Supporting Information

Real-time fluorescence turn-on assay for acetylcholinesterase activity based on the controlled release of a perylene probe from the MnO₂ nanosheets

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EXPERIMENTAL SECTION

Materials

P-4C+ was prepared as previously described.^{S1} AChE (from Electrophorus electricus) and trypsin were purchased from Sigma (St. Louis, MO, USA). Acetylthiocholine iodide, donepezil hydrochloride and 3-hydroxycarbofuran were purchased from J&K Scientific Ltd. (Beijing, China). S1 nuclease was purchased from Fermentas Inc. (MBI, Canada). Lysozyme was purchased from Dingguo Biotechnology Co., Ltd. (Beijing, China). Alkaline phosphatase (ALP) and exonuclease I (Exo I) were purchased from Takara Biotechnology Co., Ltd. (Dalian, China). All stock and buffer solutions were prepared using water purified with a Milli-Q A10 filtration system (Millipore, Billerica, MA, USA).

Instrumentation

UV-Vis absorption spectra were obtained using a Cary 50 Bio Spectrophotometer (Varian Inc., CA, USA) equipped with a xenon flash lamp. Emission spectra were recorded using a Fluoromax-4 spectrofluorometer (Horiba Jobin Yvon Inc., USA). Excitation wavelength was 442 nm. Quartz cuvettes with 10 mm path length and 2 mm window width were used for UV-vis and emission measurements. Zeta potential measurements were performed with a Zetasizer NanoZS (Malvern Instruments, USA). Transmission electron microscopy (TEM) images were obtained using a JEM-2100F high resolution transmission electron microscope (Philips, The Netherlands) operated at 200 kV. Atomic force microscopy (AFM) characterization was performed on a Multimode-V (Veeco Instruments, USA) using a tapping mode. Unless specified, all

concentrations of P-4C+, ATCh, MnO₂ nanosheets and buffer were those in the final assay solutions (total sample volume, 400 μ L), and all spectra were taken at 37 °C in 5 mM phosphate buffer at pH 7.4.

Preparation of the MnO₂ nanosheets

The MnO_2 nanosheets were prepared following the reported procedures.^{S2} In short, an aqueous solution comprised 20 mL of tetramethylammonium hydroxide (0.6 M) and H_2O_2 (3 wt%) was first prepared, and 10 mL of 0.3 M $MnCl_2$ was quickly mixed. The resulting mixture was stirred vigorously overnight in open air. The bulk MnO_2 was obtained though centrifugation, cleaned with double-distilled water and methanol and dried in an oven. MnO_2 nanosheets were obtained via ultrasonication of the aqueous solution of the bulk manganese dioxide.

Fluorescence quenching of P-4C+ by the MnO₂ nanosheets

A series of concentrations of the MnO_2 nanosheets (0, 8.75, 17.5, 26.25, 35, 43.75, 52.5, 61.25, and 70 µg/mL) were mixed with **P**-4C+ (5 µM) in 5 mM phosphate buffer (pH 7.4). Samples were mixed completely and maintained at 37 °C for 3 min. The emission intensity was then recorded.

ATCh concentration optimization

Thiocholine was prepared following the reported procedures^{S3} and characterized by ¹H-NMR (Figure S5). Different amounts of thiocholine were mixed with 5 mM phosphate buffer solution (pH 7.4) comprising 26.25 μ g/mL MnO₂ nanosheets and 50 μ M PPi. The samples were

incubated at 37 °C for 60 min. 5 μ M of **P**-4C+ was added. Samples were mixed and maintained at 37 °C for 3 min. The emission intensity was then recorded. Pyrophosphate was introduced to complex with the Mn²⁺ ions, since the in situ generated Mn²⁺ ions could attach to the surface of the MnO₂ nanosheets and caused aggregation of the nanosheets.

