

## Supporting Information

### **Cobalt Oxyhydroxide Assembled Upconversion Nanoparticles as a Ratiometric Nanoprobe for Ascorbic Acid Detecting and Imaging**

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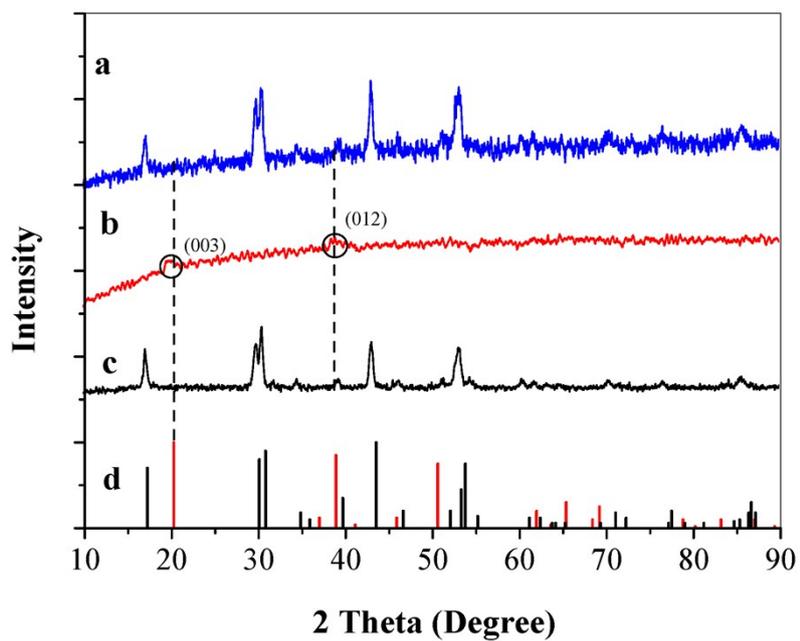
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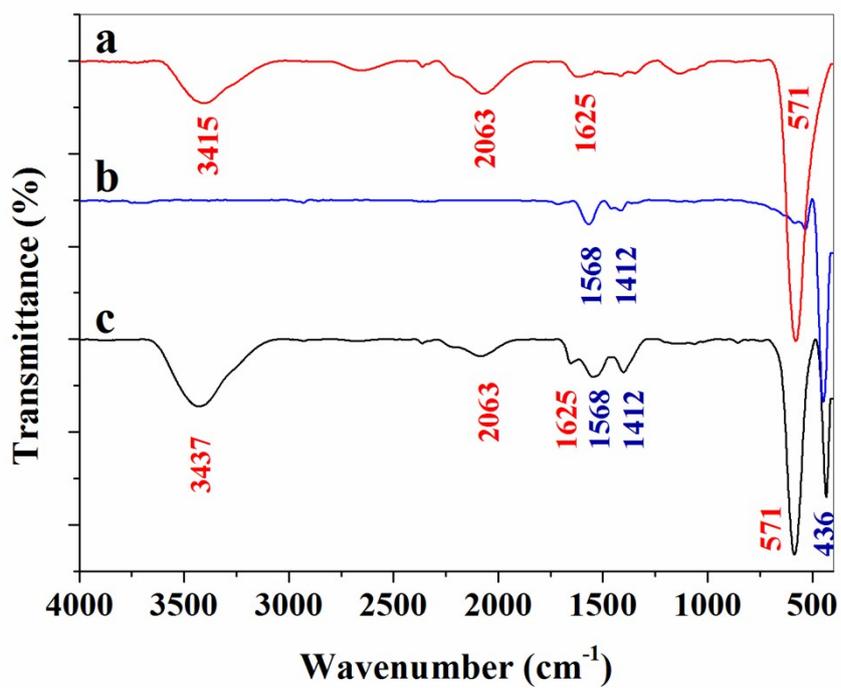
### ***Materials and instruments***

