

## Supplemental Information

### **A Two-photon Fluorescent Probe for Nitroreductase imaging in Living Cells, Tissues and Zebrafish with Hypoxia Condition**

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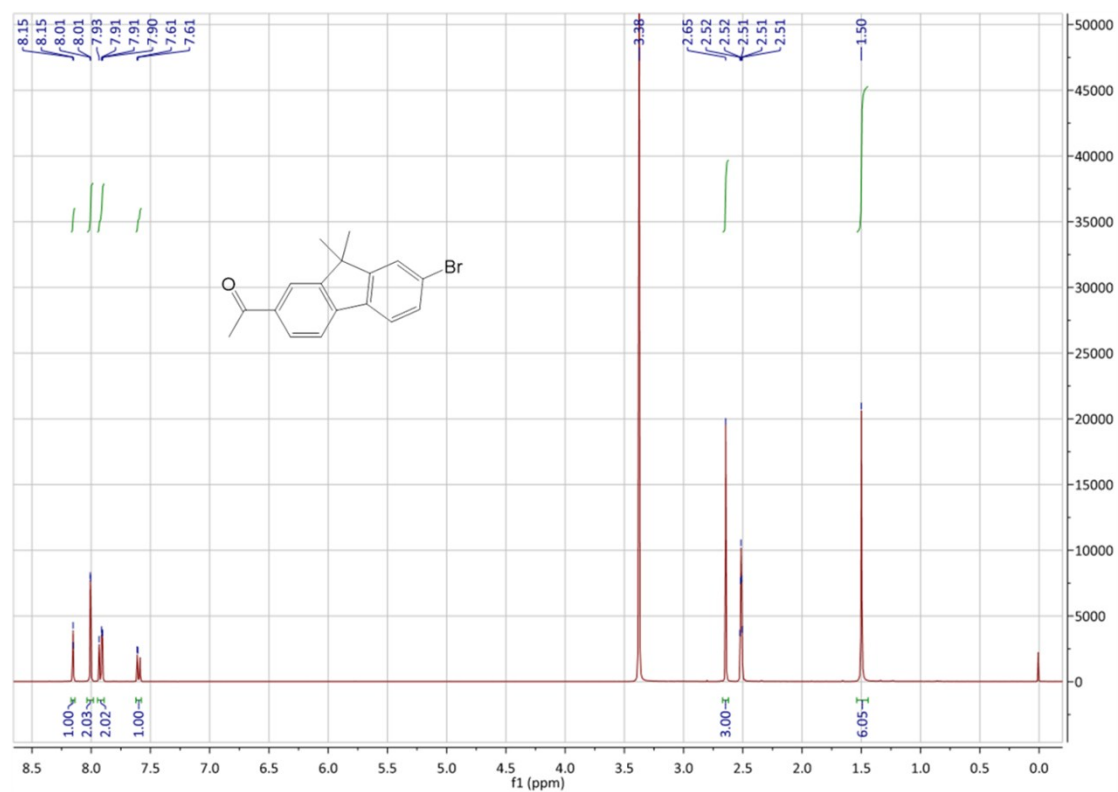
