Catalytic Asymmetric Meerwein-Ponndorf-Verley Reduction of

Glyoxylates Induced by Chiral *N*,*N*'-Dioxide/Y(OTf)₃ Complex

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Electronic Supporting Information

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(A) General information

Reactions were carried out using commercial available reagents in oven-dried apparatus. CHCl3 and iso-propanol were dried and distilled from calcium hydride under nitrogen just before use. Molecular sieves were dried at 500 °C for 4 h and restored in nitrogen before use. ¹H NMR spectra were recorded at 400 MHz. The chemical shifts were recorded in ppm relative to tetramethylsilane and with the solvent resonance as the internal standard. Data were reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, m = multiplet), coupling constants (Hz) and integration. ¹³C NMR data were collected at 100 MHz with complete proton decoupling. Chemical shifts were reported in ppm from the tetramethylsilane with the solvent resonance as internal standard. Enantiomeric excesses (ee) were determined by chiral HPLC analysis on Daicel Chiralcel IA/IC/AD-H/AS-H/OD-H and Phenomenex Lux 5u Cellulose-2 in comparison with the authentic racemates. Optical rotations were reported as follows: $[\alpha]_D^T = (c; g/100)$ mL, in solvent). ESI-HRMS spectra were recorded on a commercial apparatus and methanol or acetonitrile was used to dissolve the sample. The N,N'-dioxides were prepared according to the methods reported in the literature.^[1]

(B) Optimization of the conditions for the asymmetric MPV reaction of

glyoxylates

	OMe + ROH	L-RaPr ₂ /Y((10 mol% Al(O/Pr) ₃ , 6	$(OTf)_3$ (6) $(0 \circ C)$	OH OR
1ab				2a
Entry ^a	ROH	t (h)	$\operatorname{Yield}^{b}(\%)$	ee^{c} (%)
1	OH F₃C└CF₃	48	0	-
2	ОН	48	0	-

Table S1: Screening of the secondary alcohols



^{*a*}Unless otherwise noted, the reactions were performed with **L-RaPr**₂/Y(OTf)₃ (1:1, 10 mol%), **1ab** (0.1 mmol), Al(O*i*Pr)₃ (0.05 mmol) in ROH (0.1 M) at 60 °C for 3 h without extrusion of air. ^{*b*}Yields of the isolated products. ^{*c*}Determined by HPLC analysis using a chiral stationary phase.

OH L-RaPr₂/Y(OTf)₃ .O*i*Pr OMe (10 mol%) + *i*PrOH 0 || 0 AI(O*i*Pr)₃, T 1ab 2a Entry^a T (°C) t (h) Yield^b (%) ee^{c} (%) 1 60 3 99 63 2 50 3 99 69 3 40 5 99 71 4 30 24 99 73 5^d 30 24 99 75 6^d 0 24 _ n.r.

Table S2: Screening of the reaction temperature

^{*a*}Unless otherwise noted, the reactions were performed with **L-RaPr**₂/Y(OTf)₃ (1:1, 10 mol%), **1b** (0.1 mmol), Al(O*i*Pr)₃ (0.05 mmol) in *i*PrOH (0.1 M) without extrusion of air. ^{*b*} Yields of the isolated products. ^{*c*} Determined by HPLC analysis using a chiral stationary phase. ^{*d*}L-RaPr₃/Y(OTf)₃ was used as catalyst.

Table S3: Screening of the addictives



Entry ^a	Additive	t (h)	Yield(%) ^b	ee(%) ^c	
1	LiNTf	12	69	72	
2^d	3 Å MS	12	99	71	
3 ^e	4 Å MS	12	99	71	
4	Na_2SO_4	12	54	67	
5	TsOH	12	trace	-	
6	m-CPBA	12	trace	-	
7	TEMPO	12	74	69	
8	DMAP	12	46	67	
9	KHSO ₄	12	34	72	
10	NH ₄ Cl	12	26	26	
11	K ₂ CO ₃	12	46	71	
12	LiOH•H ₂ O	12	32	73	
13	-	12	49	72	

^{*a*}Unless otherwise noted, the reactions were performed with **L-RaPr₃**/Y(OTf)₃ (1:1, 10 mol%), **1b** (0.1 mmol), Al(O*i*Pr)₃ (0.05 mmol) and additive (0.1 mmol) in *i*PrOH (0.1 M) at 30 °C for 12 h without extrusion of air. ^{*b*}Yields of the isolated products. ^{*c*}Determined by HPLC analysis using a chiral stationary phase. ^{*d*}3 Å MS (25 mg) were added. ^{*c*}4 Å MS (25 mg) were added.



14

3

tBu (1ac)

Table S4: Screening of the ester group on the substrates

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73

99

4	Bn (1ad)	24	99	71
5^d	<i>i</i> Pr (1a)	12	85	79
6 ^e	<i>i</i> Pr (1a)	24	78	86

^{*a*}Unless otherwise noted, the reactions were performed with L-RaPr₃/Y(OTf)₃ (1:1, 10 mol%), 1 (0.1 mmol), Al(O*i*Pr)₃ (0.05 mmol), 3 Å MS (25 mg) in *i*PrOH (0.1 M) at 30 °C without extrusion of air. ^{*b*} Yields of the isolated products. ^{*c*} Determined by HPLC analysis using a chiral stationary phase. ^{*d*}Performed at 0 °C. ^{*e*}Performed at -10 °C.

Table S5: Screening of the solvent



Entry ^a	solvent	solvent volume, <i>i</i> PrOH:solvent ratio	t (h)	Yield(%) ^b	ee(%)°
1	DCM	1.0 mL, 1:3	12	82	84
2	CHCl ₃	1.0 mL, 1:3	12	70	86
3	DCE	1.0 mL, 1:3	12	75	80
4	CHCl ₂ -CHCl ₂	1.0 mL, 1:3	12	80	81
5	CH ₃ -CCl ₃	1.0 mL, 1:3	12	69	80
6	CH ₂ Cl-CHCl ₂	1.0 mL, 1:3	12	85	83
7	toluene	1.0 mL, 1:3	12	73	82
8	CHCl ₃	1 mL, 5:1	24	99	85
9	CHCl ₃	1 mL, 3:1	24	99	85
10	CHCl ₃	1 mL, 1:1	24	97	86
11	CHCl ₃	1 mL, 1:2	24	93	87
12	CHCl ₃	1 mL, 1:3	24	90	87
13	CHCl ₃	1 mL, 1:5	24	84	87

"Unless otherwise noted, the reactions were performed with L-RaPr₃/Y(OTf)₃ (1:1, 10 mol%), 1 (0.1 mmol), Al(OiPr)₃ (0.05

mmol), 3 Å MS (25 mg) in *i*PrOH/CHCl₃ (v/v = 1/2, 0.1 M) at 0 °C for 12 h without extrusion of air. ^bYields of the isolated products. ^cDetermined by HPLC analysis using a chiral stationary phase.