Yb<sub>2</sub>O<sub>3</sub> (99.9%), Tm<sub>2</sub>O<sub>3</sub> (99.9%), Y<sub>2</sub>O<sub>3</sub> (99.9%), Gd<sub>2</sub>O<sub>3</sub> (99.9%), were obtained from Tianjin Guangfu Fine Chemical Research Institute. 1-octadecene (90%), oleic acid (90%), and YCl<sub>3</sub>·6H<sub>2</sub>O were purchased from Alfa Aesar. The other reagents used without further purification were received from commercial sources. Thermo gravimetric analysis (TGA) was conducted on Netzsch STA449F3 Jupiter® thermal analyzer up to 800 °C at a heating rate of 10 °C·min<sup>-1</sup> in N<sub>2</sub> atmosphere. Transmission electron microscopy (TEM) was obtained on a Tecnai-G2-F30 (300 kV). X-ray diffraction (XRD) pattern of the synthesized products were recorded with a Rigaku D/MAX 2400 X-ray diffractometer using Cu K $\alpha$  radiation ( $\lambda = 0.154056 \text{ \AA}$ ). Fluorescence spectra were recorded on a Hitachi F-7000 spectrofluorometer equipped with a 0-5 W adjustable CW laser (980 nm, Connect Fiber Optics, China) as the excitation source. The lifetimes were determined by FLS920 of Edinburgh Instruments. Cyclic voltammograms were identified by a CHI660B electrochemical Workstation (CH Instruments, Inc., USA) using platinum (Pt) wire and Ag/AgCl (*aq.* saturated KCl) as the counter and reference electrodes, respectively. Glassy carbon electrode (3 mm) was used as the working electrode with scan rate of 5 mV s<sup>-1</sup>. All pH measurements were executed with a pH-10C digital pH meter. HeLa cells were provided by the Institute of Biochemistry and Cell Biology (China) were cultured for 48 h in culture in DMEM (Dulbecco's Modified Eagle's Medium, High Glucose) with 10% fetal bovine serum (FBS) in an atmosphere of 5% CO<sub>2</sub> and 95% at 37 °C with a humidified incubator.