AChE assay procedures

0.3 mM ATCh (20 μ L) was injected to 380 μ L of 5 mM phosphate buffer solution (pH 7.4) comprising 26.25 μ g/mL MnO₂ nanosheets, 50 μ M PPi, 5 μ M **P**-4C+ and AChE of different concentrations. The final concentrations of AChE were 0, 5, 10, 25, 50, 100, 250, 500 and 1000 mU/mL, respectively. The temperature was stabilized at 37 °C and the emission intensity of **P**-4C+ at 488 nm was recorded with data points taken every 5 seconds.

Selectivity of the AChE assay

Different enzymes including lysozyme, alkaline phosphatase (ALP), S1 nuclease, trypsin, Exo I (10.0 U/mL each) and AChE (500 mU/mL) were mixed with the solution of 0.3 mM ATCh, 50 μ M PPi and 26.25 μ g/mL MnO₂ nanosheets in 5 mM phosphate buffer (pH 7.4). The solutions were incubated at 37 °C for 60 min. 5 μ M of **P**-4C+ was added. Samples were mixed completely and maintained at 37 °C for 3 min. The emission intensity was then recorded.

AChE assay in biological fluid

Various quantities of AChE were mixed with the sample solution of 26.25 μ g/mL MnO₂ nanosheets, 0.3 mM ATCh, 50 μ M PPi and 2.5% human serum in 5 mM phosphate buffer (pH

7.4). Samples were kept at 37 °C for 15 min. 5 μ M of **P**-4C+ was then mixed with the solution. Samples were mixed completely and maintained at 37 °C for 3 min. The emission intensity was then recorded.

AChE inhibitor screening

Various concentrations of the inhibitors (donepezil or 3-hydroxycarbofuran) were mixed with the sample solution of 26.25 μ g/mL MnO₂ nanosheets, 0.3 mM ATCh, 50 μ M PPi and 100 mU/mL AChE in 5 mM phosphate buffer (pH 7.4). The samples were kept at 37 °C for 15 min. 5 μ M of **P**-4C+ was added. Samples were mixed thoroughly and maintained at 37 °C for 3 min. The emission intensity was then recorded.

The inhibition efficiency (IE) is given by: $IE = [I - I_i]/[I - I_0]$

The emission intensity values of P-4C+ at 488 nm in the absence and presence of the inhibitor are symbolized as I and I_i, I₀ is the emission intensity of P-4C+ at 488 nm in the absence of AChE.

REFERENCES

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Figure S1. Tapping-mode of the AFM image of the MnO_2 nanosheets. Inset: the height profile of the section labeled with the white line.



Figure S2. UV-vis absorption spectrum of the MnO_2 nanosheets (black curve) and the fluorescence emission spectrum of P-4C+ (blue curve).



Figure S3. Zeta potential analysis: the MnO_2 nanosheets possess a potential value of -26.2 mV before (a) and 1.29 mV after (b) mixed with P-4C+.



Figure S4. Fluorescence quenching efficiency versus MnO₂ nanosheets concentration (0, 8.75,

17.5, 26.25, 35, 43.75, 52.5, 61.25, and 70 µg/mL). Buffer: 5 mM phosphate buffer, pH 7.4.



Figure S5. ¹H-NMR spectrum of thiocholine.



Figure S6. UV-vis absorption spectra: Blue curve: the reaction mixture of MnO_2 nanosheets and thiocholine was centrifuged, the supernatant was taken and treated with sodium periodate; Green curve: the reaction mixture of sodium periodate and $MnCl_2$; Red curve: KMnO₄ aqueous solution; Black curve (blank control): the MnO₂ nanosheets containing sample solution was centrifuged, the supernatant was taken and mixed with sodium periodate.

The reduction of MnO_2 to Mn^{2+} by thiocholine could be proven by a highly specific reaction of Mn^{2+} with sodium periodate, and the resulting sample solution shows clear color changes:

$$2Mn^{2+} + 5IO_4^{-} + 3H_2O \rightarrow 2MnO_4^{-} + 5IO_3^{-} + 6H^{-}$$



Figure S7. UV-vis absorption spectra of the MnO_2 nanosheets before (red curve) and after (black curve) the enzymatic reaction of AChE with the substrate.



