

Low toxicity and accumulation of zinc oxide nanoparticles in mice after 270-day consecutive dietary supplement

Jia-Hui Liu,^{a,b} Xin Ma,^c Yingying Xu,^a Huan Tang,^a Sheng-Tao Yang,^a
Yi-Fan Yang,^c Dong-Dong Kang,^c Haifang Wang^{*c} and Yuanfang Liu^{*a}

^aBeijing National Laboratory for Molecular Sciences, College of Chemistry and Molecular Engineering, Peking University, Beijing 100871, China

^bBeijing Key Laboratory of Bioprocess, College of Life Science and Technology, Beijing University of Chemical Technology, Beijing 100029, China

^cInstitute of Nanochemistry and Nanobiology, Shanghai University, Shanghai 200444, China

1. Bioaccessibility of Zn in animal food by the RIVM method

The RIVM method was used to assess the Zn bioaccessibility of animal food containing ZnO NPs, ZnO MPs, and Zinc ion, respectively. The animal food (0.1 g) was used in this in vitro digestion. The constituents and concentrations of all synthetic juices are shown in Table S1. All digestive juices were heated to 37 °C before digestion. The digestions were carried out in a Tabletop Orbital Shaker (SPH-100B, Shanghai Shiping Laboratory Equipment Co., LTD, China) at 100 rpm. The pH values of the digestive juices were adjusted with NaHCO₃ (1M) or HCl (37% w/w). Digestion started by introducing 4 mL gastric juice into 0.1 g animal food. The mixture was adjusted to pH 1.3 ± 0.2 and shaken for 2 h. After that, 4 mL duodenal juice, 2 mL bile juice were added. The mixture was adjusted to pH 6.5 ± 0.2, and shaken for another 2 h. Then, the mixture was centrifuged at 3000 g for 10 min (MIKRO 200R, Hettich, Germany). Finally, 0.5 mL of the supernatant was used to determine Zn concentration by an atomic absorption spectroscopy (900T, PerkinElmer, USA) after digestion with 1 mL HNO₃ and 1 mL H₂O₂, and diluted to 2% HNO₃ solution. The Zn bioaccessibility in animal food was calculated following the completion of intestinal phase extraction according to the following equation:

$$\text{Zn bioaccessibility (\%)} = \text{Zn in supernatant} / \text{total Zn} \times 100$$

where Zn in supernatant is Zn (μg) extracted from 0.1 g of sample, and total Zn is Zn (μg) in 0.1 g animal food.

Table S1. Composition of the synthetic juices for the in vitro digestion model (Amounts based on 1000 mL of juice) with minor modification (R. Peters, E. Kramer, A. G. Oomen, Z. E. Herrera Rivera, G. Oegema, P. C. Tromp, R. Fokink, A. Rietveld, H. J. P. Marvin, S. Weigel, A. A. C. M. Peijnenburg, H. Bouwmeester, *ACS Nano* **2012**, 6, 2441).

| | <i>Gastric juice</i> <i>pH 1.3±0.1</i> | <i>Duodenal juice</i> <i>pH 8.1±0.1</i> | <i>Bile juice</i> <i>pH 8.2±0.1</i> |
|------------------------|--|--|--|
| Inorganic constituents | 2752 mg NaCl 306 mg NaH ₂ PO ₄ ·H ₂ O 824 mg KCl 302 mg CaCl ₂ 6.5 mL glucose | 7012 mg NaCl 3388 mg NaHCO ₃ 80 mg KH ₂ PO ₄ 564 mg KCl 50 mg MgCl ₂ ·6H ₂ O | 5259 mg NaCl 5785 mg NaHCO ₃ 376 mg KCl 150 µL HCl(37%) 250 mg urea |
| organic constituents | 650 mg glucose 20 mg glucuronic acid 85 mg urea 330 mg glucosaminhydrochloride 1 g BSA 2.5g pepsin 3 g mucin milli-Q water | 180 µL HCl(37%) 100 mg urea 151 mg CaCl ₂ 1 g BSA 4.5 g pancreatin 1.5 g lipase milli-Q water | 167.5 mg CaCl ₂ 1.8 g BSA 30 g bile milli-Q water sodium carbonate solution 84.7 g NaHCO ₃ milli-Q water |

Table S2. Zn bioaccessibility of food replenished with Zn ion, ZnO-NP and ZnO-MP by the RIVM method.

