

Supporting Information for

**Rhodium-catalyzed Asymmetric Hydrogenation of
Tetrasubstituted β -acetoxy- α -enamido Esters and Efficient
Synthesis of Droxidopa**

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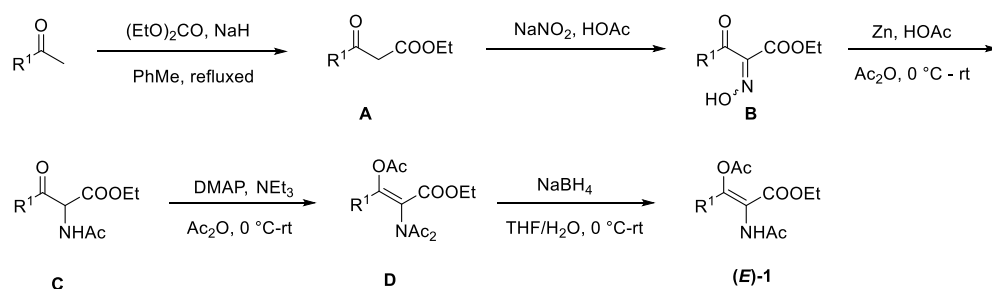
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1. General Information

Unless otherwise noted, all reagents and solvents were purchased from commercial suppliers and used without further purification. NMR spectra were recorded on Bruker ADVANCE III (400 MHz) spectrometers for ^1H NMR and ^{13}C NMR. CDCl_3 was the solvent used for the NMR analysis, with tetramethylsilane as the internal standard. Chemical shifts were reported upfield to TMS (0.00 ppm) for ^1H NMR and relative to CDCl_3 (77.0 ppm) for ^{13}C NMR. Optical rotation was determined using a Perkin Elmer 343 polarimeter. HPLC analysis was conducted on an Agilent 1260 Series instrument. Column Chromatography was performed with silica gel Merck 60 (300-400 mesh). All new products were further characterized by HRMS. A positive ion mass spectrum of sample was acquired on a Thermo LTQ-FT mass spectrometer with an electrospray ionization source.

2. General Procedure for the Synthesis of Compound 1



To a stirred solution of ketones (25 mmol) in toluene (120 mL) was added ethyl dicarbonate (75 mmol) and NaH (50 mmol, 60%). The reaction mixture was refluxed until TLC indicated the total consumption of the ketones. After cooling, the reaction mixture was quenched by ice water, acidified with 3 M HCl to pH 2~3 and extracted with EtOAc ($100\text{ mL} \times 3$). The combined organic layer was dried over sodium sulfate and evaporated under reduced pressure. The obtained-keto ester **A** was used directly for the next step without further purifications.^[1]

To a solution of β -keto ester **A** (20 mmol) in HOAc (5.7 mL, 100 mmol) was added NaNO_2 (2.07 g, 30 mmol) in H_2O (6 mL) at $0\text{ }^\circ\text{C}$. The reaction was monitored by TLC. When the reaction was completed, the reaction mixture was treated with H_2O (50 mL) and CH_2Cl_2 (50 mL), the mixture was stirred for 10 minutes and the phases were separated. The aqueous layer was extracted with CH_2Cl_2 ($3 \times 30\text{ mL}$). The combined organic solution was washed with saturated NaHCO_3 (150 mL) and brine

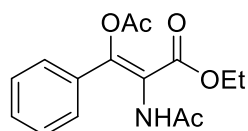
(150 mL), then dried over Na₂SO₄. After evaporation of the solvent, the residue was subjected to chromatography or crystallization to afford the desired α -hydroxyimino- β -ketoester **B**, yielding 95-99%.^[2]

To a stirred ice-cold solution of **B** (10 mmol) in Ac₂O (15-30 mL) was added Zn powder (3.25 g, 10 mmol) in small portion. Then HOAc (5.7 mL, 100 mmol) was added drop-wise from the dropping funnel to the heterogeneous solution at 0 °C. The reaction. After completion of the reaction, the mixture was filtered and the residue was washed with CH₂Cl₂ (100 mL). The filtrate was treated with saturated NaHCO₃ and separated. The organic phase was washed with brine (100 mL), then dried over Na₂SO₄. After evaporation of the solvent, the residue was subject to chromatography (PE/EA 2/1) to get the desired product α -acetamido- β -ketoester **C** in 70-92% yield.^[3]

To a stirred solution of **C** (4.26 mmol) in acetic anhydride (16 mL) was added Et₃N (1.85 mL, 13.4 mmol), and DMAP (0.0052 g, 0.043 mmol) at 0 °C. After 0.5 h at 0 °C, the reaction was kept at room temperature and monitored by TLC. The reaction mixture was treated with saturated NaHCO₃ and separated. The organic phase was washed with brine (50 mL), then dried over Na₂SO₄. After evaporation of the solvent, the residue was subject to chromatography (PE/EA 8/1) to get the desired product α -acetamido- β -ketoester **D**, yielding 78-90%.^[4]

Sodium borohydride (76.5 mg, 2.0 mmol) in water (0.6 mL) was added dropwise to a stirred solution of **D** (1 mmol) in tetrahydrofuran (40 mL) at 0 °C. After 20 min, the mixture was allowed to warm up to room temperature and then stirring was continued for 1 h. The reaction was quenched with brine (2 mL), and the resulting mixture was diluted with ethyl acetate (200 mL). The organic layer was washed with brine and dried over Na₂SO₄. Concentration of the solvent in vacuo gave a residue, which was purified by column chromatography (PE/EA 1/1) to give (*E*)-**1** as a white solid, yielding 40-70%.^[5]

(*E*)-ethyl-2-acetamido-3-acetoxy-3-phenylacrylate ((*E*)-1a)



White solid; m. p. 142-145 °C; 150 mg; ¹H NMR (400 MHz, CDCl₃) δ = 7.48-7.50 (m, 2H), 7.40-7.42 (m, 3H), 6.99 (s, 1H), 4.28 (q, *J* = 7.1 Hz, 1H), 2.23 (s, 3H), 2.00 (s, 3H), 1.33 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ = 169.22,

168.36, 163.41, 148.52, 133.16, 130.09, 128.68, 127.88, 119.03, 61.59, 22.81, 20.81, 14.14. ESI-HRMS calculated for $C_{15}H_{18}NO_5^+$ ($[M+H]^+$): 292.1179, found 292.1175. The structure of (*E*)-**1a** was determined by X-Ray (figure 1).

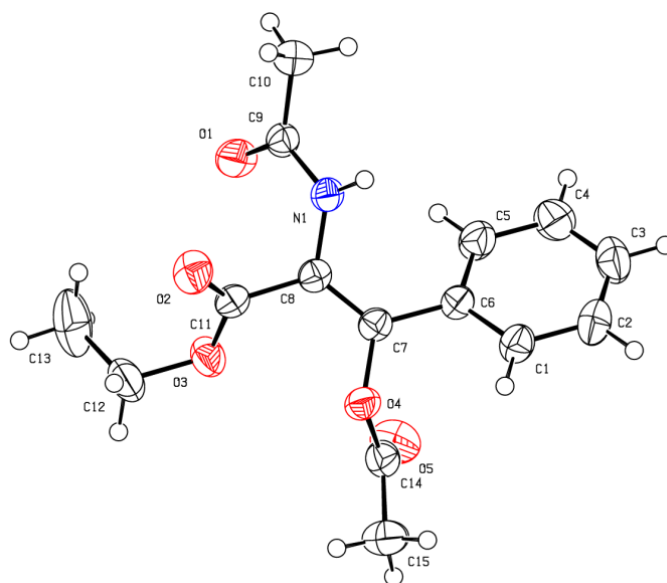
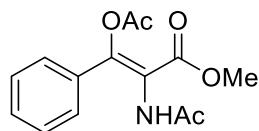


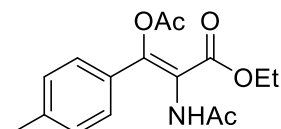
Figure 1. the X-ray structure of (*E*)-**1a**.

(*E*)-methyl-2-acetamido-3-acetoxy-3-phenylacrylate ((*E*)-1b)



White solid; m. p. 183-186 °C; 130 mg; 1H NMR (400 MHz, $CDCl_3$) δ = 7.46-7.49 (m, 2H), 7.39-7.43 (m, 3H), 6.73 (s, 1H), 3.81 (s, 3H), 2.23 (s, 3H), 2.01 (s, 3H); ^{13}C NMR (101 MHz, $CDCl_3$) δ = 169.14, 168.48, 163.78, 149.02, 133.12, 130.24, 128.78, 127.94, 118.65, 52.60, 22.94, 20.83. ESI-HRMS calculated for $C_{14}H_{16}NO_5^+$ ($[M+H]^+$): 278.1023, found 278.1016.

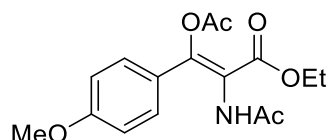
(*E*)-ethyl-2-acetamido-3-acetoxy-3-(*p*-tolyl)acrylate ((*E*)-1c)



White solid; m. p. 174-176 °C; 145 mg; 1H NMR (400 MHz, $CDCl_3$) δ = 7.36-7.38 (m, 2H), 7.19-7.21 (m, 2H), 6.72 (s, 1H), 4.27 (q, J = 7.1 Hz, 2H), 2.37 (s, 3H), 2.21 (s, 3H), 2.01 (s, 3H), 1.31 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, $CDCl_3$) δ =

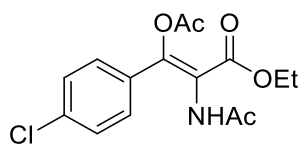
169.30, 168.39, 163.54, 148.87, 140.41, 130.24, 129.38, 127.78, 118.48, 61.50, 22.77, 21.47, 20.82, 14.14. ESI-HRMS calculated for $C_{16}H_{20}NO_5^+$ ($[M+H]^+$): 306.1336, found 306.1333.

(E)-ethyl-2-acetamido-3-acetoxy-3-(4-methoxyphenyl)acrylate ((E)-1d)



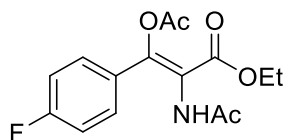
White solid; m. p. 161-164 °C; 155 mg; 1H NMR (400 MHz, $CDCl_3$) δ = 7.42-7.44 (m, 2H), 6.72-6.91 (m, 2H), 6.72 (s, 1H), 4.26 (q, J = 7.1 Hz, 2H), 3.83 (s, 3H), 2.22 (s, 3H), 2.02 (s, 3H), 1.31 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, $CDCl_3$) δ = 169.10, 168.44, 163.55, 160.91, 149.32, 129.55, 125.40, 117.72, 114.16, 61.53, 55.41, 22.99, 20.92, 14.20. ESI-HRMS calculated for $C_{16}H_{20}NO_6^+$ ($[M+H]^+$): 322.1285, found 322.1282.

(E)-ethyl-2-acetamido-3-acetoxy-3-(4-chlorophenyl)acrylate ((E)-1e)



White solid; m. p. 188-190 °C; 150 mg; 1H NMR (400 MHz, $CDCl_3$) δ = 7.41-7.43 (m, 2H), 7.35-7.38 (m, 2H), 4.27 (q, J = 7.1 Hz, 2H), 2.22 (s, 3H), 2.01 (s, 3H), 1.32 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, $CDCl_3$) δ = 168.97, 168.34, 163.24, 148.28, 136.14, 131.89, 129.25, 129.06, 119.04, 61.80, 23.01, 20.84, 14.18. ESI-HRMS calculated for $C_{15}H_{15}NO_5Cl^-$ ($[M-H]^-$): 324.0644, found 324.06447.

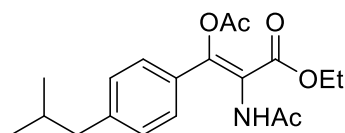
(E)-ethyl-2-acetamido-3-acetoxy-3-(4-fluorophenyl)acrylate ((E)-1f)



White solid; m. p. 135-137 °C; 132 mg; 1H NMR (400 MHz, $CDCl_3$) δ = 7.46-7.50 (m, 2H), 7.06-7.10 (m, 2H), 6.73 (s, 1H), 4.27 (q, J = 7.1 Hz, 2H), 2.22 (s, 3H), 2.00 (s, 3H), 1.32 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, $CDCl_3$) δ = 169.02, 168.35, 164.66, 163.31, 148.62, 130.13, 130.04, 129.50, 118.74, 116.06, 115.84, 61.73, 22.99,

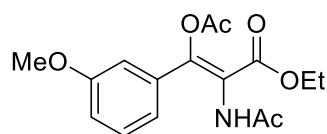
20.86, 14.18. ESI-HRMS calculated for $C_{15}H_{17}NO_5F^+$ ($[M+H]^+$): 310.1085, found 310.1077.

(E)-ethyl-2-acetamido-3-acetoxy-3-(4-isobutylphenyl)acrylate ((E)-1g)



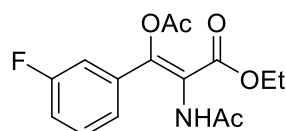
White solid; m. p. 127-130 °C; 152 mg; 1H NMR (400 MHz, $CDCl_3$) δ = 7.38 (d, J = 8.2 Hz, 2H), 7.17 (d, J = 8.2 Hz, 2H), 6.73 (s, 1H), 4.27 (q, J = 7.1 Hz, 2H), 2.48 (d, J = 7.2 Hz, 2H), 2.22 (s, 3H), 2.02 (s, 3H), 1.85-1.90 (m, 1H), 1.31 (t, J = 7.1 Hz, 3H), 0.91 (d, J = 6.6 Hz, 6H); ^{13}C NMR (101 MHz, $CDCl_3$) δ = 169.12, 168.42, 163.47, 148.78, 144.29, 130.41, 129.47, 127.70, 118.55, 61.55, 45.30, 30.13, 22.93, 22.44, 20.90, 14.18. ESI-HRMS calculated for $C_{19}H_{26}NO_5^+$ ($[M+H]^+$): 348.1805, found 348.1797.

(E)-ethyl-2-acetamido-3-acetoxy-3-(3-methoxyphenyl)acrylate ((E)-1h)



White solid; m. p. 123-125 °C; 128 mg; 1H NMR (400 MHz, $CDCl_3$) δ = 7.30-7.34 (m, 1H), 7.05-7.07 (m, 1H), 7.00-7.01 (m, 1H), 6.91-6.94 (m, 1H), 4.28 (q, J = 7.1 Hz, 2H), 3.80 (s, 3H), 2.21 (s, 3H), 2.02 (s, 3H), 1.32 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, $CDCl_3$) δ = 168.91, 168.35, 163.33, 159.68, 147.73, 134.40, 129.91, 120.23, 119.25, 115.78, 113.36, 61.67, 55.40, 22.94, 20.83, 14.18. ESI-HRMS calculated for $C_{16}H_{20}NO_6^+$ ($[M+H]^+$): 322.1285, found 322.1283.

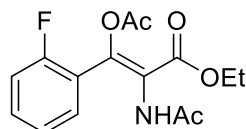
(E)-ethyl-2-acetamido-3-acetoxy-3-(3-fluorophenyl)acrylate ((E)-1i)



White solid; m. p. 112-115 °C; 123 mg; 1H NMR (400 MHz, $CDCl_3$) δ = 7.35-7.40 (m, 1H), 7.28-7.30 (m, 1H), 7.17-7.20 (m, 1H), 7.05-7.09 (m, 1H), 6.73 (s, 1H), 4.29 (q, J = 7.1 Hz, 2H), 2.23 (s, 3H), 2.03 (s, 3H), 1.32 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101

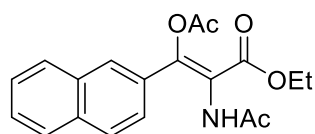
MHz, CDCl₃) δ = 168.82, 168.29, 163.17, 161.43, 130.49, 130.41, 123.62, 117.28, 117.07, 114.99, 114.76, 61.84, 23.05, 20.82, 14.18. ESI-HRMS calculated for C₁₅H₁₅NO₅F⁻ ([M-H]⁻): 308.0940, found 308.0942.

(E)-ethyl-2-acetamido-3-acetoxy-3-(2-fluorophenyl)acrylate ((E)-1j)



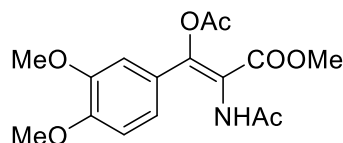
White solid; m. p. 137-139 °C; 117 mg; ¹H NMR (400 MHz, CDCl₃) δ = 7.39-7.44 (m, 2H), 7.10-7.21 (m, 2H), 6.73 (s, 1H), 4.28 (q, *J* = 7.1 Hz, 2H), 2.20 (s, 3H), 1.97 (s, 3H), 1.32 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ = 168.78, 168.08, 162.82, 160.63, 158.13, 144.12, 132.10, 132.01, 130.19, 130.17, 124.52, 124.49, 121.28, 121.02, 116.38, 116.16, 61.72, 22.85, 20.76, 14.14. ESI-HRMS calculated for C₁₅H₁₇NO₅F⁺ ([M+H]⁺): 310.1085, found 310.1077.

(E)-ethyl-2-acetamido-3-acetoxy-3-(naphthalen-2-yl)acrylate ((E)-1k)



White solid; m. p. 162-164 °C; 238 mg; ¹H NMR (400 MHz, CDCl₃) δ = 7.95 (s, 1H), 7.83-7.86 (m, 3H), 7.52-7.56 (m, 3H), 6.79 (s, 1H), 4.30 (q, *J* = 7.1 Hz, 2H), 2.24 (s, 3H), 2.00 (s, 3H), 1.33 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ = 169.03, 168.45, 163.42, 148.63, 133.78, 132.86, 130.62, 128.62, 128.53, 128.10, 127.81, 127.59, 126.85, 124.64, 119.19, 61.70, 22.96, 20.90, 14.21. ESI-HRMS calculated for C₁₉H₂₀NO₅⁺ ([M+H]⁺): 342.1336, found 342.1332.

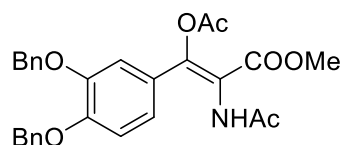
(E)-methyl-2-acetamido-3-acetoxy-3-(3,4-dimethoxyphenyl)acrylate ((E)-1l)



White solid; m. p. 174-176 °C; 150 mg; ¹H NMR (400 MHz, CDCl₃) δ = 7.06-7.09 (m, 1H), 7.00 (d, *J* = 2.0 Hz, 1H), 6.87 (d, *J* = 8.4 Hz, 1H), 6.79 (s, 1H), 3.90 (s, 3H), 3.85 (s, 3H), 3.81 (s, 3H), 2.23 (s, 3H), 2.03 (s, 3H); ¹³C NMR (101

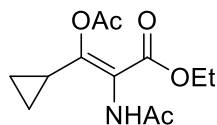
MHz, CDCl₃) δ = 169.16, 168.57, 163.88, 150.60, 149.50, 148.82, 125.44, 121.29, 117.56, 110.94, 110.66, 55.97, 52.54, 22.94, 20.87. ESI-HRMS calculated for C₁₆H₂₀NO₇⁺ ([M+H]⁺): 338.1232, found 338.1229.

(E)-methyl-2-acetamido-3-acetoxy-3-(3,4-bis(benzyloxy)phenyl)acrylate ((E)-1m)



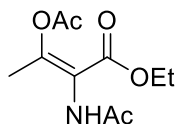
White solid; m. p. 122-125 °C; 250 mg; ¹H NMR (400 MHz, CDCl₃) δ 7.49-7.28 (m, 10 H), 7.02 (s, 1H), 6.91 (d, *J* = 8.9 Hz, 1H), 6.66 (s, 1H), 5.18 (s, 2H), 5.14 (s, 2H), 3.78 (s, 3H), 2.13 (s, 3H), 1.90 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 169.08, 168.46, 163.85, 150.53, 148.86, 148.50, 136.83, 136.64, 128.67, 128.08, 128.01, 127.21, 127.17, 125.80, 121.86, 117.66, 114.21, 113.88, 71.20, 70.91, 52.51, 22.89, 20.76. ESI-HRMS calculated for C₂₈H₂₈NO₇⁺ ([M+H]⁺): 490.1858, found 490.1855.

(E)-ethyl-2-acetamido-3-acetoxy-3-cyclopropylacrylate ((E)-1n)



White solid; 120 mg; ¹H NMR (400 MHz, CDCl₃) δ = 6.83 (s, 1H), 4.16 (q, *J* = 7.1 Hz, 2H), 2.19 (s, 3H), 2.14 (s, 3H), 1.89-1.93 (m, 1H), 1.25 (t, *J* = 7.1 Hz, 3H), 0.80-0.98 (m, 4H); ¹³C NMR (101 MHz, CDCl₃) δ = 169.61, 168.29, 162.74, 159.08, 116.06, 61.17, 23.21, 20.85, 14.19, 12.87, 6.76. ESI-HRMS calculated for C₁₂H₁₈NO₅⁺ ([M+H]⁺): 256.1179, found 256.1177.

(E)-ethyl-2-acetamido-3-acetoxybut-2-enoate ((E)-1o)



White solid; 133 mg; ¹H NMR (400 MHz, CDCl₃) δ = 6.76 (s, 1H), 4.20 (q, *J* = 7.1 Hz, 2H), 2.22 (s, 3H), 2.12 (s, 3H), 2.01 (s, 3H), 1.28 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ = 168.78, 168.37, 162.93, 155.78, 117.24, 61.46, 23.30, 20.99, 19.28, 14.19. ESI-HRMS calculated for C₁₀H₁₆NO₅⁺ ([M+H]⁺): 230.1023, found 230.1018.

3. General Procedure for Asymmetric Hydrogenation of Compound 1

A stock solution was made by mixing $[\text{Rh}(\text{COD})_2]\text{BF}_4$ with (*R_C,S_P*)-DuanPhos in a 1:1.1 molar ratio in CH_2Cl_2 at room temperature for 30 min in a nitrogen-filled glovebox. An aliquot of the catalyst solution (0.5 mL, 0.001 mmol) was transferred by syringe into the vials charged with different substrates (0.05 mmol for each). The vials were subsequently transferred into an autoclave into which hydrogen gas was charged. The reaction was then stirred under H_2 (30 atm) at room temperature for 24 h. The hydrogen gas was released slowly and carefully. The solution was concentrated and passed through a short column of silica gel (eluent: EtOAc) to remove the metal complex. The ee values of all compounds **2** were determined by HPLC analysis on a chiral stationary phase.

Procedure for Gram-scale Asymmetric Hydrogenation of (*E*)-**1a**

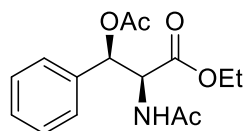
A stock solution was made by mixing $[\text{Rh}(\text{COD})_2]\text{BF}_4$ with (*R_C,S_P*)-DuanPhos in a 1:1.1 molar ratio in CH_2Cl_2 at room temperature for 45 min in a nitrogen-filled glovebox. An aliquot of the catalyst solution (2 mL, 0.068 mmol) was transferred by syringe into the vial charged with (*E*)-**1a** (3.4 mmol) in anhydrous CH_2Cl_2 (5 mL). The vials were subsequently transferred into an autoclave into which hydrogen gas was charged. The reaction was then stirred under H_2 (30 atm) at room temperature for 30 h. The hydrogen gas was released slowly and carefully. The solution was concentrated and passed through a short column of silica gel (eluent: EtOAc) to remove the metal complex. The ee values of **2a** were determined by HPLC analysis on a chiral stationary phase.

Procedure for TON study of Asymmetric Hydrogenation of (*E*)-**1a**

A stock solution was made by mixing $[\text{Rh}(\text{COD})_2]\text{BF}_4$ with (*R_C,S_P*)-DuanPhos in a 1:1.1 molar ratio in CH_2Cl_2 at room temperature for 45 min in a nitrogen-filled glovebox. An aliquot of the catalyst solution (0.2 mL, 0.0004 mmol) was transferred by syringe into the vial charged with (*E*)-**1a** (0.2 mmol) in anhydrous CH_2Cl_2 (2 mL). The vials were subsequently transferred into an autoclave into which hydrogen gas was charged. The reaction was then stirred under H_2 (70 atm) at 60 °C for 24 h. The hydrogen gas was released slowly and carefully. The solution was concentrated and passed through a short column of silica gel (eluent: EtOAc) to remove the metal

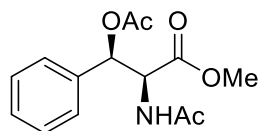
complex. The ee values of **2a** were determined by HPLC analysis on a chiral stationary phase.

(2*S*,3*R*)-ethyl-2-acetamido-3-acetoxy-3-phenylpropanoate (2a)



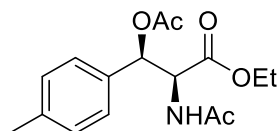
White solid; m. p. 68-70 °C; 14.6 mg, 99% yield; >99% ee; $[\alpha]_D^{20} = -36.70$ ($c = 1.00$, CHCl_3); The enantiomeric excess was determined by HPLC on Chiralpak OJ-H column, hexane: isopropanol = 95:5; flow rate = 1.0 mL/min; UV detection at 210 nm; $t_R = 21.2$ min (major), 29.7 min (minor). ^1H NMR (400 MHz, CDCl_3) $\delta = 7.32$ -7.39 (m, 5H), 6.30 (d, $J = 4.2$ Hz, 1H), 6.18 (d, $J = 9.2$ Hz, 1H), 5.09 (dd, $J = 9.2$ Hz, 4.2 Hz, 1H), 4.17 (q, $J = 7.1$ Hz, 2H), 2.16 (s, 3H), 1.97 (s, 3H), 1.22 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) $\delta = 169.85, 169.56, 169.53, 135.94, 128.66, 128.55, 126.29, 74.69, 61.97, 56.38, 23.00, 20.89, 14.02$. ESI-HRMS calculated for $\text{C}_{15}\text{H}_{20}\text{NO}_5^+$ ($[\text{M}+\text{H}]^+$): 294.1336, found 294.1334.

