

Supplementary Information

Au-Pd bimetallic nanoparticles anchored on α -Fe₂O₃ nonenzymatic hybrid nanoelectrocatalyst for simultaneous electrochemical detection of dopamine and uric acid in the presence of ascorbic acid

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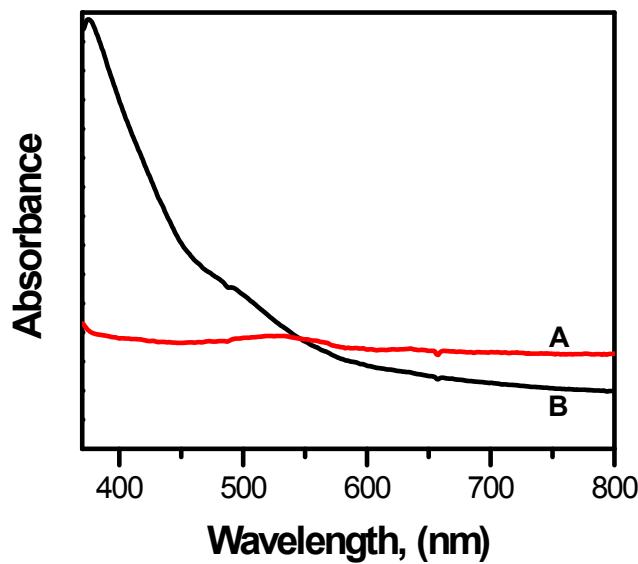


Figure S1. UV visible spectra of (A) $\alpha\text{-Fe}_2\text{O}_3$ (B) $\alpha\text{-Fe}_2\text{O}_3$ / Au-Pd hybrid nanostructure

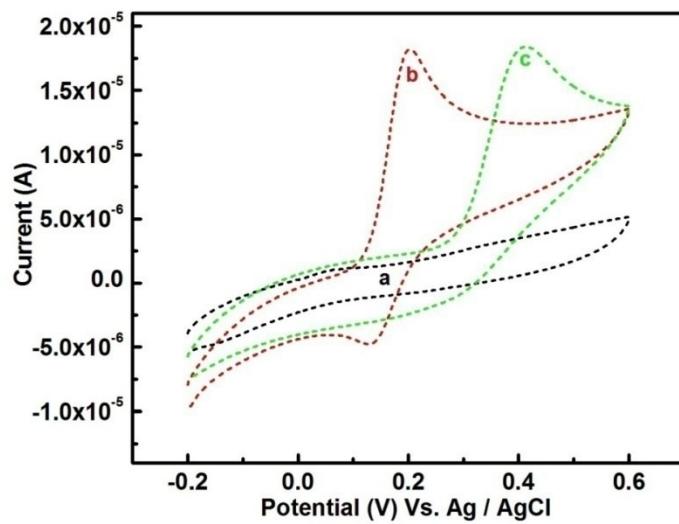


Figure S2. 1mM of (a) Ascorbic acid, (b) Dopamine (c) Uric acid in PBS (pH 7) for the $\alpha\text{-Fe}_2\text{O}_3$ / Au-Pd hybrid modified electrode

