

Supporting Information

Fabrication of Cell-Compatible Hyaluronan Hydrogels with a Wide Range of Biophysical Properties Through High Tyramine Functionalization

Claudia Loebel^{1,2,†}, Tino Stauber², Matteo D'Este¹, Mauro Alini¹, Marcy Zenobi-Wong², David Eglin^{1}*

¹AO Research Institute Davos, Clavadelerstrasse 8, Davos Platz, 7270, Switzerland

²ETH Zurich, Cartilage Engineering + Regeneration, Department of Health, Science and Technology, Otto-Stern-Weg 7, Zürich, 8093, Switzerland

† Current address: Department of Bioengineering, University of Pennsylvania, Philadelphia, Pennsylvania 19104, United States, loebelcl@seas.upenn.edu

*Corresponding author: AO Research Institute Davos, Clavadelerstrasse 8, Davos Platz, 7270, Switzerland, david.eglin@aofoundation.org

Synthesis of HA-Tyr conjugates by conventional carbodiimide chemistry

HA-Tyr was prepared by amidation of the carboxylic acid groups of HA with the amine groups of Tyr as described previous.¹ HANA (500 mg, 1.25 mmol carboxyl groups) was hydrated in MES Buffer (100 mM, pH 5.5, adjusted with NaOH 5 M) for 24 h at a final concentration of 1% (w/v). 1-Ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDC) and N-hydroxysuccinimide (NHS) (1/1 to HA) were added and stirred for 1 h at RT. Tyr (1/1 to HA) was dissolved in MES-buffer (100 mM, pH 5.5; 0.1% w/v) and added drop-wise to the HA-solution. The reaction was maintained at RT for 24 h under continuous stirring. Products were precipitated by 96% ethanol. For two-step HA-Tyr conjugation, the isolated HA-Tyr product was isolated and rehydrated in 100 mM MES-buffer (pH 5.5) and the conjugation repeated. Note, that the product after 48 h was highly viscous when dissolved in PBS (1% w/v). This may be caused by a high percentage of auto-crosslinked EDC-Tyr adducts, as shown by improved solubility in alkaline conditions (pH 8).

