

## Supporting Information

### A pH responsive AIE probe for enzyme assays

Leilei Shi,<sup>1,†</sup> Yufeng Liu,<sup>1,†</sup> Qian Wang,<sup>1</sup> Tiankuo Wang,<sup>2</sup> Yubin Ding,<sup>1</sup> Yi Cao,<sup>2</sup> Zhe Li,<sup>1</sup> and  
Hui Wei,<sup>1,3,\*</sup>

<sup>1</sup> Department of Biomedical Engineering, College of Engineering and Applied Sciences, Nanjing National Laboratory of Microstructures, Nanjing University, Nanjing, Jiangsu, 210093, China.

<sup>2</sup> School of Physics, Collaborative Innovation Center of Advanced Microstructures, Nanjing National Laboratory of Microstructures, Nanjing University, Nanjing, Jiangsu 210093, China.

<sup>3</sup> Collaborative Innovation Center of Chemistry for Life Sciences, State Key Laboratory of Analytical Chemistry for Life Science, Nanjing University, Nanjing, Jiangsu, 210093, China.

Email: weihui@nju.edu.cn; Fax: +86-25-83594648; Tel: +86-25-83593272; Web:  
<http://weilab.nju.edu.cn>.

† L.S. and Y.L. contributed equally.

## Table of Contents

**Figure S1.**  $^1\text{H}$  NMR of **TPE-NH<sub>2</sub>** in DMSO- $d_6$ .

**Figure S2.**  $^1\text{H}$  NMR of **TPE-Leu** in DMSO- $d_6$ .

**Figure S3.**  $^{13}\text{C}$  NMR of **TPE-Leu** in DMSO- $d_6$ .

**Figure S4.** HPLC Chromatogram of **TPE-Leu**.

**Figure S5.** Mass spectrum of **TPE-Leu**.

**Figure S6.** Plot of  $I/I_0$  of 10  $\mu\text{M}$  **TPE-Leu** in DMSO/PBS buffer with different volume fractions of PBS buffer, where  $I_0$  is the fluorescence intensity of **TPE-Leu** in 99.5% PBS buffer.  $\lambda_{\text{ex}}=320$  nm.

**Figure S7.** Plots of fluorescence intensity of 10  $\mu\text{M}$  **TPE-NH<sub>2</sub>** at 455 nm versus pH values.  $\lambda_{\text{ex}}=320$  nm.

**Figure S8.** Selectivity of 10  $\mu\text{M}$  **TPE-Leu** in pH=7 DMSO/buffer (1:9, v:v) over 0.1 mM  $\text{Cu}^{2+}$ ,  $\text{Fe}^{2+}$ , and  $\text{Fe}^{3+}$ .

**Figure S9.** Selectivity of 10  $\mu\text{M}$  **TPE** in pH=7 DMSO/buffer (1:9, v:v) over 0.1 mM/0.5 mM  $\text{Cu}^{2+}$ ,  $\text{Fe}^{2+}$ , and  $\text{Fe}^{3+}$ .

**Figure S10.** Selectivity of 10  $\mu\text{M}$  **TPE-NH<sub>2</sub>** in pH=7 DMSO/buffer (1:9, v:v) over 0.1 mM/0.5 mM  $\text{Cu}^{2+}$ ,  $\text{Fe}^{2+}$ , and  $\text{Fe}^{3+}$ .

**Figure S11.** Fluorescence intensity of **TPE-Leu** (10  $\mu\text{M}$ ) in pH=7 DMSO/buffer (1:9, v/v) in the absence (1) or presence of 0.1 mM  $\text{Cu}^{2+}$  (2), 0.1 mM  $\text{Cu}^{2+}$  and EDTA (3), 0.1 mM  $\text{Fe}^{3+}$  (4), and 0.1 mM  $\text{Fe}^{3+}$  and EDTA (5).  $\lambda_{\text{ex}}=320$  nm.

