## Supporting Information

# Multifunctional Catalysis: Stereoselective Construction of α-Methylidene-γ-Lactams *via* Amidation/Rauhut-Currier Sequence

Kenta Kishi, Fernando Arteaga Arteaga, Shinobu Takizawa\* and Hiroaki Sasai\*

The Institute of Scientific and Industrial Research (ISIR), Osaka University Mihogaoka, Ibaraki-shi, Osaka 567-0047 (Japan)

taki@sanken.osaka-u.ac.jp, sasai@sanken.osaka-u.ac.jp

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#### 1. General information

<sup>1</sup>H- and <sup>13</sup>C-NMR spectra were recorded with a JEOL JMN ECS400 FT NMR, JNM ECA600 FT NMR or Bruker AVANCE II (<sup>1</sup>H-NMR 400 or 600 MHz, <sup>13</sup>C-NMR 100 or 150 MHz. <sup>1</sup>H-NMR spectra are reported as follows: chemical shift in ppm relative to the chemical shift of CHCl<sub>3</sub> at 7.26 ppm, integration, multiplicities (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet), and coupling constants (Hz). <sup>13</sup>C-NMR spectra reported in ppm relative to the central line of triplet for CDCl<sub>3</sub> at 77 ppm. ESI-MS spectra were obtained with JMS-T100LC (JEOL). Optical rotations were measured with JASCO P-1030 polarimeter. HPLC analyses were performed on a JASCO HPLC system (JASCO PU 980 pump and UV-975 UV/Vis detector). FT-IR spectra were recorded on a JASCO FT-IR system (FT/IR4100). Column chromatography on SiO<sub>2</sub> was performed with Kanto Silica Gel 60 (40-100  $\mu$ m). Commercially available organic and inorganic compounds were used without further purification.

#### 2. Procedure for the preparation of 7l.



A solution of **61** (8.67 mmol) and TsCl (9.10 mmol) in pyridine (29 mL) was heated to 100 °C. After 14 h, the solution was cooled to room temperature followed by evaporation in *vacuo*. The crude mixture was dissolved with EtOAc (40 mL) and washed with 10% aq. HCl (30 mL). The separated organic layer was dried over Na2SO4, evaporated in *vacuo*, giving pure product **71** as white solid quantitatively.

 $\begin{array}{c} \textbf{71}; \ \textbf{Quant.}; \ \textbf{White solid;} \ ^1\textbf{H-NMR} \ (400 \ \text{MHz}, \ \textbf{CDCl}_3) \ \delta \ 7.60 \ (d, \ J=8.2 \ \text{Hz}, \ \textbf{2H}), \ 7.25 \ (d, \ J=8.2 \ \text{Hz}, \ \textbf{2H}), \ 6.54 \ (s, \ \textbf{2H}), \ 5.80 \ (s, \ \textbf{1H}), \ 3.76 \ (s, \ \textbf{3H}), \ 2.42 \ (s, \ \textbf{3H}), \ 1.99 \ (s, \ \textbf{6H}); \ ^{13}\textbf{C}-\textbf{NMR} \ (100 \ \text{MHz}, \ \textbf{CDCl}_3) \ \delta \ 158.4, \ 143.5, \ 139.3, \ 137.7, \ 129.6, \ 127.2, \ 125.3, \ 113.7, \ 55.2, \ 21.6, \ 19.0; \ \textbf{HRMS} \ (\textbf{ESI}) \ \textbf{calcd for} \ \textbf{C}_{16}\textbf{H}_{19}\textbf{NO}_3\textbf{SNa} \ \textbf{m/z} = 328.0978, \ \textbf{found} \ \textbf{m/z} = 328.0977 \ [(M+Na)^+]; \ \textbf{IR} \ (\textbf{KBr}): v \ 3283, \ 2361, \ 1517, \ 1327, \ 543 \ \textbf{cm}^{-1}. \end{array}$ 

## 3. Procedure for the preparation of 2l.



To a solution of **71** (3.08 mmol) in MeOH (15 mL) was added PhI(OAc)<sub>2</sub> (3.08 mmol) at 0 °C and stirred at room temperature. After 1 h, saturated NaHCO<sub>3</sub> aq. was added to the reaction solution to quench. EtOAc (50 mL) was added to the reaction mixture and washed with brine (30 mL). The organic layer was separated and dried over Na<sub>2</sub>SO<sub>4</sub>. Evaporation of solvent followed by dried in *vacuo* gave **81** as crude product. This crude product was dissolved in THF (10 mL) and reacted with MeLi (1.13M ether solution, 8.2 mL) at -78 °C. After 0.5 h, 10% aq. HCl (10 mL) was added to reaction mixture, and then it

warmed to room temperature and stirred for 17 h. The reaction mixture was dissolved with EtOAc (30 mL) and the organic phase was washed with saturated NaHCO3 aq. The separated organic layer was dried over Na2SO4, evaporated, dried in *vacuo* affording crude product which was purified by silica gel column chromatography. Pure **2l** was obtained as yellow solid in 17% overall yield from **7l**.

**2l**; 17% yield; Yellow solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) 
$$\delta$$
 7.63 (d,  $J = 8.4$  Hz, 2H), 7.26 (d,  $J = 8.4$  Hz, 2H), 6.04 (s, 2H), 5.09 (s, 1H), 2.43 (s, 3H), 1.79 (s, 6H), 1.40 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  185.0, 159.3, 144.0, 137.0, 129.4, 128.3, 127.7, 59.2, 26.3, 21.6, 18.6; HRMS (ESI) calcd for C<sub>16</sub>H<sub>19</sub>NO<sub>3</sub>SNa m/z = 328.0978, found m/z = 328.0980 [(M+Na)<sup>+</sup>]; IR (KBr): v 3087, 2867, 1670, 1607, 1455, 1330, 988, 633 cm<sup>-1</sup>.