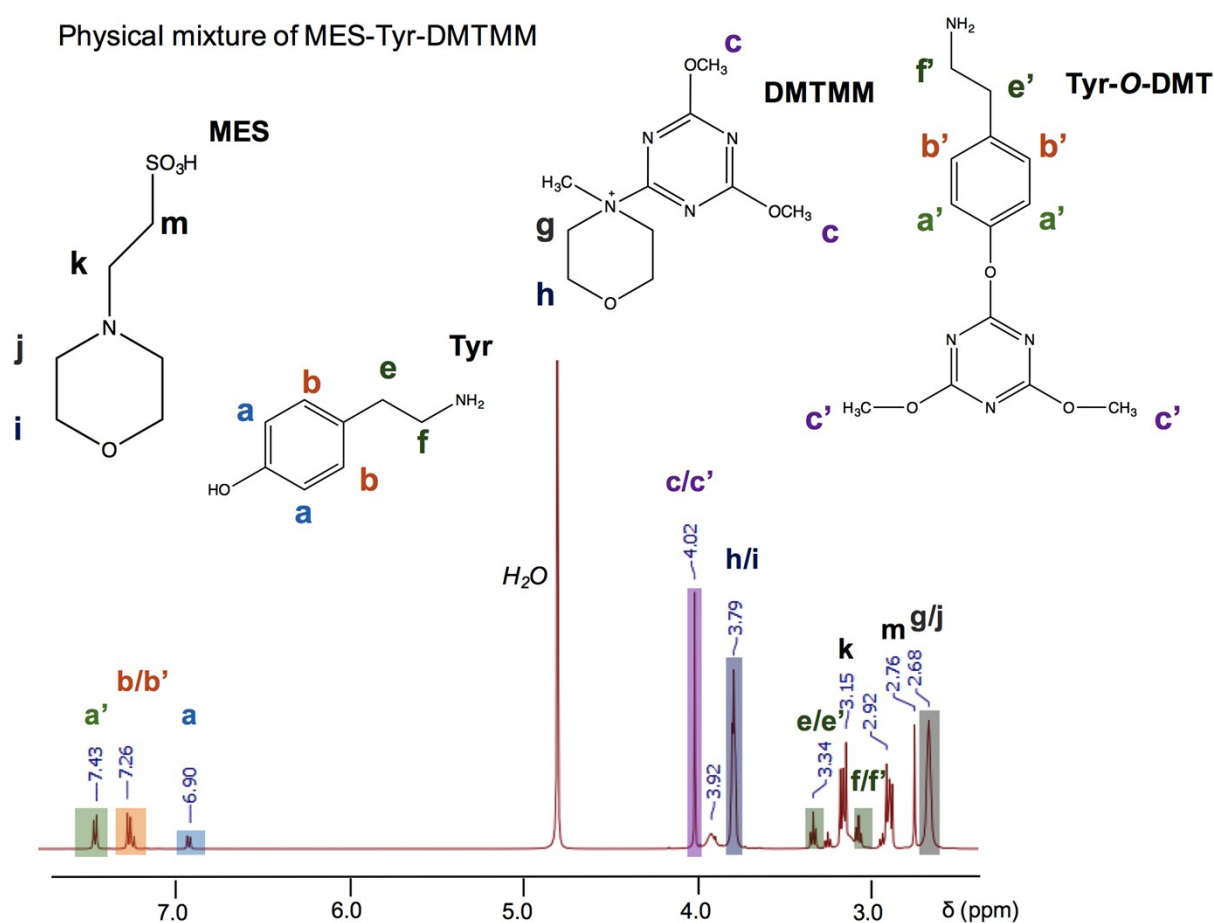


Figure S1. ¹H NMR spectrum of the physical mixture MES, DMTMM and Tyr (0.025 mmol each, without additional chemical modifications) in D₂O. Measurements were taken after 24 h incubation at RT.

