, *J* = 7.8 Hz, 3H, 1-CH₃); 1.04 (s, 3H, 20-CH₃); 1.46-1.56 (m, 2H, 2-CH₂); 2.20 (dq, *J* = 1.3 Hz, *J* = 6.7 Hz, 2H, 5-CH₂); 2.41 (t, *J* = 6.1 Hz, 2H, 11-CH₂); 3.03-3.18 (m, 2H, 7-CH₂); 3.36-3.65 (m, 6H, 8-12-17-CH₂); 3.99 (s, ¹H, 15-CH); 6.13 (m, ¹H, 5-CH); 6.17 (dt, *J* = 1.8 Hz, *J* = 15.7 Hz, ¹H, 9-NH); 6.93 (dt, *J* = 6,7 Hz, *J* = 15.3 Hz, ¹H, 4-CH); 7.32 (bt, *J* = 6.¹Hz, ¹H, 13-NH). ¹³C-NMR (CDCl₃, 100 MHz) $\delta_{\rm C}$ 13.7 (1-CH₃), 20.4 (19-CH₃); 21.1 (18-CH₃); 21.7 (2-CH₂); 28.1 (7-CH₂); 34.3 (3-CH₂); 35.1 (12-CH₂); 35.5 (11-CH₂); 39.4 (16-C); 39.8 (8-CH₂); 70.9 (17-CH₂); 77.8 (15-CH); 128.3 (5-CH); 147.0 (4-CH); 171.6 (10-CO); 173.3 (14-CO); 190.6 (6-COS). ESMS *m/z* (%): 375 [M]H⁺(100 %), 357 [M - H₂O]H⁺ (32 %). HRMS *m/z* (%): calculated 397.1773, found 397.1774 for [M]Na+.



E-2-methylhex-2-enoyl-pantetheine 12P



E-Methylhex-2-enoyl pantetheine dimethyl ketal (**12Pa**, 0.06 g, 0.003 mmol) stirred in a mixture of CH₂Cl₂ and 10 % TFA for 20 min. The reaction was followed by TLC and LCMS. After that the solvents were evaporated. 0.02 g of the product were purified by HPLC. ¹H-NMR (CDCl₃, 400 MHz) $\delta_{\rm H}$ 0.93 (s, 3H, 18-CH₃); 0.95 (t, *J* = 7.4 Hz, 3H, 1-CH₃); 1.02 (s, 3H, 19-CH₃); 1.46-1.55 (m, 2H, 2-CH₂); 1.88 (d, *J* = 1.4 Hz, 5-CH₃) 2.20 (dq, *J* = 1.3 Hz, *J* = 7.0 Hz, 2H, 3-CH₂); 2.41 (t, *J* = 5.9 Hz, 2H, 11-CH₂); 3.00-3.13 (m, 2H, 8-CH₂); 3.35-3.59 (m, 6H, 9-12-17-CH₂); 3.99 (s, ¹H, 15-CH); 6.23 (bt, *J* = 5.7 Hz, ¹H, 10-NH); 6.77 (tq, *J* = 1,4 Hz, *J* = 7.4 Hz, ¹H, 4-CH); 7.38 (bt, *J* = 5.9 Hz, ¹H, 13-NH). ¹³C-NMR (CDCl₃, 100 MHz) $\delta_{\rm C}$ 12.5 (5-CH₃), 13.9 (1-CH₃), 20.4 (19-CH₃); 21.7 (18-CH₃); 21.8 (2-CH₂); 28.3 (8-CH₂); 30.8 (3-CH₂); 35.1 (12-CH₂); 35.5 (11-CH₂); 39.4 (16-C); 39.8 (9-CH₂); 70.9 (17-CH₂); 77.7 (15-CH); 135.7 (5-C); 142.5 (4-CH); 171.6 (11-CO); 173.4 (14-CO); 194.2 (7-COS). ESMS *m*/*z* (%): 411 [M]Na⁺(28 %), 389 [M]H⁺(31 %), 371 [M - H₂O]H⁺ (88 %), 259 [M - C₆H₁₁O₃]H⁺ (100 %). HRMS *m*/*z* (%): calculated 411.1930, found 411.1931 for [M]Na⁺.



E-4RS-4-methyl-hex-2-enoyl-pantetheine 13P



E-4*RS*-4-Methylhex-2-enoyl pantetheine dimethyl ketal (**13Pa**, 0.06 g, 0.003 mmol) stirred in a mixture of CH₂Cl₂ and 10 % TFA for 20 min. The reaction was followed by TLC and LCMS. After that the solvents were evaporated. 0.02 g of the product were purified by HPLC. ¹H-NMR (CDCl₃, 400 MHz) $\delta_{\rm H}$ 0.88 (t, *J* = 7.5 Hz, 3H, 1-CH₃); 0.98 (s, 3H, 18-CH₃); 1.01 (s, 3H, 19-CH₃); 1.05 (d, *J* = 6.9 Hz, 4-CH₃); 1.38-1.47 (m, 2H, 2-CH₂); 2.19-2.27 (m, ¹H, 3-CH); 2.41 (t, *J* = 5.8 Hz, 2H, 11-CH₂); 2.94-3.15 (m, 2H, 8-CH₂); 3.33-3.58 (m, 6H, 9-12-17-CH₂); 4.00 (s, ¹H, 15-CH); 6.08 (dd, *J* = 1.4 Hz, *J* = 15.4 Hz, ¹H, 6-CH); 6.40 (bt, *J* = 5.4 Hz, ¹H, 10-NH); 6.83 (dd, *J* = 7.4 Hz, *J* = 15.4 Hz, ¹H, 5-CH); 7.40 (bt, *J* = 5.4 Hz, ¹H, 13-NH). ¹³C-NMR (CDCl₃, 100 MHz) $\delta_{\rm C}$ 11.6 (1-CH₃), 18.7 (4-CH₃), 20.4 (19-CH₃); 21.7 (18-CH₃); 28.2 (2-CH₂); 28.8 (8-CH₂); 30.7 (3-CH₂); 35.1 (12-CH₂); 35.2 (11-CH₂); 39.4 (16-C); 39.7 (9-CH₂); 70.8 (17-CH₂); 77.6 (15-CH); 126.7 (6-CH); 152.1 (5-CH); 171.7 (10-CO); 173.6 (14-CO); 190.8 (7-COS). ESMS *m/z* (%): 411 [M]Na⁺ (8 %), 389 [M]H ⁺ (65 %), 371 [M - H₂O]H ⁺ (88 %), 259 [M - C₆H₁₁O₃]H ⁺ (100 %). HRMS *m/z* (%): calculated 411.1930, found 411.1930 for [M]Na⁺.



E-4RS-2,4-dimethylhex-2-enoyl-pantetheine 14P



E-4*RS*-2,4-Dimethylhex-2-enoyl pantetheine dimethyl ketal (**14Pa**, 0.06 g, 0.003 mmol) was stirred in a mixture of CH₂Cl₂ and 10 % TFA for 20 min. The reaction was followed by TLC and LCMS. After that the solvents were evaporated. 0.01 g of the product were purified by HPLC (acetonitrile). ¹H-NMR (CDCl₃, 400 MHz) $\delta_{\rm H}$ 0.86 (t, *J* = 7.4 Hz, 3H, 1-CH₃); 0.92 (s, 3H, 18-CH₃); 1.03 (s, 3H, 19-CH₃); 1.03 (d, *J* = 2.3 Hz, 3H, 4-CH₃); 1.30-1.51 (m, 4H, 2-CH₂); 1.88 (d, *J* = 1.1 Hz, 7-CH₃), 2.41 (t, *J* = 5.6 Hz, 2H, 12-CH₂); 2.40-2.49 (m, ¹H, 3-CH); 2.99-3.14 (m, 2H, 9-CH₂); 3.35-3.59 (m, 6H, 10-13-17-CH₂); 3.99 (s, ¹H, 15-CH); 6.19 (bt, *J* = 5.7 Hz, ¹H, 11-NH); 6.54 (dq, *J* = 1.3 Hz, *J* = 9.8 Hz, ¹H, 5-CH); 7.36 (bt, *J* = 5.9 Hz, ¹H, 14-NH). ¹³C-NMR (CDCl₃, 100 MHz) $\delta_{\rm C}$ 11.9 (1-CH₃), 12.7 (4-CH₃), 19.5 (7-CH₃), 20.4 (19-CH₃); 21.7 (18-CH₃); 28.3 (9-CH₂); 29.6 (3-CH); 35.1 (13-CH₂); 35.6 (12-CH₂); 39.4 (16-C); 39.8 (10-CH₂); 71.0 (17-CH₂); 77.7 (15-CH); 134.4 (6-C); 147.8 (5-CH); 171.4 (11-CO); 173.3 (14-CO); 194.5 (8-COS). ESMS *m/z* (%): 425 [M]Na⁺(12%), 403 [M]H⁺(28%), 385 [M - H₂O]H⁺(88%), 273 [M - C₆H₁₁O₃]H⁺(100%). HRMS *m/z* (%): calculated 425.2086, found 425.2085 for [M]H+.



E-4S-2,4-dimethylhex-2-enoyl-pantetheine 15P



E-4S-2,4-Dimethylhex-2-enoyl pantetheine dimethyl ketal (**15Pa**, 0.06 g, 0.003 mmol) was stirred in a mixture of CH₂Cl₂ and 10 % TFA for 20 min. The reaction was followed by TLC and LCMS. After that the solvents were evaporated. 0.01 g of the product were purified by HPLC. ¹H-NMR (CDCl₃, 400 MHz) $\delta_{\rm H}$ 0.86 (t, *J* = 7.4 Hz, 3H, 1-CH₃); 0.92 (s, 3H, 18-CH₃); 1.03 (s, 3H, 19-CH₃); 1.03 (d, *J* = 2.3 Hz, 3H, 4-CH₃); 1.30-1.51 (m, 4H, 2-CH₂); 1.88 (d, *J* = 1.1 Hz, 7-CH₃), 2.41 (t, *J* = 5.6 Hz, 2H, 12-CH₂); 2.40-2.49 (m, ¹H, 3-CH); 2.99-3.14 (m, 2H, 9-CH₂); 3.35-3.59 (m, 6H, 10-13-17-CH₂); 3.99 (s, ¹H, 15-CH); 6.19 (bt, *J* = 5.7 Hz, ¹H, 11-NH); 6.54 (dq, *J* = 1.3 Hz, *J* = 9.8 Hz, ¹H, 5-CH); 7.36 (bt, *J* = 5.9 Hz, ¹H, 14-NH). ¹³C-NMR (CDCl₃, 100 MHz) $\delta_{\rm C}$ 11.9 (1-CH₃), 12.7 (4-CH₃), 19.5 (7-CH₃), 20.4 (19-CH₃); 21.7 (18-CH₃); 28.3 (9-CH₂); 29.6 (3-CH); 35.1 (13-CH₂); 35.6 (12-CH₂); 39.4 (16-C); 39.8 (10-CH₂); 71.0 (17-CH₂); 77.7 (15-CH); 134.4 (6-C); 147.8 (5-CH); 171.4 (11-CO); 173.3 (14-CO); 194.5 (8-COS). ESMS *m/z* (%): 425 [M]Na⁺ (11%), 403 [M]H⁺ (26%), 385 [M - H₂O]H⁺ (88%), 273 [M - C₆H₁₁O₃]H⁺ (100%). HRMS *m/z* (%): calculated 425.2086, found 425.2085 for [M]Na⁺.



