Supplementary Information for:

Neutral Iridium Catalysts with Chiral Phosphine-Carboxy Ligands for Asymmetric Hydrogenation of Unsaturated Carboxylic Acids

Shuang Yang,^a Wen Che,^a Hui-Ling Wu,^a Shou-Fei Zhu^{*a} and Qi-Lin Zhou^{*ab}

^a State Key Laboratory and Institute of Elemento-Organic Chemistry, Nankai University, Tianjin 300071, China

^b Collaborative Innovation Center of Chemical Science and Engineering (Tianjin), Tianjin 300071, China

CONTENTS:

1. Preparation and Analytical Data of Iridium Complexes with Chiral	Spiro
Phosphine-Carboxy Ligands	S3
2. Preparation and Analytical Data of Unsaturated Carboxylic Acids	S4
3. Asymmetric Hydrogenation and Analytical Data of Products	S6
4. Total Synthesis of (S)-14-Methyloctadec-1-ene	S10
5. X-ray Diffraction Analysis of (S)-2d	S11
6. NMR Spectra of New Compounds	S13
7. HPLC or GC Charts of Hydrogenation Product Derivatives	S29
8. Reference	S45

General Information. Unless otherwise noted, all reactions and manipulations were performed in an argon-filled glovebox (VAC DRI-LAB HE 493) or using standard Schlenk techniques. Melting points were measured on a RY-I apparatus and uncorrected. ¹H, ¹³C and ³¹P NMR spectra were recorded on a Bruker AV 400 spectrometer or a Varian Mercury Plus 400 spectrometer at 400 MHz (¹H NMR), 100 MHz (¹³C NMR) and 162 MHz (³¹P NMR) in CDCl₃. Chemical shifts were reported in ppm down field from internal Me₄Si and external 85% H₃PO₄, respectively. Optical rotations were determined using a Perkin Elmer 341 MC polarimeter. IR spectra were obtained with a Perkin-Elmer spectrometer in KBr disks. HRMS were recorded on IonSpec FT-ICR mass spectrometer with ESI or MALDI resource. Enantiomeric excesses of the asymmetric hydrogenation products were determined by chiral HPLC or GC. HPLC analyses were performed using a Waters 2996 instruments or a Hewlett Packard Model HP 1100 instruments. GC analyses were performed using a Hewlett Packard Model HP 6890 Series instruments. Anhydrous Et₂O, THF and toluene were distilled from sodium benzophenone ketyl, anhydrous CH₂Cl₂, NEt₃, DMF, and pyridine were freshly distilled from calcium hydride under nitrogen atmosphere. Absolute MeOH, EtOH, "PrOH, "BuOH, PrOH, and 'BuOH were distilled from magnesium under nitrogen atmosphere. Hydrogen gas (99.999%) was purchased from Boc Gas Inc., Tianjin. [Ir(COD)Cl]₂ was prepared from IrCl₃ 3H₂O according to the literature procedure.¹

1. Preparation and Analytical Data of Iridium Complexes with Chiral Spiro Phosphine-Carboxy Ligands

(S)-2d



Typical procedure: (S)-1d (112 mg, 0.250 mmol), [Ir(COD)Cl]₂ (84 mg, 0.125 mmol) and Na₂CO₃ (13 mg, 0.125 mmol) were mixed in CH₂Cl₂ (2 mL) in a Schlenk tube under argon atmosphere. The resulting suspension was heated to 40 °C till that the TLC analysis showed no free ligand existed. After cooling to room temperature, the mixture was concentrated under reduced pressure and the residue was purified by a flash column chromatography on silica gel with petroleum ether/ethyl acedate (PE/EA = 1:1, v/v) to offer (S)-2d (133 mg, yield: 71%) as an orange-yellow solid, mp: 190–191 °C. [α]_D²⁵ +277 (c 0.5, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 8.20 (d, J = 5.2 Hz, 1H), 7.64 (s, 2H), 7.48–7.24 (m, 10H), 7.12 (t, J = 7.6 Hz, 1H), 6.69 (s, 2H), 4.41 (s, 1H), 3.55 (d, J = 6.0 Hz, 1H), 3.26 (s, 1H), 2.92–2.75 (m, 5H), 2.43–2.31 (m, 2H), 2.19 (dd, J = 15.7 and 9.3 Hz, 1H), 1.91 (d, J = 2.4 Hz, 1H), 1.55–1.46 (m, 3H), 1.15–1.14 (m, 1H), 0.97-0.96 (m, 1H), 0.78-0.76 (m, 1H), 0.62-0.60 (m, 1H), 0.12-0.09 (m, 1H); ³¹P NMR (161 MHz, CDCl₃) δ 8.9 (s); ¹³C NMR (101 MHz, CDCl₃) δ 174.5, 150.0, 149.9, 146.4, 146.3, 142.9, 140.5, 136.2, 135.7, 132.33, 132.30, 131.7, 131.1, 130.6, 130.3, 129.0, 128.9, 127.7, 127.6, 127.4, 127.3, 127.2, 127.1, 126.0, 125.6, 119.1, 74.4, 64.5, 64.2, 64.1, 63.0, 61.0, 39.7, 35.0 34.9, 34.2, 31.1, 31.0, 30.6, 30.4, 27.1, 27.0. HRMS (ESI) Calcd for [C₃₈H₃₆IrNaO₂P, M + Na]⁺: 771.1974, Found: 771.1977.