HA-Tyr synthesized in 100 mM NaCl

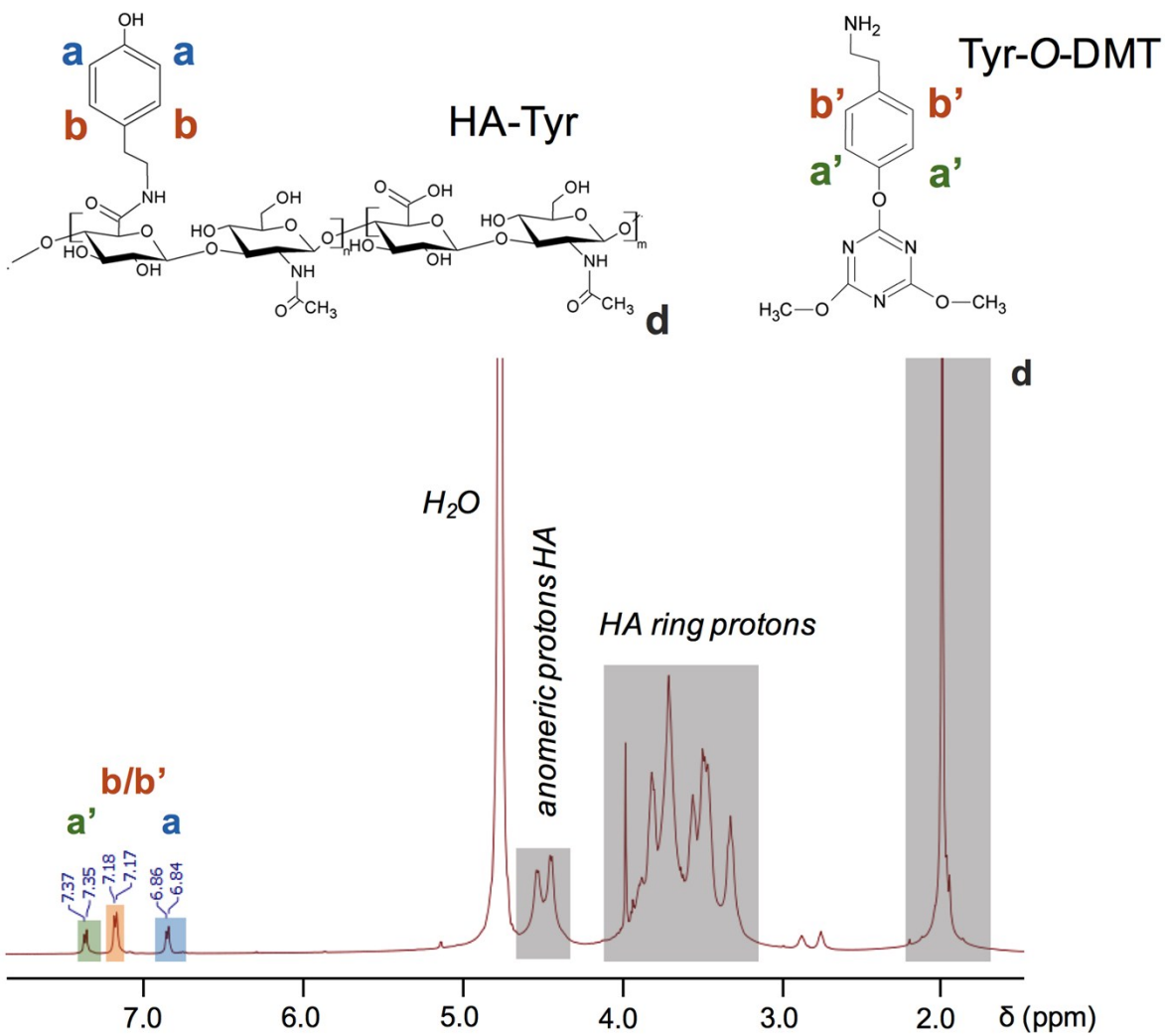


Figure S2. ¹H NMR spectrum of the conjugate HA-Tyr synthesized in 100 mM NaCl at 37 °C for 24 h.

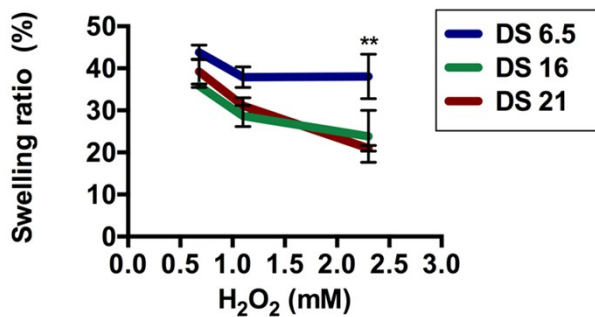


Figure S3. Swelling of HA-Tyr hydrogels with different DS as a function of H₂O₂ concentration. Hydrogels were formed with 0.5 U/ml HRP. Data shown as mean ± SD for n = 3 constructs, **p<0.01 for DS 6.5% compared to DS 16 and 21%.

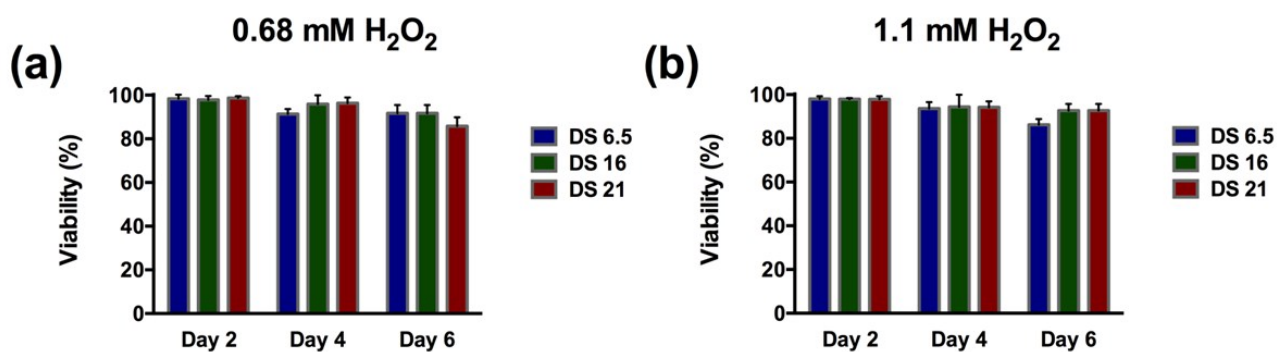


Figure S4. Quantification of cell viability of MSCs (5×10^6 /ml) encapsulated in HA-Tyr hydrogels crosslinked with (a) 0.68 mM H₂O₂ and (b) 1.1 mM H₂O₂, after 2, 4 and 6 days of culture in basal medium (DMEM plus 10% FBS), mean + SD of four randomly chosen locations per hydrogel. One representative donor shown with mean + SD of n = 4 randomly chosen locations within the constructs.