(2*S*,3*R*)-methyl-2-acetamido-3-acetoxy-3-phenylpropanoate (2b)



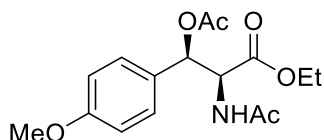
White solid; m. p. 135-136 °C; 13.9 mg; 99% yield; >99% ee; $[\alpha]_D^{20} = -29.33$ ($c = 0.60$, CHCl_3); The enantiomeric excess was determined by HPLC on Chiralpak OJ-H column, hexane: isopropanol = 95:5; flow rate = 1.0 mL/min; UV detection at 210 nm; $t_R = 39.4$ min (major), 54.8 min (minor). ^1H NMR (400 MHz, CDCl_3) $\delta = 7.28$ -7.37 (m, 5H), 6.28 (d, $J = 3.9$ Hz, 1H), 6.13 (d, $J = 9.2$ Hz, 1H), 5.07 (dd, $J = 9.3$ Hz, 3.9 Hz, 1H), 3.71 (s, 3H), 2.14 (s, 3H), 1.95 (s, 3H); ^{13}C NMR (101 MHz, CDCl_3) $\delta = 170.10, 169.83, 169.53, 135.91, 128.68, 128.60, 126.16, 74.53, 56.29, 52.80, 23.01, 20.92$. ESI-HRMS calculated for $\text{C}_{14}\text{H}_{18}\text{NO}_5^+$ ($[\text{M}+\text{H}]^+$): 280.1179, found 280.1173.

(2*S*,3*R*)-ethyl-2-acetamido-3-acetoxy-3-(*p*-tolyl)propanoate (2c)



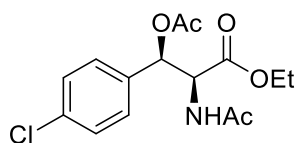
White solid; m. p. 123-158 °C; 15.1 mg; 98% yield; >99% ee; $[\alpha]_D^{20} = -14.08$ ($c = 0.71$, CHCl_3); The enantiomeric excess was determined by HPLC on Chiralpak OJ-H column, hexane: isopropanol = 95:5; flow rate = 1.0 mL/min; UV detection at 210 nm; $t_R = 17.6$ min (major), 23.9 min (minor). ^1H NMR (400 MHz, CDCl_3) $\delta = 7.13$ -7.20 (m, 4H), 6.24 (d, $J = 4.2$ Hz, 1H), 6.13 (d, $J = 9.3$ Hz, 1H), 5.04 (dd, $J = 9.3$ Hz, 4.2 Hz, 1H), 4.15 (q, $J = 7.1$ Hz, 2H), 2.33 (s, 3H), 2.12 (s, 3H), 1.96 (s, 3H), 1.20 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) $\delta = 169.88$, 169.63, 169.58, 138.47, 132.93, 129.25, 126.22, 74.62, 61.93, 56.37, 23.05, 21.23, 20.92, 14.02. ESI-HRMS calculated for $\text{C}_{16}\text{H}_{22}\text{NO}_5^+$ ($[\text{M}+\text{H}]^+$): 308.1492, found 308.1491.

(2S,3R)-ethyl-2-acetamido-3-acetoxy-3-(4-methoxyphenyl)propanoate (2d)



White solid; m. p. 129-131 °C; 15.7 mg; 97% yield; >99% ee; $[\alpha]_D^{20} = -2.96$ ($c = 0.30$, CHCl_3); The enantiomeric excess was determined by HPLC on Chiralpak OJ-H column, hexane: isopropanol = 94:6; flow rate = 1.0 mL/min; UV detection at 210 nm; $t_R = 34.9$ min (major), 49.1 min (minor). ^1H NMR (400 MHz, CDCl_3) $\delta = 7.22$ -7.26 (m, 2H), 6.85 – 6.88 (m, 2H), 6.21 (d, $J = 4.5$ Hz, 1H), 6.17 (d, $J = 9.3$ Hz, 1H), 5.03 (dd, $J = 9.3$ Hz, 4.5 Hz, 1H), 4.13 (q, $J = 7.1$ Hz, 2H), 3.79 (s, 3H), 2.11 (s, 3H), 1.97 (s, 3H), 1.19 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) $\delta = 169.85$, 169.62, 159.73, 127.95, 127.75, 113.89, 74.42, 61.87, 56.42, 55.26, 23.04, 20.92, 14.01. ESI-HRMS calculated for $\text{C}_{16}\text{H}_{22}\text{NO}_6^+$ ($[\text{M}+\text{H}]^+$): 324.1442, found 324.1437.

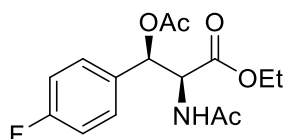
(2S,3R)-ethyl-2-acetamido-3-acetoxy-3-(4-chlorophenyl)propanoate (2e)



White solid; 16.1 mg; 98% yield; >99% ee; $[\alpha]_D^{20} = -14.61$ ($c = 0.70$, CHCl_3); The enantiomeric excess was determined by HPLC on Chiralpak OD-H column,

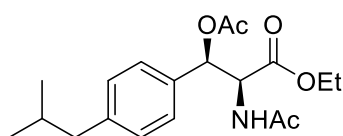
hexane: isopropanol = 95:5; flow rate = 1.0 mL/min; UV detection at 210 nm; t_R = 21.8 min (major), 29.3 min (minor). ^1H NMR (400 MHz, CDCl_3) δ = 7.30-7.33 (m, 2H), 7.22-7.25 (m, 2H), 6.24 (d, J = 4.0 Hz, 1H), 6.11 (d, J = 9.3 Hz, 1H), 5.05 (dd, J = 9.4 Hz, 4.0 Hz, 1H), 4.16 (q, J = 7.1 Hz, 2H), 2.12 (s, 3H), 1.95 (s, 3H), 1.22 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ = 169.83, 169.38, 169.35, 134.57, 134.55, 128.79, 127.69, 74.14, 62.14, 56.05, 23.02, 20.85, 14.06. ESI-HRMS calculated for $\text{C}_{15}\text{H}_{19}\text{NO}_5\text{Cl}^+$ ($[\text{M}+\text{H}]^+$): 328.0946, found 328.0943.

(2*S*,3*R*)-ethyl-2-acetamido-3-acetoxy-3-(4-fluorophenyl)propanoate (2f)



White solid; m. p. 167-169 °C; 15.4 mg; 99% yield; >99% ee; $[\alpha]_D^{20}$ = -17.43 (c = 0.70, CHCl_3); The enantiomeric excess was determined by HPLC on Chiralpak OJ-H column, hexane: isopropanol = 95:5; flow rate = 1.0 mL/min; UV detection at 210 nm; t_R = 23.6 min (major), 32.7 min (minor). ^1H NMR (400 MHz, CDCl_3) δ = 7.34 – 7.27 (m, 1H), 7.15 – 6.98 (m, 1H), 6.25 (d, J =4.2, 1H), 6.13 (d, J =9.3, 1H), 5.05 (dd, J =9.4, 4.2, 1H), 4.16 (q, J =7.1, 1H), 2.13 (s, 1H), 1.96 (s, 1H), 1.21 (t, J =7.1, 1H); ^{13}C NMR (101 MHz, CDCl_3) δ = 169.80, 169.45, 169.42, 163.96, 161.50, 131.87, 131.84, 128.22, 128.13, 115.68, 115.46, 74.15, 62.07, 56.23, 23.01, 20.87, 14.04. ESI-HRMS calculated for $\text{C}_{15}\text{H}_{19}\text{NO}_5\text{F}^+$ ($[\text{M}+\text{H}]^+$): 312.1242, found 312.1233.

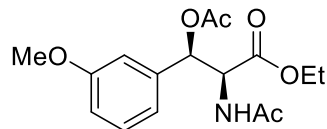
(2*S*,3*R*)-ethyl-2-acetamido-3-acetoxy-3-(4-isobutylphenyl)propanoate (2g)



White solid; 16.8 mg; 96% yield; >99% ee; $[\alpha]_D^{20}$ = -10.32 (c = 0.53, CHCl_3); The enantiomeric excess was determined by HPLC on Chiralpak OJ-H column, hexane: isopropanol = 97:3; flow rate = 1.0 mL/min; UV detection at 210 nm; t_R = 19.1 min (major), 23.5 min (minor). ^1H NMR (400 MHz, CDCl_3) δ = 7.21 (d, J = 8.1 Hz, 2H), 7.11 (d, J = 8.1 Hz, 2H), 6.25 (d, J = 4.5 Hz, 1H), 6.12 (d, J = 9.2 Hz, 1H), 5.04 (dd, J = 9.3 Hz, 4.5 Hz, 1H), 4.12 (q, J = 7.1 Hz, 2H), 2.45 (d, J = 7.2 Hz, 2H), 2.13 (s, 3H), 1.95 (s, 3H), 1.81-1.85 (m, 1H), 1.18 (t, J = 7.1 Hz, 1H), 0.87-0.89 (m, 6H); ^{13}C NMR (101 MHz, CDCl_3) δ = 169.84, 169.67, 142.28, 133.12, 129.27,

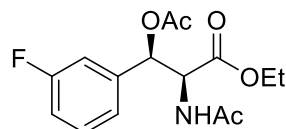
126.16, 74.63, 61.89, 56.46, 45.11, 30.20, 23.05, 22.39, 22.35, 20.95, 14.01.
ESI-HRMS calculated for $C_{19}H_{28}NO_5^+$ ($[M+H]^+$): 350.1962, found 350.1954.

(2*S*,3*R*)-ethyl-2-acetamido-3-acetoxy-3-(3-methoxyphenyl)propanoate (2h)



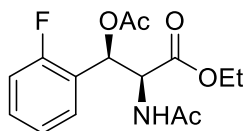
White solid; m. p. 134-135 °C; 15.7 mg; 97% yield; >99% ee; $[\alpha]_D^{20} = -16.13$ ($c = 0.50$, $CHCl_3$); The enantiomeric excess was determined by HPLC on Chiralpak OJ-H column, hexane: isopropanol = 95:5; flow rate = 1.0 mL/min; UV detection at 210 nm; $t_R = 31.1$ min (major), 37.4 min (minor). 1H NMR (400 MHz, $CDCl_3$) $\delta = 7.24$ -7.28 (m, 1H), 6.83-6.90 (m, 3H), 6.25 (d, $J = 4.0$ Hz, 1H), 6.10 (d, $J = 9.3$ Hz, 1H), 5.06 (dd, $J = 9.3$ Hz, 4.0 Hz, 1H), 4.16 (q, $J = 7.1$ Hz, 2H), 3.80 (s, 3H), 2.13 (s, 3H), 1.96 (s, 3H), 1.22 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (101 MHz, $CDCl_3$) $\delta = 168.78$, 168.46, 168.41, 158.59, 136.45, 128.60, 117.47, 112.66, 111.12, 73.41, 60.92, 55.24, 54.25, 21.99, 19.82, 12.98. ESI-HRMS calculated for $C_{16}H_{22}NO_6^+$ ($[M+H]^+$): 324.1442, found 324.1439.

(2*S*,3*R*)-ethyl-2-acetamido-3-acetoxy-3-(3-fluorophenyl)propanoate (2i)



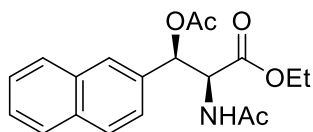
Pale oil; 14.9 mg; 96% yield; >99% ee; $[\alpha]_D^{20} = -27.87$ ($c = 0.93$, $CHCl_3$); The enantiomeric excess was determined by HPLC on Chiralpak OD-H column, hexane: isopropanol = 95:5; flow rate = 1.0 mL/min; UV detection at 210 nm; $t_R = 23.4$ min (major), 32.2 min (minor). 1H NMR (400 MHz, $CDCl_3$) $\delta = 7.29$ -7.33 (m, 1H), 7.10 (d, $J = 7.8$ Hz, 1H), 6.99-7.09 (m, 2H), 6.27 (d, $J = 3.8$ Hz, 1H), 6.14 (d, $J = 9.3$ Hz, 1H), 5.07 (dd, $J = 9.4$ Hz, 3.8 Hz, 1H), 4.18 (q, $J = 7.1$ Hz, 2H), 2.14 (s, 3H), 1.96 (s, 3H), 1.24 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (101 MHz, $CDCl_3$) $\delta = 169.85$, 169.31, 163.97, 161.52, 138.63, 138.56, 130.26, 130.18, 122.01, 121.98, 115.68, 115.47, 113.43, 113.20, 74.04, 62.14, 56.09, 22.97, 20.81, 14.05. ESI-HRMS calculated for $C_{15}H_{19}NO_5F^+$ ($[M+H]^+$): 312.1242, found 312.1238.

(2*S*,3*R*)-ethyl-2-acetamido-3-acetoxy-3-(2-fluorophenyl)propanoate (2j)



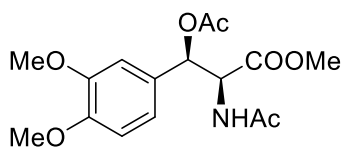
Pale oil; 15.1 mg; 97% yield; >99% ee; $[\alpha]_D^{20} = -26.43$ ($c = 0.70$, CHCl_3); The enantiomeric excess was determined by HPLC on Chiralpak OJ-H column, hexane: isopropanol = 95:5; flow rate = 1.0 mL/min; UV detection at 210 nm; $t_R = 25.5$ min (major), 30.6 min (minor). ^1H NMR (400 MHz, CDCl_3) $\delta = 7.25\text{--}7.29$ (m, 2H), 7.06–7.13 (m, 2H), 6.55 (d, $J = 3.6$ Hz, 1H), 6.16 (d, $J = 9.3$ Hz, 1H), 5.15 (dd, $J = 9.4$ Hz, 3.6 Hz, 1H), 4.20 (q, $J = 7.1$ Hz, 2H), 2.13 (s, 3H), 1.92 (s, 3H), 1.25 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) $\delta = 169.74, 169.29, 169.13, 161.00, 158.54, 130.34, 130.26, 127.28, 127.25, 124.04, 124.01, 123.69, 123.56, 115.79, 115.58, 69.85, 69.83, 62.12, 55.06, 22.94, 20.76, 14.07$. ESI-HRMS calculated for $\text{C}_{15}\text{H}_{19}\text{NO}_5\text{F}^+$ ($[\text{M}+\text{H}]^+$): 312.1242, found 312.1233.

(2S,3R)-ethyl-2-acetamido-3-acetoxy-3-(naphthalen-2-yl)propanoate (2k)



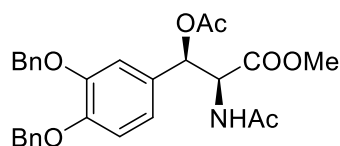
White solid; 17.1 mg; 99% yield; >99% ee; $[\alpha]_D^{20} = -4.88$ ($c = 0.47$, CHCl_3); The enantiomeric excess was determined by HPLC on Chiralpak OD-H column, hexane: isopropanol = 95:5; flow rate = 1.0 mL/min; UV detection at 210 nm; $t_R = 32.6$ min (major), 46.9 min (minor). ^1H NMR (400 MHz, CDCl_3) $\delta = 7.81\text{--}7.85$ (m, 3H), 7.75 (s, 1H), 7.42–7.50 (m, 3H), 6.45 (d, $J = 4.2$ Hz, 1H), 6.18 (d, $J = 9.3$ Hz, 1H), 5.18 (dd, $J = 9.3$ Hz, 4.2 Hz, 1H), 4.14 (q, $J = 7.1$ Hz, 2H), 2.18 (s, 3H), 1.93 (s, 3H), 1.16 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) $\delta = 169.88, 169.59, 169.58, 133.34, 133.26, 132.96, 128.52, 128.05, 127.81, 126.50, 125.61, 123.78, 74.81, 62.01, 56.33, 23.03, 20.95, 14.00$. ESI-HRMS calculated for $\text{C}_{19}\text{H}_{22}\text{NO}_5^+$ ($[\text{M}+\text{H}]^+$): 344.1492, found 344.1490.

(2S,3R)-methyl-2-acetamido-3-acetoxy-3-(3,4-dimethoxyphenyl)propanoate (2l)



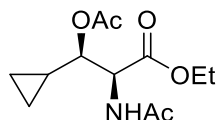
White solid; m. p. 139-140 °C; 16.6 mg; 98% yield; >99% ee; $[\alpha]_D^{20} = -7.75$ ($c = 0.80$, CHCl_3); The enantiomeric excess was determined by HPLC on Chiralpak OJ-H column, hexane: isopropanol = 80:20; flow rate = 1.0 mL/min; UV detection at 210 nm; $t_R = 23.0$ min (minor), 35.8 min (major). ^1H NMR (400 MHz, CDCl_3) δ 6.81-6.87 (m, 3H), 6.20 (d, $J = 4.6$ Hz, 1H), 6.17 (d, $J = 9.4$ Hz, 1H), 5.06 (dd, $J = 9.3$, 4.6 Hz, 1H), 3.88 (s, 3H), 3.87 (s, 3H), 3.68 (s, 3H), 2.13 (s, 3H), 1.99 (s, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 170.16, 169.86, 169.71, 149.24, 148.97, 128.24, 118.93, 110.85, 109.48, 74.38, 56.32, 55.99, 55.86, 52.74, 23.06, 20.95. ESI-HRMS calculated for $\text{C}_{16}\text{H}_{22}\text{NO}_7^+$ ($[\text{M}+\text{H}]^+$): 340.1388, found 340.1386.

(2*S*,3*R*)-methyl -2-acetamido-3-acetoxy-3-(3,4-bis(benzyloxy)phenyl)propanoate (2m)



White solid; m. p. 89-94 °C; 24.3 mg; 99% yield; >99% ee; $[\alpha]_D^{20} = -4.60$ ($c = 1.00$, CHCl_3); The enantiomeric excess was determined by HPLC on Chiralpak OD-H column, hexane: isopropanol = 80:20; flow rate = 1.0 mL/min; UV detection at 210 nm; $t_R = 19.8$ min (major), 36.3 min (minor). ^1H NMR (400 MHz, CDCl_3) δ 7.42-7.45 (m, 4H), 7.34-7.39 (m, 4H), 7.30-7.32 (m, 2H), 6.87-6.89 (m, 2H), 6.81 (dd, $J = 8.4$, 1.9 Hz, 1H), 6.14 (d, $J = 4.5$ Hz, 1H), 6.08 (d, $J = 9.2$ Hz, 1H), 5.14 (s, 1H), 5.00 (dd, $J = 9.2$, 4.5 Hz, 1H), 3.63 (s, 3H), 2.06 (s, 3H), 1.93 (s, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 170.11, 169.85, 169.59, 149.21, 148.80, 137.08, 137.01, 128.87, 128.56, 128.54, 127.91, 127.89, 127.35, 127.28, 119.72, 114.44, 113.30, 74.17, 71.46, 71.07, 56.31, 52.70, 23.04, 20.89. ESI-HRMS calculated for $\text{C}_{28}\text{H}_{30}\text{NO}_7^+$ ($[\text{M}+\text{H}]^+$): 492.2014, found 492.2012.

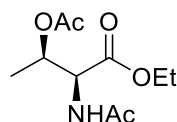
(2*S*,3*R*)-ethyl-2-acetamido-3-acetoxy-3-cyclopropylpropanoate (2n)



White solid; 12.5 mg; 97% yield; >99% ee; $[\alpha]_D^{20} = -61.80$ ($c = 0.23$, CHCl_3); The enantiomeric excess was determined by GC on Gamma-Dex 225 column; flow rate = 2.0 mL/min; 160 °C isothermal; $t_R = 33.4$ min (major), 33.7 min (minor). ^1H NMR (400 MHz, CDCl_3) δ 6.22 (d, $J = 9.1$ Hz, 1H), 4.97 (dd, $J = 9.4$ Hz, 3.0 Hz, 1H),

4.65 (dd, $J = 9.5, 3.0$ Hz, 1H), 4.16 (q, $J = 7.2$ Hz, 2H), 2.11 (s, 3H), 2.06 (s, 3H), 1.25 (t, $J = 7.1$ Hz, 3H), 1.05-1.11 (m, 1H), 0.39-0.66 (m, 4H); ^{13}C NMR (101 MHz, CDCl_3) δ 170.28, 169.97, 169.95, 78.02, 61.91, 55.28, 23.26, 20.91, 14.08, 12.79, 4.26, 3.47. ESI-HRMS calculated for $\text{C}_{12}\text{H}_{20}\text{NO}_5^+$ ($[\text{M}+\text{H}]^+$): 258.1336, found 258.1335.

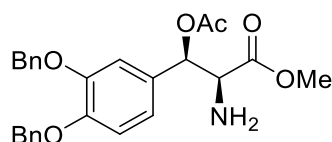
ethyl *N,O*-diacetyl-L-threoninate (**2o**)



Yellow oil; 11.3 mg; 98% yield; >99% ee; $[\alpha]_{\text{D}}^{20} = -62.33$ ($c = 0.65$, CHCl_3); The enantiomeric excess was determined by HPLC on Chiralpak OJ-H column, hexane: isopropanol = 95:5; flow rate = 1.0 mL/min; UV detection at 210 nm; $t_{\text{R}} = 14.1$ min (major), 16.5 min (minor). ^1H NMR (400 MHz, CDCl_3) δ 6.17 (d, $J = 8.9$ Hz, 1H), 5.45 (m, 1H), 4.79 (dd, $J = 9.4$ Hz, 2.6 Hz, 1H), 4.19 (q, $J = 7.1$ Hz, 2H), 2.11 (s, 3H), 2.01 (s, 3H), 1.25-1.28 (m, 6H); ^{13}C NMR (101 MHz, CDCl_3) δ = 170.42, 169.95, 169.71, 70.56, 61.91, 55.40, 23.18, 20.92, 17.07, 14.07. MS calculated for $\text{C}_{10}\text{H}_{18}\text{NO}_5^+$ ($[\text{M}+\text{H}]^+$): 232.1, found 232.1.

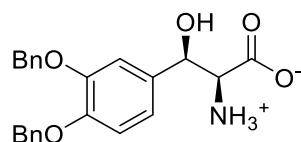
4. Procedure for the Synthesis of Compound 4

(2*S*,3*R*)-methyl -3-acetoxy-2-amino-3-(3,4-bis(benzyloxy)phenyl)propanoate (**3**)



3H); ^{13}C NMR (101 MHz, CDCl_3) δ 171.45, 166.93, 149.27, 149.21, 137.02, 136.97, 132.44, 128.56, 127.95, 127.91, 127.42, 127.24, 119.08, 114.84, 112.50, 83.03, 76.19, 71.42, 71.19, 52.80, 14.16. ESI-HRMS calculated for $\text{C}_{26}\text{H}_{27}\text{NO}_6\text{Na}^+$ ($[\text{M}+\text{Na}]^+$): 472.1731, found 472.1715.

(2*S*,3*R*)-2-ammonio-3-(3,4-bis(benzyloxy)phenyl)-3-hydroxypropanoate (4)



To a solution of **3** (37 mg, 0.08 mmol) in MeOH (2.0 mL) was added dropwise 2 N NaOH (2.0 mL) at 0 °C. The heterogeneous milky reaction mixture was stirred overnight at 40 °C. Then the mixture was cooled to 0 °C, and neutralized to pH 6-7 by slowly addition of 1 N HCl. The resulting white precipitate was filtered and washed with cold water and ether to give 30 mg (92%) of **4**. $[\alpha]_{\text{D}}^{25} = +2.00$ ($c = 0.20$, DMSO). ^1H NMR (400 MHz, DMSO- d_6) δ 10.08 (d, $J = 10.2$ Hz, 1H), 7.46 (m, 4H), 7.38 (m, 6H), 7.08 (d, $J = 1.8$ Hz, 1H), 6.95 (d, $J = 8.3$ Hz, 1H), 6.81 (dd, $J = 8.3, 1.8$ Hz, 1H), 6.02 (d, $J = 14.7$ Hz, 1H), 5.15 (s, 2H), 5.10 (s, 2H), 5.09 (s, 1H); ^{13}C NMR (101 MHz, DMSO) δ 167.59, 149.00, 147.28, 137.87, 137.84, 130.54, 128.94, 128.83, 128.19, 128.03, 127.93, 122.91, 118.91, 115.24, 111.43, 111.19, 70.56, 70.42, 23.18, 22.89. ESI-HRMS calculated for $\text{C}_{23}\text{H}_{24}\text{NO}_5^+$ ($[\text{M}+\text{H}]^+$): 394.1649, found 394.1637.

