

Supporting Information

T-shaped monopyridazinotetrathiafulvalene-amino acid diad based chiral organogels with aggregation-induced fluorescence emission

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1. Instrumentation

Gelation study

A measured amount of gelator **1** and a measured volume of the solvent were placed in a sealed test tube and heated to obtain a clear solution. And then, the system was left at room temperature. The transition temperatures (T_{gel}) were determined by ball-drop method. An inverted gel was immersed in a water bath initially at or below room temperature, and then was heated (about 2 °C min⁻¹) slowly up to the point at which the gel fell due to the force of gravity, i.e. the T_{gel} .

NMR experiments

All solution state NMR studies were carried out on Bruker AV-300 Spectrometer (300 MHz for ¹H and 75 MHz for ¹³C) and chemical shifts were referenced relative to tetramethylsilane ($\delta_{\text{H}}/\delta_{\text{C}}=0$).

FT-IR spectroscopy

IR spectra were recorded on a Shimadzu FT-IR Prestige-21 instrument with the KBr disk technique.

MALDI-TOF-MS spectrometry

Mass spectra were performed on a Shimadzu Axima CFRTM Plus using a 1,8,9-anthracenetriol (DITH) and β -phenylacrylic acid (CHCA) matrix.

Cyclic voltammetry

Cyclic voltammetry was performed on a CHI660D instrument in benzonitrile (10⁻³ M) with 0.1 M Bu₄NPF₆ as the supporting electrolyte and at a scan rate of 100mVs⁻¹. Counter and working electrodes were made of Pt and glass carbon, respectively, and Ag/AgCl was used as the reference electrode. For xerogel measurements, a small amount of the gel was carefully put on the glass carbon electrode, which was left in air for 24 h. This modified glass carbon electrode (as working electrode), together with Pt as the counter electrode and Ag/AgCl as the reference electrode, was put into the benzonitrile solution containing 0.1 M Bu₄NPF₆, and the cyclic voltammograms were recorded at a scanning rate of 100 mV s⁻¹.

UV-Vis spectroscopy

UV-vis spectra were recorded on a Hitachi U-3010 spectrophotometer.

Fluorescence spectroscopy

Fluorescence spectra were recorded on a Shimadzu RF-5301PC fluorescence spectrophotometer.

Circular dichroism (CD) spectroscopy

CD spectra were obtained on Chirascan spectrometer using a 1 mm path-length cell.

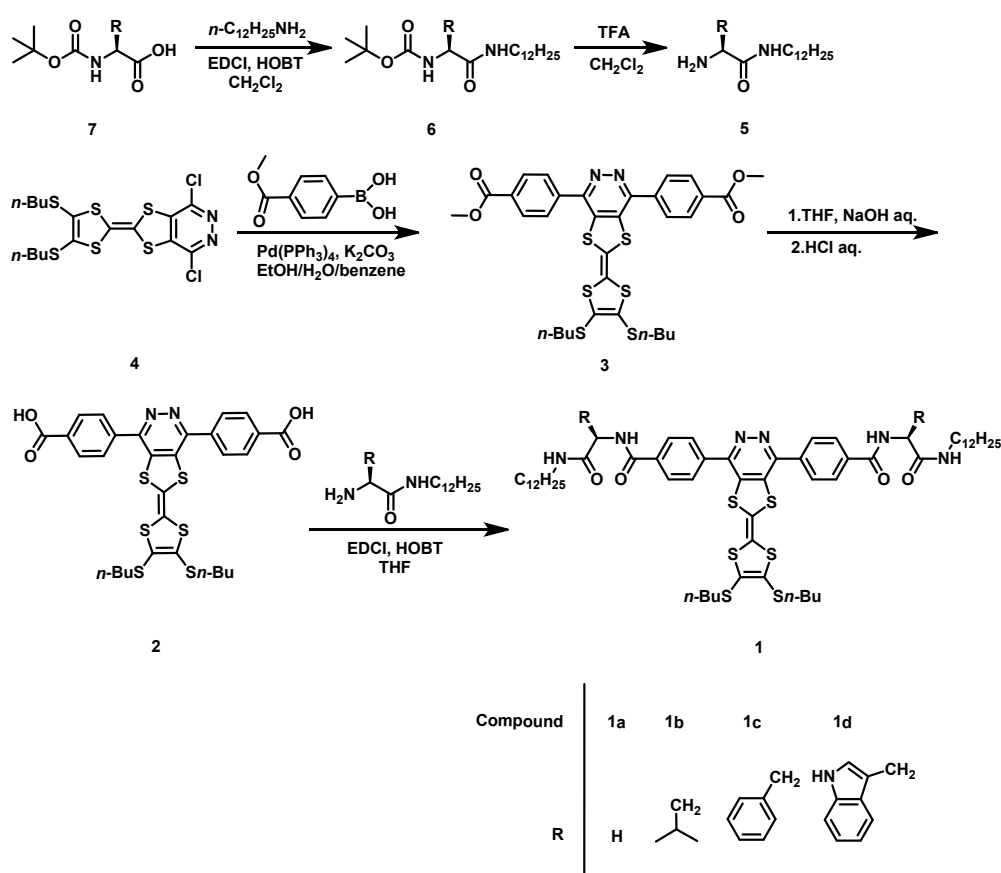
Atomic force microscopy (AFM)

For AFM experiments, 10 μ L of sample solution (diluted gels) was drop-casted onto a freshly cleaved mica surface. Each sample was air-dried 48 h in a dust-free environment prior to AFM imaging. The images were obtained by scanning the mica surfaces in air under ambient conditions using Agilent-5500 in tapping mode.

Small-angle X-ray diffracting

Small-angle X-ray scattering (SAXS) measurements were carried out at 298 K on a beamline 1W2A synchrotron radiation X-ray small angle system at Beijing Synchrotron Radiation Facility ($\lambda = 1.54 \text{ \AA}$).

2. Synthesis and characterization of organic compounds



Scheme S1. Synthetic routes of compounds 1a-d.

All solvents were purified by the standard procedures before use. *p*-(Methoxycarbonyl) phenylboronic acid, Pd(PPh₃)₄, 1-(3-(dimethyl-amino)propyl)-3-ethyl-carbodiimide hydrochloride (EDCI) and 1H-benzo[d][1,2,3]triazol-1-ol (HOBT) were purchased from Adamas, and were used as received without further purification. The Boc protection compounds were synthesized according to the traditional methods, and compounds 4, 6a-c were synthesized according to the literature procedure.^[1]

Typical procedure for compound 6d

The mixture of **7d** (4.0 g, 0.01314 mol), EDCI (2.7715 g, 0.01446 mol), HOBT (1.9535 g, 0.01446 mol) and *n*-dodecylamine (2.4361 g, 0.01314 mol) was suspended in CH₂Cl₂ (45 mL). The reaction mixture was stirred at room temperature for one day. The solution was successively washed with aqueous HCl (1 M), aqueous NaOH (1 M) and brine. After drying and filtering, the solvent was eliminated and the residue was purified by column chromatography on silica gel using CH₂Cl₂ : ethyl acetate at 20 : 1 to give a white solid (4.4 g, 71%). m.p.: 70-71 °C. ¹H NMR (300 MHz, CDCl₃) δ: 0.88 (t, *J* = 6Hz, 3H, -CH₂), 1.287-1.049 (m, 20H, -CH₂), 1.43 (br, 9H, -CH₃), 3.344-3.034 (m, 4H, -CH₂), 4.437- 4.319 (m, 1H, -CH), 5.19 (br, 1H, -NH), 5.61 (br, 1H, -NH), 7.06 (s, 1H, *Pγ*-NH), 7.165-7.101 (m, 1H, *Ar*), 7.233-7.17 (m, 1H, *Ar*), 7.36 (d, *J* = 9Hz, 1H, *Ar*), 7.67 (d, *J* = 9Hz, 1H, *Ar*), 8.10 (s, 1H, -NH); ¹³C NMR (75 MHz, CDCl₃) δ: 166.39, 151.48, 140.79, 140.50, 131.72, 130.23, 128.10, 127.83, 117.09, 103.82, 52.57, 36.09, 31.71, 21.61, 13.59; MALDI-TOF MS *m/z* calcd for C₂₈H₄₅N₃O₃: 471.35. Found: 471.9 ([M]⁺, 100)

Cleavage of the N-Boc protecting group and get compounds 5

Trifluoroacetic acid (TFA, 10 mL) was added to a solution of the corresponding N-Boc-protected compound (ca. 2 g) in dichloromethane (15 mL). The solution was stirred at room temperature for 2 h and evaporated. The residue obtained was repeatedly dissolved in dichloromethane and the solvent evaporated to yield the crude trifluoroacetate salt, which was redissolved in dichloromethane (60 mL) and washed with saturated aqueous NaHCO₃ (3 x 40 mL) and brine to neutrality. After drying and filtering, evaporation of the solvent afforded the corresponding amine which was used in the next step without further purification.

Typical procedure for compound 3

A mixture of **4** (462.9 mg, 0.9228 mmol), *p*-(methoxycarbonyl)phenyl-boronic acid (664.3 mg, 3.6912 mmol), Pd(PPh₃)₄ (511.9 mg, 0.4429 mmol) and K₂CO₃ (918.3 mg, 6.442 mmol) in a mixture of EtOH-H₂O-C₆H₆ at 1 : 1 : 2 (40 mL) was stirred under N₂ at 90 °C for 12 h. After

cooling, the resulting mixture was dissolved in CH_2Cl_2 , washed with water and dried over anhydrous Na_2SO_4 . After evaporation of the solvent, the crude product was purified by column chromatography on silica gel using CH_2Cl_2 : ethyl acetate at 30 : 1 to give a yellow solid (429.5 mg, 66.4%). m.p.: 176-177°C. ^1H NMR (300 MHz, CDCl_3) δ : 0.93 (t, 6H, J = 9Hz, $-\text{CH}_3$), 1.485-1.411 (m, 4H, $-\text{CH}_2$), 1.663-1.575 (m, 4H, $-\text{CH}_2$), 2.83 (t, J = 9Hz, 4H, $-\text{CH}_2$), 4.00 (s, 6H, $-\text{CH}_3$), 8.01(d, J = 9Hz, 4H, *Ar*), 8.27 (d, J = 9Hz, 4H, *Ar*); ^{13}C NMR (75 MHz, CDCl_3) δ : 166.39, 151.48, 140.79, 140.50, 131.72, 130.23, 128.10, 127.83, 117.09, 103.82, 52.57, 36.09, 31.71, 21.61, 13.59; MALDI-TOF MS m/z calcd for $\text{C}_{32}\text{H}_{32}\text{N}_2\text{O}_4\text{S}_6$: 700.07. Found: 700.08. ($[\text{M}]^+$, 100).