0 0 1a	, ^{OiPr} + iPrOH	L-RaPr ₃ /Y(OTf) ₃ (10 mol%) Al(OiPr) ₃ , -10 °C 3 Å MS, solvent	OH 2a	OiPr	
Entry ^a	3 Å MS (mg)	Al(O <i>i</i> Pr) ₃ (mmol)	t (h)	Yield(%) ^b	ee(%)°
1	25	0.1	24	82	86
2	25	0.05	24	78	86
3	25	0.02	24	70	86
4	25	0.01	24	64	86
5	100	0.1	24	71	84
6	50	0.1	24	67	84
7	10	0.1	24	72	85
8	5	0.1	24	68	86

Table S6: Screening of the amount of addictives

"Unless otherwise noted, the reactions were performed with L-RaPr₃/Y(OTf)₃ (1:1, 10 mol%), 1 (0.1 mmol), Al(OiPr)₃, 3 Å MS in iPrOH and solvent at 0 °C for 12 h without extrusion of air. ^bYields of the isolated products. ^cDetermined by HPLC analysis using a chiral stationary phase.

0 0 1a	OiPr + iPrOH	L-RaPr ₃ /Y(OTf) ₃ (10 mol%) Al(OR) ₃ , -10 °C 3 Å MS, solvent	OH IIII 2a	O/Pr ⊖ O
Entry ^a	Al(OR) ₃	t (h)	Yield(%) ^b	ee(%) ^c
1	Al(OEt) ₃	24	77	85
2	Al(O <i>i</i> Pr) ₃	24	78	86
			S	-6

Table S7: Screening of the aluminium alkoxides

3	Al(OtBu) ₃	24	79	91
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^{*a*}Unless otherwise noted, the reactions were performed with L-RaPr₃/Y(OTf)₃ (1:1, 10 mol%), 1 (0.1 mmol), Al(O*i*Pr)₃ (0.05 mmol), 3 Å MS (25 mg) in *i*PrOH and solvent at 0 °C for 12 h without extrusion of air. ^{*b*}Yields of the isolated products. ^{*c*}Determined by HPLC analysis using a chiral stationary phase.

Table S8: Probing of the effect of the additives.



Entry ^a	T (°C)	t (h)	Additive	Yield ^b (%)	Ee ^c (%)
1	30	24	-	trace	n.d.
2	30	24	3 Å MS	82	73
3	30	14	$Al(OtBu)_3$	99	73
4	30	3	3 Å MS, Al(O t Bu) ₃	99	73
5	0	24	-	n.r.	-
6	0	24	3 Å MS	n.r.	-
7	0	24	Al(OtBu) ₃	n.r.	-
8	0	24	3 Å MS, Al(OtBu) ₃	99	87

^{*a*}Unless otherwise noticed, the reactions were performed with **L-RaPr₃**-Metal (1:1, 10 mol%), **1a** (0.10 mmol) and 3Å MS (25 mg) in *i*PrOH/CHCl₃ (v/v = 1/2, 0.1 M) in air. ^{*b*}Yields of the isolated products. ^{*c*}Determined by HPLC analysis using a chiral stationary phase.

(C) Optimization of the conditions for the asymmetric MPV reaction of other

ketones.



With the success in the reduction of gloxylates, we were encouraged to apply such method to a variety of substrates. After a serious of condition screening, we established the MPV reaction of α -bromoacetophenone **1v** with *iso*-propanolin the presence of 3 Å MS and Al(O*i*Pr)₃by employing **L-PrPr**₂/Zn(OTf)₂ complex as the catalyst at 0 °C for 120 h, and obtained 95% yield and 81% ee (Scheme 3). Other types of substrates were also under investigation. Simple ketones such as **1u** and **1w** had low reaction reactivity and often require high reaction temperature. The 2,2,2-trifluoacetophenone (**1x**) gave a higher reactivity as the trifluomethyl group serves as a efficient electrodrawing group. However, the stereo control was not satisfying after a serious of screening. For substrates **1y** and **1z**, which more inclined to form an enol, the MPV reduction is reluctant, either stablized via a hydrogen bond (**1y**), or a conjugating effect (**1z**). Enlightened by previous work,^{10e} we also expanded substrate scope to ketimines **1aa** and **1ab**, yet no desired product was formed. Such facts portrayed the property of chemical specificity towards carbonyl groups in the MPV reaction, and enols or imines were nonreactive.

Table S9: Screening of the metal salts

	Br + <i>i</i> PrOH	L-PrPr ₂ /M (10 mol ⁴ Al(O/Pr) ₃ , 3 Å M	1etal %) 30 °C S	OH Br
iu.				20
Entry ^a	Metal	t (h)	Yield(%) ^b	ee(%) ^c
1	Sc(OTf) ₃	24	99	61
2	Y(OTf) ₃	24	99	67
3	La(OTf) ₃	24	0	-
4	Yb(OTf) ₃	48	99	58
5	Zn(OTf) ₂	24	99	73
6	Ba(OTf) ₂	24	99	71
7	Zn(NTf) ₂	72	0	-
8	Zn(ClO ₄) ₂	72	0	-
9	Zn(BF ₄) ₂	72	0	-
10	ZnBr ₂	72	0	-

^{*a*}Unless otherwise noted, the reactions were performed with **L-PrPr₂-metal** (1:1, 10 mol%), **1** (0.1 mmol), Al(O*i*Pr)₃ (0.1 mmol), 3 Å MS (25 mg) in *i*PrOH and solvent at 30 °C without extrusion of air. ^{*b*}Yields of the isolated products. ^cDetermined by HPLC analysis using a chiral stationary phase.

Table S10: Screening of the ligands



L-PrPr₂: $R = 2,6-iPr_2C_6H_3$, n = 1 **L-PrPr**₃: $R = 2,6-iPr_3C_6H_2$, n = 1 **L-PiPr**₂: $R = 2,6-iPr_3C_6H_2$, n = 1**L-PiPr**₂: $R = 2,6-iPr_2C_6H_3$, n = 2

 $\begin{array}{l} \textbf{L-RaPr_2:} \ R = 2,6\text{-}i\text{-}Pr_2C_6H_3, \ m = 1 \\ \textbf{L-OIPr_2:} \ R = 2,6\text{-}i\text{-}Pr_2C_6H_3, \ m = 2 \end{array}$

Entry ^a	Metal	t (h)	$\operatorname{Yield}^{b}(\%)$	ee ^c (%)
1	L-PrEt ₂	24	trace	50
2	L-PrPr ₂	24	99	73
3	L- PrPr ₃	24	99	64
4	L-PiPr ₂	24	99	69
5	L- RaPr ₂	24	99	43
6 ^{<i>d</i>}	L-PrPr ₂	120	90	80
$7^{d,e}$	L-PrPr ₂	120	95	81

^{*a*}Unless otherwise noted, the reactions were performed with L-Zn(OTf)₂ (1:1, 10 mol%), **1** (0.1 mmol), Al(O*i*Pr)₃ (0.1 mmol), 3 Å MS (25 mg) in *i*PrOH at 30 °C without extrusion of air. ^{*b*}Yields of the isolated products. ^{*c*}Determined by HPLC analysis using a chiral stationary phase. ^{*d*}The reaction proceed at 0 °C. ^{*e*}Al(O*t*Bu)₃ (0.1 mmol) were used instead of Al(O*i*Pr)₃.