E-2-ethylhex-2-enoyl-pantetheine 16P



E-2-Ethylhex-2-enoyl pantetheine dimethyl ketal (**16Pa**, 300 mg) was stirred in a mixture of CH₂Cl₂ and 10 % TFA for 20 min. The reaction was followed by TLC and LCMS. After that the solvents were evaporated. 0.027 g of the product were purified by HPLC. ¹H-NMR (CDCl₃, 400 MHz) $\delta_{\rm H}$ 0.85-0.99 (m, 12H, 1-7-18-19-CH₃); 1.38-1.53 (m, 2H, 2-CH₂); 2.16-2.35 (m, 4H, 3-6-CH₂); 2.41 (t, *J* = 6.4 Hz, 2H, 12-CH₂); 2.97-3.06 (m, 2H, 9-CH₂); 3.33-3.56 (m, 6H, 10-13-17-CH₂); 3.98 (s, ¹H, 15-CH); 5.63 (t, *J* = 7.6 Hz, ¹H, 4-CH_z); 6.56 (bt, *J* = 5.7 Hz, ¹H, 11-NH); 6.70 (t, *J* = 7.6 Hz, ¹H, 4-CH_E); 7.46 (bt, *J* = 5.9 Hz, ¹H, 14-NH). ¹³C-NMR (CDCl₃, 100 MHz) $\delta_{\rm C}$ 13.4 (1-CH₃), 13.9 (7-CH₃), 20.4 (19-CH₃); 21.5 (18-CH₃); 22.0 (6-CH₂); 22.8 (2-CH₂); 28.2 (9-CH₂); 30.4 (3-CH₂); 35.1 (13-CH₂); 35.5 (12-CH₂); 39.4 (16-C); 39.8 (10-CH₂); 70.9 (17-CH₂); 77.7 (15-CH); 136.1 (4-CH_z); 140.5 (5-C); 142.2 (4-CH_E); 171.6 (11-CO); 173.4 (14-CO); 193.9 (8-COS). ESMS *m/z* (%): 425 [M]Na⁺ (78 %), 403 [M]H⁺ (13 %), 385 [M - H₂O]H⁺ (56 %), 273 [M - C₆H₁₁O₃]H⁺ (100 %). HRMS *m/z* (%): calculated 425.2086, found 425.2086 for [M]Na⁺.







E-Hept-2-enoyl chloride was prepared from *E*-hept-2-enoic acid (**17**, 0.11 g, 0.90 mmol) with thionyl chloride (0.11 g, 1.00 mmol) in THF (0.5 ml). This solution was stirred for 24 h under nitrogen, followed by evaporation of solvent.

A solution of pantetheine (0.30 g, 0.54 mmol) and DTT (88 mg, 0.54 mmol) in THF (3 ml) was stirred in a nitrogen atmosphäre at 30 °C for 3 h. Pyridine (0.3 ml) and DMAP (12 mg, 0.09 mmol) were added to the reaction and stirred for 15 min. The reaction was cooled to 0 °C and hept-2-enoyl chloride was added. The mixture was stirred for one hour at this temperature. The solvents were removed under a nitrogen flow. 6 mg of the product were purified by HPLC. ¹H-NMR (CDCl₃, 400 MHz) $\delta_{\rm H}$ 0.94 (t, *J* = 7.1 Hz, 3H, 1-CH₃); 0.95 (s, 3H, 18-CH₃); 1.06 (s, 3H, 17-CH₃); 1.28-1.42 (m, 4-H, 2-3-CH₂); 2.24 (dq, *J* = 1.6 Hz, *J* = 7.2 Hz, 2H, 4-CH₂); 2.43 (t, *J* = 7.2 Hz, 2H, 11-CH₂); 3.05-3.25 (m, 2H, 8-CH₂); 3.38-3.62 (m, 6H, 16-12-9-CH₂); 4.01 (s, ¹H, 14-CH); 6.13 (m, ¹H, 6-CH); 6.15 (dt, *J* = 1.5 Hz, *J* = 15.4 Hz, ¹H, 10-NH); 6.96 (dt, *J* = 7,0 Hz, *J* = 15.5 Hz, ¹H, 5-CH); 7.34 (bt, *J* = 6.0 Hz, ¹H, 13-NH). ¹³C-NMR (CDCl₃, 100 MHz) $\delta_{\rm C}$ 13.8 (1-CH₃), 20.4 (17-CH₃); 21.7 (18-CH₃); 22.3 (2-CH₂); 28.1 (8-CH₂); 29.9 (3-CH₂); 31.9 (4-CH₂); 35.1 (12-CH₂); 35.6 (11-CH₂); 39.3 (15-C); 39.9 (9-CH₂); 70.9 (16-CH₂); 77.8 (14-CH); 128.1 (6-CH); 147.3 (5-CH); 171.6 (10-CO); 173.2 (13-CO); 190.6 (7-COS). ESMS *m/z* (%):777.9 [M₂]H⁺ (28 %), 389 [M]H⁺ (100 %). HRMS *m/z* (%): calculated 411.1930, found 411.1929 for [M]Na+.



E-6-methylhept-2-enoyl pantetheine 18P



E-6-Methylhept-2-enoyl pantetheine dimethyl ketal (**18Pa**, 0.06 g, 0.003 mmol) was stirred in a mixture of CH₂Cl₂ and 10 % TFA for 20 min. The reaction was followed by TLC and LCMS. After that the solvents were evaporated. 0.01 g of the product were purified by HPLC. ¹H-NMR (CDCl₃, 400 MHz) $\delta_{\rm H}$ 0.86-0.92 (m, 9H, 1-3-18-CH₃); 1.01 (s, 3H, 19-CH₃); 1.14-1.38 (m, 2H, 4-CH₂); 1.47-1.62 (m, ¹H, 2-CH); 2.22 (q, *J* = 7.0 Hz, 2H, 5-CH₂); 2.41 (t, *J* = 6.2 Hz, 2H, 12-CH₂); 3.01-3.15 (m, 2H, 9-CH₂); 3.35-3.58 (m, 6H, 10-13-17-CH₂); 3.99 (s, ¹H, 15-CH); 6.13 (dt, *J* = 1.5 Hz, *J* = 15.2 Hz, ¹H, 7-CH); 6.39 (bt, *J* = 5.6 Hz, ¹H, 11-NH); 6.93 (dt, *J* = 6.5 Hz, *J* = 15.5 Hz, ¹H, 6-CH); 7.39 (bt, *J* = 5.9 Hz, ¹H, 14-NH). ¹³C-NMR (CDCl₃, 100 MHz) $\delta_{\rm C}$ 20.4 (19-CH₃); 21.7 (18-CH₃); 22.4 (1-3-CH₃); 27.6 (2-CH); 28.1 (9-CH₂); 30.2 (4-CH₂); 35.1 (13-CH₂); 35.6 (12-CH₂); 36.9 (5-CH₂); 39.4 (16-C); 39.8 (10-CH₂); 70.9 (17-CH₂); 77.7 (15-CH); 128.0 (7-CH); 147.4 (6-CH); 171.7 (11-CO); 173.5 (14-CO); 190.6 (8-COS). ESMS *m/z* (%): 425 [M]Na⁺ (12%), 403 [M]H⁺ (30%), 385 [M - H₂O]H⁺ (88%), 273 [M - C₆H₁₁O₃]H⁺ (100%). HRMS *m/z* (%): calculated 425.2086, found 425.2089 for [M]Na⁺.





E-Oct-2-enoyl chloride was prepared from *E*-oct-2-enoic acid (**19**, 0.27g, 2.00 mmol) with thionyl chloride (0.22 g, 3.20 mmol) in THF (0.5 ml). This solution stirred for 24 h under nitrogen and solvent removed *in vacuo*.

A solution of pantetheine (0.45 g, 0.81 mmol) and DTT (0.12 g, 0.81 mmol) in THF (3 ml) was stirred in a nitrogen atmosphere at 30 °C for 3 h. After that pyridine (0.3 ml) and DMAP (12 mg, 0.09 mmol) were added to the reaction and stirred for 15 min. Then the reaction was cooled to 0 °C and oct-2-enoyl chloride was added. The mixture was stirred for one hour at this temperature. The solvents were removed under a nitrogen flow. 6 mg of the product were purified by HPLC. ¹H-NMR (CDCl₃, 400 MHz) $\delta_{\rm H}$ 0.91 (t, *J* = 6.7 Hz, 3H, 1-CH₃); 0.95 (s, 3H, 21-CH₃); 1.06 (s, 3H, 20-CH₃); 1.28-1.61 (m, 6H, 2-3-4-CH₂); 2.21 (dq, *J* = 1.4 Hz, *J* = 7.3 Hz, 2H, 5-CH₂); 2.43 (t, *J* = 6.0 Hz, 2H, 13-CH₂); 3.05-3.20 (m, 2H, 9-CH₂); 3.38-3.62 (m, 6H, 10-14-19-CH₂); 4.01 (s, ¹H, 17-CH); 6.13 (t, *J* = 4.07, ¹H, 7-CH); 6.17 (dt, *J* = 1.4 Hz, *J* = 15.4 Hz, ¹H, 11-NH); 6.96 (dt, *J* = 6.9 Hz, *J* = 15.4 Hz, ¹H, 6-CH); 7.34 (bt, *J* = 5.82 Hz, ¹H, 15-NH). ¹³C-NMR (CDCl₃, 100 MHz) $\delta_{\rm C}$ 13.9 (1-CH₃), 20.4 (21-CH₃); 39.3 (18-C); 39.9 (10-CH₂); 70.9 (19-CH₂); 77.8 (17-CH); 128.1 (7-CH); 147.3 (6-CH); 171.6 (12-CO); 173.2 (16-CO); 190.6 (8-COS). ESMS *m/z* (%): 805.9 [M₂]H⁺ (14 %), 403 [M]H⁺ (100 %). HRMS *m/z* (%): calculated 425.2086, found 425.2089 for [M]Na+.