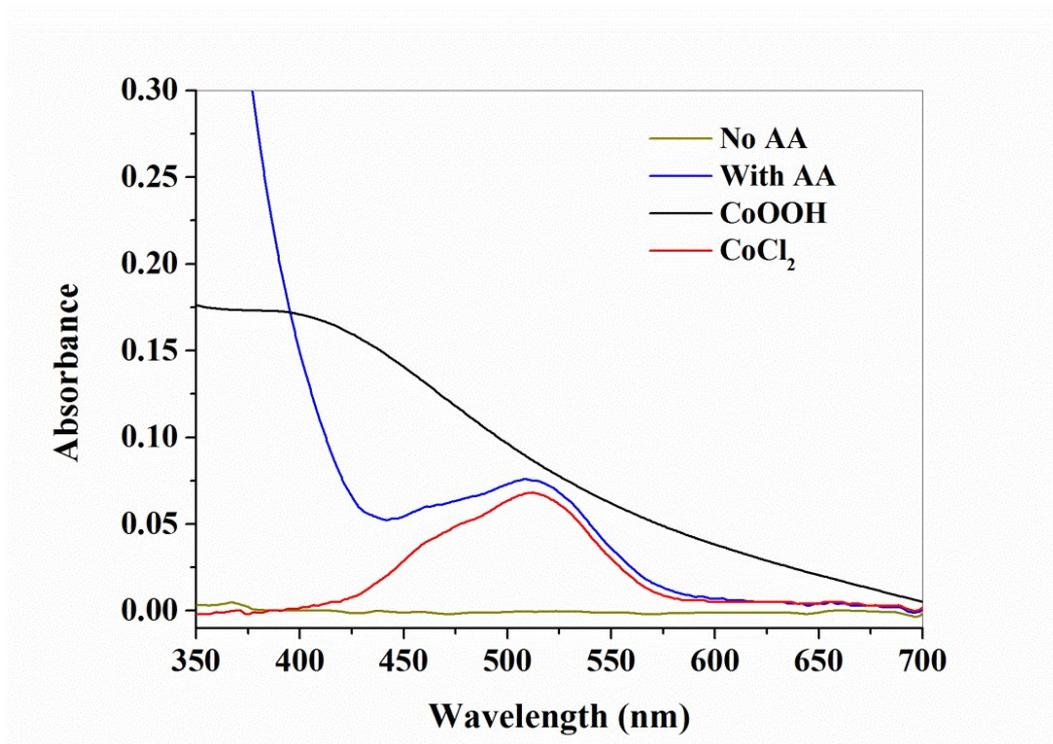
*Supporting Figures:*



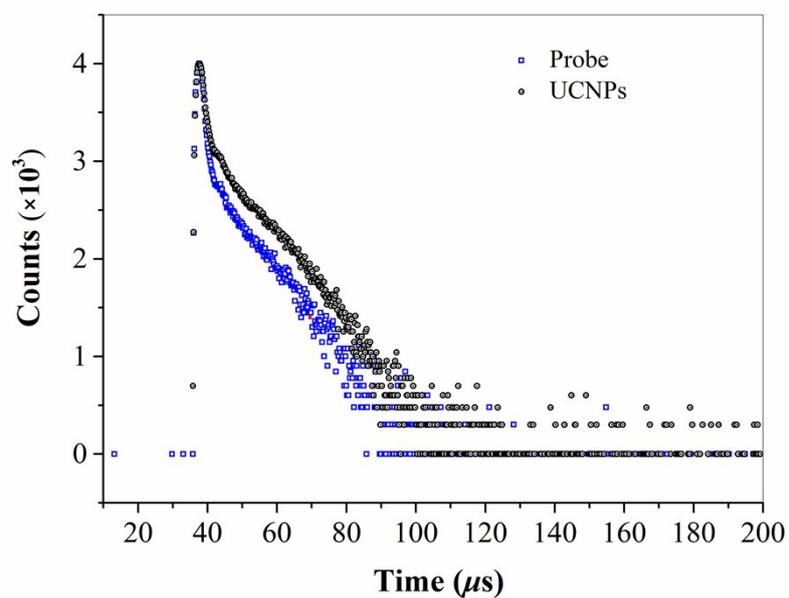
**Fig. S1.** XRD patterns of ligand-free core-shell UCNPs (a), CoOOH (b), and CoOOH-modified UCNPs (c), standard XRD patterns for CoOOH nanocrystals (JCPDS, Card No.07-0169) (red) and NaYF<sub>4</sub> UCNPs (JCPDS, Card No.16-0334) (black) (d).



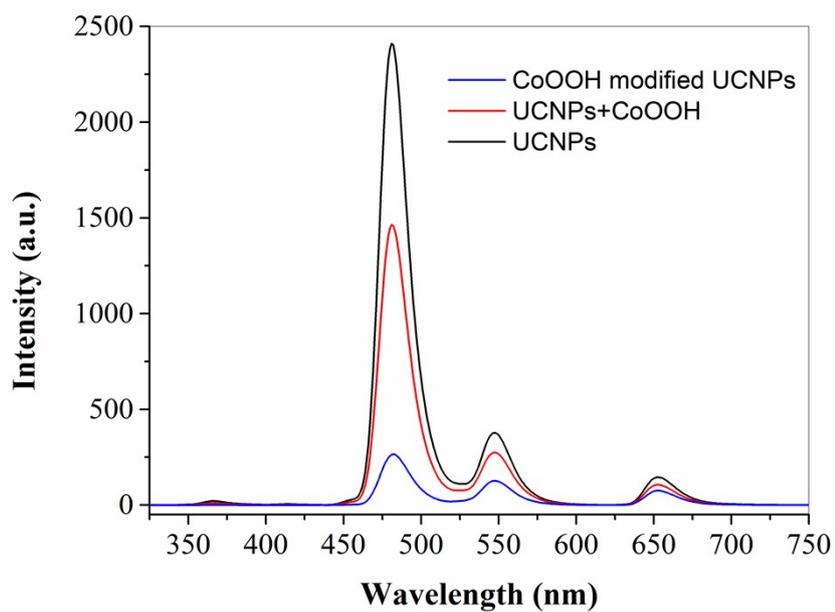
**Fig. S2.** FT-IR spectra of CoOOH (a), ligand-free core-shell UCNPs (b), and CoOOH modified UCNPs (c).



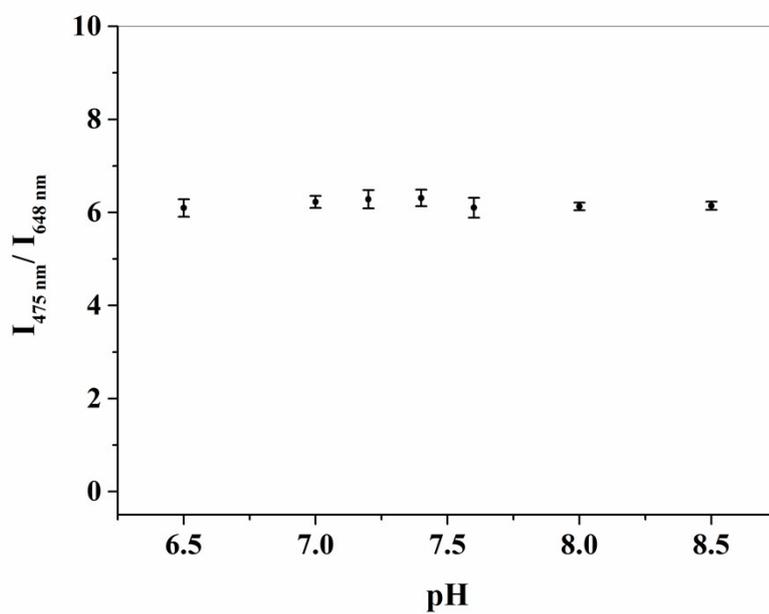
**Fig. S3.** UV-vis absorption of aqueous solution of CoCl<sub>2</sub> (red line), CoOOH solution (black line), supernates of CoOOH-modified UCNPs solution (brown line) and CoOOH-modified UCNPs solution with AA (blue line).