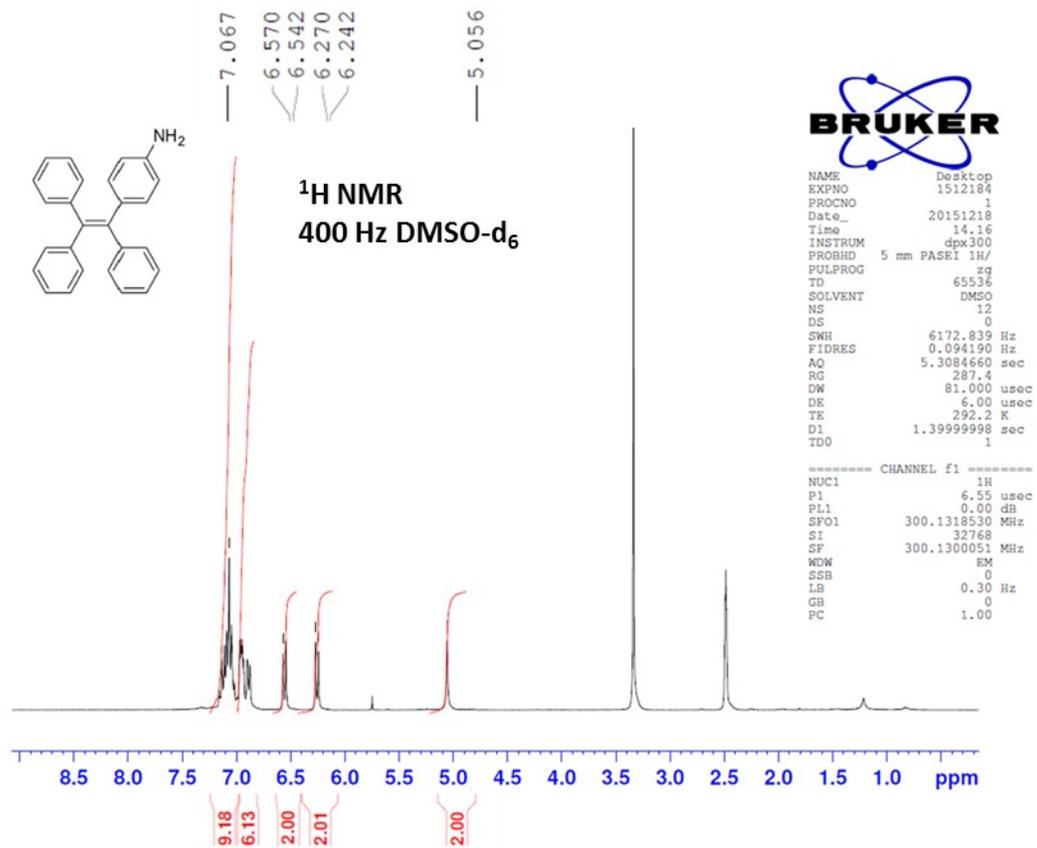
**Figure S12.** Photo of **TPE-Leu** in different pH buffers.

**Figure S13.** AFM images of (A) **TPE-Leu** in DMSO/buffer (5 mM pH=4 NaOAc) (1:9, v:v) and (B) **TPE-Leu** in DMSO/buffer (5 mM pH=10 PBS) (1:9, v:v). (C) and (D) were the corresponding cross-sectional profiles.

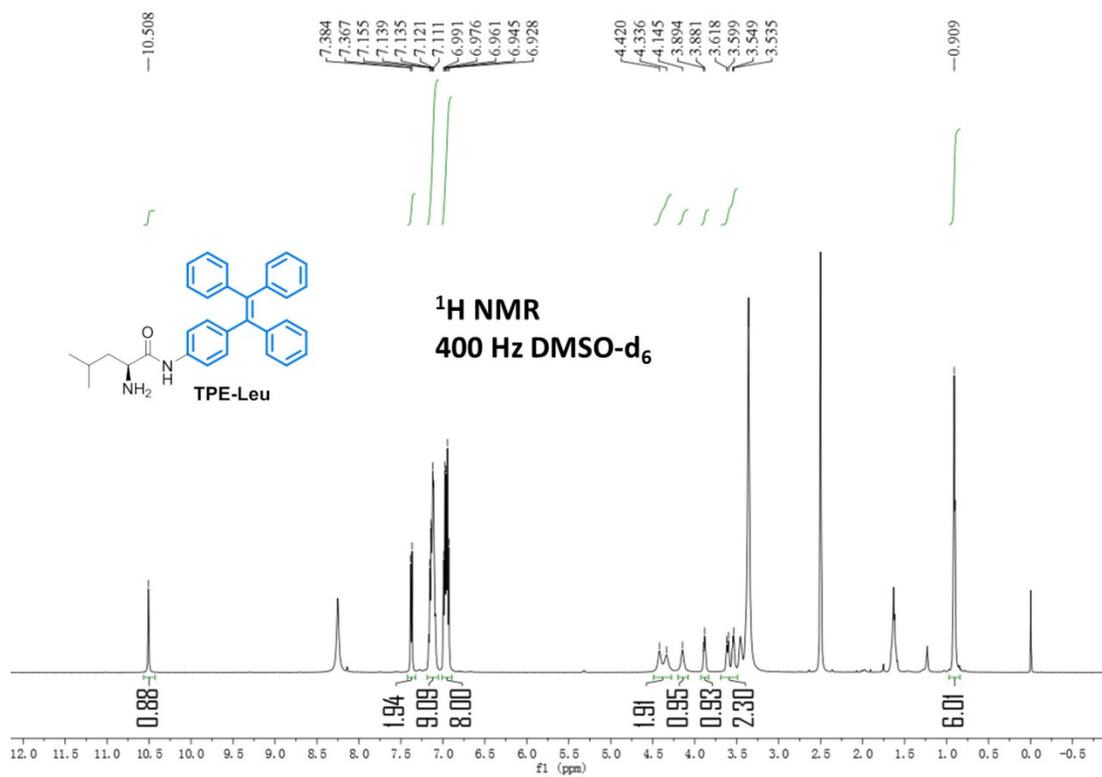
**Figure S14.** Fluorescence responses of 10  $\mu\text{M}$  **TPE-Leu** to different pH values and fluorescence quantum yields were measured with hymecromone ( $\Phi = 0.74$  in pH 5.98) as the reference.