Figure S8. Changes in emission intensity of P-4C+ (5 μ M) at 488 nm upon the addition of

increasing concentrations of free thiocholine. Conditions: 5 mM phosphate buffer (pH 7.4), 26.25 μ g/mL MnO₂ nanosheets and 50 μ M PPi.



Figure S9. Selectivity study. Columns A–G: AChE, lysozyme, ALP, S1 nuclease, trypsin and Exo I nuclease. AChE: 500 mU/mL; the other enzymes: 10 U/mL each. 50 μ M EDTA was used instead of pyrophosphate. I and I₀ represent the emission intensity of **P**-4C+ in the presence and absence of the enzyme.



Figure S10. Probing AChE activity in the presence of different biothiols (Cys, GSH, Hcy and Na₂S). Black columns: no AChE added. Red columns: all contains 100 mU/mL AChE. Columns a - d: different concentrations of ATCh were added (0, 50, 100, 150 \Box M, respectively). All samples contain 5 mM phosphate buffer (pH 7.4), 5 μ M P-4C+, 26.25 μ g/mL MnO₂ nanosheets, 50 μ M biothiol (Cys, GSH, Hcy, or Na₂S), and the emission intensity changes of P-4C+ at 488 nm were monitored.



Figure S11. Changes in emission intensity of **P**-4C+ at 488 nm (5 μ M) in the presence of the mround reductive substances. Green column: background emission of **P**-4C+ in the presence of the MnO₂ nanosheets and ATCh only; Red column: emission of **P**-4C+ in the presence of the MnO₂ nanosheets, ATCh, and Cys (or GSH, Hcy, Na₂S); Blue column: samples of Cys, GSH, Hcy, and Na₂S were passed through a filter (MWCO 30K), wash with water, the supernatant was taken, and mixed with **P**-4C+, MnO₂ nanosheets, and ATCh; Black column: samples of Cys, GSH, Hcy, and Na₂S were mixed with AChE, passed through the filter, and mixed with **P**-4C+, MnO₂ nanosheets, and ATCh; Conditions: 5 mM phosphate buffer (pH 7.4), 26.25 μ g/mL MnO₂ nanosheets, 0.3 mM ATCh, 50 μ M PPi, 0.3 mM Cys (or GSH, Hcy, Na₂S), 0.5 U/mL AChE.



Figure S12. Emission intensity changes of **P**-4C+ (5 μ M) at 488 nm in dilute human serum (2.5%) with time. Conditions: 5 mM phosphate buffer (pH 7.4), 50 μ M PPi, 26.25 μ g/mL MnO₂ nanosheets. The results show that the assay mixture is stable enough for the enzyme activity quantification (Figure 3).



Figure S13. Changes in emission intensity of **P**-4C+ at 488 nm (5 μ M) with AChE concentration in dilute human serum (2.5%). Conditions: 5 mM phosphate buffer (pH 7.4), 26.25 μ g/mL MnO₂ nanosheets, 0.3 mM ATCh, 50 μ M PPi.



Figure S14. Changes in emission spectrum (a) and the corresponding emission intensity at 488 nm (b) of **P**-4C+ upon the addition of increasing concentrations of donepezil (0 – 200 nM). Conditions: 5 mM phosphate buffer (pH 7.4), 26.25 μ g/mL MnO₂ nanosheets, 0.3 mM ATCh, 50 μ M PPi and 100 mU/mL AChE.



Figure S15. Changes in emission spectrum (a) and the corresponding emission intensity at 488 nm (b) of **P**-4C+ upon the addition of increasing concentrations of 3-hydroxycarbofuran (0 – 400 nM). Conditions: 5 mM phosphate buffer (pH 7.4), 26.25 μ g/mL MnO₂ nanosheets, 0.3 mM ATCh, 50 μ M PPi and 100 mU/mL AChE.