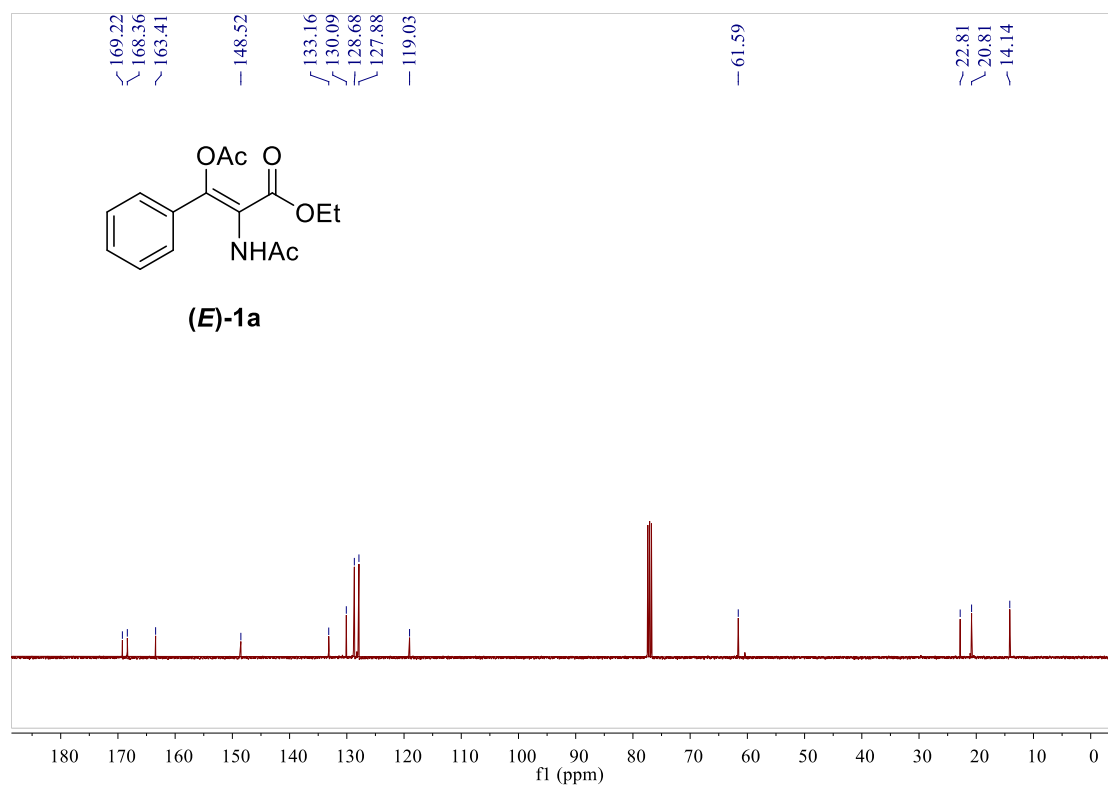
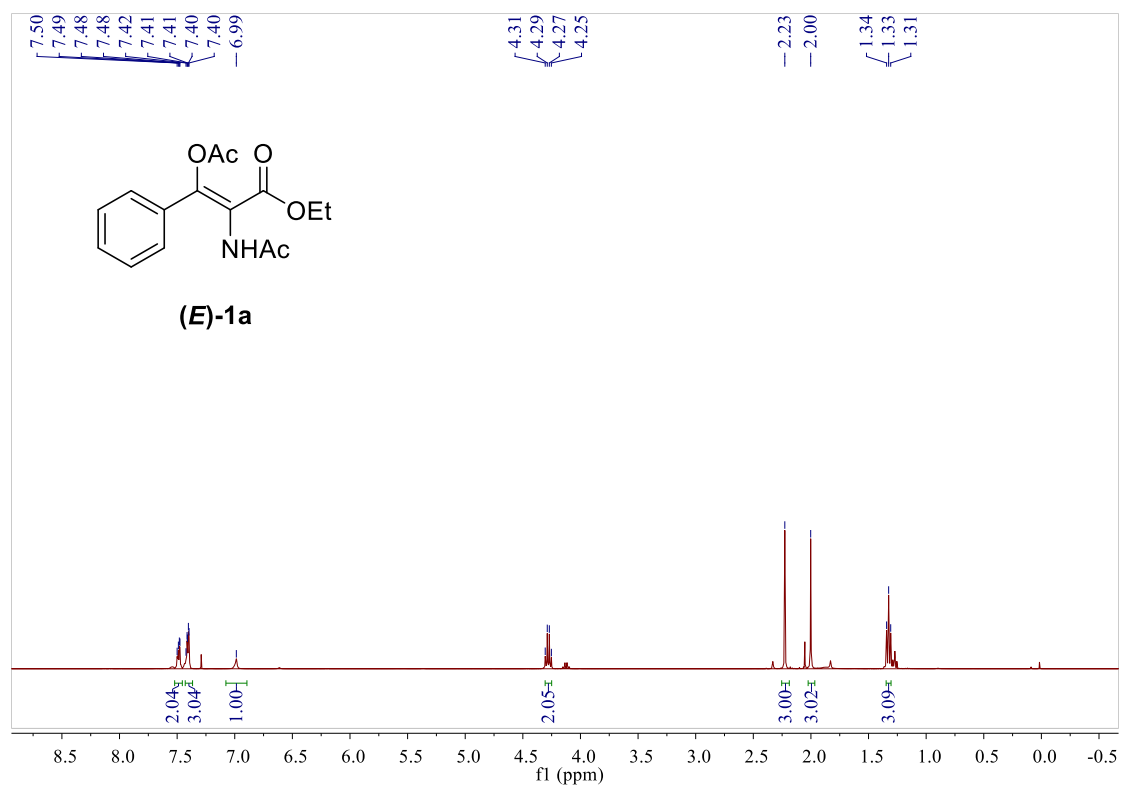
Compared the optical rotation of **2m** with the literature^[7], the absolute configuration of **2** is determined as (2*S*,3*R*).

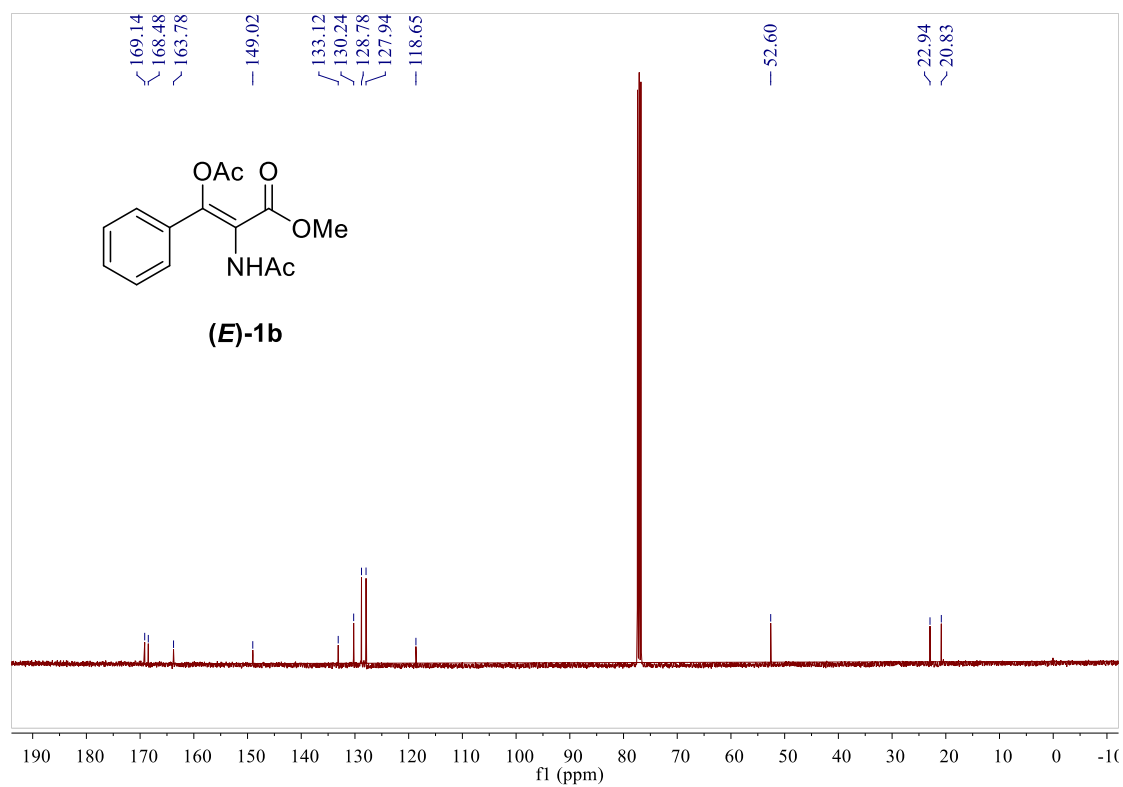
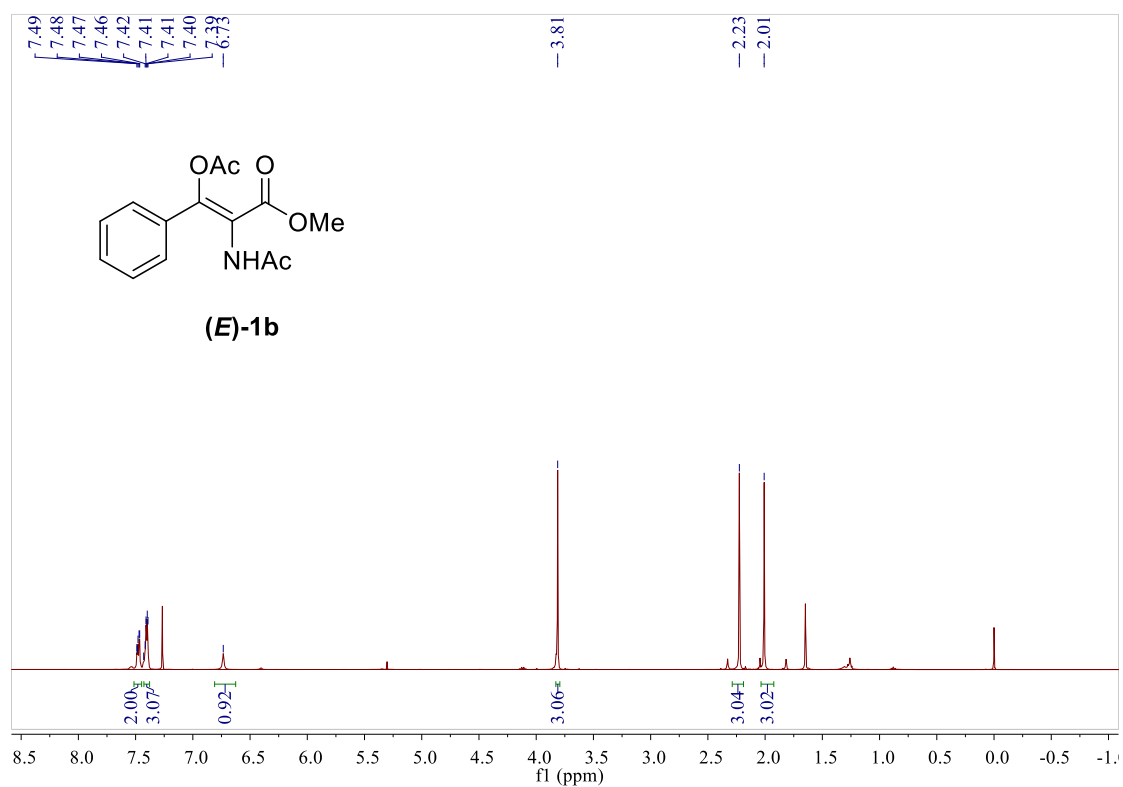
5. References

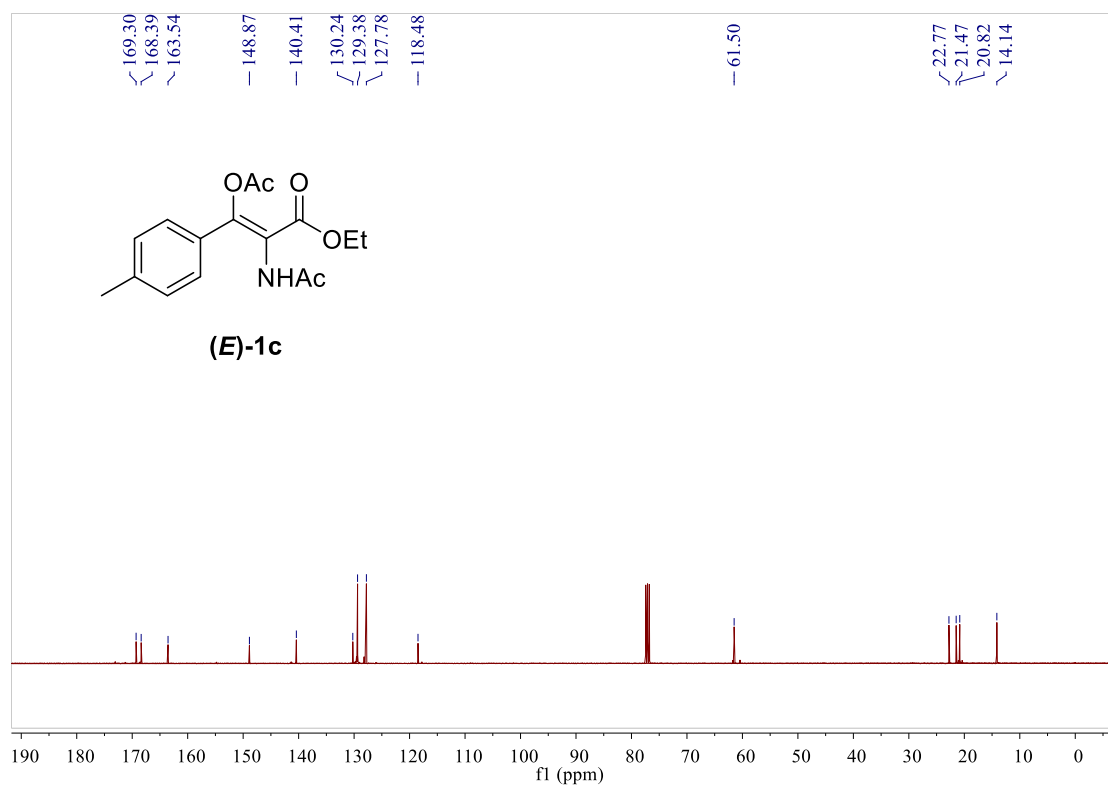
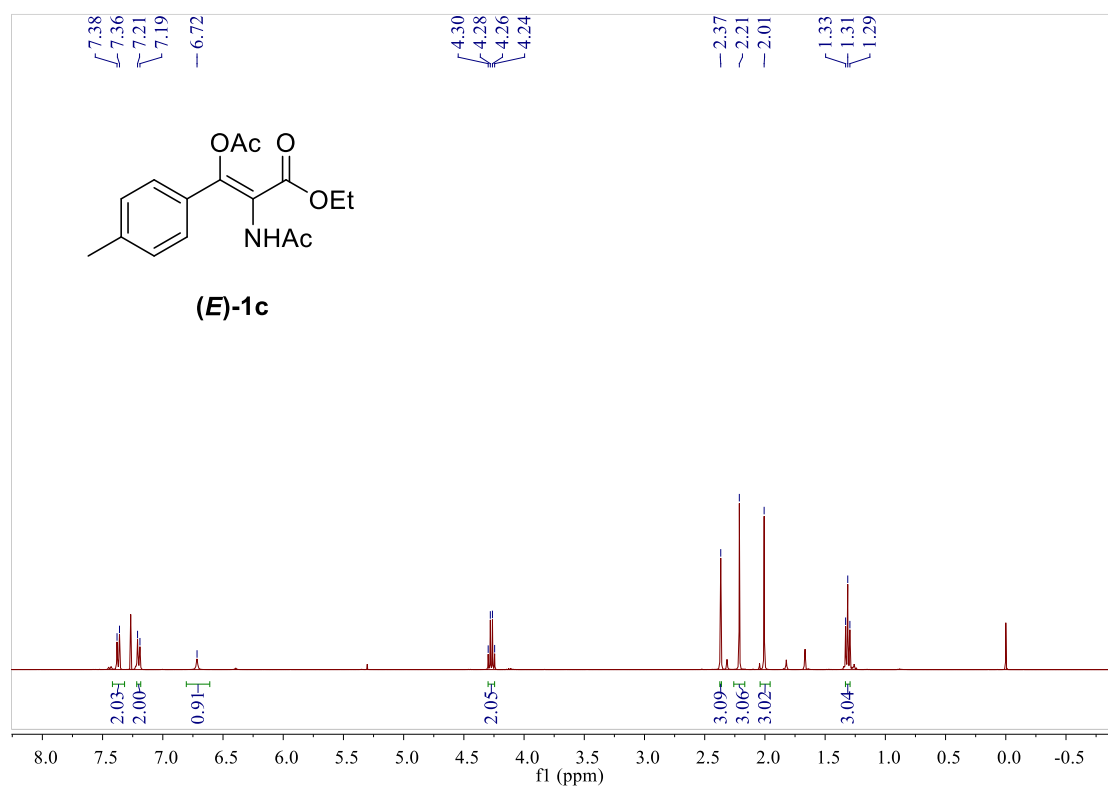
- [1] P. Borowiecki, M. Bretner, *Tetrahedron: Asymmetry* **2013**, 24, 925-936.
- [2] J. P. Genet, C. Pinel, S. Mallart, S. Juge, S. Thorimbert, J. A. Laffitte, *Tetrahedron-Asymmetry* **1991**, 2, 555-567.
- [3] H. Ryu, H. Kuriyama, H. Miyazato, S. Minakata, M. Komatsu, J. Y. Yoon, S. Kim, *Bull. Chem. Soc. Jpn.* **2004**, 77, 1407.
- [4] J. P. Burkhardt, J. R. Koehl, S. Mehdi, S. L. Durham, M. J. Janusz, E. W. Huber, et al. *J. Med. Chem.* **1995**, 38, 223-233.
- [5] T. Katoh, E. Itoh, T. Yoshino, S. Terashima, *Tetrahedron*, **1997**, 53, 10229-10238.
- [6] S. Hanessian, *Tetrahedron Lett.* **1967**, 1549-1552.

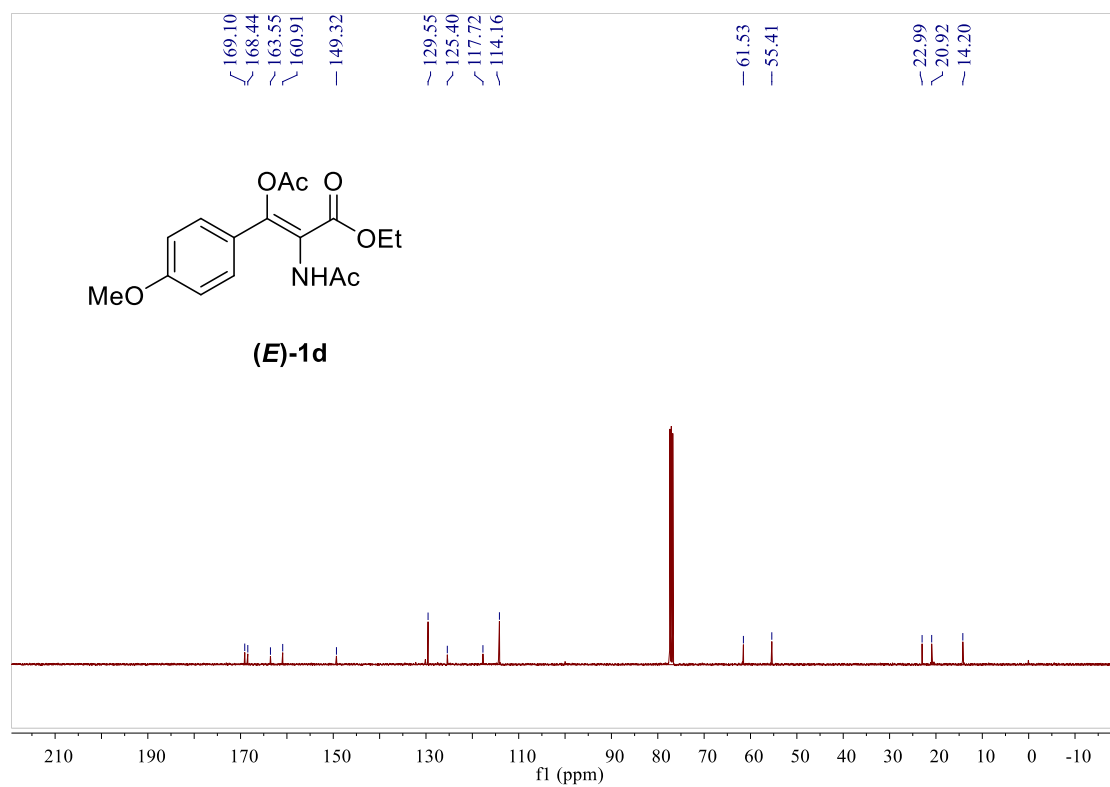
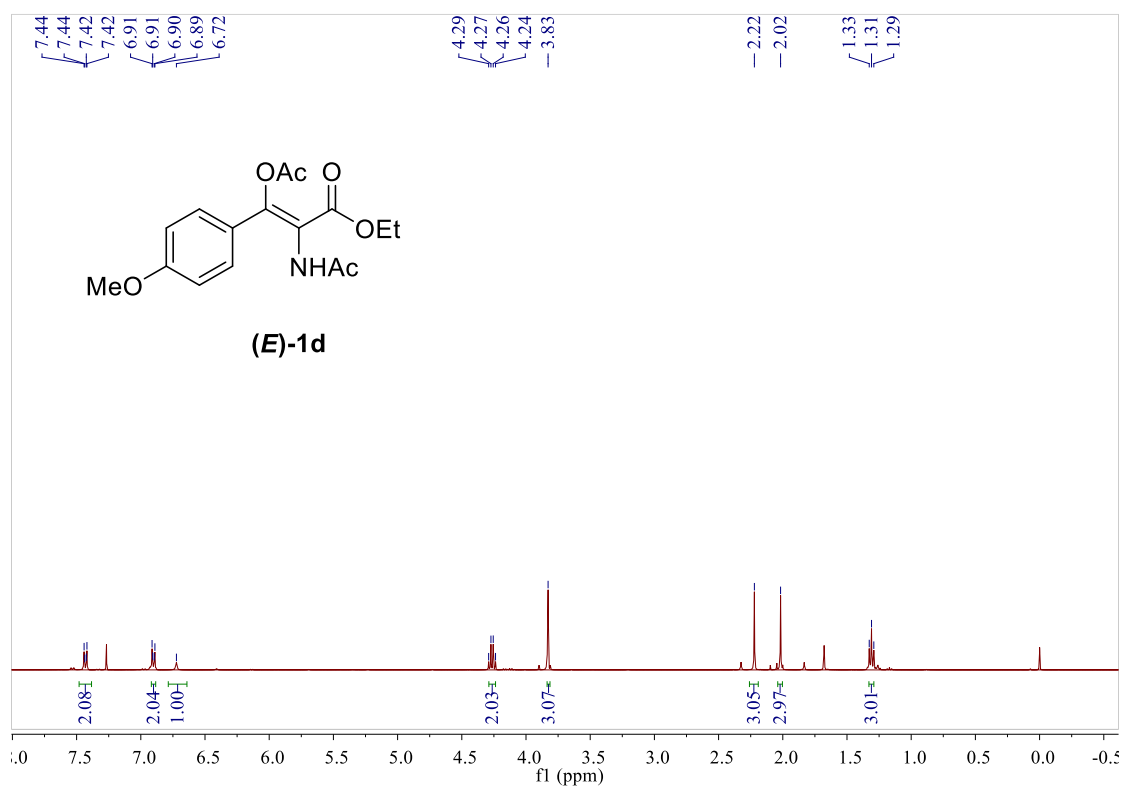
[7] B. Herbert, I. H. Kim and K. L. Kirk, *J. Org. Chem.* **2001**, 66, 4892-4897.

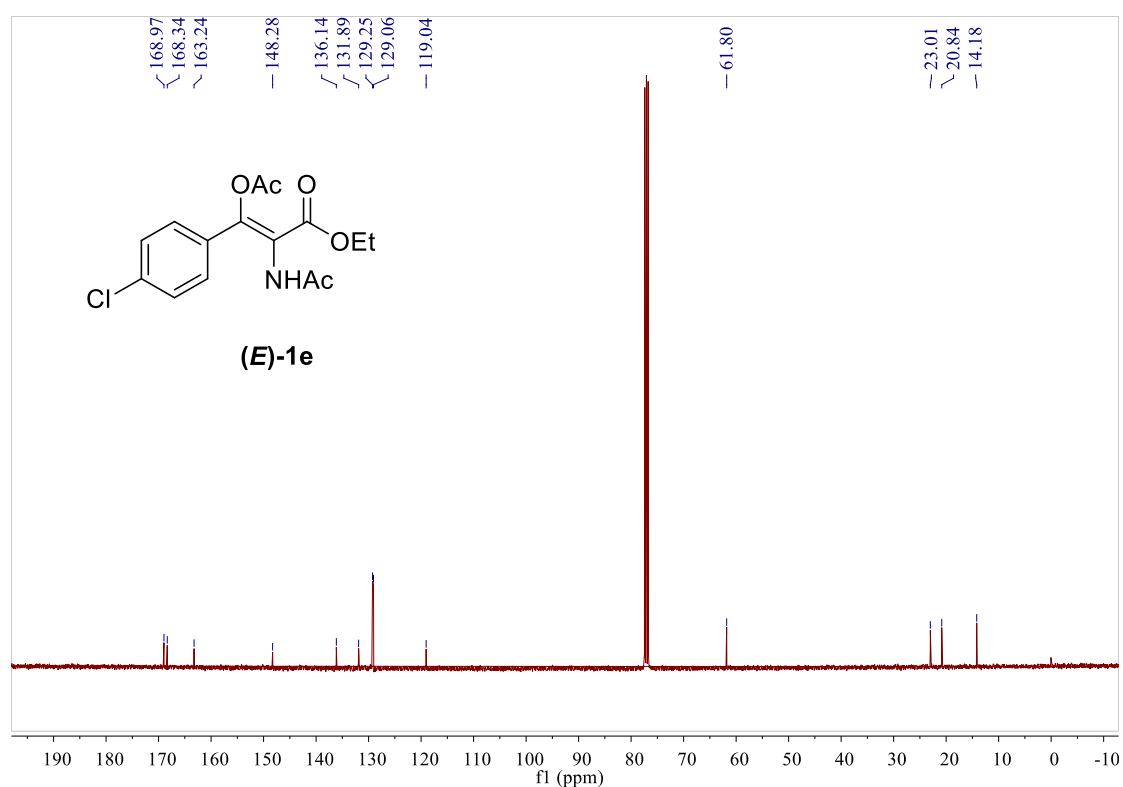
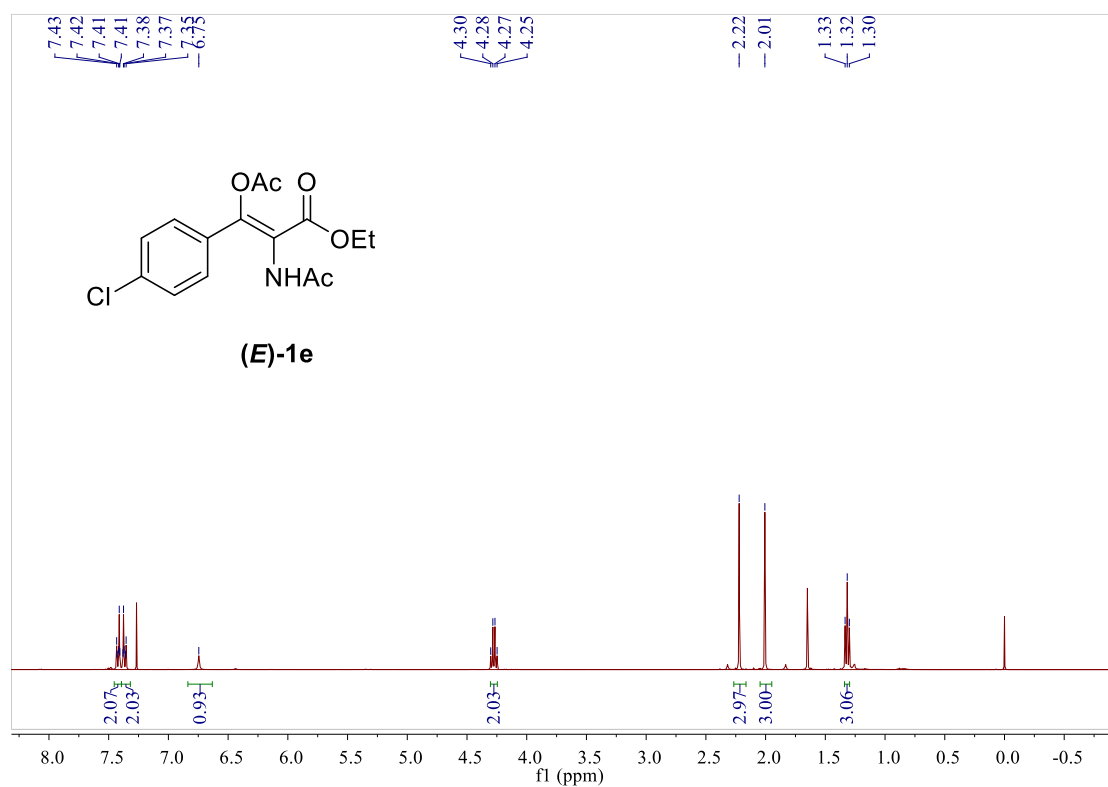
6. NMR spectra of (*E*)-1, 2 and 3, HPLC spectra or GC spectra of 2