(S)-2b



Yield: 75%, orange-yellow solid, mp: 205–206 °C. $[\alpha]_D^{25}$ +318 (*c* 0.5, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 8.24 (d, *J* = 6.0 Hz, 1H), 7.48 (t, *J* = 8.0 Hz, 1H), 7.37–7.32 (m, 3H), 7.27–7.23 (m, 2H), 7.11 (t, *J* = 8.0 Hz, 2H), 7.04 (s, 2H), 5.73 (s, 1H), 4.40 (td, *J* = 7.6 and 2.2 Hz, 1H), 3.57–3.52 (m, 1H), 3.20 (t, *J* = 7.2 Hz, 1H), 2.92–2.67 (m, 5H), 2.46–2.38 (m, 2H), 2.31 (s,

6H), 2.22 (s, 6H), 2.09 (dd, J = 16.0 and 9.2 Hz, 1H), 1.95–1.88 (m, 1H), 1.58–1.42 (m, 3H), 1.23–1.15 (m, 1H), 1.05–0.96 (m, 1H), 0.79–0.73 (m, 1H), 0.60–0.50 (m, 1H), 0.12 (dd, J = 22.1 and 10.1 Hz, 1H); ³¹P NMR (161 MHz, CDCl₃) δ 9.9 (s); ¹³C NMR (101 MHz, CDCl₃) δ 174.5, 149.9, 149.8, 146.2, 146.1, 143.2, 140.5, 138.3, 137.0, 136.9, 135.6, 135.2, 133.4, 132.7, 132.42, 132.38, 132.04, 132.02, 130.4, 129.7, 129.3, 127.9, 127.6, 127.1, 127.0, 125.8, 125.2, 118.6, 73.7, 64.1, 63.9, 63.8, 63.0, 61.1, 39.6, 34.83, 34.77, 34.1, 31.1, 30.8, 30.6, 30.3, 27.4, 27.3, 21.5, 21.4. HRMS (ESI) Calcd for [C₄₂H₄₄IrNaO₂P, M + Na]⁺: 827.2600, Found: 827.2603.

(S)-2c



Yield: 78%, orange-yellow solid, mp: 195–196 °C. $[\alpha]_D^{25}$ +273 (*c* 0.5, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 8.19 (dd, *J* = 5.5 and 2.3 Hz, 1H), 7.57 (t, *J* = 10.4 Hz, 2H), 7.39–7.33 (m, 3H), 7.25–7.23 (m, 1H), 7.13–7.06

(m, 2H), 6.93 (d, J = 10.0 Hz, 2H), 6.81 (d, J = 10.4 Hz, 2H), 6.59 (s, 1H), 4.45 (dd, J = 7.3 and 5.5 Hz, 1H), 3.86 (s, 3H), 3.81 (s, 3H), 3.57–3.52 (m, 1H), 3.23–3.20 (m, 1H), 2.92–2.73 (m, 5H), 2.40–2.18 (m, 3H), 1.95–1.87 (m, 1H), 1.60–1.21 (m, 4H), 1.07–1.01 (m, 1H), 0.87–0.66 (m, 2H), 0.25 (dd, J = 22.2 and 10.3 Hz, 1H); ³¹P NMR (161 MHz, CDCl₃) δ 6.4 (s); ¹³C NMR (101 MHz, CDCl₃) δ 174.5, 161.7, 160.2, 149.8, 149.7, 146.2, 146.1, 143.0, 140.6, 137.2, 134.3, 132.1, 132.0, 131.9, 129.6, 128.7, 128.3, 127.4, 127.1, 127.0, 126.9, 125.9, 125.6, 121.3, 120.9, 119.2, 114.5, 113.2, 113.1, 74.4, 64.3, 64.0, 63.6, 63.0, 60.8, 55.5, 55.4, 39.8, 34.8, 34,7, 34.2, 31.3, 31.1, 30.6, 30.5, 27.1, 27.0. HRMS (ESI) Calcd for [C₄₀H₄₀IrNaO₄P, M + Na]⁺: 831.2186, Found: 831.2182.

(S)-2a



Yield: 83%, orange-yellow solid, mp: 212–213 °C. $[\alpha]_D^{25}$ +200 (*c* 0.5, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 8.17 (d, *J* = 6.8 Hz, 1H), 7.98 (s, 1H), 7.62 (t, *J* = 8.4 Hz, 1H), 7.43 (s, 1H), 7.38 (s, 1H), 7.34–7.26 (m, 3H), 7.15 (t, *J* = 7.2 Hz, 1H), 6.92 (s, 1H), 6.38 (s, 1H), 6.05 (s, 1H), 4.37 (t, *J* = 6.4 Hz, 1H), 3.53 (d, *J* = 7.2 Hz, 1H), 3.34 (s, 1H), 2.94–2.74 (m, 5H), 2.43–

2.32 (m, 2H), 2.09 (dd, J = 15.8 and 9.3 Hz, 1H), 1.90 (dd, J = 9.9 and 4.9 Hz, 1H), 1.54–0.81 (m, 41H), 0.73 (d, J = 10.8 Hz, 1H), 0.53–0.37 (m, 2H); ³¹P NMR (161 MHz, CDCl₃) δ 11.1 (s); ¹³C NMR (101 MHz, CDCl₃) δ 174.7, 149.7, 149.6, 146.1, 146.0, 142.5, 140.6, 135.9, 135.4, 132.5, 131.6, 130.6, 130.2, 127.7, 127.4, 127.1, 126.7, 126.6, 125.6, 125.4, 124.7, 121.9, 120.1, 73.0, 64.1, 63.8, 63.3, 63.0, 60.3, 39.7, 34.9, 34.8, 33.8, 31.3, 31.2, 30.8, 30.5, 30.1, 27.4, 27.3. HRMS (ESI) Calcd for [C₅₄H₆₈IrNaO₂P, M + Na]⁺: 995.4478, Found: 995.4480.