E-2-methyloct-2-enoyl-pantetheine 20P



E-2-Methyloct-2-enoyl pantetheine dimethyl ketal (**20Pa**, 0.06 g, 0.003 mmol) was stirred in a mixture of CH₂Cl₂ and 10 % TFA for 20 min. The reaction was followed by TLC and LCMS. After that the solvents were evaporated. 0.01 g of the product were purified by HPLC. ¹H-NMR (CDCl₃, 400 MHz) $\delta_{\rm H}$ 0.90 (tr, *J* = 6.9 Hz, 3H, 1-CH₃); 0.91 (s, 3H, 19-CH₃); 1.01 (s, 3H, 20-CH₃); 1.25-1.36 (m, 4H, 2-4-CH₂); 1.41-1.50 (m, 2H, 3-CH₂); 1.85 (s, 3H, 8-CH₃); 2.20 (q, *J* = 7.5 Hz, 2H, 5-CH₂); 2.41 (t, *J* = 6.6 Hz, 2H, 13-CH₂); 2.99-3.12 (m, 2H, 10-CH₂); 3.35-3.58 (m, 6H, 11-14-18-CH₂); 3.99 (s, ¹H, 16-CH); 6.37 (bt, *J* = 5.7 Hz, ¹H, 12-NH); 6.77 (tq, *J* = 1.3 Hz, *J* = 7.6 Hz, ¹H, 6-CH); 7.41 (bt, *J* = 6.2 Hz, ¹H, 15-NH). ¹³C-NMR (CDCl₃, 100 MHz) $\delta_{\rm C}$ 12.5 (8-CH₃); 14.1 (1-CH₃); 20.4 (19-CH₃); 21.6 (20-CH₃); 22.5 (2-CH₂); 28.1 (5-CH₂); 28.2 (4-CH₂); 28.8 (10-CH₂); 31.6 (3-CH₂); 35.1 (14-CH₂); 35.6 (13-CH₂); 39.4 (17-C); 39.8 (11-CH₂); 70.9 (18-CH₂); 77.8 (16-CH); 135.6 (7-C); 142.7 (6-CH); 171.7 (12-CO); 173.2 (15-CO); 194.2 (9-COS). ESMS *m/z* (%): 439 [M]Na⁺ (2%), 417 [M]H⁺ (41%), 399 [M - H₂O]H⁺ (85%), 287 [M - C₆H₁₁O₃]H⁺ (100%). HRMS *m/z* (%): calculated 439.2240 for [M]Na⁺.



E-7-methyloct-2-enoyl-pantetheine 21P



E-7-Methyloct-2-enoyl pantetheine dimethyl ketal (**21Pa**, 0.06 g, 0.003 mmol) was stirred in a mixture of CH₂Cl₂ and 10 % TFA for 20 min. The reaction was followed by TLC and LCMS. After that the solvents were evaporated. 0.01 g of the product were purified by HPLC. ¹H-NMR (CDCl₃, 400 MHz) $\delta_{\rm H}$ 0.86 (d, *J* = 6.6 Hz, 6H, 1-3-CH₃); 0.93 (s, 3H, 18-CH₃); 1.03 (s, 3H, 19-CH₃); 1.14-1.30 (m, 2H, 3-4-CH₂); 1.42-1.60 (m, ¹H, 2-CH); 2.22 (q, *J* = 7.4 Hz, 2H, 5-CH₂); 2.41 (t, *J* = 5.8 Hz, 2H, 12-CH₂); 3.01-3.18 (m, 2H, 9-CH₂); 3.35-3.59 (m, 6H, 10-13-17-CH₂); 3.99 (s, ¹H, 15-CH); 6.13 (dt, *J* = 1.5 Hz, *J* = 15.5 Hz, ¹H, 7-CH); 6.18 (bt, *J* = 5.8 Hz, ¹H, 11-NH); 6.93 (dt, *J* = 6.9 Hz, *J* = 15.5 Hz, ¹H, 6-CH); 7.34 (bt, *J* = 5.8 Hz, ¹H, 14-NH). ¹³C-NMR (CDCl₃, 100 MHz) $\delta_{\rm C}$: 20.4 (19-CH₃); 21.7 (18-CH₃); 22.5 (1-1-CH₃); 25.8 (4-CH); 27.8 (2-CH); 28.1 (9-CH₂); 32.6 (3-CH₂); 35.1 (13-CH₂); 35.6 (12-CH₂); 38.5 (5-CH₂); 39.4 (16-C); 39.8 (10-CH₂); 70.9 (17-CH₂); 77.8 (15-CH); 128.1 (7-CH); 147.3 (6-CH); 171.6 (11-CO); 173.3 (14-CO); 190.6 (8-COS). ESMS *m/z* (%): 417 [M]H⁺(15%), 399 [M - H₂O]H⁺(60%), 287 [M - C₆H₁₁O₃]H⁺(100%). HRMS *m/z* (%): calculated 439.2243, found 439.2242 for [M]Na⁺.



E-4RS,6RS-4,6-dimethyl-oct-2-enoyl-pantetheine 22P



E-4*RS*,6*RS*-4,6-dimethyl-oct-2-enoyl pantetheine dimethyl ketal (**22Pa**, 300 mg) stirred in a mixture of CH₂Cl₂ and 10 % TFA for 20 min. The reaction was followed by TLC and LCMS. After that the solvents were evaporated. 0.027 g of the product were purified by HPLC. ¹H-NMR (CDCl₃, 400 MHz) $\delta_{\rm H}$ 0.81-0.90 (m, 6H, 1-3-CH₃), 0.92 (s, 3H, 20-21-CH₃), 1.03 (s, 3H, 20-CH₃), 1.00-1.07 (m, 3H, 7-CH₃), 1.07-1.44 (m, 6H, 2-5-CH₂, 4-CH), 2.41 (t, 2H, *J* = 6.0 Hz, 14-CH₂), 2.34-2.46 (m, ¹H, 6-CH), 2.95-3.17 (m, 2H, 11-CH₂), 3.33-3.66 (m, 6H, 12-15-19-CH₂), 3.99 (s, ¹H, 17-CH), 6.08 (dt, ¹H, *J* = 1.4 Hz, *J* = 15.6 Hz, 8-CH), $\delta_{\rm C}$ 6.23 (bt, *J* = 6.0 Hz, ¹H, NH), 6.75-6.88 (m, ¹H, 9-CH); 7.36 (bt, *J* = 6.0 Hz, ¹H, NH). ¹³C-NMR (CDCl₃, 100 MHz) 11.3 (22-CH₃ and 24-CH₃), 19.2 (23-CH₃), 20.5 (20-CH₃), 21.9 (21-CH₃), 28.3 (11-CH₂), 29.5 (5-CH₂), 29.8 (19-CH₂), 31.9 (4-CH), 34.3 (6-CH), 35.3 (15-CH₂), 35.7 (14-CH₂), 39.5 (18-C), 40.0 (12-CH₂), 71.1 (19-CH₂), 77.9 (17-CH), 126.2 (9-CH), 152.5 (8-CH), 171.8 (13-CO), 173.5 (16-CO), 190.9 (10-CO). ESMS *m/z* (%): 453 [M]Na⁺ (11 %), 431 [M]H⁺ (62 %), 413 [M - H₂O]H⁺ (85 %), 301 [M - C₆H₁₁O₃]H⁺ (100 %). HRMS *m/z* (%): calculated 453.2396, found 453.2396 for [M]Na⁺.



E-4S,6S-4,6-dimethyl-oct-2-enoyl pantetheine 1P



E-4*S*,6*S*-4,6-dimethyloct-2-enoyl pantetheine dimethyl ketal (**1Pa**, 300 mg) was stirred in a mixture of CH₂Cl₂ and 10 % TFA for 20 min. The reaction was followed by TLC and LCMS. After that the solvents were evaporated. 0.027 g of the product were purified by HPLC (acetonitrile). ¹H-NMR (CDCl₃, 400 MHz) $\delta_{\rm H}$ 0.81-0.90 (m, 6H, 1-3-CH₃), 0.92 (s, 3H, 21-CH₃), 1.03 (s, 3H, 20-CH₃), 1.05 (m, 3H, 7-CH₃), 1.10-1.44 (m, 6H, 2-5-CH₂, 4-CH), 2.42 (t, 2H, *J* = 6.0 Hz, 14-CH₂), 2.37-2.41 (m, ¹H, 6-CH), 3.03-3.17 (m, 2H, 11-CH₂), 3.36-3.58 (m, 6H, 12-15-19-CH₂), 3.99 (s, ¹H, 17-CH), 6.08 (dd, ¹H, *J* = 1.4 Hz, *J* = 15.6 Hz, 8-CH), 6.23 (bt, *J* = 6.0 Hz, ¹H, NH), 6.79 (dd, *J* = 7.6 Hz, ¹H, 9-CH); 7.37 (bt, *J* = 6.0 Hz, ¹H, NH). ¹³C-NMR (CDCl₃, 100 MHz) $\delta_{\rm C}$ 11.3 (1-CH₃), 18.8 (3-CH₃), 20.2 (20-CH₃), 20.5 (21-CH₃), 21.9 (7-CH₃), 28.3 (2-CH₂), 28.5 (11-CH₂), 29.5 (4-CH), 34.3 (6-CH), 35.1 (15-CH₂), 35.9 (14-CH₂), 39.5 (18-C), 39.8 (12-CH₂), 43.3 (5-CH₂), 71.1 (19-CH₂), 77.9 (17-CH), 126.2 (9-CH), 152.5 (8-CH), 171.8 (13-CO), 173.5 (16-CO), 190.9 (10-CO). ESMS *m/z* (%): 453 [M]Na⁺ (8 %), 431 [M]H⁺ (65 %), 413 [M - H₂O]H⁺ (88 %), 301 [M - C₆H₁₁O₃]H ⁺ (100 %). HRMS *m/z* (%): calculated 453.2399, found 453.2392 for [M]Na⁺.