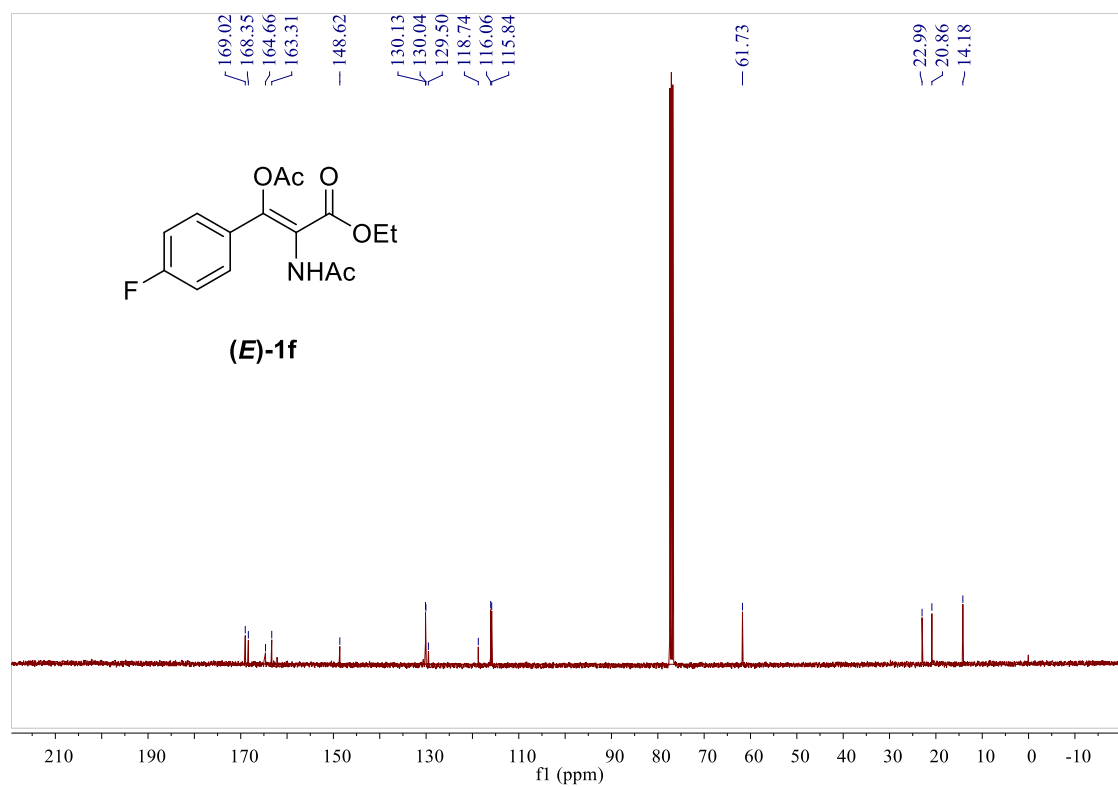
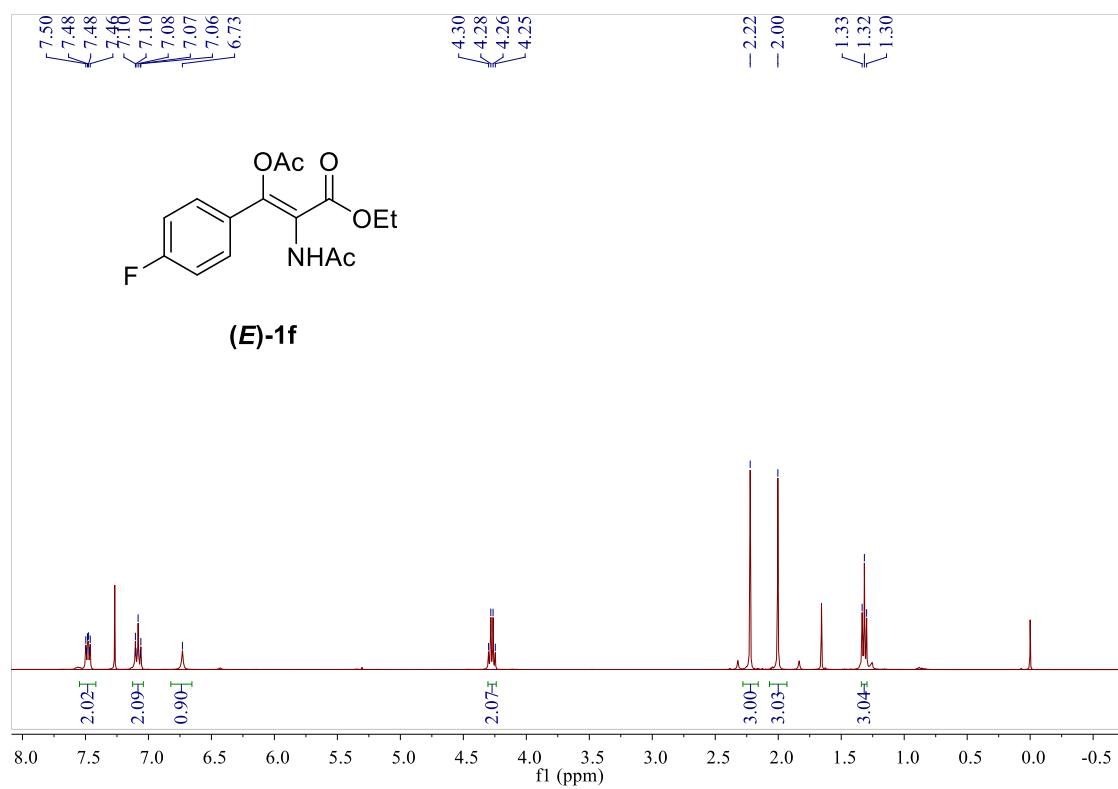


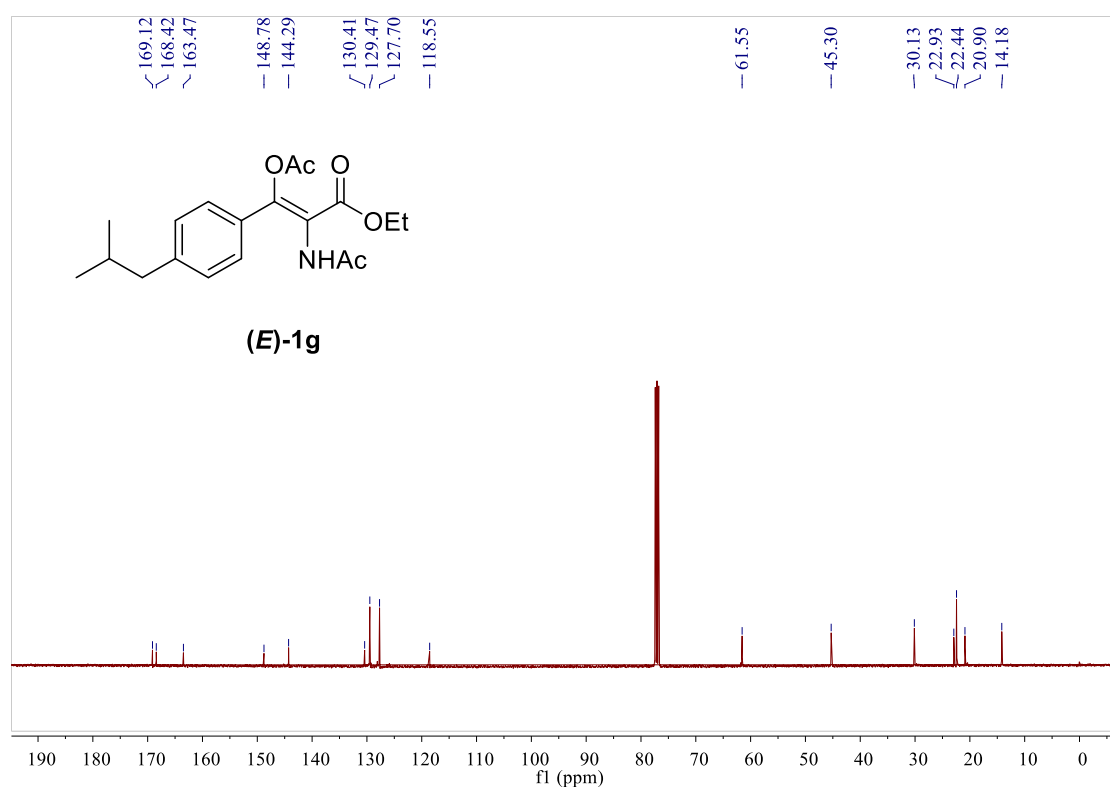
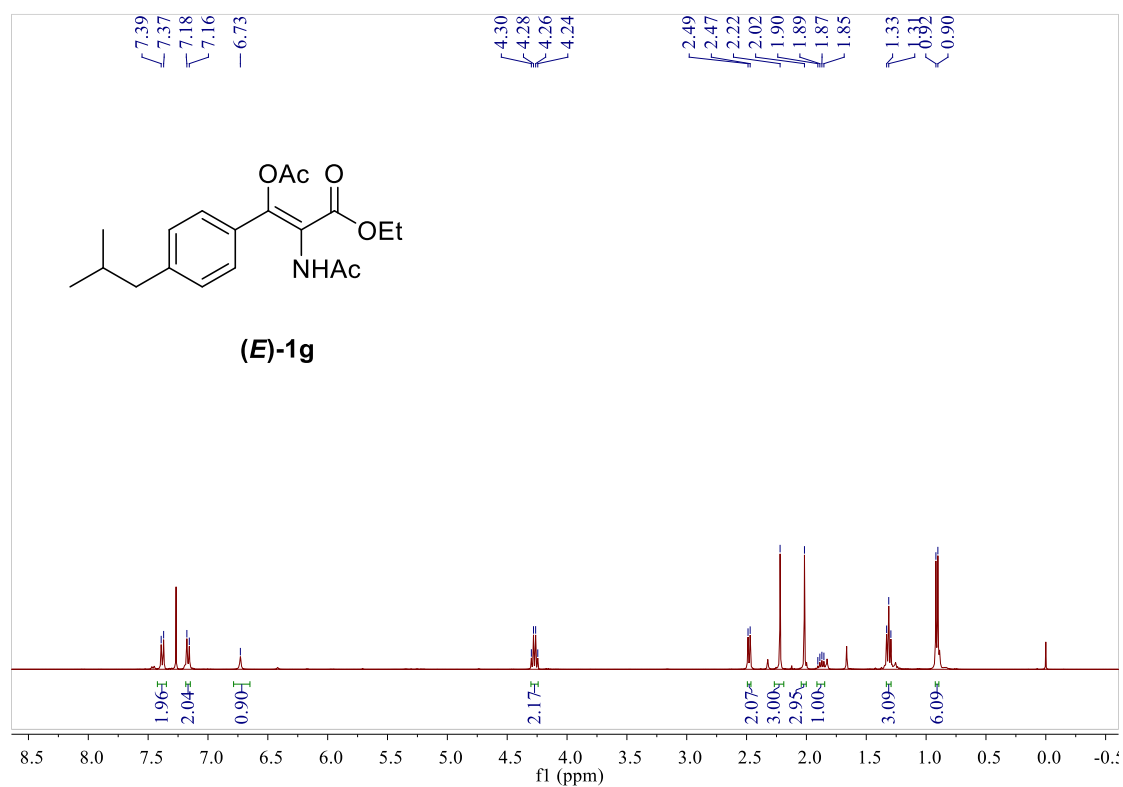


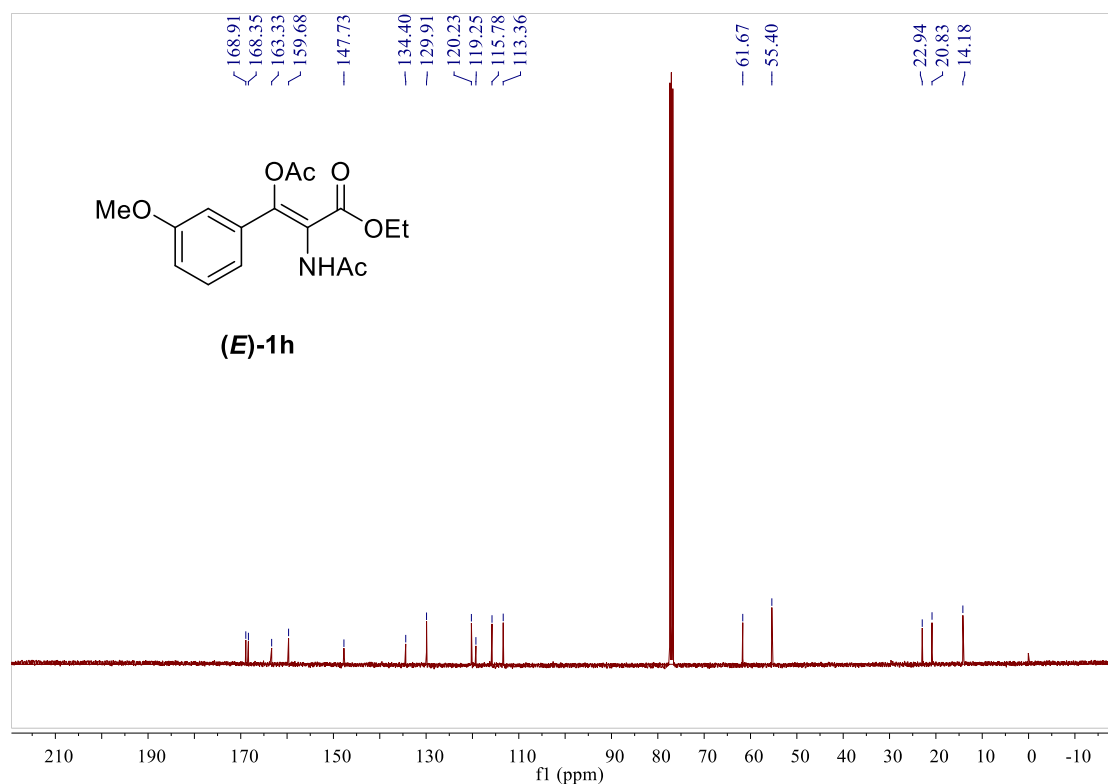
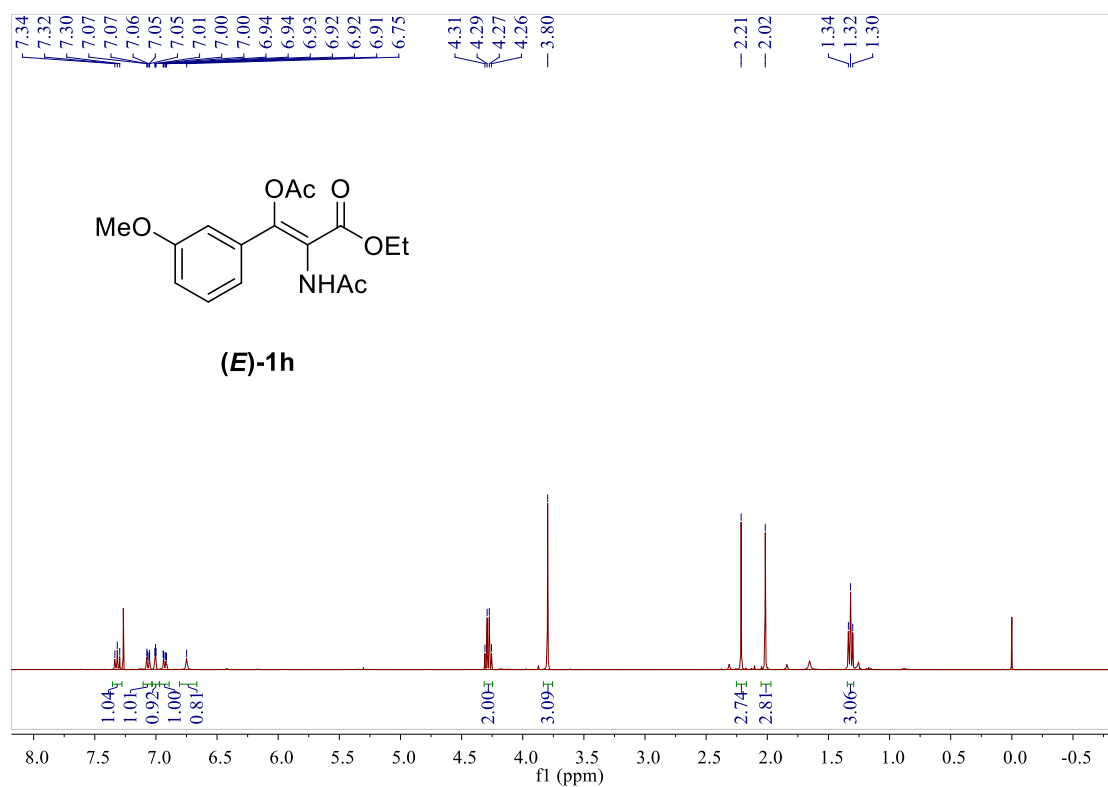


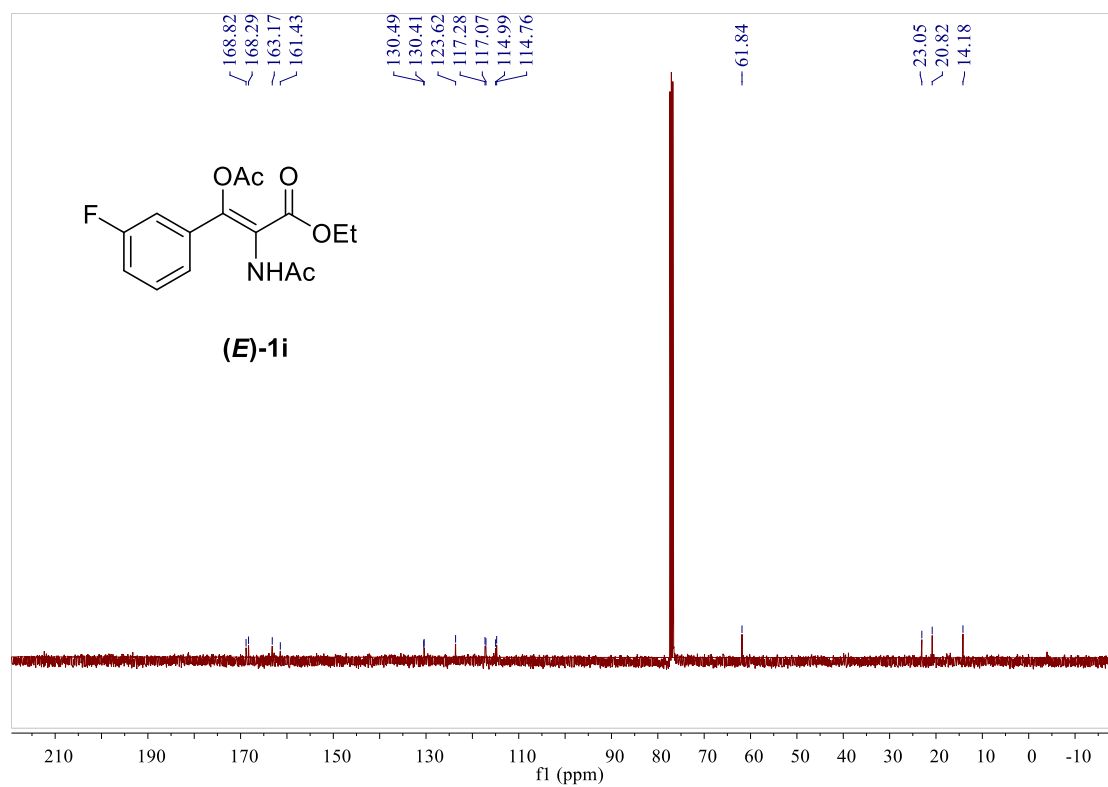
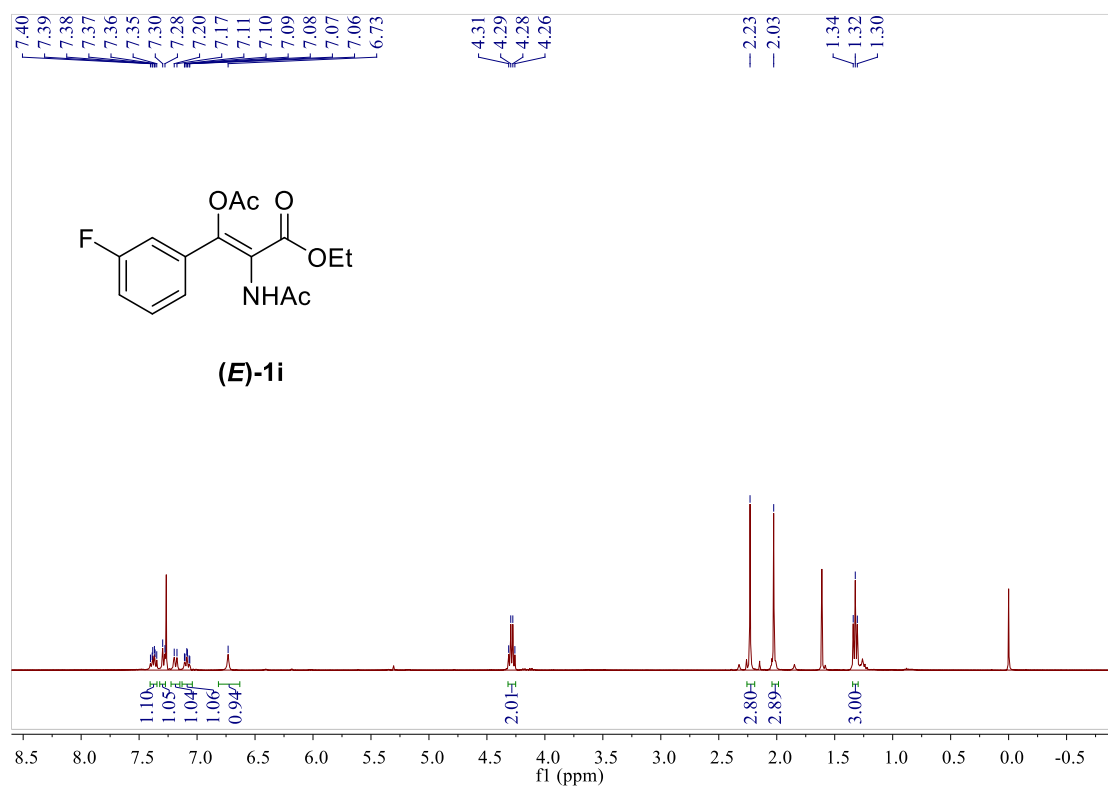


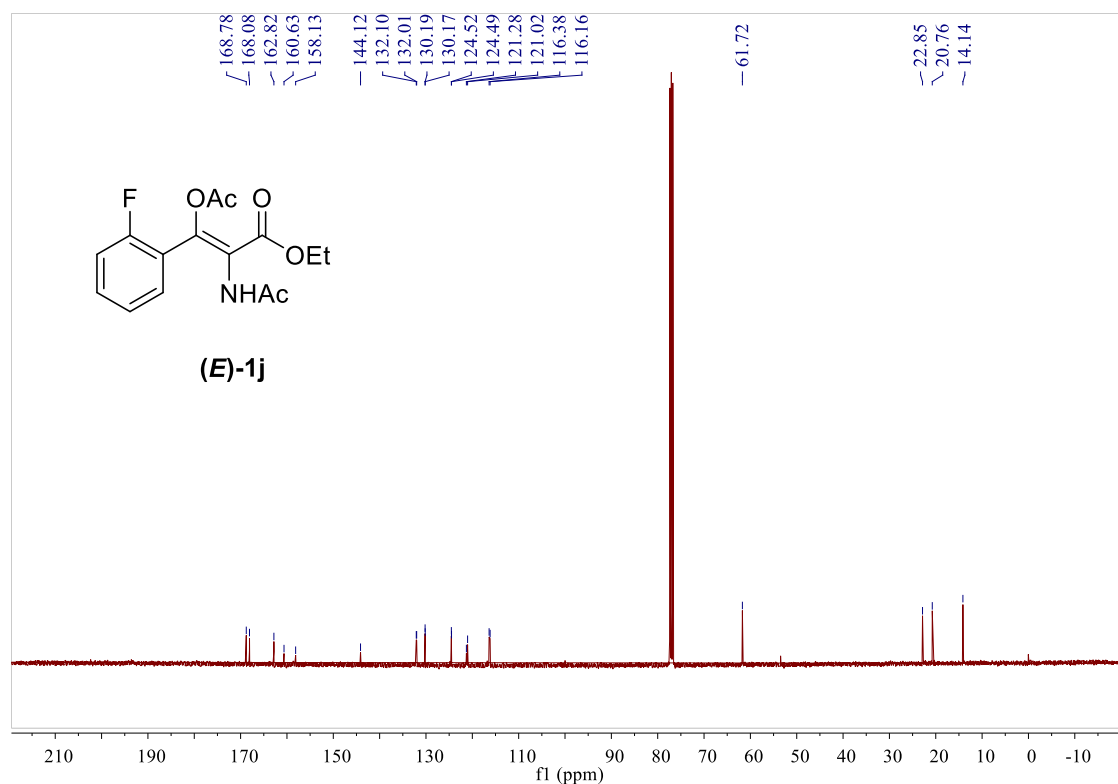
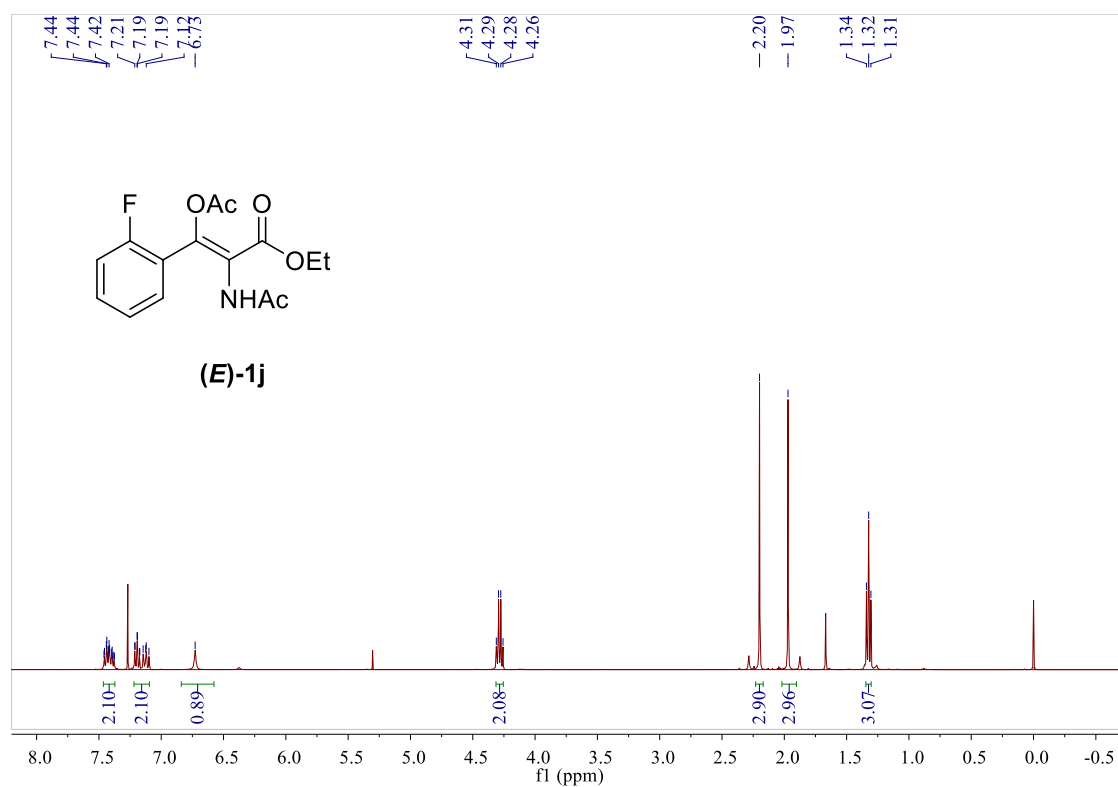