#### 4. General procedure for the preparation of dienones 2.



A solution of **8** (5.0 mmol) in THF (10 mL) was added to a solution of the corresponding Grignard reagents (7.5 mmol) in THF (7.5 mL) at -78 °C. After 0.5 h, the reaction mixture was acidified by aq. HCl, then, increased to room temperature. After 3 h, the organic layer was extracted with EtOAc and dried in *vacuo*. The resulting crude product was purified by silica gel column chromatography, followed by recrystallization to provide dienones **2** as a solid. Starting material **8a** (PG = Ts), **8b** (PG = Ms), known dienones **2a**, and **2g** were synthesized according to the following literatures.<sup>1-4</sup>

**2b**; 63% yield; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.66 (d, *J* = 8.2 Hz, 2H), 7.31 (d, *J* = 8.2 Hz, 2H), 7.26 (d, *J* = 8.2 Hz, 2H), 7.16 (d, *J* = 8.2 Hz, 2H), 6.75 (d, *J* = 10.1 Hz, 2H), 6.05 (d, *J* = 10.1 Hz, 2H), 5.27 (s, 1H), 2.43 (s, 3H), 2.33 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  185.0, 148.9, 144.2, 139.3, 137.4, 134.7, 130.1, 129.6, 127.8, 127.7, 125.6, 59.6, 21.6, 21.0; HRMS (ESI) calcd for C<sub>20</sub>H<sub>19</sub>NO<sub>3</sub>SNa m/z = 376.0978; found m/z = 376.0969 [(M+Na)<sup>+</sup>]; IR (KBr): v 3092, 2893, 1661, 1616, 1338, 1163, 962 cm<sup>-1</sup>.



2b

**2c**; 73% yield; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.62 (dt, *J* = 8.5, 1.8 Hz, 2H), 7.31 (dd, *J* = 8.0, 1.1 Hz, 1H), 7.19-7.27 (m, 4H), 7.11-7.15 (t, *J* = 8.0 Hz, 1H), 6.98 (dt, *J* = 11.0, 2.5 Hz, 2H), 6.07 (dt, *J* = 11.0, 2.5 Hz, 2H), 5.10 (s, 1H), 2.61 (s, 3H), 2.42 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  184.7, 147.2, 144.3, 137.5, 137.2, 135.7, 134.0, 129.7, 129.5, 128.3, 127.9, 126.9, 126.7, 60.4, 22.0, 21.7; HRMS (ESI) calcd for C<sub>20</sub>H<sub>19</sub>NO<sub>3</sub>SNa m/z = 376.0978; found m/z = 376.0980 [(M+Na)<sup>+</sup>]; IR (KBr): v 3115, 2875, 1662,

1616, 1334, 753 cm<sup>-1</sup>.

TsHN *t*Bu 2d **2d**; 51% yield; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.65 (d, *J* = 8.2 Hz, 2H), 7.34 (s, 4H), 7.25 (d, *J* = 8.2 Hz, 3H), 6.78 (d, *J* = 10.1 Hz, 2H), 6.05 (d, *J* = 10.1 Hz, 2H), 5.44 (s, 1H), 2.42 (s, 3H), 1.28 (s, 9H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  184.9, 152.5, 148.9, 144.2, 137.4, 134.5, 129.6, 127.8, 127.7, 126.4, 125.5, 59.5, 34.6, 31.1, 21.6 HRMS (ESI) calcd for C<sub>23</sub>H<sub>25</sub>NO<sub>3</sub>SNa m/z = 418.1447; found m/z = 418.1440 [(M+Na)<sup>+</sup>]; IR (KBr): v 3095, 2953, 1664, 1618, 1340, 722 cm<sup>-1</sup>.



**2e**; 45% yield; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.63 (d, *J* = 8.2 Hz, 2H), 7.43 (d, *J* = 8.7 Hz, 2H), 7.24-7.30 (m, 4H), 6.76 (d, *J* = 10.5 Hz, 2H), 6.05 (d, *J* = 10.5 Hz, 2H), 5.82 (s, 1H), 2.43 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  184.6, 147.9, 144.4, 137.1, 136.8, 132.4, 129.7, 128.0, 127.8, 127.6, 123.3, 59.4, 21.6; HRMS (ESI) calcd for C<sub>19</sub>H<sub>16</sub>BrNO<sub>3</sub>SNa m/z = 439.9926; found m/z = 439.9921 [(M+Na)<sup>+</sup>]; IR (KBr): v 3107, 2899, 1662, 1617, 1335, 712 cm<sup>-1</sup>.



**2f**; 40% yield; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.64 (d, J = 8.2 Hz, 2H), 7.40 (d, J = 1.8 Hz, 1H), 7.25-7.34 (m, 5H), 6.77 (d, J = 10.1 Hz, 2H), 6.09 (d, J = 10.1 Hz, 2H), 5.41 (s, 1H), 2.43 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  184.5, 147.7, 144.5, 139.7, 137.2, 135.4, 130.6, 129.7, 129.4, 128.3, 127.8, 126.2, 124.0, 59.4, 21.6; HRMS (ESI) calcd for C<sub>19</sub>H<sub>16</sub>ClNO<sub>3</sub>SNa m/z = 396.0432; found m/z = 396.0432 [(M+Na)<sup>+</sup>]; IR (KBr): v 3094, 2881, 1656, 1619, 1336, 1155, 857 cm<sup>-1</sup>.



2h

**2h**; 43% yield; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.66 (d, J = 8.2 Hz, 2H), 7.26 (d, J = 8.2 Hz, 2H), 6.53 (d, J = 10.1 Hz, 2H), 6.09 (d, J = 10.1 Hz, 1H), 5.25 (s, 1H), 2.42 (s, 3H), 1.76 (q, J = 7.6 Hz, 2H), 0.79 (t, J = 7.6 Hz, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  185.0, 149.2, 144.2, 137.4, 129.6, 127.8, 58.1, 33.4, 21.6, 7.5; HRMS (ESI) calcd for C<sub>15</sub>H<sub>17</sub>NO<sub>3</sub>SNa m/z = 314.0821; found m/z = 314.0811

[(M+Na)<sup>+</sup>]; IR (KBr): v 3133, 2971, 2775, 1661, 1615, 1320, 1181, 869 cm<sup>-1</sup>.



**2i**; 13% yield; White solid; <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.69 (d, J = 8.4 Hz, 2H), 7.27 (d, J = 8.4 Hz, 2H), 6.65 (d, J = 10.8 Hz, 2H), 6.06 (d, J = 10.8 Hz, 2H), 5.89 (s, 1H), 5.62 (dd, J = 17.2, 10.3 Hz, 1H), 5.38 (d, J = 17.2 Hz, 1H), 5.24 (d, J = 10.3 Hz, 1H), 2.42 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  184.9, 147.8, 144.3, 137.2, 135.1, 129.6, 128.2, 127.7, 117.7, 58.4, 21.5 HRMS (ESI) calcd for C<sub>15</sub>H<sub>15</sub>NO<sub>3</sub>SNa m/z =

312.0665; found m/z = 312.0659 [(M+Na)<sup>+</sup>]; IR (KBr): v 3088, 2878, 1661, 1616, 1334, 1160, 998, 555 cm<sup>-1</sup>.



