## Electronic Supplementary Information (ESI) for

## A Ratiometric Fluorescent pH Probe Based on Keto-Enol Tautomerization for Imaging of Living Cells in Extreme Acidity

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Figure S1. <sup>1</sup>H NMR spectrum of compound 1 in CDCl<sub>3</sub>



Figure S2.  $^{\rm 13}\rm C$  NMR spectrum of compound 1 in  $\rm CDCl_3$ 



00 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -1 f1 (ppm)

Figure S4. <sup>13</sup>C NMR spectrum of probe DDXC in CDCl<sub>3</sub>



Figure S5. HRMS of probe DDXC



**Figure S6.** UV-vis absorption spectra of probe DDXC (30 µM) recorded at different pH values from 6.8 to 1.2. Inset: Sigmoidal fitting of pH-dependent absorbance at 400 nm



**Figure S7.** a) HRMS of probe DDXC (1.0  $\mu$ M) toward CF<sub>3</sub>COOH (CF<sub>3</sub>COOH/MeOH = 1.0%, v/v); b) the partial enlarged drawing of a).



**Figure S8.** Fluorescence emission spectra of DDXC (10  $\mu$ M) in phosphate-citrate buffer (10 mM Na<sub>2</sub>HPO<sub>4</sub> and 10 mM citric acid) as the pH value decreased from 6.8 to 1.2.  $\lambda_{ex}$  = 490 nm, excitation and emission bandwidths were both set at 10 nm.



Figure S9. The time courses of fluorescence intensity of DDXC (10  $\mu$ M) in buffer pH 6.8, 3.3 and 1.2, respectively,  $\lambda_{ex/em} = 490/580$  nm.



**Figure S10.** Cell cytotoxicity assay of probe DDXC: the cell viability values (%) are determined by incubated Hela cells with DDXC of varying concentrations (0, 2, 5, 10 and 20  $\mu$ M) for 12 h.