

Direct Electron Transfer-Type Dual Gas-Diffusion H₂/O₂ Biofuel Cells

Keisei So,^a Yuki Kitazumi,^a Osamu Shirai,^a Koji Nishikawa,^b Yoshiki Higuchi,^b and Kenji Kano^{*a}

Supporting Information

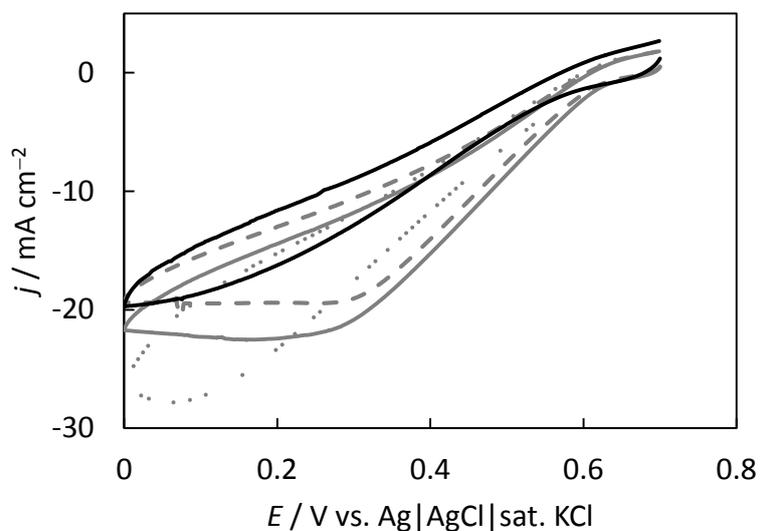


Fig. S1 CVs at the BOD-adsorbed KB/PTFE/WPCC (black line) and the BOD-adsorbed BL/KB/PTFE/WPCC (gray line). The scan rate was 10 mV s⁻¹. The bilirubin concentrations were 1 (dotted line), 3 (solid line), and 5 mM (broken line). Measurements were carried out in 1.5 M citrate buffer (pH 5) at 40 °C under quiescent and O₂-atmospheric conditions.

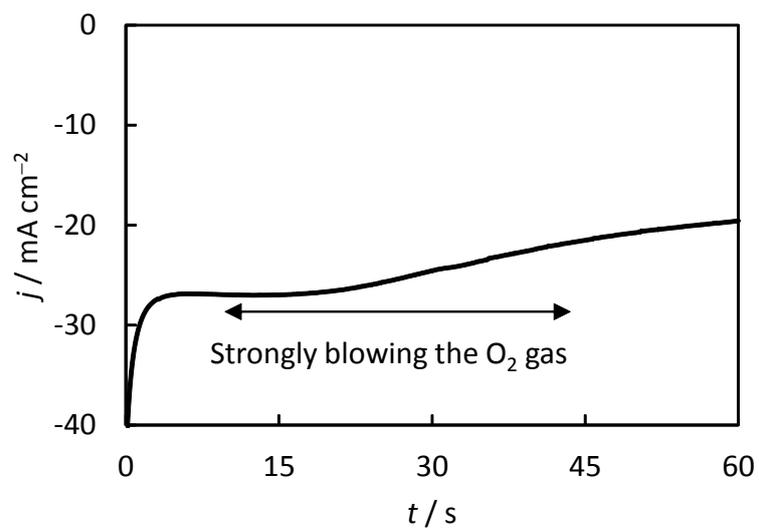


Fig. S2 CA at the BOD-adsorbed KB/PTFE/WPCC. The electrode potential is 0 V. O₂ gas was strongly blown on the surface of the gas-diffusion-type electrode. Measurements were carried out in 1.5 M citrate buffer (pH 5) at 40 °C under quiescent and O₂-atmospheric conditions. The PTFE content was 50%.

