

## *Supporting Information*

# **Multi-stimuli Responsive Nanoparticulate SN38 Prodrug for Cancer Chemotherapy**

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## **Methods**

### ***High performance liquid chromatography (HPLC)***

A Waters HPLC system consisting of a 1525 binary HPLC Pump, a 2475 multi  $\lambda$  Fluorescence Detector, a 2998 Photodiode Array Detector and a SunFire™ C18 (4.6×250 mm, 5  $\mu$ m) column. Data were acquired and analyzed using Breeze software. The mobile phase was a gradient of 60-90% of methanol/water solution at a total flow rate of 0.7 mL min<sup>-1</sup>. 20  $\mu$ L sample was injected into the column at 35°C. The UV absorption from 200 nm to 400 nm of the elution was recorded for analysis, while the fluorescence detectors were set as 360 nm for excitation and 400-700 nm for emission, respectively. The linear calibration curves of SN38 were also constructed using the peak areas by linear regression analysis.

### ***Size and size distribution measurements***

The size and size distribution of the nanoparticles were measured using a Zetasizer Nano-ZS (Malvern Instruments, UK) with a 633 nm laser light at a scattering angle of 173°. The disposable sizing cuvettes were used for measurements. The results were processed with Dispersion Technology Software version 5.1.

### ***Transmission electron microscope (TEM) observation***

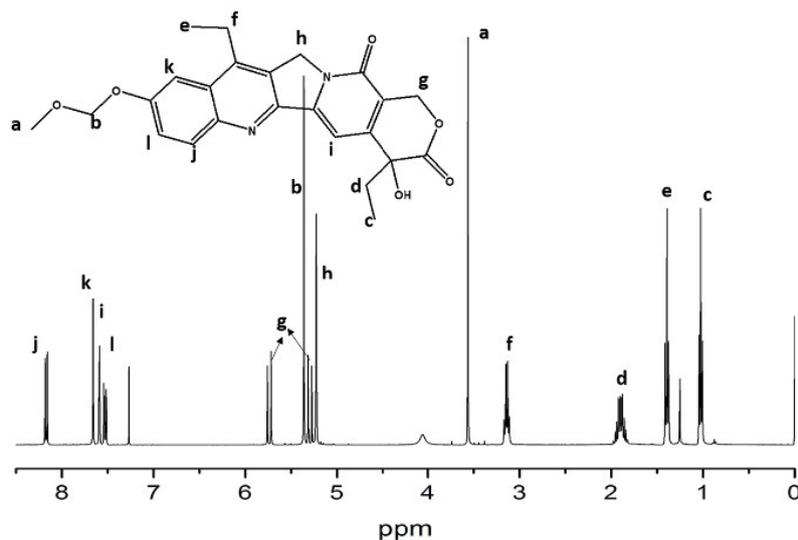
PEG-S-S-SN38 nanoparticles were dissolved in water at a concentration of 0.5 mg mL<sup>-1</sup>. Then a copper grid covered with a nitrocellulose membrane was dipped into the solution and air-dried. Observation was carried out on a Hitachi (H-7000) TEM.

### ***Determination of biodistribution***

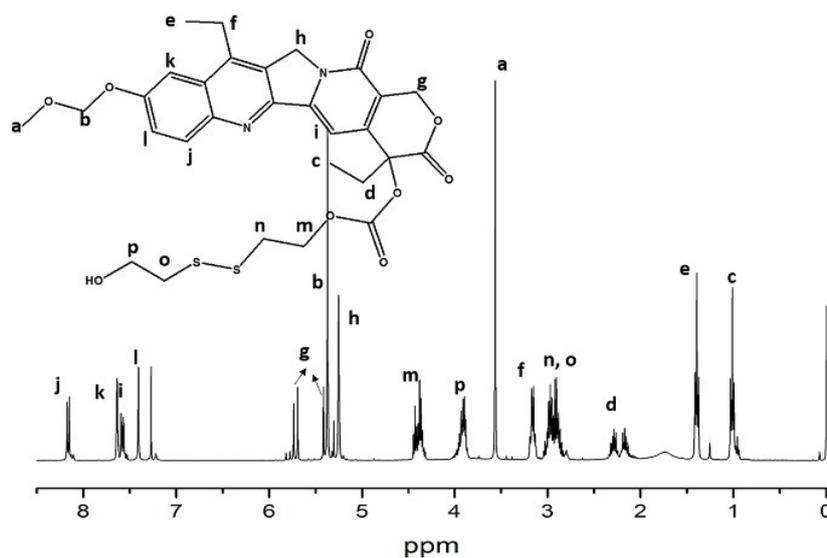
To predict the possible toxicity to normal organs or tissues and evaluate how the nanoparticles