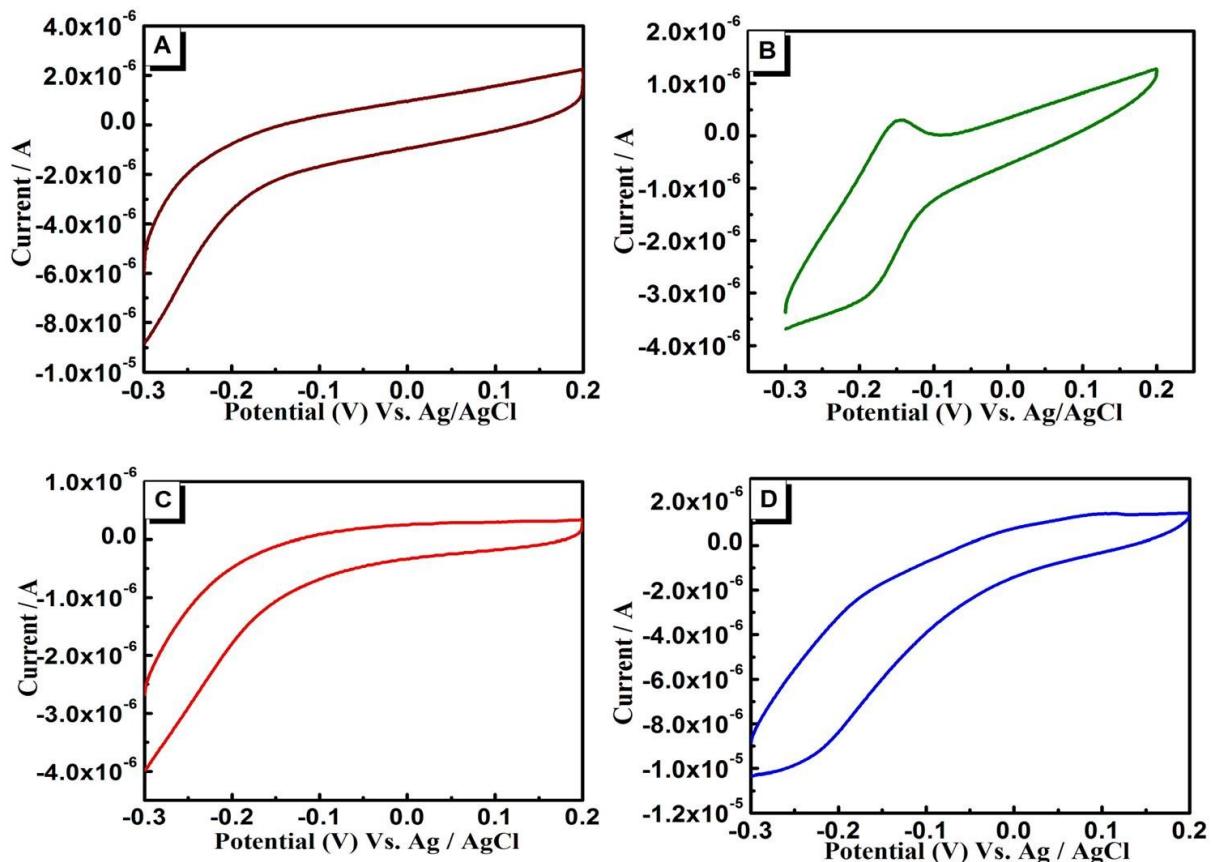


Figure S3. CVs obtained for (1mM) PBS at a 50 mV s⁻¹ with addition 25 μ M of Methylene blue at a (A) bare (B) α -Fe₂O₃ (C) Au-Pd (D) α -Fe₂O₃/Au-Pd hybrid at a potential between -0.3 and 0.2 V (pH 7).

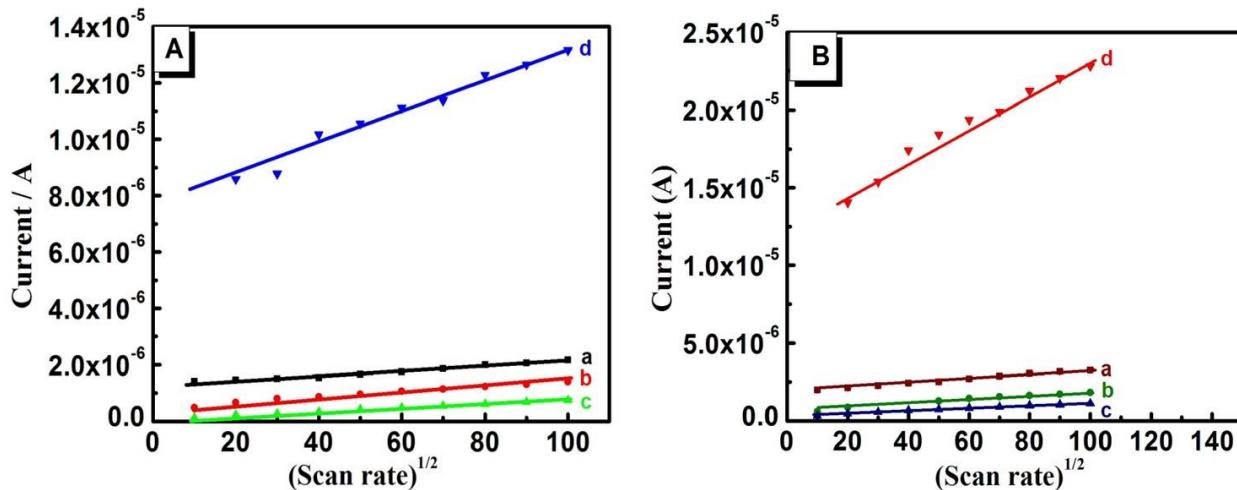


Figure S4. (A) CV studies recorded at different scan rate from ($10 - 100 \text{ mV s}^{-1}$) (a) Bare GC (b) $\alpha\text{-Fe}_2\text{O}_3$ (c) Au/ Pd (d) $\alpha\text{-Fe}_2\text{O}_3$ /Au-Pd hybrid for dopamine. Fig (B) differ scan rate for ($10 - 100 \text{ mV s}^{-1}$) (a) Bare GC (b) $\alpha\text{-Fe}_2\text{O}_3$ (c) Au/Pd (d) $\alpha\text{-Fe}_2\text{O}_3$ /Au-Pd hybrid for uric acid.