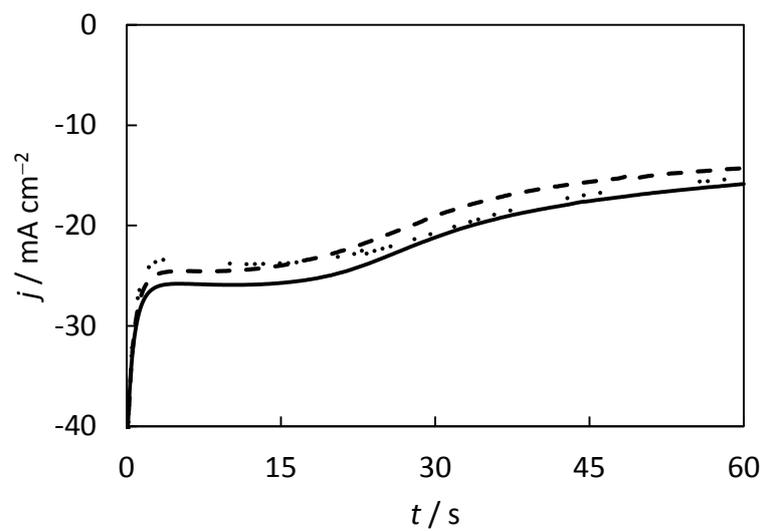


Fig. S3 Repeated CAs at the BOD-adsorbed KB/PTFE/WPCC. The electrode potential is 0 V. Solid, broken, and dotted lines indicate the first, second, and third measurements. The electrode was left in open-circuit for 5 min before the second and third measurements. Measurements were carried out in 1.5 M citrate buffer (pH 5) at 40 °C under quiescent and O₂-atmospheric conditions. The PTFE content was 50%.

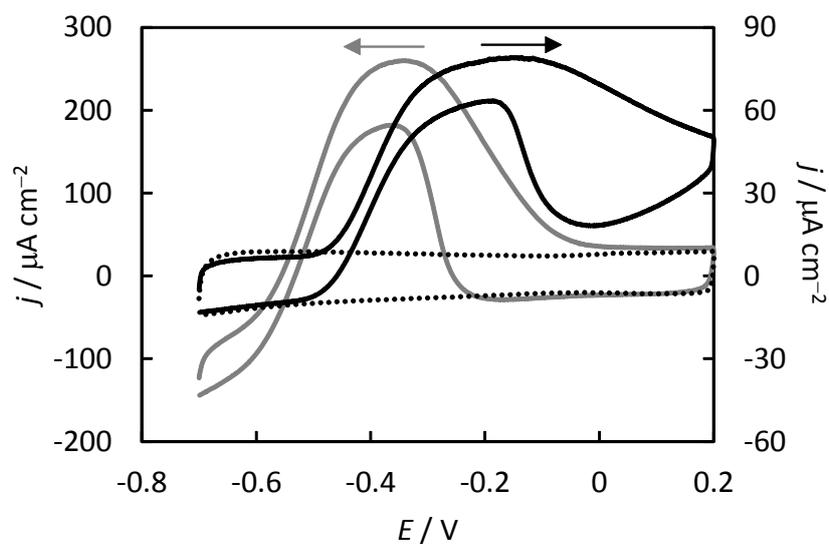


Fig. S4 Rotating disk cyclic voltammograms at *HmMBH*-adsorbed (black solid line, right axis) and *DvMF*-adsorbed (gray solid line, left axis) KB-modified glassy carbon electrodes. The dotted black line is the results at KB-modified glassy carbon electrode without enzymes (with the right axis). The scan rate was 10 mV s^{-1} . Measurements were carried out in 0.1 M phosphate buffer (pH 6) at $40 \text{ }^\circ\text{C}$, a rotating rate of 4000 rpm under H_2 -atmospheric conditions. The electrodes were prepared according to the literature.⁸⁰