were cleared, we further investigated the biodistribution of PEG-S-S-SN38 nanoparticles. Nine female nude mice were implanted with  $1 \times 10^6$  BCap37 cells on their right flanks subcutaneously. When the tumors grew up to a mean volume of around  $90 \text{ mm}^3$ , the tumor-bearing mice were injected *via* tail vein with PEG-S-S-SN38 nanoparticles at a dosage equivalent to  $10 \text{ mg kg}^{-1}$  SN38. Then mice were sacrificed at 0.5 h, 2 h and 10 h after treatments, respectively (n=3).  $50 \text{ }\mu\text{L}$  orbital bleeds were collected into tubes, and mixed gently with  $50 \text{ }\mu\text{L}$  0.1 N NaOH. Major organs including heart, kidney, spleen, lung, liver and tumors were excised and washed with 0.9% saline before weighed. All the tissues were cut into small pieces and homogenized. 2 mL PBS solution was added into the sample of liver, then mixed with  $200 \text{ }\mu\text{L}$  0.1 N NaOH. Other tissues were immersed in  $200 \text{ }\mu\text{L}$  PBS solution respectively, then  $100 \text{ }\mu\text{L}$  0.1 N NaOH was added. SN38 in liver sample was extracted with 6 mL acetonitrile, while those in bloods and other tissues were extracted with 1 mL acetonitrile. The obtained mixture was centrifuged at  $14,000 \times g$  for 10 min and  $100 \text{ }\mu\text{L}$  of supernatant was transferred to an eppendorf tube. Then  $100 \text{ }\mu\text{L}$  0.1 N HCl was added, and the solution was centrifuged at  $14,000 \times g$  for 5 min to obtain the supernatant. Finally,  $20 \text{ }\mu\text{L}$  of the supernatant was injected for HPLC test. The drug concentrations were determined according to the linear calibration curves.