Typical procedure for compound 2

A mixture of **3** (278.7 mg, 0.3976 mmol) and NaOH (4 M, 20 mL) aqueous solution in THF (30 mL) was stirred at 50 °C overnight. After cooling and evaporation of the solvent, the pH was adjusted to 2-3. Thereafter, the mixture was filtered and washed with water. The filter cake was dried in vacuum to obtain a brownish yellow solid (215.5 mg, 80.5%). m.p.: 233-234°C. MALDI-TOF MS m/z calcd for $\text{C}_{30}\text{H}_{28}\text{N}_2\text{O}_4\text{S}_6$: 672.04. Found: 672.0. ($[\text{M}]^+$, 100).

Typical procedure for compounds 1

The mixture of **2** (0.06 mmol), EDCI (0.13 mmol), HOBT (0.13 mmol) and the corresponding compounds **5** (0.12 mmol) was suspended in THF (10 mL). The reaction mixture was stirred at room temperature for one day. The solvent was eliminated and the residue was taken up in CHCl_3 . The solution was successively washed with aqueous HCl (1 M), aqueous NaOH (1 M) and brine. After drying and filtering, the solvent was eliminated and the residue was purified by column chromatography on silica gel using CH_2Cl_2 : methanol at 30 : 1 to give a orange-yellow solid.

Compound 1a: Yield 72.5%, m.p.: 189-190°C. ^1H NMR (300 MHz, CDCl_3) δ : 0.955-0.818 (m, 12H, $-\text{CH}_3$), 1.464-1.241 (m, 40H, $-\text{CH}_2$), 1.646-1.516 (m, 8H, $-\text{CH}_2$), 1.82 (br, 2H, $-\text{NH}$), 2.987-2.662 (m, 4H, $-\text{CH}_2$), 3.31 (q, J = 6 Hz, 4H, $-\text{CH}_2$), 4.267-4.120 (m, 4H, $-\text{CH}_2$), 6.81 (br, 1H, $-\text{NH}$), 7.79 (br, 1H, $-\text{NH}$), 7.97 (d, J = 9Hz, 4H, *Ar*), 7.87 (d, J = 9Hz, 4H, *Ar*); ^{13}C NMR (75 MHz, CDCl_3) δ : 169.05, 167.37, 151.16, 140.68, 139.43, 135.06, 128.32, 127.93, 127.72, 103.65, 44.13, 39.85, 36.04, 31.93, 31.71, 29.68, 29.66, 29.63, 29.59, 29.49, 29.37, 29.33, 26.97, 22.70, 21.63, 14.14, 13.61; MALDI-TOF MS m/z calcd for $\text{C}_{58}\text{H}_{84}\text{N}_6\text{O}_4\text{S}_6$: 1120.49. Found: 1121.5. ($[\text{M}+\text{H}]^+$, 100).

Compound 1b: Yield 68.2%, m.p.: 231-232°C. ^1H NMR (300 MHz, CDCl_3) δ : 0.999-0.814 (m, 12H, $-\text{CH}_3$), 1.177-1.009 (m, 12H, $-\text{CH}_3$), 1.496-1.246 (m, 38H, $-\text{CH}_2$), 1.688-1.529 (m, 8H, $-\text{CH}_2$), 1.987-1.700 (m, 8H, $-\text{CH}_2$), 2.81(t, J = 9Hz, 4H, $-\text{CH}_2$), 3.22-3.070 (m, 2H, $-\text{CH}_2$), 3.501-3.356

(m, 2H, $-CH_2$), 5.058-4.889 (m, 2H, $-CH$), 7.3 (br, 1H, $-NH$), 7.45 (br, 2H, $-NH$), 7.70 (d, $J = 9\text{ Hz}$, 4H, *Ar*), 7.82 (d, $J = 9\text{ Hz}$, 4H, *Ar*), 7.97 (br, 1H, $-NH$); ^{13}C NMR (75 MHz, CDCl_3) δ : 172.11, 167.22, 150.90, 135.67, 128.25, 128.20, 128.05, 127.96, 127.56, 52.25, 41.04, 39.79, 39.72, 35.98, 31.94, 31.76, 31.72, 29.69, 29.51, 29.41, 27.11, 25.09, 22.85, 22.71, 21.68, 14.14, 13.63; MALDI-TOF MS m/z calcd for $\text{C}_{66}\text{H}_{100}\text{N}_6\text{O}_4\text{S}_6$: 1232.61. Found: 1233.9. ($[\text{M}+\text{H}]^+$, 100).

Compound 1c: Yield 67.3%, m.p.: 226-227°C. ^1H NMR (300 MHz, CDCl_3) δ : 0.96-0.827 (m, 12H, $-CH_3$), 1.459-1.243 (m, 44H, $-CH_2$), 1.651-1.557 (m, 4H, $-CH_2$), 1.77 (br, 1H, $-NH$), 2.906-2.710 (m, 4H, $-CH_2$), 3.363-3.021 (m, 8H, $-CH_2$), 5.236-5.100 (m, 2H, $-CH$), 7.00 (br, 2H, $-NH$), 7.4-7.287 (m, 10H, *Ar*), 7.71 (d, $J = 9\text{ Hz}$, 4H, *Ar*), 7.79 (d, $J = 9\text{ Hz}$, 4H, *Ar*), 7.88 (br, 1H, $-NH$); ^{13}C NMR (75 MHz, CDCl_3) δ : 175.47, 170.87, 166.92, 137.06, 135.52, 129.52, 128.60, 128.53, 128.44, 128.28, 128.25, 128.19, 128.03, 127.94, 127.67, 126.88, 55.32, 39.72, 38.85, 38.77, 36.02, 32.00, 31.95, 31.77, 31.72, 29.73, 29.70, 29.63, 29.47, 29.40, 29.34, 27.06, 26.98, 22.72, 21.66, 14.15, 13.64; MALDI-TOF MS m/z calcd for $\text{C}_{72}\text{H}_{96}\text{N}_6\text{O}_4\text{S}_6$: 1300.58. Found: 1301.6. ($[\text{M}+\text{H}]^+$, 100).

Compound 1d: Yield 61.4%, m.p.: 224-225°C. ^1H NMR (300 MHz, CDCl_3) δ : 0.915-0.742 (m, 12H, $-CH_3$), 1.4-1.175 (m, 44H, $-CH_2$), 1.537-1.433 (m, 4H, $-CH_2$), 3.272-3.023 (m, 8H, $-CH_2$), 4.792-4.673 (m, 2H, $-CH$), 7.102-6.952 (m, 4H, *Ar*), 7.24 (s, 2H, *Py-NH*), 7.315 (d, $J = 9\text{ Hz}$, 2H, *Ar*), 7.695 (d, $J = 9\text{ Hz}$, 2H, *Ar*), 7.89 (d, $J = 9\text{ Hz}$, 4H, *Ar*), 8.05 (d, $J = 6\text{ Hz}$, 4H, *Ar*), 8.73 (d, $J = 9\text{ Hz}$, 2H, $-NH$), 10.80 (br, 1H, $-NH$); ^{13}C NMR (75 MHz, DMSO) δ : 171.78, 165.88, 151.88, 140.27, 138.94, 136.53, 136.28, 128.73, 128.08, 127.74, 127.46, 124.07, 121.31, 118.98, 118.63, 111.77, 110.99, 54.92, 35.61, 34.84, 31.77, 31.67, 30.87, 29.59, 29.52, 29.48, 29.23, 26.74, 22.57, 21.27, 14.41, 13.85; MALDI-TOF MS m/z calcd for $\text{C}_{76}\text{H}_{98}\text{N}_8\text{O}_4\text{S}_6$: 1378.60. Found: 1377.3 ($[\text{M}+\text{H}]^+$, 100).

Reference:

- (a) N. Zheng, H. Li, G. Sun, K. Zhong, B. Yin, *Org. Biomol. Chem.* **2013**, *11*, 5100-5108; (b) S. Debnath, A. Shome, S. Dutta, P. K. Das, *Chem. Eur. J.* **2008**, *14*, 6870-6881; (c) Y. Dai, X. Zhao, X. Su, G. Li, A. Zhang, *Macromol. Rapid. Commun.* **2014**, *35*, 1326-1331; (d) S. Basak, N. Nandi, A. Baral, A. Banerjee, *Chem. Commun.* **2015**, *51*, 780-783.

3. Additional data

Table S1 Gelation properties of **1a-d** in various solvents.

Solvents	1a	1b	1c	1d
CH ₂ Cl ₂	P	S	TG(1.7)	IS
CHCl ₃	S	S	S	IS
THF	S	S	TG(3.8)	IS
1,4-dioxane	OG ^a (6.0 ^b)	P	OG(2.0)	OG(6.0)
CCl ₄	TG (4.2)	S	TG (5.0)	sS
Ethyl acetate	IS	IS	IS	IS
Benzene	TG(10.0)	S	TG(10)	sS
Toluene	TG(6.0)	S	TG (5)	sS
Xylene	TG(5.0)	S	TG(3.7)	sS
Phemethylol	S	S	S	S
<i>n</i> -Hexane	IS	IS	PG	IS
Cyclohexane	IS	IS	PG	IS
PE	IS	IS	IS	IS
Acetone	IS	IS	IS	IS
Acetonitrile	IS	IS	IS	IS
Aether	IS	IS	IS	IS
Methanol	IS	IS	IS	IS
Ethanol	IS	IS	IS	IS
DMF	SG(5.0)	SG(5.0)	SG(5.0)	S
DMSO	OG(1.6)	S	OG(1.8)	TG(5.0)
Glacial acetic acid	OG (4.0)	OG(4.0)	OG (4.0)	OG(4.0)

^aOG = Opaque gel; TG = Transparent gel; P = Precipitation; S = Soluble; IS = Insoluble; SG = Sonication gel

^bCGC = the critical gelation concentrations (mg/mL) at room temperature.