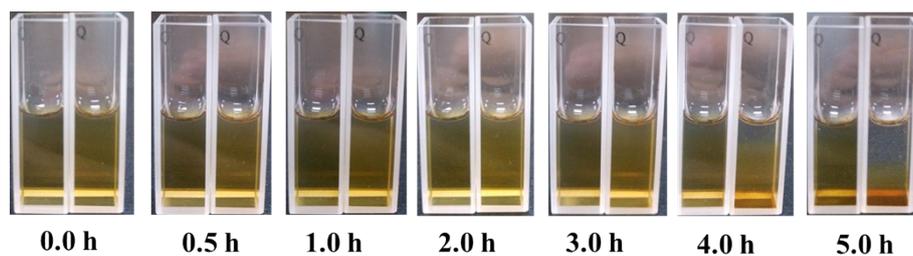
**Fig. S4.** The luminescence (475 nm) lifetime decay profile of bare UCNPs (black dots) and CoOOH-modified UCNPs (blue dots).



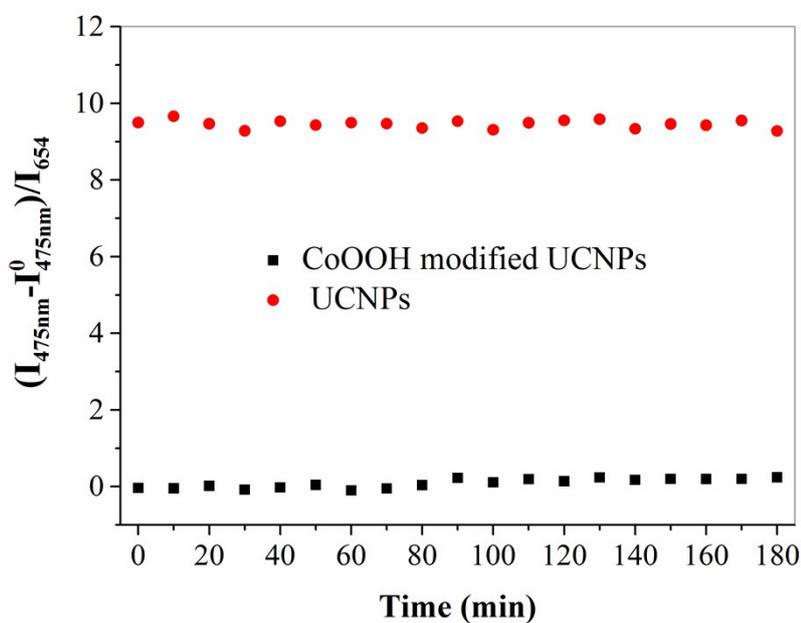
**Fig. S5.** UCL spectra of aqueous solutions of UCNPs (1mg/mL, black line), physical mixture of CoOOH (2 mmol) and UCNPs (blue line), and CoOOH-modified UCNPs (prepared with 2.0 mmol  $\text{CoCl}_2$ , red line).



