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Efficient Phosphine-Mediated Formal C(sp³)-C(sp³) Coupling Reactions of Alkyl Halides in Batch and Flow

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Supporting Information

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General Methods

Reactions, unless otherwise stated, were conducted under a positive pressure of argon in oven-dried glassware. Toluene, CH_2Cl_2 , tetrahydrofuran (THF), and acetonitrile were dried with an SPS apparatus. Commercially available reagents were used as purchased unless otherwise noted. Polymer supported triphenylphosphine (CAS # 39319-11-4, catalogue # 93093) was purchased from Sigma-Alrich and used as is. Analytical thin layer chromatography was performed using aluminium plates precoated with silica gel 60 F_{254} (0.2 mm). Flash chromatography employed 230-400 mesh silica gel. Solvents used for chromatography are quoted as volume/volume ratios.

NMR spectroscopy was performed at 298 K using an Avance III HD 400 (400.1 MHz, ¹H; 100.6 MHz, ¹³C, 376.5 MHz, ¹⁹F) or an Avance III 300 (300 MHz, ¹H; 75 MHz, ¹³C; 282.5 MHz, ¹⁹F). Data is expressed in parts per million (ppm) downfield shift from tetramethylsilane with residual solvent as an internal reference (δ 7.26 ppm for chloroform, 5.27 ppm for dichloromethane, 1.94 ppm for acetonitrile, and 2.09 ppm for the toluene methyl group) and is reported as position (δ in ppm), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet), coupling constant (*J* in Hz) and integration (number of protons). ¹³C NMR spectra were recorded at 298 K with complete proton decoupling. Data is expressed in parts per million (ppm) downfield shift relative to the internal reference (δ 77.2 ppm for the central peak of deuterated chloroform).

Infrared spectra were obtained on a ThermoNicolet Avatar 370 FT-IR spectrometer and are reported in wavenumbers (cm⁻¹). HRMS were performed at the Bioanalytical Mass Spectrometry Facility within the Mark Wainwright Analytical Centre at the University of New South Wales on an Orbitrap LTQ XL (Thermo Fisher Scientific, San Jose, CA, USA) ion trap mass spectrometer. LCMS and GCMS analyses were carried out on Shimadzu LCMS-2010 EV and GCMS-QP2010 Ultra, respectively.

Microwave reactions were carried out in 35 mL microwave vials on CEM Discover – SP W/ACTIVENT 909155 or 30 mL vials on Anton Paar Monowave 300.

Phosphine Auxiliary Screening and Optimization of the C(sp³)-C(sp³) Coupling Reaction

General procedure for Table 1: A mixture of benzylbromide (1.0 mmol) and phosphine **3** (1.0 mmol) was taken up in dry solvent (3 mL) in a reaction flask loaded with a stirrer bar under argon atmosphere. The reaction mixture was heated to the indicated temperature then cooled to room temperature. A 2 mL solution of benzylbromide (1.0 mmol) and base (1.0 mmol) was subsequently added. The reaction mixture was heated to the indicated temperature again then cooled to room temperature. An aqueous solution of NaOH (indicated molar concentration, 1 mL) was then added and the reaction mixture was heated to the indicated temperature was reacted with ethyl acetate (3 x 5 mL). The combined organic phases was washed with brine, dried over Na₂SO₄ and concentrated under reduced pressure. The products were purified from the residues by column chromatography (silica-gel, 100% hexanes \rightarrow hexane/EtOAc = 95/5).



Table	1.	Dhag	nhina	anniliam	, correcting and	antimization	of the reaction
I able	1:	Phos	pnine	auxillaly	screening and	opumization	of the reaction