**2j**; 31% yield; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.47-7.52 (m, 2H), 7.33-7.41 (m, 3H), 7.08-7.12 (m, 2H), 6.32-6.36 (m, 2H), 5.75 (s, 1H), 3.02 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  184.8, 148.9, 137.5, 129.4, 129.2, 128.2, 125.8, 59.8, 43.0; HRMS (ESI) calcd for C<sub>13</sub>H<sub>13</sub>NO<sub>3</sub>SNa m/z = 286.0508; found m/z = 286.0510 [(M+Na)<sup>+</sup>]; IR (KBr): v 3153, 2881, 1661, 1613, 1336, 1161, 980, 756 cm<sup>-1</sup>.



**2k**; 57% yield; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.96 (d, J = 10.4 Hz, 2H), 6.29 (d, J = 10.4 Hz, 2H), 5.30 (s, 1H), 3.00 (s, 3H), 1.56 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  184.5, 150.5, 128.6, 54.6, 43.0, 27.4; HRMS (ESI) calcd for C<sub>8</sub>H<sub>11</sub>NO<sub>3</sub>SNa m/z = 224.0352; found m/z = 224.0348 [(M+Na)<sup>+</sup>]; IR (KBr): v 3293, 2986, 1712, 1671, 1631, 1139, 594 cm<sup>-1</sup>.

**2m**; 12% yield; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.67 (d, J = 8.2 Hz, 2H), 7.25 (d, J = 8.2 Hz, 2H), 6.56 (d, J = 10.1 Hz, 2H), 6.05 (d, J = 10.1 Hz, 2H), 5.68 (s, 1H), 2.41 (s, 3H), 1.70-1.66 (m, 2H), 1.24-1.10 (m, 4H), 0.80 (t, J = 7.3 Hz, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  185.2, 149.7, 144.1, 137.4, 129.6, 129.2, 127.7, 57.6, 40.1, 25.1, 22.4, 21.6, 13.7; HRMS (ESI) calcd for C<sub>17</sub>H<sub>21</sub>NO<sub>3</sub>SNa m/z = 342.1134; found m/z = 342.1128 [(M+Na)<sup>+</sup>]; IR (KBr): v 3117, 2934, 1659, 1614, 1343, 1161, 873 cm<sup>-1</sup>.

5. General procedure for the preparation of acrylamide 3.



A round bottom flask was charged with a dichloromethane solution of dienone 2 (10 mL, 1.0 mmol) and powdered NaOH (3.0 mmol). Then, acrolylchloride (1.5 mmol) was added to the reaction vesicle at 0 °C. The reaction mixture was stirred vigorously 1 h at 0 °C. The reaction was then quenched with water, extracted with dichloromethane, and dried over Na<sub>2</sub>SO<sub>4</sub>. The combined solvent was removed in *vacuo* and the obtained crude product was quickly purified by silica gel column chromatography using hexane/ethyl acetate as an eluent to give the desired product **3** as a yellow oil or white solid. (**3** is not so stable in silica gel)



**3a**; 63% yield; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.61-7.64 (m, 2H), 7.24-7.33 (m, 9H), 6.94 (dd, J = 16.9, 10.1 Hz, 1H), 6.21 (dd, J = 16.9, 1.1 Hz, 1H), 6.04 (d, J = 10.4 Hz, 2H), 5.78 (dd, J = 10.1, 1.1 Hz, 1H), 2.44 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  184.7, 169.4, 147.9, 145.7, 138.9, 134.7, 133.2, 130.9, 129.8, 129.4, 128.6, 128.5, 127.6, 125.0, 65.6, 21.7; HRMS (ESI) calcd for C<sub>22</sub>H<sub>19</sub>NO<sub>4</sub>SNa m/z = 416.0927; found m/z = 416.0927 [(M+Na)<sup>+</sup>]; IR (KBr): v 3030, 2368, 1702, 1670, 1397, 1349, 1191, 1176, 989, 751,

661 cm<sup>-1</sup>.



**3b**; 12% yield; Pale yellow oil; <sup>1</sup>H-NMR (400 MHz CDCl<sub>3</sub>)  $\delta$  7.62 (d, J = 8.2 Hz, 2H), 7.31-7.26 (m, 4H), 7.17 (d, J = 8.2 Hz, 2H), 7.07 (d, J = 8.2 Hz, 2H), 6.92 (dd, J = 17.4, 10.1 Hz, 1H), 6.23 (dd, J = 17.4, 0.9 Hz, 1H), 6.02 (d, J = 10.1 Hz, 2H), 5.78 (dd, J = 10.1, 0.9 Hz, 1H), 2.44 (s, 3H), 2.28 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  184.8, 169.5, 148.1, 145.6, 138.6, 136.4, 135.5, 133.3, 130.8, 130.1, 129.7, 128.5, 127.4, 124.9, 65.4, 21.7, 21.0; HRMS (ESI) calcd for C<sub>23</sub>H<sub>21</sub>NO<sub>4</sub>SNa m/z = 430.1083; found m/z = 430.1082 [(M+Na)<sup>+</sup>]; IR (KBr): v 3033, 2921, 1703, 1666, 1496, 1355, 1177, 984 cm<sup>-1</sup>.



**3f**; 4% yield; Pale yellow oil; <sup>1</sup>H-NMR (400 MHz CDCl<sub>3</sub>)  $\delta$  7.61 (d, J = 8.2 Hz, 2H), 7.18-7.31 (m, 8H), 6.96 (dd, J = 16.9, 10.1 Hz, 1H), 6.24 (dd, J = 16.9, 1.1 Hz, 1H), 6.06 (d, J = 10.5 Hz, 2H), 5.83 (dd, J = 10.1, 1.1 Hz, 1H), 2.45 (s, 3H) <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  184.4, 169.3, 147.2, 145.9, 140.7, 136.2, 135.3, 133.0, 131.3, 130.5, 129.9, 128.8, 128.4, 128.1, 125.3, 123.1, 65.1, 21.7; HRMS (ESI) calcd for

 $C_{22}H_{18}CINO_4SNa m/z = 450.0537$ ; found m/z = 450.0536 [(M+Na)<sup>+</sup>]; IR (KBr): v 3035, 2925, 1705, 1660, 1354, 1179, 980 cm<sup>-1</sup>.



**3g**; 65% yield; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.61 (d, J = 8.2 Hz, 2H), 7.25-7.27 (m, 2H), 7.07-7.11 (m, 2H), 6.75 (dd, J = 16.9, 10.1 Hz, 1H), 6.45 (dd, J = 16.9, 1.4 Hz, 1H), 5.92-6.00 (m, 3H), 2.41 (s, 3H), 1.54 (s, 3H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  184.2, 170.1, 150.2, 145.2, 136.4, 133.7, 131.5, 129.6, 128.1, 127.2, 60.1, 26.7, 21.5; HRMS (ESI) calcd for C<sub>17</sub>H<sub>17</sub>NO<sub>4</sub>SNa, m/z = 354.0770; found m/z =

354.0774 [(M+Na)<sup>+</sup>]; IR (KBr): v 3044, 1672, 1345, 1182, 973, 866, 666, 592 cm<sup>-1</sup>.