(D) Methods for the preparation of α-keto esters.



Glyoxal monohydrides 5a-5u were prepared according to the methods reported in the literature.^[2] Into a dry flask were added SeO₂ 4 (11 mmol), 2,6-dioxane (7.5 mL) and H₂O (0.6 mL) and the reaction mixture was stirred at 50 °C until 4 dissolve

completely. Then acetyl compound **3** (10 mmol) was added and the reaction mixture was stirred at 100 $^{\circ}$ C. The reaction was monitored under TLC.

The reaction mixture was filtered through a pad of celite and the solvent was removed under reduced pressure. Purification was accomplished by flash column chromatography and recrystallization with water, and a white to pale yellow solid **5** was obtained.

$$R^{1} \xrightarrow[OH]{OH} + R^{2}OH \xrightarrow[I_{2}, K_{2}CO_{3}]{} R^{1} \xrightarrow[OH]{COOR^{2}}$$

$$5 \quad 6 \qquad 1$$

 α -Keto esters **1a–1u**, **1ab**, **1ac** and **1ad** were prepared according to the methods reported in the literature.^[3] Into a dry flask were added **5** (5 mmol), toluene (10 mL) and **6** (7.5 mmol) and the reaction mixture was stirred room temperature until **5** dissolve completely. Then I₂ (5 mmol) and K₂CO₃ (10 mmol) were added to the reaction mixture and the reaction was monitored under TLC.

The reaction mixture was washed with aqueous Na₂SO₃ solution and the solvent was removed under reduced pressure. Purification was accomplished by flash column chromatography and distillation, and a colorless to pale yellow liquid **1** was obtained.

(E) Typical procedure for the asymmetric MPV reaction



Chiral *N*,*N'*-dioxide **L-RaPr**₃ (10 mol%), Y(OTf)₃ (10 mol%), 3 Å MS (25 mg), and Al(O*t*Bu)₃ (50 mol%)were added in a dry reaction tube, then *i*PrOH/CHCl₃ (v/v = 1/2, 1 mL) was added in air. The mixture was stirred at 30°C for 30 min. Then, **1** (0.1 mmol) were added. The reaction was stirred vigorously at -10°C for 72 h and at 0 °C for 48 h. The mixture was purified by column chromatography on silica gel to afford the desired product **2**. The yields of **2** were calculated according to the amount of **1**.

(F) Gram-scale experiment



Chiral *N*,*N'*-dioxide **L-RaPr₃** (10 mol%), Y(OTf)₃ (10 mol%), 3 Å MS (1.750 g), and Al(O*t*Bu)₃ (50 mol%) were added in a dry reaction vessel, then *i*PrOH/CHCl₃ (v/v = 1/2, 70 mL) was added in air. The mixture was stirred at 30 °C for 30 min. Then, **1a** (0.1 mmol) were added. The reaction was stirred vigorously at -10 °C for 72 h and at 0 °C for 48 h. The mixture was purified by column chromatography on silica gel to afford the desired product **2a**. The yield of **2a** was calculated according to the amount of **1a**.

(G) Kinetic information.

1. General procedures.



Chiral *N*,*N'*-dioxide **L-RaPr₃** (10 mol%), Y(OTf)₃ (10 mol%), 3 Å MS (25 mg), and Al(O*t*Bu)₃ (50 mol%) were added in a dry reaction tube, then *i*PrOH/CHCl₃ (1 mL) was added in air. The mixture was stirred at 20°C for 30 min. Then, **1a** (0.1 mmol) were added. The reaction was stirred vigorously at 20 °C minor portion (approx. 20 μ L) of the reaction mixture at certain reaction time. Each portion of the mixture was filtered through a pad of celite. After removal of solvent, the residue was analysized via HPLC.

In the HPLC, the amount of substance N is proportional to the chromatographic peak area.

$$N_i = f_i A_i \,.$$

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In the equation, A_i refers to chromatographic peak area of the substance, f_i refers to the correction factors of peak area. For **1a** and **2a**, it is deduced that,

$$f_{2a}/f_{1a} = (N_{2a}A_{1a})/(N_{1a}A_{2a})$$

Entry	N_{2a}/N_{1a}	A_{1a}/A_{2a}	f_{2a}/f_{1a}
1	0.01	276.7	2.77
2	0.03	88.29	2.65
3	0.05	54.87	2.74
4	0.1	26.32	2.63
5	0.15	18.88	2.83
6	0.2	13.16	2.63
7	0.25	10.20	2.55
8	0.3	8.49	2.55
9	0.4	6.45	2.58
10	0.6	4.37	2.62
11	0.8	3.34	2.68
			2.66 ^{<i>a</i>}

In order to obtain f_{Ia}/f_{2a} , a serious of mixture of **1a** and **2a** were analyzed.

^{*a*}Average of entry 1 to entry 11.

It is estimated that the for **1a** and **2a** is 2.66. As a result, the HPLC analysis of portion of the reaction mixture at certain reaction time would give out the mole ratio of **1a** and **2a**, thus the yield of **2a** at certain reaction time can be calculated. The set of experiment was implemented under given alcohol concentration.

For $t = 0.5(t_i + t_{i+1})$, the reaction rate *r* of **1a** was estimated using an approximate formula,

$$r = (c_i - c_{i+1})/(\mathbf{t}_{i+1} - \mathbf{t}_i) \quad .$$

2. Data collection.

Solvent: $iPrOH/CHCl_3$ (v/v = 1/5, 1 mL)

Enter	t (min)	4 / 4	Ea of 2 a	Yield of 2a
Entry	t (mm)	A_{1a}/A_{2a}	Ee 01 2 2	(%)
1	0	-	-	0
2	1	22.04	82	10.8
3	2	12.83	82	17.2
4	3	10.00	82	21.0
5	4	8.35	82	24.2
6	5	7.24	82	26.9



Solvent: iPrOH/CHCl₃ (v/v = 1/2, 1 mL)

Enter t(min) 4/4 East		Ea of 2 a	Yield of 2a
t (mm)	$A_{1a'}A_{2a}$		(%)
0	-	-	0
1	13.86	82	16.1
2	8.53	82	23.8
3	6.87	81	27.9
4	5.73	82	31.7
5	4.98	83	34.8
	t (min) 0 1 2 3 4 5	t (min) A_{Ia}/A_{2a} 0 - 1 13.86 2 8.53 3 6.87 4 5.73 5 4.98	t (min) A_{Id}/A_{2a} Ee of $2a$ 0 - - 1 13.86 82 2 8.53 82 3 6.87 81 4 5.73 82 5 4.98 83



Solvent: $iPrOH/CHCl_3$ (v/v = 1/1, 1 mL)

			F 60	Yield of 2a
Entry	t (min)	A_{1a}/A_{2a}	Ee of 2a	(%)
1	0	-	-	0
2	1	10.96	82	19.5
3	2	6.76	82	28.3
4	3	5.25	82	33.6
5	4	4.33	82	38.0
6	5	3.71	82	41.8



Solvent: *i*PrOH (1 mL)

				Yield of 2a
Entry	t (min)	A_{1a}/A_{2a}	Ee of 2a	(%)
				(/0)



t (min)

3. Data analyses and explanations.

As we can observed from charts and graphs above, the following can be concluded.