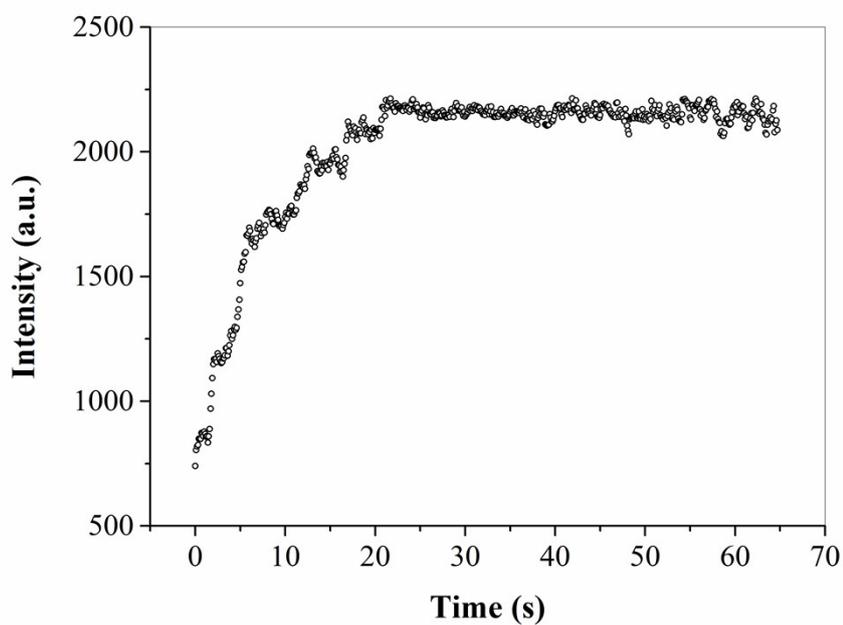
**Fig. S6.** UCL intensity of CoOOH modified UCNPs (1mg/mL) in different pH solutions in the absence (black dots) and presence (red dots) of AA (100  $\mu\text{M}$ ).



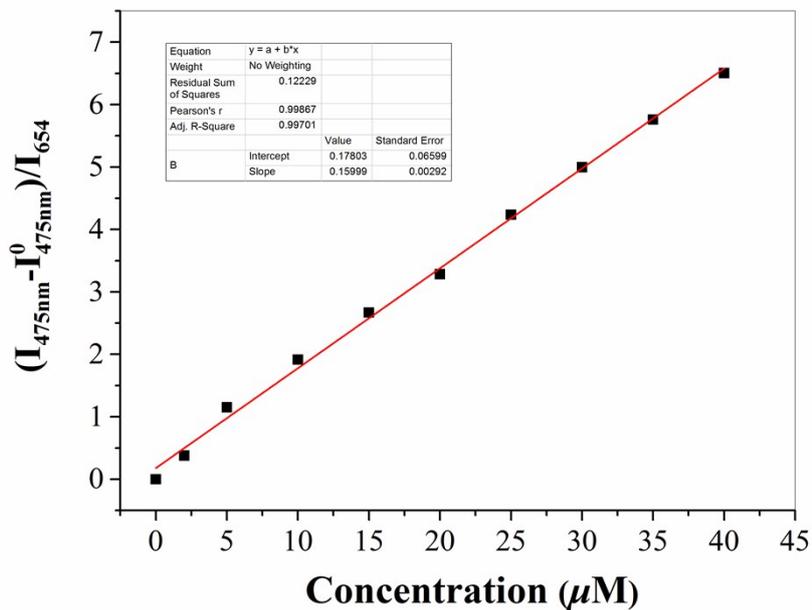
**Fig. S7.** Photograph changes of CoOOH-modified UCNPs (1 mg/mL) solution with (left) and without (right) 10% ethylene glycol (EG) over time. EG was also introduced as the dispersant into sensing system to improve the stability of a probe. Due to the small size of CoOOH-modified UCNPs, the improvement in our probe is limited. In this regard, EG has not been introduced in the sensing study of CoOOH-modified UCNPs nano-system.



**Fig. S8.** Time-dependent UCL intensity ratio changes of CoOOH-modified UCNPs (1 mg/mL) (black dots) and UCNPs (red dots) depending on time in HEPES (10 mM, pH = 7.4).



**Fig. S9.** Time-dependent UCL intensity (475 nm) changes of CoOOH-modified UCNPs (1 mg/mL) with AA (100  $\mu$ M)) in HEPES solution (10 mM, pH = 7.4).



**Fig. S10.** UCL intensity ratio  $I_{475}/I_{468}$  changes of CoOOH modified UCNPs as a function of AA concentration in HEPES (10 mM, pH = 7.4).