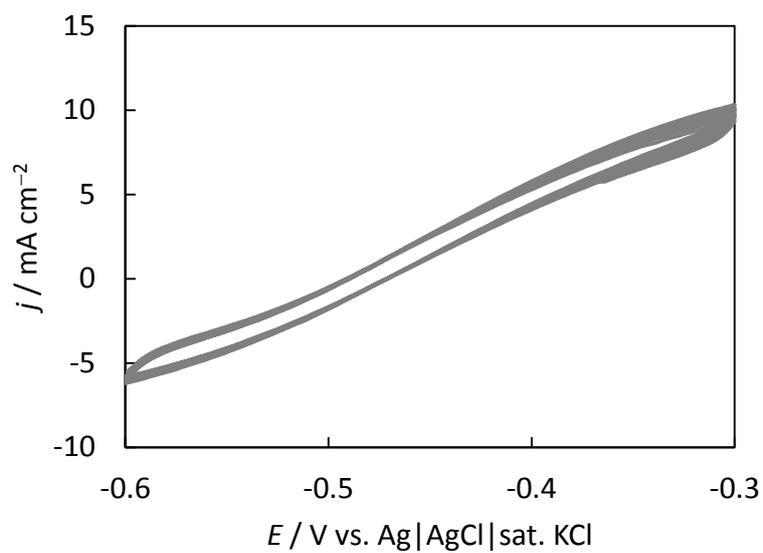


Fig. S5 Multi-swept CVs at the DvMF-adsorbed KB/PTFE/WPCC for 10 cycles. The scan rate was 10 mV s^{-1} . Measurements were carried out in 1.5 M citrate buffer (pH 5) at $40 \text{ }^\circ\text{C}$ under quiescent and H_2 -atmospheric conditions. The PTFE content was 50%.