E-Dec-2-enoyl chloride was prepared from *E*-dec-2-enoic acid (**24**, 0.138 g, 1.00 mmol,) with thionyl chloride (0.110 g, 1.00 mmol) in THF (0.5 ml). This solution stirred for 24 h under nitrogen, and solvent removed *in vacuo*. A solution of pantetheine (0.30 g, 0.54 mmol,) and DTT (0.08 mg, 0.54 mmol) in THF (3 ml) was stirred in a nitrogenatmosphäre at 30 °C for 3 h. After that pyridine (0.3 ml) and DMAP (12 mg, 0.09 mmol,) were added to the reaction and stirred for 15 min. Then the reaction was cooled to 0 °C and *E*-dec-2-enoyl chloride was added to the reaction. The mixture was stirred for one hour at this temperature. The solvents were removed under a nitrogen flow. 6 mg of the product were purified by HPLC. ¹H-NMR (CDCl₃, 400 MHz) $\delta_{\rm H}$ 0.91 (t, *J* = 7.1 Hz, 3H, 1-CH₃); 0.95 (s, 3H, 21-CH₃); 1.06 (s, 3H, 20-CH₃); 1.28-1.33 (m, 8H, 3,4,5,6-CH₂); 1.45-1.53 (m, 2H, 2-CH₂); 2.23 (dq, *J* = 1.6 Hz, *J* = 7.2 Hz, 2H, 7-CH₂); 2.43 (t, *J* = 7.2 Hz, 2H, 15-CH₂); 3,05-3.20 (m, 2H, 11-CH₂); 3.7-3.62 (m, 6H, 12-16-23-CH₂); 4.01 (s, ¹H, 19-CH); 6.13 (m, ¹H, 9-CH); 6.17 (dt, *J* = 1.5 Hz, *J* = 15.4 Hz, ¹H, 13-NH); 6.96 (dt, *J* = 7,0 Hz, *J* = 15.5 Hz, ¹H, 8-CH); 7.34 (bt, *J* = 6.0 Hz, ¹H, 17-NH). ¹³C-NMR (CDCl₃, 100 MHz) $\delta_{\rm C}$ 14.1 (1-CH₃), 20.4 (21-CH₃); 21.7 (20-CH₃); 22.6 (2-CH₂); 28.3 (11-CH₂); 29.1 (4-CH₂); 31.7 (6-CH₂); 32.3 (7-CH₂); 35.1 (16-CH₂); 35.6 (15-CH₂); 39.9 (12-CH₂); 70.9 (23-CH₂); 77.8 (19-CH); 128.1 (9-CH); 147.7 (8-CH); 171.6 (14-CO); 173.3 (18-CO); 190.9 (10-COS). ESMS *m*/*z* (%):862.9 [M₂]H⁺(28 %), 431 [M]H⁺(100 %), 413 [M - H₂O]H⁺(1 %). HRMS *m*/*z* (%): calculated 453.2399, found 453.2399 for [M]Na+.





E-2-methyldec-2-enoyl pantetheine 25P



E-2-Methyldec-2-enoyl pantetheine dimethyl ketal (**25Pa**, 0.06 g, 0.003 mmol) was stirred in a mixture of CH₂Cl₂ and 10 % TFA for 20 min. The reaction was followed by TLC and LCMS. After that the solvents were evaporated. 0.01 g of the product were purified by HPLC. ¹H-NMR (CDCl₃, 400 MHz) $\delta_{\rm H}$ 0.88 (tr, *J* = 7.2 Hz, 3H, 1-CH₃); 0.93 (s, 3H, 22-CH₃); 1.04 (s, 3H, 21-CH₃); 1.28-1.31 (m, 8H, 2-3-4-6-CH₂); 1.43-1.48 (m, 2H, 5-CH₂); 1.87 (s, 3H, 10-CH₃); 2.22 (q, *J* = 7.2 Hz, 2H, 7-CH₂); 2.41 (t, *J* = 6.2 Hz, 2H, 12-CH₂); 3.00-3.14 (m, 2H, 12-CH₂); 3.35-3.63 (m, 6H, 13-16-20-CH₂); 3.99 (s, ¹H, 18-CH); 6.09 (bt, *J* = 5.6 Hz, ¹H, 14-NH); 6.78 (tq, *J* = 1.3 Hz, *J* = 7.4 Hz, ¹H, 8-CH); 7.38 (bt, *J* = 6.6 Hz, ¹H, 17-NH). ¹³C-NMR (CDCl₃, 100 MHz) $\delta_{\rm C}$ 12.5 (10-CH₃); 14.1 (1-CH₃); 20.4 (21-CH₃); 21.7 (22-CH₃); 22.6 (2-CH₂); 28.3 (7-CH₂); 28.5 (12-CH₂); 28.8 (4-CH₂); 29.1 (5-CH₂); 29.4 (6-CH₂); 31.8 (3-CH₂); 35.1 (16-CH₂); 35.6 (15-CH₂); 39.4 (19-C); 39.8 (13-CH₂); 70.9 (20-CH₂); 77.8 (18-CH); 135.6 (9-C); 142.8 (8-CH); 171.7 (14-CO); 173.2 (17-CO); 194.2 (11-COS). ESMS *m/z* (%): 467 [M]Na⁺ (12%), 445 [M]H⁺ (27%), 427 [M - H₂O]H⁺ (67%), 315 [M - C₆H₁₁O₃]H⁺ (100%). HRMS *m/z* (%): calculated 467.2556, found 467.2556 for [M]Na⁺.



1.1 Synthesis of Compounds for Stereochemical Investigations

1.1.1 Testing the stereochemistry of the α -methyl of the reduction of the diketide substrate

NMR assays were performed with tiglyl-SNAC **5S** (2.0 mg, 10 mmol), NADPH (8 mg, 10 mmol), tris buffer pH 8.5 (50 μ l, 1M stock), glycerol (20% v/v) and the isolated ER domain (380 mU, 100 μ l of stock) in water (total volume 1 ml). The progress of the reaction was monitored by LCMS. After 24 h further NADPH (8 mg, 10 mmol) was added to the reaction. After a further 17 h the protein was precipitated with CH₂Cl₂ (1 ml) and centrifuged (3000 rpm, 20 min). The organic layer was removed and the aqueous layer was further extracted with CH₂Cl₂ (2 × 1 ml). The organic fractions were combined and the solvent was removed under a flow of dry N₂ and then dried in *vacuo* for a further 2 h. To the dried sample was added water (450 μ l) and aqueous NaOH (2M, 50 μ l). The sample was left for 90 min at RT and then acidified with aqueous HCl (2M, 100 μ l) to pH 3 and extracted with CDCl₃ (4 × 200 μ l). The organics were combined, dried with MgSO₄ and then filtered through MgSO₄ directly into an NMR tube. (1*R*,2*R*)-(+)-1,2 diphenylethylenediamine was titrated (100 mM stock) to optimize resolution of the obtained spectra.

1.1.2 Production of NADPD stereoisomers

4'S-NADPD 27²⁹

A solution of sodium bicarbonate (6.3 mg, 75 μ mol), D₇-glucose (8.0 mg, 45 μ mol) and NADP + (16.7 mg, 23 μ mol) were titrated to a pH 7.5 in water (1.5 ml). Glucose dehydrogenase from *Pseudomonas sp* (35 U) was added and the reaction was incubated at RT and the progress of the reaction was monitored by UV (340 nm). On completion the reaction was confirmed by MS (747 (100, [M]H+) and was used without further purification (final concentration of NADPD 15 mM).

4'*R***-NADPD 28**³⁰

A stock solution of alcohol dehydrogenase NADP + dependent, from *thermoanerobium brockii* (TbADH) was prepared in tris pH 7.2, 25 mM (0.085 U/µl) and was added (25 µl) to a solution of NADP + disodium trihydrate (17. mg, 20.8 µmol), *iso*propanol (300 µl) and tris buffer pH 9 (25 mM, 7.5 ml), this solution was incubated at 43 °C and the progress of the reaction was monitored by UV (340 nm). When no further change in absorption was observed the acetone and remaining *iso*propanol was removed *in vacuo* and the product was confirmed by MS (747 (100, [M]H+). This was used without and further purification (final concentration of NADPD 3.1 mM).

1.1.3 Synthesis of mandelate ester standards

RS-2-methyl butyryl methyl S-mandelate 30³¹

2-Methylbutyric acid (102 mg, 1.0 mmol) and S-methyl mandelate (208 mg, 1.3 mmol) were stirred at 0 °C in CH₂Cl₂ (3 ml). To this was added DMAP (4 mg) and EDCI (240 mg, 1.25 mmol) and this was then stirred at RT and followed by TLC. On completion, aqueous HCl (0.5 M, 5 ml) was added and this was extracted with CH₂Cl₂ (3 × 5 ml). The organic fractions were combined and concentrated in *vacuo*. The resulting oil was purified by column chromatography (hexane:ethyl acetate 3:1) to yield the desired product as a inseparable mixture of diastereomer (A:B 1:1, colourless oil, 206 mg, 832 µmol, 83%, Rf 0.46, hexane:ethyl acetate, 3:1): $\delta_{\rm H}$ (400 MHz, CDCl₃); 7.50-7.45 (2H, m, H₂-10 A and B), 7.41-7.36 (3H, m, H₂-11 H-12 A and B), 5.93 (0.5H, s, H-6 A), 5.92 (0.5H, s, H-6 B), 3.72 (1.5H, s, H₃-8 A), 3.72 (1.5H, s, H₃-8 B), 2.54 (1H, qt *J* 7.0, H-2), 1.85-1.69 (1H, m, H-3), 1.63-1.47 (1H, m, H-3), 1.24 (1.5H, d *J* 7.0, H₃-5 A), 1.19 (1.5H, d *J* 7.0, H₃-5 B), 0.99 (1.5H, t *J* 7.4, H₃-4 B), 0.92 (1.5H, t *J* 7.4, H₃-4 A); $\delta_{\rm C}$ (100 MHz, CDCl₃); 176.1 (C-1B), 176.0 (C-1A), 169.5 (C-7 A and B), 134.0 (C-9 129.1 (C-12), 128.8 (C-11), 127.7 (C-10), 74.2 (C-6B), 74.1 (C-6A), 52.6 (C-8A), 52.5 (C-8B), 40.9 (C-2A), 40.3 (C-2B), 26.8 (C-3B), 26.6 (C-3A), 16.6 (C-5A), 16.3 (C-5B), 11.5 (C-4A), 11.4 (C-4B); MS (ES +): *m*/z (%) 523 (13, [M₂ + Na]+), 501 (10, [M₂ + H]+), 251 (63, [M]H+), 191 (81, [M + H - COOHCH₃]+), 149 (100, CH₃CHCH₃COOH]+), 212 (23, [M + H₂ - CH₃CH₂CHCH₃COOHOCH₃]+).

Ethyl 3*R*-hydroxyl 2*R*-methylbutyrate 32³²

BuLi (1.6 M in hexanes, 4.2 ml, 6.7 mmol), was added dropwise to a solution of freshly distilled diisopropylamine (894 μ l, 6.4 mmol) in THF (6.5 ml) at -78 °C. The resulting solution was stirred for 1 hour. A solution of ethyl (*R*)-3-hydroxybutyrate (400 mg, 3.0 mmol) and hexamethylphosphoramide (900 μ l) in THF (2 ml) was added dropwise to the

reaction over 15 min. The mixture was then warmed to RT for 20 min and then cooled back to -78 °C. Iodomethane (236 µl, 3.79 mmol) was added to the reaction and this was stirred at 0 °C for 2 h. On completion saturated NH₄Cl (8 ml) was added to the reaction and then was acidified with aqueous HCl (2 M) until pH 7 was reached. The solution was then extracted with diethyl ether (3 × 10 ml) and the organics were combined, dried (MgSO₄) and concentrated *in vacuo*. The resulting oil was purified by silica chromatography (ethyl acetate:hexane 1:1) to yield the desired product (a pale yellow oil, 306 mg, 2.1 mmol, 69%, Rf 0.47, ethyl acetate:hexane 1:1); $\delta_{\rm H}$ (200 MHz, CDCl₃); 4.17 (2H, q *J* 7.1, H₂-7), 3.88 (1H, qdd *J* 6.4 6.4 6.4, H-3), 2.69 (1H, br d *J* 5.8, H-5), 2.44 (1H, p *J* 7.2, H-2), 1.31-1.16 (9H, m, H₃-4 H₃-6 H₃-8); $\delta_{\rm C}$ (100 MHz, CDCl₃); 176.1 (C-1), 69.6 (C-3), 60.7 (C-7), 47.1 (C-2), 20.9 (C-4), 14.3 (C-8), 14.3 (C-6); MS (ES +): *m/z* (%) 147 (100, [M]H+), 129 (11, [M + H - H₂O]+), 101 (8, [M + H - CH₃CH₂OH]+).