10	(2-furyl) ₃ P	KHMDS/THF	2 M/70 °C	64/15
11	(2,4,6-MeO ₃ Ph) ₃ P	KHMDS/THF	2 M/70 °C	51/n.d.
12	$(C_6F_5)_3P$	KHMDS/THF	2 M/70 °C	trace/n.d.
13	Ph ₃ P	LDA/THF	2 M/70 °C	58/18
14	Ph ₃ P	KO ^t Bu/THF	2 M/70 °C	34/14
15	Ph ₃ P	KHMDS/Tol	2 M/70 °C	49/trace
16	Ph ₃ P	KHMDS/MeCN	2 M/70 °C	66/trace
17^{d}	Ph ₃ P-resin	KHMDS/MeCN	2 M/70 °C	65/5
	-			
18	Ph ₃ P	KHMDS/MeCN	1 M/70 °C	54/trace
18 19	Ph ₃ P Ph ₃ P	KHMDS/MeCN KHMDS/MeCN	1 M/70 °C 5 M/70 °C	54/trace 51/20
18 19 20	Ph ₃ P Ph ₃ P Ph ₃ P	KHMDS/MeCN KHMDS/MeCN KHMDS/MeCN	1 M/70 °C 5 M/70 °C 2 M/rt	54/trace 51/20 12/trace
18 19 20 21	Ph ₃ P Ph ₃ P Ph ₃ P Ph ₃ P	KHMDS/MeCN KHMDS/MeCN KHMDS/MeCN KHMDS/MeCN	1 M/70 °C 5 M/70 °C 2 M/rt 2 M/100 °C	54/trace 51/20 12/trace 84/trace
18 19 20 21 22 ^e	Ph ₃ P Ph ₃ P Ph ₃ P Ph ₃ P Ph₃P	KHMDS/MeCN KHMDS/MeCN KHMDS/MeCN KHMDS/MeCN KHMDS/MeCN	1 M/70 °C 5 M/70 °C 2 M/rt 2 M/100 °C 2 M/150 °C	54/trace 51/20 12/trace 84/trace 94/trace

^{*a*} Reaction conditions: halide **1a** (1.0 mmol) and phosphine **3** (1.0 mmol) in THF (3 mL), followed by the addition of **1a** (1.0 mmol) and base (1.0 mmol), finally aq. NaOH (1 mL) with conventional heating; ^{*b*} Overall yield of isolated product; ^{*c*} Phosphonium tetrafluoroborate was deprotonated with 1 equiv of base in step 1; ^{*d*} Ph₃P on polymer support (~3 mmol/g) was used; ^{*e*} All steps were carried out in MW reactor (reaction time: step 1 = 2 h at 150 °C, step 2 = 3 h at 150 °C, step 3 = 1 h at 100 °C).

Substrate Scope under Microwave-Assisted Conditions

General procedure for Table 2: A mixture of halide **1** (1.0 mmol) and triphenylphosphine (1.0 mmol) was taken up in dry acetonitrile (3 mL) in a microwave vial loaded with a stirrer bar. The reaction mixture was heated to 150 °C in microwave reactor (ramp-up time 2 min, holding time 120 min) then cooled to room temperature. A 2 mL solution of halide **2** (1.0 mmol) and KHMDS (1.0 mmol) was subsequently added to the reaction vial. The reaction mixture was heated to 150 °C again (ramp-up time 2 min, holding time 180 min) then cooled to room temperature. An aqueous solution of NaOH (2 M, 1 mL, 2 mmol) was then added and the reaction mixture was heated to 100 °C (ramp-up time 2 min, holding time 60 min) before cooled to room temperature. The reaction mixture was diluted with water (10 mL) and extracted with ethyl acetate (3 x 5 mL). The combined organic phases was washed with brine, dried over Na₂SO₄ and concentrated under reduced pressure. The product was purified from the residues by column chromatography (silica-gel, 100% hexanes \rightarrow hexane/EtOAc = 95/5).



Table 2: Substrate scope under microwave-assisted conditions^a

^{*a*} Reaction conditions: halide **1** (1.0 mmol) and Ph₃P (1.0 mmol) in MeCN (3 mL), followed by the addition of **2** (1.0 mmol) and KHMDS (1.0 mmol), finally aq. NaOH (2 M, 1 mL). All steps were carried out in MW reactor (reaction time: step 1 = 2 h at 150 °C, step 2 = 3 h at 150 °C, step 3 = 1 h at 100 °C); ^{*b*} From benzyl bromide and 1,5-diiodopentane.

Characterization Data of C(*sp*³)-C(*sp*³) Coupled Products (Table 2)



¹**H** NMR (300 MHz, CDCl₃): δ = 7.31-7.26 (4H, m, Ar*H*), 7.21-7.17 (6H, m, Ar-*H*), 2.95 (4H, s, ArC*H*₂) ppm.