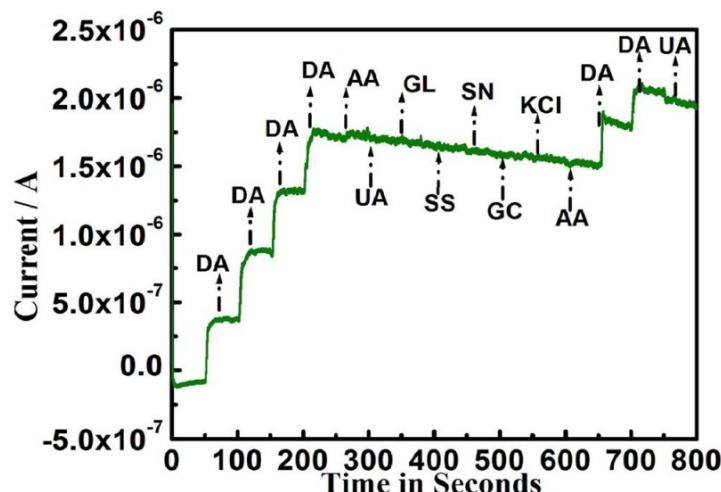


Figure S5. Amperometry (Time Vs Current) for modified $\alpha\text{-Fe}_2\text{O}_3$ /Au-Pd hybrid GC for dopamine & uric acid of $50 \mu\text{M}$ concentration while other interfering analytes of 5 mM concentration

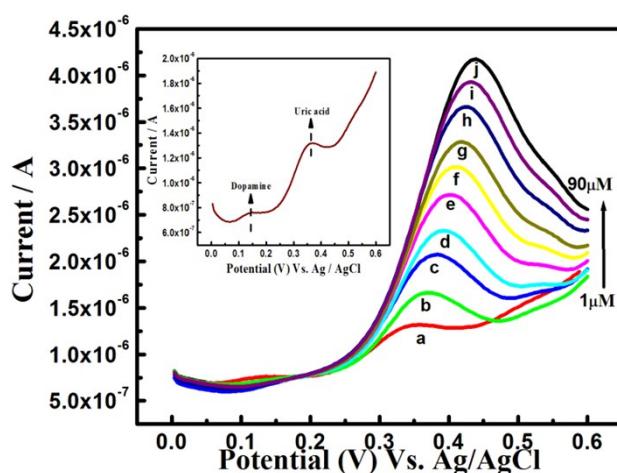


Figure S6. shows the response of $\alpha\text{-Fe}_2\text{O}_3$ /Au-Pd hybrid GC at different concentration of urine sample 1 (a) 1 μM (b) 3 μM , (c) 5 μM , (d) 6 μM , (e) 10 μM , (f) 20 μM , (g) 40 μM , (h) 60 μM , (i) 70 μM (j) 90 μM in 0.1 M PBS containing pH 7.0.

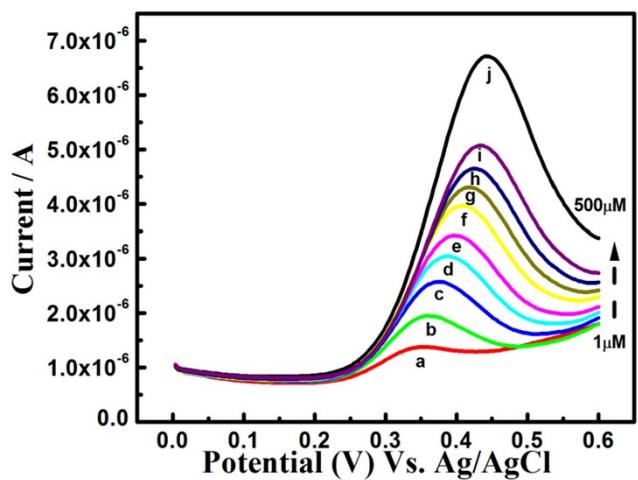


Figure S7. shows the response of $\alpha\text{-Fe}_2\text{O}_3$ /Au-Pd hybrid GC at different concentration of urine sample 2(a) 1 μM , (b) 2 μM , (c) 5 μM , (d) 20 μM , (e) 70 μM , (f) 80 μM , (g) 90 μM , (h) 200 μM , (i) 300 μM , (j) 500 μM in 0.1 M PBS containing pH 7.0