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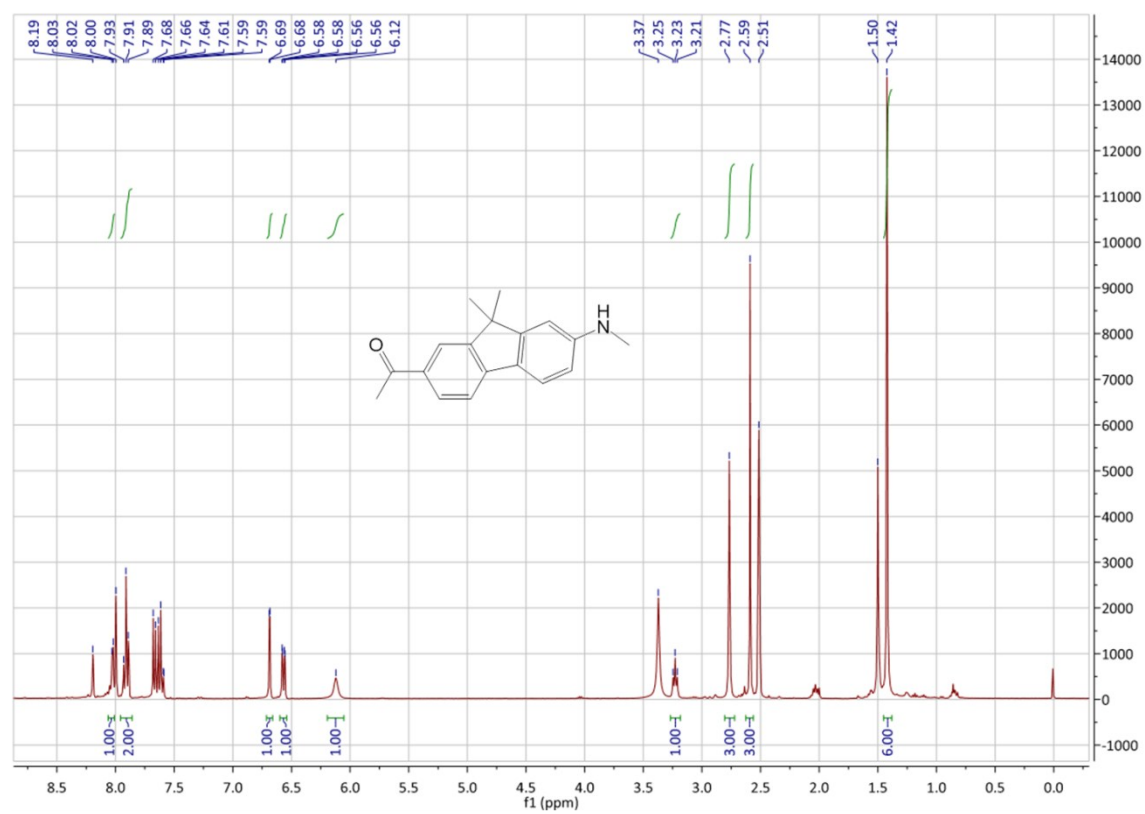
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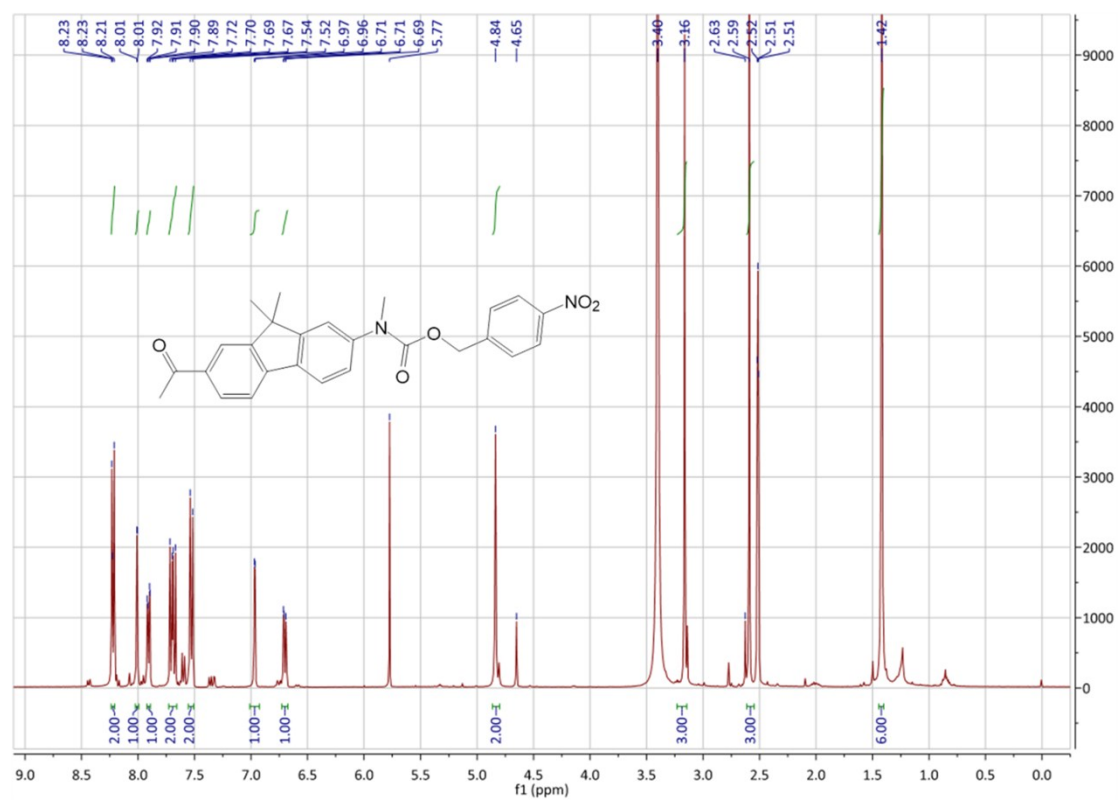
E-mail address: zhhlui@whu.edu.cn