| Sample | Gastric | | Intestine | |
|----------|--------------------------------|-------------------------|--------------------------------|-------------------------|
| | Dissolved content (µg/0.1g) | Bioaccessibility (%) | Dissolved content (µg/0.1g) | Bioaccessibility (%) |
| Zinc ion | 119.9±5.3 | 74.9 | 15.9±2.0 | 9.9 |
| ZnO-NP | 104.0±1.1 | 65.0 | 15.1±1.8 | 9.4 |
| ZnO-MP | 110.5±2.0 | 69.1 | 15.3±1.2 | 9.5 |

2. XRD patterns of ZnO particles

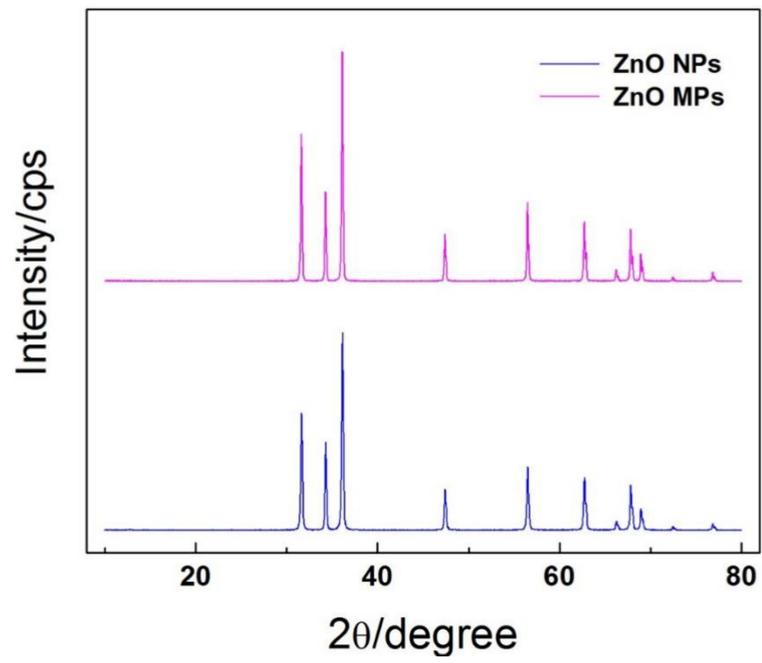


Fig. S1 XRD spectra of ZnO NPs and ZnO MPs.