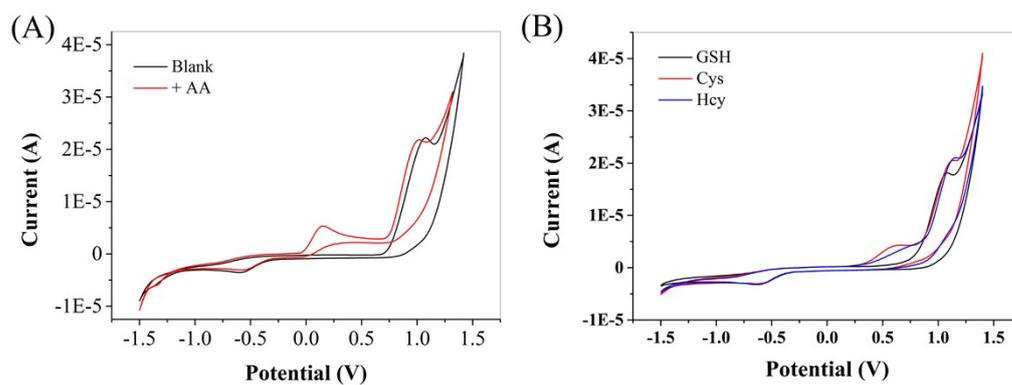


Fig. S11. Cyclic voltammograms of 1mM AA, GSH, Cys and Hcy in HEPES solution (10 mM, pH = 7.4).

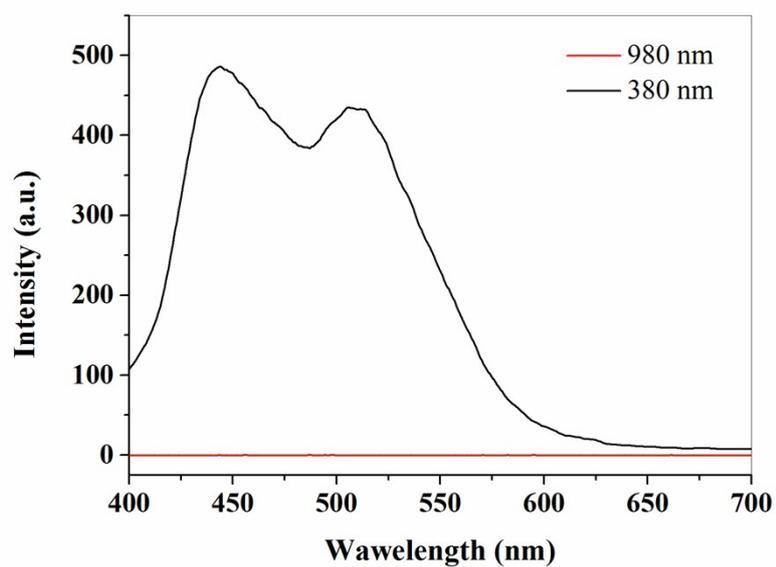


Fig. S12. UCL spectra of diluted (25 %) fetal bovine serum in the absence AA under excitation at 380 nm (black) and 980 nm (red).

# EDX

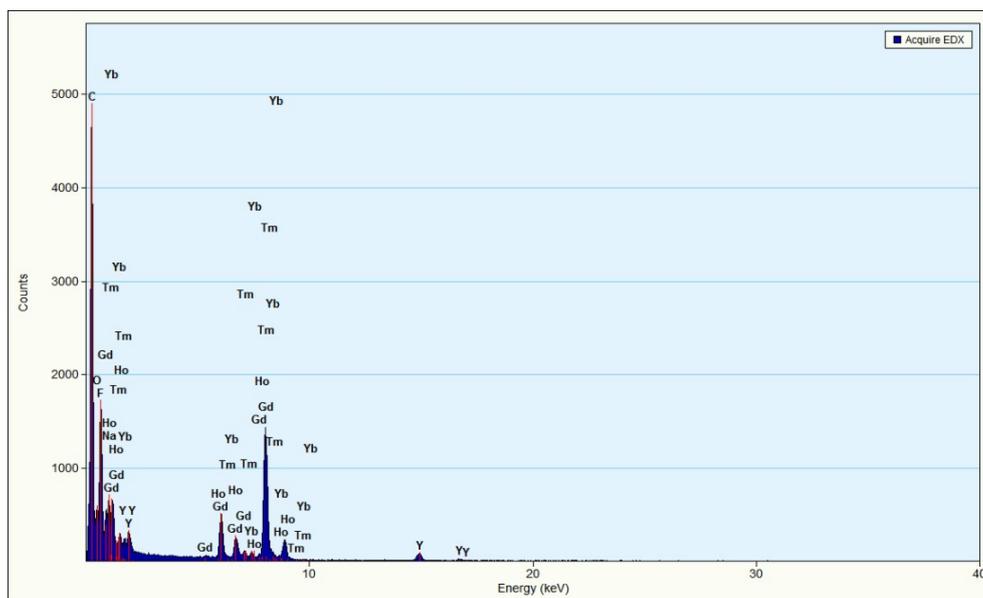


Fig. S13. EDX spectrum of UCNPs.