Ethyl 2*R*-methyl 3*R*-(methylsulfonyl)oxybutanoate 33

Mesyl chloride (325 µl, 4.2 mmol) was slowly added to a solution of ethyl 3-(*R*)- hydroxy-2-(*R*)-methyl butyrate (307 mg, 2.1 mmol) and pyridine (1.7 ml, 10.5 mmol) in CH₂Cl₂ (5 ml) at 0 °C. The reaction was stirred at 0 °C and followed by TLC and on completion water (10 ml) was added to the solution and then this was extracted with CH₂Cl₂ (3 × 10 ml). The organics were combined, dried (MgSO₄) and concentrated *in vacuo*. The resultant oil was further purified by silica column chromatography (hexane:ethyl acetate 3:1) to yield the desired product (colourless oil, 360mg, 1.6 mmol, 76%, Rf 0.22 hexane:ethyl acetate 3:1); $\delta_{\rm H}$ (400 MHz, CDCl₃); 4.97 (1H, dq *J* 7.8 6.3, H-3), 4.16 (2H, q *J* 7.1, H₂-7), 2.99 (3H, s, H₃-5), 2.77 (1H, qd *J* 7.2 7.2, H-2), 1.44 (3H, d *J* 6.4, H₃-4), 1.27 (3H, t *J* 7.1, H₃-8), 1.20 (3H, d *J* 7.2, H₃-6); $\delta_{\rm C}$ (100 MHz, CDCl₃); 173.4 (C-1), 80.3 (C-3), 61.1 (C-7), 45.3 (C-2), 38.5 (C-5), 18.5 (C-4), 14.3 (C-8), 12.9 (C-4); MS (ES +): *m/z* (%) 471 (9, [M₂]Na+), 449 (25, [M₂]H+), 225 (8, [M]H+), 170 (14), 129 (100, [M + H - MsOH]+); ν_{max} (oil)/cm⁻¹ 2986 (C-H), 2953 (C-H), 1729 (C=O), 1458 (C-H), 1351 (S=O), 1333 (S=O), 1205 (S=O), 1169 (S=O), 1073 (C- O), 1027 (C-O); HRMS (ESI +): calcd for [M]H+ , C₈H₁₆NO₅SNa: 255.0797: found 255.0799.

$2R, 3S-[3-^{2}H]-30$

Ethyl 2-methyl-3-(methylsulfonyl)oxybutanoate (273 mg, 1.2 mmol) was dissolved in anhydrous diethyl ether (1 ml) and was added to $LiAlD_4$ (76 mg, 1.8 mmol) under dry N₂. The reaction was warmed to RT and stirred for a further 2 h. On completion saturated sodium sulfate (3 ml) was added and the solution was extracted with diethyl ether (3 × 5 ml), dried (MgSO₄) and concentrated carefully in *vacuo*. This yielded 189 mg of the crude material (colourless oil) that was used without further purification or characterization.

Jones reagent was prepared by taking chromium oxide (1.6 g) and adding conc. H_2SO_4 (1.3 ml) and then the solution was diluted to a final volume of 6 ml with water. The freshly prepared Jones reagent was added to the previously produced oil (189 mg) in acetone (4 ml) at 0 °C until a permanent red colour was seen. The reaction was stirred for 1 hour and then excess Jones reagent was quenched by slowly adding *iso*-propanol. The acetone was removed in *vacuo* and the remaining green precipitate was redissolved in water (5 ml) and extracted with chloroform (3 × 5 ml). The organics were dried and concentrated *in vacuo*. This yielded a colourless oil (117 mg, Rf 0.42, hexane:ethyl acetate 8:2) which was used without further purification or characterization.

The oil (117 mg) was then dissolved in CH₂Cl₂ (2 ml) and cooled to 0 °C and to it was added *S*-methyl mandelate (237 mg, 1.4 mmol), DMAP (5 mg) and EDCI (234 mg, 1.4 mmol). The reaction was warmed to RT and followed by TLC. On completion aqueous HCl (2M, 2 ml) was added and the reaction was extracted with CH₂Cl₂ (3 × 3 ml), dried (MgSO₄) and concentrated *in vacuo*. The resulting oil was purified by silica flash chromatography (hexane:ethyl acetate 3:1) and yielded the desired product (a pale yellow oil, 42 mg, 167 µmol, 14 % across the three steps, Rf 0.48, hexane:ethyl acetate 3:1): $\delta_{\rm H}$ (400 MHz, CDCl₃); 7.49-7.46 (2H, m, H₂-10), 7.40-7.37 (3H, m, H₂-11 H-12), 5.93 (1H, s, H-6), 3.72 (3H, s, H₃-8), 2.53 (1H, p *J* 7.1, H-2), 1.73 (1H, p *J* 7.4 *JHD* 1.8, H-3), 1.24 (3H, d *J* 7.0, H₃-5), 0.91 (3H, d *J* 7.1, H₃-4); $\delta_{\rm C}$ (100 MHz, CDCl₃); 176.1 (C-1), 169.5 (C-7), 134.1 (C-9), 129.3 (C-12), 128.9 (C-11), 127.7 (C-10), 74.2 (C-6), 52.7 (C-8), 40.9 (C-2), 26.4 (C-3, *J*CD 19.6), 16.6 (C-5), 11.6 (C-4); MS (ES +): *m/z* (%) 252 (83, [M]H+), 192 (11, [M + H - COOHCH₃]+), 149 (100, [M + H - CH₃CHDCHCH₃COOH]+); vmax (oil/cm⁻¹ 2963 (C-H), 2925 (C-H), 2851 (C-H), 1755 (C=O), 1737 (C=O), 1498 (C=C-C), 1455 (C=C-C), 1436 (C=C-C), 1215 (C-H aromatic), 1533 (C-H aromatic), 1124 (C-H aromatic), 1084 (C-H aromatic), 732 (C-H aromatic), 696 (C-H aromatic); HRMS (ESI +): calcd for [M]Na+, C1₄H₁₇O₄DNa: 274.1166: found 274.1166.

2.0 Production and Purification of ER protein

All media and Gilson tips were autoclaved (steam, 121 °C, 10 min) before use. Media used was LB agar [Agar (3 g), tryptone (2 g), yeast extract (1 g), NaCl (1 g) and water (200 ml)] and 2TY media [Tryptone (16 g), yeast extract (10 g), NaCl (5 g) and water (1 L)] and kanamycin was added to the media (100 μ L of stock (30 mg/ ml) per 100 ml of media).

2.1 SQTKS isolated ER domain

The sequence of SQTKS¹⁴ was used to design an *E. coli* optimised sequence which was used as the template for the amplification of the ER domain. Primers for the ER domain with added bases in bold and restriction sites underlined. FW-Nde1: **CAC CAT ATG** GAA CCC TTT CAT CAG CCG GGG AAG C RV-BamH1: **AAA GGA TCC** TTA TGG CGC GGT GAT GAC AAT TTT GC. The PCR product was cloned into pET28a and the plasmid transformed into *E. coli* BL21 DE3 (Novagen), 100 μ l of the culture was spread on a LB agar plate with kanamycin and this was incubated overnight at 37 °C. A single colony was taken to form a starter culture (50 ml 2TY media with kanamycin added), which was incubated overnight at 37°C with shaking. The starter culture was then used to inoculate 2TY media (1:100 starter culture:media dilution per flask). The OD was monitored and when it reached ~0.6 A the flasks were cooled to 16 °C and IPTG solution (1 M, 50 μ l, 0.5 mM final IPTG concentration) and the flasks were left overnight at 16 °C with shaking (250 rpm).

On completion the media was centrifuged (7000 rpm, 15 min) and the pellet was collected. The cells were resuspended in nickel column wash buffer (50 ml buffer per 1 l of media, [50 mM Tris pH 8, 150 mM NaCl, 10% glycerol (v/v) and 20 mM imidazole]). The suspension was then sonicated (6.5 min, 30 seconds 20% power, 30 seconds rest) on ice and then centrifuged (17000 rpm, 30 min). The supernatant was taken, filtered (0.45 μ m filter) and purified on a Ni²⁺ affinity column using a linear gradient with the elution buffer [50 mM Tris pH 8, 150 mM NaCl, 10% glycerol (v/v) and 0.5 M imidazole] to 100%. The fractions were analyzed by SDS page and fractions containing protein of approximately the right mass were combined and concentrated to ~2 ml using centrifugal ultrafiltration (10 kDa cut-off).

The concentrated protein was purified using size exclusion chromatography. The protein was loaded directly onto the column and eluted in the size exclusion elution buffer [50 mM Tris pH 8, 150 mM NaCl, 20% glycerol (v/v)]. The fractions were analyzed by SDS page and fractions containing protein of approximately the right mass and high purity were combined together and concentrated to ~2 ml using centrifugal ultrafiltration (10 kDa cutoff). The concentration of protein was estimated using a Bradford assay and the activity (U) of the enzyme was measure. The protein was then divided into aliquots (0.2 U/µl) and stored long term at -80 °C.



Fig S2.1.1. Nickel affinity purification of SQTKS ER domain: Lane 1, molecular weight markers / kDa; Lane 2, cell-free lysate; lanes 3-9 fractions eluted with imidazole gradient.



Figure S2.1.2 SDS-PAGE showing the purification of isolated ER domain by size exclusion chromatography.

The identity og the protein was confirmed by MALDI analysis of digested protein fragments obtained directly from the SDS-PAGE gel.

MEPFHQPGKLLQMGIKTPGLIDTLQFSKTDATDNLPNDYIEIEPKAFGLNFRDV MVAMGQLEESIMGFECAGVVRRVGPSSAGHNIKVGDRVCALLGGQWTNTVR VHWHSVAPIPQAMDWETAASIPIVFVTAYISLVKIARMQAGETVLIHAASGGVG QAAIILAKHVGAEIFATVGTDEKRDLLIKEYKIPDDHIFSSRNALFAKSIRQRTNG KGVDVVLNCLAGGLLQESFDCLADFGRFIEIGKRDIELNHCLNMGMFARSATFT AVDLIAIGRDRSYMFAEALPKIMTLLQEKAIRPVTPISIYKIGDIETAFRLMQAGK HMGKIVITAP

Figure 2.1.3. Red text shows peptide sequences detected by MS from MALDI analysis of digested protein.