**Figure S 1** <sup>1</sup>H-NMR spectrum of **2** (400 MHz, *d*<sub>6</sub>-DMSO)

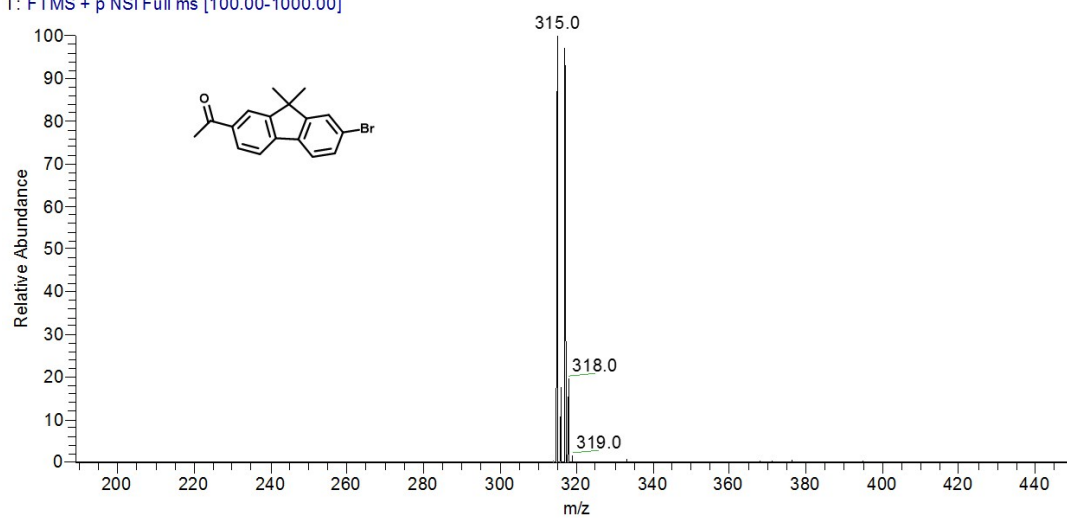


**Figure S 2**  $^1\text{H-NMR}$  spectrum of **1** (400 MHz,  $d_6$ -DMSO)

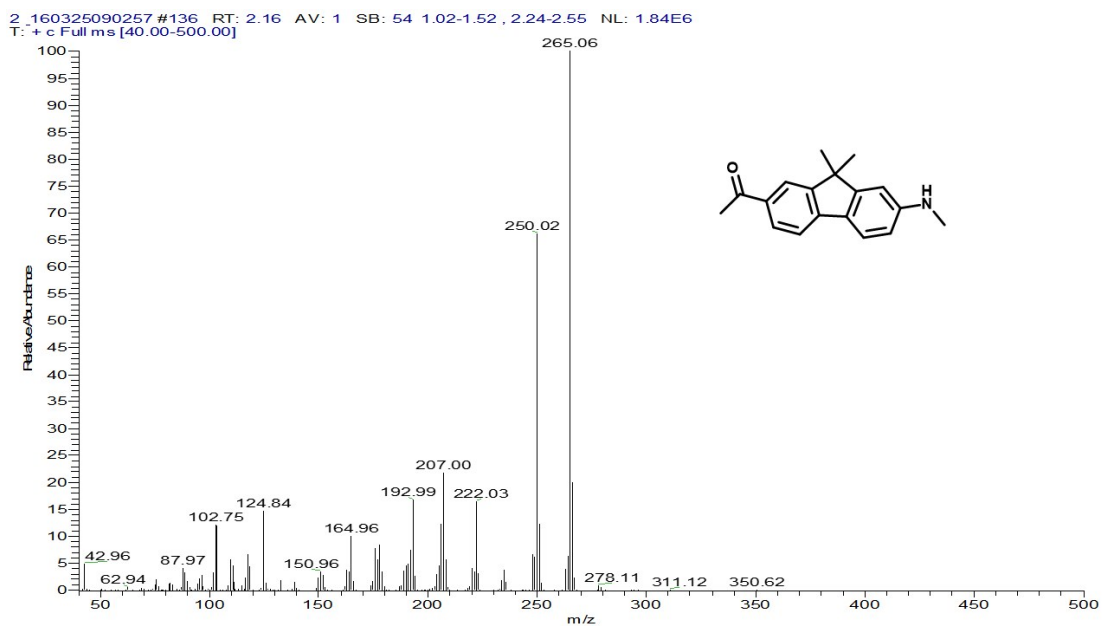


**Figure S 3**  $^1\text{H-NMR}$  spectrum of FNTR (400 MHz,  $d_6$ -DMSO)

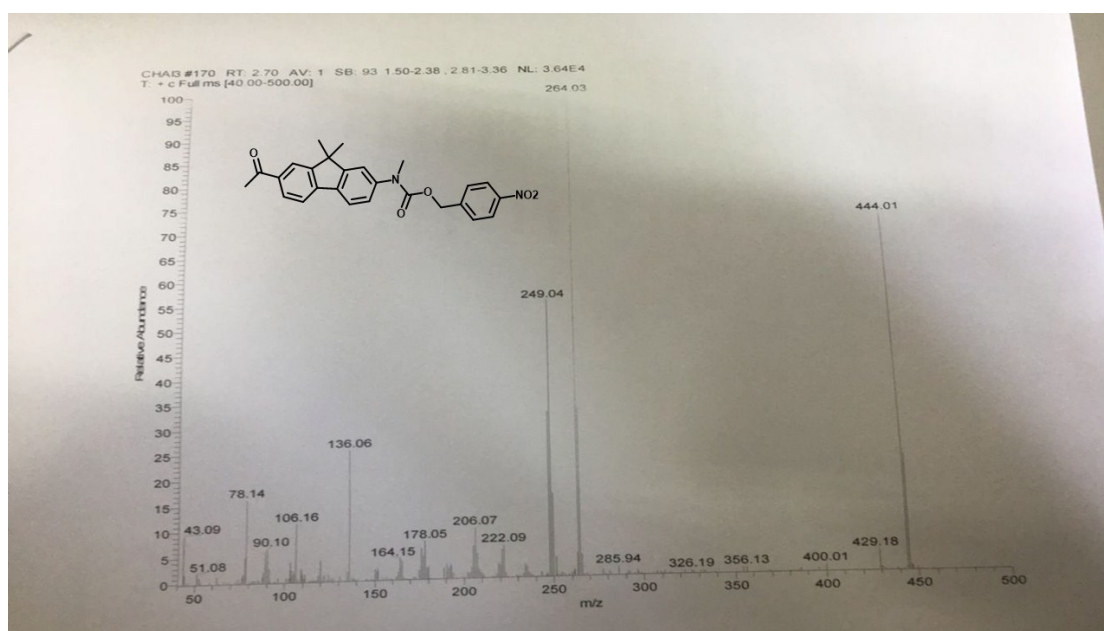
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**Figure S 4** HRMS (MALDI) spectrum of **2**