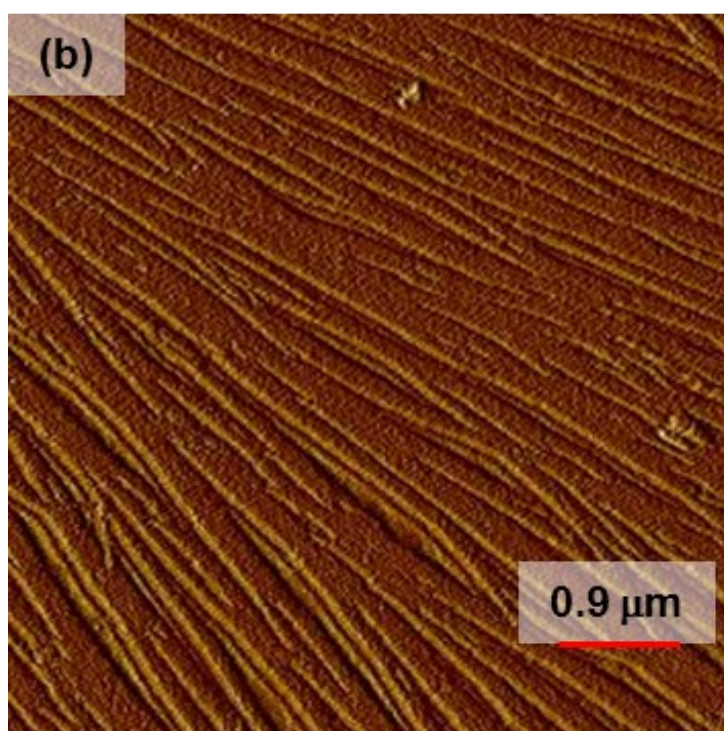
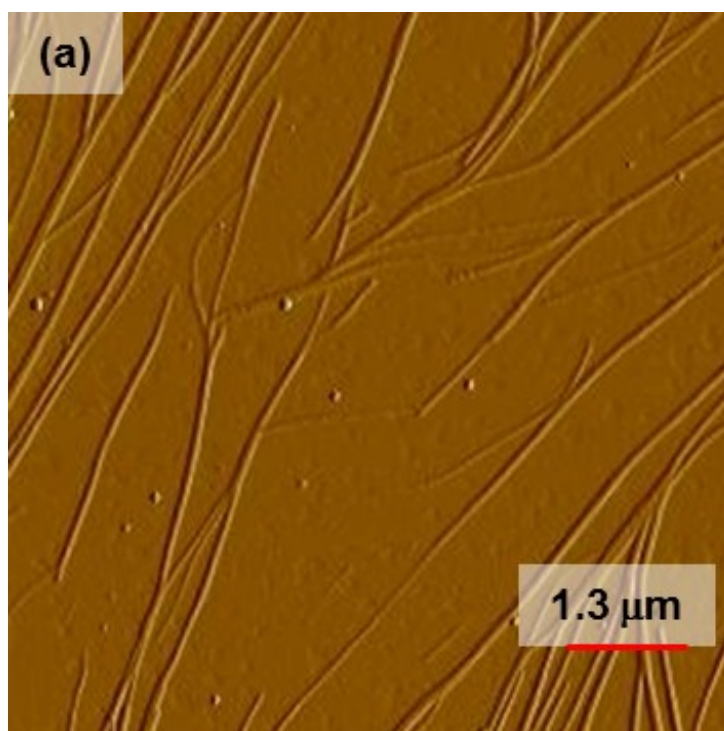


Figure S1. AFM images of organogels of **1c** and **1d** in DMSO (a, b).

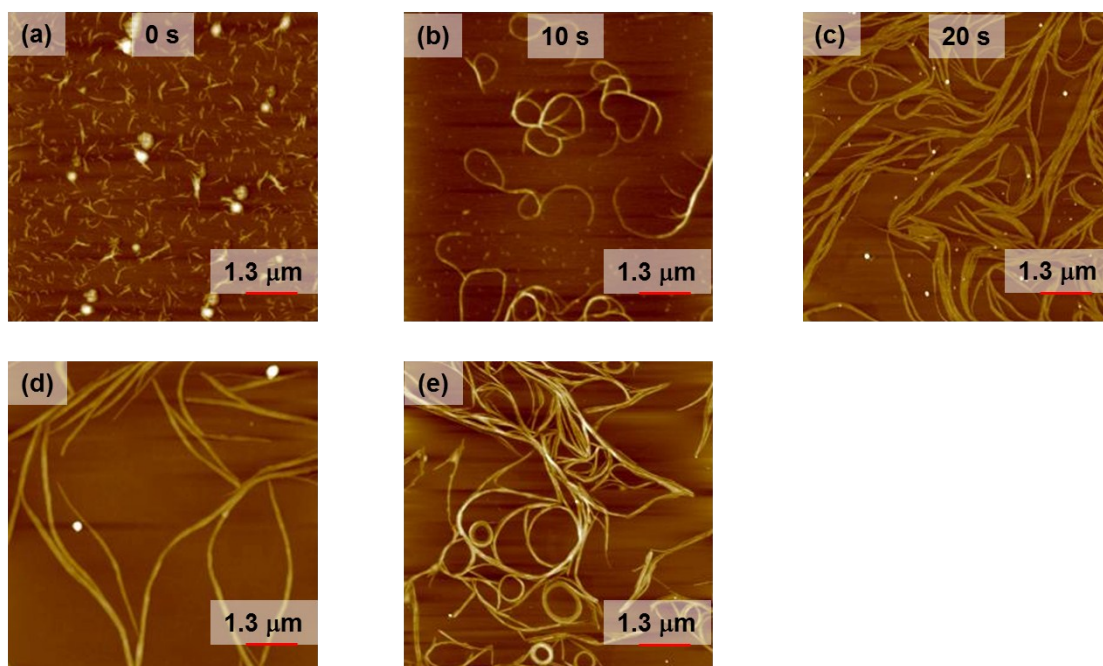


Figure S2. AFM observations of the conversion process of the gelator **1b** in DMF by sonication for different times (a-c), and AFM images of sonication organogels of **1a** and **1c** in DMF.

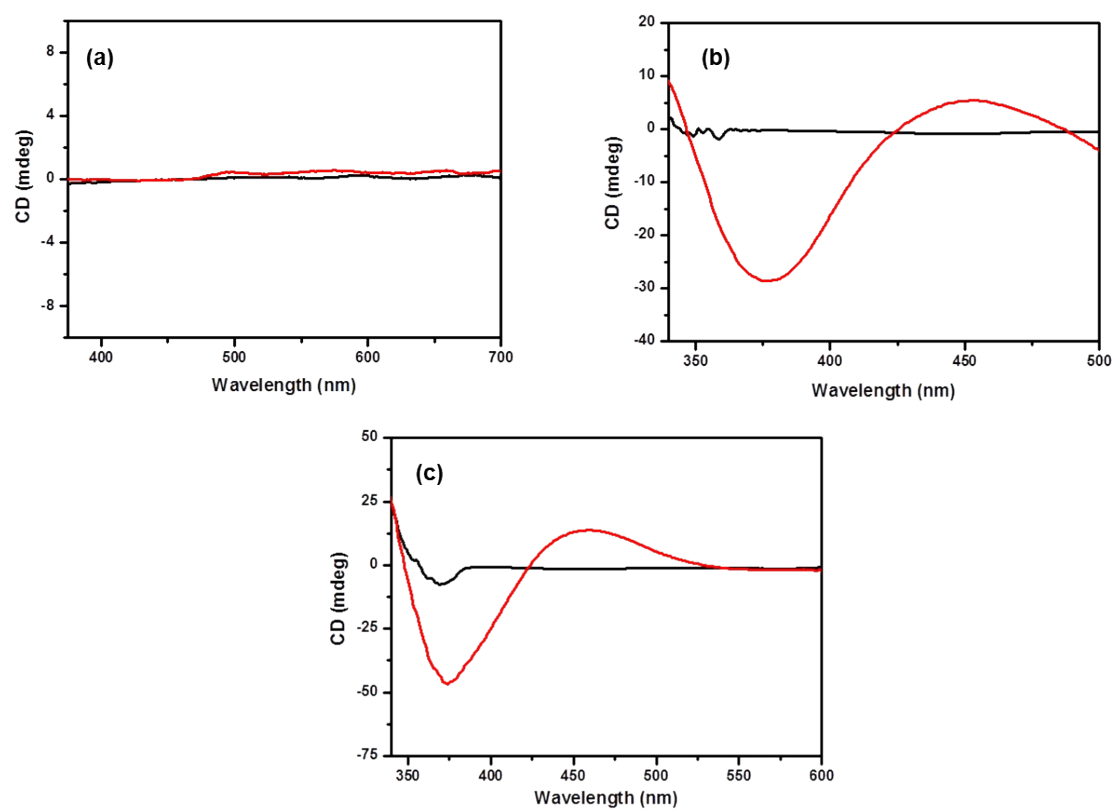


Figure S3. CD spectra of **1a** (a), **1c** (b) and **1d** (c) in CHCl_3 solution and 1,4-dioxane gels. Solutions are black lines and gels are red lines.

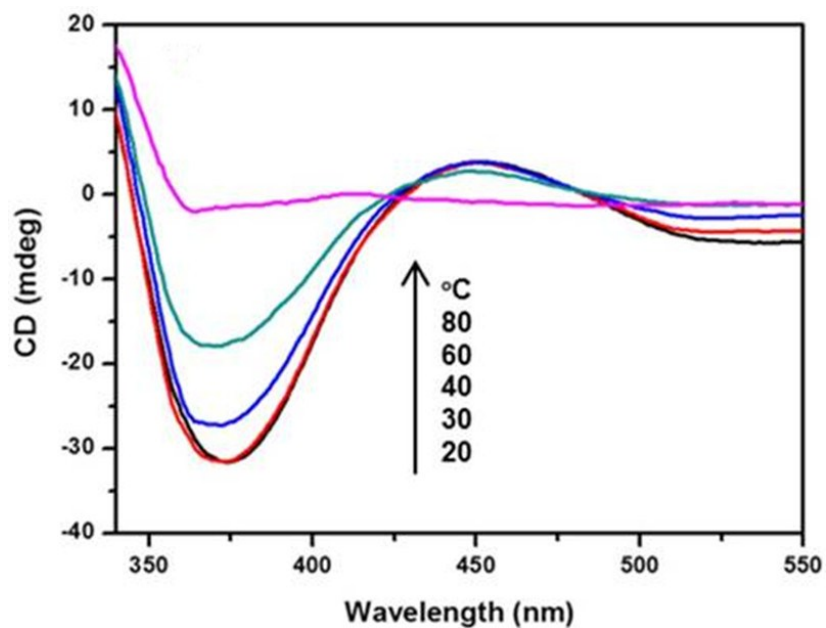


Figure S4. Temperature-dependence of the CD spectra of the organogel of **1c** in 1,4-dioxane. The concentration is 10 mg/mL.

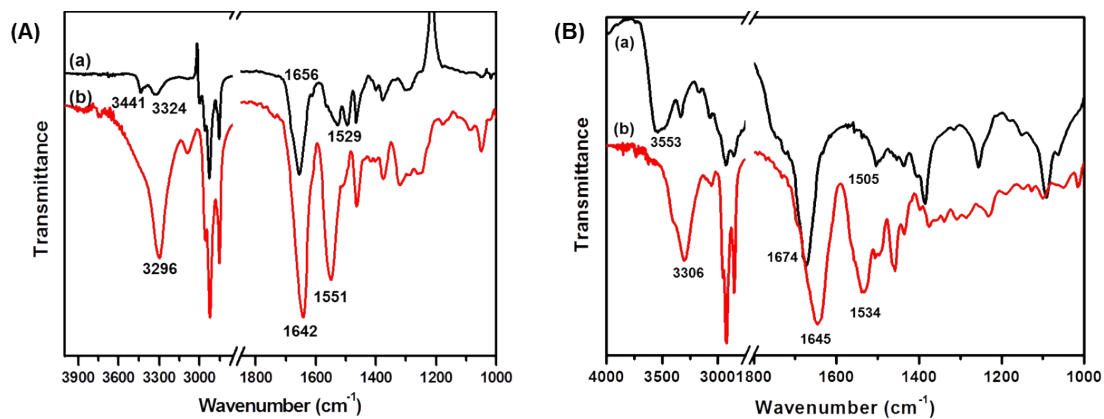


Figure S5. FTIR spectra of (A) **1a** and (B) **1d** in CHCl_3 solution (a), xerogel from 1,4-dioxane (b).