2. Preparation and Analytical Data of Unsaturated Carboxylic Acids

The 3-alkyl-3-methylene-carboxylic acids were prepared according to literature procedure.² The acids **5a**², **5b**³ and **5k**² are known compounds. The other α , β -unsaturated carboxylic acids **7a** and **7b** are commercially available; **7c**,⁴ **7d**,⁵ **7e**⁶ and **7f**⁷ were prepared according to the reported procedures.

3-Methylenedecanoic acid (5c)

Yield: 80%, colorless oil. ¹H NMR (400 MHz, CDCl₃) δ 4.96 (s, 1H), *n*-Hept COOH 4.93 (s, 1H), 3.08 (s, 2H), 2.12 (t, *J* = 7.6 Hz, 2H), 1.48–1.41 (m, 2H), 1.32–1.28 (m, 8H), 0.88 (t, *J* = 6.4 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 177.7, 142.1, 114.0, 41.6, 35.9, 31.8, 29.2, 29.1, 27.4, 22.7, 14.1. HRMS (ESI) Calcd for [C₁₁H₁₉O₂, M – H]⁻: 183.1391, Found: 183.1390.

4-Methyl-3-methylenepentanoic acid (5d)

Yield: 76%, colorless oil. ¹H NMR (400 MHz, CDCl₃) δ 4.99 (s, 1H), 4.92 (s, ^{*i*}Pr COOH 1H), 3.10 (s, 2H), 2.39–2.32 (m, 1H), 1.06 (d, J = 6.8 Hz, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 178.5, 147.8, 112.0, 40.2, 33.8, 21.4. IR (KBr): v_{max} 3449, 2957, 2924, 2873, 1709, 1644, 1459, 1293, 1164, 901, 742, 699. HRMS (ESI) Calcd for [C₇H₁₁O₂, M – H]⁻: 127.0765, Found: 127.0766.

4-Ethyl-3-methylenehexanoic acid (5e)



Yield: 70%, colorless oil. ¹H NMR (400 MHz, CDCl₃) δ 5.05 (s, 1H), 4.96 (s, 1H), 2.97 (s, 2H), 1.96–1.89 (m, 1H), 1.46–1.30 (m, 4H), 0.83 (t, *J* = 7.6 Hz, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 178.4, 143.7, 114.5, 49.7, 38.8,

25.7, 11.7. IR (KBr): v_{max} 3498, 2963, 2930, 2875, 1711, 1643, 1457, 1293, 900. HRMS (ESI) Calcd for $[C_9H_{15}O_2, M - H]^-$: 155.1078, Found: 155.1076.

5-Methyl-3-methylenehexanoic acid (5f)

Me Me Me COOH Me Me COOH Me COCI ME C

115.6, 45.5, 41.4, 25.9, 22.4. HRMS (ESI) Calcd for $[C_8H_{13}O_2, M - H]^-$: 141.0921, Found: 141.0923.

3-Cyclohexylbut-3-enoic acid (5g)

СООН

Yield: 70%, colorless oil. ¹H NMR (400 MHz, CDCl₃) δ 4.97 (s, 1H), 4.92 (s, 1H), 3.10 (s, 2H), 1.98–1.92 (m, 1H), 1.83–1.75 (m, 4H), 1.70–1.66 (m, 1H), 1.33–1.07 (m, 5H); ¹³C NMR (101 MHz, CDCl₃) δ 178.6, 147.1, 112.4, 43.9, 40.6, 32.0, 26.6, 26.2. IR (KBr): v_{max} 3088,

2927, 2853, 1710, 1643, 1448, 1409, 1294, 1216, 891, 735, 620. HRMS (ESI) Calcd for $[C_{10}H_{15}O_2, M - H]^-$: 167.1078, Found: 167.1077.

3-Methylene-5-phenylpentanoic acid (5h)

Ph

Yield: 72%, colorless oil. ¹H NMR (400 MHz, CDCl₃) δ 7.30–7.26 (m, 2H), 7.20–7.17 (m, 3H), 5.01 (s, 1H), 4.99 (s, 1H), 3.12 (s, 2H), 2.79 (t, *J* = 7.6 Hz, 2H), 2.45 (t, *J* = 8.4 Hz, 2H); ¹³C NMR (101 MHz, CDCl₃) δ

178.0, 141.5, 141.2, 128.4, 128.3, 126.0, 114.8, 41.9, 37.5, 33.9. IR (KBr): v_{max} 3643, 3084, 3027, 2929, 2857, 1709, 1648, 1409, 1294, 1217, 902, 744, 698. HRMS (ESI) Calcd for [C₁₂H₁₃O₂, M – H]⁻: 189.0921, Found: 189.0923.