**Figure S 5** HRMS (MALDI) spectrum of **1**



**Figure S 6** HRMS (MALDI) spectrum of **FNTR**

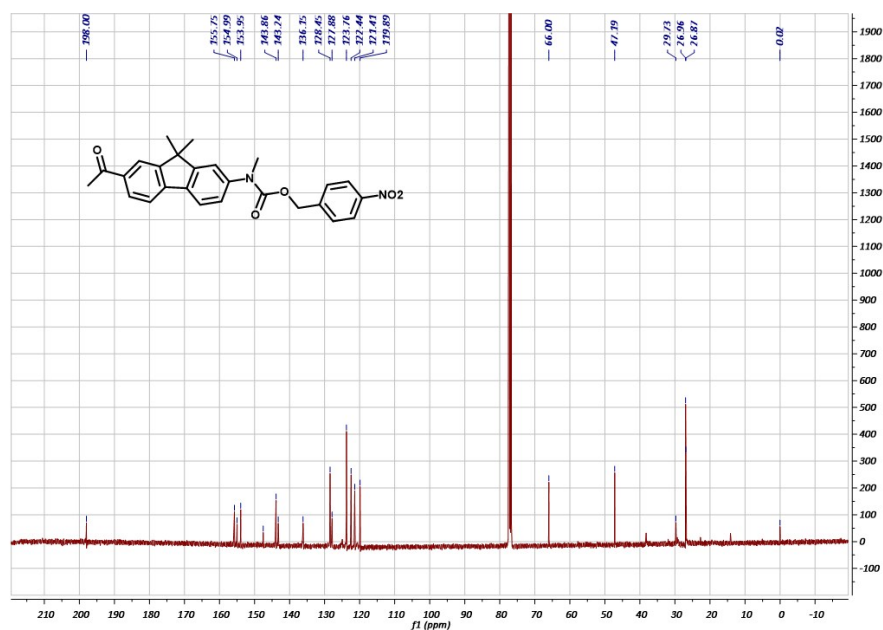


Figure S 7  $^{13}\text{C}$ -NMR spectrum of FNTR (100 MHz,  $d_6$ -DMSO)