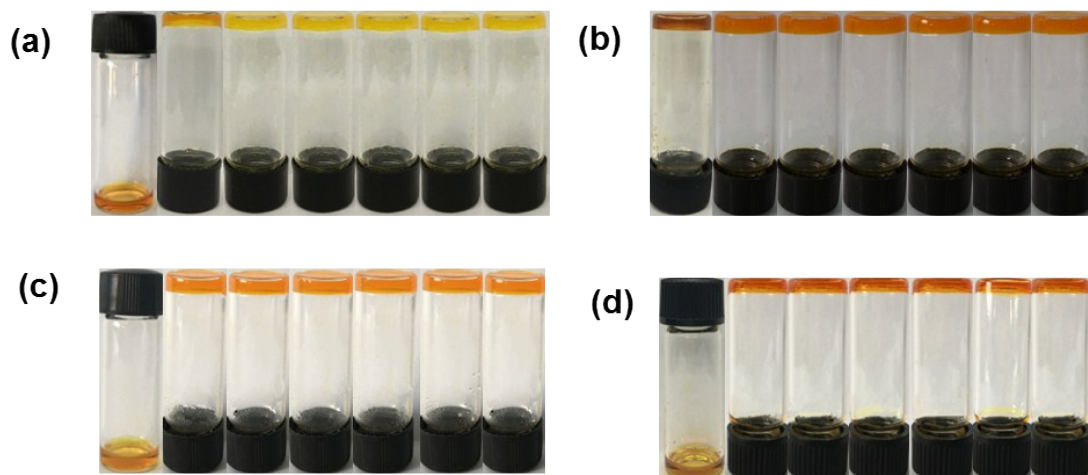


Figure S6. Photographs of the **1c** and **1d** organogels in 1,4-dioxane (a, c) and DMSO (b, d) upon the addition of 5.0 equiv. of each anion. From left to right is addition of F⁻, Cl⁻, Br⁻, I⁻, AcO⁻, HSO₄⁻ and H₂PO₄⁻.

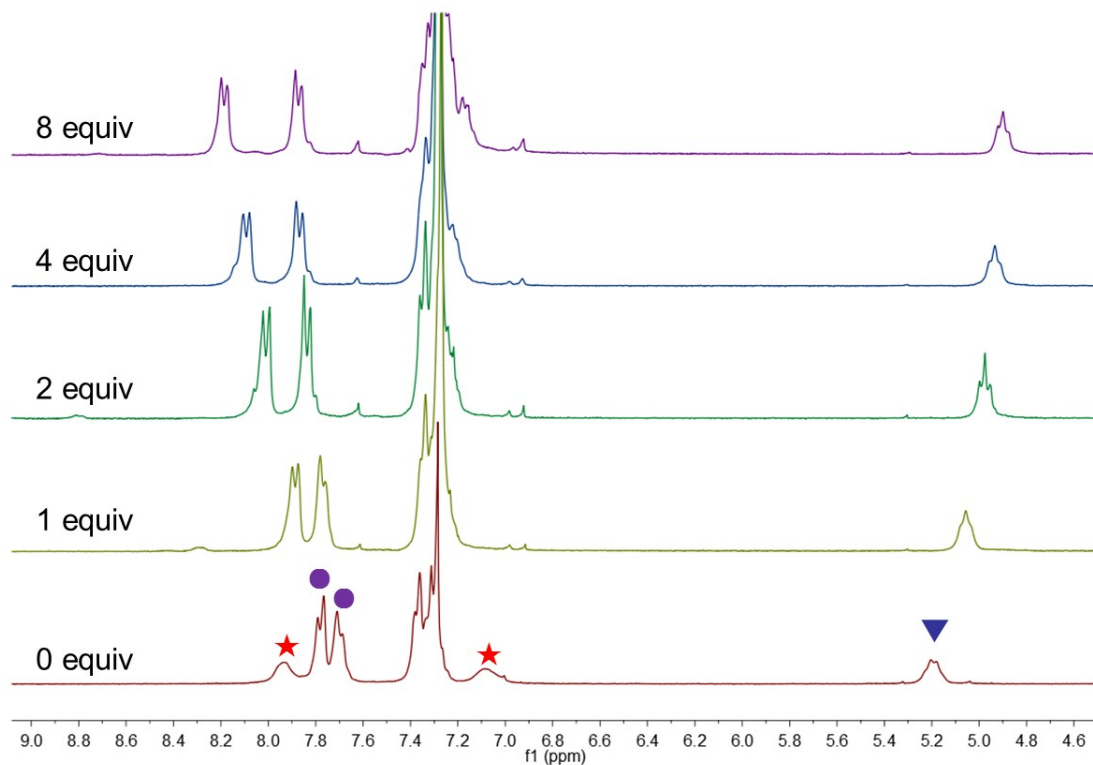


Figure S7. Partial ¹H NMR spectra of **1c** in CDCl₃ (5 mg/mL) upon the addition of F⁻. ★: the amide N-H; ▼: the C-H on the Phe residues; ●: the aromatic proton on T-shaped skeleton.

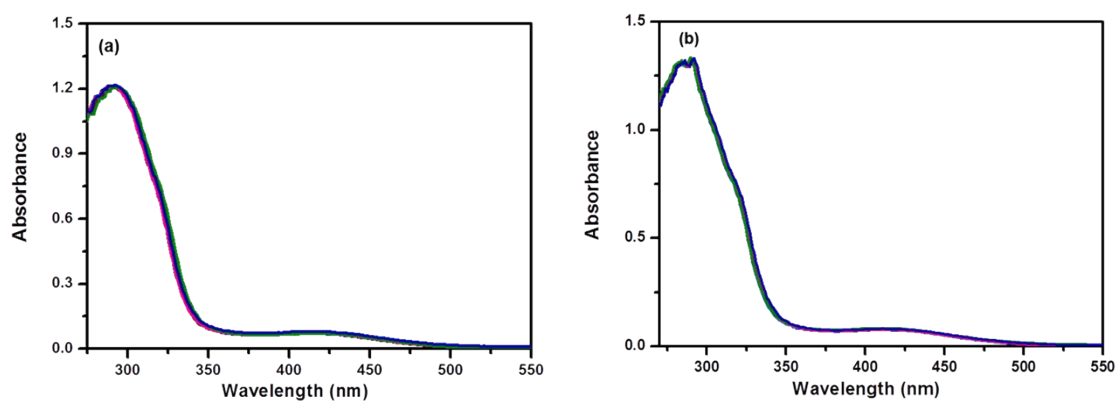


Figure S8. UV-Vis spectra of (a) **1c** in 1,4-dioxane (3×10^{-5} M) and (b) **1d** in DMSO (3×10^{-5} M) with increasing addition of F⁻ (0-20 equiv).

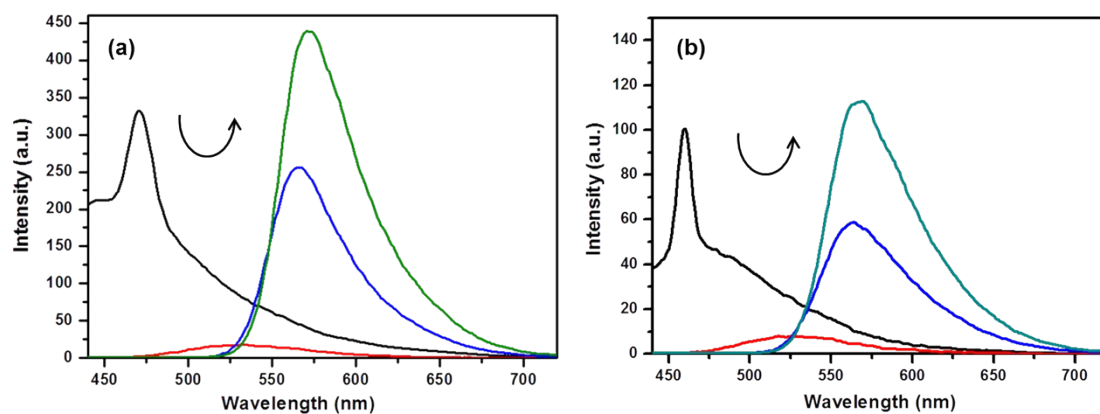


Figure S9. Fluorescence spectra of **1a** (a) and **1d** (b) in 1,4-dioxane at different concentrations. 1×10^{-5} M (black line), 1×10^{-3} M (red line), 5×10^{-3} M (CGC, blue line), 8×10^{-3} M (green line).

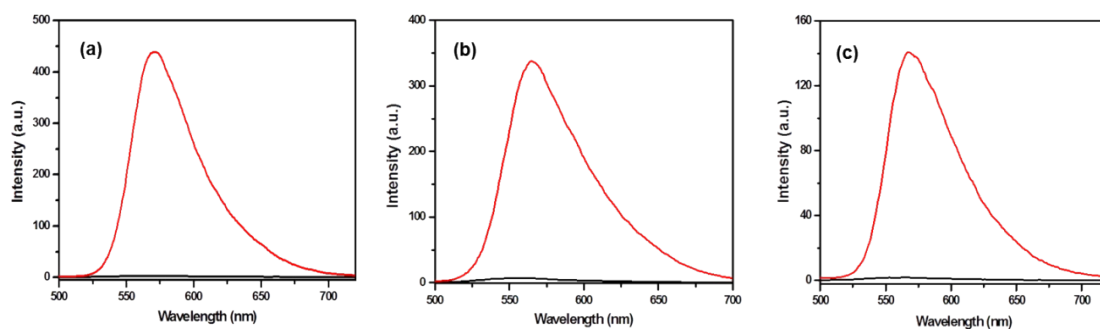


Figure S10. Fluorescence spectra of **1a** (a), **1c** (b) and **1d** (c) in the gel phase (red) and hot solution (black) in 1,4-dioxane (10 mg/mL).

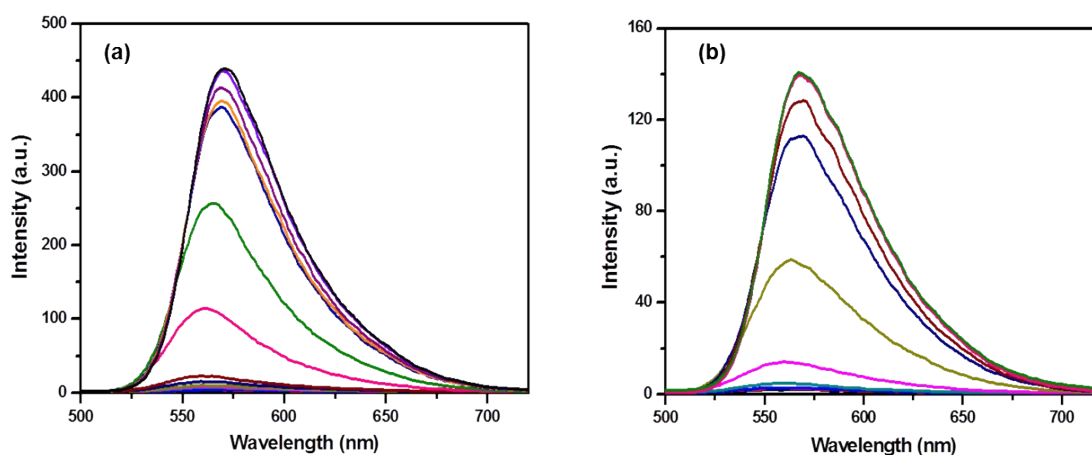


Figure S11. Time-dependent fluorescence changes of gelator **1a** (a) and **1d** (d) in 1,4-dioxane (10 mg/mL).