- In every individual experiment, the ee value stayed unchanged at the first 5 min of the reaction. We believe consumption of 1a or formation of optically active 2a have no influence of the ee of the product 2a.
- 2) For mixed reaction solvents, the reaction rate increased as the concentration of *i*PrOH increase. We calculated the reaction rate of *i*PrOH by using r_0 at different *i*PrOH concentration. The reaction order of *i*PrOH is about 0.54.

Entry	<i>i</i> PrOH/CHCl ₃	c(iPrOH) (mol·L ⁻¹)	r_0 (mol·L ⁻¹ ·min ⁻¹)	ln(c)	$\ln(r_0)$
1	1/5	2.18	0.0108	0.338	-1.97
2	1/2	4.36	0.0161	0.639	-1.79
3	1/1	6.54	0.0195	0.816	-1.71
				S-16	



Further calculation did not elicit the reaction order of 1a at different *i*PrOH concentrations. We assume that in such catalytic system, formula for elementary reaction is not feasible for 1a.

- 3) When *i*PrOH was used as solvent the reaction rate at the first 5 min of was slightly slower than the mixed reaction solvent of *i*PrOH and chloroform (v/v = 1:1). Moreover, the ee value was lower than the mixed reaction solvent.
- 4) Such low reaction order of *i*PrOH and solvent effect suggested that it might be not *i*PrOH itself but the formed aluminium alkoxide that served as the true reactant, as proposed in the manuscript.

(H) HRMS data of the reaction.



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Y(OTf)₃ + L-RaPr₃ +1a + Al(OtBu)₃ (0.1:0.1:1:0.5) :

[Al³⁺+2*i*PrO⁻+2**a**] Calcd for : 339.1752, found : 339.0927;

[Al³⁺+2*i*PrO⁻+[**2a**-H⁺]⁻+Na⁺] Calcd for : 361.1572, found : 361.1881;

[Y³⁺+L-RaPr₃-H⁺] Calcd for : 436.2424, found : 436.2404;

[Y³⁺+**L-RaPr₃**+*i*PrO⁻] Calcd for : 466.2711, found : 466.2669;

[Y³⁺+**L-RaPr₃**+**2a**-H⁺] Calcd for : 533.2895, found : 533.2891;

[L-RaPr₃+ H⁺] Calcd for : 785.5945, found : 785.6016;

[Y³⁺+**L-RaPr**₃+TfO⁻+[**2a**-H⁺]⁻] Calcd for : 1215.5310, found : 1215.5674;

[Y³⁺+L-RaPr₃+2TfO⁻] Calcd for : 1171.3966, found : 1171.4941;

[Y³⁺+**L-RaPr₃+2a-**2H⁺] Calcd for : 1065.5711, found : 1065.5967;

[Y³⁺+**L-RaPr**₃+TfO⁻-H⁺] Calcd for : 1021.4875, found : 1021.4367.

The peak for species $Al^{3+}+2iPrO^{-}+2a$, $Y^{3+}+L-RaPr_{3}+iPrO^{-}$, $Y^{3+}+L-RaPr_{3}+2a-H^{+}$ and $Y^{3+}+L-RaPr_{3}+TfO^{-}+[2a-H^{+}]^{-}$ were found. The find of the peak of $Al^{3+}+2iPrO^{-}+2a$ species allowed us to presume that aluminium alkoxide should have a direct influence in the forming of 2a. Moreover, any form of $Y^{3+}+L-RaPr_{3}+iPrO^{-}+2a$, indicating *i*PrOH may serves as direct reductant in the catalytic cycle was not found.

(I) Spectral characterization data and HPLC conditions for the products

(S)-iso-propyl mandelate (2a)



(C12H16O3) a white solid; 98% yield, 90% ee. $[\alpha]_D^{20} = +100.81$ (c = 0.248, in CHCl₃), {Lit.^[4] $[\alpha]_D^{25} = -96.1$ (c = 1.15, in CHCl₃), conf. (R)}. HPLC Phenomenex Lux 5u Cellulose-2, 2-propanol/*n*-hexane = 10/90, flow rate = 1.0 mL/min, $\lambda = 210$ nm, retention time: 8.76 min, 9.92 min. ¹H NMR (400 MHz, CDCl₃) δ 7.50 – 7.28 (m, 5H), 5.17 – 4.98 (m, 2H), 3.57 – 3.44 (m, 1H), 1.28 (d, J = 6.3 Hz, 3H), 1.11 (d, J = 6.3 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) $\delta = 173.25$, 138.56, 128.50, 128.30, 126.43, 72.89, 70.20, 21.72, 21.41. ESI-HRMS: calcd for C₁₁H₁₄NaO₃+ [M+Na⁺] 217.0835, found 217.0833.



	Retention Time	%Area
1	8.757	5.14
2	9.916	94.86

(S)-*iso*-propyl α-hydroxy-α-(2-methylphenyl)acetate (2b)



(C13H18O3) a colorless oil; 99% yield, 91% ee. $[\alpha]_D^{21} = +117.86$ (*c* = 0.328, in CH₂Cl₂). HPLC Phenomenex Lux 5u Cellulose-2, 2-propanol/*n*-hexane = 5/95, flow rate = 1.0 mL/min, λ = 210 nm, retention time: 13.04 min, 14.43 min. ¹H NMR (400 MHz, CDCl3) δ 7.29 – 7.00 (m, 4H), 5.25 (d, J = 5.3 Hz, 1H), 5.00 (dt, J = 12.5, 6.3 Hz, 1H), 3.39 (d, J = 5.3 Hz, 1H), 2.36 (s, 3H), 1.19 (d, J = 6.3 Hz, 3H), 1.03 (d, J = 6.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ = 172.71, 135.85, 135.33, 129.69, 127.25, 125.51, 125.15, 69.36, 69.08, 20.67, 20.39, 18.28. ESI-HRMS: calcd for C12H16NaO₃⁺ [M+Na⁺] 231.0992, found 231.0992.



