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Supporting information

One-step analysis of glucose and acetylcholine based on intrinsic peroxidase-like

activity of Ni/Co LDHs microspheres in water

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Figure S1. XRD pattern of Ni/Co LDHs.



Figure S2. SAED pattern of Ni/Co LDHs.



Figure S3. EDX spectrum of Ni/Co LDHs.



Figure S4. Nitrogen adsorption-desorption isotherm of Ni/Co LDHs.



Figure S5. The peroxidase-like activity of Ni/Co LDHs is dependent on temperature. The error bars represent the standard deviation of three measurements. The maximum point in each curve was set as 100%.



Figure S6. Steady-state kinetic assay and catalytic mechanism of Ni/Co LDHs (a–d). The velocity (v) of the reaction was measured using 0.1 mL 1.0 mg mL⁻¹ Ni/Co LDHs in 2.7 mL deionized water at 37 °C. The error bars represent the standard error derived from three repeated measurements. The concentration of ABTS was 0.53 mM and H_2O_2 concentration was varied (a). The concentration of H_2O_2 was 1.67 mM and ABTS concentration was varied (b). Double reciprocal plots of activity of Ni/Co LDHs with the concentration of one substrate (ABTS or H_2O_2) fixed and the other varied (c, d).



Figure S7. Determination of the selectivity of glucose detection (from left to right: blank, 0.5×10^{-3} mol L⁻¹ glucose, 3.3×10^{-3} mol L⁻¹ maltose, 3.3×10^{-3} mol L⁻¹ lactose, and 3.3×10^{-3} mol L⁻¹ fructose) at 417 nm ($\Delta A = A_{total} - A_{blank}$).



Figure S8. Selectivity analysis for ACh detection. (from left to right: $2.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ ACh}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Glycine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Tryptophan}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Tryptophan}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Tryptophan}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Tryptophan}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.10 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{L}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{C}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{C}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{C}^{-1} \text{ Cystine}$, $8.0 \times 10^{-4} \text{ mol } \text{C}^{-1} \text{ Cysti$

| Catalyst | Substance | <i>K_m</i> /mM | $V_{max}/10^{-8} \text{ M}\cdot\text{s}^{-1}$ |
|------------|-----------|--------------------------|---|
| | ABTS | 1.56 | - |
| IIIXi | H_2O_2 | 0.24 | - |
| | ABTS 3.43 | 3.43 | 3.29 |
| NI/C0 LDHS | H_2O_2 | 13.2 | 3.24 |

Table S1. Michaelis-Menten parameters of Ni/Co LDHs.

| Sample | Spiked (uM) | The proposed method | | The GOD- | The GOD-PAP method | |
|---------------|-------------|---------------------|--------------|------------|--------------------|--|
| Sumpro | 2p (p) | Found (µM) | Recovery (%) | Found (µM) | Recovery (%) | |
| Human serum 1 | 5.0 | 4.8 | 96.0 | 5.1 | 102.0 | |
| | 25.0 | 24.6 | 98.4 | 24.9 | 99.6 | |
| | 50.0 | 51.4 | 103.0 | 50.8 | 101.6 | |
| Human serum 2 | 5.0 | 5.3 | 106.0 | 5.1 | 102.0 | |
| | 25.0 | 26.1 | 104.4 | 25.4 | 101.6 | |
| | 50.0 | 48.4 | 96.8 | 49.1 | 98.2 | |
| Human serum 3 | 5.0 | 4.7 | 94.0 | 5.2 | 104.0 | |
| | 25.0 | 24.2 | 96.8 | 24.8 | 99.2 | |
| | 50.0 | 52.4 | 104.8 | 51.3 | 102.6 | |

 Table S2. Determination of glucose in real samples.

| Milk sample | Added (µM) | Found (µM) | Recovery (%) |
|-------------|------------|------------|--------------|
| 1 | 25.0 | 23.8±0.82 | 95.2 |
| 2 | 75.0 | 78.6±0.75 | 104.8 |
| 3 | 125.0 | 112.8±0.61 | 90.2 |

 Table S3. Determination of ACh in milk sample.

References

1. H. Gallati, Clinical Chemistry and Laboratory Medicine, 1979, 17, 1-8.