Fig 2.1.4. Combined ES+ spectrum from digestion of ER and matched peptides. Matched Peptides are highlighted in red and cleavage sites are underlined.

2.2 Defining and measuring the activity units for the isolated ER domain

1 unit of activity is defined as 1 μ M·min⁻¹ tigloyl pantetheine reduction by NADPH at 30 °C. The freshly produced protein was analyzed by a standard UV assay (section 3.2.2) and the produced enzyme was diluted to the appropriate concentration (typically 0.2 mU/µl) with the size exclusion buffer.

2.3 Bradford Assay

Standard solutions of bovine serum albumin (0.1-2 ml/ ml) in size exclusion buffer [50 mM Tris pH 8, 150 mM NaCl, 20% glycerol (v/v)] were prepared by serial dilution. 100 μ l of the standards were mixed with Bradford dye reagent (1 ml) and incubated for 15 min at RT. The absorption of each sample was measured at 595 nm against a standard (size exclusion buffer 100 μ l, Bradford dye reagent, 1 ml) to construct a standard concentration curve. A sample of the protein to be quantified (20 μ l) was diluted in size exclusion buffer (80 μ l) and treated with Bradford dye reagent (1 ml). This was incubated at room temperature for 15 min and then the absorption was measured at 595 nm. This was compared to the previously prepared concentration curve to calculate the amount of protein that had been produced.

2.4 Evidence for Dimeric State of ER in Solution

The protein solution was loaded onto the column (GE Healthcare HiLoad 26/600 Superdex 200 pg) and eluted with 50 mM Tris pH 8, 150 mM NaCl, 20% glycerol (v/v). In the run no concentration change take place. The flow of the FPLC was constant 1mL/min over 300 min.



Calibration:

For the calibration a mixture of three different proteins (Carbonic Anhydrase, BSA, Apoferritin) was loaded on the column. The mixture was loaded onto the column (GE Healthcare HiLoad 26/600 Superdex 200 pg) and eluted with 50 mM Tris pH 8, 150 mM NaCl, 20% glycerol (ν/ν). The flow of the FPLC was constant 1mL/min over 300 min.



3.0 Kinetic and Inhibition Assay procedures

3.1 LCMS assay of ER

A sample of reaction mixture (20 μ l) was mixed with acetonitrile (80 μ l) to precipitate the protein and then centrifuged (13,000 rpm, 1 min). The supernatant was analyzed directly by LCMS using a standard LCMS profile (Section 6.1).



Fig S3.1.1. Enzyme reactions contained 0.5 mM tiglyI-SNAC (6.73 min peak), 0.5 mM NAD(P)H, 0.1 mg SQTKS ER per 200 µL reaction, and 50 mM Tris-HCI pH 7.5. After an overnight incubation at 30 °C, protein was precipitated from the reaction mixtures by the addition of 10 µL of assay mixture to 40 µL of acetonitrile and removed by centrifugation. A portion of the supernatant was analysed by LCMS. Use of NADH (top chromatogram) gave no new product. Use of NADPH (bottom chromatogram) resulted in the formation of product (7.06 min).



Fig S3.1.2. ESMS spectra for 6.7 min peak (tigloyl SNAC, top) and 7.06 min peak (methylbutyryal SNAC, bottom).

3.2 Kinetic Assays

3.2.1 Comparative Kinetic Assays

Comparative assays of SNAC and Pantetheine activity were conducted using a Spectramax Plus-382 96-well plate reader at 340 nm over several hours at 30 °C. Four times more enzyme was used for SNAC assays than for Pantetheine assays in order to observe meaningful conversion rates.

Assays with acyl-SNAC substrates were performed using stocks of acyl-SNAC (10 mM), NADPH (10 mM), Tris pH 8.5 (1 M), glycerol (50% v/v) and SQTKS ER (0.2 mU/µl). Assays were conducted at (final concentration, volume of stock): acyl-SNAC (0.5 mM, 10 µl), NADPH (0.75 mM, 15 µl), Tris buffer pH 8.5 (50 mM, 10 µl), glycerol (20% v/v, 80 µl), SQTKS ER (4 mU, 20 µl) and water (65 µl), to a total volume of 200 µl.

Assays with acyl-PANT substrates were performed using stocks of acyl-PANT (10 mM), NADPH (10 mM), Tris pH 8.5 (1 M), glycerol (50% v/v) and SQTKS ER (0.2 mU/ μ l). Assays were conducted at (final concentration, volume of stock): acyl-PANT (0.5 mM, 10 μ l), NADPH (0.75 mM, 15 μ l), Tris buffer pH 8.5 (50 mM, 10 μ l), glycerol (20% v/v, 80 μ l), SQTKS ER (1 mU, 5 μ l) and water (80 μ l), to a total volume of 200 μ l.



Fig S3.2.1. Kinetic analysis of the substrate activity of 5P vs 5S. Note that the 5S assay contained 4 x as much enzyme as the 5P assay.

3.2.2 Quantitative Kinetic Assays

Assays were run in 500 μ l quartz cuvettes with a path length of 1 cm and the absorption was measured at 340 nm against a standard cuvette at 30 °C. Assays with acyl-PANT substrates were performed using stocks of acyl-PANT (5 mM), NADPH (10 mM), Tris pH 8.5 (1 M), glycerol (50% ν/ν) and SQTKS ER (0.08 U/ μ l). Assays were conducted at (final concentration, volume of stock): acyl-PANT (62.5 μ M - 500 μ M, 5 - 40 μ l); NADPH (250 μ M, 10 μ l); SQTKS ER (20 μ L, 4 mU); Buffer (330 - 365 μ l) in a total volume 400 μ l. Reactions were run in triplicate. Initial rates were plotted *vs* substrate concentration (section 3.4).

3.4 Kinetic Data for Substrate Variants

Initial rate data was collected over a range of substrate concentrations (0-100 μ M as appropriate) and kinetic parameters were obtained by direct fit of the data to the Michaelis Menten equation (see below). All data is *apparent* at fixed NADPH concentration.

	Substrate	<i>K_M</i> / μM	<i>k_{cat}</i> / s ⁻¹	$\mathbf{k}_{cat} / \mathbf{K}_{M} \mathrm{s}^{-1} \mu \mathrm{M}^{-1}$
7D	0	0.12	3.7	30.8
71	R			
= 0	0	0.69	8.9	12.9
5P	R			
	I Q	0.44	11.4	25.9
8P				
	0=	0	0	0
9P	R			
	0	0.16	2.58	16.1
10P	R			
		0.00		110 (
11P	∼ ~ R	0.23	25.9	112.6
12P	O R	0.54	415	768.5
	0	0.18	5.9	32.8
13P	R	0.10	5.7	52.0
14P		1.85	169	91.4
	0	0.41	40.0	110.0
15P	\bigwedge	0.41	48.8	119.0
4.60		0.19	4.71	24.8
16P	R			
17P		0.18	7.11	39.5
18P	→→→ R	0.52	5.6	10.8
19P		0.6	9	15
20P		1.70	15.7	9.2
21P		4.70	0.85	0.2
22P		0.18	2	11.1
1 D		0	0	0
24P	γ Υ Υ [™] κ	0.26	10	27.0
24P 25D		0.30	6	27.0 30
23P		0.2	0	50

Table S3.4: Kinetic data different pantetheine	substrates
R = S-Pantetheine residue	







9P 2-3-dimethylbut-2-enoylpantethein

It was not possible to measure any rate. No product observed.

10P *E*-2-Ethylbut-2-enoylpantetheine



11P

E-Hex-2-enoylpantetheine



12P *E*-2-methylhex-2-enoylpantetheine









15P *E*-4*S*-2,4-dimethylhex-2-enoylpantetheine



16P E-2-Ethylhex-2-enoylpantetheine



17P E-hept-2-enoylpantetheine



18P E-6-methylhept-2-enoylpantetheine



19P *E*-oct-2-enoylpantetheine



20P E-2-methyloct-2-enoylpantetheine



21P E-7-methyloct-2-enoylpantetheine



22P 4RS,6RS-E-4,6-dimethyloct-2-enoylpantetheine



1P *E*-4*S*-6*S*-4,6-dimethyloct-2-enoylpantethein It was not possible to measure any rate.

24P E-dec-2-enoyl pantetheine



25P E-2-methyldec-2-enoyl pantetheine



3.3 Inhibition Assay

Procedere as for Section **3.2**. Assays were performed in total volume of 400 µl; including: 5 - 40 µl Substrate (5 mmol); 10 µl NADPH (10 mmol); 20 µl Enzyme (def. 0.02 mU); 320-355 µl Buffer. (PANT); 5-10 µl SQTK (5 mmol).



4.0 Stereochemical Assay Procedures

4.1 Analysis using diphenylethylene diamine

NMR assays were performed with tiglyl-SNAC **5S** (2.0 mg, 10 mmol), NADPH (8 mg, 10 mmol), tris buffer pH 8.5 (50 μ l, 1M stock), glycerol (20% v/v) and the isolated ER domain (380 μ U, 100 μ l of stock) in water (total volume 1 ml). The progress of the reaction was monitored by LCMS. After 24 h further NADPH (8 mg, 10 mmol) was added to the reaction. After a further 17 h the protein was precipitated with CH₂Cl₂ (1 ml) and centrifuged (3000 rpm, 20 min). The organic layer was removed and the aqueous layer was further extracted with CH₂Cl₂ (2 × 1 ml). The organic fractions were combined and the solvent was removed under a flow of N₂ and then dried in *vacuo* for a further 2 h. To the dried sample was added water (450 μ l) and aqueous NaOH (2M, 50 μ l). The sample was left for 90 mins at RT and then acidified with aqueous HCl (2M, 100 μ l) to pH 3 and extracted with CDCl₃ (4 × 200 μ l). The organics were combined, dried with MgSO₄ and then filtered through MgSO₄ directly into an NMR tube. (1*R*, 2*R*)-(+)-1, 2 diphenylethylenediamine was titrated (100 mM stock) to optimize resolution of the obtained spectra.

4.2 Formation and Analysis of Methyl Mandelates

A solution of tiglyl/angelic-PANT **5P/8P** (3.6 mg, 10 mmol), NADPH (856 μ l of 12.5 mM stock), tris buffer pH 8.5 (50 μ l 1M stock), glycerol (500 μ l, 50% v/v) and SQTKS ER (80 mU, 400 μ l, 0.2 mU/ μ l stock) was incubated at 30 °C and the progress of the reaction was monitored by LCMS. After 24 h further SQTKS ER (200 μ l) was added and the reaction mixture was incubated at 30 °C for 24 h. On completion CH₂Cl₂ (1 ml) was added and the sample was centrifuged (3000 rpm, 20 min) and the organics removed. The aqueous layer was extracted with CH₂Cl₂ (2 × 2 ml) and the organics were combined and concentrated under a flow of N₂ and then further concentrated *in vacuo* for 2 h.