**3h**; 43% yield; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.58 (d, J = 8.2 Hz, 2H), 7.24 (d, J = 8.2 Hz, 2H), 7.06 (d, J = 10.1 Hz, 2H), 6.81 (dd, J = 16.9, 10.1 Hz, 1H), 6.49 (dd, J = 16.9, 0.9 Hz, 1H), 6.01 (d, J = 10.1 Hz, 2H), 5.96 (dd, J = 10.1, 0.9 Hz, 1H), 2.41 (s, 3H), 1.93 (q, J = 7.3 Hz, 2H), 0.74 (t, J = 7.3 Hz, 3H,); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  184.6, 170.8, 148.6, 145.4, 136.5, 134.5, 131.5, 129.6, 128.8, 128.5,

64.1, 31.0, 21.6, 8.2; HRMS (ESI) calcd for  $C_{18}H_{19}NO_4SNa m/z = 368.0927$ ; found  $m/z = 368.0912 [(M+Na)^+]$ ; IR (KBr): v 3042, 1677, 1340, 1180, 865, 656, 590 cm<sup>-1</sup>.



3i; 53% yield; White solid; <sup>1</sup>H-NMR (600 MHz CDCl<sub>3</sub>) δ 7.62 (d, J = 8.4 Hz, 2H), 7.27 (d, J = 8.4 Hz, 2H), 7.06 (d, J = 10.0 Hz, 2H), 6.84 (dd, J = 16.9, 10.1 Hz, 1H), 6.42 (dd, J = 16.9, 1.4 Hz, 1H), 6.05 (d, J = 10.0 Hz, 2H), 5.89 (dd, J = 10.1, 1.4 Hz, 1H), 5.69 (dd, J = 17.2, 10.3 Hz, 1H), 5.21 (d, J = 17.2 Hz, 1H), 5.12 (d, J = 10.3 Hz, 1H), 2.43 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>) δ 184.7, 169.3, 147.0, 145.5, 136.5, 134.9,

133.3, 131.3, 129.8, 128.3, 128.0, 117.1, 64.2, 21.7; HRMS (ESI) calcd for  $C_{18}H_{17}NO_4SNa m/z = 366.0770$ ; found  $m/z = 366.0766 [(M+Na)^+]$ ; IR (KBr): v 3063, 2252, 1699, 1670, 1631, 1400, 1177, 663 cm<sup>-1</sup>



**3k**; 12% yield; White solid; <sup>1</sup>H-NMR (400 MHz CDCl<sub>3</sub>)  $\delta$  7.19 (d, *J* = 10.1 Hz, 2H), 6.44-6.46 (m, 2H), 6.26 (d, *J* = 10.1 Hz, 2H), 5.90 (dd, *J* = 8.2, 3.2 Hz, 1H), 3.26 (s, 3H), 1.79 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  184.1, 169.3, 150.3, 132.5, 131.9, 128.1, 60.7, 44.5, 26.8; HRMS (ESI) calcd for C<sub>11</sub>H<sub>13</sub>NO<sub>4</sub>SNa m/z = 278.0457; found m/z = 278.0453 [(M+Na)<sup>+</sup>]; IR (KBr): v 3025, 2937, 1702, 1667, 1626, 1350, 1180,

862 cm<sup>-1</sup>.



**3I**; 45% yield; White solid; <sup>1</sup>H-NMR (400 MHz CDCl<sub>3</sub> at 60 °C) δ 7.87 (br, 2H), 7.36 (d, *J* = 8.2 Hz, 2H), 6.79 (dd, *J* = 16.3, 10.3 Hz, 1H), 6.36 (d, *J* = 16.3 Hz, 1H), 6.04 (s, 2H), 5.76 (d, *J* = 10.3 Hz, 1H), 2.45 (s, 3H), 1.92 (s, 6H), 1.82 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>) δ 184.5, 162.1, 145.2, 137.4, 131.4, 129.8, 128.4, 126.9, 77.3, 77.0, 76.7, 27.5, 21.2, 19.7 (Some peaks are broad out or overlapped); HRMS (ESI)

calcd for  $C_{19}H_{21}NO_4SNa m/z = 382.1083$ ; found  $m/z = 382.1086 [(M+Na)^+]$ ; IR (KBr): v 3092, 2991, 1666, 1623, 1353, 1190, 964, 819 cm<sup>-1</sup>.



**3m**; 33% yield; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.55-7.58 (m, 2H), 7.23 (d, *J* = 8.2 Hz, 2H), 7.08 (dt, *J* = 11.0, 2.5 Hz, 2H), 6.81 (dd, *J* = 16.9, 11.0 Hz, 1H), 6.48 (dd, *J* = 16.9, 2.5 Hz, 1H), 5.94-6.01 (m, 3H), 2.41 (s, 3H), 1.83-1.87 (m, 2H), 1.06-1.20 (m, 4H), 0.78 (t, *J* = 7.1 Hz, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  184.7, 170.8, 149.0, 145.4, 136.5, 134.6, 131.4, 129.6, 128.6, 128.5, 63.6, 37.7, 25.7, 22.4, 21.6,

13.7; HRMS (ESI) calcd for  $C_{20}H_{23}NO_4SNa m/z = 396.1240$ ; found  $m/z = 396.1239 [(M+Na)^+]$ ; IR (KBr): v 3253, 3122, 2958, 1662, 1616, 1512, 1338, 1159, 814 cm<sup>-1</sup>.

#### 6. General procedure for the preparation of $\alpha$ -methylidene- $\gamma$ -lactam 4.



Acryloylchloride **1** (0.12 mmol) was added to a mixture of dienone **2** (0.10 mmol), DIPEA (0.15 mmol) and catalyst **5a** (0.02 mmol, 20 mol%) in dichloromethane (0.5 mL) under reflux conditions. After the full conversion of **2** as determined by TLC, crude reaction mixture was directly purified by silica gel column chromatography using hexane-ethyl acetate as an eluent to provide the corresponding product **4** as a white solid.

Stepwise process (Method B)



The chiral amine catalyst **5a** (0.02 mmol, 20 mol%) was added to a dichloromethane solution of acrylamide **3** (0.10 mmol, 0.5 mL) at 25 °C. After the full conversion of **3** on TLC, the reaction mixture was directly purified by silica-gel column chromatography using hexane-ethyl acetate as eluents to provide the corresponding product **4** as a white solid.