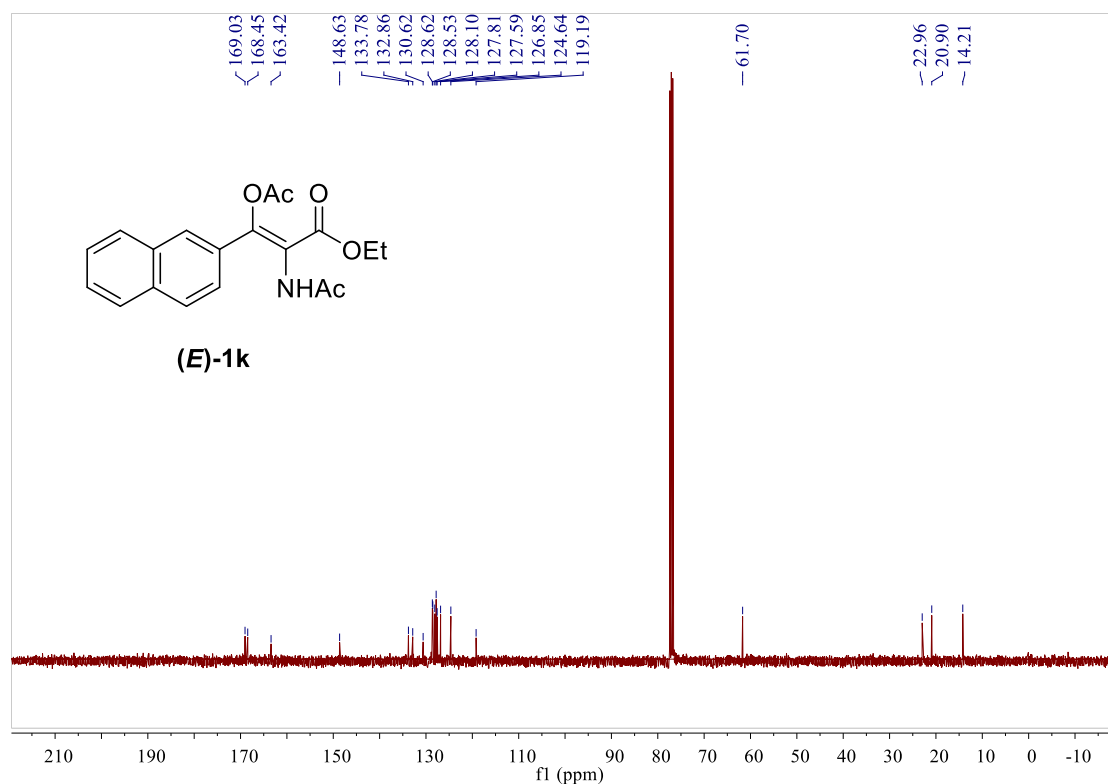
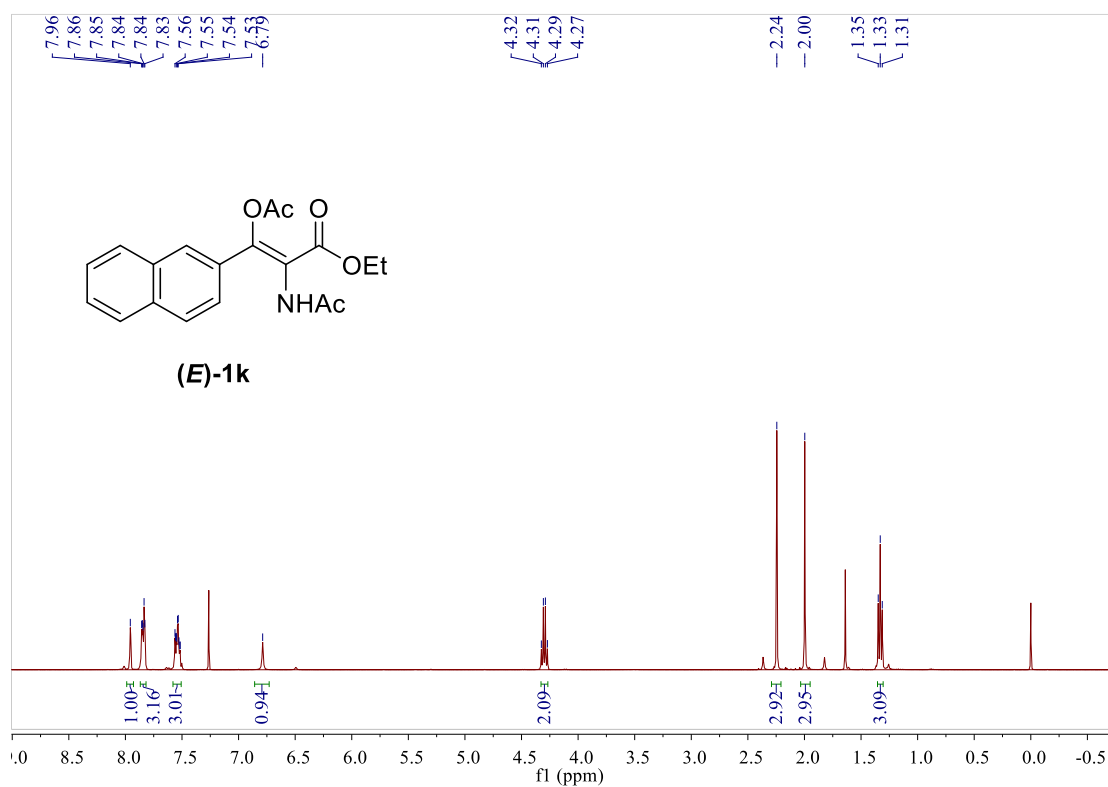


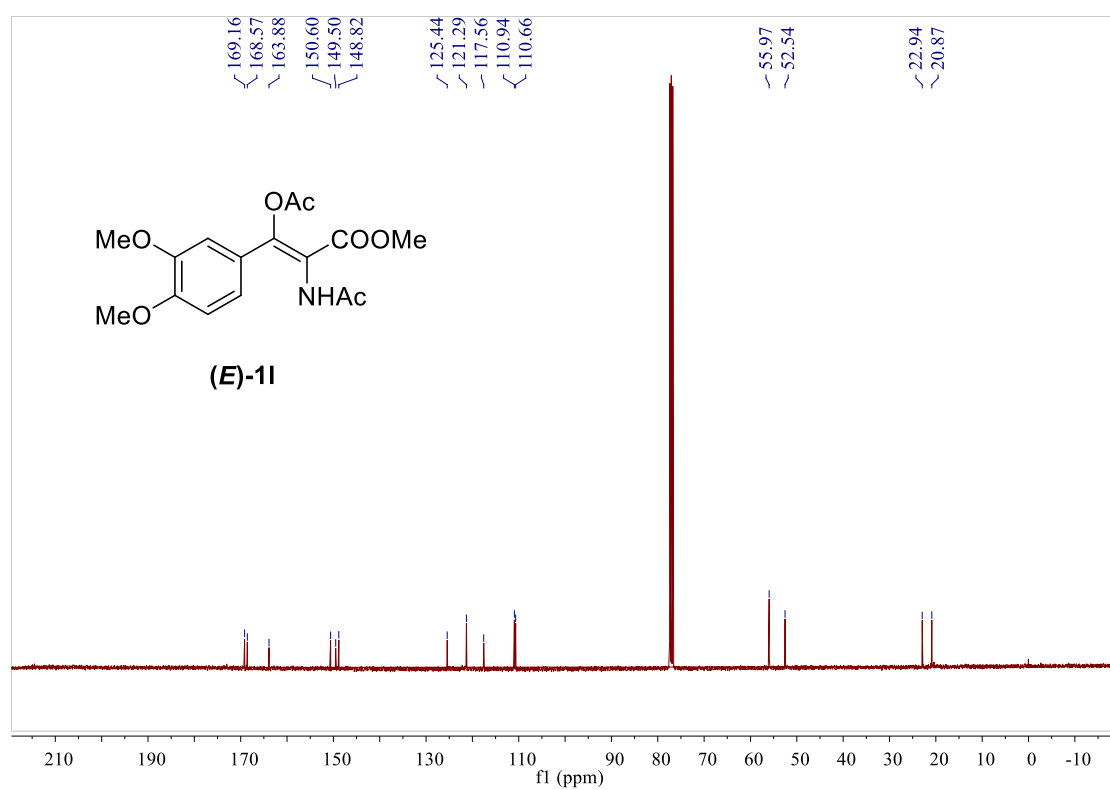
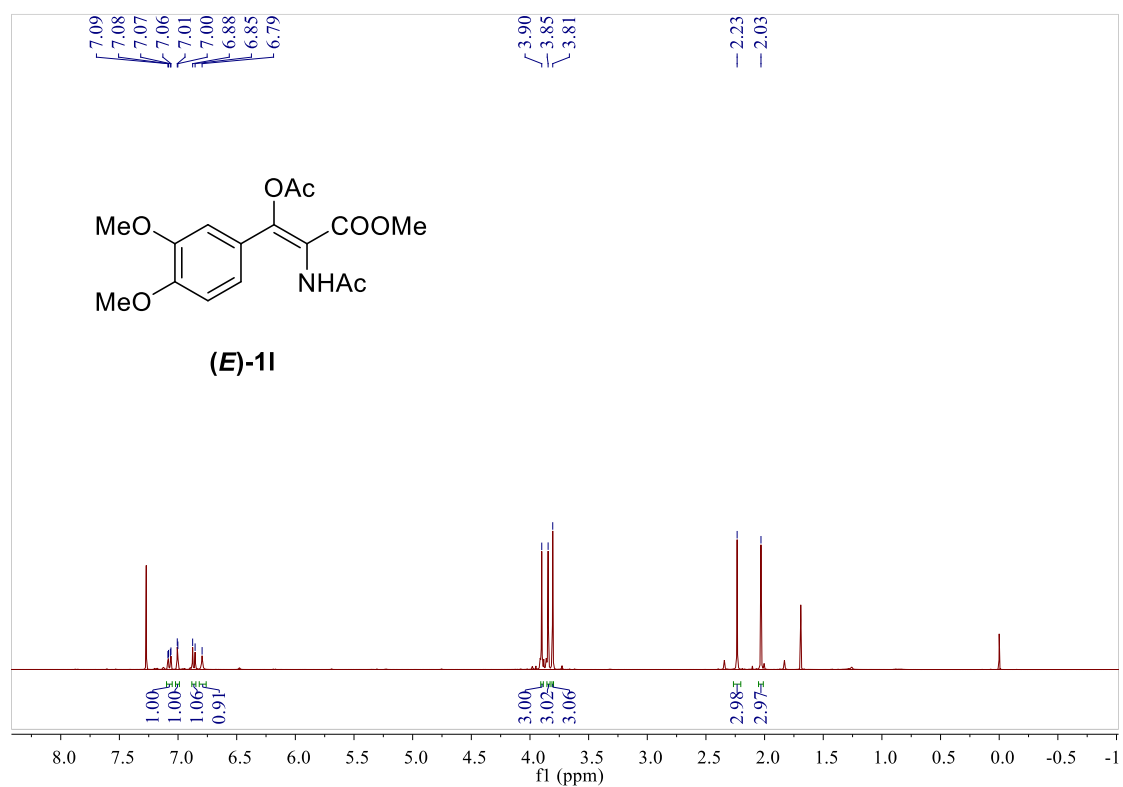


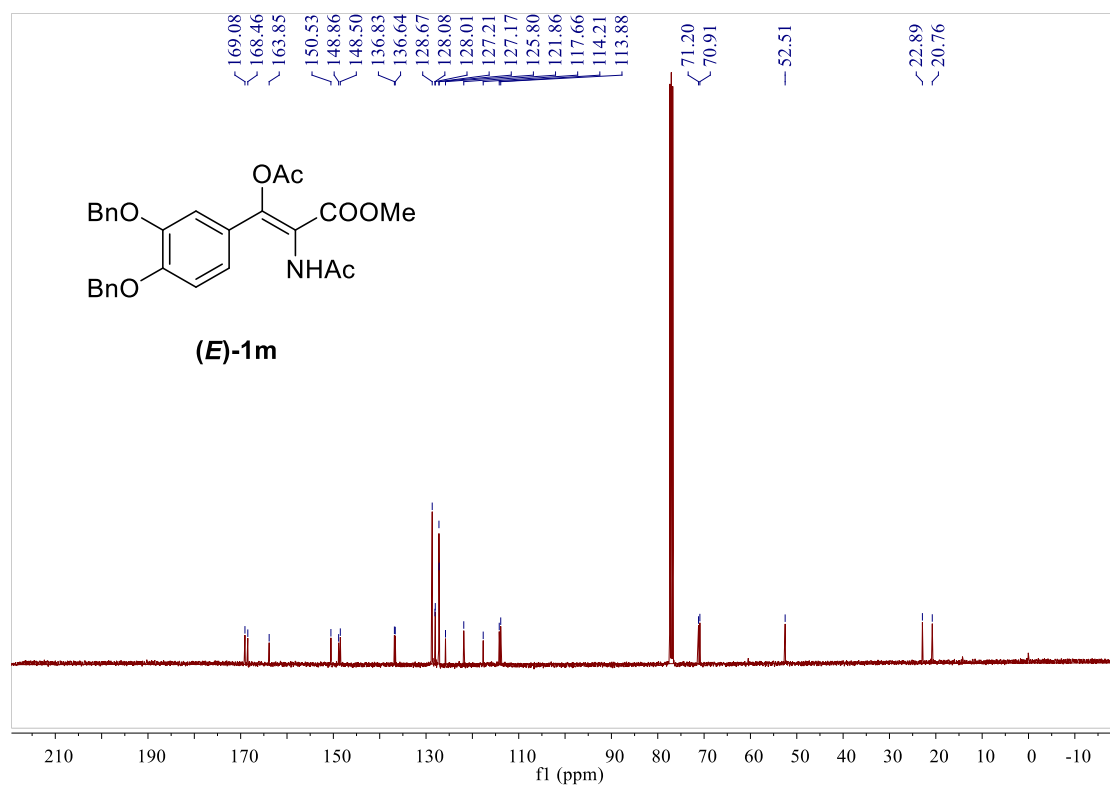
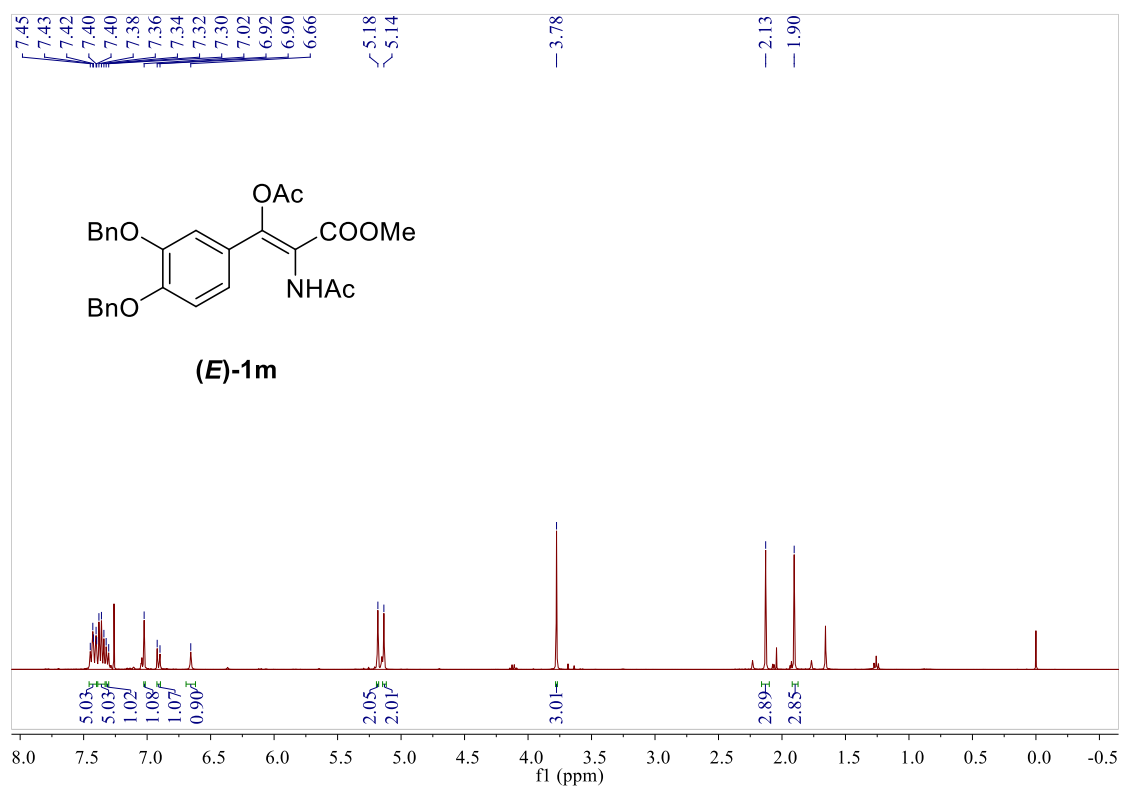


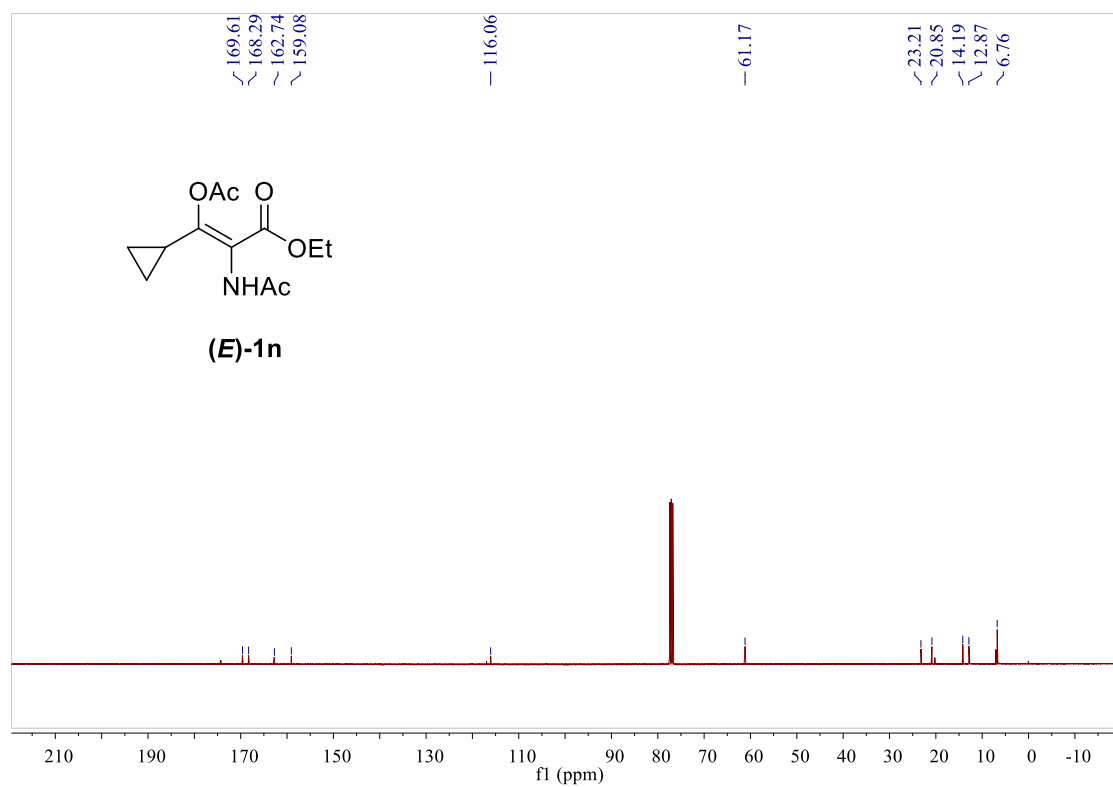
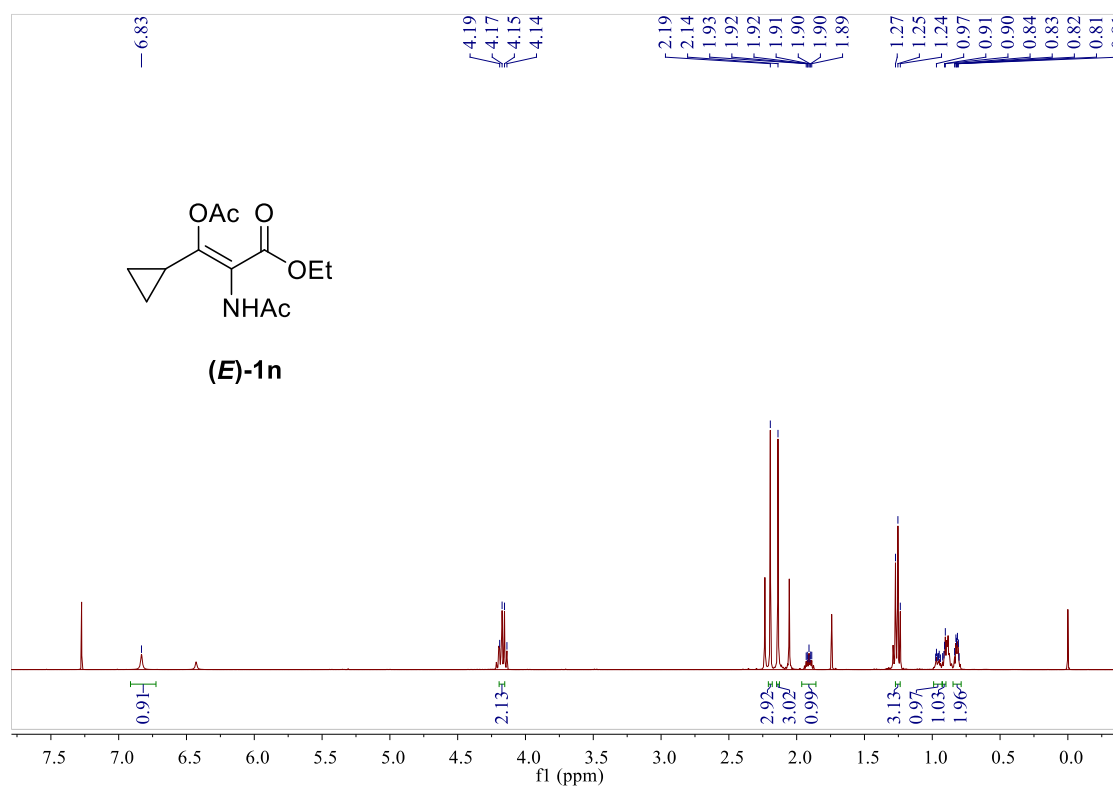