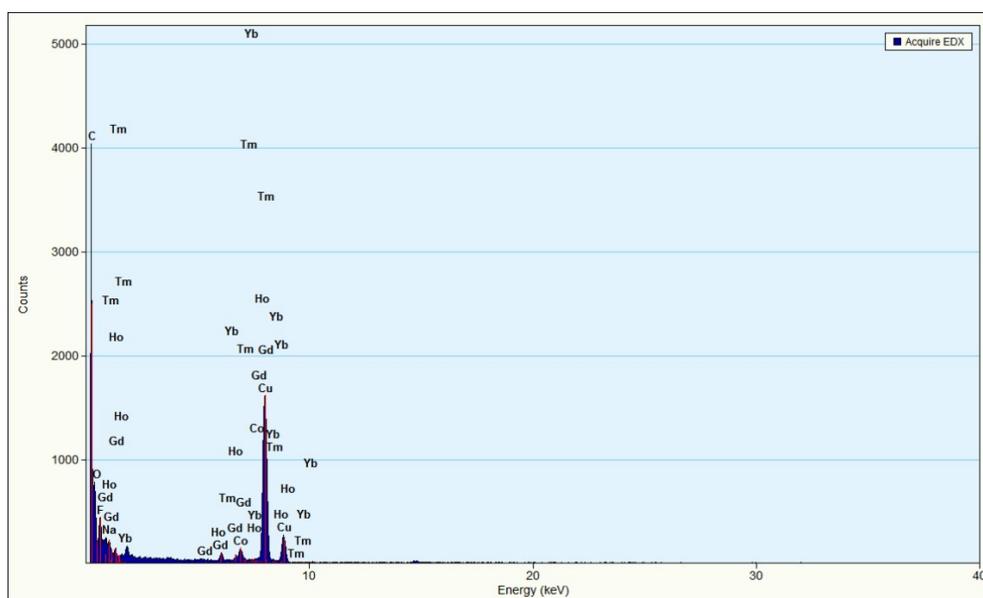


Fig. S14. EDX spectrum of CoOOH modified UCNPs.

**TG-DSC:**

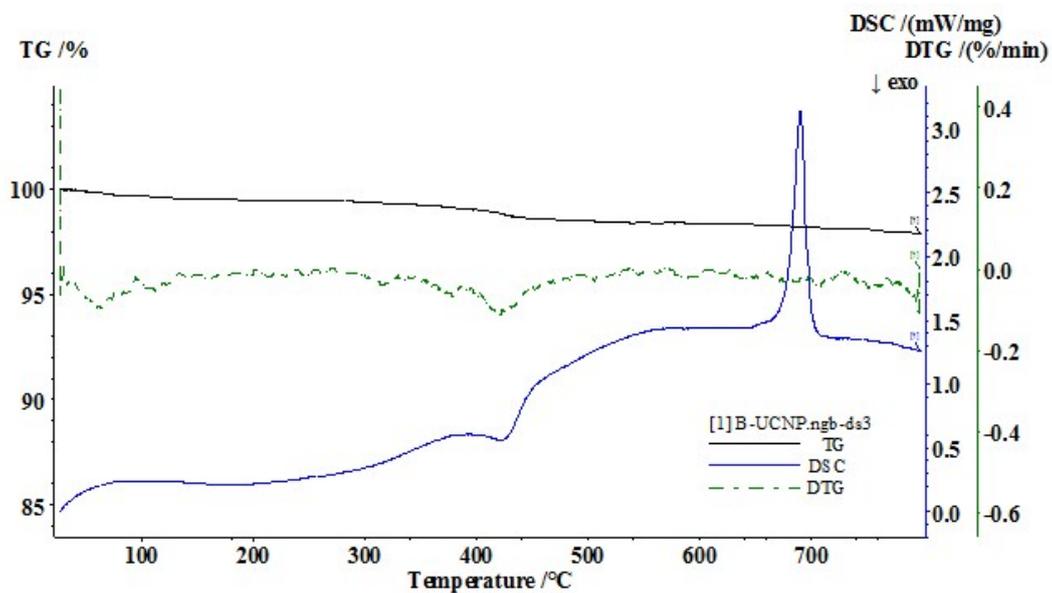


Fig. S15. TG-DSC result of ligand-free core-shell UCNPs.