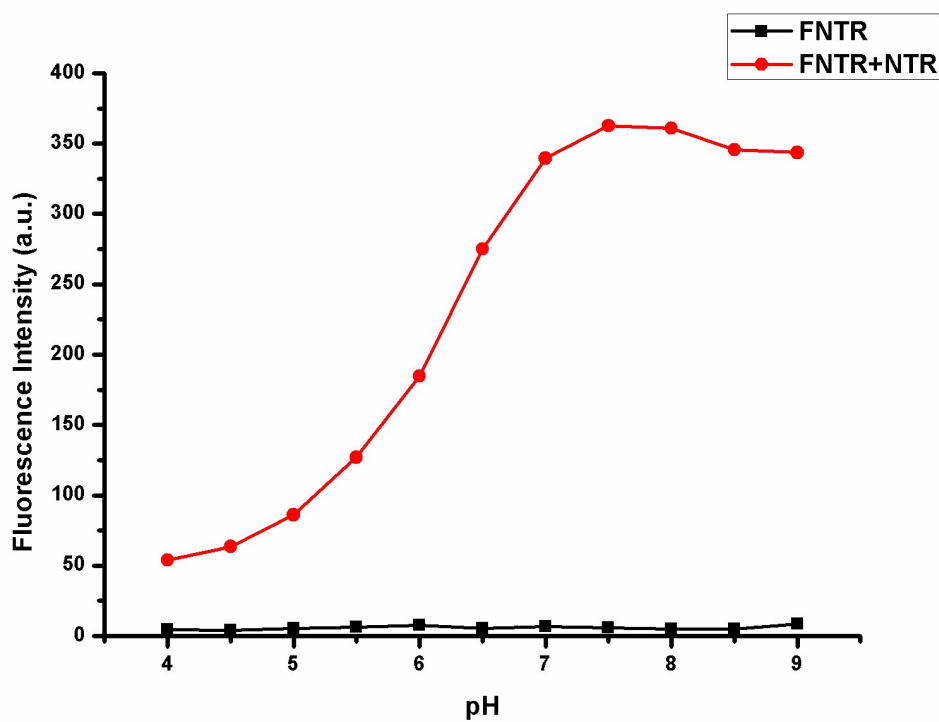
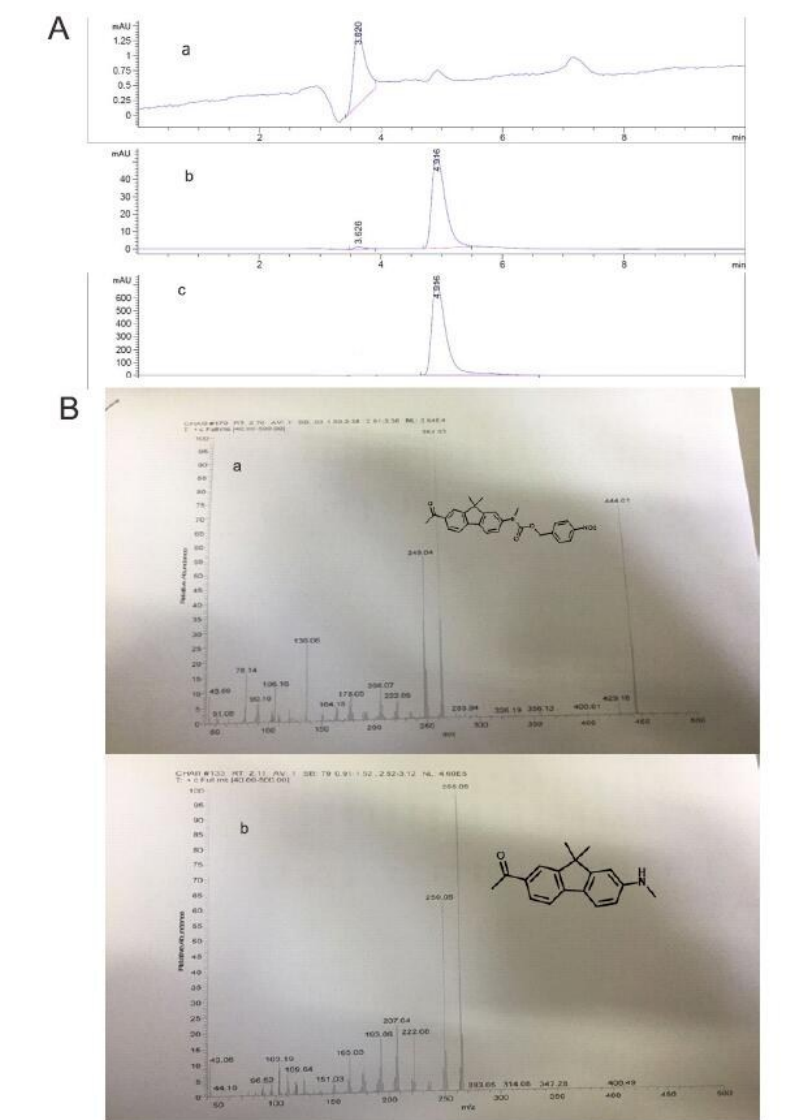
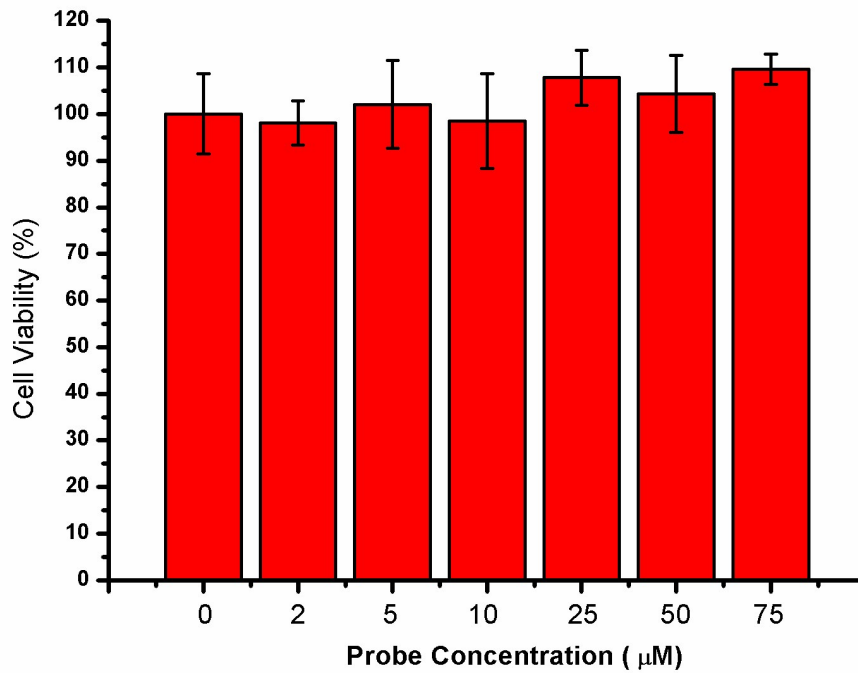