7-Methoxy-3-methyleneheptanoic acid (5i)

7-Methyl-3-methyleneoct-6-enoic acid (5j)

 $\label{eq:coord} \begin{array}{c} \mbox{Me} & \mbox{Yield: 62\%, colorless oil. 1H NMR (400 MHz, CDCl_3) δ} \\ \mbox{Me} & \mbox{COOH} & \mbox{5.12-5.07 (m, 1H), 4.98 (s, 1H), 4.96 (s, 1H), 3.09 (s, 2H),} \\ \mbox{2.15-2.14 (m, 4H), 1.68 (s, 3H), 1.61 (s, 3H); 13C NMR (101 MHz, CDCl_3) δ 178.1, 141.7, 132.1, 123.5, 114.3, 41.8, 35.8, 26.1, 25.7, 17.7. HRMS (ESI) Calcd for [C_{10}H_{15}O_2, M - H]^-: 167.1078, Found: 167.1080. \end{array}$

3. Asymmetric Hydrogenation and Analytical Data of Products

General procedure of hydrogention

$$R \xrightarrow{\text{COOH}} H_2 (3 \text{ atm}) \xrightarrow{1 \text{ mol } \% (S)-2d} \xrightarrow{\text{Me}} R \xrightarrow{\text{Me}} COOH$$

$$n^{\text{BuOH, 65 °C, 0.5 h}} \xrightarrow{\text{Me}} R^{\text{Me}}$$

A hydrogenation tube was charged with a stir bar, β -alkyl- β , γ -unsaturated acids **5** (0.5 mmol), catalyst (*S*)-**2d** (3.7 mg, 0.005 mmol), Cs₂CO₃ (82 mg, 0.25 mmol) in an argon-filled glovebox. Then 2 mL ^{*n*}BuOH was injected into the hydrogenation tube by a syringe with stirring. The hydrogenation tube was put into an autoclave. The argon in the autoclave was replaced with hydrogen for 3 times, and was finally charged with hydrogen to 3 atm. The reaction mixture was stirred at 65 °C for specified time before releasing the hydrogen.

After releasing hydrogen, the reaction mixture was added with 20 mg NaOH and concentrated under reduced pressure. The mixture was added 25 mL water and washed with Et₂O. The aqueous layer was acidified with conc. HCl, and extracted with Et₂O. The organic layer was dried with anhydrous Na₂SO₄. The conversion of substrate was determined by ¹H NMR analysis. The crude product was purified by a flash chromatography on silica gel column to give pure product **6**. The acid **6** (0.5 mmol) was reacted with aniline (50 μ L, 0.55 mmol) in the presence of *N*,*N*-4-dimethylaminopyridine (DMAP, 4 mg, 0.033 mmol) and dicyclohexylcarbodiimide (DCC, 110 mg, 0.53 mmol) in 2.0 mL THF for 30 min. The filtrate was concentrated under reduced pressure and the residue passed through a flash chromatography on Al₂O₃ column with PE/EA (4:1, v/v) as eluent to afford the corresponding amide. The ee value of amide was determined by HPLC or GC.

(*R*)-3-Methylheptanoic acid (6a)²

Yield: 99%, colorless oil. 93% ee (*R*), $[\alpha]_D^{25}$ +4.70 (*c* 0.4, CH₂Cl₂), HPLC condition for corresponding amide: Chiralpak AD-H column (25 cm × 0.46 cm ID), *n*-hexane/2-propanol = 85:15, 1.0 mL/min, 254 nm UV detector, t_R

= 5.43 min for (*S*)-enantiomer and $t_{\rm R}$ = 5.94 min for (*R*)-enantiomer. ¹H NMR (400 MHz, CDCl₃) δ 2.35 (dd, *J* = 15.2 and 6.0 Hz, 1H), 2.14 (dd, *J* = 14.8 and 8.0 Hz, 1H), 1.99–1.89 (m, 1H), 1.37–1.17 (m, 6H), 0.96 (d, *J* = 6.8 Hz, 3H), 0.89 (t, *J* = 6.8 Hz, 3H). IR (KBr): $v_{\rm max}$ 3450, 2960, 2925, 2861, 1708, 1413, 1292, 936.

(R)-3-Methylpentanoic acid (6b)⁸

Me Et COOH Et COOH Et COOH COOH

3-Methyldecanoic acid (6c)

Me *n*-Hept *n*-Hept Yield: 99%, colorless oil. 94% ee, $[\alpha]_D^{25}$ +7.62 (*c* 1.0, CHCl₃), HPLC condition for corresponding amide: Chiralpak AD-H column (25 cm × 0.46 cm ID), *n*-hexane/2-propanol = 90:10, 1.0 mL/min, 254 nm UV detector, t_R = 6.78 min for the minor isomer and t_R = 7.70 min for the major isomer. ¹H NMR (400 MHz, CDCl₃) δ 2.35 (dd, J = 14.8 and 5.6 Hz, 1H), 2.14 (dd, J = 14.8 and 8.0 Hz, 1H), 1.97–1.94 (m, 1H), 1.31–1.21 (m, 12H), 0.96 (d, J = 6.8 Hz, 3H), 0.88 (t, J = 6.4 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 180.1, 41.6, 36.7, 31.9, 30.2, 29.7, 29.3, 26.9, 22.7, 19.7, 14.1. HRMS (ESI) Calcd for [C₁₁H₂₁O₂, M – H]⁻: 185.1547, Found: 185.1545.