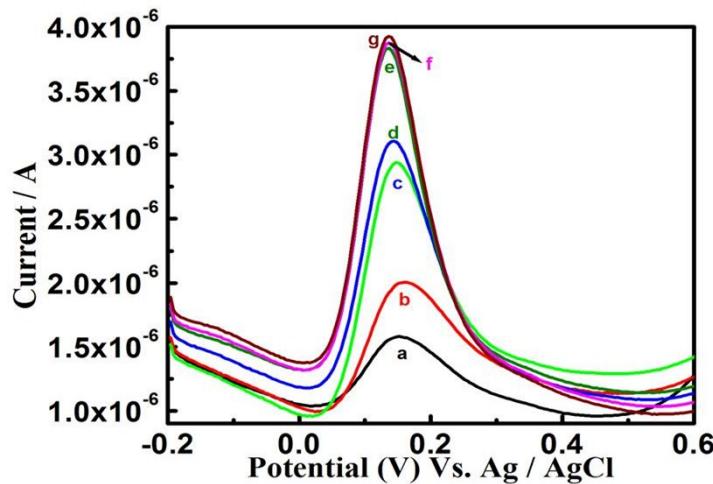


Figure S8. shows the response of $\alpha\text{-Fe}_2\text{O}_3$ /Au-Pd hybrid GC at different concentration of fresh human blood serum (a) 300 nM (b) 1 μ M (c) 4 μ M (d) 5 μ M (e) 10 μ M (f) 25 μ M (g) 50 μ M containing 0.1M of PBS (pH 7.0).

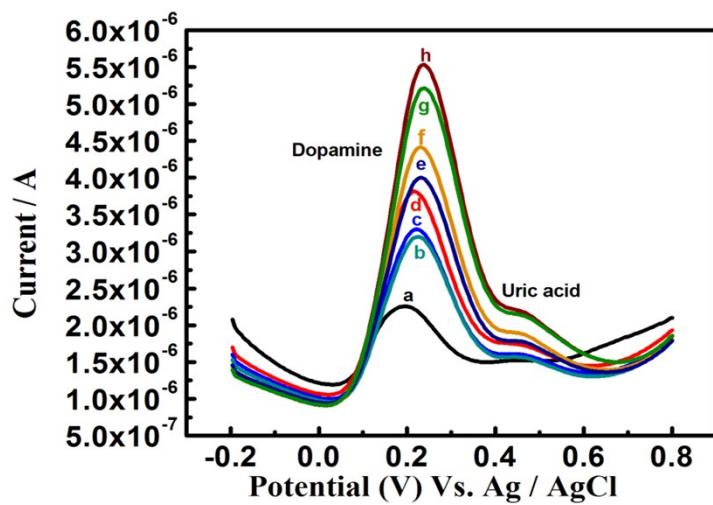


Figure S9. shows the response of $\alpha\text{-Fe}_2\text{O}_3$ /Au-Pd hybrid GC at different concentration of dopamine hydrochloride injection (a) 450 nM, (b) 4 μ M, (c) 6 μ M, (d)10 μ M, (e) 15 μ M, (f) 25 μ M, (g) 50 μ M, (h) 60 μ M containing 0.1M of PBS (pH 7.0)