	Retention Time	%Area
1	12.860	50.25
2	14.189	49.75



	Retention Time	%Area
1	13.037	95.62
2	14.425	4.38

(S)-iso-propyl α-hydroxy-α-(3-methylphenyl)acetate (2c)



(C13H18O3) a colorless oil; 98% yield, 85% ee. $[\alpha]_D^{21} = +78.67$ (c = 0.286, in CH₂Cl₂). HPLC Phenomenex Lux 5u Cellulose-2, 2-propanol/*n*-hexane = 10/90, flow rate = 1.0 mL/min, $\lambda = 210$ nm, retention time: 8.04 min, 9.06 min. ¹H NMR (400 MHz, CDCl₃) δ 7.20 – 7.02 (m, 4H), 5.10 – 4.86 (m, 2H), 3.39 (d, J = 6.1 Hz, 1H), 2.28 (s, 3H), 1.21 (d, J = 6.3 Hz, 3H), 1.05 (d, J = 6.3 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) $\delta = 172.29$, 137.43, 137.19, 128.03, 127.36, 126.02, 122.55, 71.89, 69.08, 20.69, 20.39. ESI-HRMS: calcd for C12H16NaO3⁺ [M+Na⁺] 231.0992, found 231.0997.



(S)-iso-propyl α-hydroxy-α-(4-methylphenyl)acetate (2d)

(C13H18O3) a white solid; 98% yield, 90% ee. $[\alpha]_D^{23} = +92.31$ (c = 0.260, in CH₂Cl₂). HPLC Phenomenex Lux 5u Cellulose-2, 2-propanol/*n*-hexane = 5/95, flow rate = 1.0 mL/min, $\lambda = 210$ nm, retention time: 9.27 min, 9.90 min. ¹H NMR (400 MHz, CDCl₃) δ 7.27 – 7.15 (m, 2H), 7.13 – 7.02 (m, 2H), 5.10 – 4.86 (m, 2H), 3.38 (d, J = 6.1 Hz, 1H), 2.27 (s, 3H), 1.20 (d, J = 6.3 Hz, 3H), 1.04 (d, J = 6.3 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) $\delta = 172.36$, 137.01, 134.61, 128.17, 125.33, 71.72, 69.02, 20.68, 20.39, 20.14. ESI-HRMS: calcd for C12H16NaO3⁺ [M+Na⁺] 231.0992, found 231.1000.



	Retention Time	%Area
1	9.272	4.96
2	9.904	95.04

(S)-iso-propyl α-hydroxy-α-(4-(tert-butyl)phenyl)acetate (2e)

(C₁₆H₂₄O₃) a white solid; 99% yield, 91% ee. $[\alpha]_D{}^{12} = +71.93$ (c = 0.228, in CH₂Cl₂). HPLC DAICEL CHIRALCEL IA, 2-propanol/*n*-hexane = 10/90, flow rate = 1.0 mL/min, $\lambda = 210$ nm, retention time: 6.73 min, 7.84 min. ¹H NMR (400 MHz, CDCl₃) S-23

δ 7.36 – 7.21 (m, 4H), 5.01 (dd, J = 12.5, 6.2 Hz, 2H), 3.34 (d, J = 6.2 Hz, 1H), 1.28 – 1.18 (m, 12H), 1.07 (d, J = 6.2 Hz, 3H). ¹³C NMR (101 MHz, CDC13) δ = 172.30, 150.22, 134.53, 125.07, 124.43, 71.67, 69.02, 33.54, 30.27, 20.71, 20.45. ESI-HRMS: calcd for C₁₅H₂₂NaO₃⁺ [M+Na⁺] 273.1467, found 273.1465.



(S)-iso-propyl α-hydroxy-α-(4-phenylphenyl)acetate (2f)

4.66



7.842

2

(C17H18O3) a white solid; 99% yield, 89% ee. $[\alpha]_D^{18} = +114.97$ (*c* = 0.294, in CH₂Cl₂). HPLC DAICEL CHIRALCEL IA, 2-propanol/*n*-hexane = 10/90, flow rate = 1.0 mL/min, $\lambda = 210$ nm, retention time: 9.14 min, 10.21 min. ¹H NMR (400 MHz, CDCl₃) δ 7.55 – 7.49 (m, 4H), 7.44 – 7.33 (m, 4H), 7.27 (t, *J* = 7.3 Hz, 1H), 5.09 (s, 2H), 5.01 (dt, *J* = 12.5, 6.3 Hz, 1H), 3.47 (s, 1H), 1.22 (d, *J* = 6.3 Hz, 3H), 1.06 (d, *J* = 6.3 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ = 173.03, 151.04, 135.93, 126.06, 125.39, 82.98, 72.83, 34.56, 31.33, 27.92. ¹³C NMR (101 MHz, CDCl₃) δ = 173.22, 141.18, 140.62, 137.56, 128.82, 127.46, 127.27, 127.12, 126.88, 72.68, 70.32, 21.76, 21.49. ESI-HRMS: calcd for C17H18NaO3⁺ [M+Na⁺] 293.1148, found 293.1158.



	Retention Time	%Area
1	9.137	94.69
2	10.209	5.31

(S)-*iso*-propyl α-hydroxy-α-(4-methoxylphenyl)acetate (2g)

(C₁₃H₁₈O₄) a white solid; 90% yield, 89% ee. $[\alpha]_D^{22} = +92.25$ (*c* = 0.258, in CH₂Cl₂). HPLC DAICEL CHIRALCEL IA, 2-propanol/*n*-hexane = 10/90, flow rate = 1.0 mL/min, λ = 210 nm, retention time: 5.19 min, 5.99 min. ¹H NMR (400 MHz, CDCl₃) δ 7.32 (dd, *J* = 9.1, 2.4 Hz, 2H), 6.96 – 6.83 (m, 2H), 5.15 – 4.99 (m, 2H), 3.80 (s, 3H), 3.44 (dd, *J* = 5.8, 2.6 Hz, 1H), 1.27 (d, *J* = 6.3 Hz, 3H), 1.11 (d, *J* = 6.3 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ = 173.43, 159.60, 130.82, 127.73, 113.92, 72.51, 70.00, 55.27, 21.70, 21.43. ESI-HRMS: calcd for C₁₃H₁₈NaO₄⁺ [M+Na⁺] 261.1097, found 261.1098.



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	Retention Time	%Area
1	5.193	94.53
2	5.985	5.47

(S)-iso-propyl α-hydroxy-α-(3,4-methylenedioxylphenyl)acetate (2h)



(C₁₂H₁₄O₅) a white solid; 99% yield, 90% ee. $[\alpha]_D{}^{12} = +78.40$ (c = 0.250, in CH₂Cl₂). HPLC Phenomenex Lux 5u Cellulose-2, 2-propanol/*n*-hexane = 10/90, flow rate = 1.0 mL/min, $\lambda = 210$ nm, retention time: 18.66 min, 21.94 min. ¹H NMR (400 MHz, CDCl₃) δ 6.89 (dd, J = 5.8, 1.6 Hz, 2H), 6.83 – 6.74 (m, 1H), 5.96 (s, 2H), 5.14 – 4.97 (m, 2H), 3.43 (d, J = 5.8 Hz, 1H), 1.28 (d, J = 6.3 Hz, 3H), 1.14 (d, J = 6.3 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) $\delta = 173.23$, 147.81, 147.62, 132.48, 120.25, 108.23, 106.88, 101.17, 72.64, 70.23, 21.71, 21.46. ESI-HRMS: calcd for C₁₂H₁₄NaO₅⁺ [M+Na⁺] 261.0733, found 261.0740.