(S)-3,4-Dimethylpentanoic acid (6d)⁹

Me *i*Pr *COOH i*Pr *i*Pr *COOH i*Pr *i*Pr

4-Ethyl-3-methylhexanoic acid (6e)

major isomer. ¹H NMR (400 MHz, CDCl₃) δ 2.36–2.31 (m, 1H), 2.16–2.08 (m, 2H), 1.33–1.22 (m, 5H), 0.90–0.86 (m, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 180.4, 45.9, 39.1, 31.2, 23.0, 22.3, 15.9, 12.2, 12.1. IR (KBr): v_{max} 3788, 2964, 2930, 2875, 1708, 1412, 1296, 1203, 930, 745. HRMS (ESI) Calcd for [C₉H₁₇O₂, M – H]⁻: 157.1234, Found: 157.1223.

(*R*)-3,5-Dimethylhexanoic acid (6f)

Me Me Yield: 99%, colorless oil. 96% ee (R), $[\alpha]_D^{25}$ +12.8 (c 5.0, CHCl₃),¹⁰ HPLC condition for corresponding amide: Chiralpak AD-H column (25 cm × 0.46 cm ID), n-hexane/2-propanol = 95:5, 1.0 mL/min, 254 nm

UV detector, $t_{\rm R} = 15.93$ min for (*S*)-enantiomer and $t_{\rm R} = 16.61$ min for (*R*)-enantiomer. ¹H NMR (400 MHz, CDCl₃) δ 2.32 (dd, J = 14.4 and 5.2 Hz, 1H), 2.13–1.98 (m, 2H), 1.68–1.58 (m, 1H), 1.18–1.05 (m, 2H), 0.94 (d, J = 6.4 Hz, 3H), 0.87 (t, J = 6.8 Hz, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 180.0, 46.2, 42.2, 27.9, 25.2, 23.2, 22.1, 19.7. HRMS (ESI) Calcd for [C₈H₁₅O₂, M – H]⁻: 143.1078, Found: 143.1076.

3-Cyclohexylbutanoic acid (6g)¹¹



Yield: 99%, colorless oil. 98% ee, $[\alpha]_D^{25}$ +9.97 (*c* 1.0, CHCl₃), HPLC condition for corresponding amide: Chiralpak AD-H column (25 cm × 0.46 cm ID), *n*-hexane/2-propanol = 85:15, 1.0 mL/min, 254 nm UV detector, $t_R = 6.32$ min for the minor isomer and $t_R = 7.06$ min for the

major isomer. ¹H NMR (400 MHz, CDCl₃) δ 2.41 (dd, J = 14.8 and 4.8 Hz, 1H), 2.08 (dd, J = 14.8 and 9.6 Hz, 1H), 1.87–1.85 (m, 1H), 1.75–1.73 (m, 2H), 1.65–1.62 (m, 3H), 1.25–0.94 (m, 6H), 0.91 (d, J = 6.4 Hz, 3H). IR (KBr): v_{max} 3480, 2925, 2853, 1708, 1449, 1288, 938.

(*R*)-3-Methyl-5-phenylpentanoic acid (6h)¹²

Me Ph COOH Yield: 99%, colorless oil. 93% ee (R), $[\alpha]_D^{25}$ +67.6 (c 1.0, C_6H_6), HPLC condition for corresponding amide: Chiralpak AD-H column (25 cm × 0.46 cm ID), n-hexane/2-propanol = 85:15, 1.0 mL/min, 254 nm UV detector, t_R = 7.52 min for (S)-enantiomer and t_R = 8.35 min for (R)-enantiomer. ¹H NMR (400 MHz, CDCl₃) δ 7.31–7.26 (m, 2H), 7.21–7.19 (m, 3H), 2.73–2.57 (m, 2H), 2.43 (dd, J = 15.2 and 6.0 Hz, 1H), 2.23 (dd, J = 14.8 and 8.0 Hz, 1H), 2.09–2.00 (m, 1H), 1.76–1.67 (m, 1H), 1.60–1.51 (m, 1H), 1.06 (d, J = 6.8 Hz, 3H). IR (KBr): v_{max} 3479, 3028, 2961, 2926, 1707, 1454, 1299, 746, 698.

7-Methoxy-3-methylheptanoic acid (6i)

(*R*)-3,7-Dimethyloct-6-enoic acid (6j)¹³

Me Me Yield: 99%, colorless oil. 94% ee (*R*), $[\alpha]_D^{25}$ +9.31 (*c* 1.0, CHCl₃), HPLC condition for corresponding amide: Chiralpak AD-H column (25 cm × 0.46 cm ID), *n*-hexane/2-propanol = 90:10, 1.0 mL/min, 254 nm UV detector, t_R = 15.53 min for (*S*)-enantiomer and t_R = 16.73 min for (*R*)enantiomer. ¹H NMR (400 MHz, CDCl₃) δ 5.09 (t, *J* = 7.2 Hz, 1H), 2.36 (dd, *J* = 15.2 and 6.0 Hz, 1H), 2.15 (dd, *J* = 14.8 and 8.4 Hz, 1H), 2.07–1.95 (m, 3H), 1.68 (s, 3H), 1.60 (s, 3H), 1.41–1.34 (m, 1H), 1.29–1.20 (m, 1H), 0.98 (d, *J* = 6.4 Hz, 3H).