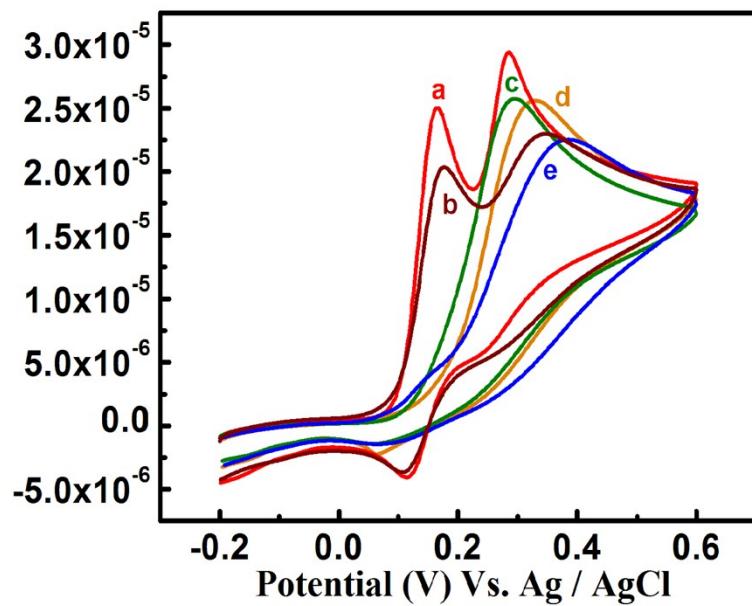


Figure S 10. shows different ratio for Au/Pd bimetal (a)1:1 (b) 1:0.5 (c) 0.5:1 (d) 1:0.25 (e) 0.25:1 ratio containing 600 μM of dopamine & uric acid in 0.1 M of PBS(pH 7.0)

Table S1 - Voltammetric DA,UA in presence of AA recovery tests performed in Human Urine, Serum and Dopamine hydrochloride injection with $\alpha-\text{Fe}_2\text{O}_3$ / Au@Pd , pH = 7.0

Sample	Added [DA, UA] μM	Obtained [DA, UA] μM	Recovery (%) [DA, UA]
Urine 1	-- , 5.00	-- , 5.06	-- , 101.2
Urine 2	-- , 5.00	-- , 5.30	-- , 106.0
Serum	4.50, --	4.29, --	95.3 , --
Dopamine hydrochloride injection	4.00,1.00	3.70,1.03	92.5,103.0

Table S2 Summary of various nanomaterial-based electrochemical sensors for DA and UA in presence of Ascorbic acid

		DA	UA	DA	UA	
1	Pretreated pencil graphite	0.15-15μM	0.3-150μM	0.033μM	0.12μM	1
2	Poly(acrylic acid)- multiwalled carbon-nanotube composite-covered glassy-carbon	40nM-3μM	0.3μM-10μM	20nM	110nM	2
3	Multi-walled carbon nanotube-chitosan/poly(amidoamine)/DNA nanocomposite modified gold electrode	0.2-10μM & 10-100μM	0.5-100μM	0.03μM	0.07μM	3
4	Pt/reduced graphene oxide(Pt/RGO) modified glassy carbon electrode	10-170μM	10-130μM	0.25μM	0.45μM	4
5	Gold nanoparticle/choline (GNP/Ch) coated glassy carbon electrode	-	-	-	-	5
6	2,2'-azino-bis(3-ethylbenzthiazoline-6-sulfonic acid)(ABTS)-immobilized carbon nanotube (CNT) electrode	0.90-10μM & 1.87-20μM	2.16-240μM & 3.07-400μM	-	-	6
7	Electrochemically preanodized clay-modified electrodes	0-6μM	0.5-10 & 10-100μM	2.7nM	0.2μM	7
8	DNA/Poly(<i>p</i> -aminobenzensulfonic acid) composite bi-layer modified glassy	0.19–13 μM	0.4–23 μM	88 nM	0.19 μM	8

	carbon electrode					
9	RNA modified electrode	0.37 to 36 µM	0.74 to 73 µM	0.2 µM	0.36 µM	9
10	poly(orthanilic acid)- multiwalled carbon nanotubes composite film-modified glassy carbon electrode	9–48 µM	6–55 µM	0.21 µM	0.44 µM	10
11	multi-walled carbon nanotubes with methylene blue composite film-modified electrode	0.4–10.0 µM	2.0–20.0 and 20.0–200.0 µM	0.2 µM	1.0 µM	11
12	Glassy carbon electrode modified with poly(dibromofluorescein)	0.2 to 200 µM L ⁻¹	1.0 to 250 µM L ⁻¹	0.03 µM L ⁻¹	0.2 µM L ⁻¹	12
13	functionalized ordered mesoporous carbon/ionic liquid gel modified electrode	0.1 to 500 µM	0.1 to 100 µM	4.1nM	2.5 nM	13
14	Ionic Liquid Functionalized Graphene-Based electrode	1–400 µM	1–600 µM	0.679 µM	0.323 µM	14
15	Indole-3-Carboxaldehyde Modified Glassy Carbon Electrode	10 µM -100 µM	10 µM -100 µM	1.70 µM	4.99 µM	15
16	DNA/Pt Nanocluster Modified Electrode	1.1×10^{-7} to 3.8×10^{-5} M L ⁻¹	3.0×10^{-7} to 5.7×10^{-5} M L ⁻¹	3.6×10^{-8} M L ⁻¹	1.0×10^{-7} M L ⁻¹	16

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