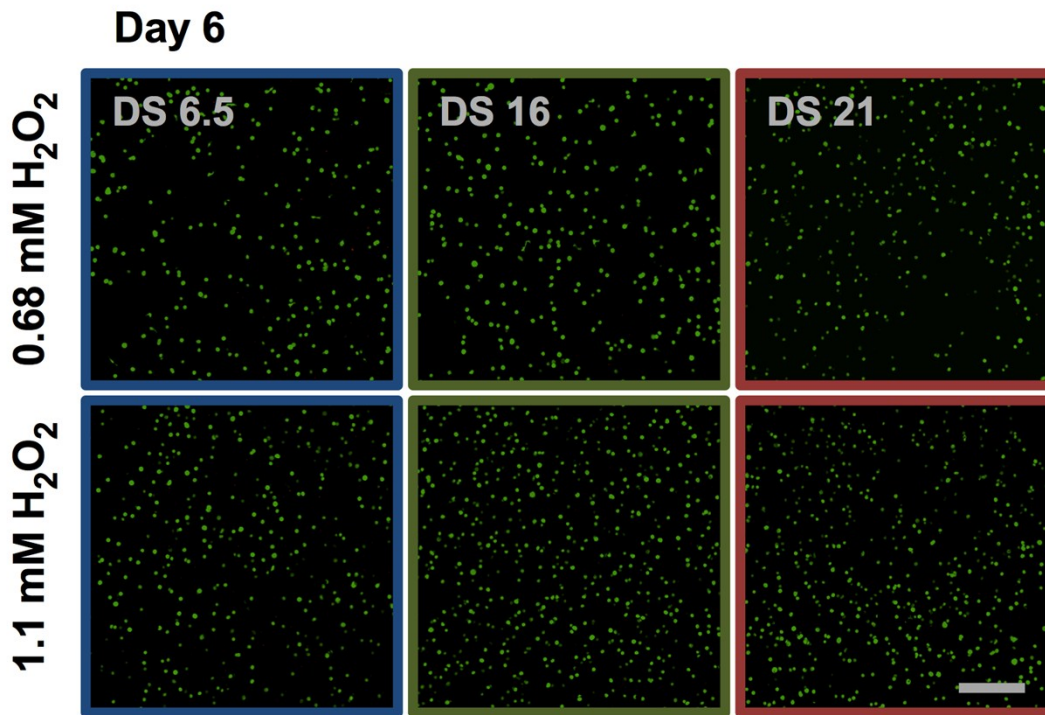


Figure S5. Representative images of hMSCs encapsulated in HA-Tyr hydrogels ($5 \times 10^6/\text{ml}$) crosslinked with 0.68 mM and 1.1 mM H₂O₂. Calcein (5 μM , green) /Ethidium homodimer (8 μM , red) staining after 6 days of culture. Images are shown as 3D rendered z-stacks with a thickness of 200 μm , Scale bar 200 μm .

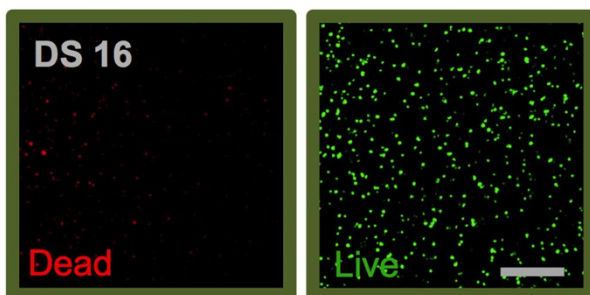


Figure S6. Representative images of encapsulated hMSCs in HA-Tyr (DS 16, 0.68 mM H₂O₂) after 6 days of culture without FBS and staining with Calcein (5 μM , green) /Ethidium homodimer (8 μM , red). Images are shown as 3D rendered z-stacks with a thickness of 200 μm . Scale bar 200 μm .

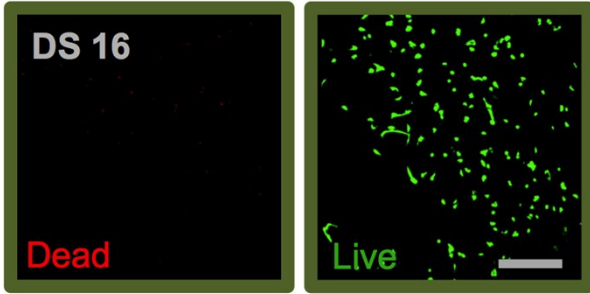


Figure S7. Representative images of encapsulated hMSCs in HA-Tyr *in-situ* modified with 500 μM RGD-Tyrosine (Genscript) after 6 days of culture and staining with Calcein (5 μM , green) /Ethidium homodimer (8 μM , red). Images are shown as 3D rendered z-stacks with a thickness of 200 μm . Scale bar 200 μm .