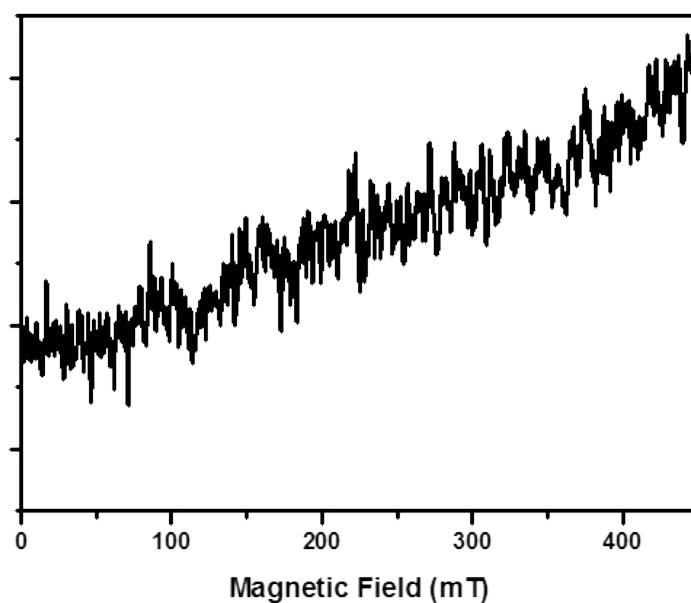


Figure S12. EPR spectrum of the binary gel of **1c** with 0.5 equiv C_{60} .

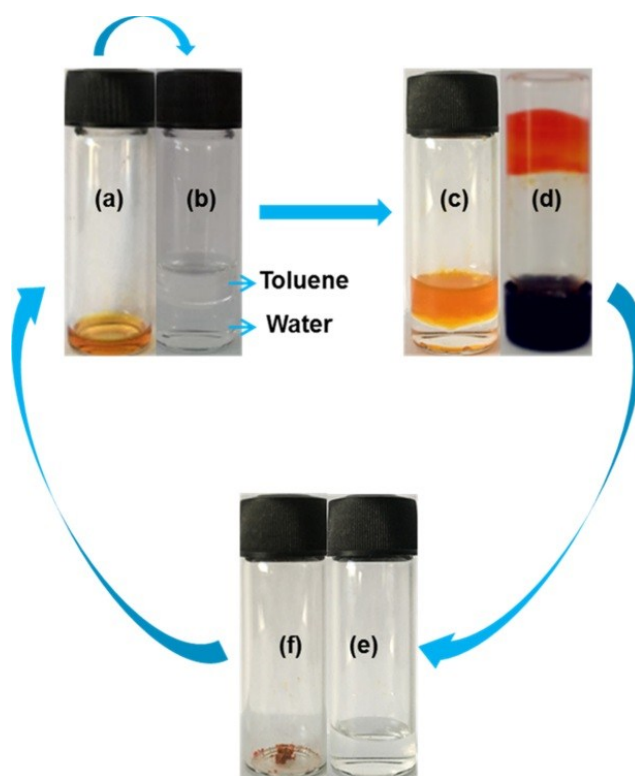


Figure S13. Room-temperature phase-selective gelation and recovery of gelator **1c**: (a) 100 μL CHCl_3 solution of **1c** (14 mg), (b) 400 μL of water and 300 μL of the toluene mixture, (c) the mixture of (a) and (b) induces the phase-selective gelation at room temperature, (d) the inclined tube after the selective gelation, (e) the separated water and (f) recovered xerogel of **1c**.