**4a**; 92% yield for **Method A**, 95% yield for **Method B**; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.85 (d, J = 8.6 Hz, 2H), 7.51 (dd, J = 10.4, 1.6 Hz, 1H), 7.48-7.37 (m, 5H), 7.33 (d, J = 8.6 Hz, 2H), 6.33 (d, J = 10.4 Hz, 1H), 6.24 (d, J = 3.7 Hz, 1H), 5.44 (d, J = 2.7 Hz, 1H), 3.45-3.37 (m, 1H), 2.74-2.54 (m, 2H), 2.45 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  194.6, 165.8, 145.7, 144.6, 139.4, 138.3, 135.4, 130.3, 129.3,

<sup>4a</sup> 129.2, 128.9, 128.8, 125.8, 120.7, 69.9, 48.7, 35.0, 21.7; HRMS (ESI) calcd for C<sub>22</sub>H<sub>19</sub>NO<sub>4</sub>SNa m/z = 416.0927; found m/z = 416.0921 [(M+Na)<sup>+</sup>]; IR (KBr): v 3056, 2925, 2300, 1720, 1691, 1366, 1240, 1172, 1062, 909, 700 cm<sup>-1</sup>; [α]<sub>D</sub><sup>24</sup> = -39 (c 0.71, CHCl<sub>3</sub>) for 96% ee; Enantiomeric excess: 80% for **Method A**, 96% for **Method B**, determined by HPLC (Daicel Chiralpak IC, hexane/EtOH = 2/1; flow rate 1.0 ml/min; 25°C; 225 nm) first peak (Minor):  $t_R$  = 9.5 min, second peak (Major):  $t_R$  = 14.1 min.



**4b**; 50% yield for **Method A**, 85% yield for **Method B**; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.86 (d, J = 8.7 Hz, 2H), 7.50 (dd, J = 10.5, 1.8 Hz, 1H), 7.24-7.34 (m, 6H), 6.31 (d, J = 10.5 Hz, 1H), 6.22 (d, J = 3.2 Hz, 1H), 5.43 (d, J = 3.2 Hz, 1H), 3.38-3.40 (m, 1H), 2.56-2.70 (m, 2H), 2.45 (s, 3H), 2.41 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  194.6, 165.8, 145.6, 144.8, 138.8, 138.5, 136.5, 135.6, 130.1, 129.6, 129.3, 129.2, 125.7, 120.5, 77.3, 77.0, 76.7, 69.9, 48.8, 35.1, 21.7, 21.2; HRMS (ESI) calcd for C<sub>23</sub>H<sub>21</sub>NO<sub>4</sub>SNa m/z = 430.1083; found m/z = 430.1084 [(M+Na)<sup>+</sup>]; IR (KBr): v 2925, 2852, 1725, 1691, 1359, 660, 544

cm<sup>-1</sup>;  $[\alpha]_D^{24} = -42$  (c 0.85, CHCl<sub>3</sub>) for 92% ee; Enantiomeric excess: 78% for **Method A**, 92% for **Method B**, determined by HPLC (Daicel Chiralpak AD-H, hexane/EtOH = 2/1; flow rate 1.0 ml/min; 25°C; 220 nm) first peak (Minor): t<sub>R</sub> = 18.8 min, second peak (Major): t<sub>R</sub> = 32.8 min.



**4c**; 81% yield for **Method A**; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.84 (d, J = 8.2 Hz, 2H), 7.61 (dd, J = 10.5, 1.8 Hz, 1H), 7.28-7.38 (m, 6H), 6.29 (d, J = 3.2 Hz, 1H), 6.24 (d, J = 10.5 Hz, 1H), 5.49 (d, J = 3.2 Hz, 1H), 3.75 (m, 1H), 2.58-2.73 (m, 2H), 2.43 (s, 3H), 2.25 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  194.7, 165.4, 146.0, 145.7, 138.3, 136.2, 135.4, 135.2, 133.8, 129.4, 129.2, 129.1, 128.8, 128.0, 126.3, 121.5, 70.3, 44.2, 35.6, 21.7, 21.1; HRMS (ESI) calcd for C<sub>23</sub>H<sub>21</sub>NO<sub>4</sub>SNa m/z = 430.1083; found m/z =

430.1083 [(M+Na)<sup>+</sup>]; IR (KBr): v 1725, 1693, 1341, 1155, 575 cm<sup>-1</sup>;  $[\alpha]_D^{24} = -35$  (c 1.0, CHCl<sub>3</sub>) for 86% ee; Enantiomeric excess: 86%, determined by HPLC (Daicel Chiralpak AD-H, hexane/EtOH = 2/1; flow rate 1.0 ml/min; 25°C; 221 nm) first peak (Minor): t<sub>R</sub> = 12.6 min, second peak (Major): t<sub>R</sub> = 22.3 min.



**4d**; 61% yield for **Method A**; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.82 (d, J = 8.2 Hz, 2H), 7.52 (dd, J = 10.5, 1.8 Hz, 1H), 7.43 (d, J = 8.7 Hz, 2H), 7.29-7.32 (m, 4H), 6.31 (d, J = 10.5 Hz, 1H), 6.22 (d, J = 3.2 Hz, 1H), 5.42 (d, J = 3.2 Hz, 1H), 3.42 (t, J = 1.6 Hz, 1H), 2.56-2.70 (m, 2H), 2.45 (s, 3H), 1.36 (s, 9H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  194.7, 165.8, 151.8, 145.5, 144.9, 138.6, 136.3, 135.7, 130.1, 129.3, 129.1, 125.8, 125.5, 120.4, 69.8, 48.7, 35.2, 34.7, 31.3, 21.7; HRMS (ESI) calcd for C<sub>26</sub>H<sub>27</sub>NO<sub>4</sub>SNa m/z = 472.1553; found m/z = 472.1551 [(M+Na)<sup>+</sup>]; IR (KBr): v 1732, 1693, 1359, 1150, 580 cm<sup>-1</sup>;  $[\alpha]_D^{27} = -46$ 

(c 4.1, CHCl<sub>3</sub>) for 76% ee; Enantiomeric excess: 76%, determined by HPLC (Daicel Chiralpak IC, hexane/EtOH = 2/1; flow rate 1.0 ml/min; 25°C; 225 nm) first peak (Minor):  $t_R = 6.9$  min, second peak (Major):  $t_R = 14.8$  min.