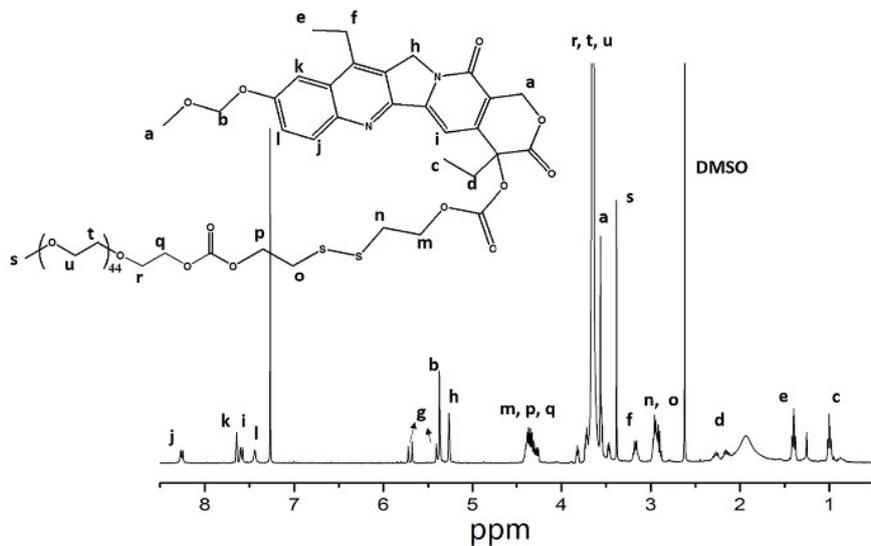
## Figures



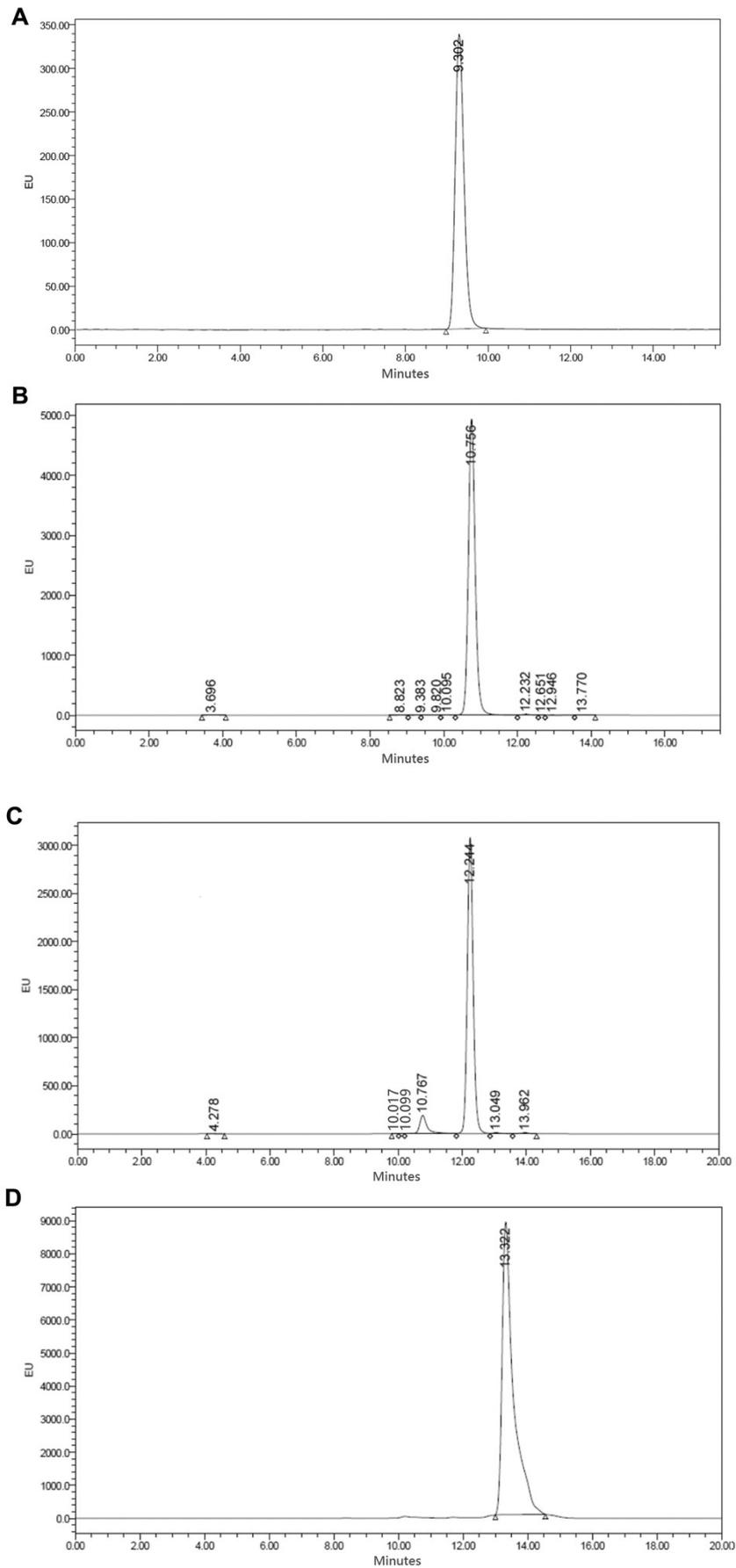
**Figure S1.**  $^1\text{H-NMR}$  spectrum of MOM-SN38 (400 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.16 (d,  $J=9.2$  Hz, 1H), 7.66 (s, 1H), 7.59 (d,  $J=2.8$  Hz, 1H), 7.54 (q,  $J_1=9.2$  Hz,  $J_2=2.4$  Hz, 1H), 5.72 (d,  $J=16.4$  Hz, 1H), 5.36 (s, 2H), 5.32 (d,  $J=16.4$  Hz, 1H), 5.23 (s, 2H), 3.56 (s, 3H), 3.14 (q,  $J=7.6$  Hz, 2H), 1.9 (m, 2H), 1.39 (t,  $J=7.6$  Hz, 3H), 1.02 (t,  $J=7.6$  Hz, 3H).