After concentration, water (450 µl) and aqueous NaOH (2M, 50 µl) were added. This was left for 90 min and then acidified with aqueous HCl (2M, 100 µl) and then aqueous CuSO₄ (saturated, 400 µl) was added to remove panthetheine. The copper solution was extracted with CH_2Cl_2 (3 × 1 ml) and the organics were dried (MgSO₄) and concentrated to 1 ml under a flow of N₂. To the concentrated solution was added *S*-methyl mandelate (2.1 mg, 12.6 µmol), EDCI (2.4 mg 12.6 µmol) and DMAP (0.1 mg) and the reaction was followed by TLC. On completion aqueous HCl (2M, 0.5 ml) was added and the solution was extracted with CH_2Cl_2 (3 × 0.5 ml). The organics were dried (MgSO₄) and concentrated with N₂ and redisolved in CDCl₃ and were analyzed directly by NMR.

4.3 Analysis of use of 4'-2H NADPH isomers



Fig S4.3.1. MS analysis of reduction of tigloyl pantethiene by isotopically labelled ²H-isomers of NADPH.

5. Stereochemical Assignment of Proton Chemical Shifts of 2-Methyl Butyric Acid.



1. Synthetic 2R material identifies chemical shift of 4-methyl at 0.92 ppm. Therefore chemical shift of 4-methyl in 2S material is 0.98 ppm.

2. Synthetic 2R material identifies chemical shift of anti 3-H at 1.75 ppm (green); thus syn 3-H resonates at 1.52 ppm (orange).

3. Reduction of *E*-2-methyl butyrate by ER gives only two products.* One is the 2*R*,3*R* (orange) diastereomer showing 3-*Re* face attack by D. The other is 2*S*, and H is therefore *anti*, thus 3*R* (purple).



4. The final ¹H resonance in the COSY must therefore arise from the 2S3-syn proton (blue).



5. Reduction of *Z*-2-methyl butyrate by ER gives two products. These are asigned as the 2*S* 3-*syn* (blue) and 2*R* 3-*anti* (green) diastereomers and this shows that D addition occurs at the 3-*Si* face, *i.e* with the same absolute sense as reduction of the *E*-substrate.



* Since it is known that the reprotonation step gives racemate at C-2, the origin of the 2-diastereomers observed must be due to the two configurations at C-2. Thus the configurations at C-3 must be identical. The substrate must be bound in a fixed orientation for only two products to be observed.

6. LCMS conditions

6.1 Analytical LCMS

LC-MS data were obtained with using a Waters LCMS system comprising of a Waters 2767 autosampler, Waters 2545 pump system, a Phenomenex Kinetex column (2.6 μ , C₁₈, 100 Å, 4.6 × 100 mm) equipped with a Phenomenex Security Guard precolumn (Luna C₅ 300 Å) eluted at 1 ml/min. Detection was by Waters 2998 Diode Array detector between 200 and 600 nm; Waters 2424 ELSD and Waters SQD-2 mass detector operating simultaneously in ES + and ES- modes between 100 *m/z* and 650 *m/z*. Solvents were: A, HPLC grade H₂O containing 0.05% formic acid; B, HPLC grade MeOH containing 0.045% formic acid; and C, HPLC grade CH₃CN containing 0.045% formic acid. Gradients were as follows. *Method 1*. Kinetex/CH₃CN: 0 min, 10% C; 10 min, 90% C; 12 min, 90% C; 13 min, 10% C; 15 min, 10% C.

6.2 Semi-Preparative LCMS and Compound Purification.

Purification of all pantetheines was generally achieved using a Waters mass-directed autopurification system comprising of a Waters 2767 autosampler, Waters 2545 pump system, a Phenomenex Kinetex Axia column (5 μ , C₁₈, 100 Å, 21.2 × 250 mm) equipped with a Phenomenex Security Guard precolumn (Luna C₅ 300 Å) eluted at 20 ml/min at ambient temperature. Solvent A, HPLC grade H₂O + 0.05% formic acid; Solvent B, HPLC grade CH₃CN + 0.045% formic acid. The post-column flow was split (100:1) and the minority flow was made up with HPLC grade MeOH + 0.045% formic acid to 1 ml·min⁻¹ for simultaneous analysis by diode array (Waters 2998), evaporative light scattering (Waters 2424) and ESI mass spectrometry in positive and negative modes (Waters SQD-2). Detected peaks were collected into glass test tubes. Combined tubes were evaporated (vacuum centrifuge), weighed, and residues dissolved directly in solvent for use or analysis.

7.0 Multiple alignment of ER sequences

SQTKS Azanigerone Spin Geldanmycin Borrelidin erythromycin Megalomicin vFAS RHFPLE	EPFHQ IHCRFRD QLALRRGGAH QVALRRGLLW ELVPPAG RLSPLAP RLTPLAG Q	APRLAGLGSD VPRLVRSP	PGKL AARP DVLPVPDGTG QGLAVPAHEH ADA -SALTLPAGT -PVPAVADRA -DRPEKQTEH	LQMGIKTPGL LRMKIGTPGL WRLEATRPGS WYLDVSEKGS WRLEITEPGT QRLVPGDG-A YRLVPGNGGS AF <mark>V</mark> NVLSRGD	I TLQFS-KT I TLVFHDDL LDGLALVDEP LENIVLRPDP FINLTLGVYP I SVAFEPAP I RAVAFAPVP LSTRWVCSP	DATDN-LPND DAKSDPLPAD TATAP-LGDG EATAP-LATG HAEKT-LADN DVEQP-LRAG DADRP-LAPE LHYALPASCQ	1911 1860 239 2519 1429 2914 2843 1561		
SQTKS Azanigerone Spin Geldanmycin Borrelidin erythromycin Megalomicin vFAS	YIEIEPKAFG WIEFDPTAFG EVRIAMRAAG QVRIEVRAAG EVRVAVHAGG EVRVDVRATG DRLCSVYYT <mark>S</mark>	LNFRDVMVAM LNFRDVMVAM VNFRDALIAL QNFRDVLVAL LNFHDVVAAL VNFRDVLLAL LNFRDVLLAL LNFRDVLLAL	Substrate GQLEE-SIMG GQLEANAIMG GMYPGVASLG GGVAGQEGLG GMVEDDLTLG GMYPQKADMG GMYPEPAEMG GKLSPDSIPG	Binding FECAGVVRRV FECAGTIVRL SEGAGVVVEF GEGAGVVTEV REAAGVVVEV TEAAGVVTAV TEASGVVTEV KWLT-RDCM	GPSSAGHNIK GATAAAKGFA GPGVTGLA GPGVEGLA GDAVPDLI GPDVDAFA GSGVRRFT G-MEFSGRDA	VGDRVCALLG VGDRVCTLLR PGDRVMGNIP VGDRVMGLFP PGDHVMGLS PGDRVLGLFQ PGQAVTGLFQ SGRRVMGVP	1970 1920 297 2577 1487 2972 2901 1618		
SQTKS Azanigerone Spin Geldanmycin Borrelidin erythromycin Megalomicin vFAS	GQ-WINTVRV GH-WAIRERA KA-FGELAVA RS-FGELAIA SG-FGELAVT GA-FAEIEVT GA-FGEVAVA AEG <mark>LAISV</mark> LL	HWHSVAPIPQ PWTSVMRIPQ DHRMVTRIPA DARTVAPIPE DHRYLARMPE DHRLLARVPD DHRLLTPVPD LQHATWEVPS	AMDWETAASI HISDQEAASF GWSFARAASV GWSFARAASV GWTFAQAASV GWSDADAAAV GWRAVDAAAV FWTLEEAASV	CC PIVEVTAYIS PTVEATAYIA PIVELTAYYA PVAYLTALYG PAAELTAYYG PIAETTAEYA PIAETTAEYA PIVYTTAYYS	DÍACTOT LVXIARMQAG LHETARLORG LVDLAGLROPG LRDLGTVOPG LCDLGGIRAG LHDLAGLRAG LHDLAGLQAG LVVRGRMQPG	ROSS ETVLIHAASG ESILIHAAG ESILVHSAAG ETVLVHAAAG OSVLIHAAG OSVLIHAAG SVLIHAAG SVLIHAAG	2029 1979 356 2636 1546 3031 2960 1678		
SQTKS Azanigerone Spin Geldanmycin Borrelidin erythromycin Megalomicin vFAS	MANN GVGQAAIQLA GVGQAAIQLA GVGMAAIQLA GVGMAAVQLA GVGMAAVALA GVGMAAVALA GVGQAAIALA	KHVGAEIFAT OLIGAEIYAT RHLGAEVYAT RHLGAEVYAT RHLGAEVFGT RRAGAEVLAT RRAGAEVFAT LSRGCRVFTT	Cofac VGTDEKRD-L ASTPAKRQ-L ASE-DK AHP-SKHH ASP RKWG ASP AKHG ASP AKHP VGSAEKRAYL	LIKEYK <mark>I</mark> PDD LHETYGIPEN -WQAVELSRE VLTALGVPEG ALRALGLDDA TLRALGLDDE TLRALGLDDE QARFPQLDET	HIFSSRNALF NIFSSRDPSF HLASSRTCDF HLSSSRTLDF HISSSRTLDF HIASSRETGF HIASSRESGF GFANSRDTSF	AKSIRQRTNG ATDVHLRTDG EQQFLGATGG ASAFP EQEFLDATDG ARKFRERTGG GERFAARTGG EQHVLRHTAG	2088 2038 410 2688 1603 3088 3017 1738		
SQTKS Azanigerone Spin Geldanmycin Borrelidin erythromycin Megalomicin vFAS	KGVDVVLNCL RGVDVVLNSL RGVDVVLNSL -ALDVVLNSL RGVDLVLNSL RGVDVVLNSL KGVDVVLNSL KGVDLVLNSL	AGGLLQESFD AGRILQESFN AGEFADASLR IGEYVDASLG AREFVDASLR IGELLDESAD IGDLLDESAR AEEKLQASVR	CLADFGRFIE CLAEFGRMVE MLPRGGRFVE LLGTGGRFVE LMPGGGRFVD LLAEDGVFVE LLADGGVFVE CLAQHGRFLE	IGKRDIELNH IGKRDLEQHS LGKTDVRDPV MGKNDIRDPA MGKTDIRPE MGKTDLRD MGKTDLRP IGKFDLSNNH	CLNMGMFARS GLDMYPFTRN EVADAHPG SVAAHPG QVAEDHGG AGDFRS ALGMAVFLKN	ATETAVDLIA VSFSSVDLLT VSYQAFDT VGYQAFDL R-YAPFDL R-YVPFDL VTFHGTLLDS !**.	2148 2098 466 2744 1559 3139 3068 1798		
SQTKS Azanigerone Spin Geldanmycin Borrelidin erythromycin Megalomicin vFAS	IGRDRSYMFA WQSRRGADIS -VEAGPQRIG -GDAGPDRIR -VEAGPQRTG -GBAGDDRLG -ABAGPDRLG LFEEGGATWQ	EALPKINTLL CVLQSLSKLL EMLHELVELF ELLAELVELF EMLAEIVRLF EILREVVGLL EILEEVVGLL EVSELLKAGI	C-termin QEKAIRPVTP GEKKIMPVYP EGRVIEPLP- EAGRIEPLP- QAGAFRPLP- GAGELDRLP- QEGVVQPLK-	nal ISI <mark>Y</mark> KIGD <mark>IE</mark> LTLYPITQIE VTAWDVRQAP IRHWDVTQAP IRQWDVRRAP VSAWELGSAP VSVWELSAAP CTV <mark>F</mark> PRTK <mark>VE</mark>	TAFRLMOAGK KAFRTMOTGQ EAFRHLSQAR TAFRWMSQGR EAFRHISQAK AALQHMSRGR AALTHMSRGR AAFRYMAQGK	HMGKIVITAP HMGKIIISVG HVGKLVITMP HTGKIVITLP HIGKIVITQP HVGKLVITQP HIGKVVIQVR	2208 2158 524 2802 1717 3197 3126 1857		
= residues = residues substrate.	identified contacting	by Leadlay, NADPH; <mark>*</mark> =	Ban and o residues co	thers as pos ontacting th	ssibly invo ne pantethe:	lved in enoi ine; <mark>î</mark> = res	l(ate) reprotonation; sidues contacting the		
colour code. A = 8/8 conserved; A = 7/8 conserved; A = 6/8 conserved; = 5/8 conserved;									
Domain colours (blue, red, green, grey) and descriptions correspond to those in Figs 4, 5 and 6.									
8.0 Modelling and Docking Procedures