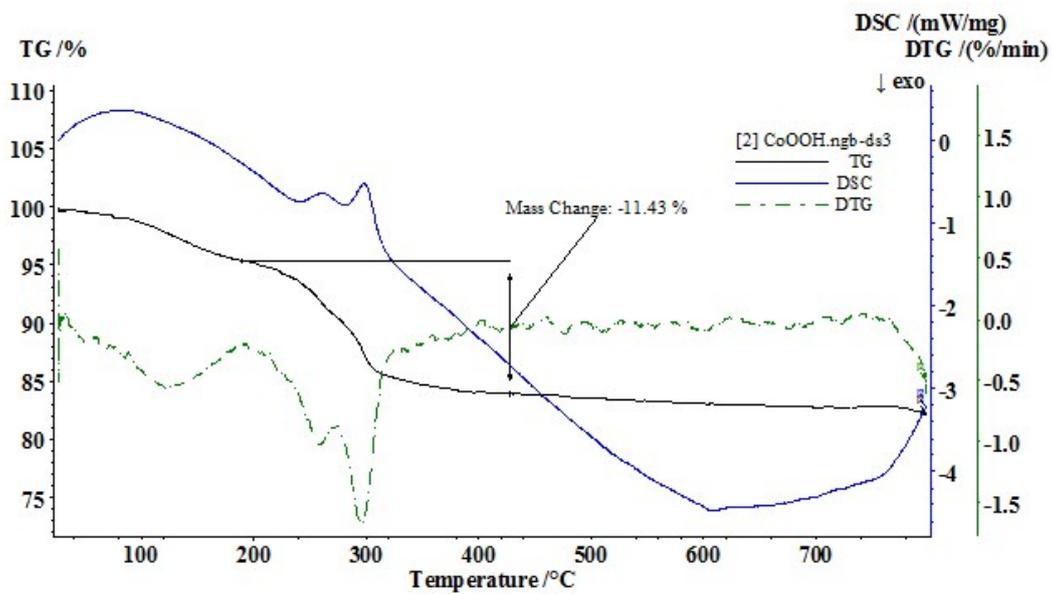


Fig. S16. TG-DSC result of CoOOH.

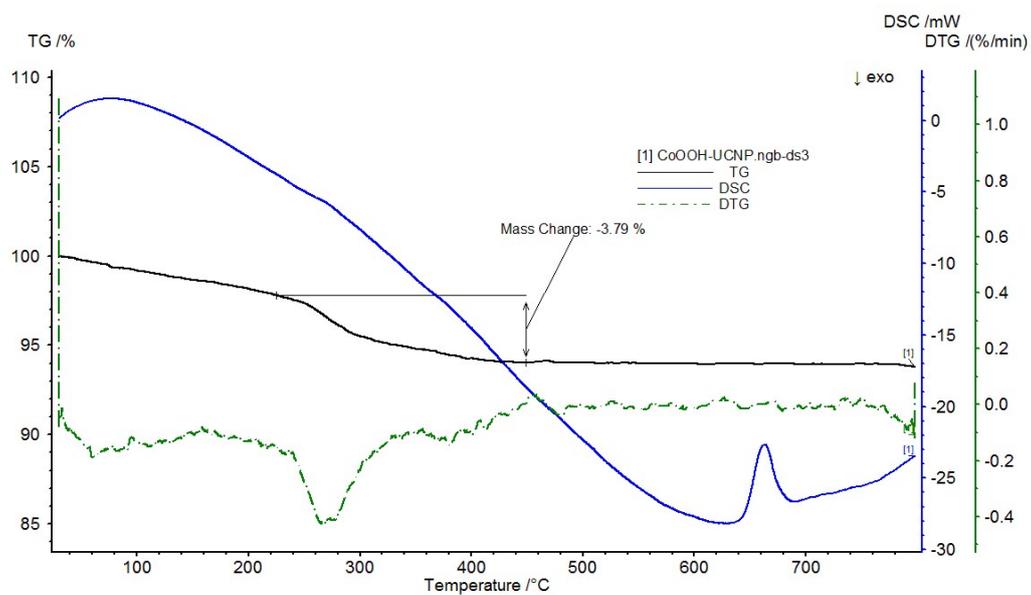


Fig. S17. TG-DSC result of CoOOH modified UCNPs.