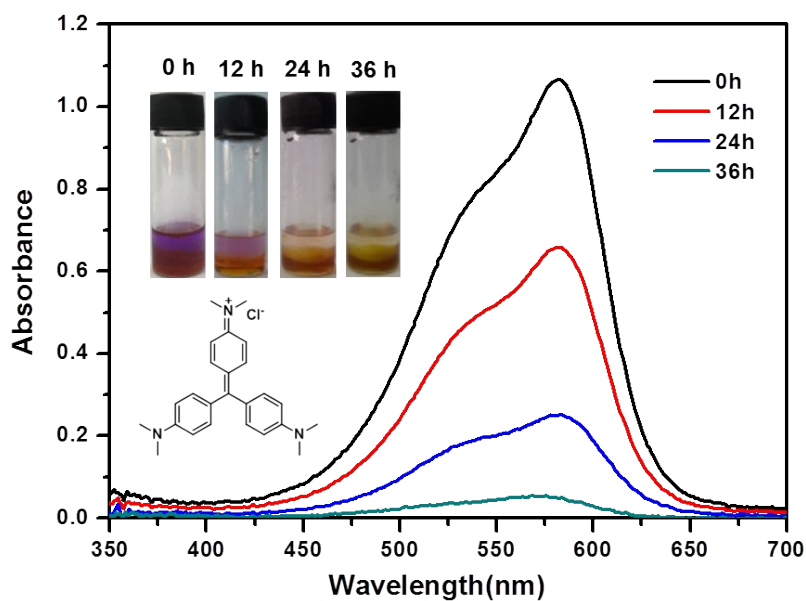
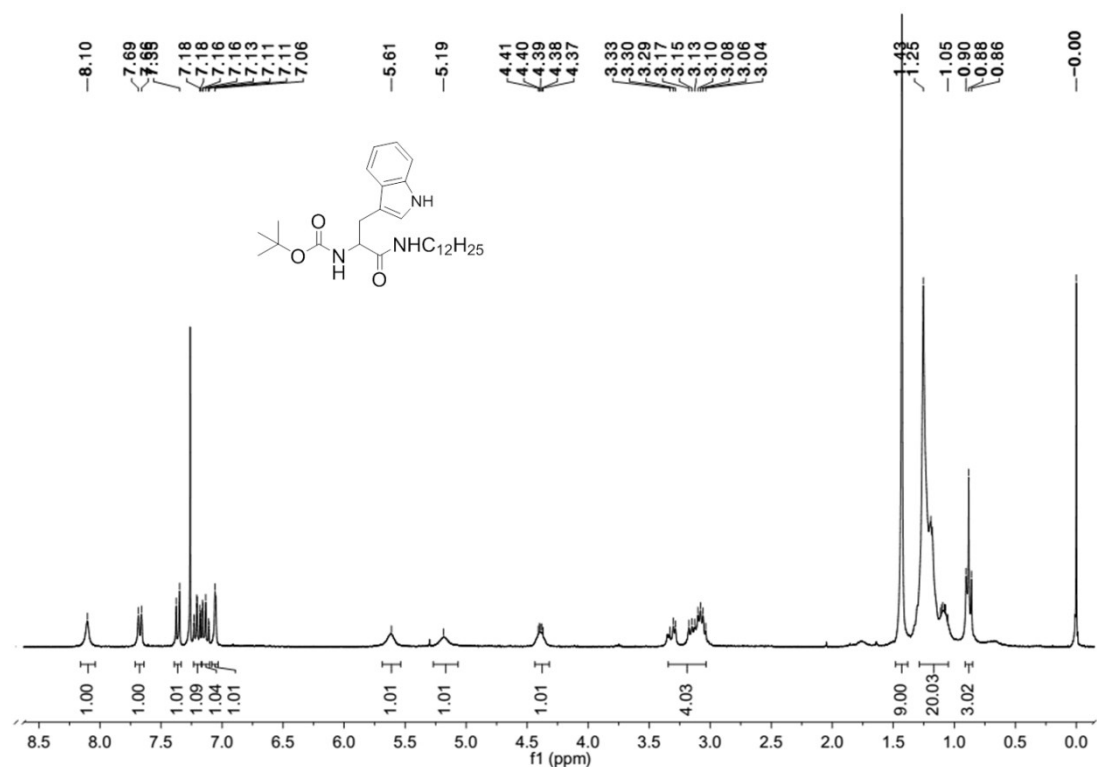
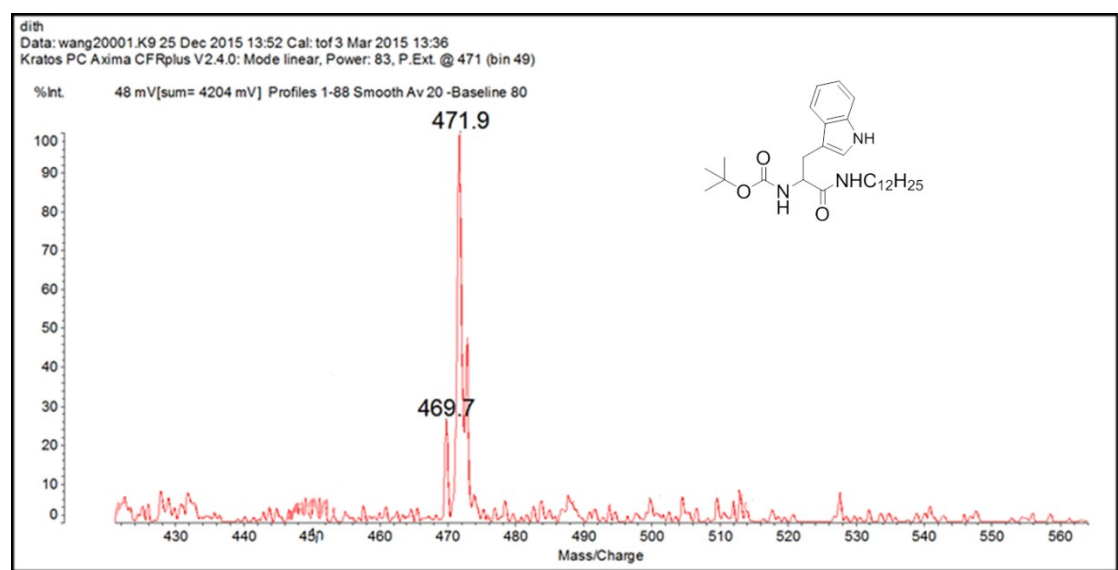
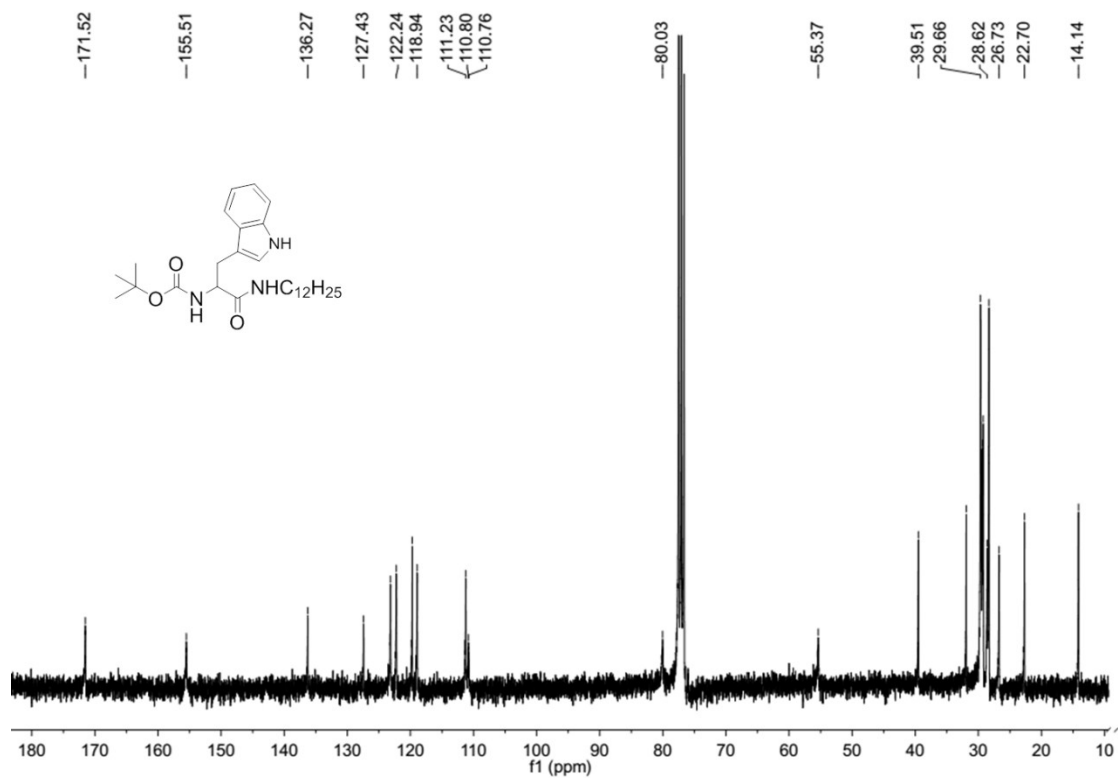
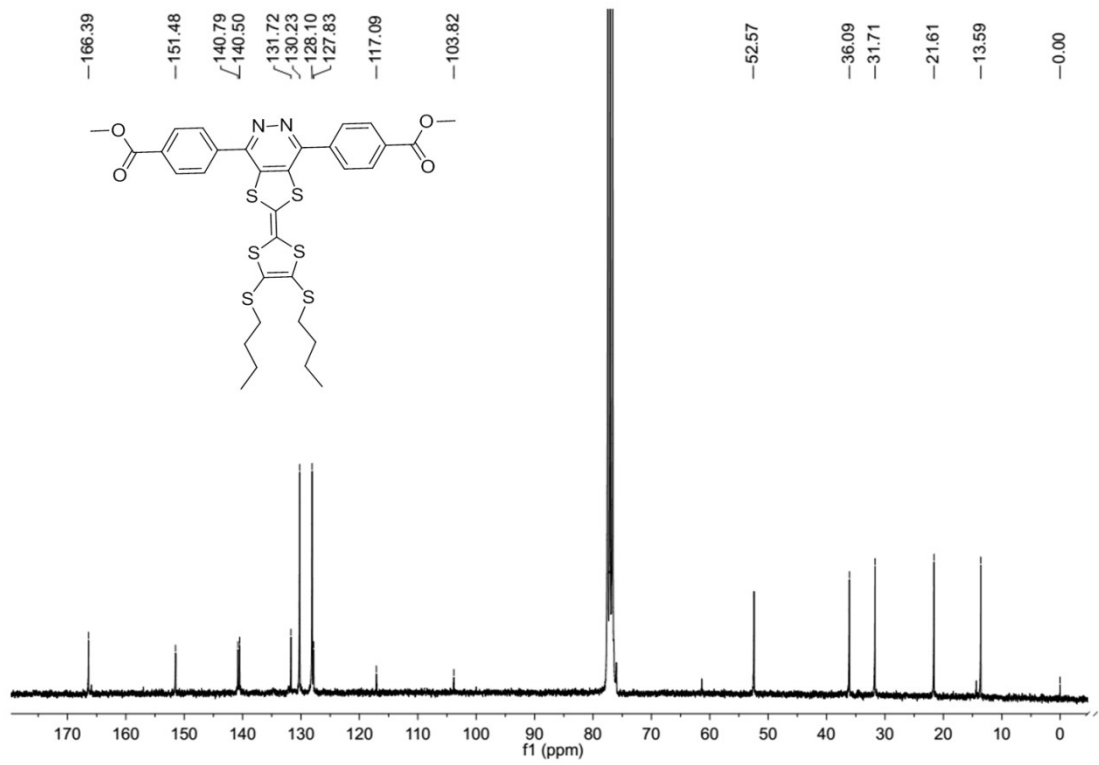
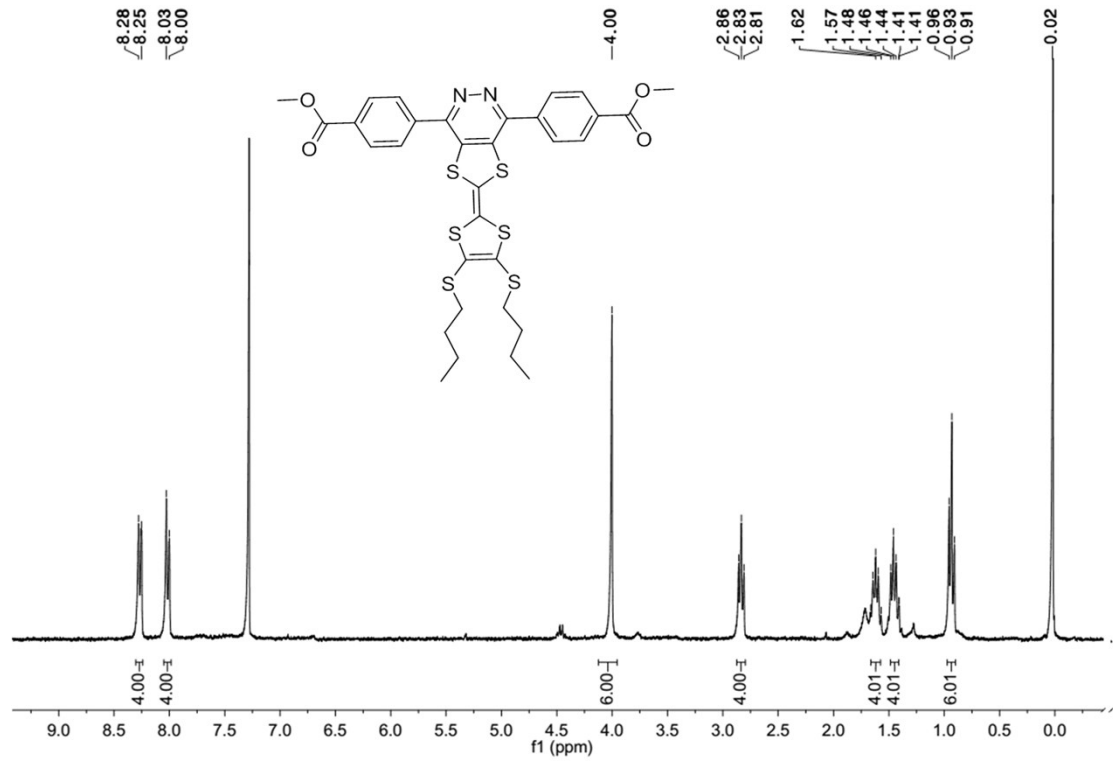


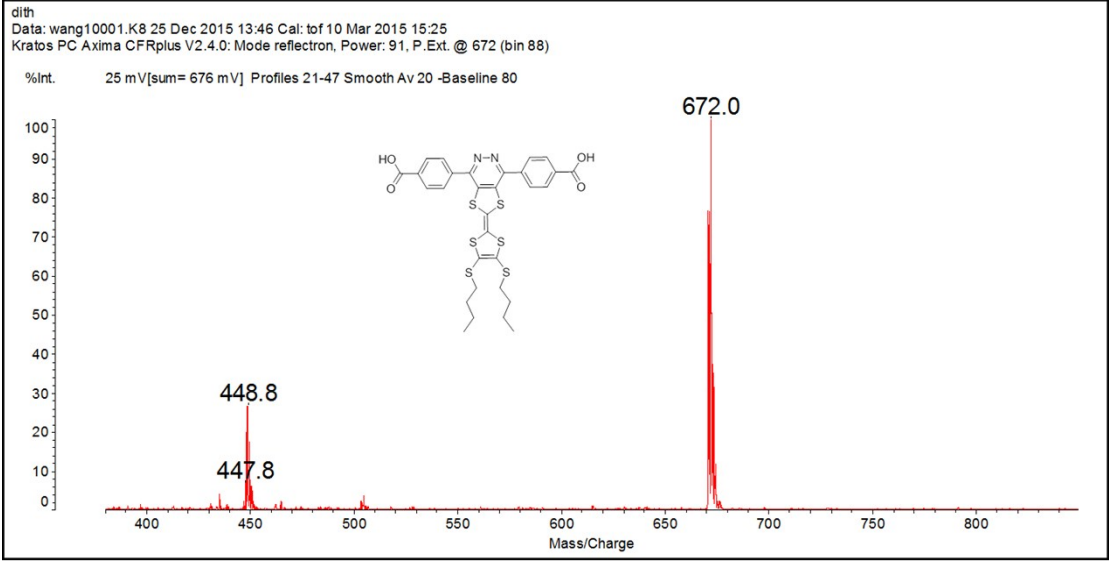
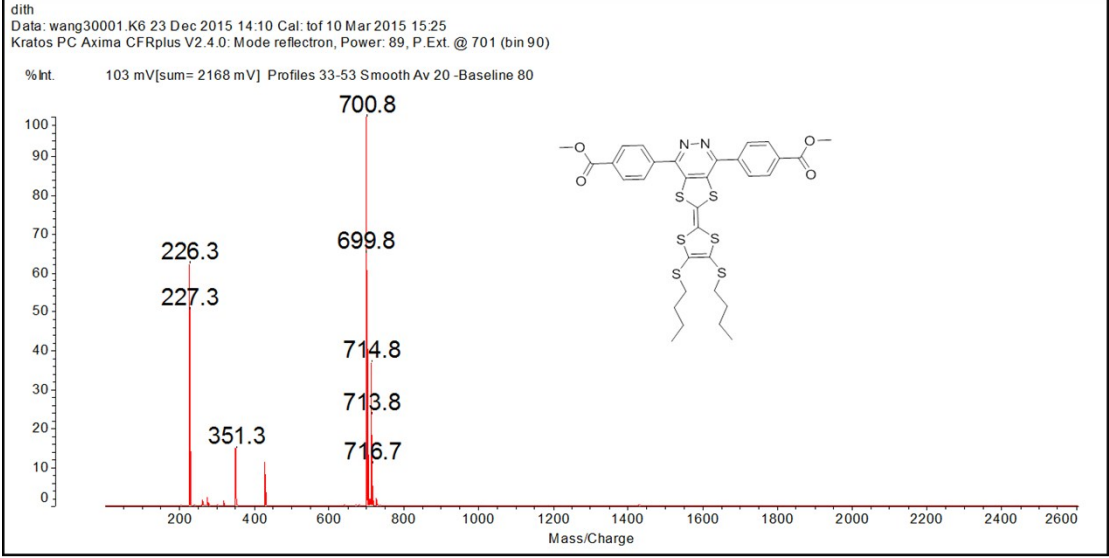
Figure S14. Absorption spectra of aqueous solutions crystal violet after adsorption by organogel **1c** in 1,4-dioxane for different times; 1.0 mL of crystal violet solution (2×10^{-4} M) was added on the top of 0.4 mL gel and the samples were tested at room temperature.