(S)-2-Methyl-3-phenylpropionic acid (8a)¹⁴

Ph Me Yield: 99%, colorless oil. 99.4% ee (*S*), $[\alpha]_D^{25}$ +31.2 (*c* 0.8, CHCl₃), HPLC condition for corresponding amide: Chiralpak AS column (25 cm × 0.46 cm ID), *n*-hexane/2-propanol = 95:5, 1.0 mL/min, 254 nm UV detector, t_R = 14.53 min for (*S*)-enantiomer and t_R = 17.72 min for (*R*)-enantiomer. ¹H NMR (400 MHz, CDCl₃) δ 7.30–7.17 (m, 5H), 3.08 (dd, *J* = 17.2 and 8.0 Hz, 1H), 2.80–2.63 (m, 2H), 1.17 (d, *J* = 9.2 Hz, 3H). IR (KBr): v_{max} 3664, 3062, 2972, 2927, 2855, 1707, 1496, 1236, 910, 742, 699.

(S)-2-Methylbutanoic acid (8b)¹⁵

Me COOH Me Ne Vield: 98%, colorless oil. 97% ee (*S*), $[\alpha]_D^{25}$ +20.7 (*c* 0.8, C₂H₅OH), HPLC condition for corresponding amide: Chiralpak AD-H column (25 cm × 0.46 cm ID), *n*-hexane/2-propanol = 95:5, 1.0 mL/min, 254 nm UV detector, t_R =

17.23 min for (S)-enantiomer and $t_R = 18.94$ min for (R)-enantiomer. ¹H NMR (400 MHz, CDCl₃)

δ 2.45–2.34 (m, 1H), 1.76–1.64 (m, 1H), 1.54–1.45 (m, 1H), 1.17 (d, J = 9.2 Hz, 3H), 0.94 (t, J = 10.0 Hz, 3H). IR (KBr): v_{max} 3670, 2962, 2928, 2872, 1708, 1550, 1459, 1024.

(R)-2-Phenylbutanoic acid (8c)¹⁶

Me COOH Ph Yield: 98%, white solid, mp: 40–42 °C. 94% ee (*R*), $[\alpha]_D^{25}$ –70.4 (*c* 1.0, CHCl₃), HPLC condition for corresponding amide: Chiralpak AD-H column (25 cm × 0.46 cm ID), *n*-hexane/2-propanol = 80:20, flow rate = 1.0 mL/min, 254 nm UV detector, t_R = 6.41 min for (*S*)-enantiomer and t_R = 7.67 min for (*R*)-enantiomer. ¹H NMR (400 MHz, CDCl₃) δ 7.33–7.22 (m, 5H), 3.45 (t, *J* = 8.0 Hz, 1H), 2.15–2.04 (m, 1H), 1.86– 1.75 (m, 1H), 0.90 (t, *J* = 7.6 Hz, 3H).

(S)-2-(Benzyloxy)-3-phenylpropionic acid (8d)⁵

Ph (OBn) Vield: 98%, white solid, mp: 59–60 °C. 98% ee (*S*), $[\alpha]_D^{20}$ –83.2 (*c* 2.2, C₂H₅OH), ¹⁷ HPLC condition for corresponding amide: Chiralpak AD-H column (25 cm × 0.46 cm ID), *n*-hexane/2-propanol = 80:20, flow rate = 1.0 mL/min, 254 nm UV detector, t_R = 10.19 min for (*S*)-enantiomer and t_R = 10.91 min for (*R*)-enantiomer. ¹H NMR (400 MHz, CDCl₃) δ 7.33–7.26 (m, 8H), 7.16–7.14 (m, 2H), 4.67 (d, *J* = 15.6 Hz, 1H), 4.40 (d, *J* = 15.6 Hz, 1H), 4.18 (dd, *J* = 11.2 and 5.6 Hz, 1H), 3.20–3.01 (m, 2H).

(*R*)-2-(4-Isobutylphenyl)propionic acid [(*R*)-ibuprofen] (8e)¹⁸



Yield: 99%, white solid, mp: 53–54 °C. 96% ee (*R*), $[a]_D^{25}$ –51.4 (*c* 2.0, C₂H₅OH), GC condition for corresponding methyl ester: Coating CP Chirasil-DEX CB, CP7502, df = 0.25 µm, 0.25 mm i.d. x 25 m, carrier gas: N₂ (1.3 mL/min), inject temperature: 230

°C, initial temperature: 120 °C, hold 60 min then temperature programmed, programming rate: 1.0 °C /min, final temperature: 180 °C. $t_{\rm R}$ = 38.26 min for (*S*)-enantiomer and $t_{\rm R}$ = 38.88 min for (*R*)-enantiomer. ¹H NMR (400 MHz, CDCl₃) δ 7.22 (d, *J* = 8.0 Hz, 2H), 7.11 (d, *J* = 8.0 Hz, 2H), 3.71 (q, *J* = 7.2 Hz, 1H), 2.45 (d, *J* = 6.8 Hz, 2H), 1.90–1.80 (m, 1H), 1.50 (d, *J* = 7.2 Hz, 3H), 0.90 (d, *J* = 6.8 Hz, 6H).