3. TEM images of ZnO particles separated from the animal food samples

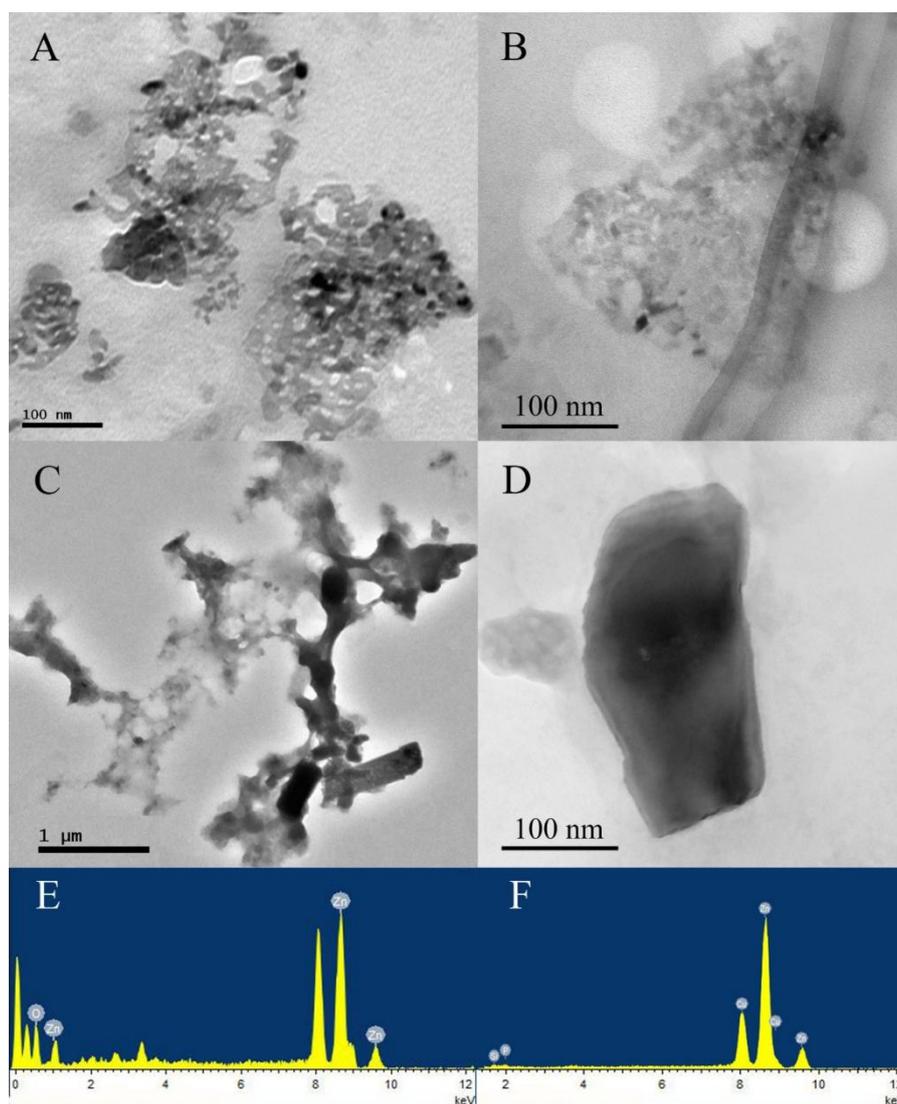


Fig. S2 The morphology and size of ZnO-NPs and ZnO-MPs separated from the animal food. (A&B) TEM images of ZnO-NPs; (C&D) TEM images of ZnO-MPs; (E) EDS data confirming the chemicals from image B; (F) EDS data confirming the chemicals from image D.

4. Ultra-thin pathological photos of stomach of female mice

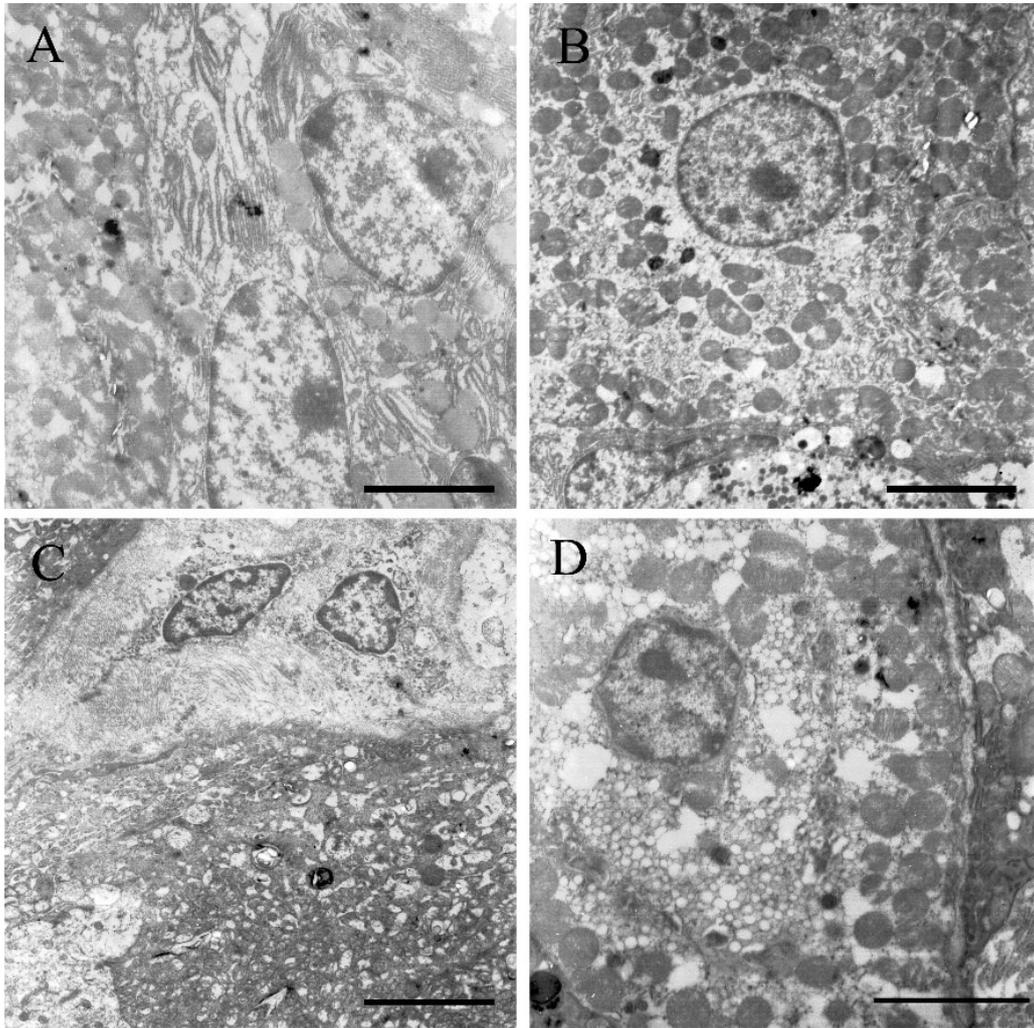


Fig. S3 Representative ultra-thin pathological photos of stomach of female mice after 270 days consecutive zinc dietary supplement. (A) control group, (B) Zn ion group, (C) ZnO-NPs group, (D) ZnO-MPs group. The scale bar is 2.5 μm.