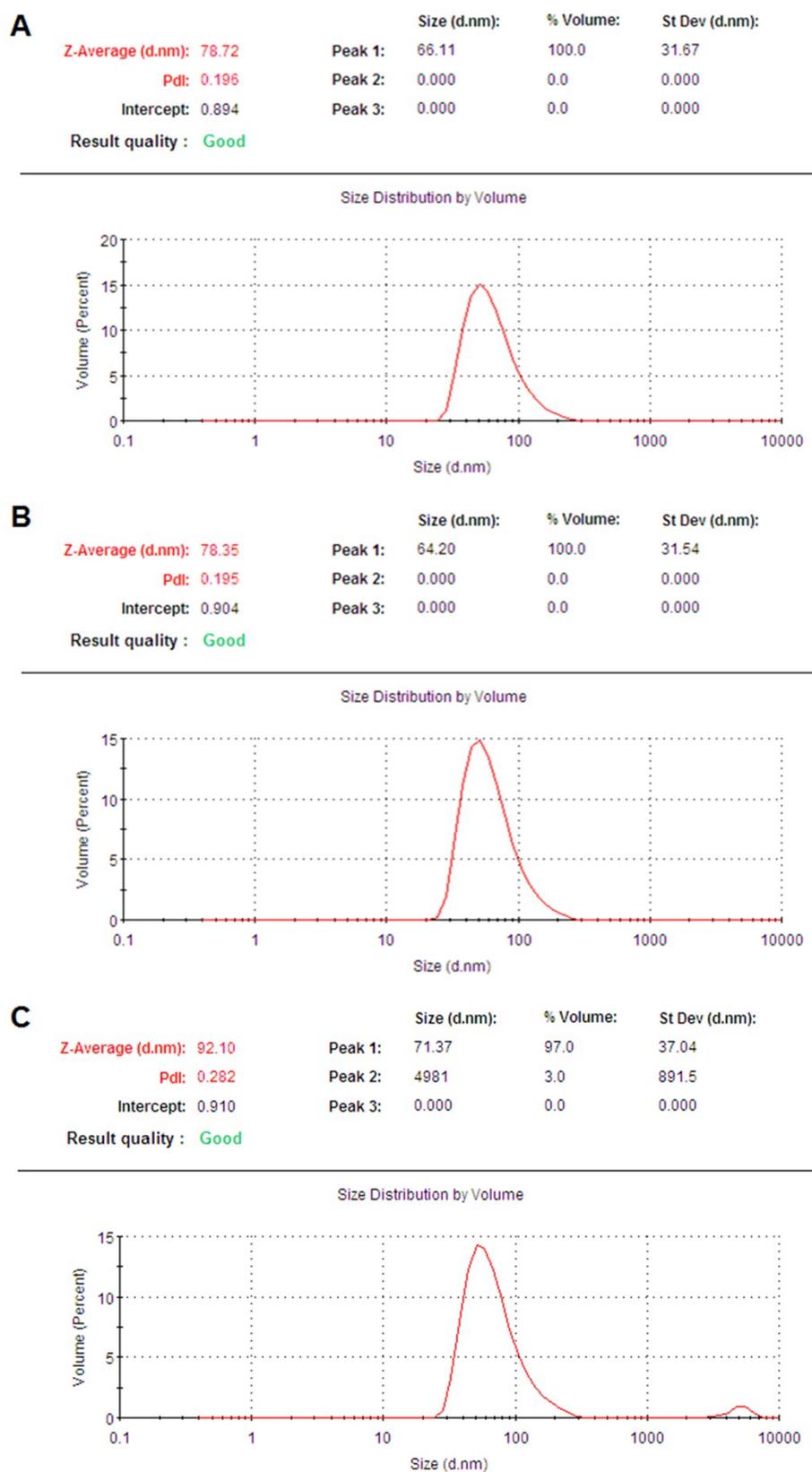
**Figure S2.**  $^1\text{H-NMR}$  spectrum of MOM-SN38-S-S-OH (400 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.15 (d,  $J=9.2$  Hz, 1H), 7.64 (d,  $J=2.8$  Hz, 1H), 7.59 (q,  $J_1=9.2$  Hz,  $J_2=2.4$  Hz, 1H), 7.4 (s, 1H), 5.7 (d,  $J=16.4$  Hz, 1H), 5.41 (d,  $J=16.4$  Hz, 1H), 5.37 (s, 2H), 5.25 (s, 2H), 4.37 (m, 2H), 3.9 (m, 2H), 3.56 (s, 3H), 3.17 (q,  $J=7.6$  Hz, 2H), 2.96 (m, 4H), 2.28 (m, 2H), 1.39 (t,  $J=7.6$  Hz, 3H), 1.01 (t,  $J=7.6$  Hz, 3H).