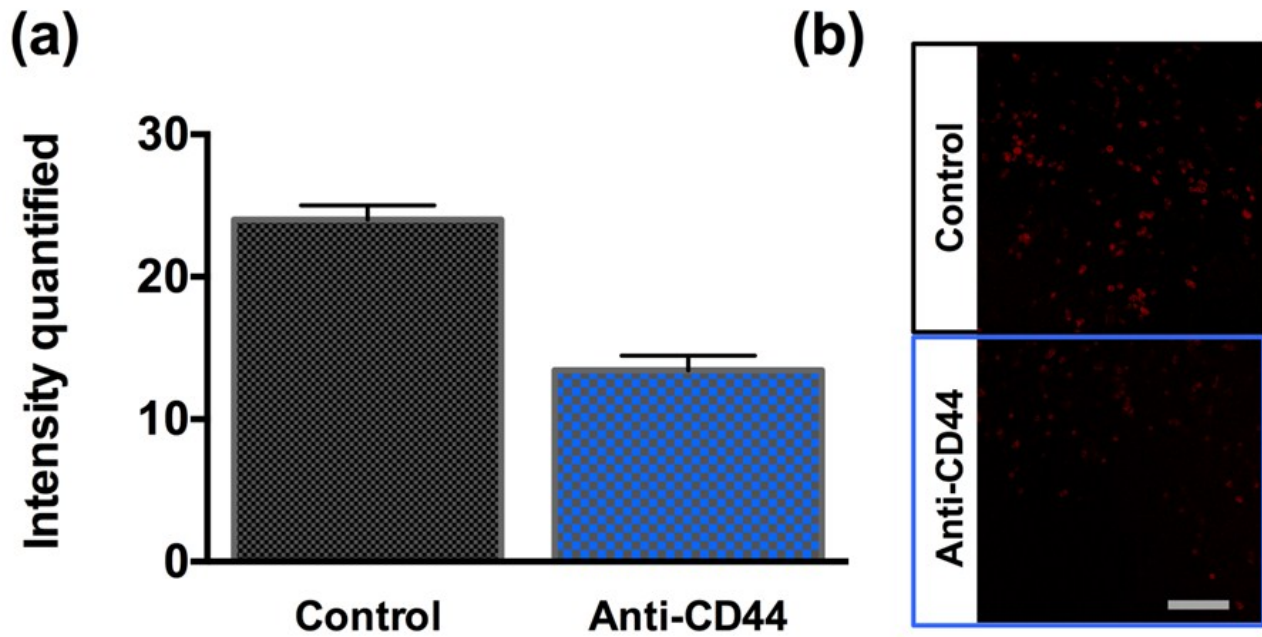


Figure S8. (a) Quantification of CD44 expression intensity and (b) representative images of hMCS after incubation with anti-CD44 antibody (abcam 10-44-2). 5×10^6 cells/ml, encapsulated in DS 16% HA-Tyr hydrogels crosslinked with 1 U/ml HRP and 0.68 mM H_2O_2 . Cell-laden hydrogels were fixed 30 min after encapsulation with 4% paraformaldehyde (overnight at 4 $^\circ\text{C}$) and washed three times prior incubation with anti-CD44-Allophycocyanin (APC) antibody (Miltenyi Biotec 130-095-177). Confocal images were taken using a Zeiss LSM510 (10x objective). Scale bar 200 μm .