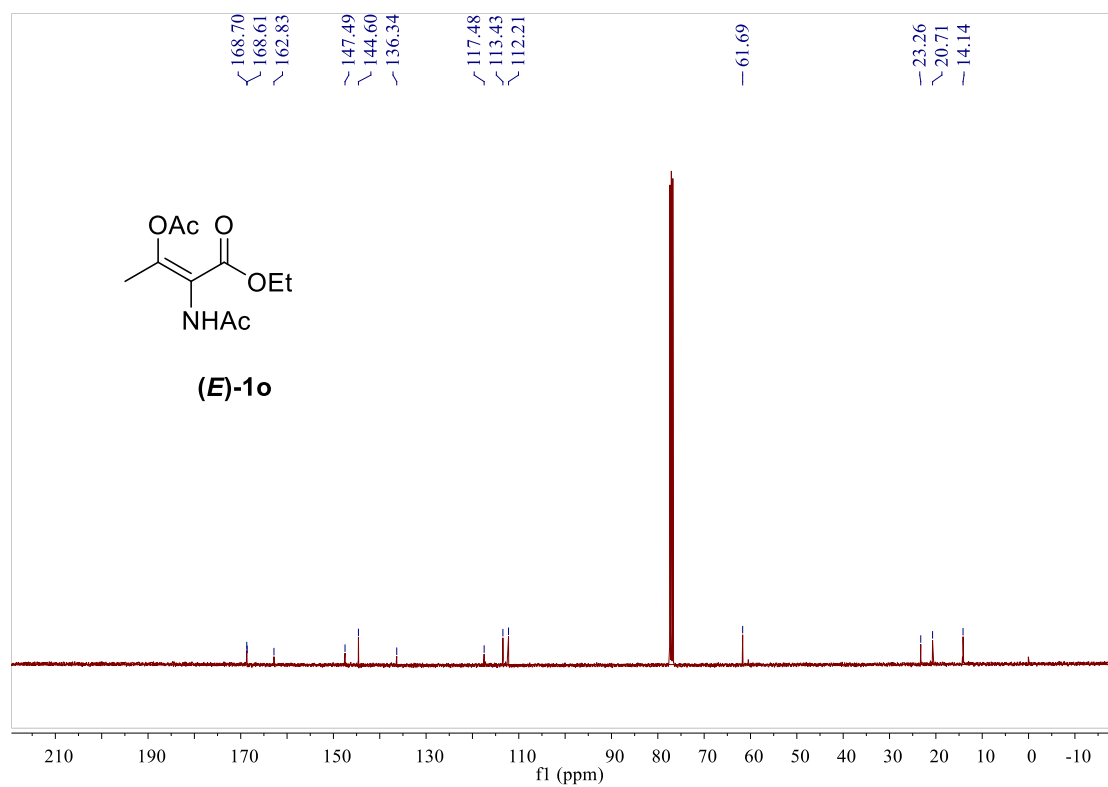
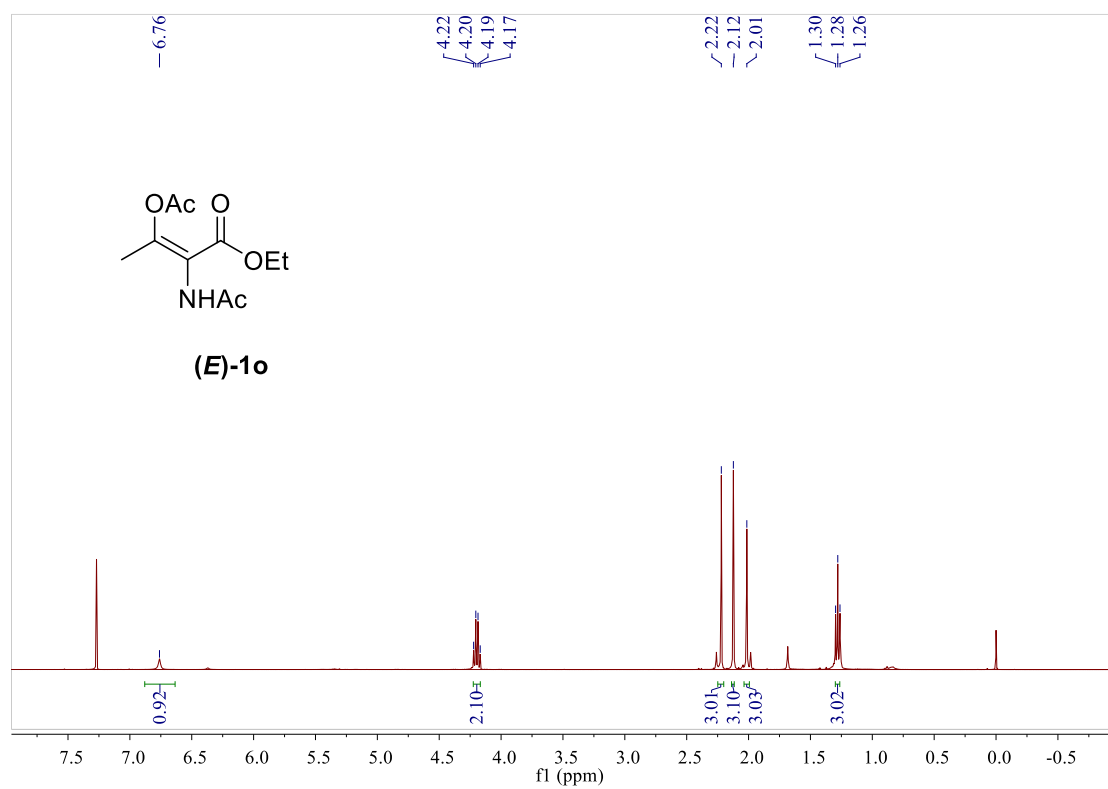


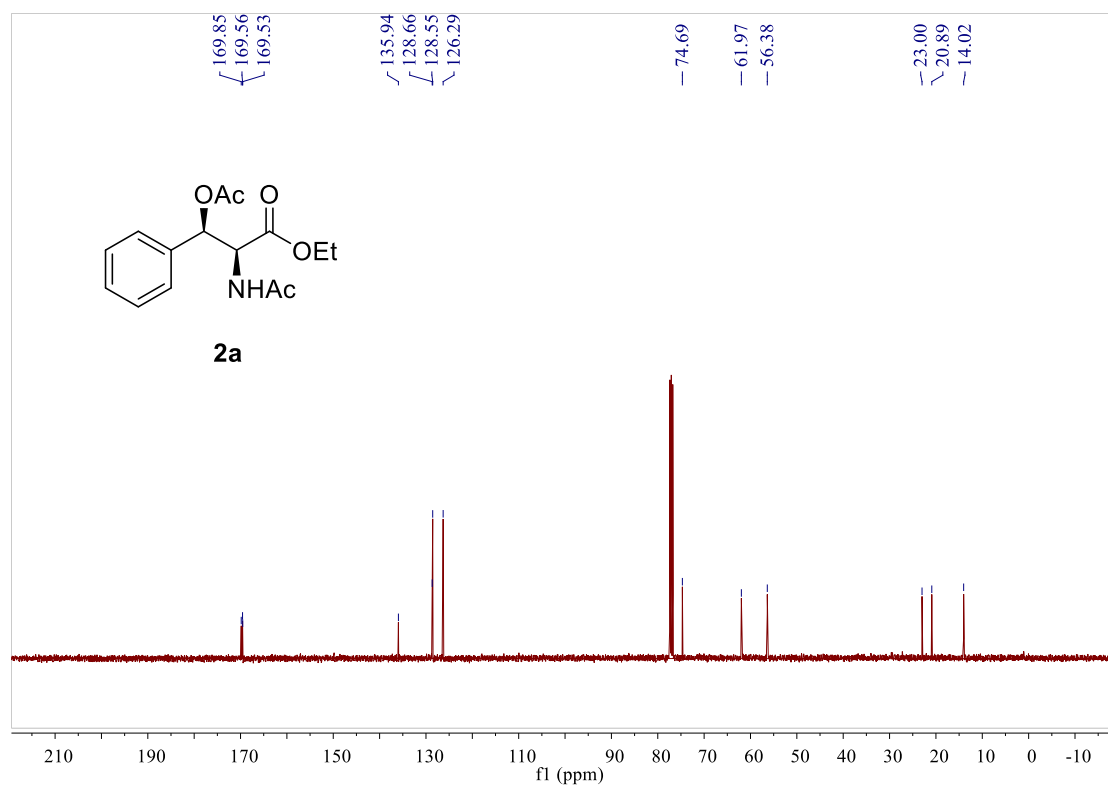
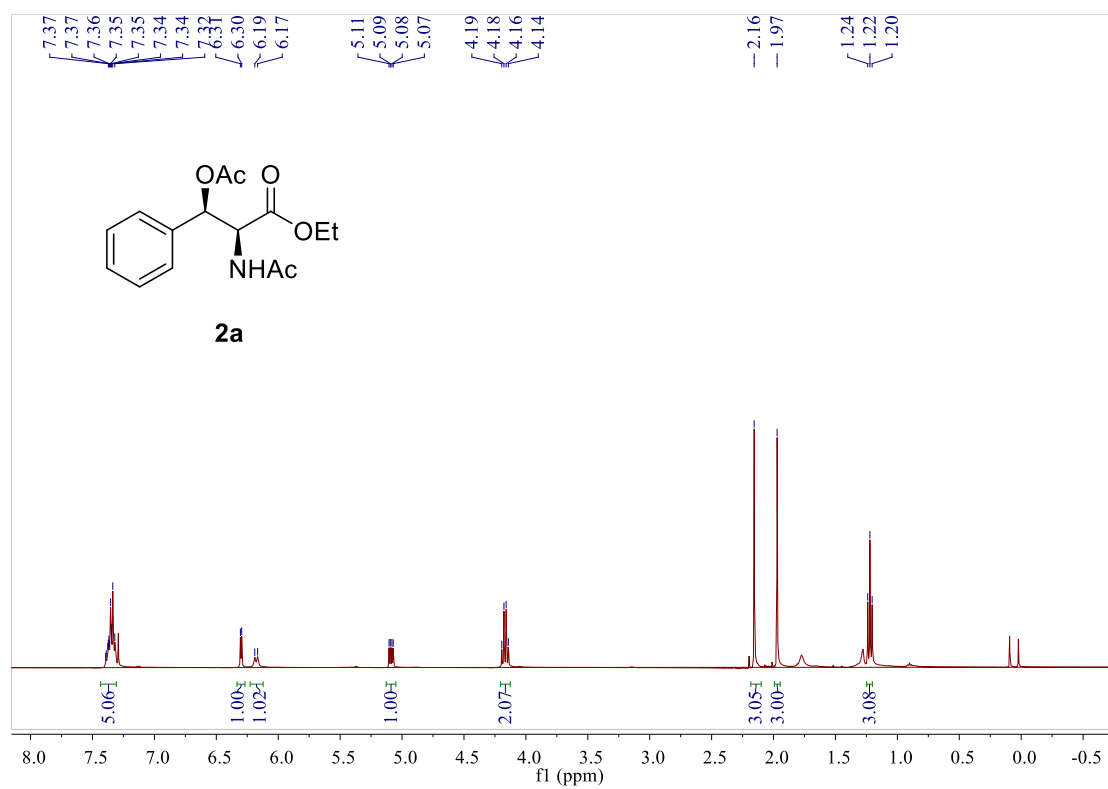


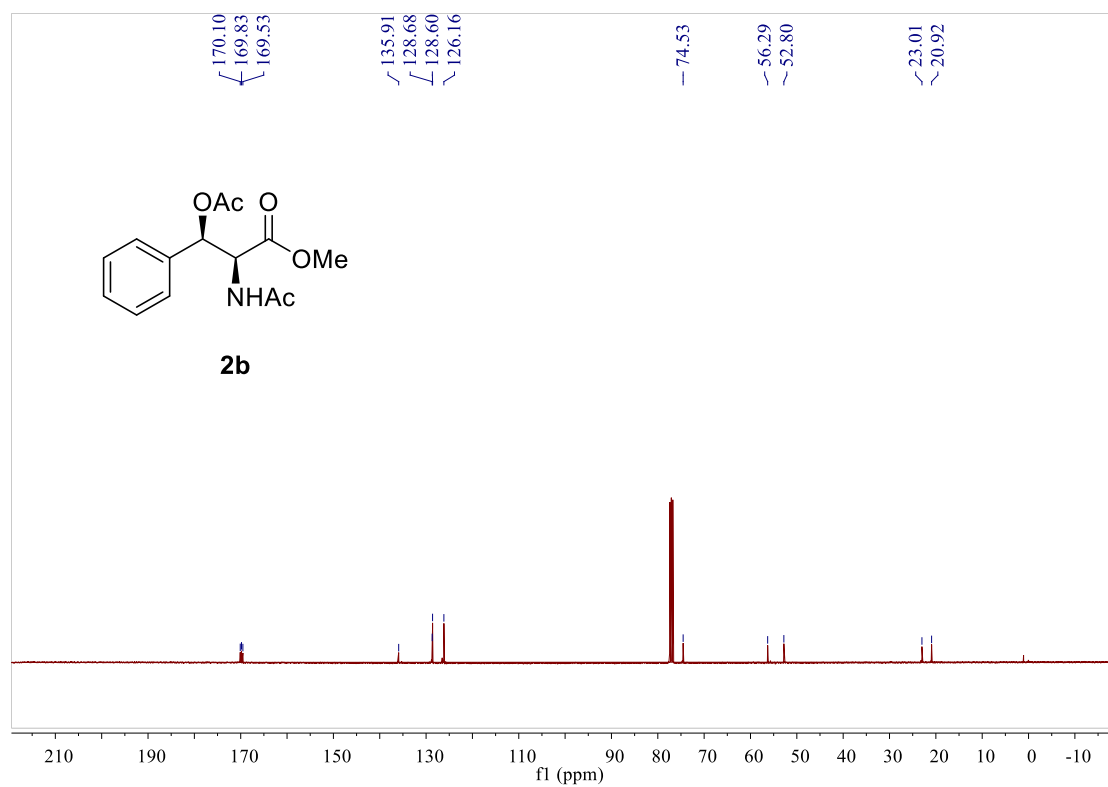
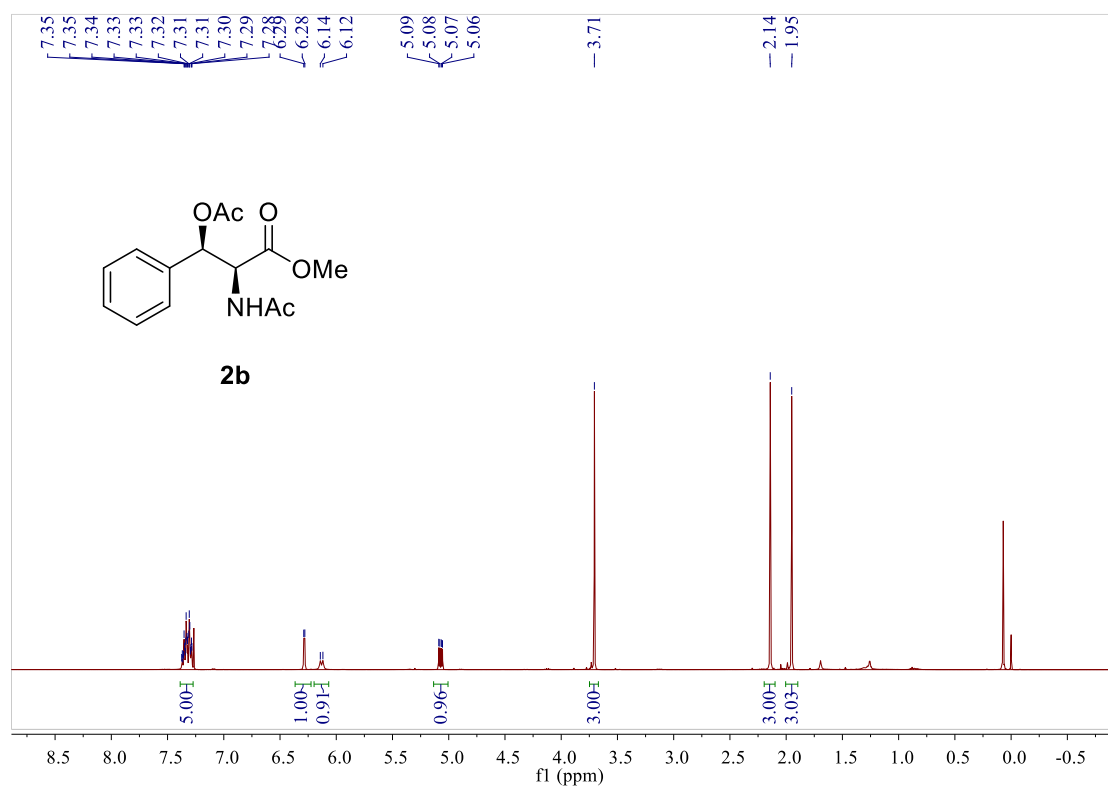


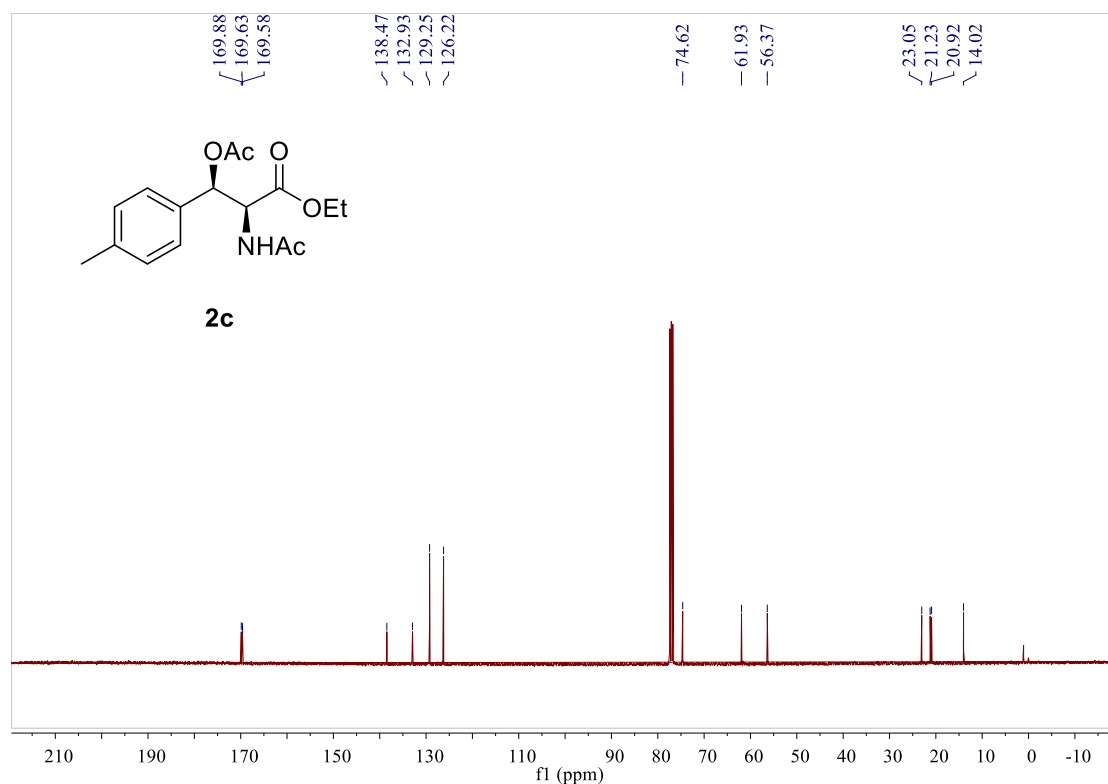
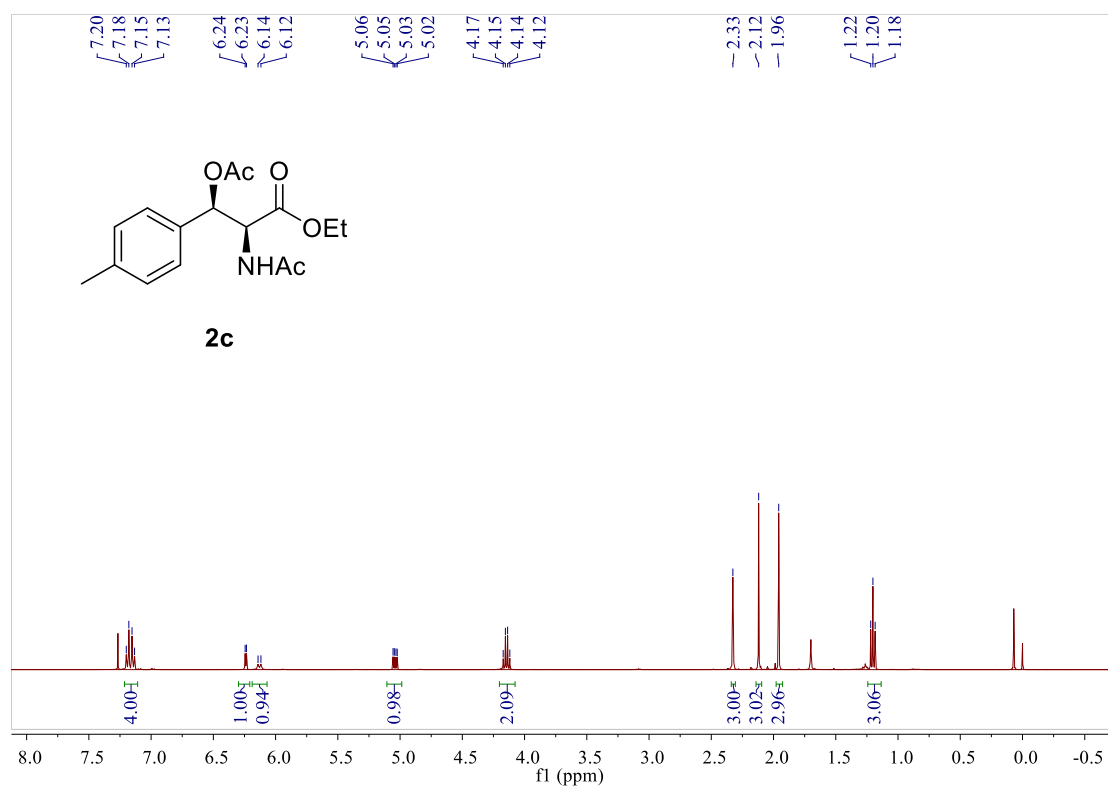


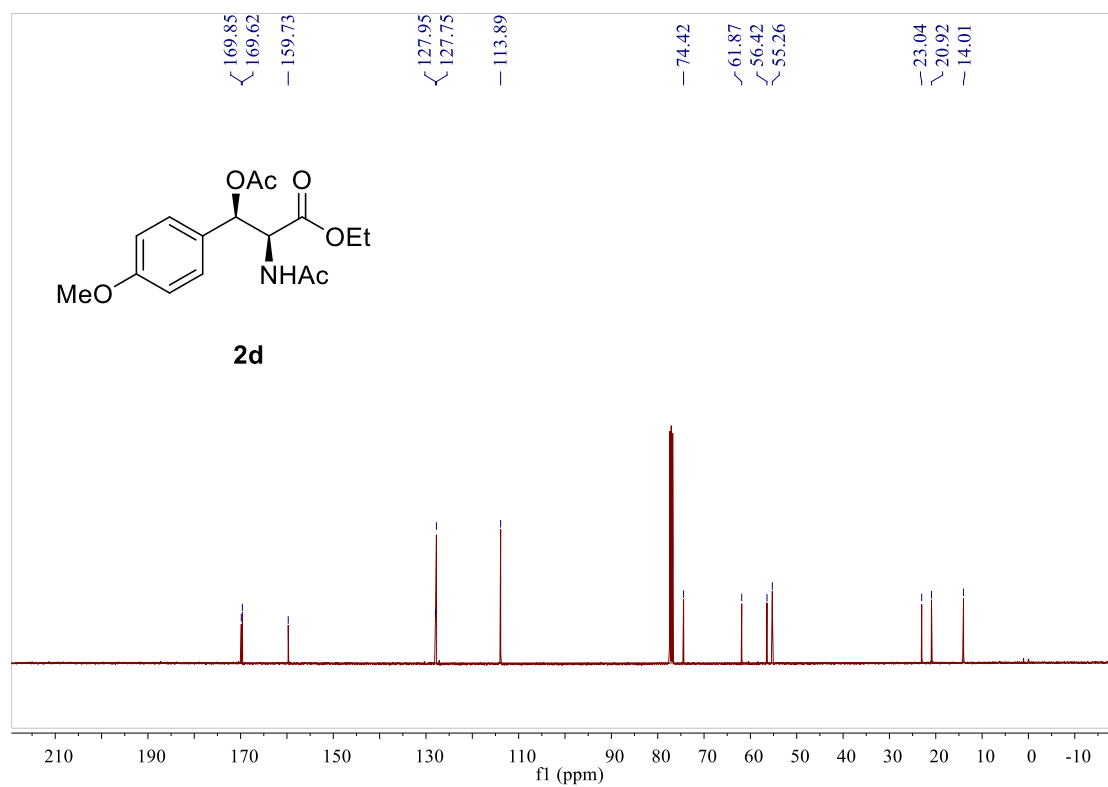
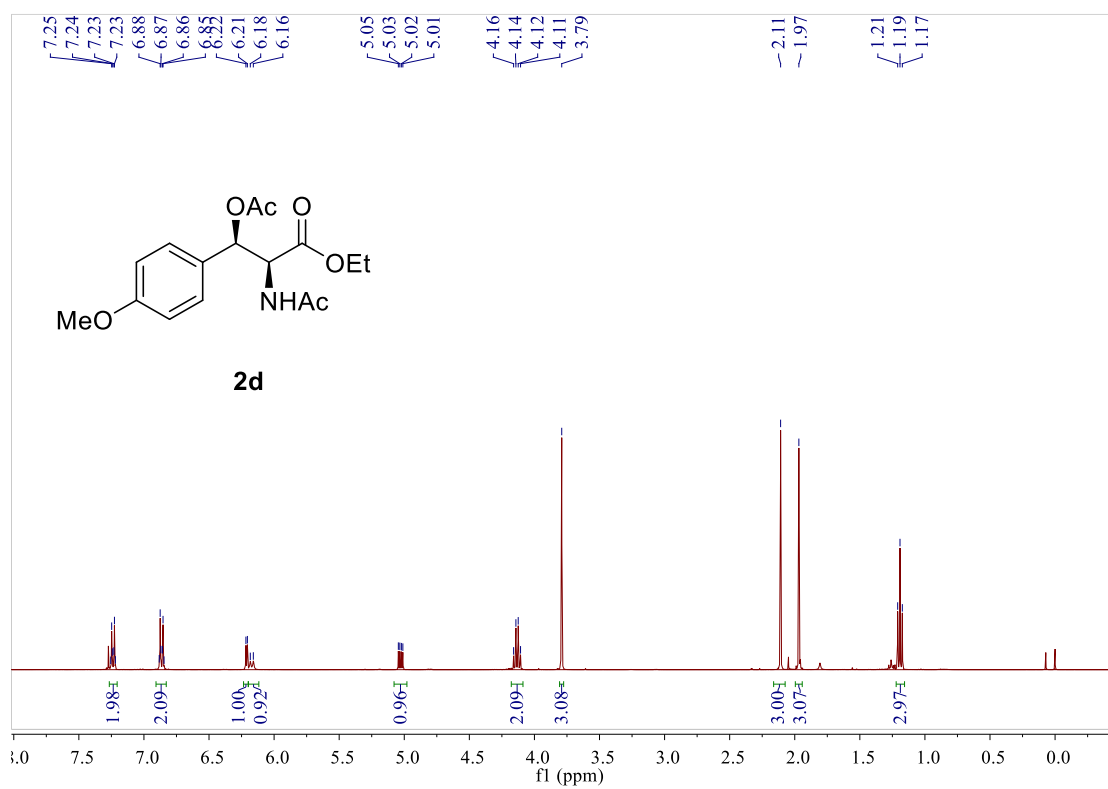