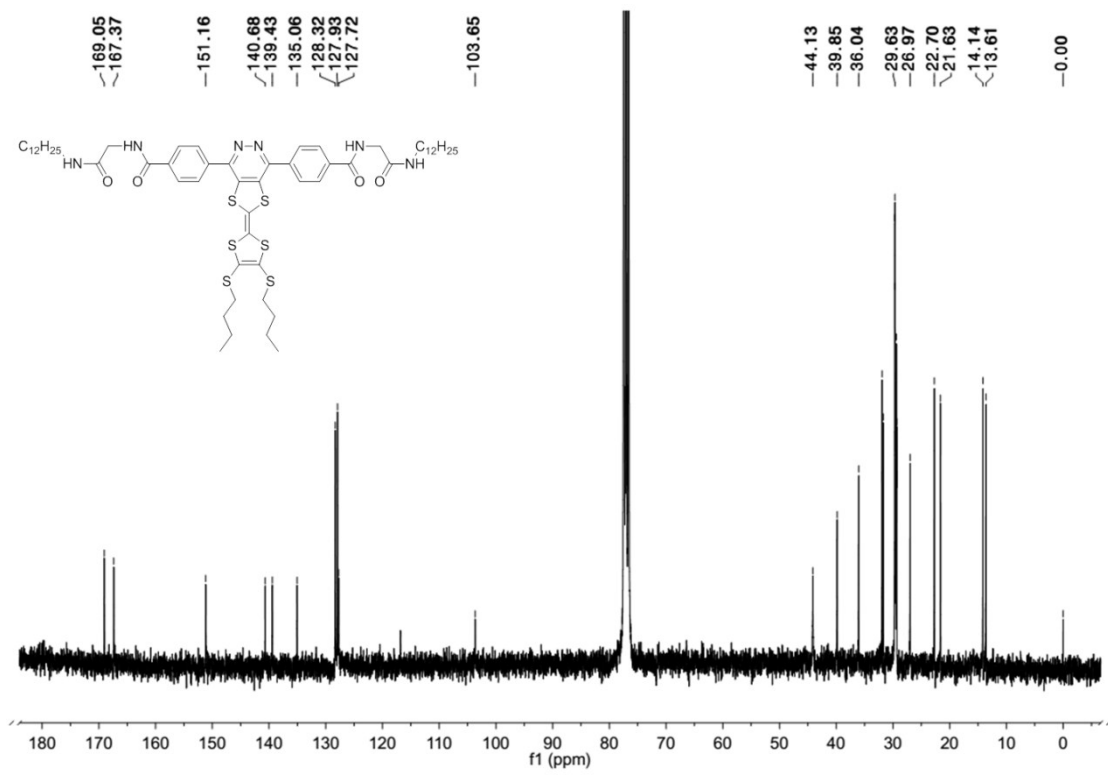
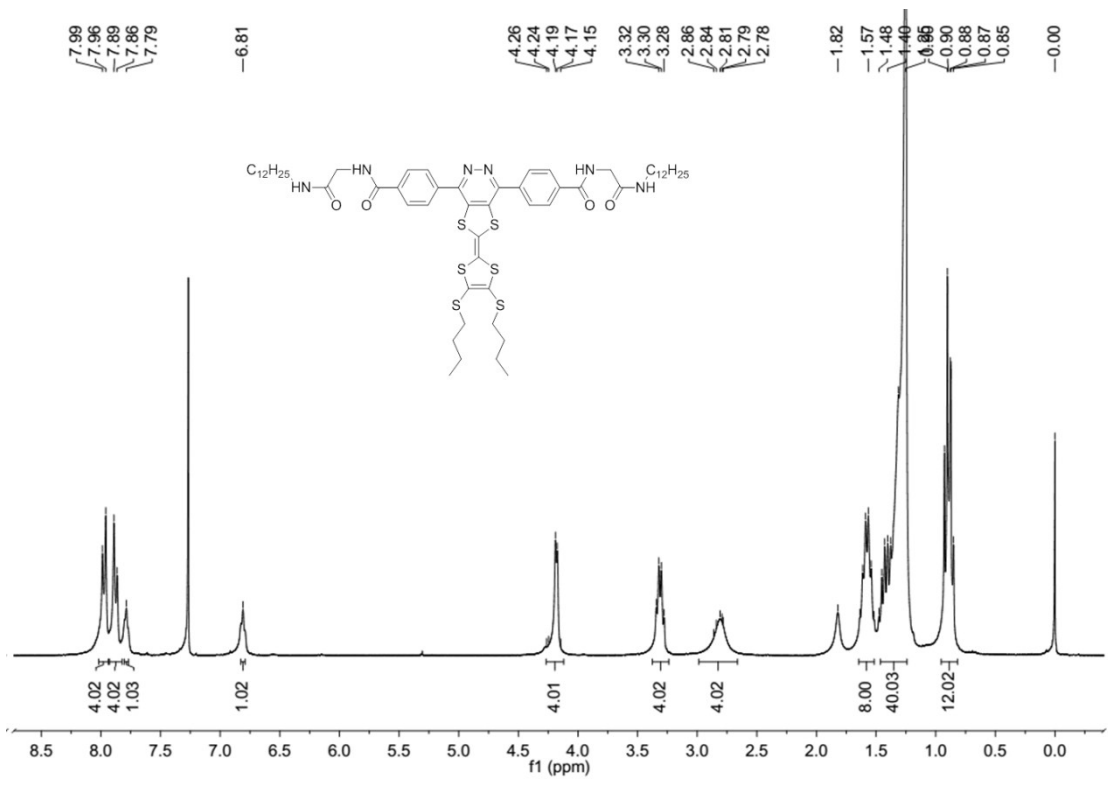
3. ^1H , ^{13}C NMR and MALDI-TOF-MS Spectra

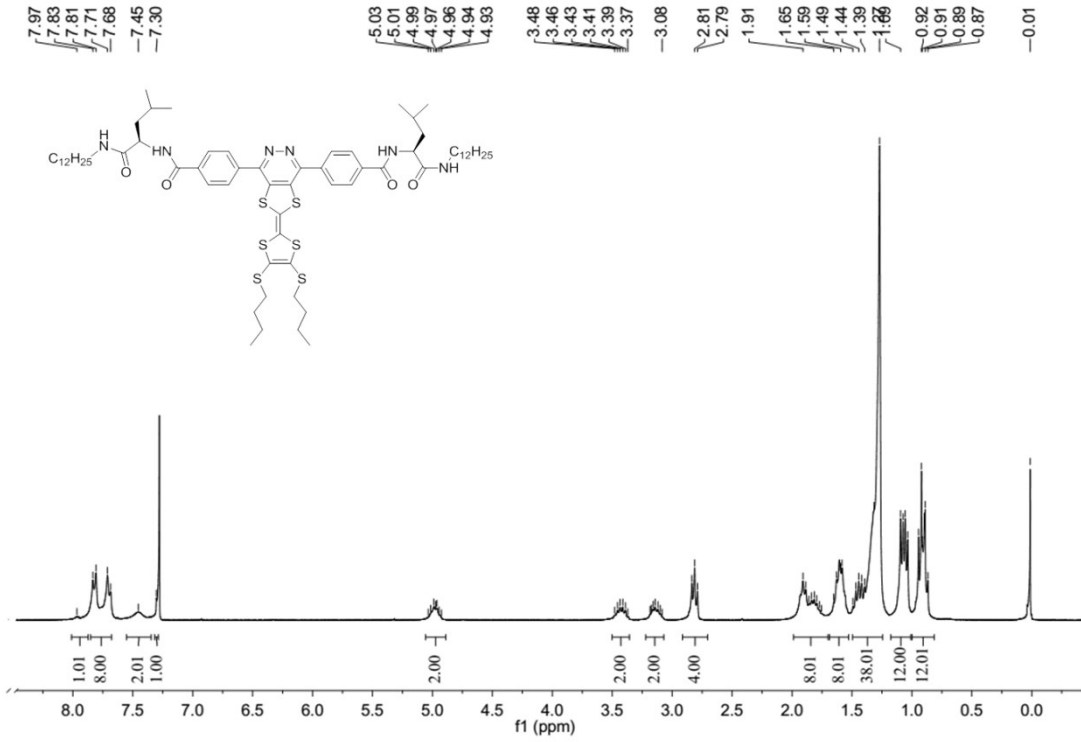
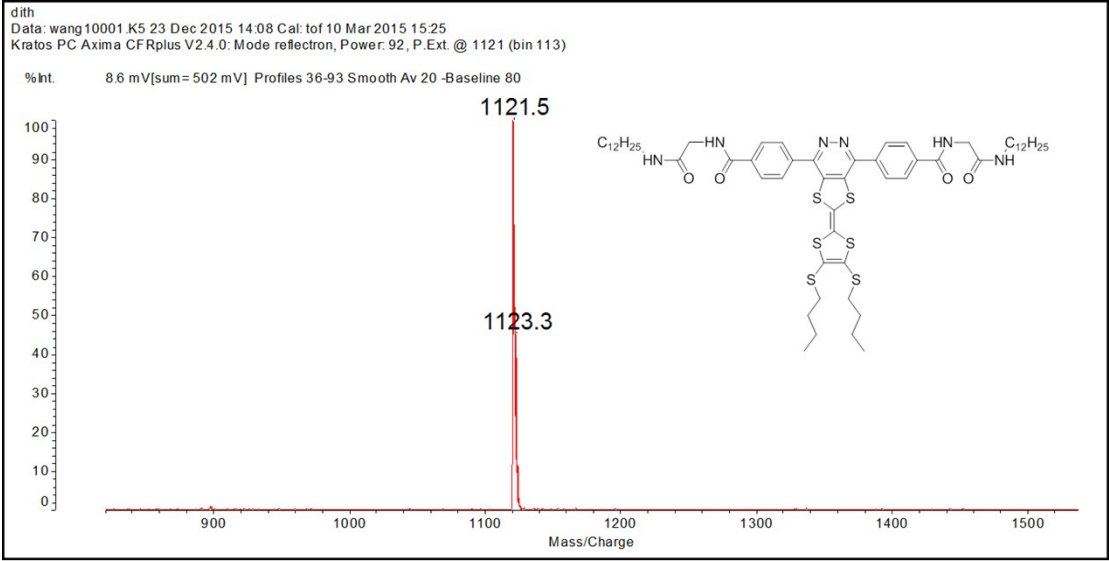


