

Electronic supplementary information

## **Simple and rapid colorimetric detection of melanoma circulating tumor cells with bifunctional magnetic nanoparticles**

Junrong Li,<sup>a†</sup> Jing Wang,<sup>a†</sup> Yuling Wang<sup>b\*</sup> and Matt Trau<sup>a,c\*</sup>

<sup>a</sup>Centre for Personalized Nanomedicine, Australian Institute for Bioengineering and Nanotechnology (AIBN), The University of Queensland, Brisbane, QLD 4072, Australia

<sup>b</sup>Department of Molecular Sciences, ARC Centre of Excellence for Nanoscale BioPhotonics, Faculty of Science and Engineering, Macquarie University, Sydney, NSW 2109, Australia

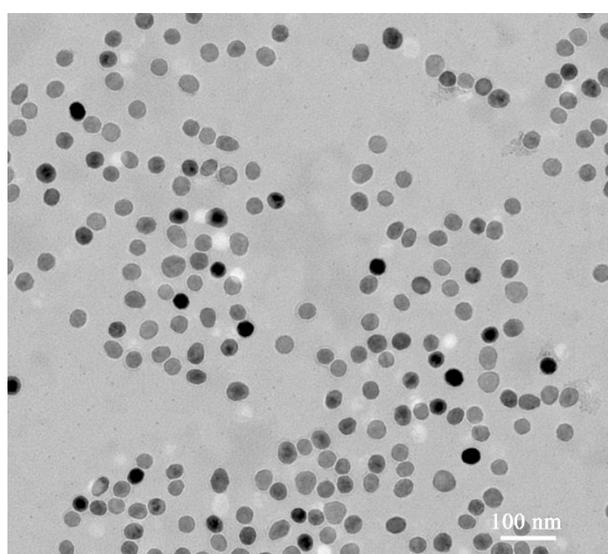
<sup>c</sup>School of Chemistry and Molecular Biosciences, The University of Queensland, Brisbane, QLD 4072, Australia

E-mail: [yuling.wang@mq.edu.au](mailto:yuling.wang@mq.edu.au); [m.trau@uq.edu.au](mailto:m.trau@uq.edu.au)

†These two authors contributed equally.



**Fig. S1.** Nanozyme activities of 30 nm and 1 μm magnetic nanoparticles, respectively.



**Fig. S2.** TEM image of MNPs

**Table S1.** Capture efficiency and nonspecific binding from spiked cells in PBS solution

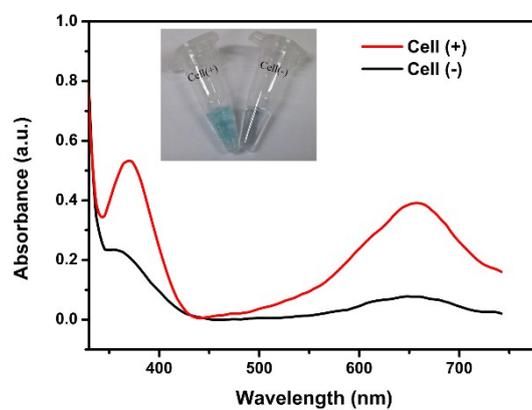
	Spiked	Detected 1	Detected 2	Detected 3	Average Recovery (%)	RSD (%)
LM-MEL-33	1000	673	771	662	70.20	6.00
MDA-MB-231	10000	360	260	233	2.84	0.67

$$\text{Capture efficiency} = \frac{\text{Number of detected positive cells}}{\text{Number of spiked positive cells}} \times 100\%$$

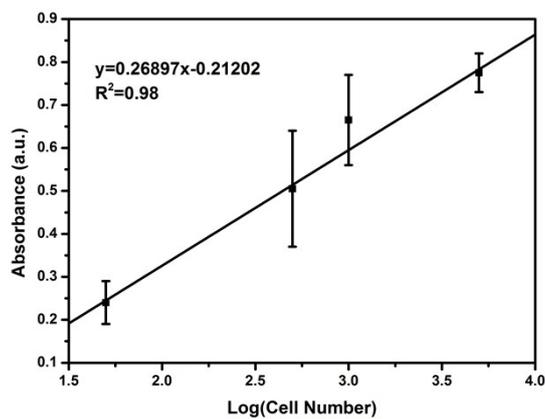
$$\text{Non-specific recovery} = \frac{\text{Number of detected negative cells}}{\text{Number of spiked negative cells}} \times 100\%$$

Positive cells: LM-MEL-33

Negative cells: MDA-MB-231



**Fig. S3.** Cells alone in catalyzing TMB in the presence of H<sub>2</sub>O<sub>2</sub>. Cell (+): 10<sup>6</sup> cell, Cell (-): 0 cell.



**Fig. S4.** The absorbance at 652 nm against natural logarithm over the different numbers of LM-MEL-33 cells in 7.5 mL of blood samples.