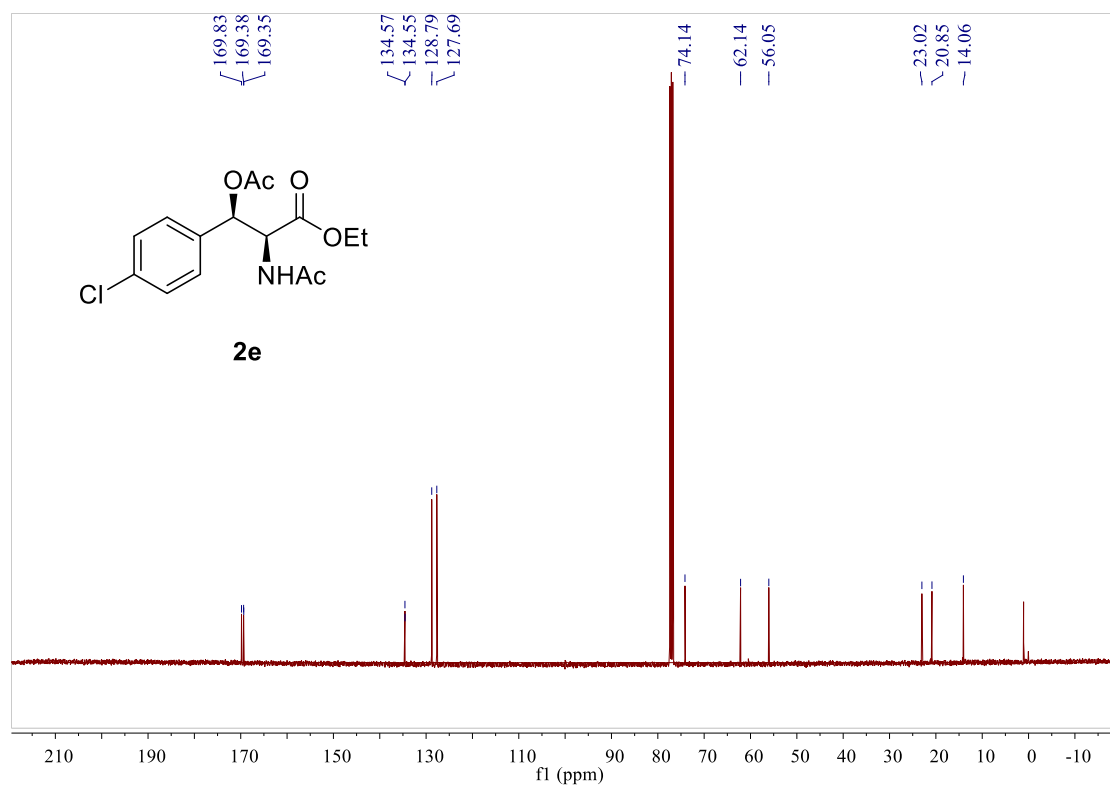
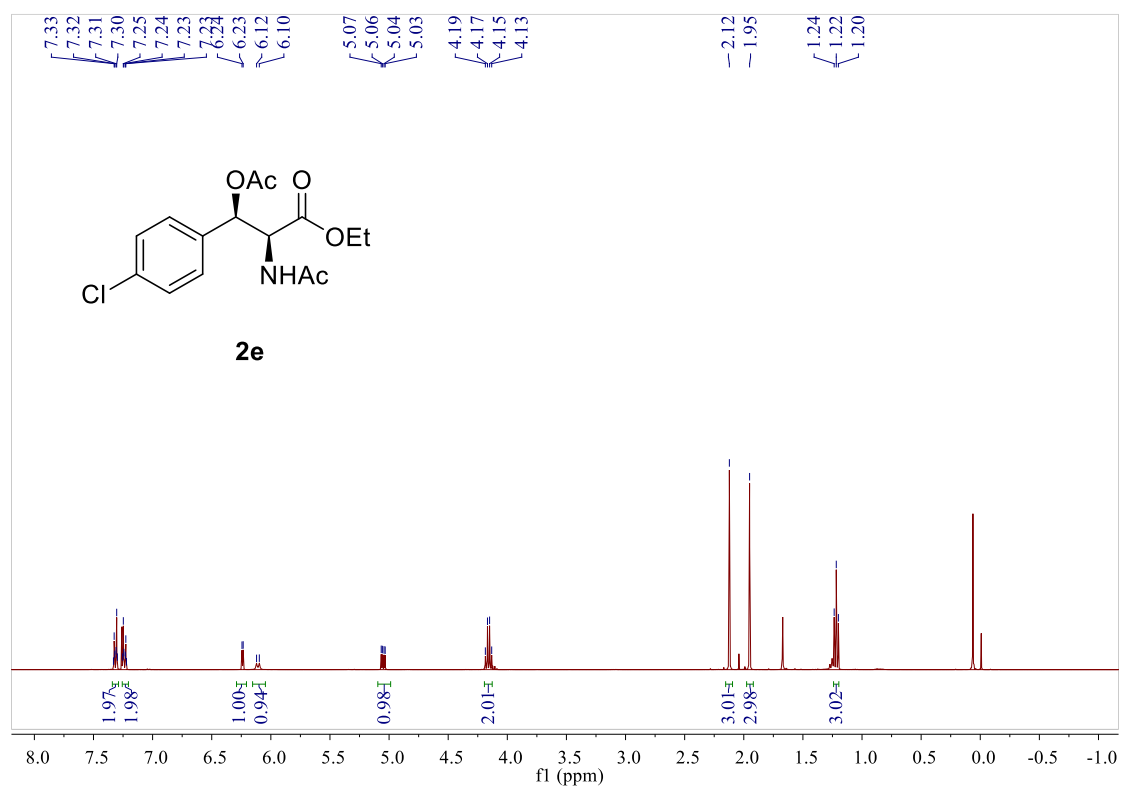


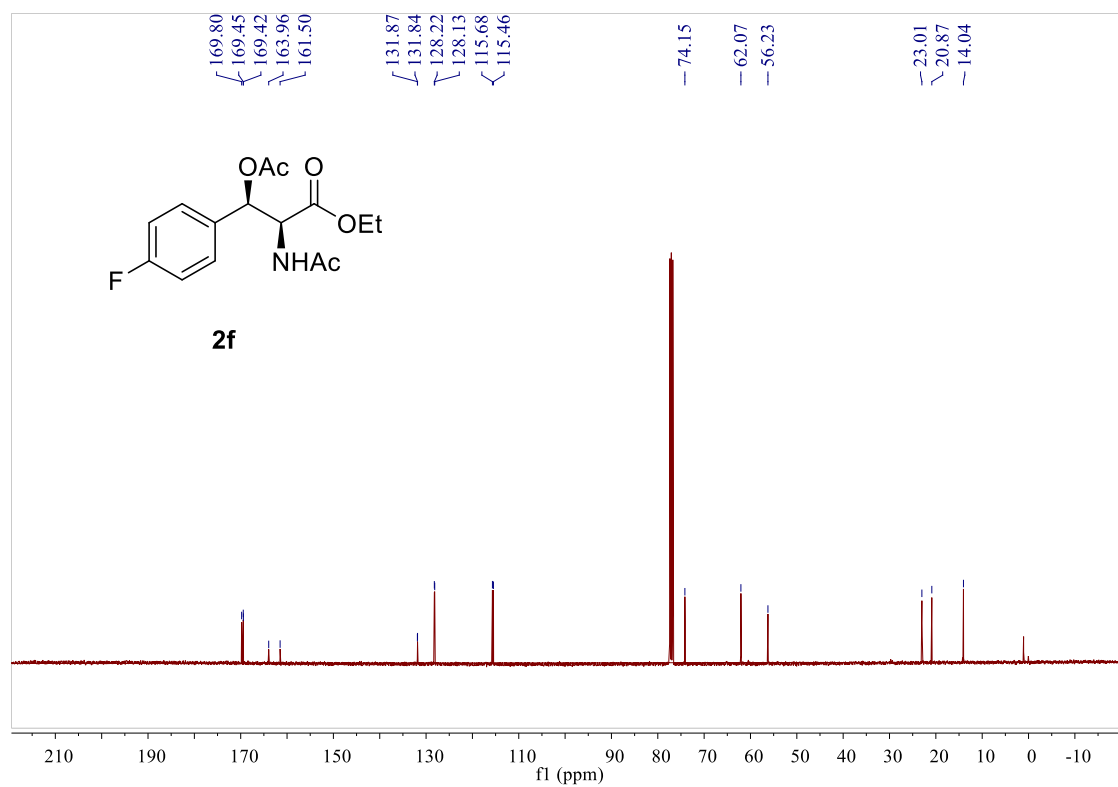
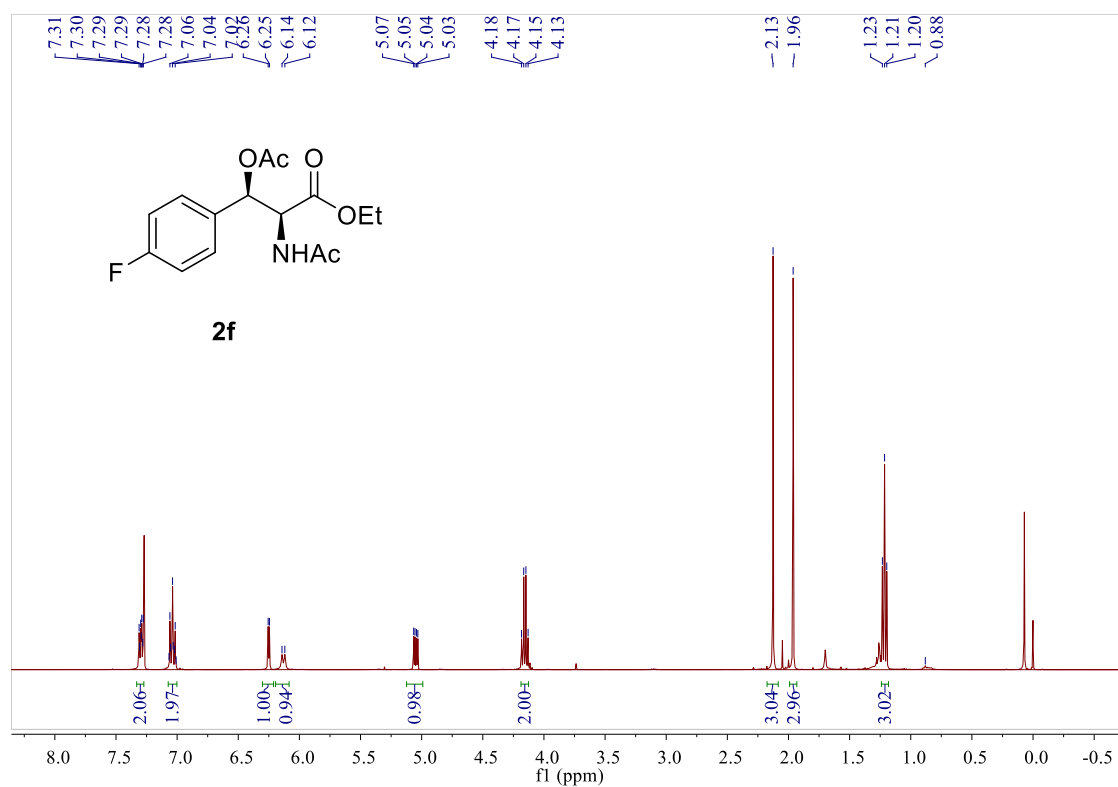


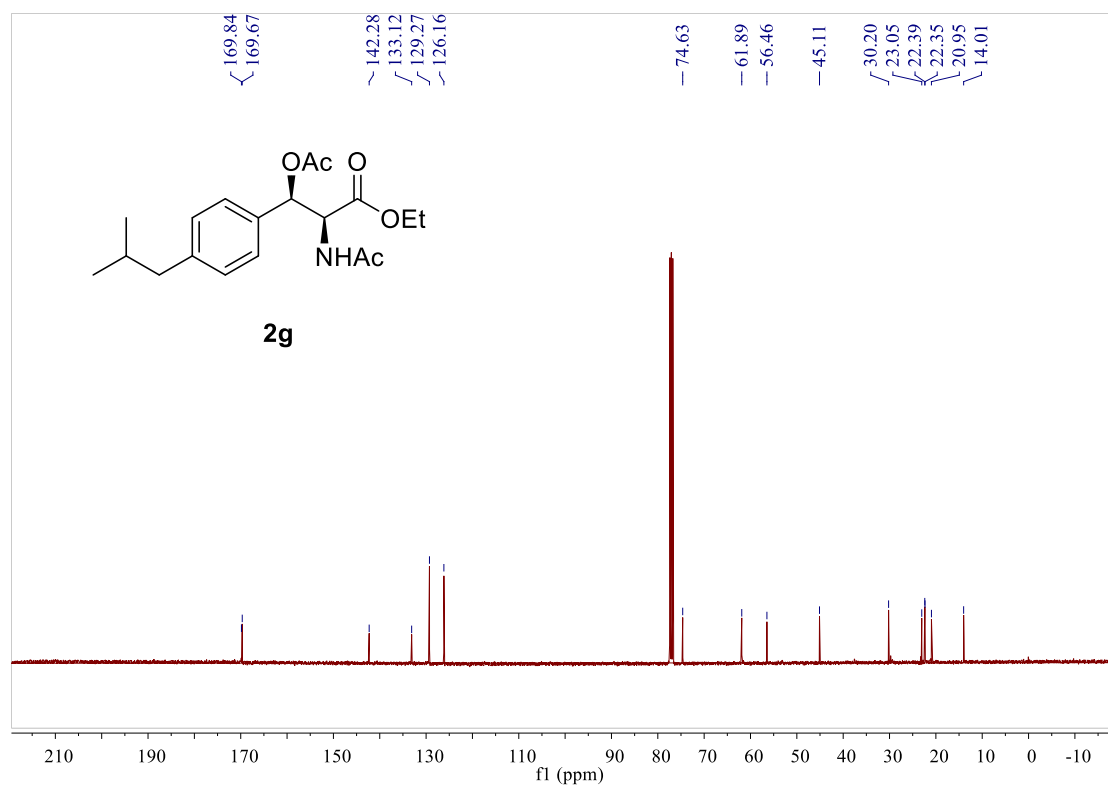
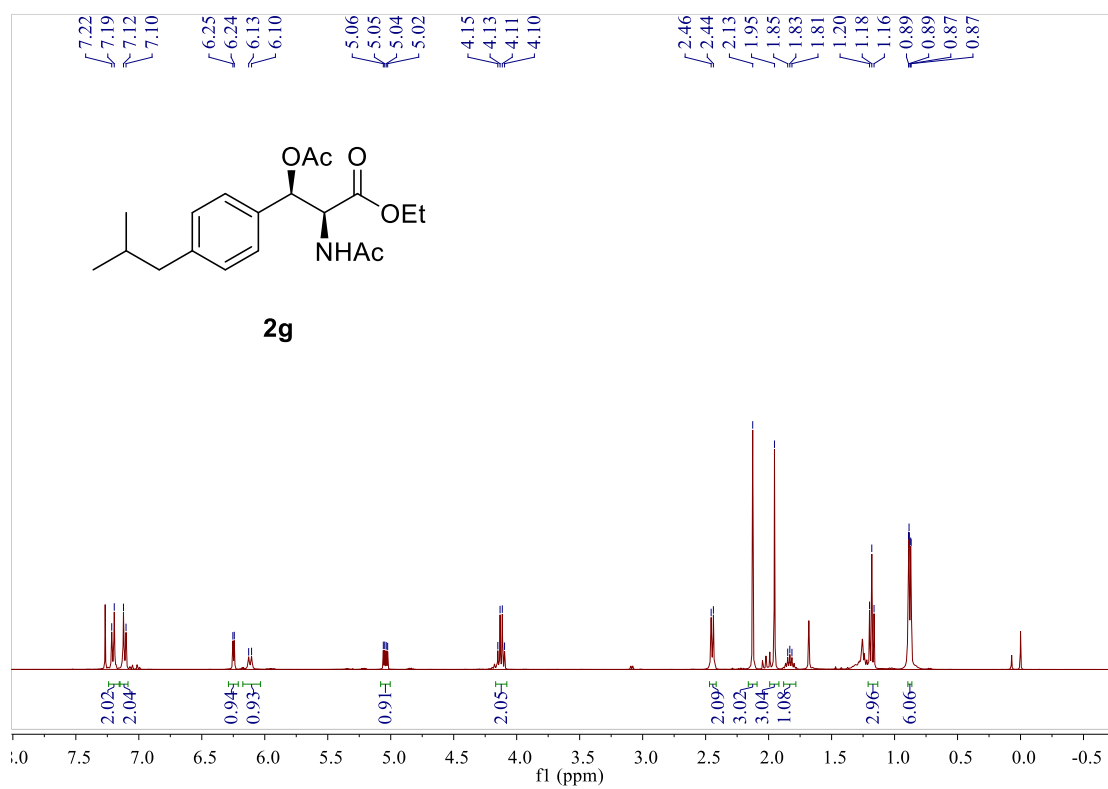


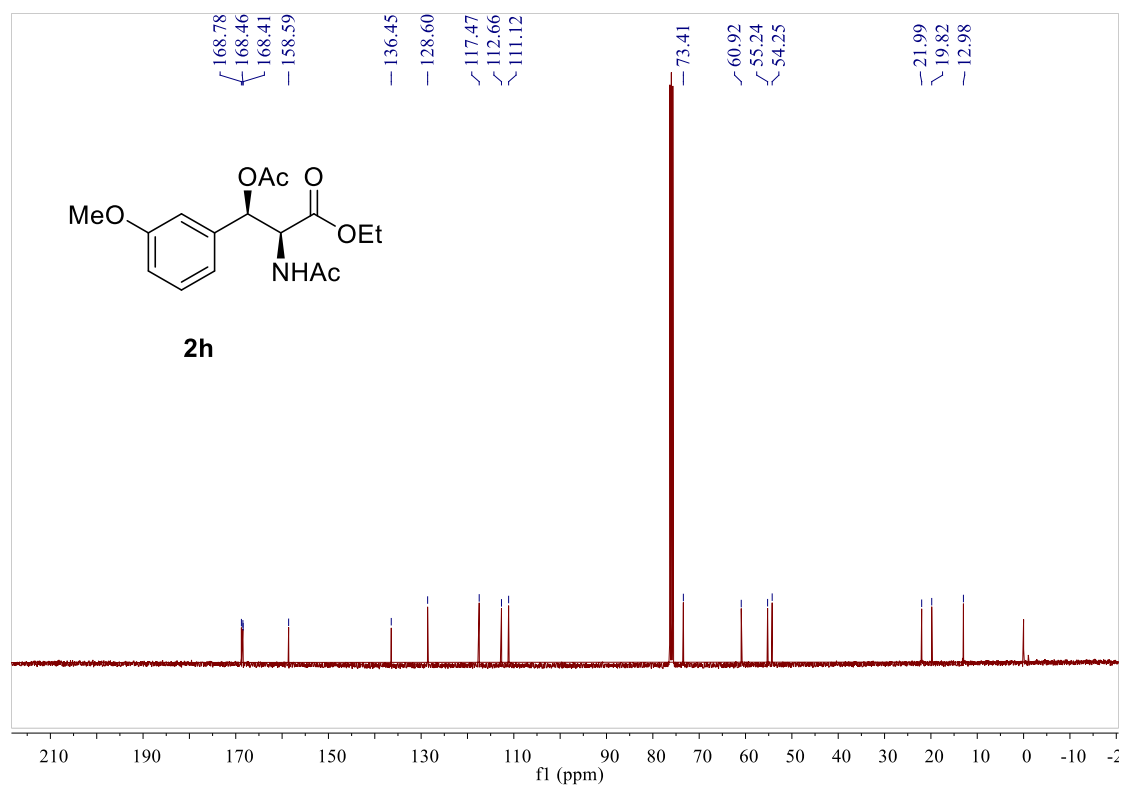
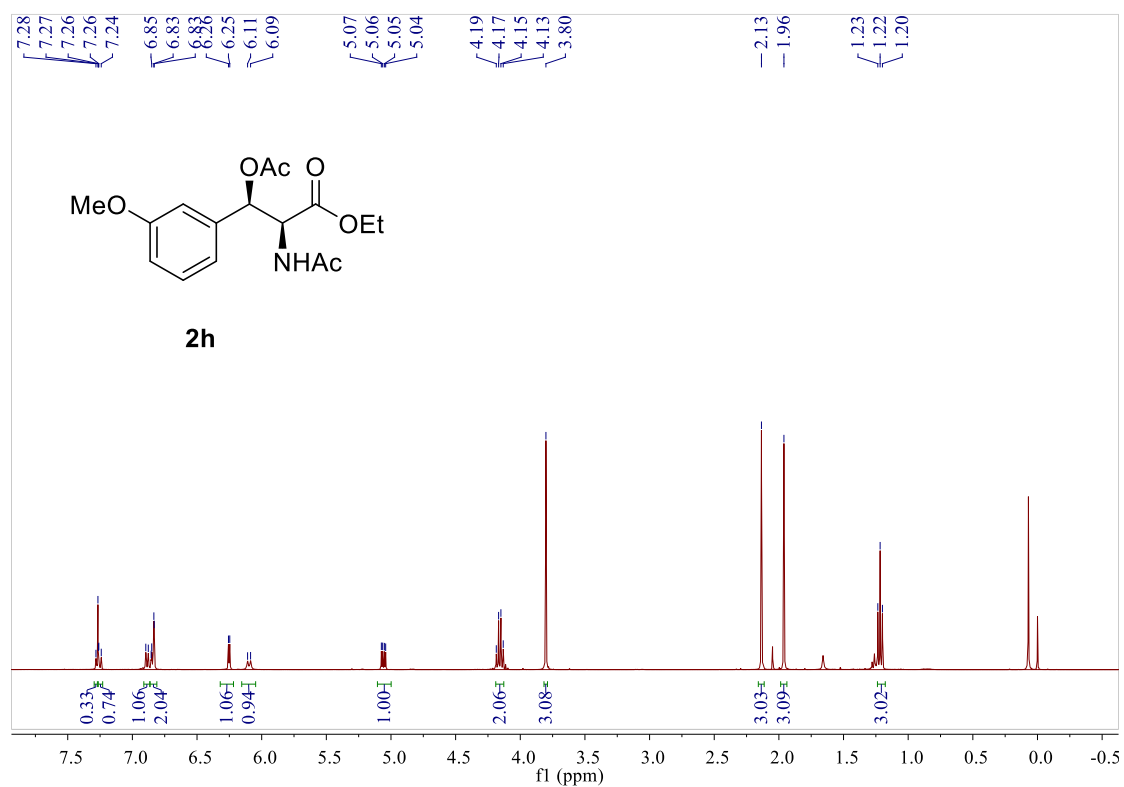


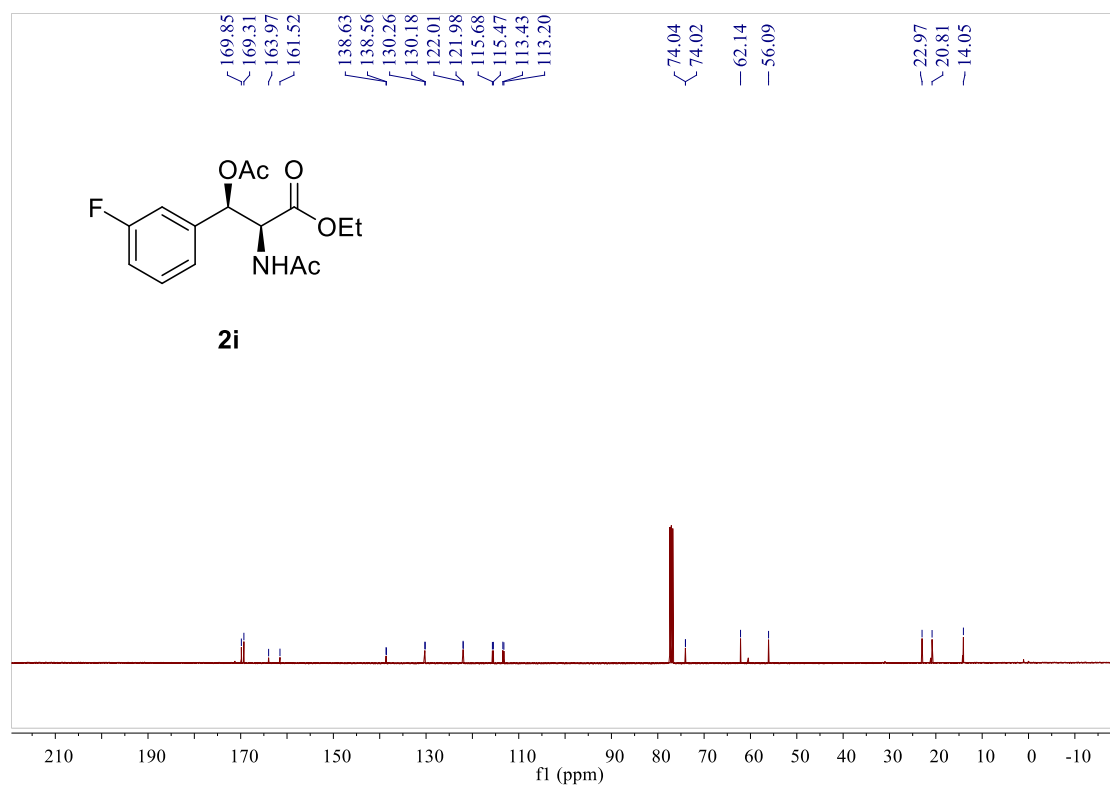
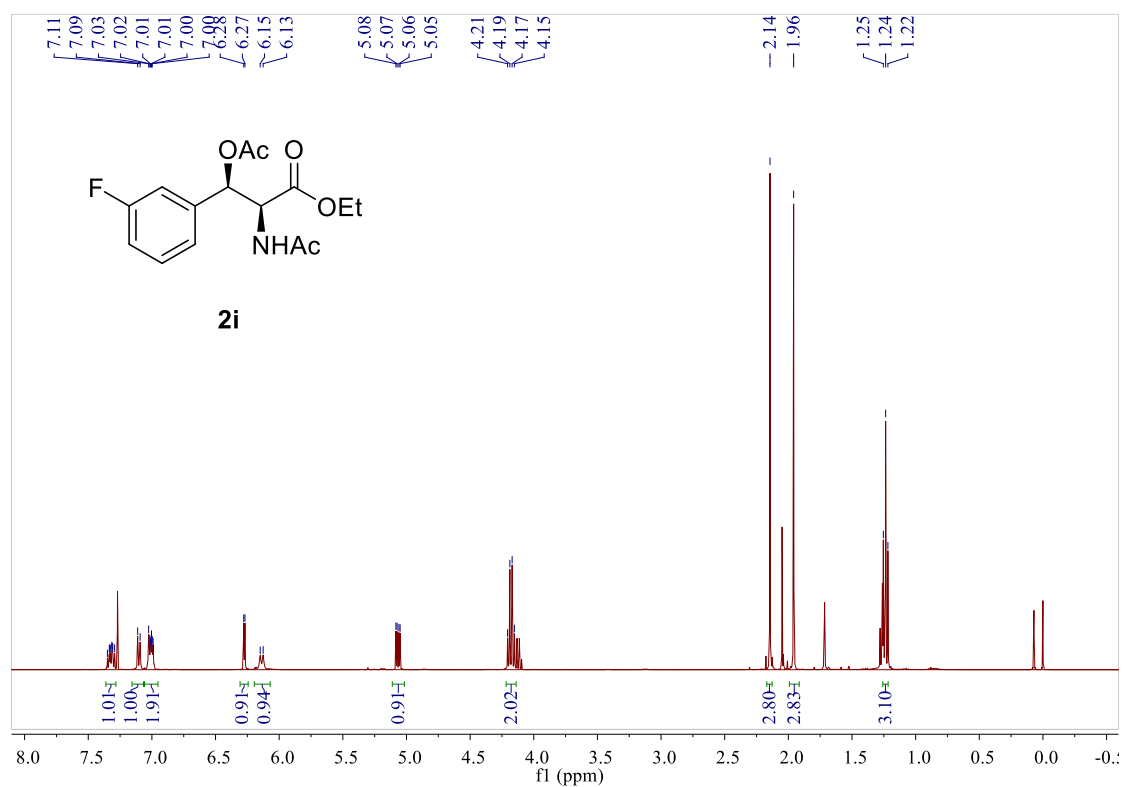