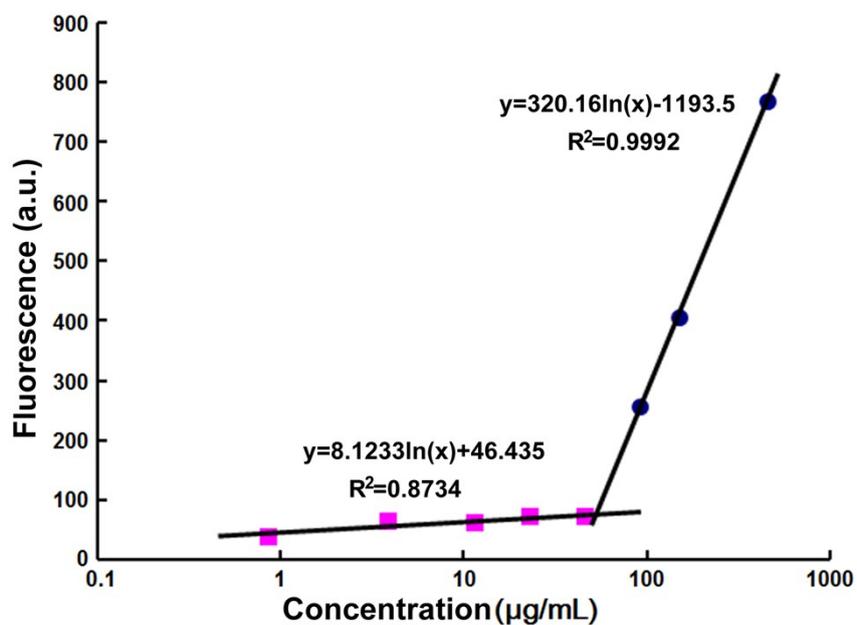
**Figure S3.**  $^1\text{H-NMR}$  spectrum of PEG-S-S-SN38 (400 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.25 (d,  $J=9.2$  Hz, 1H), 7.64 (d,  $J=2.8$  Hz, 1H), 7.59 (q,  $J_1=9.2$  Hz,  $J_2=2.4$  Hz, 1H), 7.44 (s, 1H), 5.7 (d,  $J=16.4$  Hz, 1H), 5.41 (d,  $J=16.4$  Hz, 1H), 5.37 (s, 2H), 5.26 (s, 2H), 4.36 (m, 6H), 3.41-3.85 (m, 178H), 3.38 (s, 3H), 3.17 (q,  $J=7.6$  Hz, 2H), 2.94 (m, 4H), 2.25 (m, 2H), 1.39 (t,  $J=7.6$  Hz, 3H), 1.01 (t,  $J=7.6$  Hz, 3H).



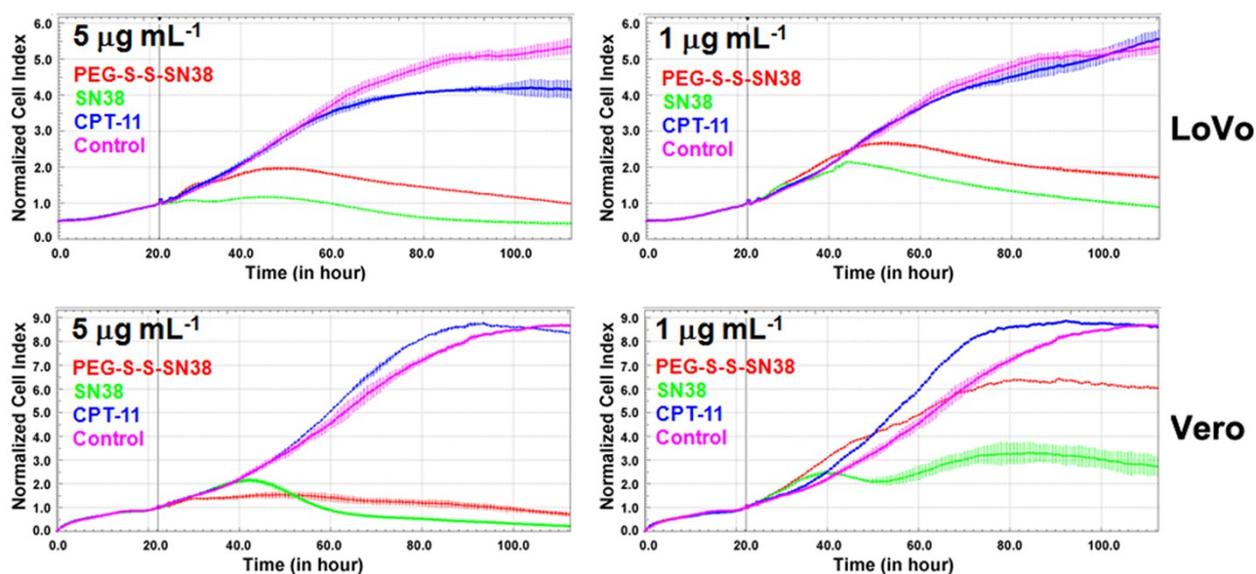
**Figure S4.** The HPLC traces of SN38 (A), MOM-SN38 (B), MOM-SN38-S-S-OH (C) and PEG-S-S-SN38 (D).