**4e**; 87% yield for **Method A**; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.86 (d, J = 8.2 Hz, 2H), 7.58 (d, J = 8.2 Hz, 2H), 7.46 (dd, J = 8.2, 1.8 Hz, 1H), 7.32 (dd, J = 19.0, 8.2 Hz, 4H), 6.32-6.34 (m, 1H), 6.24 (d, J = 3.2 Hz, 1H), 5.45 (d, J = 3.2 Hz, 1H), 3.34-3.36 (m, 1H), 2.55-2.72 (m, 2H), 2.46 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  194.1, 165.6, 145.9, 143.8, 138.8, 138.0, 135.3, 132.2, 130.6, 129.5, 129.2, 127.5, 123.0, 121.0, 69.4, 48.6, 34.9, 21.8; HRMS (ESI) calcd for C<sub>22</sub>H<sub>18</sub>BrNO<sub>4</sub>SNa m/z = 494.0032; found m/z = 494.0021 [(M+Na)<sup>+</sup>]; IR (KBr): v 1732, 1693, 1359, 1150, 580 cm<sup>-1</sup>; [ $\alpha$ ]D<sup>27</sup> = -70 (c 4.1, CHCl<sub>3</sub>) for 74%

ee; Enantiomeric excess: 74%, determined by HPLC (Daicel Chiralpak IC, hexane/EtOH = 2/1; flow rate 1.0 ml/min; 25°C; 225 nm) first peak (Minor):  $t_R = 6.9$  min, second peak (Major):  $t_R = 12.2$  min.



**4f**; 81% yield for **Method A**, 93% yield for **Method B**; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.86 (dd, *J* = 8.5, 2.1 Hz, 2H), 7.31-7.48 (m, 7H), 6.34 (dd, *J* = 10.5, 0.9 Hz, 1H), 6.26 (d, *J* = 3.0 Hz, 1H), 5.46 (d, *J* = 3.0 Hz, 1H), 3.36-3.38 (m, 1H), 2.57-2.74 (m, 2H), 2.46 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  194.0, 165.5, 145.9, 143.8, 141.6, 138.0, 135.3, 135.1, 130.6, 130.3, 129.5, 129.1, 126.1, 124.1, 120.9, 69.4, 48.6, 34.9, 21.7; HRMS (ESI) calcd for C<sub>22</sub>H<sub>18</sub>ClNO<sub>4</sub>SNa m/z = 450.0537; found m/z = 450.0537

 $[(M+Na)^+]$ ; IR (KBr): v 2364, 1725, 1699, 1358, 1150, 782 cm<sup>-1</sup>;  $[\alpha]_D^{24} = -33$  (c 1.0, CHCl<sub>3</sub>) for 94% ee; Enantiomeric excess: 79% for **Method A**, 94% for **Method B**, determined by HPLC (Daicel Chiralpak AD-H, hexane/EtOH = 2/1; flow rate 1.0 ml/min; 25°C; 220 nm) first peak (Minor): t<sub>R</sub> = 14.4 min, second peak (Major): t<sub>R</sub> = 40.1 min.



**4g**; 66% yield for **Method A**, 95% yield for **Method B**; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.98 (d, 2H, *J* = 8.4 Hz), 7.35 (d, *J* = 8.4 Hz, 2H), 7.24 (dd, *J* = 10.6, 1.4 Hz, 1H), 6.17 (d, *J* = 2.8 Hz, 1H), 6.02 (d, *J* = 10.6 Hz, 1H), 5.45 (d, *J* = 2.8 Hz, 1H), 3.14-3.21 (m, 1H), 2.70-2.81 (m, 2H), 2.44 (s, 3H), 1.98 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  194.3, 165.3, 146.8, 145.5, 138.4, 135.9, 129.6, 128.6, 128.1, 120.8,

64.7, 45.2, 36.2, 25.1, 21.7; HRMS (ESI) calcd for  $C_{17}H_{17}NO_4SNa m/z = 354.0770$ ; found  $m/z = 354.0775 [(M+Na)^+]$ ; IR (KBr): v 2967, 1722, 1684, 1350, 1167, 805, 661, 583 cm<sup>-1</sup>;  $[\alpha]_D^{25} = -58$  (c 1.1, CHCl<sub>3</sub>) for 94% ee; Enantiomeric excess: 82% for Method A, 94% for Method B, determined by HPLC (Daicel Chiralpak AD-H, hexane/EtOH = 2/1; flow rate 1.0 ml/min; 25°C; 215 nm) first peak (Major):  $t_R = 18.1$  min, second peak (Minor):  $t_R = 28.1$  min.



**4h**; 71% yield for **Method A**, 90% yield for **Method B**; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.96 (d, J = 8.2 Hz, 2H), 7.33 (d, J = 8.2 Hz, 2H), 7.27 (d, J = 10.5 Hz, 1H), 6.16 (d, J = 2.4 Hz, 1H), 6.12 (d, J = 10.5 Hz, 1H), 5.47 (d, J = 2.4 Hz, 1H), 3.30-3.39 (m, 1H), 2.55-2.70 (m, 2H), 2.37-2.48 (m, 4H), 2.11-2.22 (m, 1H), 1.09 (t, J = 7.6 Hz, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  194.9, 165.7, 146.5, 145.5, 139.0,

135.6, 129.5, 129.3, 128.9, 120.9, 68.2, 40.5, 37.9, 30.3, 21.7, 8.8; HRMS (ESI) calcd for  $C_{18}H_{19}NO_4SNa m/z = 368.0927$ ; found m/z = 368.0926 [(M+Na)<sup>+</sup>]; IR (KBr): v 2972, 2359, 1725, 1688, 1353, 1155, 665 cm<sup>-1</sup>;  $[\alpha]_D^{23} = -48$  (c 2.5, CHCl<sub>3</sub>) for 92% ee; Enantiomeric excess: 70% for Method A, 92% for Method B, determined by HPLC (Daicel Chiralpak AD-H, hexane/EtOH = 2/1; flow rate 1.0 ml/min; 25°C; 221 nm) first peak (Major):  $t_R = 14.6$  min, second peak (Minor):  $t_R = 18.4$ min.



**4i**; 51% yield for **Method A**, 95% yield for **Method B**; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.00 (d, J = 8.2 Hz, 2H), 7.36 (d, J = 8.2 Hz, 2H), 7.13 (dd, J = 10.7, 1.7 Hz, 1H), 6.17-6.24 (m, 3H), 5.50 ( J = 10.7 Hz, 1H), 5.44 (d, J = 2.7 Hz, 1H), 5.35 (d, J = 17.2 Hz, 1H), 3.18 (m, 1H), 2.75 (m, 2H), 2.45 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>) δ 194.4, 165.1, 145.6, 143.6, 138.0, 136.8, 135.8, 130.4, 129.7, 128.8, 120.9, 117.3, 68.3, 44.1, 34.5, 21.7; HRMS (ESI) calcd for  $C_{18}H_{17}NO_4SNa m/z = 366.0770$ ; found  $m/z = 366.0760 [(M+Na)^+]$ ; IR (KBr): v 3003, 2357, 1735, 1690, 1515, 1362, 1173, 666 cm<sup>-1</sup>;  $[\alpha]_D^{22} = -71$  (c 0.38, CHCl<sub>3</sub>) for 96% ee; Enantiomeric

excess: 80% for Method A, 96% for Method B, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 9/1; flow rate 1.0 ml/min; 25°C; 225 nm) first peak (Major):  $t_R = 29.4$  min, second peak (Minor):  $t_R = 33.7$  min.