(*R*)-2-Methylheptanoic acid (8f)¹⁹

Me *n*-Pent COOH Wield: 97%, colorless oil. 99.3% ee (*R*), $[\alpha]_D^{25}$ –16.8 (*c* 0.6, C₂H₅OH), HPLC condition for corresponding amide: Chiralpak AD-H column (25 cm × 0.46 cm ID), *n*-hexane/2-propanol = 95:5, flow rate = 1.0 mL/min, 254 nm UV detector, $t_R = 11.46$ min for (*S*)-enantiomer and $t_R = 12.43$ min for (*R*)-enantiomer. ¹H NMR (400 MHz, CDCl₃) δ 2.49–2.41 (m, 1H), 1.72–1.63 (m, 1H), 1.47–1.38 (m, 1H), 1.36–1.24 (m, 6H), 1.17 (d, J = 7.2 Hz, 3H), 0.88 (t, J = 6.4 Hz, 3H).

4. Total Synthesis of (S)-14-Methyloctadec-1-ene



(S)-3-Methylheptan-1-ol (9)²⁰

Me ⁿBu OH A solution of acid **6a** (72 mg, 0.50 mmol) in 1 mL of anhydrous THF was added to a stirred suspension of LiAlH₄ (38 mg, 1.0 mmol) in 1 mL of anhydrous THF at 0 °C. The reaction mixture was warmed to room temperature, and stirred for 2 h. After cooling to 0 °C, 1 mL of water and 1 mL of aq. NaOH (10% solution) were added with stirring. The organic layer was separated, and the aqueous layer was extracted with Et₂O. The combined organic layer was dried with Na₂SO₄ and concentrated under reduced pressure. The residue was purified by a flash column chromatography on silica gel with PE/EA (4:1) to offer **9** (61 mg, 94%) as a colorless oil. $[\alpha]_D^{25}$ –2.73 (*c* 3.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 3.69–3.63 (m, 2H), 1.66–1.54 (m, 2H), 1.37–1.24 (m, 7H), 0.89 (t, *J* = 6.8 Hz, 3H), 0.86 (t, *J* = 6.6 Hz, 3H).

(S)-3-Methylheptyl 4-methylbenzenesulfonate (10)²⁰

Tosyl chloride (70 mg, 0.38 mmol) was added to a solution of alcohol **9** (50 mg, 0.35 mmol) in 1 mL of anhydrous pyridine at 0 $^{\circ}$ C and the mixture was stirred for 16 h at the same temperature. The mixture was added 20 mL of

 Et_2O and washed successively with brine, saturated solution of $CuSO_4$ and $NaHCO_3$, and brine. The organic phase was dried with Na_2SO_4 , and concentrated to give tosylate **10**. The product was used in the next step without further purification.

(S)-14-Methyloctadec-1-ene²⁰

OTs

Me

ⁿBu⁻

Me ⁿBu ⁿB

5. X-ray Diffraction Analysis of (S)-2d

The fine yellow crystals of (*S*)-2d suitable for the X-ray diffraction analyses grow slowly on the interface of a solution of (*S*)-2d (30 mg) in dichloromethane (0.5 mL) and *n*-hexane (1.5 mL).



Table S1. Crystal data and structure refinement for (S)-2d

Empirical formula	C39 H36 Cl2 Ir O2 P
Moiety formula	$C_{39}H_{36}Cl_2IrO_2P$
Formula weight	830.75
Temperature	113(2) K
Wavelength	0.71073 Å
Crystal system	Monoclinic
Space group	C2
Unit cell dimensions	a = 19.385(3) Å alpha = 90° b = 16.220(3) Å beta = 107.035(18)° c = 10.9215(15) Å gamma = 90°
Volume	3283.2(10) Å ³
Z	4
Calculated density	1.681 Mg/m ³
Absorption coefficient	4.313 mm ⁻¹

F(000)	1648
Crystal size	0.20 x 0.18 x 0.12 mm
Theta range for data collection	3.18 to 25.01°
Limiting indices	-23<=h<=23, -19<=k<=18, -12<=l<=12
Reflections collected / unique	17815 / 5559 [R(int) = 0.0734]
Completeness to theta $= 25.01$	99.7 %
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	0.6256 and 0.4792
Refinement method	Full-matrix least-squares on F ²
Data / restraints / parameters	5559 / 71 / 434
Goodness-of-fit on F ²	1.041
Final R indices [I>2sigma(I)]	$R_1 = 0.0317, wR_2 = 0.0748$
R indices (all data)	$R_1 = 0.0321, wR_2 = 0.0750$
Absolute structure parameter	0.010(7)
Largest diff. peak and hole	2.781 and -1.354 e.Å ⁻³

6. NMR Spectra of New Compounds

(S)-2a









3-Methylenedecanoic acid (5c)



4-Methyl-3-methylenepentanoic acid (5d)



4-Ethyl-3-methylenehexanoic acid (5e)



5-Methyl-3-methylenehexanoic acid (5f)



3-Cyclohexylbut-3-enoic acid (5g)



3-Methylene-5-phenylpentanoic acid (5h)



7-Methoxy-3-methyleneheptanoic acid (5i)



7-Methyl-3-methyleneoct-6-enoic acid (5j)



3-Methyldecanoic acid (6c)



4-Ethyl-3-methylhexanoic acid (6e)



(R)-3,5-Dimethylhexanoic acid (6f)



7-Methoxy-3-methylheptanoic acid (6i)