**Figure S15.** (A) Plots of normalized fluorescence intensity of 10  $\mu\text{M}$  **TPE-Leu** at 455 nm in the presence of different concentrations of AChE in pH=9.5 DMSO/buffer (5 mM pH=9.5 PBS) (1:9, v:v). (B) Plots of normalized fluorescence intensity of 10  $\mu\text{M}$  **TPE-Leu** at 455 nm in the presence of different concentrations of urease in pH=5.5 DMSO/buffer (5 mM pH=5.5 NaOAc) (1:9, v:v). (C) and (D) were the linear fit of (A) and (B), respectively.

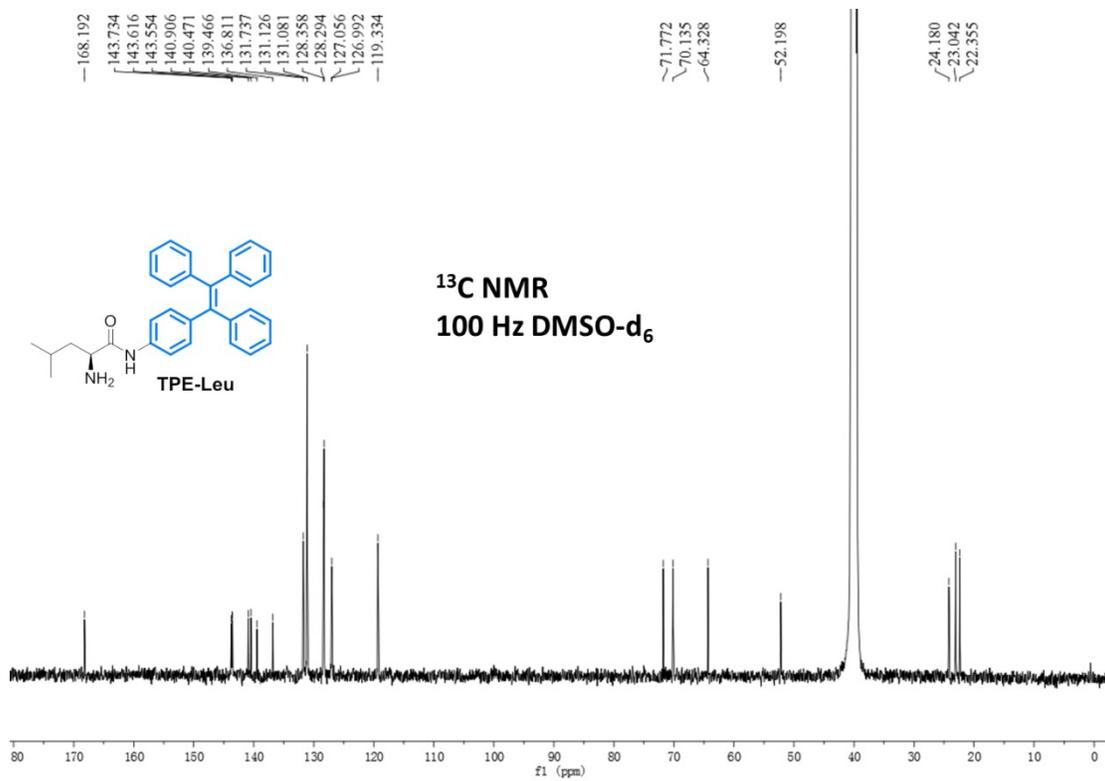
**Table S1.** Comparison of the current AIE probe with reported methods.



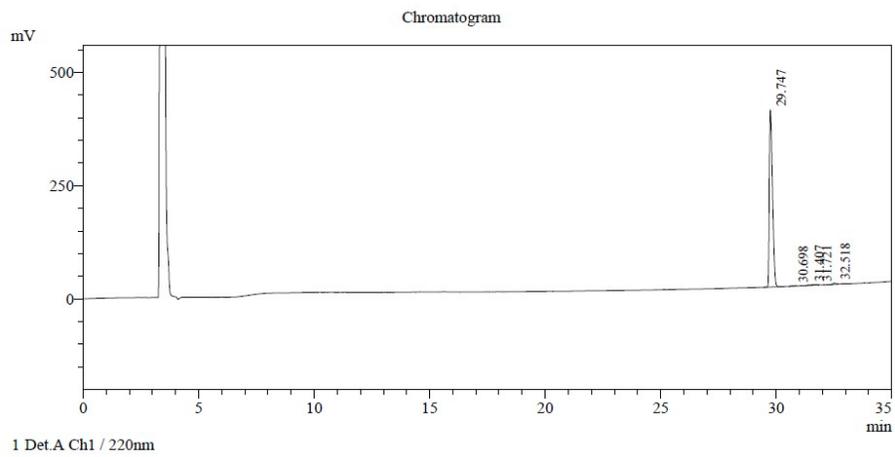
**Figure S1.** <sup>1</sup>H NMR of TPE-NH<sub>2</sub> in DMSO-d<sub>6</sub>.