¹³C NMR (75 MHz, CDCl₃): δ = 141.8, 128.5 (2C), 128.4 (2C), 126.0, 38.0 ppm.



4b²

¹H NMR (400 MHz, CDCl₃): δ = 7.21–7.05 (m, 9H, ArH), 2.91–2.81 (m, 4H, 2CH₂) ppm.
¹³C NMR (101 MHz, CDCl₃): δ = 142.6, 141.1, 132.0, 131.9, 128.7, 128.5 (4C), 126.2, 125.2, 124.4, 122.8, 37.7, 37.6 ppm.

¹⁹**F NMR** (282.5 MHz, CDCl₃): δ = -62.61 (d, J = 53.7 Hz) ppm.



¹**H NMR** (300 MHz, CDCl₃): δ = 7.38-7.33 (m, 4H), 7.26 (d, J = 7.1 Hz, 6H, Ar-H), 2.67 (t, J = 8.1 Hz, 4H, Ar-CH₂), 2.03 (quintet, J = 7.3 Hz, 2H, Ar-CH₂-CH₂-CH₂-Ar) ppm. ¹³**C NMR** (75 MHz, CDCl₃): δ = 142.4, 128.6 (2C), 128.4 (2C), 125.9, 35.6, 33.1 ppm.



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¹ W. Kerr, R. Mudd, J. Brown, Chem. Eur. J. 2016, 22, 4738-4742

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⁴ W. Guo, Z. Wang, *Tetrahedron* **2013**, *69*, 9580-9585

¹**H NMR** (300 MHz, CDCl₃): δ = 8.01-7.98 (m, 1H), 7.89-7.86 (m, 1H), 7.74 (d, J=8.0 Hz, 1H), 7.54-7.40 (m, 3H), 7.37-7.02 (m, 6H), 3.14 (t, J = 7.9 Hz, 2H), 2.79 (t, J = 7.6 Hz, 2H), 2.18-2.10 (m, 2H) ppm.

¹³**C NMR** (75 MHz, CDCl₃): δ = 142.2, 138.5, 133.9, 132.2, 128.8, 128.5 (2C), 128.3 (2C), 126.6, 125.9, 125.8, 125.7, 125.5, 125.4, 123.8, 35.9, 32.6, 32.3 ppm.



¹**H NMR** (400 MHz, CDCl₃): δ = 7.64 – 7.54 (m, 2H, Ar*H*), 7.39 – 7.21 (m, 5H, Ar*H*), 7.19 – 7.12 (m, 2H, Ar*H*), 3.06 – 2.92 (m, 4H, Ar-C*H*₂) ppm.

¹³**C** NMR (101 MHz, CDCl₃): $\delta = 147.3$, 140.6, 132.2, 129.4, 128.8, 128.5, 128.5, 128.2, 126.3, 119.1, 109.9, 37.9, 37.2 ppm.



¹**H NMR** (400 MHz, CDCl₃): δ = 7.48 – 7.41 (m, 2H, Ar*H*), 7.37 – 7.16 (m, 5H, Ar*H*), 7.12 – 7.05 (m, 2H, Ar*H*), 2.93 (s, 2H, Ar-C*H*₂) ppm.

¹³**C NMR** (101 MHz, CDCl₃): δ = 141.3, 140.7, 131.4, 130.3, 128.5 (2C), 128.4 (2C), 126.1, 119.7, 37.7, 37.3 ppm.



 $4g^7$

¹**H NMR** (300 MHz, CDCl₃): δ = 7.61 – 7.51 (m, 2H, Ar*H*), 7.39 – 7.21 (m, 5H, Ar*H*), 7.21 – 7.15 (m, 2H, Ar*H*), 3.15 – 2.83 (m, 4H, Ar-C*H*₂) ppm. ¹³**C NMR** (101 MHz, CDCl₃): δ = 145.77, 141.05, 128.81 (2C), 128.45 (2C), 126.17, 125.26

(q, J = 3.8 Hz), 37.67, 37.52 ppm.

⁵ Shen, Z.-L., Goh, K. K. K., Yang, Y.-S., Lai, Y.-C., Wong, C. H. A., Cheong, H.-L. and Loh, T.-P. *Angew. Chem. Int. Ed.* **2011**, *50*, 511–514.