7. HPLC or GC Charts of Hydrogenation Product Derivatives

(R)-3-Methyl-N-phenylheptanamide (6a)



(*R*)-3-Methyl-N-phenylpentanamide (6b)





Peak	RetTime	Туре	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	90
1	6.101	BV	0.1160	551.33026	73.09827	6.8041
2	6.461	VB	0.1252	7551.61914	944.85028	93.1959

3-Methyl-N-phenyldecanamide (6c)



(S)-3,4-Dimethyl-N-phenylpentanamide (6d)



S32





----|-----|-----|------|------| 1 5.426 BB 0.1094 162.20784 22.68942 0.9221 2 5.943 BB 0.1317 1.74287e4 2039.17822 99.0779

(R)-3,5-Dimethyl-N-phenylhexanamide (6f)



3-Cyclohexyl-N-phenylbutanamide (6g)



S35

(R)-3-Methyl-N,5-diphenylpentanamide (6h)



7-Methoxy-3-methyl-N-phenylheptanamide (6i)



1 23.019 BV 0.4654 1639.00330 54.72998 3.5834 2 24.184 VB 0.5454 4.40996e4 1256.79285 96.4166

(R)-3,7-Dimethyl-N-phenyloct-6-enamide (6j)



(S)-2-Methyl-N,3-diphenylpropanamide (8a)



S39

(S)-2-Methyl-N-phenylbutanamide (8b)



S40

2 18.941 VB 0.3908 503.29144 19.63482 1.3289





(S)-2-(Benzyloxy)-N,3-diphenylpropanamide (8d)



(*R*)-Methyl 2-(4-isobutylphenyl)propanoate (8e)



1	38.260	BV	0.2450	32.48083	1.65475	1.79662
2	38.884	VB	0.6741	1775.40771	32.05692	98.20338

(R)-2-Methyl-N-phenylheptanamide (8f)



8. Reference:

- 1 J. L. Herde, J. C. Lambert and C. V. Senoff, Inorg. Synth., 1974, 15, 18.
- 2 X. Sun, L. Zhou, C.-J. Wang and X. Zhang, Angew. Chem. Int. Ed., 2007, 46, 2623.
- 3 S. G. Alcock, J. E. Baldwin, R. Bohlmann, L. M. Harwood and J. I. Seeman, *J. Org. Chem.*, 1985, **50**, 3526.
- 4 Y. Zhang, Z.-B. Han, F.-Y. Li, K.-L. Ding and A. Zhang, *Chem. Commun.*, 2010, **46**, 156.
- 5 S. Li, S.-F. Zhu, J.-H. Xie, S. Song, C.-M. Zhang and Q.-L. Zhou, J. Am. Chem. Soc., 2010, **132**, 1172.
- 6 R. R. Kurtz and D. J. Houser, J. Org. Chem., 1981, 46, 202.
- 7 D. V. Kummer, W. J. Chain, M. R. Morales, O. Quiroga and A. G. Myers, *J. Am. Chem. Soc.*, 2008, **130**, 13231.
- 8 B.-F. Li, R. M. Hughes, J. Le, K. McGee, D. J. Gallagher, R. S. Gross, D. Provencal, J. P. Reddy, P. Wang, L. Zegelman, Y. Zhao and S. E. Zook, *Org. Process Res. Dev.*, 2009, **13**, 463.
- 9 E. Reyes, J. L. Vicario, L. Carrillo, D. Badia, U. Uria and A. Iza, *J. Org. Chem.*, 2006, **71**, 7763.
- 10 K. C. Rice, J. Org. Chem., 1982, 47, 3617.
- 11 J. M. Garcia, A. Gonzalez, B. G. Kardak, J. M. Odriozola, M. Oiaarbide, J. Razkin and C. Palomo, *Chem. Eur. J.*, 2008, **14**, 8768.
- 12 S. Sugiyama and T. Satoh, Tetrahedron: Asymmetry, 2005, 16, 665.
- 13 P. Heretsch, S. Rabe and A. Giannis, Org. Lett., 2009, 11, 5410.
- 14 S. G. Davies, D. J. Dixon, G. J.-M. Doisneau, J. C. Prodger and H. J. Sanganee, *Tetrahedron: Asymmetry*, 2002, **13**, 647.
- 15 S. Li, S.-F. Zhu, C.-M. Zhang, S. Song and Q.-L. Zhou, J. Am. Chem. Soc., 2010, 132, 1172.
- 16 I. Shiina, K. Nakata, K.Ono, Y. Onda and M. Itagaki, J. Am. Chem. Soc., 2010, 132, 11629.
- 17 V. Gopalsamuthiram, R. Huang and W. D. Wulff, Chem. Commun., 2010, 46, 8213.
- 18 T. Fujiwara, M. Sasaki, K. Omata, C. Kabuto, K. Kabuto and Y. Takeuchi, *Tetrahedron: Asymmetry*, 2004, **15**, 555.
- 19 N. Harrington-Frost, H. Leuser, M. I. Calaza, F. F. Kneisel and P. Knochel, *Org. Lett.*, 2003, **5**, 2111.
- 20 R. Y. Kharisov, E. R. Latypova, R. F. Talipov, R. R. Muslukhov, G. Y. Ishmuratov and G. A. Tolstikov, *Russ. Chem. Bull. Int. Ed.*, 2003, **52**, 2267.