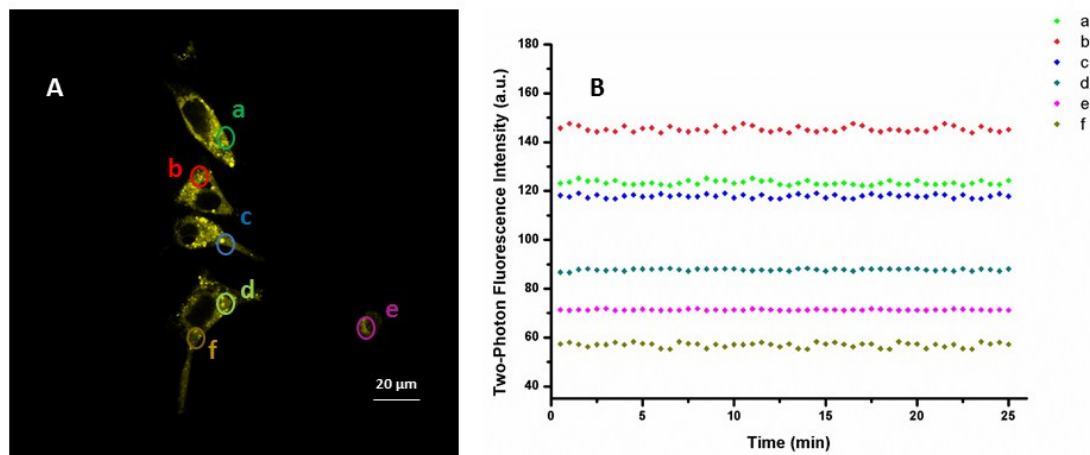
Figure S 8 Effects of pH on the fluorescence ( $\lambda_{\text{ex/em}} = 410/560 \text{ nm}$ ) of FNTR ( $5 \mu\text{M}$ ) and its reaction product with nitroreductase ( $1 \mu\text{g/mL}$ ) in the presence of  $500 \mu\text{M}$  NADH.



**Figure S 9** (A) Chromatograms of different reaction systems. (A) 10  $\mu$ M **FNTR** (a); 10  $\mu$ M **1** (b); the reaction products of 10  $\mu$ M **FNTR** with 20  $\mu$ g/mL nitroreductase in the presence of 100  $\mu$ M NADH (c). Detection: UV-vis (365 nm) detector. Flow rate: 0.3 mL/min. T: 20  $^{\circ}$ C. Injection volume: 100  $\mu$ L. Mobile phase: acetonitrile–water, 50:50 (v/v). (B) The Chromatograms-mass of the production with the standard substances of **FNTR** (a) and **1** (b)

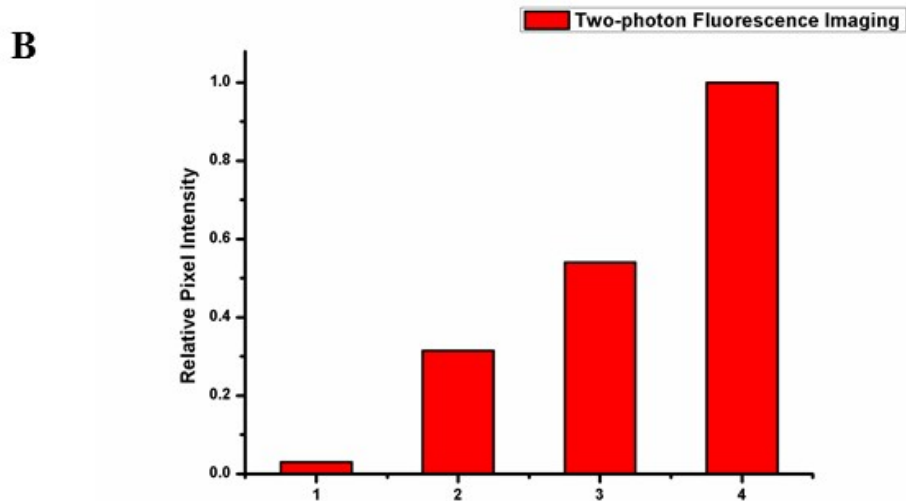
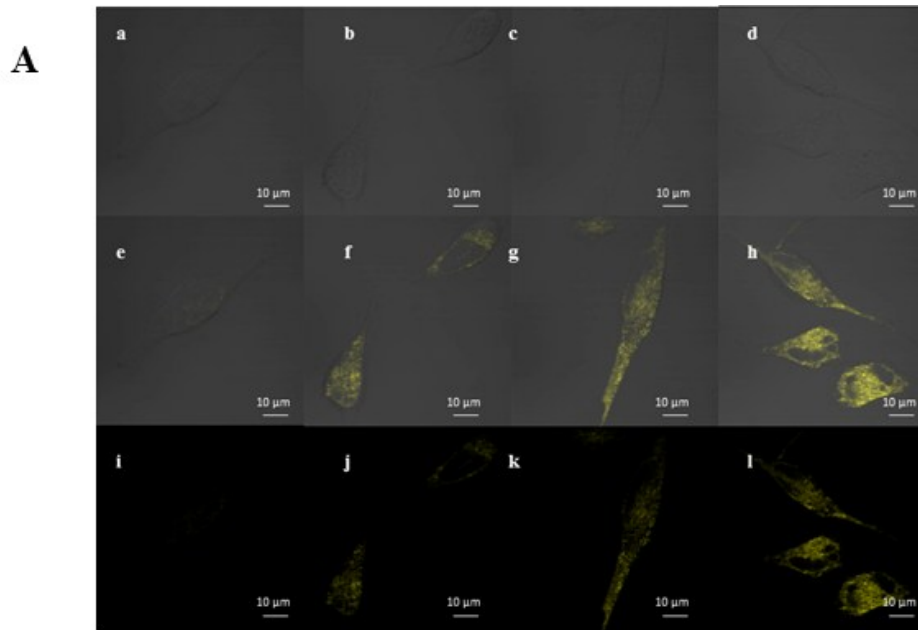