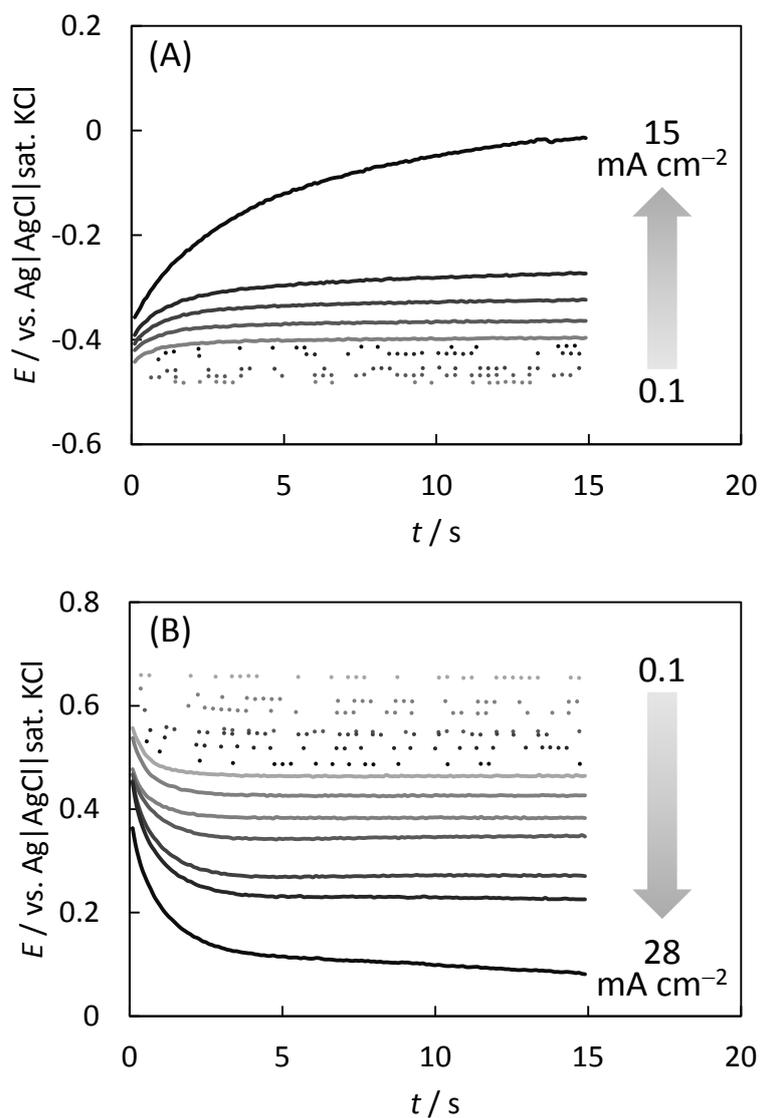


Fig. S6 (A) Chronopotentiograms at the $DvMF$ -adsorbed $KB/PTFE/WPCC$. The oxidation current densities were $0.1, 1, 2, 4, 5, 6, 8, 10, 12,$ and 15 mA cm^{-2} (from bottom to top). (B) Chronopotentiograms at the BOD -adsorbed $BL/KB/PTFE/WPCC$. The reduction current densities were $0.1, 1, 2, 4, 5, 6, 8, 10, 12, 15, 16, 20, 22,$ and 28 mA cm^{-2} (from top to bottom). Measurements were carried out in 1.5 M citrate buffer ($\text{pH } 5$) at $40 \text{ }^\circ\text{C}$ under quiescent and (A) H_2 - or (B) O_2 -atmospheric conditions. The PTFE content and the bilirubin concentration were 50% and 3 mM , respectively.

Table S1 Details of recent development in power of biofuel cells

H ₂ /O ₂ Biofuel Cells						
Date	Power density / mW cm ⁻²	Anode catalyst	Cathode catalyst	Conditions	Notes	Ref.
2001	0.4	DvMF cells	MvBOD	pH 7, room temperature (RT), 100% H ₂ , 100% O ₂	MV and ABTS are used as mediators. Using a membrane as a separator	17
2002	0.32	Hase	LAC	pH 8 (anode), pH 4.2 (cathode)	Using a membrane as a separator	18
2006	0.005	Rm CH34 MBH	TvLAC	pH 5, 100 mM citrate, 3% H ₂ in air	Low H ₂ concentration was selected for avoiding an explosion	19
2010	0.063	<i>E. coli</i> Hyd1	MvBOD	pH 5, 100 mM citrate, RT, 100% H ₂ , 100% O ₂	Using a membrane as a separator	20
2012	0.12	<i>E. coli</i> Hyd1	MvBOD	pH 5, 100 mM citrate, RT, H ₂ /air (80:20 mixture), under quiescent conditions	N.A	21
2012	0.3	AaMBH	MvBOD	pH 6.8, 50 mM Hepes, 60 °C (anode), 25 °C (cathode), 100% H ₂ , 100% O ₂	Using a membrane as a separator	22
2013	a) 0.56 b) 1.67	<i>E. coli</i> Hyd1	MvBOD	pH 6, 100 mM phosphate, 25 °C 78% H ₂ , 22% air, under quiescent conditions	a) The cathode surface area is equal to anode. b) The cathode surface area is 3-times larger than anode.	23
2014	1.5	AaMBH	BpBOD	pH 6, 100 mM phosphate, 60 °C 100% H ₂ , 100% O ₂ , under quiescent conditions.	Using a membrane as a separator	24
2015	0.72	AaMBH	BpBOD	pH7.2, phosphate buffer, 45 °C Air-breathing/H ₂ under quiescent conditions	Gas-diffusion-type electrode is used for the cathode	25
2016	8.4	DvMF	MvBOD	pH 5, 1.5 M citrate, 40 °C, 100% H ₂ , 100% O ₂ , under quiescent conditions	Dual gas-diffusion-system	This study
DET-type Biofuel Cells with other substrates						
2007	a) 0.4 b) 0.85	FDH	TsLAC	pH 5, McIlvaine, RT with 200 mM fructose, O ₂ saturated	a) Under quiescent conditions b) With stirring	89
2009	a) 0.66 b) 0.87	FDH	MvBOD	pH 6, 100 mM acetate, 25 °C with 200 mM fructose, O ₂ saturated	a) Under quiescent conditions b) With stirring	90
2011	1.3	GOD and catalase	TvLAC	pH 7, 100 mM phosphate, with 50 mM glucose, air saturated	No information about stirring.	91
2011	1.8	FDH	TsLAC	pH 5, McIlvaine, with 200 mM fructose, under O ₂ saturated stirring conditions	N.A.	59
2013	a) 0.95 b) 2.55	FDH	MvBOD	pH 5, McIlvaine, with 500 mM fructose, air saturated, under quiescent conditions.	a) Single-layer cell, b) Triple-layer cell Gas-diffusion-type electrode is used for the cathode.	83
2014	2.6	FDH	MvBOD	pH 5, 1.0 M citrate, RT, with 500 mM fructose, air saturated, under quiescent conditions	Gas-diffusion-type electrode is used for the cathode.	56