**Figure S2.** <sup>1</sup>H NMR of TPE-Leu in DMSO-d<sub>6</sub>.



**Figure S3.** <sup>13</sup>C NMR of TPE-Leu in DMSO-d<sub>6</sub>.



Peak Table

Peak#	Ret. Time	Area	Height	Area %
1	29.747	3771142	391837	98.755
2	30.698	8876	991	0.232
3	31.407	8164	1008	0.214
4	31.721	6091	758	0.160
5	32.518	24425	2846	0.640
Total		3818699	397441	100.000

**Figure S4. HPLC Chromatogram of TPE-Leu.**

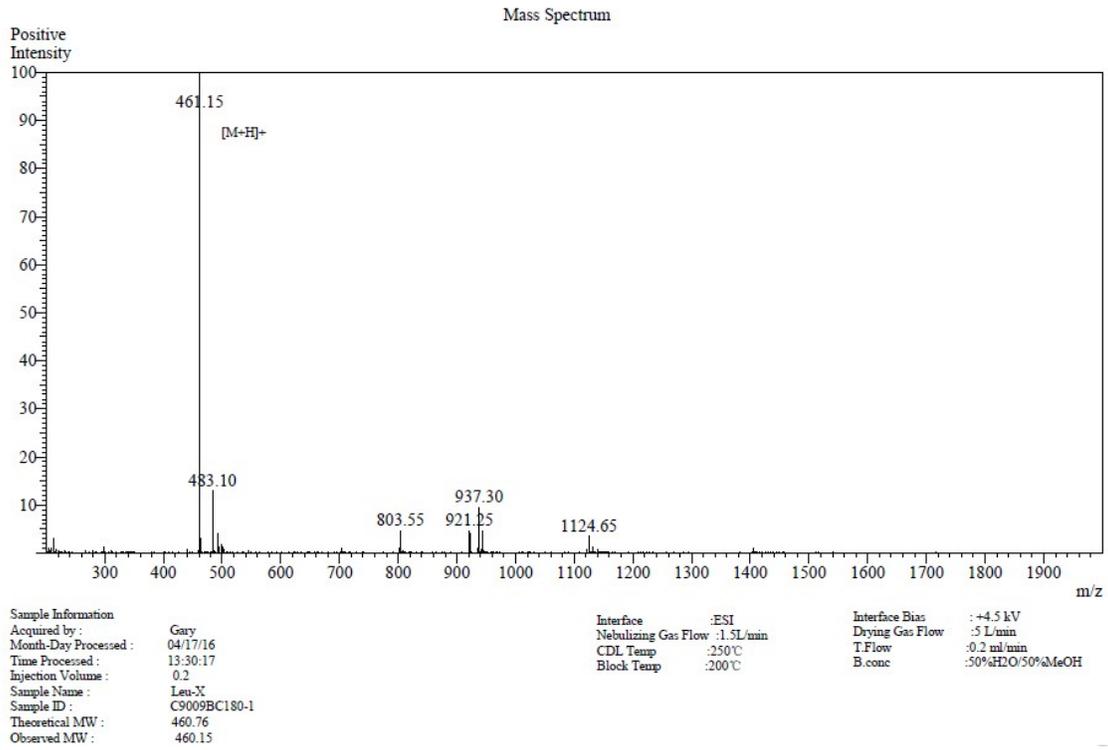
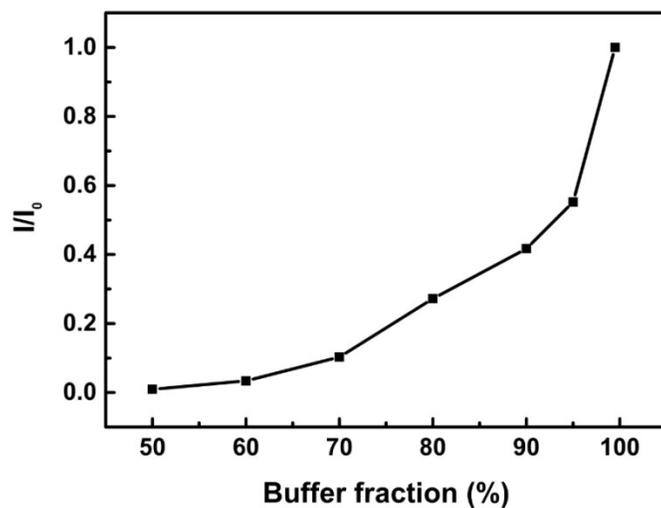
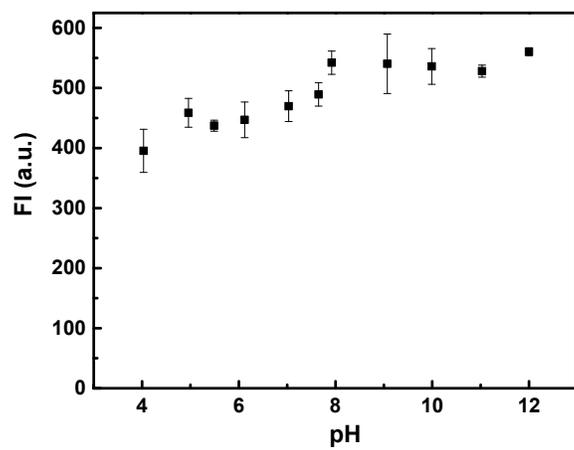