**Figure S 10.** Cell survival rate of control groups (without FNTR) and experimental group (with 2,, 5, 10, 25, 50, 75 μM of FNTR). All groups contain 1 % DMSO in 100 μL DMEM)



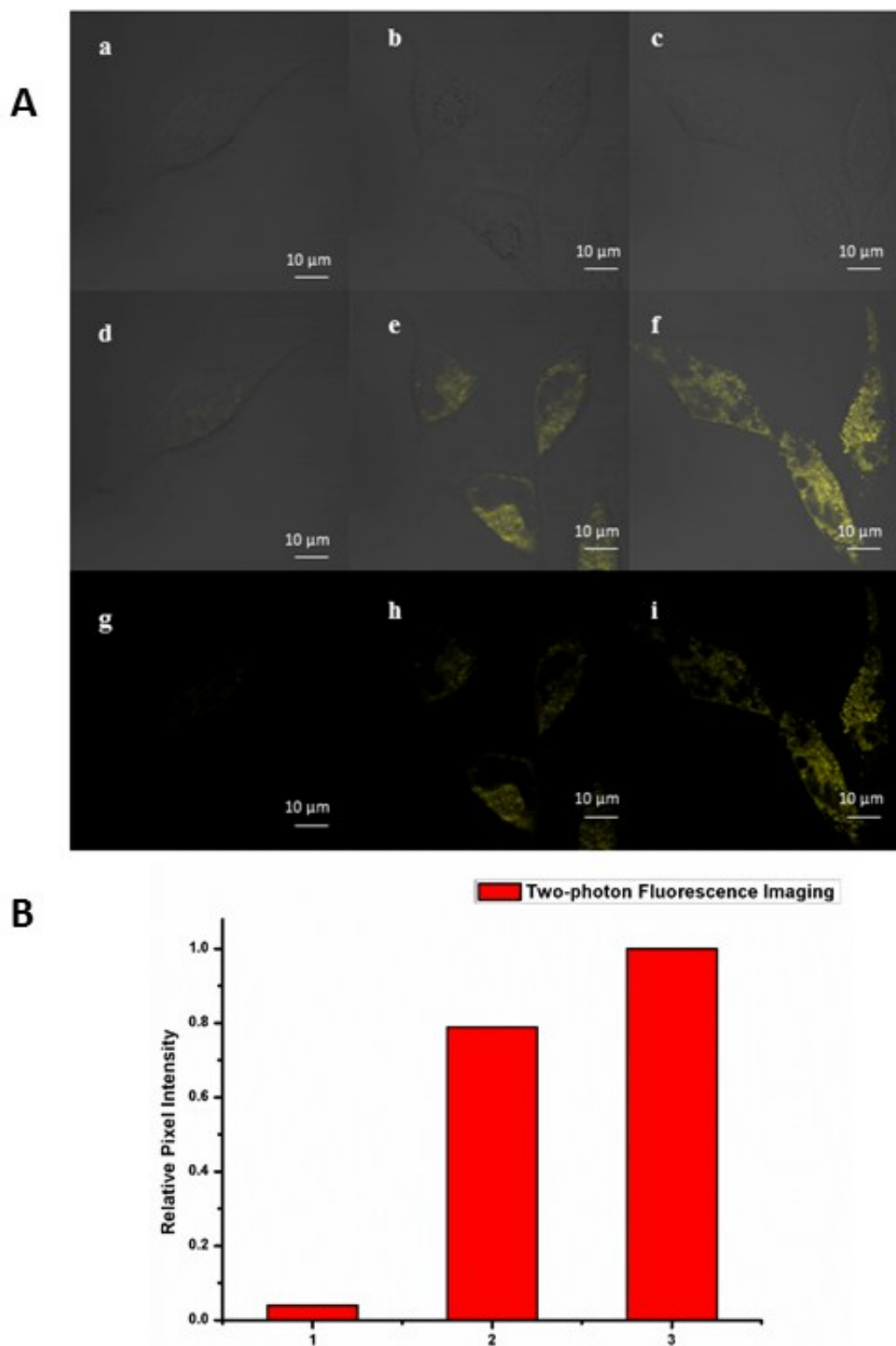
**Figure S 11.** (A) TPM images of HeLa cells labeled with 200 μM CoCl<sub>2</sub> for 48 h and further incubated with FNTR for 30 min separately. (B) Two-photon fluorescence intensity from circle a-f as a function of time. The two-photon fluorescence intensity was collected with 15 sec intervals for the duration of 25 min under *xyt* mode. Scale bar: 20 μm