	Retention Time	%Area
1	18.664	5.20
2	21.936	94.80

(S)-iso-propyl α-hydroxy-α-(2,4-dimethoxylphenyl)acetate (2i)

(C₁₃H₁₈O₄)a white solid; 99% yield, 90% ee. $[\alpha]_D{}^{19}$ = +83.70 (*c* = 0.270, in CH₂Cl₂). HPLC DAICEL CHIRALCEL IA, 2-propanol/*n*-hexane = 10/90, flow rate = 1.0 mL/min, λ = 210 nm, retention time: 5.26 min, 8.97 min. ¹H NMR (400 MHz, CDCl3) δ 7.12 – 7.05 (m, 1H), 6.41 – 6.36 (m, 2H), 5.07 (s, 1H), 5.02 (dt, J = 12.5, 6.3 Hz, 1H), 3.73 (d, J = 5.0 Hz, 6H), 3.41 (s, 1H), 1.16 (d, J = 6.3 Hz, 3H), 1.07 (d, J = 6.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl3) δ = 173.58, 161.15, 158.24, 130.29, 120.03, 104.21, 98.90, 70.07, 69.39, 55.37, 21.69, 21.47. ESI-HRMS: calcd for C₁₃H₁₈NaO₄⁺ [M+Na⁺] 261.1097, found 261.1102.



	Retention Time	%Area
1	15.559	95.00
2	18.194	5.00

(S)-iso-propyl α-hydroxy-α-(2-flurophenyl)acetate (2j)

(C₁₂H₁₅FO₃) a white solid; 99% yield, 92% ee. $[\alpha]_D{}^{12} = +100.00$ (*c* = 0.306, in CH₂Cl₂). HPLC Phenomenex Lux 5u Cellulose-2, 2-propanol/*n*-hexane = 10/90, flow rate = 1.0 mL/min, λ = 210 nm, retention time: 8.55 min, 9.63 min. ¹H NMR (400 MHz, CDCl₃) δ 7.45 – 7.28 (m, 2H), 7.21 – 7.00 (m, 2H), 5.34 (d, *J* = 5.2 Hz, 1H), 5.21 – 5.01 (m, 1H), 3.56 (d, *J* = 5.3 Hz, 1H), 1.19 (dd, *J* = 59.9, 6.1 Hz, 6H). ¹³C NMR (101 MHz, CDCl₃) δ = 172.78, 161.75, 159.28, 130.21, 130.13, 128.71, 128.67, 126.13, 125.99, 124.31, 124.27, 115.78, 115.56, 70.43, 67.74, 21.62, 21.34. ESI-HRMS: calcd for C₁₁H₁₃FNaO₃⁺ [M+Na⁺] 235.0741, found 235.0741.



	Retention Time	%Area
1	8.546	3.84
2	9.627	96.16

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(S)-iso-propyl α-hydroxy-α-(4-flurophenyl)acetate (2k)

(C₁₁H₁₃FO₃) a white solid; 98% yield, 91% ee. $[\alpha]_D^{19} = +103.17$ (*c* = 0.252, in CH₂Cl₂). HPLC DAICEL CHIRALCEL IA, 2-propanol/*n*-hexane = 5/95, flow rate = 1.0 mL/min, $\lambda = 210$ nm, retention time: 8.66 min, 9.34 min. ¹H NMR (400 MHz, CDCl₃) δ 7.47 – 7.23 (m, 2H), 7.03 – 6.23 (m, 2H), 5.10 – 4.87 (m, 2H), 3.48 (d, *J* = 5.6 Hz, 1H), 1.21 (d, *J* = 6.3 Hz, 3H), 1.04 (d, *J* = 6.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ = 172.04, 162.86, 160.41, 133.32, 133.28, 127.18, 127.10, 114.48, 114.27, 71.17, 69.35, 20.66, 20.37. ESI-HRMS: calcd for C₁₁H₁₃FNaO₃⁺ [M+Na⁺] 235.0741, found 235.0747.





	Retention Time	%Area
1	8.664	95.40
2	9.344	4.60

(S)-iso-propyl α-hydroxy-α-(2-chlorophenyl)acetate (2l)



(C₁₁H₁₃ClO₃) a white solid; 99% yield, 91% ee. $[\alpha]_D^{20} = +99.15$ (*c* = 0.234, in CH₂Cl₂), HPLC DAICEL CHIRALCEL IA, 2-propanol/*n*-hexane = 10/90, flow rate = 1.0 mL/min, $\lambda = 210$ nm, retention time: 6.73 min, 7.84 min. ¹H NMR (400 MHz, CDCl₃) δ 7.35 – 7.27 (m, 2H), 7.22 – 7.17 (m, 2H), 5.43 (d, *J* = 2.9 Hz, 1H), 5.01 (d, *J* = 6.3 Hz, 1H), 3.57 (d, J = 4.0 Hz, 1H), 1.19 (d, *J* = 6.3 Hz, 3H), 1.04 (d, *J* = 6.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ = 171.73, 135.28, 132.55, 128.86, 128.58, 127.66, 126.02, 69.45, 69.42, 20.59, 20.36. ESI-HRMS: calcd for C₁₁H₁₃Cl^{34.9689}NaO₃⁺ [M+Na⁺] 251.0445, found 251.0449; calcd for C₁₁H₁₃Cl^{36.9659}NaO₃⁺ [M+Na⁺] 253.0416, found 253.0422.



	Retention Time	%Area
1	6.732	95.48
2	7.839	4.52

(S)-iso-propyl α-hydroxy-α-(4-chlorophenyl)acetate (2m)

(C11H13ClO3) a white solid; 98% yield, 90% ee. $[\alpha]_D{}^{18} = +95.93$ (c = 0.246, in CH₂Cl₂). HPLC Phenomenex Lux 5u Cellulose-2, 2-propanol/*n*-hexane = 5/95, flow rate = 1.0 mL/min, $\lambda = 210$ nm, retention time: 9.93 min, 10.51 min. ¹H NMR (400

MHz, CDCl₃) δ 7.38 – 7.11 (m, 4H), 5.13 – 4.81 (m, 2H), 3.50 (d, J = 5.6 Hz, 1H), 1.21 (d, J = 6.3 Hz, 3H), 1.04 (d, J = 6.3 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ = 171.82, 135.98, 133.09, 127.62, 126.76, 71.15, 69.48, 20.66, 20.38. ESI-HRMS: calcd for C₁₁H₁₃Cl^{34.9689}NaO₃⁺ [M+Na⁺] 251.0445, found 251.0449; calcd for C₁₁H₁₃Cl^{36.9659}NaO₃⁺ [M+Na⁺] 253.0416, found 253.0391.