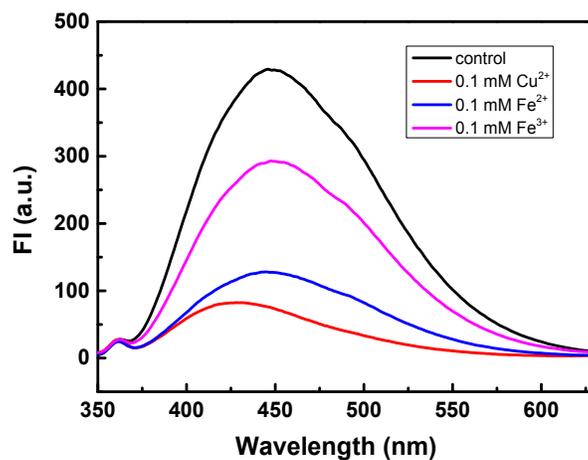
Figure S5. Mass spectrum of TPE-Leu.



**Figure S6.** Plot of  $I/I_0$  of 10  $\mu\text{M}$  TPE-Leu in DMSO/PBS buffer with different volume fractions of PBS buffer, where  $I_0$  is the fluorescence intensity of TPE-Leu in 99.5% PBS buffer.  $\lambda_{\text{ex}}=320$  nm.

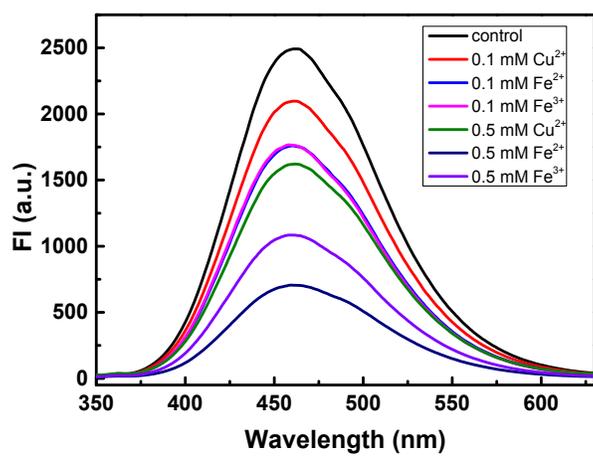


**Figure S7.** Plots of fluorescence intensity of 10  $\mu\text{M}$  TPE-NH<sub>2</sub> at 455 nm versus pH values.  $\lambda_{\text{ex}}=320$  nm.

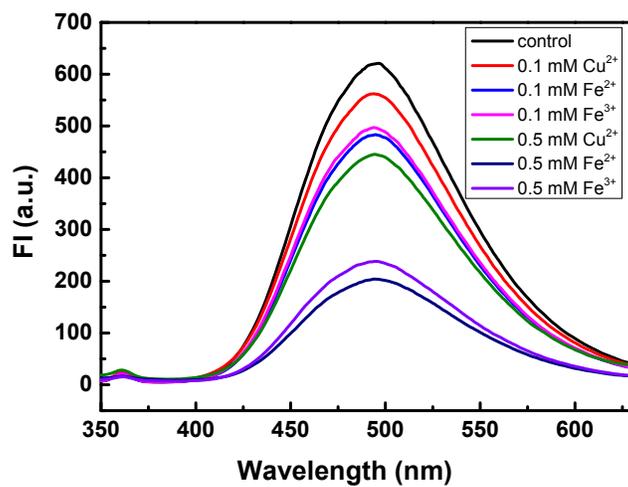


**Figure S8.** Selectivity of 10  $\mu\text{M}$  **TPE-Leu** in pH=7 DMSO/buffer (1:9, v:v) over 0.1 mM  $\text{Cu}^{2+}$ ,  $\text{Fe}^{2+}$ , and  $\text{Fe}^{3+}$ .