ER Homology Model:

A homology of the ER domain with NADP bound was generated using the SWISS-MODEL protein structure homology-modelling server in the fully automated mode.³³ The PDB entry 2VZ9 was used as the template structure.³⁴

Pantetheine Docking:

Ligand structures were generated using Chem3D Pro 13.0 (Perkin Elmer). The ligands were docked manually into the active site using PyMOL (The PyMOL Molecular Graphics System, Version 1.3 Schrodinger, LLC.) and the resulting protein-ligand complex was energy minimised using the YASARA energy minimization server.³⁵ The docked ligand was removed from the resulting protein model and re-docked using Autdock Vina 1.1.2.³⁶ The ligand files and ER PDB file were converted into PDBQT format using Autdock Tools $1.5.6^{37}$ to add polar hydrogens and set rotatable bonds. Ligands were docked using a grid of 44 Å (x) x 22 Å (y) x 16 Å (z) centred on the active site at position 45.8 (x), 143 (y), 76.556 (z). Images were processed using PyMol.

pdb files of representative docked structures are appended as supplementary information.

9.0 Integration of Angelic Mandelate Spectrum



Integration of crosspeaks for **30** from reduction of **8P** was achieved using MestreNova 9.0 Software using standard parameters.

9.0 References

- 1. Jackson, David A.; Rey, Max; Dreiding, Andre S., *Helv. Chim. Acta*, **1983**, 66, 2330 2341.
- 2. Ramamoorthy, G.; Acevedo, C. M.; Alvira, E.; Lipton, M. A., Tetrahedron: Asymmetry 2008, 19, 2546.
- 3. Hadrami, El, M.; Lavergne, J. P.; Viallefont, P.; Itto, M., Tetrahedron Letters 1991, 32, 3985.
- 4. Rossi, R.; Carpita, A.; Cossi, P. *Tetrahedron* **1992**, *48*, 8801.
- 5. Das, Biswanath; Banerjee, Joydeep; Chowdhury, Nikhil; Majhi, Anjoy; Mahender, Gurram, *Helv. Chimi. Acta*, **2006**, *89*, 876 883
- 6. T. Mimura, Y. Kimura and T. Nakai, *Chem. Lett.*, **1979**, 1361–1364.
- 7. Blanc, P.-Y. et al., *Helv. Chim. Acta*, **1964**, *47*, 567 575
- 8. Concellón, J. M.; Concellón, C., J. Org. Chem. 2006, 71, 1728.
- 9. H. H. Wasserman, A. K. Petersen and M. Xia, *Tetrahedron*, **2003**, *59*, 6771.
- 10. T. E. Lightburn, O. A. De Paolis, K. H. Cheng, and K. L. Tan, *Org. Lett.*, **2011**, *13*, 2686.
- 11. F. Bargiggia and O. Piva, *Tetrahedron: Asymmetry*, **2003**, *14*, 6771.
- 12. Ahad, S. M.; Ange, A. L.; Bates, R. B.; Bell, B. L.;Bodour, A. A.; Bourne, B. R.; Contreras, C. G.; Goldberg, E. L.; Gunatilaka, A. A. L.; King, S.; Lee, A. K.; Low, R. L.; Maier, R. M.; Marlor, K. M.; Marron, M. T.; Scolnik, R. C.; Streeter, M. J.; Strelczuk, M.; Trinh, L. N.; Truong, V. K.; Vissering, S. P.; Weick, M. C.; Williams, M. T., *Tetrahedron*, **2010**, *66*, 9107.
- 13. M. Morr, V. Wray, J. Fortkamp and R. D. Schmid, *Liebigs Ann. der Chem.*, **1992**, 433.
- 14. Cox, R. J.; Glod, F.; Hurley, D.; Lazarus, C. M.; Nicholson, T. P.; Rudd, B. A. M.; Simpson, T. J.; Wilkinson, B.; Zhang, Y., *Chem. Commun.*, **2004**, 2260 2261.
- 15. B. Bonsch, V. Belt, C. Bartel, N. Duensing, M. Koziol, C. M. Lazarus, A. M. Bailey, T. J. Simpson and R. J. Cox, *Chem. Commun.*, **2016**, *52*, 6777–6780.
- 16. L. Raffier and O. Piva, *Beilstein J. Org. Chem.*, **2011**, *7*, 151.
- 17. Lee, M. S.; Qin, G.; Nakanishi, K., J. Am. Chem. Soc., 1989, 11, 6234.
- 18. Moore, Bradley S.; Seng, Dieter, *Tetrahedron Letters*, **1998**, 39, 3915 3918.
- 19. F. Wang, Y. Wang, J. Ji, Z. Zhou, J. Yu, H. Zhu, Z. Su, L. Zhang and J. Zheng, *ACS Chem. Biol.*, 2015, **10**, 1017–1025.
- 20. Jensen, Katja; Niederkrueger, Holger; Zimmermann, Katrin; Vagstad, Anna L.; Moldenhauer, Jana; Brendel, Nicole; Frank, Sarah; Poeplau, Petra; Kohlhaas, Christoph; Townsend, Craig A.; Oldiges, Marco; Hertweck, Christian; Piel, Joern, *Chemistry and Biology*, **2012**, *19*, 329 - 339
- 21. Speir; Barnsley, *Biochem. J.*, **1971**, *125*, 267 269.
- 22. Winter, Jaclyn M.; Cascio, Duilio; Dietrich, David; Sato, Michio; Watanabe, Kenji; Sawaya, Michael R.; Vederas, John C.; Tang, Yi, *J. Am. Chem. Soc.*, **2015**, *137*, 9885 9893.
- 23. Mizugaki, Michinao; Hoshino, Toshiaka; Ito, Yoko; Sakamoto, Takao; Shiraishi, Takayuki; Yamanaka, Hiroshi, Chem. Pharm. Bull., **1980**, *28*, 2347 2350.
- 24. Singh, Renu; Reynolds, Kevin A., ChemBioChem, 2015, 16, 631 640.
- 25. Schwab, John M.; Klassen, John B., J. Am. Chem. Soc., **1984**, 106, 7217 7227.
- 26. Agarwal, Vinayak; Diethelm, Stefan; Ray, Lauren; Garg, Neha; Awakawa, Takayoshi; Dorrestein, Pieter C.; Moore, Bradley S., *Org. Lett.*, **2015**, *17*, 4452 4455.
- 27. M. Koller, M. Karpf and A. S. Dreiding, *Helv. Chim. Acta.*, **1986**, *69*, 560 579.
- 28. Gay, Darren C.; Spear, Philip J.; Keatinge-Clay, Adrian T., *ACS Chem. Biol.*, **2014**, *9*, 2374 2381.
- 29. Ottolina, G.; Riva, S.; Carrea, G.; Danieli, B.; Buckmann, A. F. Biochimica et Biophysica Acta (BBA) Protein Structure and Molecular Enzymology 1989, 998, 173.; Podschun, B.; Jahnke, K.; Schnackerz, K. D.; Cook, P. F. J. Biol. Chem., **1993**, *268*, 3407.
- 30. Jeong, S. S.; Gready, J. E., Anal. Biochem., **1994**, 221, 273.
- 31. Kashima; Fukuchi; Hosomi, J. Org. Chem., **1994**, 59, 7821 7824.
- 32. Micoine, K.; Fürstner, A., J. Am. Chem. Soc., **2010**, 132, 14064.

- M. Biasini, S. Bienert, A. Waterhouse, K. Arnold, G. Studer, T. Schmidt, F. Kiefer, T. G. Cassarino, M. Bertoni, L. Bordoli, T. Schwede, *Nucleic Acids Res.*, **2014**, *42*, W252 - W258; Arnold K., Bordoli L., Kopp J., and Schwede T., *Bioinformatics*, **2006**, *22*, 195 - 201; Kiefer F, Arnold K, Künzli M, Bordoli L, Schwede T, *Nucleic Acids Res.*, **2009**, *37*, D387 - D392; Guex, N., Peitsch, M.C., Schwede, T., *Electrophoresis*, **2009**, *30*, S162 - S173.
- 34. T. Maier, M. Leibundgut and N. Ban, *Science*, **2008**, *321*, 1315-1322.
- 35. E. Krieger, K. Joo, J. Lee, J. Lee, S. Raman, J. Thompson, M. Tyka, D. Baker and K. Karplus, *Proteins*, **2009**, 77, 114 122.
- 36. O. Trott and A. J. Olson, J. Comp. Chem., 2010, 31, 455 461.
- 37. Morris, G. M., Huey, R., Lindstrom, W., Sanner, M. F., Belew, R. K., Goodsell, D. S. and Olson, A. J., J. *Comp. Chem.*, **2009**, *16*, 2785 - 2791; M. F. Sanner, *J. Mol. Graphics Mod.*, **1999**, *17*, 57 - 61.