⁶ T. Di Franco; N.Boutin; X. Hu, *Synthesis* **2013**, 45, 2949-2958

⁷ P. J. Rushworth, D. G. Hulcoop, D. J. Fox, *J. Org. Chem.* **2013**, *78*, 9517-9521.

¹⁹**F NMR** (377 MHz, CDCl₃): δ = -62.28 ppm.



¹**H NMR** (400 MHz, CDCl₃): δ = 8.16-8.13 (m, 2H, ArH), 7.33-7.28 (m, 4H, ArH), 7.26-7.22 (m, 1H, ArH), 7.17-7.15 (m, 2H, ArH), 3.04-3.08 (m, 2H, CH₂), 2.97-3.00 (m, 2H, CH₂) ppm. ¹³**C NMR** (101 MHz, CDCl₃): δ = 149.4, 146.5, 140.5, 129.4 (2C), 128.5 (2C), 128.4 (2C), 126.3, 123.6 (2C), 37.7, 37.2.



4i⁹

¹**H NMR** (400 MHz, CDCl₃): $\delta = 7.37 - 7.16$ (m, 5H, Ar*H*), 3.30 - 2.73 (m, 4H, Ar-C*H*₂) ppm.

¹³**C NMR** (101 MHz, CDCl₃): δ = 140.04, 128.56 (2C), 128.32 (2C), 126.55, 35.36, 24.46 ppm.

¹⁹**F NMR** (377 MHz, CDCl₃): δ = -144.31 (dd, *J* = 22.1, 8.4 Hz), -157.55 (d, *J* = 21.1 Hz), -162.75 - -162.98 (m) ppm.



¹**H NMR** (400 MHz, CDCl₃): δ = 7.39-7.23 (m, 5H, ArH), 3.01-3.95 (m, 1H, CH), 1.36-1.33 (m, 6H, 2CH₃) ppm.

¹³C NMR (101 MHz, CDCl₃): δ = 148.9, 128.4 (2C), 126.5 (2C), 125.8, 34.2, 24.1 (2C) ppm

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⁹ S. Xu, G. Wu, F. Ye, X. Wang, H. Li, X. Zhao, Y. Zhang, J. Wang, *Angew. Chem. Int. Ed.* 2015, 54, 4669-4672

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¹**H NMR** (300 MHz, CDCl₃): δ = 7.23-7.09 (m, 5H, ArH), 2.51 (t, J = 7.7 Hz, 2H, Ar-CH₂), 1.59-1.45 (m, 3H, ArCH₂-CH₂), 1.19-1.12 (m, 2H, CH₂-CH-(CH₃)₂), 0.80 (d, 6H, J = 6.6 Hz, 2CH₃) ppm

¹³C NMR (75 MHz, CDCl₃): δ = 143, 128.4 (2C), 128.2 (2C), 125.6, 38.7, 36.3, 29.4, 27.9, 22.6 (2C) ppm.



4n¹²

¹**H NMR** (300 MHz, CDCl₃): δ = 7.20-7.09 (m, 5H, Ar-H), 2.53 (t, J= 7.6 Hz, 2H, CH₂-Ar), 1.59-1.51 (m, 2H, ArCH₂-CH₂), 1.26-1.18 (m, 6H, 3CH₂), 0.81 (t, J= 7.1 Hz, 3H, CH₃) ppm. ¹³**C NMR** (75 MHz, CDCl₃): δ = δ 143.0, 128.4 (2C), 128.2 (2C), 125.6, 36.0, 31.8, 31.5, 29.1, 22.7, 14.1 ppm.



¹**H NMR** (400 MHz, CDCl₃): δ = 7.22-7.06 (m, 5H, Ar-H), 2.52 (t, J = 7.8 Hz, 2H, Ar-CH₂), 1.56-1.51 (quint, J = 7.5 Hz, 2H, Ar-CH₂-CH₂), 1.26-1.18 (m, 8H, 4CH₂), 0.81 (t, J = 7.0 Hz,

3H, CH₃) ppm.