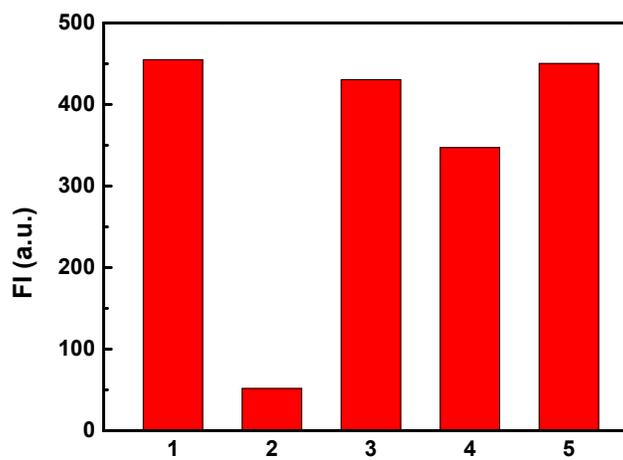
We noticed that  $\text{Cu}^{2+}$ ,  $\text{Fe}^{2+}$  and  $\text{Fe}^{3+}$  influenced the fluorescence intensity of **TPE-Leu**. From the emission spectra, it was observed that  $\text{Cu}^{2+}$ ,  $\text{Fe}^{2+}$ , and  $\text{Fe}^{3+}$  decreased the fluorescence intensity of **TPE-Leu** (Figure S8). More interestingly, the fluorescence intensity of **TPE** and **TPE-NH<sub>2</sub>** was also decreased in the presence of  $\text{Cu}^{2+}$ ,  $\text{Fe}^{2+}$  and  $\text{Fe}^{3+}$ , especially in high concentrations (Figure S9 and S10). The reasons for these phenomena were still not clear currently and is still under investigation. However, we demonstrated that the interference of  $\text{Cu}^{2+}$  and  $\text{Fe}^{3+}$  could be eliminated by using ethylene diamine tetraacetic acid (EDTA) (Figure S11). Since  $\text{Fe}^{2+}$  is not very stable and easily to be oxidized into  $\text{Fe}^{3+}$ , its interference could be eliminated by bubbling oxygen gas. All of these results indicated that **TPE-Leu** was highly selective to pH over other potential competing species and thus could be applied in the complicated biosystems for bioanalysis.



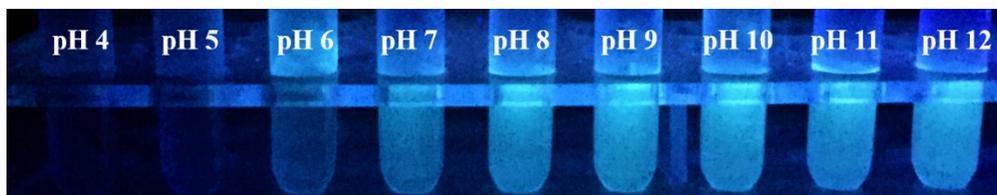
**Figure S9.** Selectivity of 10  $\mu\text{M}$  TPE in pH=7 DMSO/buffer (1:9, v:v) over 0.1 mM/0.5 mM Cu<sup>2+</sup>, Fe<sup>2+</sup>, and Fe<sup>3+</sup>.



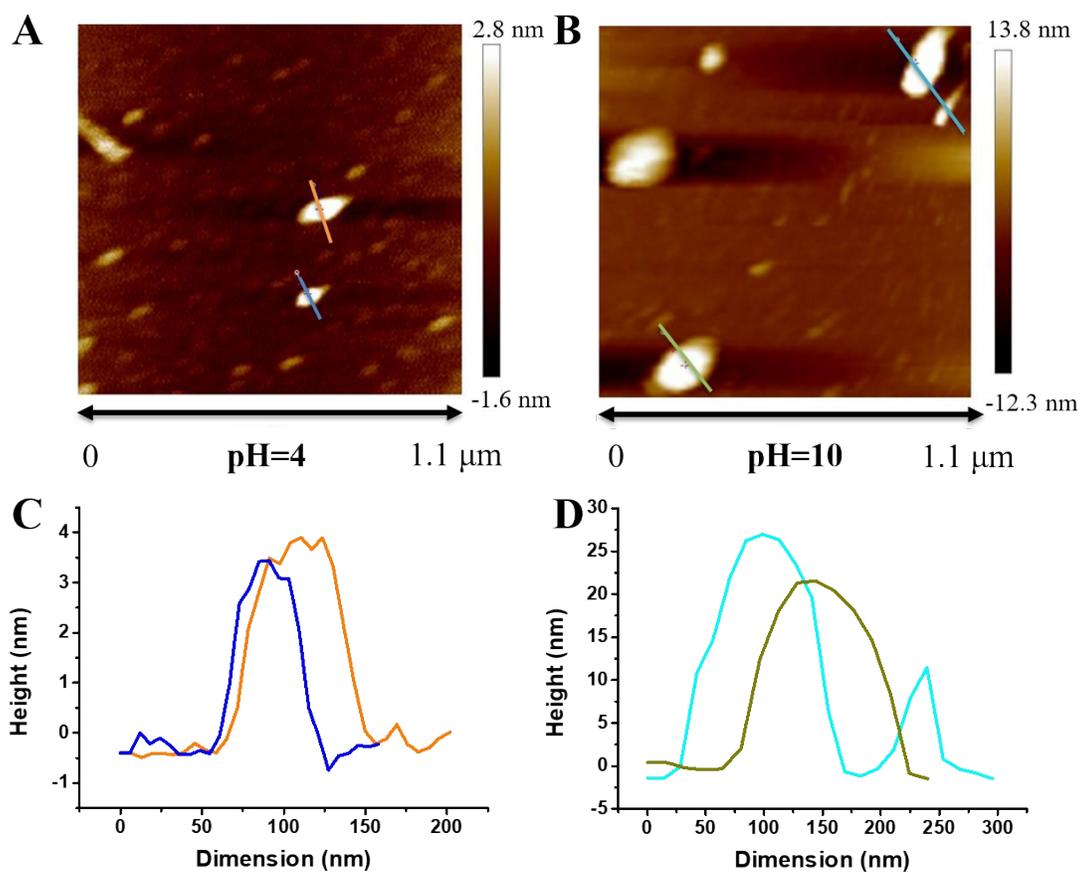
**Figure S10.** Selectivity of 10  $\mu\text{M}$  TPE-NH<sub>2</sub> in pH=7 DMSO/buffer (1:9, v:v) over 0.1 mM/0.5 mM Cu<sup>2+</sup>, Fe<sup>2+</sup>, and Fe<sup>3+</sup>.