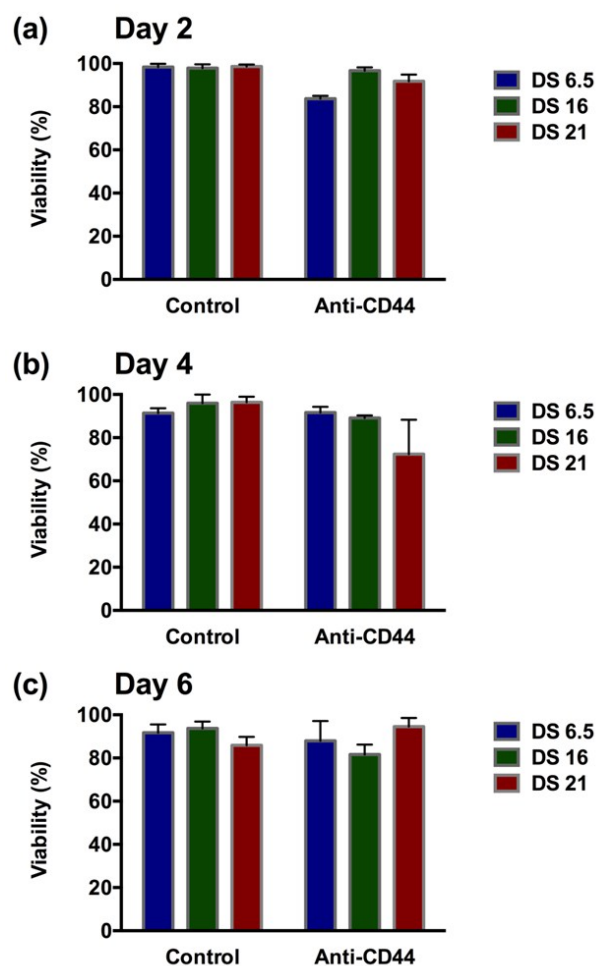


Figure S9. Quantification of cell viability of encapsulated hMSCs ($5 \times 10^6/\text{ml}$, $0.68 \text{ mM H}_2\text{O}_2$), blocked with CD44 antibody prior encapsulation compared to un-treated MSCs (control), after **(a)** 2 days, **(b)** 4 days and **(c)** 6 days of culture in basal medium (DMEM plus 10% FBS). One representative donor shown with mean + SD of $n = 4$ randomly chosen locations within the construct.

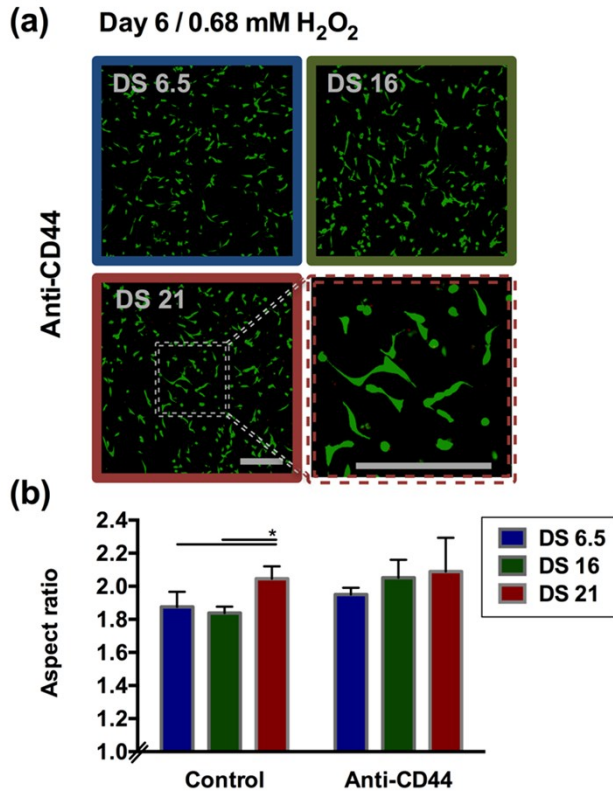


Figure S10. (a) Representative images of encapsulated hMSCs treated with CD44 antibody prior encapsulation and stained with Calcein (5 μ M, green) /Ethidium homodimer (8 μ M, red) after 6 days of culture. Images are shown as 3D rendered z-stacks with a thickness of 200 μ m. Dashed box shows a zoom-in of the corresponding area, scale bars are 200 μ m. (b) Quantification of cell aspect ratio of encapsulated hMSCs (5×10^6 /ml, 0.68 mM H₂O₂), blocked with CD44 antibody prior encapsulation compared to un-treated MSCs (control), after 6 days of culture in basal medium (DMEM plus 10% FBS). One representative donor shown with mean + SD of n = 4 randomly chosen locations within the construct (≥ 50 cells per image), *p<0.05.

REFERENCES

1. C. Loebel, M. D'Este, M. Alini, M. Zenobi-Wong and D. Eglin, *Carbohydr Polym*, 2015, **115**, 325-333.