	Retention Time	%Area
1	9.850	5.18
2	10.646	94.82

(S)-iso-propyl α-hydroxy-α-(2,3,4-trichlorophenyl)acetate (2n)

(C₁₁H₁₁Cl₃O₃) a white solid; 99% yield, 89% ee. $[\alpha]_D{}^{16} = +127.69$ (*c* = 0.242, in CH₂Cl₂). HPLC Phenomenex Lux 5u Cellulose-2, 2-propanol/*n*-hexane = 5/95, flow rate = 1.0 mL/min, $\lambda = 210$ nm, retention time: 9.93 min, 11.08 min. ¹H NMR (400 MHz, CDCl₃) δ 7.40 (d, *J* = 8.4 Hz, 1H), 7.27 (d, *J* = 8.2 Hz, 1H), 5.49 (d, *J* = 3.6 Hz, 1H), 5.08 (dt, *J* = 12.0, 6.0 Hz, 1H), 3.77 (d, *J* = 4.0 Hz, 1H), 1.20 (dd, *J* = 54.4, 6.2 Hz, 6H). ¹³C NMR (101 MHz, CDCl₃) $\delta = 172.01$, 136.95, 134.14, 133.62, 132.26,

128.44, 126.78, 70.99, 70.77, 21.58, 21.42. ESI-HRMS: calcd for C11H11Cl3^{34.9689}NaO3⁺ [M+Na⁺] 318.9671, found 318.9676; calcd for C11H11Cl2^{34.9689}Cl^{36.9659}NaO3⁺ [M+Na⁺] 320.9642, found 320.9656.



1	9.926	5.75
2	11.079	94.25

(S)-iso-propyl α-hydroxy-α-(2-bromophenyl)acetate (20)



(C₁₁H₁₃BrO₃) a white solid; 98% yield, 90% ee. $[\alpha]_D{}^{20}$ = +79.11 (*c* = 0.316, in CH₂Cl₂). HPLC Phenomenex Lux 5u Cellulose-2, 2-propanol/*n*-hexane = 2/98, flow rate = 1.0 mL/min, λ = 210 nm, retention time: 19.11 min, 20.39 min. ¹H NMR (400 MHz, CDCl₃) δ 7.46 – 7.33 (m, 2H), 7.27 – 7.17 (m, 2H), 5.13 – 4.84 (m, 2H), 3.52 (d, = 5.6 Hz, 1H), 1.21 (d, *J* = 6.3 Hz, 3H), 1.04 (d, *J* = 6.3 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ = 172.41, 137.97, 131.52, 128.08, 122.11, 83.55, 72.35, 27.83. ¹³C NMR (101 MHz, CDCl₃) δ = 172.77, 137.53, 131.60, 128.13, 122.31, 72.24, 70.54, 21.70,

21.42. ESI-HRMS: calcd for $C_{11}H_{13}Br^{78.9183}NaO_3^+$ [M+Na⁺] 294.9940, found 294.9948; calcd for $C_{11}H_{13}Br^{80.9163}NaO_3^+$ [M+Na⁺] 296.9920, found 296.9900.



(S)-iso-propyl α-hydroxy-α-(4-(trifluoromethyl)phenyl)acetate (2p)

(C13H15F3O3) a white solid; 99% yield, 85% ee. $[\alpha]_D^{17} = +73.62$ (c = 0.254, in CH₂Cl₂). HPLC Phenomenex Lux 5u Cellulose-2, 2-propanol/*n*-hexane = 5/95, flow rate = 1.0 mL/min, $\lambda = 210$ nm, retention time: 7.48 min, 8.28 min. ¹H NMR (400 MHz, CDCl3) δ 7.69 – 7.46 (m, 4H), 5.11 (d, J = 5.2 Hz, 1H), 5.00 (dt, J = 12.5, 6.3 Hz, 2H), 3.56 (d, J = 5.5 Hz, 1H), 1.23 (d, J = 6.3 Hz, 3H), 1.04 (d, J = 6.3 Hz, 3H). ¹³C NMR (101 MHz, CDCl3) δ = 171.50, 141.29, 129.59, 129.26, 125.71, 124.44, 124.40, 124.36, 124.32, 121.64, 71.24, 69.78, 20.65, 20.36. ESI-HRMS: calcd for C12H13F3NaO5⁺ [M+Na⁺] 285.0709, found 285.0719.



	Retention Time	%Area
1	7.483	7.51
2	8.278	92.49

(S)-iso-propyl α-hydroxy-α-(naphthalen-1-yl)acetate (2q)

 $(C_{16}H_{18}O_3)$ a white solid; 99% yield, 86% ee. $[\alpha]_D^{25} = +93.18$ (c = 0.264, in CH₂Cl₂). HPLC Phenomenex Lux 5u Cellulose-2, 2-propanol/*n*-hexane = 10/90, flow rate = 1.0 mL/min, $\lambda = 210$ nm, retention time: 13.56 min, 15.76 min. ¹H NMR (400 MHz, CDCl3) δ 8.18 – 8.02 (m, 1H), 7.85 – 7.71 (m, 2H), 7.51 – 7.31 (m, 4H), 5.69 (s, 1H), 5.02 (dt, J = 12.5, 6.3 Hz, 1H), 3.53 (s, 1H), 1.16 (d, J = 6.3 Hz, 3H), 0.92 (d, J = 6.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl3) $\delta = 173.77$, 134.35, 134.02, 131.10, 129.31, 128.74, 126.40, 125.84, 125.63, 125.20, 123.85, 71.40, 70.32, 21.68, 21.36. ESI-HRMS: calcd for C₁₆H₁₈NaO₃⁺ [M+Na⁺] 267.0997, found 267.0998.



	Retention Time	%Area
1	13.699	50.00
2	15.867	50.00



	Retention Time	%Area
1	13.559	93.37
2	15.764	6.63

(S)-iso-propyl α-hydroxy-α-(naphthalen-2-yl)acetate (2r)



(C₁₆H₁₈O₃) a white solid; 99% yield, 86% ee. $[\alpha]_D^{25} = +85.71$ (c = 0.308, in CH₂Cl₂). HPLC Phenomenex Lux 5u Cellulose-2, 2-propanol/*n*-hexane = 10/90, flow rate = 1.0 mL/min, $\lambda = 210$ nm, retention time: 11.86 min, 13.28 min. ¹H NMR (400 MHz, CDCl₃) δ 7.83 (s, 1H), 7.80 – 7.67 (m, 3H), 7.49 – 7.35 (m, 3H), 5.21 (d, J = 5.2 Hz, 1H), 5.00 (dt, J = 12.5, 6.3 Hz, 1H), 3.59 (d, J = 5.7 Hz, 1H), 1.20 (d, J = 6.3 Hz, 3H). 1.00 (d, J = 6.3 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) $\delta = 173.27$, 135.94, 133.24, 133.20, 128.34, 128.16, 127.71, 126.28, 126.27, 125.79, 124.14, 73.06, 70.36, 21.75, 21.45. ESI-HRMS: calcd for C₁₆H₁₈NaO₃⁺ [M+Na⁺] 267.0997, found 267.0995.