**Figure S11.** Fluorescence intensity of **TPE-Leu** (10  $\mu$ M) in pH=7 DMSO/buffer (1:9, v/v) in the absence (1) or presence of 0.1 mM  $\text{Cu}^{2+}$  (2), 0.1 mM  $\text{Cu}^{2+}$  and EDTA (3), 0.1 mM  $\text{Fe}^{3+}$  (4), and 0.1 mM  $\text{Fe}^{3+}$  and EDTA (5).  $\lambda_{\text{ex}}$ =320 nm.

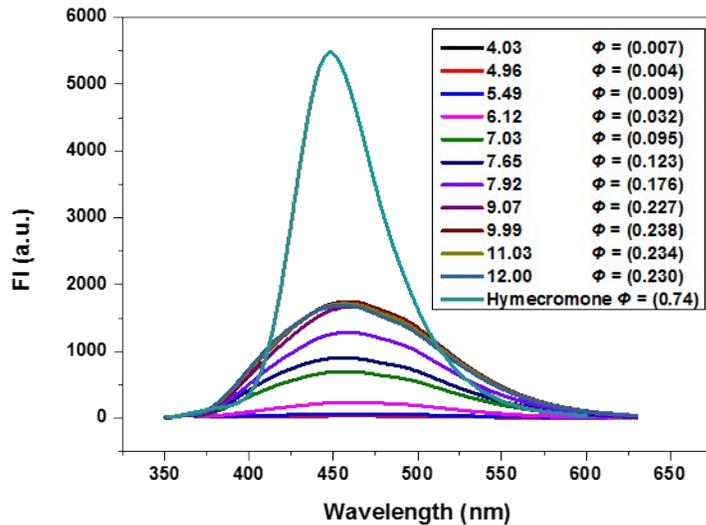


**Figure S12.** Photo of TPE-Leu in different pH buffers.



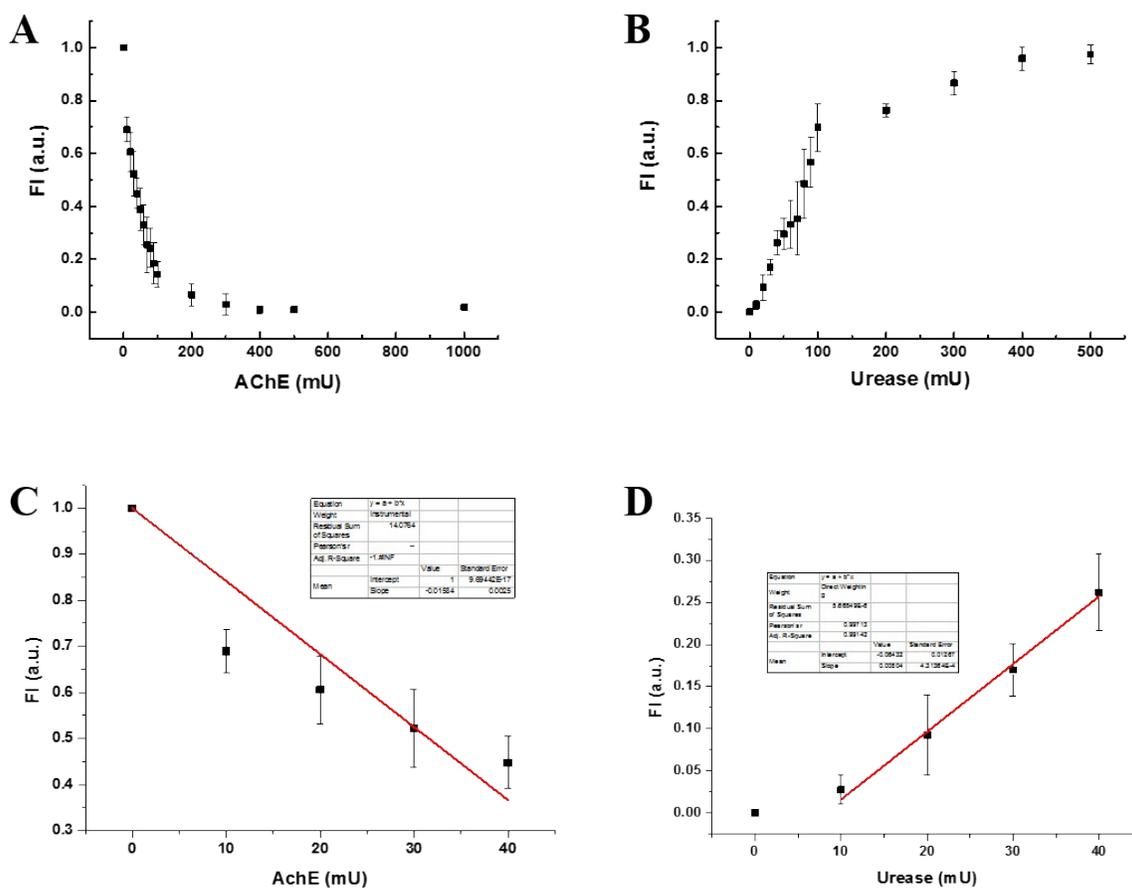
**Figure S13.** AFM images of (A) **TPE-Leu** in DMSO/buffer (5 mM pH=4 NaOAc) (1:9, v:v) and (B) **TPE-Leu** in DMSO/buffer (5 mM pH=10 PBS) (1:9, v:v). (C) and (D) were the corresponding cross-sectional profiles.