**4j**; 93% yield for **Method A**; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.38-7.47 (m, 5H), 7.26 (d, J = 10.4 Hz, 1H), 6.31-6.37 (m, 2H), 5.56 (d, J = 2.7 Hz, 1H), 3.47 (q, J = 2.7 Hz, 1H), 3.36 (s, 3H), 2.63-2.78 (m, 2H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>) δ 194.3, 166.8, 143.5, 139.4, 138.1, 130.6, 129.2, 128.8, 125.1, 121.5, 69.7, 48.6, 43.1, 34.7; HRMS (ESI) calcd for  $C_{16}H_{15}NO_4SNa m/z = 340.0614$ ; found m/z =  $340.0616 [(M+Na)^+]$ ; IR (KBr): v 2354, 1731, 1672, 1353, 1145, 750 cm<sup>-1</sup>;  $[\alpha]_D^{24} = -90$  (c 1.1, CHCl<sub>3</sub>) for

73% ee; Enantiomeric excess: 73%, determined by HPLC (Daicel Chiralpak AD-H, hexane/EtOH = 2/1; flow rate 1.0 ml/min; 25°C; 216 nm) first peak (Minor):  $t_R = 13.6$  min, second peak (Major):  $t_R = 20.8$  min.



**4k** 74% yield for **Method A**, 83% yield for **Method B**; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) δ 7.04 (dd, J = 10.5, 1.6 Hz, 1H), 6.32 (d, J = 3.2 Hz, 1H), 6.01 (d, J = 10.5 Hz, 1H), 5.57 (d, J = 3.2 Hz, 1H),3.39 (s, 3H), 3.26 (qd, J = 3.2, 1.6 Hz, 1H), 2.83 (d, J = 5.0 Hz, 2H), 1.93 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>) δ 194.1, 166.6, 145.9, 138.1, 128.4, 121.7, 64.9, 45.1, 43.1, 36.1, 25.2; HRMS (ESI) calcd for

 $C_{11}H_{13}NO_4SNa m/z = 278.0463$ ; found m/z = 278.0449 [(M+Na)<sup>+</sup>]; IR (KBr): v 3013, 2926, 2357, 1722, 1684, 1357, 1231, 1164, 971 cm<sup>-1</sup>;  $[\alpha]_D^{22} = -130$  (c 0.21, CHCl<sub>3</sub>) for 90% ee; Enantiomeric excess: 63% for **Method A**, 90% for **Method B**, determined by HPLC (Daicel Chiralpak AD-H, hexane/EtOH = 2/1; flow rate 1.0 ml/min; 25°C; 215 nm) first peak (Major):  $t_R = 17.9$  min, second peak (Minor):  $t_R = 46.5$  min.



4l; 53% yield for Method A, 81% yield for Method B; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) δ 7.98 (d, J = 8.2 Hz, 2H), 7.35 (d, J = 8.2 Hz, 2H), 6.01 (s, 1H), 5.84 (s, 1H), 5.29 (s, 1H), 2.62 (m, 2H), 2.44 (s, 3H), 2.31 (d, J = 1.4 Hz, 3H), 1.88 (s, 3H), 0.95 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  194.4, 160.1, 145.4, 144.0, 135.5, 129.5, 128.7, 127.3, 118.4, 73.2, 48.5, 41.7, 24.8, 21.7, 21.3, 17.6; HRMS (ESI) calcd for  $C_{19}H_{21}NO_4SNa m/z = 382.1083$ ; found  $m/z = 382.1080 [(M+Na)^+]$ ; IR (KBr): v 3743, 3650, 2971,

2366, 1741, 1677, 1356, 1173, 1089, 816, 663 cm<sup>-1</sup>;  $[\alpha]_D^{26} = -87$  (c 0.31, CHCl<sub>3</sub>) for 98% ee; Enantiomeric excess: 84% for Method A, 98% for Method B, determined by HPLC (Daicel Chiralpak AD-H, hexane/EtOH = 2/1; flow rate 1.0 ml/min; 25°C; 216 nm) first peak (Major):  $t_R = 10.1$  min, second peak (Minor):  $t_R = 11.9$  min.

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**4m**; 90% yield for **Method B**; Colorless oil; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) δ 7.96 (d, *J* = 8.0 Hz, 2H), 7.34 (d, J = 8.0 Hz, 2H), 7.29 (d, J = 0.9 Hz, 1H), 6.16 (d, J = 2.3 Hz, 1H), 6.10 (d, J = 10.5 Hz, 1H), 5.46 (d, J = 2.3 Hz, 1H), 3..40-3.33 (m, 1H), 2.72-2.55 (m, 2H), 2.44 (s, 3H), 2.41-2.29 (m, 1H), 2.21-2.02 (m, 1H), 1.49-1.22 (m, 4H), 0.95 (t, J = 7.1 Hz, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  194.9, 165.7, 146.7, 145.5,

139.1, 135.6, 129.5, 129.1, 128.9, 120.9, 67.8, 41.1, 38.0, 37.3, 26.4, 22.8, 21.7, 13.9; HRMS (ESI) calcd for C<sub>20</sub>H<sub>23</sub>NO<sub>4</sub>SNa m/z = 396.1240; found  $m/z = 396.1236 [(M+Na)^+]$ ; IR (KBr): v 2922, 2853, 2361, 1729, 1697, 1355, 1086, 802, 590 cm<sup>-1</sup>;  $[\alpha]_D^{25} = -39$  (c 0.36, CHCl<sub>3</sub>) for 89% ee; Enantiomeric excess: 89%, determined by HPLC (Daicel Chiralpak ID, hexane/EtOH = 2/1; flow rate 1.0 ml/min; 25°C; 215 nm) first peak (Minor): t<sub>R</sub> = 11.5 min, second peak (Major): t<sub>R</sub> = 14.3 min.

#### 7. Screening of chiral catalysts.

[c] Isolated yield.

[d] Polymerization of 3g mainly proceeded.