¹³**C NMR** (101 MHz, CDCl₃): δ = 143.0, 128.4 (2C), 128.2 (2C), 125.6, 36.0, 31.9, 31.6, 29.3, 29.2, 22.7, 14.1 ppm.



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¹**H NMR** (300 MHz, CDCl₃): δ = 7.22-7.06 (m, 5H, Ar-H), 2.52 (t, J = 7.5 Hz, 2 H, CH₂-Ar), 1.64 (quint, J = 7.6 Hz, 2 H, Ar-CH₂-CH₂), 1.33–1.29 (m, 10 H), 0.91 (t, J = 7.0 Hz, 3H, CH₃) ppm.

¹³**C NMR** (75 MHz, CDCl₃): δ = 143.0, 128.4 (2C), 128.2 (2C), 125.6, 36.0, 31.9, 31.5, 29.5, 29.4, 29.3, 22.7, 14.2 ppm.



¹**H NMR** (300 MHz, CDCl₃): δ = 7.33-7.17 (m, 5H, ArH), 2.63 (t, J = 7.5 Hz, 2H, CH₂-Ar), 1.64 (quint, J = 7.4 Hz, 2H, Ar-CH₂-CH₂), 1.30-1.26 (m, 14H, 7CH₂), 0.91 (t, J = 7.0 Hz, 3H, CH₃) ppm.

¹³**C NMR** (76 MHz, CDCl₃): δ = 143.1, 128.4(2C), 128.2 (2C), 125.5, 36.0, 31.9, 31.5, 29.7, 29.63, 29.61, 29.5, 29.4, 22.7, 14.1 ppm.



¹**H NMR** (300 MHz, CDCl₃): δ = 7.31-7.20 (m, 5H, ArH), 2.64 (t, J = 7.5Hz, 2H, Ar-CH₂), 1.65 (quint, J = 7.3 Hz, 2H, Ar-CH₂-CH₂), 1.41-1.30 (m, 16H, 8CH₂), 0.89 (t, J=7.2 Hz, 3H, CH₃) ppm.

¹³**C NMR** (76 MHz, CDCl₃): δ = 143.0, 128.4 (2C), 128.2 (2C), 125.5, 36.0 (C₁), 31.9 (C₉), 31.5 (C₂), 29.68, 29.64, 29.61, 29.5, 29.4 (2C), 22.7, 14.1 ppm



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¹**H NMR** (300 MHz, CDCl₃): δ = 7.22-7.09 (m, 5H, ArH), 2.53 (t, J= 7.5 Hz, 2H, Ar-CH₂), 1.54 (quint, J = 7.3 Hz, 2H, Ar-CH₂-CH₂), 1.25-1.18 (m, 28H, 14CH₂), 0.81 (t, J = 6.5 Hz, 3H, CH₃)

¹³C NMR (75 MHz, CDCl₃): 143.0, 128.4 (2C), 128.2 (2C), 125.5, 36.0, 31.9, 31.5, 29.7 (9C), 29.6, 29.5, 29.4, 22.7, 14.1 ppm



This compound was produced according to the general procedure with benzylbromide as halide **1** and 1,5-diiodopentane as halide **2**. Two molar equivalents of KHMDS were used to facililate the second ylide formation and the cyclization of the six-membered ring. Presumably, the reaction proceeded through intermediates depicted below:



¹**H NMR** (400 MHz, CDCl₃): δ = 7.37-7.22 (m, 5H, ArH), 2.59-2.53 (m, 1H, Ar-C*H*), 1.96-1.80 (m, 5H, 5C*H*₂), 1.55-1.30 (m, 5H, 5C*H*₂) ppm.

¹³C NMR (101 MHz, CDCl₃): δ = 148.1, 128.3 (2C), 126.9 (2C), 125.8, 44.7, 34.5(2C), 27.0 (2C), 26.2 ppm



¹**H NMR** (400 MHz, CDCl₃): 7.38-7.24 (m, 5H, ArH), 2.70 (t, J = 7.7 Hz, 2H, Ar-CH₂), 1.71 (quint, J = 7.1 Hz, 2H, Ar-CH₂-CH₂), 1.45-1.36 (m, 10H, 5CH₂), 1.00-0.97 (t, J = 7.0 Hz, 3H, CH₃) ppm