As shown in Figure S13, the thickness of **TPE-Leu** in acidic or basic condition was estimated to be ca. 3 – 4 nm and 20 – 25 nm, respectively, indicating the deaggregation/aggregation of **TPE-Leu**.



**Figure S14.** Fluorescence responses of 10  $\mu\text{M}$  TPE-Leu to different pH values and fluorescence quantum yields were measured with hymecromone ( $\Phi = 0.74$  in pH 5.98) as the reference.

Fluorescence quantum yields ( $\Phi$ ) of TPE-Leu in different pH buffers were measured with hymecromone ( $\Phi = 0.74$  in pH 5.98) as the reference<sup>1</sup>.



**Figure S15.** (A) Plots of normalized fluorescence intensity of 10  $\mu$ M **TPE-Leu** at 455 nm in the presence of different concentrations of AChE in pH=9.5 DMSO/buffer (5 mM pH=9.5 PBS) (1:9, v:v). (B) Plots of normalized fluorescence intensity of 10  $\mu$ M **TPE-Leu** at 455 nm in the presence of different concentrations of urease in pH=5.5 DMSO/buffer (5 mM pH=5.5 NaOAc) (1:9, v:v). (C) and (D) were the linear fit of (A) and (B), respectively.

**Table S1.** Comparison of the current AIE probe with reported methods.

Strategy	probe	Dynamic range (mU/mL)	Detection limit (mU/mL)	Ref.
Fluorescence	AuNCs-Cu <sup>2+</sup>	AChE: 0.05 – 2.5	AChE: 0.05	2
	C-dots-AgNPs	AChE: 0.025 – 2	AChE: 0.021	3
	AuNCs	Urease: 2.2 – 55	Urease: 0.55	4
	TPE-Leu	AChE: 0 – 1000 Urease: 0 – 500	AChE: 8.71 Urease: 6.39	<b>this work</b>
Nanozyme	AuNCs	Urease: 1.8 – 90	Urease: 1.8	5
	PAA-CeO <sub>2</sub>	AChE: 0.263 – 50	AChE: 0.263	6
	Citrate-CeO <sub>2</sub>	AChE: 0 – 1400 Urease: 0 – 1500	AChE: 3.5 Urease: 2.5	7

**Abbreviations**

AgNPs: Ag nanoparticles

AuNCs: gold nanoclusters

C-dots: carbon dots

PAA: poly (acrylic acid)

## References

1. L. J. Xie, Y. H. Chen, W. T. Wu, H. M. Guo, J. Z. Zhao, X. R. Yu, *Dyes Pigments*, 2012, 92, 1361-1369.
2. J. Sun, X. R. Yang, *Biosens. Bioelectron.*, 2015, 74, 177-182.
3. D. Zhao, C. X. Chen, J. Sun, X. R. Yang, *Analyst*, 2016, 141, 3280-3288.
4. H. H. Deng, G. W. Wu, Z. Q. Zou, H. P. Peng, A. L. Liu, X. H. Lin, X. H. Xia, W. Chen, *Chem. Commun.*, 2015, 51, 7847-7850.
5. H. H. Deng, G. L. Hong, F. L. Lin, A. H. Liu, X. H. Xia, W. Chen, *Anal. Chim. Acta*, 2016, 915, 74-80.
6. S. X. Zhang, S. F. Xue, J. J. Deng, M. Zhang, G. Y. Shi, T. S. Zhou, *Biosens. Bioelectron.*, 2016, 85, 457-463.
7. H. J. Chen, S. L. Lin, F. Muhammad, Y. W. Lin, H. Wei, *ACS Sens.*, 2016, 1, 1336-1343.