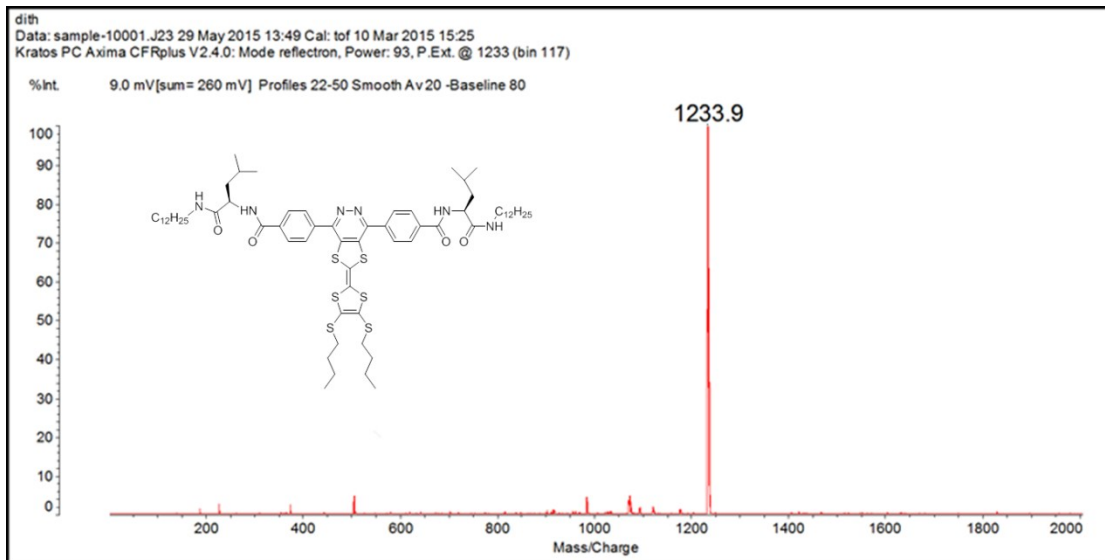
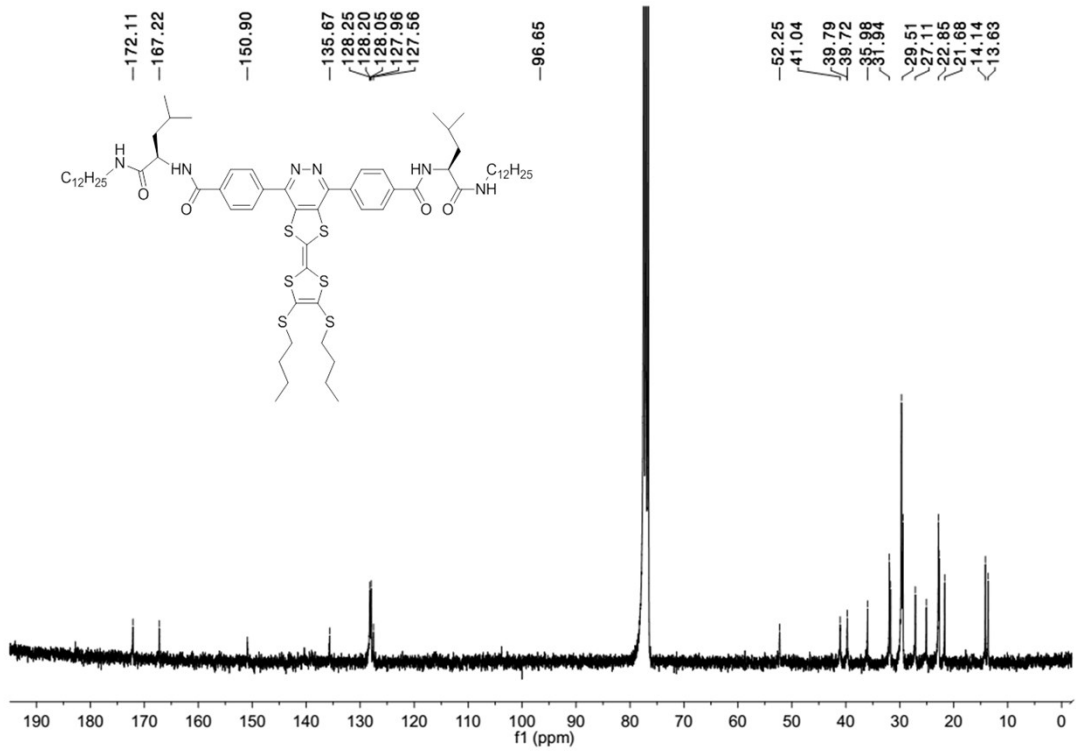


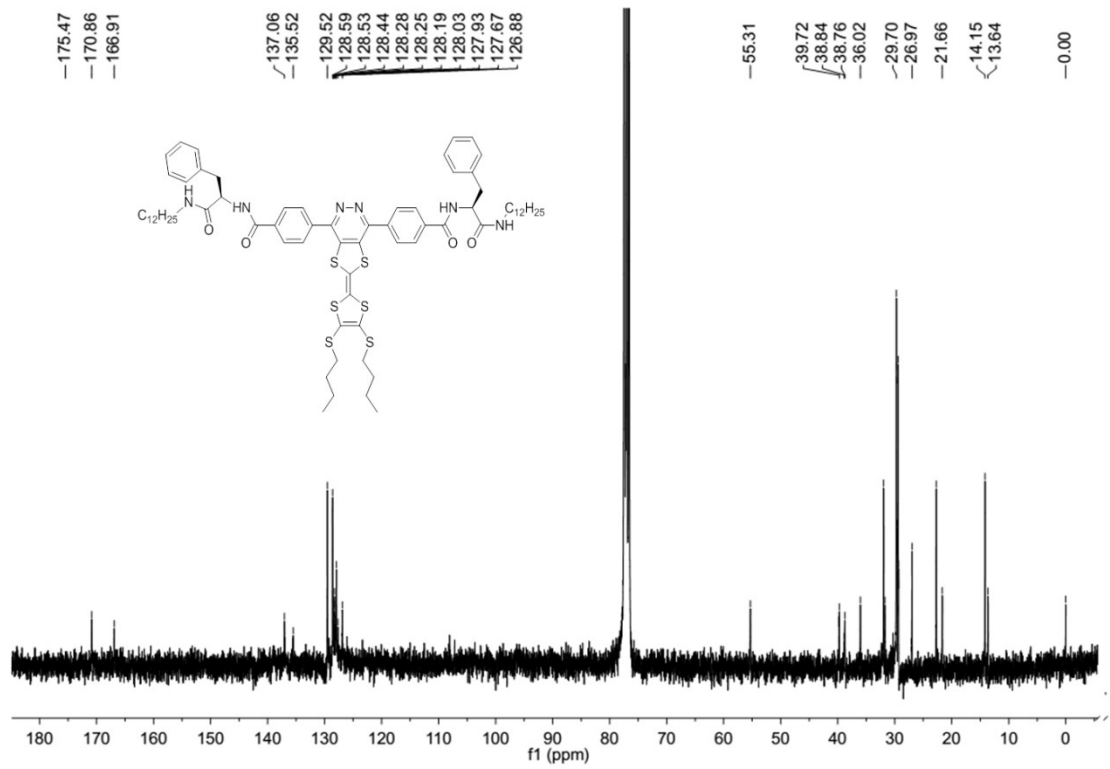
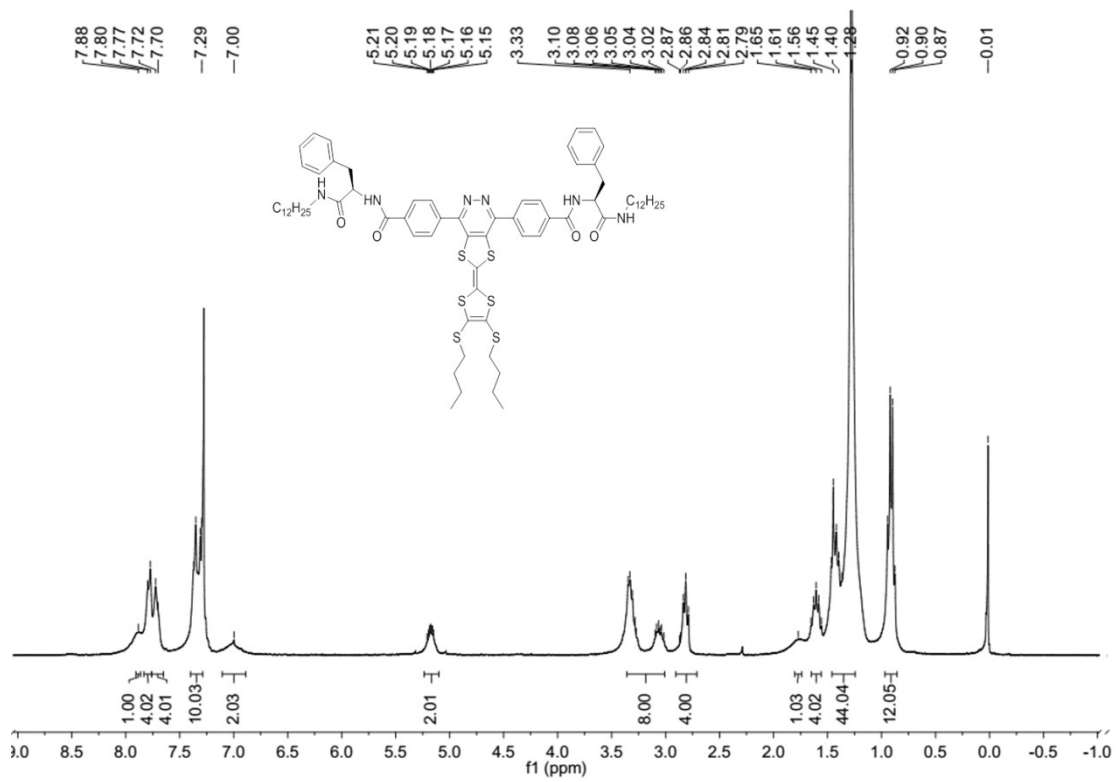












dith
Data: wang20001.K6 23 Dec 2015 14:09 Cal: tof 3 Mar 2015 13:36
Kratos PC Axima CFRplus V2.4.0: Mode linear, Power: 79, P.Ext. @ 1301 (bin 61)

%Int. 31 mV[sum= 1551 mV] Profiles 1-50 Smooth Av 20 -Baseline 80