¹³C NMR (101 MHz, CDCl₃): 143.0, 128.5 (2C), 128.3 (2C), 125.6, 36.1, 32.0, 31.6, 29.7, 29.6, 29.4 (2C), 22.8, 14.2 ppm

¹⁹ C. Indranil, O. Martin, Org. Lett. 2016, 18, 2463-2466

²⁰ P. Tuhin, A. Soumitra, Akanksha, M. Debabrata, Chem. Commun. 2013, 49, 69-71



This compound was produced according to the general procedure with ethyl bromoacetate as halide 1 and pentadecyl tosylate as halide 2. The ethyl ester group was hydrolyzed under the reaction conditions.

¹**H NMR** (400 MHz, CDCl₃): 3.69 (s, 1H, HO-CO), 2.32 (t, J = 7.5 Hz, 2H, CO-C*H*₂), 1.66-1.59 (m, 2H, COCH₂-C*H*₂), 1.33-1.27 (m, 27H, 13(C*H*₂)-CH₃), 0.9 (t, J = 6.6 Hz, 3H, CH₂-*CH*₃) ppm.

¹³C NMR (101 MHz, CDCl₃): 174.4, 51.4, 34.1, 31.9, 29.7 (5C), 29.6, 29.5, 29.4, 29.3, 29.2, 25.0, 22.7, 14.1 ppm.

²¹ E. Bengsch, B. Perly, C. Deleuze, A. Valero., J. Magn. Reson. 1986, 68, 1-13

General Settings for C(sp³)-C(sp³) Coupling Reactions in Flow



Typical reactions and conditions: i) Pump 1: alkyl bromide 1 (2.4 equiv.) in acetonitrile (0.08 M), 0.3 mL/min, column T = 120 °C, 5 h; ii) Pump 2: 1.0 M KHMDS (5.0 equiv.), THF, 0.3 mL/min, column T = 20 °C, 50 min; iii) Pump 3: alkyl bromide 2 (2.4 equiv.) in acetonitrile (0.08 M), 0.3 mL/min, column T = 120 °C, 5 h; iv) Pump 4: 2.0 M NaOH_(aq), 0.3 mL/min, column T = 20 °C, 4 h; v) Pump 5 (optional for regeneration of TPP-resin): Ph₂SiH₂ (2.4 equiv.) in acetonitrile (0.08 M), 0.3 mL/min, column T = 120 °C, 5 h. Yields based on molar amount of Ph₃P on TPP-resin used (3 mmol on 1 g).

TPP-resin: triphenylphosphine, styrene polymer-bound, 100-200 mesh, ~3.0 mmol/g loading, 2% cross-linked with divinylbenzene. This resin was purchased from Sigma-Alrich and used as is. For **4a**, 2 g of TPP-resin was used; for other flow setups, 1 g TPP-resin was used.

Equipment: Pumps = Little Things Factory GmbH Mr. Q continuous syringe pump; Column = Omnifit®, $L \times I.D.$ 100 mm × 10 mm, bed volume 5.6 mL, fixed ends; Heat block for column = FRX volcano; Pressure regulator = Upchurch ScientificTM 100 psi.





Recyclability of TPP-resin and Solid State ³¹**P NMR Studies**

We carried out solid-state ³¹P NMR (121.4 MHz, 25 °C) studies of TPP-resin on a Bruker Avance III 700 MHz Solid State NMR spectrometer after each step of the homocoupling reaction of **1a** to **4a**. We could not obtain a solid state ³¹P NMR spectrum for the polymer-bound phosphonium ylide intermediate due to the unstable nature of this species. The presence of these phosphine, phosphine oxide and phosphonium salts intermediates in the reaction was confirmed by the characteristic solid-state ³¹P NMR signals.



NMR Spectra of C(sp³)-C(sp³) Coupled Products















82	0.05.00
S S	-8.0E+08
	-7.5E+08
	-7.0E+08
	-6.5E+08
	-6.0E+08
	-5.5E+08
	-5.0E+08
	-4.5E+08
	-4.0E+08
	-3.5E+08
	-3.0E+08
	-2.5E+08
	-2.0E+08
	-1.5E+08
	-1.0E+08
	-5.0E+07
	-0.0E+00
	5.0E+07
10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)	











