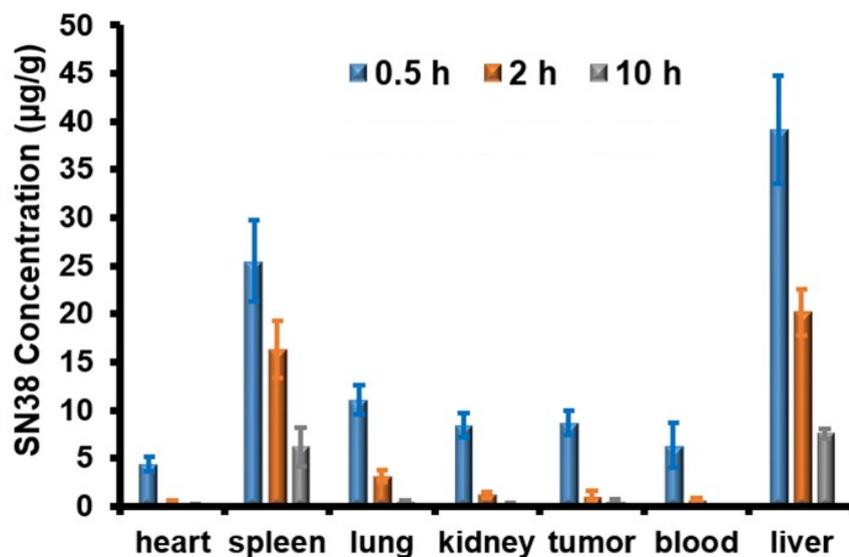
**Figure S5.** The stability of PEG-S-S-SN38 nanoparticles in 40% fetal bovine serum solution. Samples were analyzed by DLS after incubation at 37°C for 2.5 h (A), 6 h (B) and 32 h (C).



**Figure S6.** The fluorescence intensity of Nile red as a function of the PEG-S-S-SN38 concentration.



**Figure S7.** *In vitro* cytotoxicity of CPT-11, SN38 and PEG-S-S-SN38 NPs against LoVo and Vero cell lines evaluated by xCELLigence system real time cellular analysis (RTCA), at representative concentrations equivalent to 5 µg mL<sup>-1</sup> and 1 µg mL<sup>-1</sup> SN38, respectively.



**Figure S8.** The biodistribution of SN38 in various organs, blood and BCap37 xenograft tumors after tumor-bearing nude mice were administrated with PEG-S-S-SN38 nanoparticles for 0.5 h, 2 h and 10 h, respectively, at a dosage equivalent to 10 mg kg<sup>-1</sup> SN38.

**Table S1. IC<sub>50</sub> values in a panel of cancer cell lines as determined by MTT assays**

IC <sub>50</sub> (µg/mL)	BCap37	SKOV3	KB	KBv200	MCF-7	MCF-7/ADR
CPT-11	28.43	13.39	50.25	>100 *	53.75	>100 *
SN38	0.2707	0.4515	0.1863	0.1435	0.2873	5.49
PEG-S-S-SN38	0.9079	0.5041	0.4511	0.2147	1.598	58.21

\* Exact values could not be calculated based on the software used.