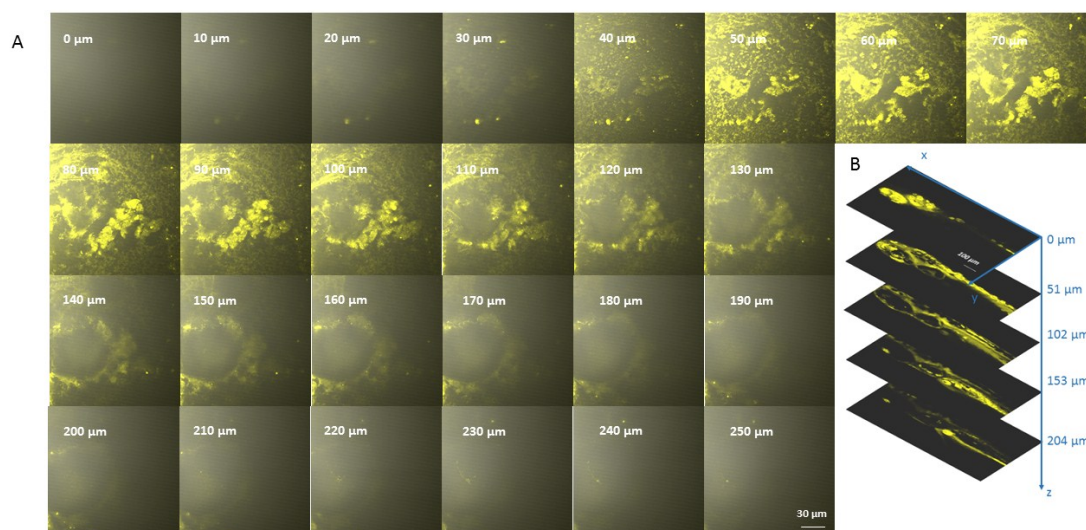
**Fig. S 12** (A) TPM images of HeLa cells under normoxic, 200  $\mu\text{M}$   $\text{CoCl}_2$  for 4, 24, 48 h, respectively. (a-d) The differential interference contrast (DIC) images. (e-h) Merged images of TPM and DIC. (i-l) TP fluorescence images of HeLa cells under normoxic and different hypoxic conditions.  $\lambda_{\text{ex}} = 750$  nm; yellow channel:  $\lambda_{\text{em}} = 480\text{-}680$  nm. Scale bar: 10  $\mu\text{m}$ . (B) Relative pixel fluorescence intensity of the HeLa cells images. These cells were incubated normoxic (0  $\mu\text{M}$   $\text{CoCl}_2$ ) and 200  $\mu\text{M}$   $\text{CoCl}_2$  for 4, 24 and 48 h, respectively. And then incubated with 10  $\mu\text{M}$  **FNTR** for 30 min. The strongest pixel

signal intensity from the images of HeLa cells under 200  $\mu\text{M}$   $\text{CoCl}_2$  for 48 h is defined as 1.0.



**Fig. S 13** (A) TPM images of HeLa cells under normoxic, 400  $\mu\text{M}$   $\text{CoCl}_2$  for 4, 24 h, (a-c) The differential interference contrast (DIC) images. (d-f) Merged

images of TPM and DIC. (g-i) TP fluorescence images of HeLa cells under normoxic and different hypoxic conditions.  $\lambda_{\text{ex}} = 750 \text{ nm}$ ; yellow channel:  $\lambda_{\text{em}} = 480\text{-}680 \text{ nm}$ . Scale bar:  $10 \mu\text{m}$ . (B) Relative pixel fluorescence intensity of the HeLa cells images. These cells were incubated normoxic ( $0 \mu\text{M CoCl}_2$ ) and  $400 \mu\text{M CoCl}_2$  for 4, 24 h, respectively. And then incubated with  $10 \mu\text{M FNTR}$  for 30 min. The strongest pixel signal intensity from the images of HeLa cells under  $400 \mu\text{M CoCl}_2$  for 24 h is defined as 1.0.



**Fig. S 14** Separate images at different  $z$ -axis depth of the **FNTR**-labeled rat liver tissue and zebrafish . The fluorescence intensity was reflected by color. Scale bar:  $30 \mu\text{m}$  and  $100 \mu\text{m}$ , respectively.