	Retention Time	%Area
1	11.896	49.78



	Retention Time	%Area
1	11.859	6.95
2	13.283	93.05

(S)-2-bromo-1-phenylethanol (2v)

(C12H16O3) a white solid; 95% yield, 81% ee. $[\alpha]_D^{27} = +44.36$ (c = 0.780, in CHCl3), {Lit.^[5] $[\alpha]_D^{20} = -45.5$ (c = 1.0, CHCl3), conf. (R)}. HPLC DAICEL CHIRALCEL ODH, 2-propanol/*n*-hexane = 5/95, flow rate = 1.0 mL/min, $\lambda = 210$ nm, retention time: 13.47 min, 14.75 min. ¹H NMR (400 MHz, CDCl3) δ 7.39 – 7.27 (m, 5H), 4.93 – 4.81 (m, 1H), 3.58 (dd, J = 10.5, 3.5 Hz, 1H), 3.49 (dd, J = 10.4, 8.8 Hz, 1H), 2.94 (d, J = 2.8 Hz, 1H). ¹³C NMR (101 MHz, CDCl3) $\delta = 140.44$, 128.72, 128.50, 126.06, 73.83, 40.07. ESI-HRMS: calcd for C11H14Br^{78.9183}NaO3⁺ [M+Na⁺] 222.9734, found 222.9739. C11H14Br^{80.9163}NaO3⁺ [M+Na⁺] 224.9714, found 224.9725.


	Retention Time	%Area
1	13.469	9.74
2	14.753	90.26

(J) Spectral characterization data and HPLC conditions for the alkyl products



2s-2t (x mmol) obtained in asymmetric intramolecular Cannizzaro reaction, benzoic anhydride (1.1x mmol), pyridine (x mmol), DMAP (x mmol) and CH_2Cl_2 (1 mL) were added in the reaction tube sequently. The reaction was stirred vigorously at 30°C (monitored by TLC). The mixture was purified by column chromatography on silica gel (petroleum ether/ethyl acetate) to afford the desired product **2s-2t**. The ee of **2s-2t** was determined according to the ee of **3s-3t**.

(S)-iso-propyl 2-hydroxyhexanoate (2s)



(C₉H₁₈O₃) a colorless oil; 99% yield. $[\alpha]_D{}^{19} = +15.08$ (*c* = 0.252, in CH₂Cl₂). ESI-HRMS: calcd for C₉H₁₈NaO₃⁺ [M+Na⁺] 117.1154, found 117.1161.

(S)-iso-propyl 2-benzoxyhexanoate (3s)



(C₁₆H₂₂O₄) a colorless oil; 90% ee. $[\alpha]_D^{18} = +67.75$ (c = 0.306, in CH₂Cl₂). HPLC Phenomenex Lux 5u Cellulose-2, 2-propanol/*n*-hexane = 5/95, flow rate = 1.0

mL/min, $\lambda = 210$ nm, retention time: 5.05 min, 5.53 min. ¹H NMR (400 MHz, CDCl₃) $\delta 8.11 - 7.97$ (m, 2H), 7.51 (t, J = 7.4 Hz, 1H), 7.45 - 7.34 (m, 2H), 5.22 - 5.06 (m, 1H), 5.02 (dt, J = 12.5, 6.3 Hz, 1H), 2.00 - 1.83 (m, 2H), 1.45 - 1.29 (m, 4H), 1.21 (d, J = 6.3 Hz, 3H), 1.17 (d, J = 6.3 Hz, 3H), 0.87 (t, J = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) $\delta = 169.91$, 166.18, 133.22, 129.82, 129.68, 128.40, 73.05, 68.96, 30.93, 27.36, 22.32, 21.75, 21.70, 13.90.ESI-HRMS: calcd for C₁₆H₂₂NaO₄⁺ [M+Na⁺] 301.1416, found 301.1414.



(S)-iso-propyl 2-hydroxy-2-(adamantan-1-yl)acetate (2u)

(C₁₅H₂₄O₃) a colorless oil; 99% yield. $[\alpha]_D{}^{19} = +81.20$ (c = 0.266, in CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 5.06 (dt, J = 12.5, 6.3 Hz, 1H), 3.53 (s, 1H), 2.63 (s, 1H), 1.93 (s, 3H), 1.64 (d, J = 12.0 Hz, 3H), 1.56 (d, J = 9.8 Hz, 9H), 1.23 (t, J = 6.4 Hz, 6H). ¹³C NMR (101 MHz, CDCl₃) $\delta = 173.49$, 78.80, 69.35, 37.96, 37.15, 36.94, 28.26, 22.08, 21.89. ESI-HRMS: calcd for $C_{15}H_{24}NaO_3^+$ [M+Na⁺] 275.1623, found 275.1615.

(S)-iso-propyl 2-benzoxy-2-(adamantan-1-yl)acetate (3u)

OBz ¥°∕

(C₂₂H₂₈O₃) a colorless oil; 84% ee. $[\alpha]_D^{18}$ = +90.78 (*c* = 0.326, in CH₂Cl₂). HPLC DAICEL CHIRALCEL IC, 2-propanol/*n*-hexane = 5/95, flow rate = 1.0 mL/min, λ = 210 nm, retention time: 7.63 min, 8.25 min. ESI-HRMS: calcd for C₂₂H₂₈NaO₃⁺ [M+Na⁺] 379.1885, found 379.1888.



	Retention Time	%Area
1	7.632	91.99
2	8.254	8.01

(K) Copies of NMR spectra for catalysts and products



(S)-iso-propyl mandelate (2a)





(S)-iso-propyl α-hydroxy-α-(2-methylphenyl)acetate (2b)





(S)-*iso*-propyl α-hydroxy-α-(3-methylphenyl)acetate (2c)





(S)-iso-propyl α-hydroxy-α-(4-methylphenyl)acetate (2d)





(S)-iso-propyl α-hydroxy-α-(4-(*tert*-butyl)phenyl)acetate (2e)





(S)-iso-propyl α-hydroxy-α-(4-phenylphenyl)acetate (2f)











(S)-*iso*-propyl α-hydroxy-α-(3,4-methylenedioxylphenyl)acetate (2h)









(S)-iso-propyl α-hydroxy-α-(2-flurophenyl)acetate (2j)





(S)-*iso*-propyl α-hydroxy-α-(4-flurophenyl)acetate (2k)





(S)-iso-propyl α-hydroxy-α-(2-chlorophenyl)acetate (2l)





(S)-iso-propyl α-hydroxy-α-(4-chlorophenyl)acetate (2m)











(S)-iso-propyl α-hydroxy-α-(2-bromophenyl)acetate (20)











(S)-iso-propyl α-hydroxy-α-(naphthalen-1-yl)acetate (2q)




(S)-*iso*-propyl α-hydroxy-α-(naphthalen-2-yl)acetate (2r)



(S)-2-bromo-1-phenylethanol (2u)





(S)-iso-propyl 2-benzoxyhexanoate (3s)







9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 f1 (ppm)

10.5

(S)-iso-propyl 2-hydroxy-2-(adamantan-1-yl)acetate (2t)



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