	~~°	Chiral catalyst		~=0					
N Ts		CH₂Cl₂, 25 °C ≥90% conv	N Ts	<i></i>	Et <sub>2</sub> N	NHBz	Ph <sub>2</sub> P NHBz	: Ph <sub>2</sub> P	NHTs
3g		· 00 /0 00111.	4g		5a		5b		5c
entry	chiral amine	time (h)	NMR yield (%) <sup>[a]</sup>	ee (%) <sup>[b]</sup>		Δ.	Δ.		$\mathbf{X}$
1	5a	48	97 (95) <sup>[c]</sup>	94					¥
2	5b	0.1	18 <sup>[d]</sup>	90	Et <sub>o</sub> N	NHBoc		Et <sub>o</sub> N	NHBZ
3	5c	0.1	12 <sup>[d]</sup>	90					-1 ()
4	5d	48	87	67	50 (	(Ref. 4)	<b>5e</b> (Ref. 5)	<b>5</b> 1 (Re	er. 6)
5	5e	48	95	39				/	
6	5f	48	93	90	0—	$\neg$	$\land$		
7	β-ICD	96	87	70		$-\beta$	N-	$\prec$	
8	9	1.5	97	7		•N~/	ОН	-N	$\square$
9	10	96	40	15	N ≥		ОН		N∕∽⊂CO₂H
[a] 1.3.5-Tr	imethoxvbenzene v	vas used as an int	ernal standard.						Ме
[b] Determi	ned by HPLC (Daid	el Chiralpak AD-F	1)				<b>9</b> (Re	f. 8)	<b>10</b> (Ref. 9)

β-ICD (Ref. 7)

#### 8. Further optimization of reaction conditions.

	TsHN Ph	) mol%) base blvent, temp, ti	(1.5 eq.) me ►	O= N Ts Ph		
<b>1</b> (1.2 eq.)	2a			4a single diaste	reomer	3a
(				on gro and ore		not observed
entry	base	solvent	temp. (°C)	time (h)	NMR yiel	d (%) <sup>[a]</sup> ee (%) <sup>[b]</sup>
1 <sup>[c]</sup>	-	$CH_2CI_2$	25	17	84	85
2	proton sponge	$CH_2CI_2$	25	72	51	64
3	proton sponge	CHCI <sub>3</sub>	25	72	44	63
4	proton sponge	toluene	25	72	0	-
5	proton sponge	THF	25	72	0	-
6	tetramethylguanidine	$CH_2CI_2$	25	48	5>	-
7	DBU	$CH_2CI_2$	25	12	25	0
8	DIPEA	CH <sub>2</sub> Cl <sub>2</sub>	25	12	82	76
9	DIPEA	CHCI <sub>3</sub>	25	12	24	83
10	DIPEA	PhCI	25	12	67	59
11	DIPEA	$CH_2CI_2$	reflux	8	95 (93	2) <sup>[d]</sup> 80
12 <sup>[e]</sup>	DIPEA	$CH_2CI_2$	reflux	24	85	78
13	DIPEA	CH <sub>2</sub> Cl <sub>2</sub>	-20	60	72	81

[a] 1,3,5-Trimethoxybenzene was used as an internal standard.

[b] Determined by HPLC (Daicel Chiralpak IC).

[c] 150 mol% of **5a** was used.

[d] Isolated yield.

[e] 10 mol% of 5a was used.

#### 9. Synthesis of amine catalyst 5a



A solution of **5e** (0.32 mmol) and triethylamine (0.35 mmol) in dichloromethane (1.6 mL) was treated with benzoylchloride (0.35 mmol) at 0 °C. The reaction mixture was warmed to room temperature and stirred for 0.5 h. The reaction was quenched with saturated NaHCO3 aq. (3 mL) followed by extraction with EtOAc (3 mL). The separated organic layer was dried over Na2SO4 and reduced in *vacuo*. The obtained residue was purified by silica gel column chromatography using hexane, ethyl acetate, and triethylamine (10:10:1) mixed eluent, giving **5a** in 66% yield as a white solid.

**5a**; White solid; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.76-7.78 (m, 2H), 7.40-7.50 (m, 3H), 6.42 (d, *J* = 6.0 Hz, 1H), 4.03 (qd, *J* = 7.3, 5.0 Hz, 1H), 2.45-2.62 (m, 6H), 2.15-2.23 (m, 1H), 0.95-1.00 (m, 12H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  167.6, 135.3, 131.1, 128.5, 126.8, 53.0, 52.3, 47.0, 29.8, 18.6, 17.9, 11.8; HRMS **5a** (ESI) calcd for C<sub>16</sub>H<sub>26</sub>NO<sub>4</sub>SNa m/z = 285.1937; found m/z = 285.1938 [(M+Na)<sup>+</sup>]; IR (KBr): v 3313, 2964, 1634, 1546, 1187, 696 cm<sup>-1</sup>; [ $\alpha$ ]<sub>D</sub><sup>26</sup> = +18 (c 1.0, CHCl<sub>3</sub>) for >99% ee.

#### 10. Procedure for reuse of catalyst 5a in the stereoselective RC reaction of 3g<sup>a,b</sup>

	30	cat. <b>5a</b>	) 	a	
	Jg	CH <sub>2</sub> Cl <sub>2</sub> ,	3h 4	9	
Cycle	1st	2nd	3rd	4th	5th
Results	95% 94% ee	95% 94% ee	95% 94% ee	95% 94% ee	95% 94% ee

[a] Reaction conditions: 3e (0.10 mmol), cat. 5a (20 mol%) and CH<sub>2</sub>Cl<sub>2</sub> (0.5 mL).

[b] Determined by HPLC (Daicel Chiralpak AD-H).

After the full conversion of **3g**, diethyl ether (1.5 mL) and saturated aq.  $NH_4Cl(2.0 \text{ mL})$  was added to the reaction mixture. Evaporation of the separated organic layer gave product **4g**. To the aqueous layer was basified with saturated aq.  $NaHCO_3$  followed by extraction using CHCl<sub>3</sub>-MeOH (4:1) mixed solvent (2 mL×2). Evaporation of the organic layer gave catalyst **5a**, which can be used for the next reaction without further purification.

#### 11. References

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#	Name	RT Hei	ight [uAU]	Area[uAU.Sec]	%Area	#	Name	RT	Height[uAU]	Area[uAU.Sec]	%Area
1 2		13. 560 20. 827	267630 166091	8452742. 926 8412256. 925	50. 12 49. 88	1 2		13. 6 20. 9	527 71818 960 252684	2230404. 033 14117326. 610	13.64 86.36

![](_page_57_Figure_0.jpeg)

![](_page_58_Figure_0.jpeg)

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