5. Copper distribution in mice after exposed to zinc oxide particles and zinc ion

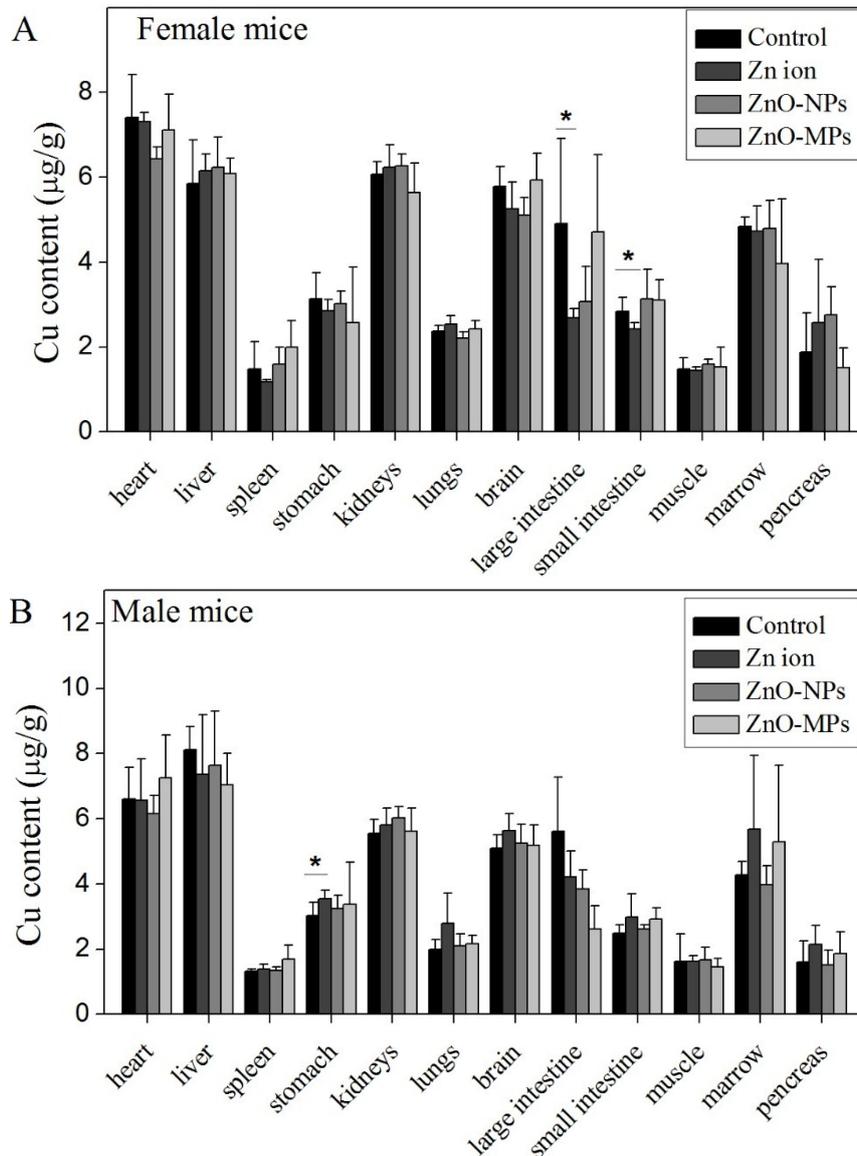


Fig. S4 Copper contents in tissues in the control, Zn ion, ZnO-NPs, and ZnO-MPs mice after 270 days consecutive exposure to zinc oxide particles or zinc ion (n=5). * $p < 0.05$, compared to the control. (A) female mice; (B) male mice.

The consumption of excessive zinc ion increased the copper content in stomach in male mouse, but decreased the copper content in intestine in female mouse. However, the exposure of ZnO-NPs and ZnO MPs didn't influence the copper distribution in mice.