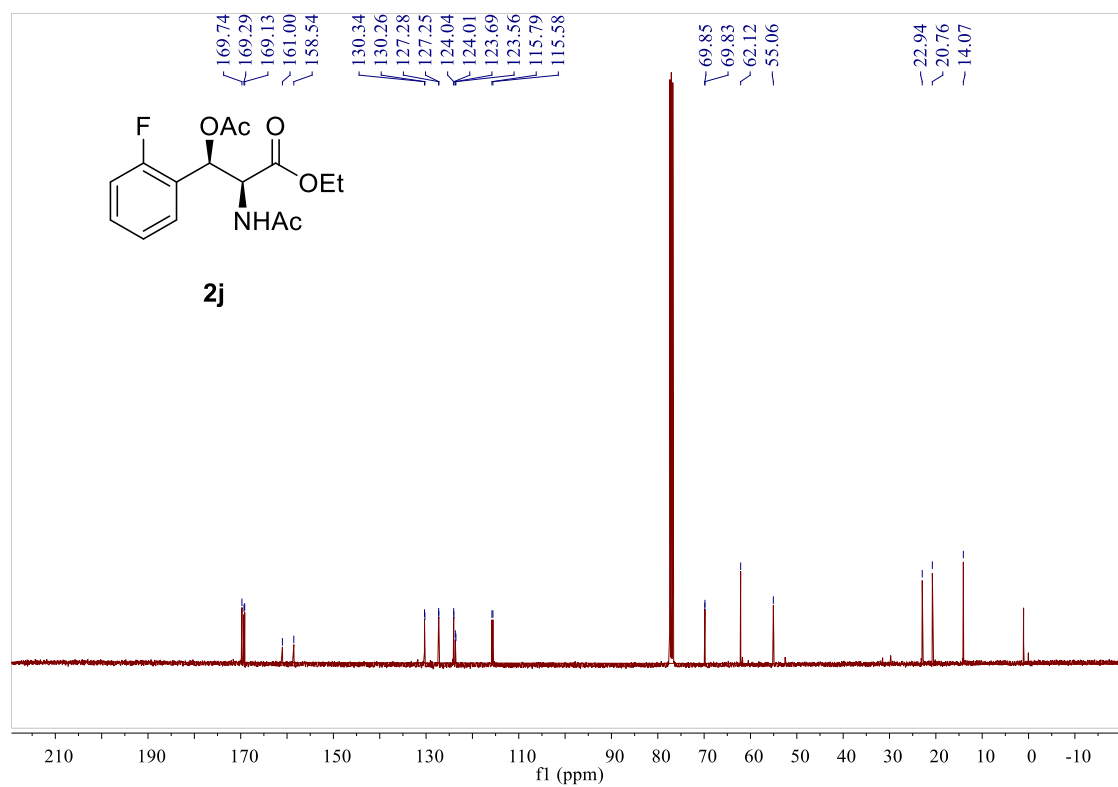
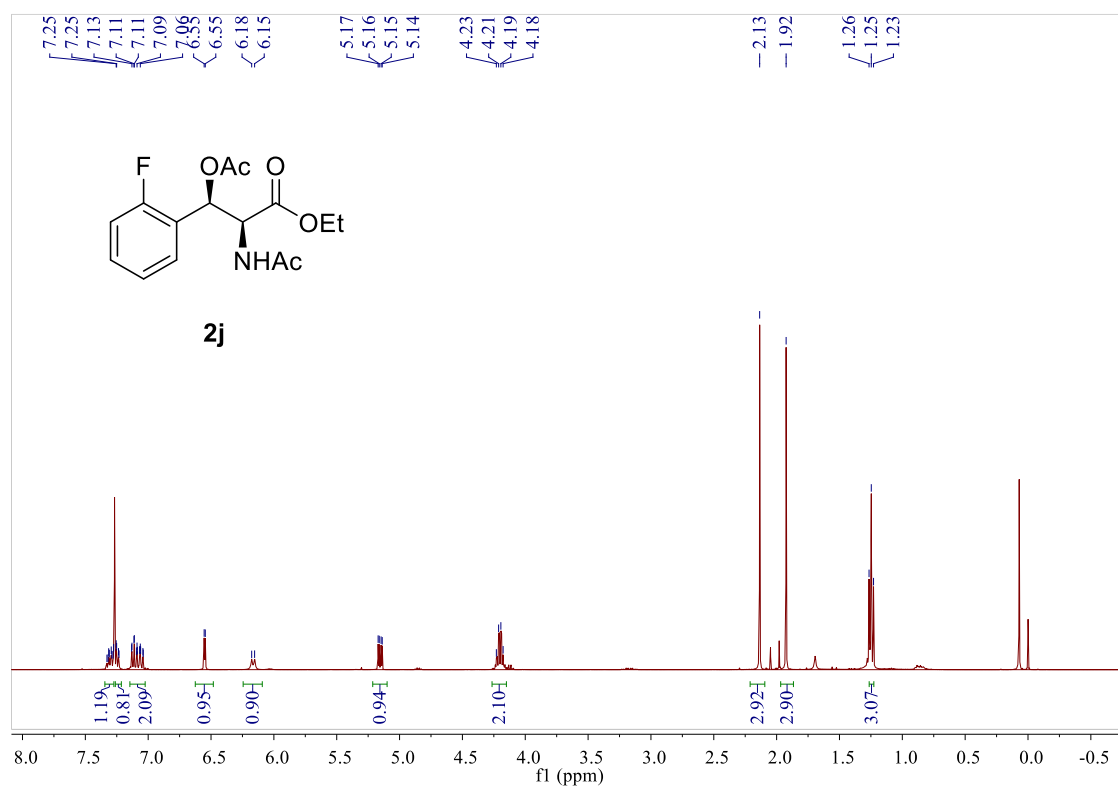


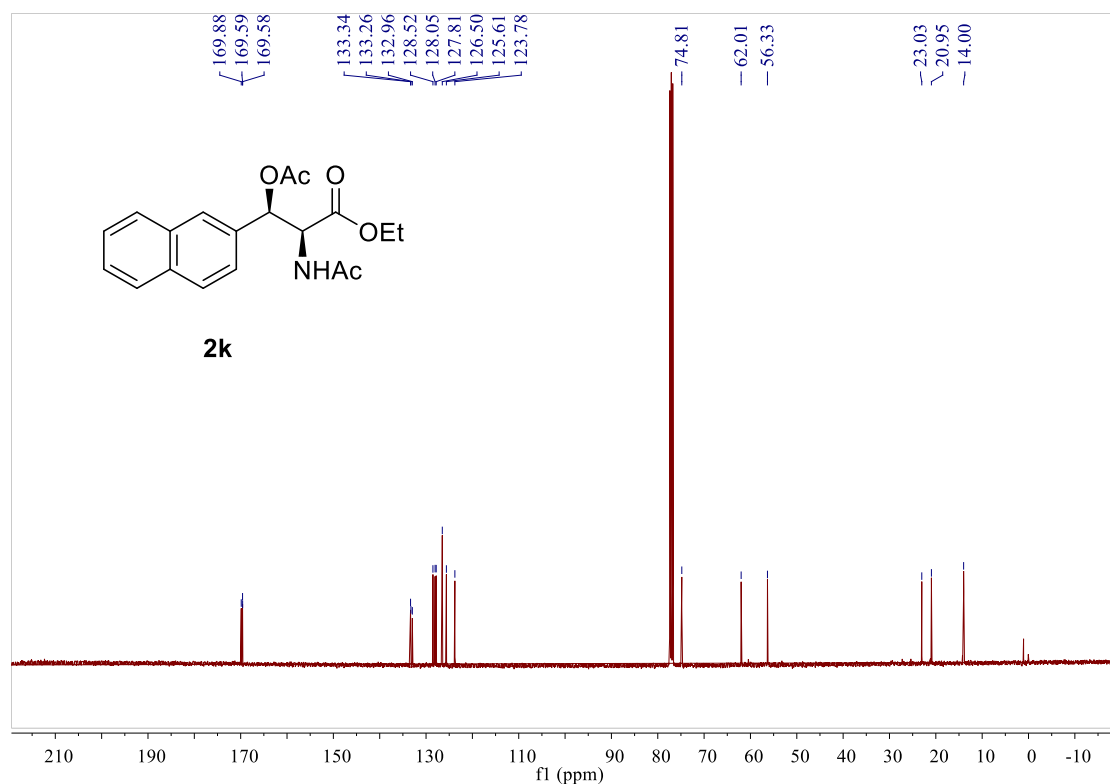
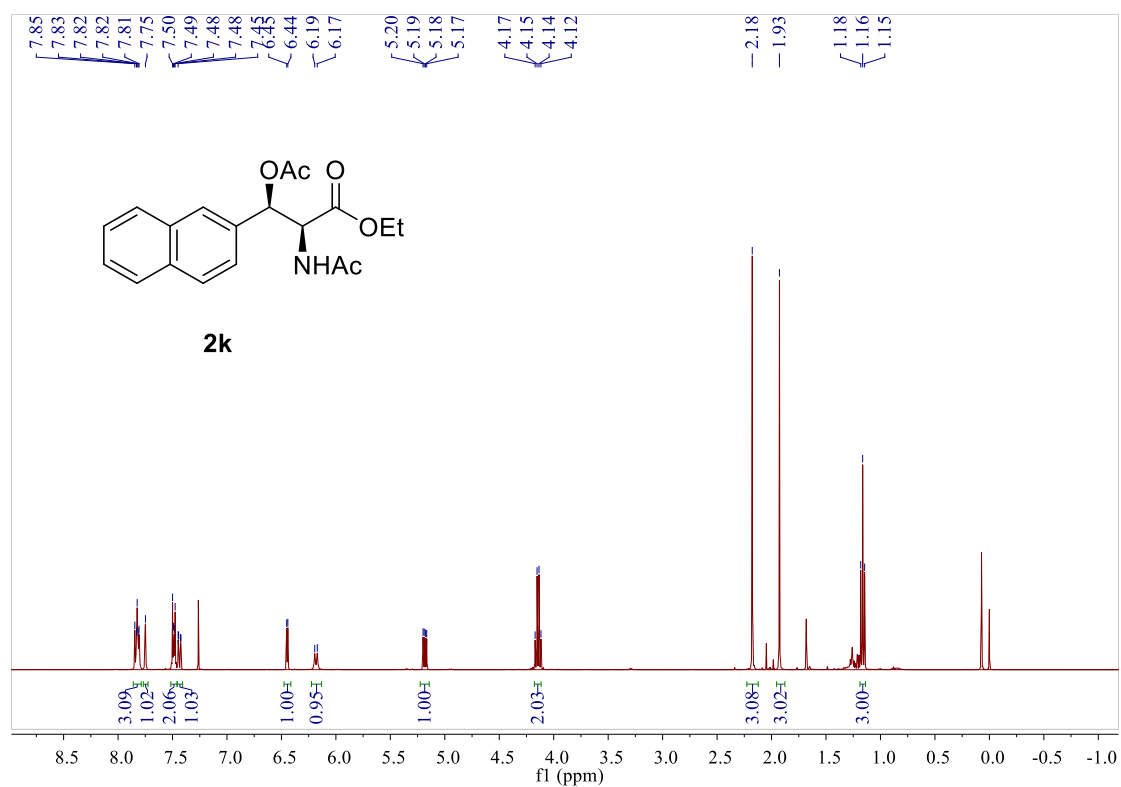


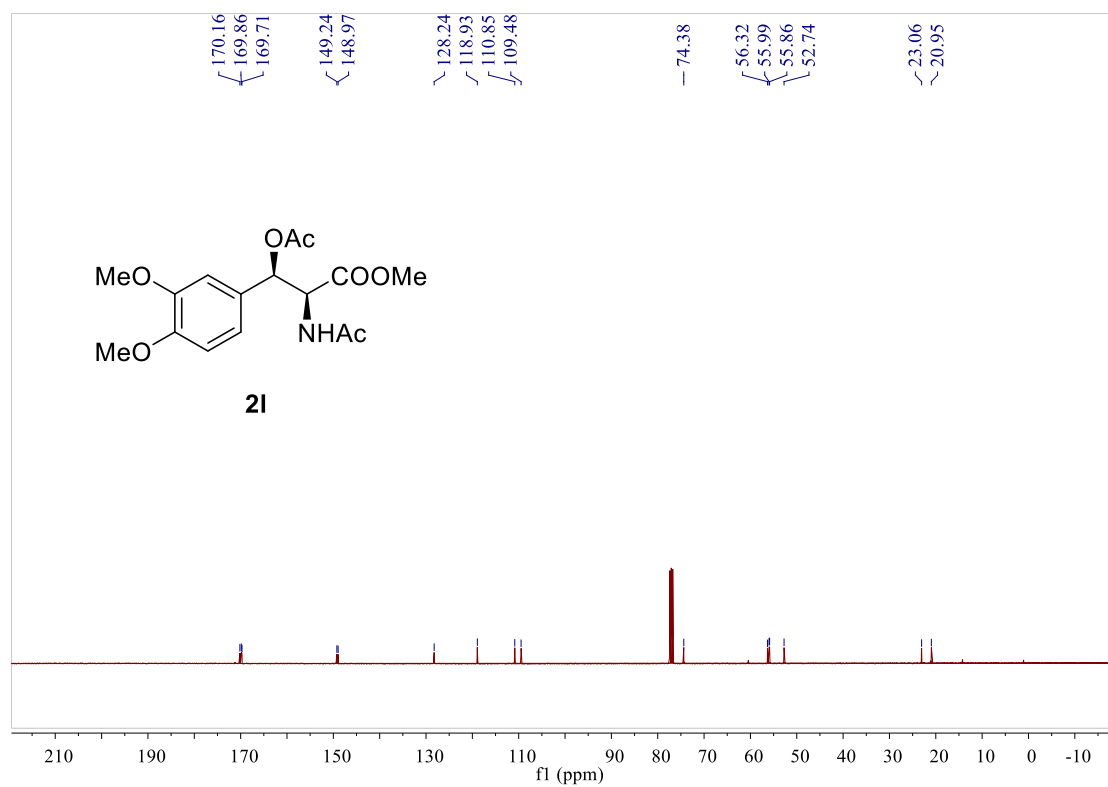
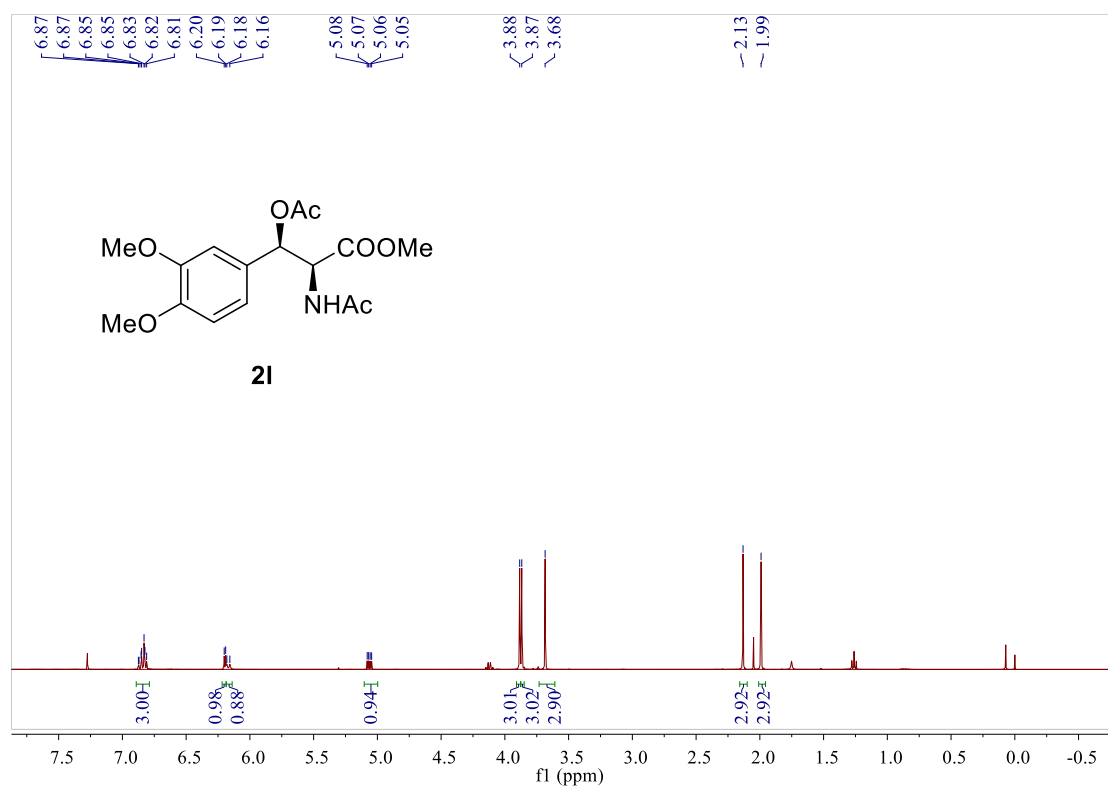


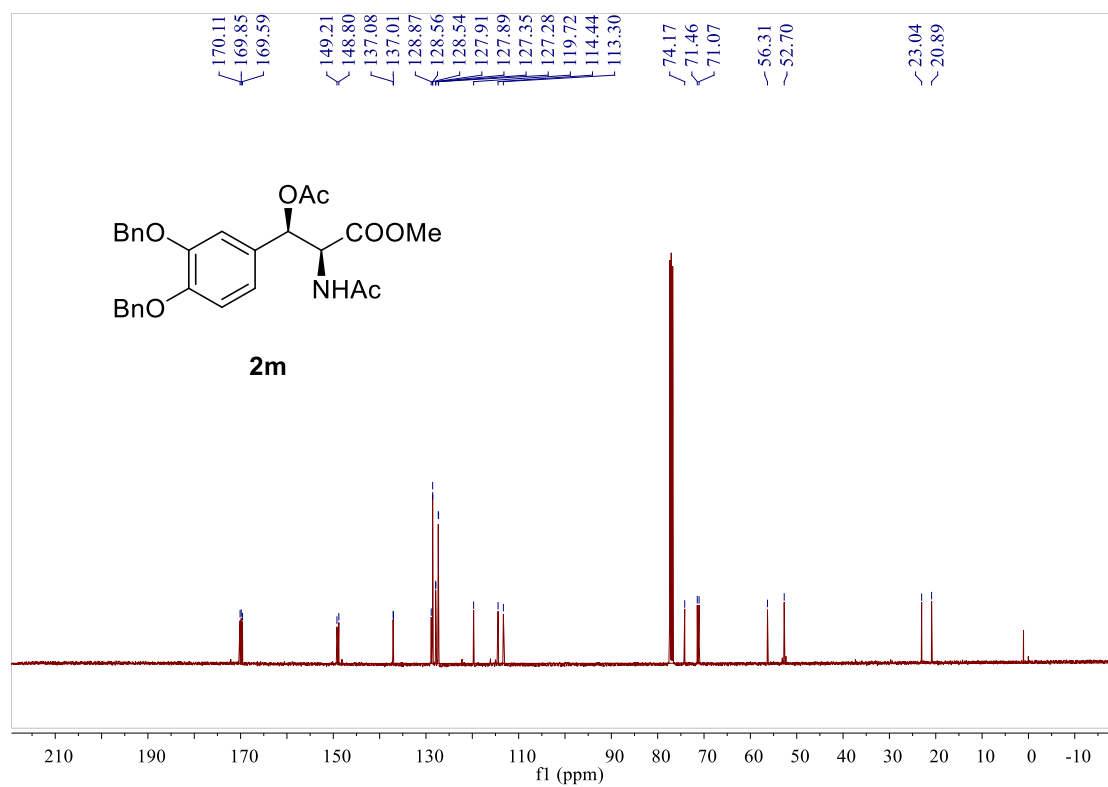
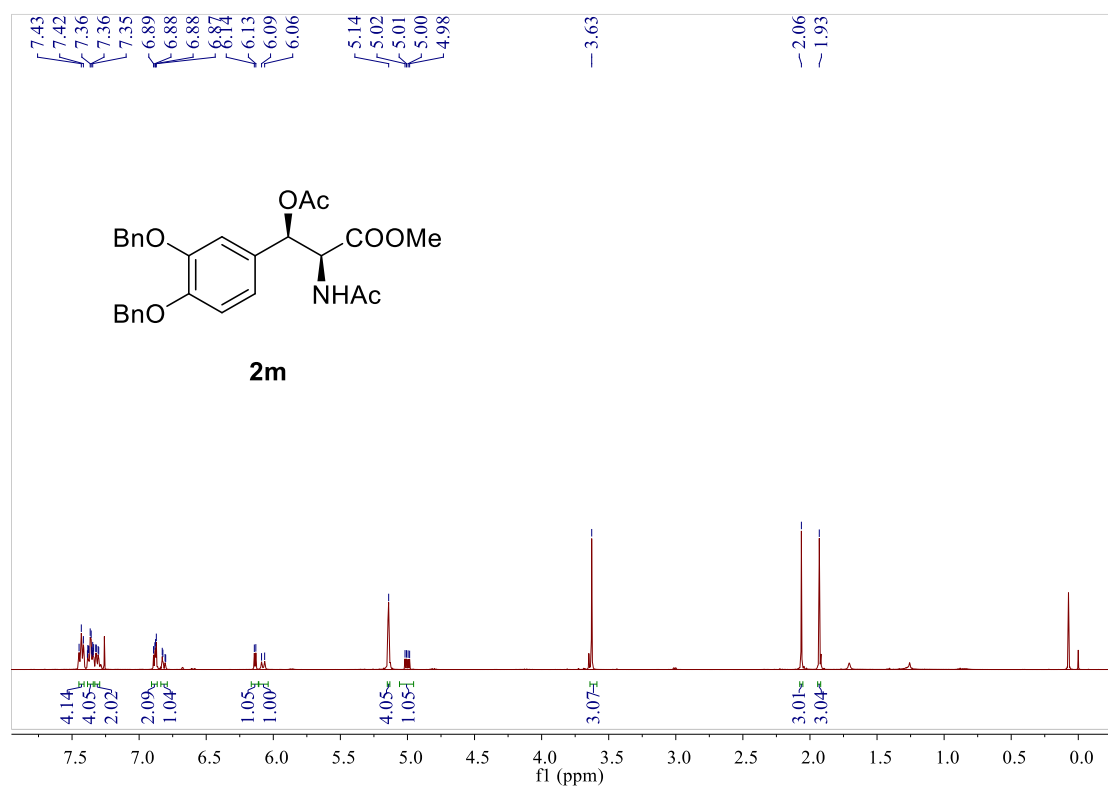


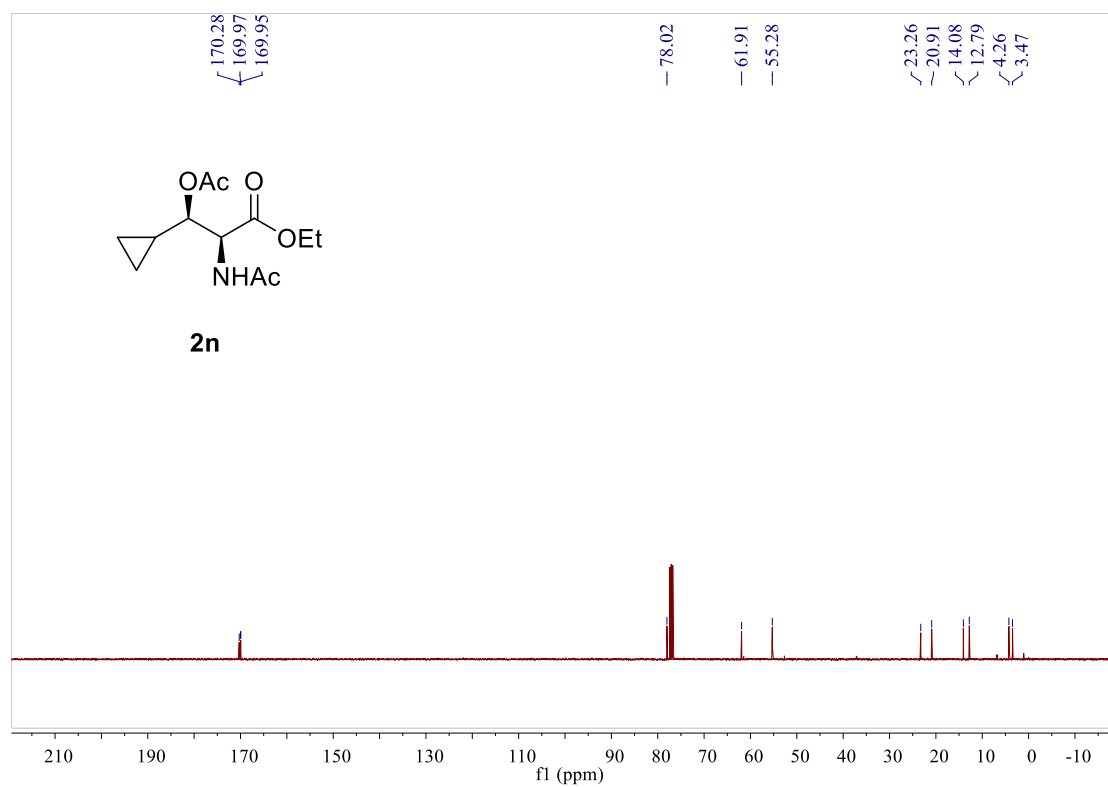
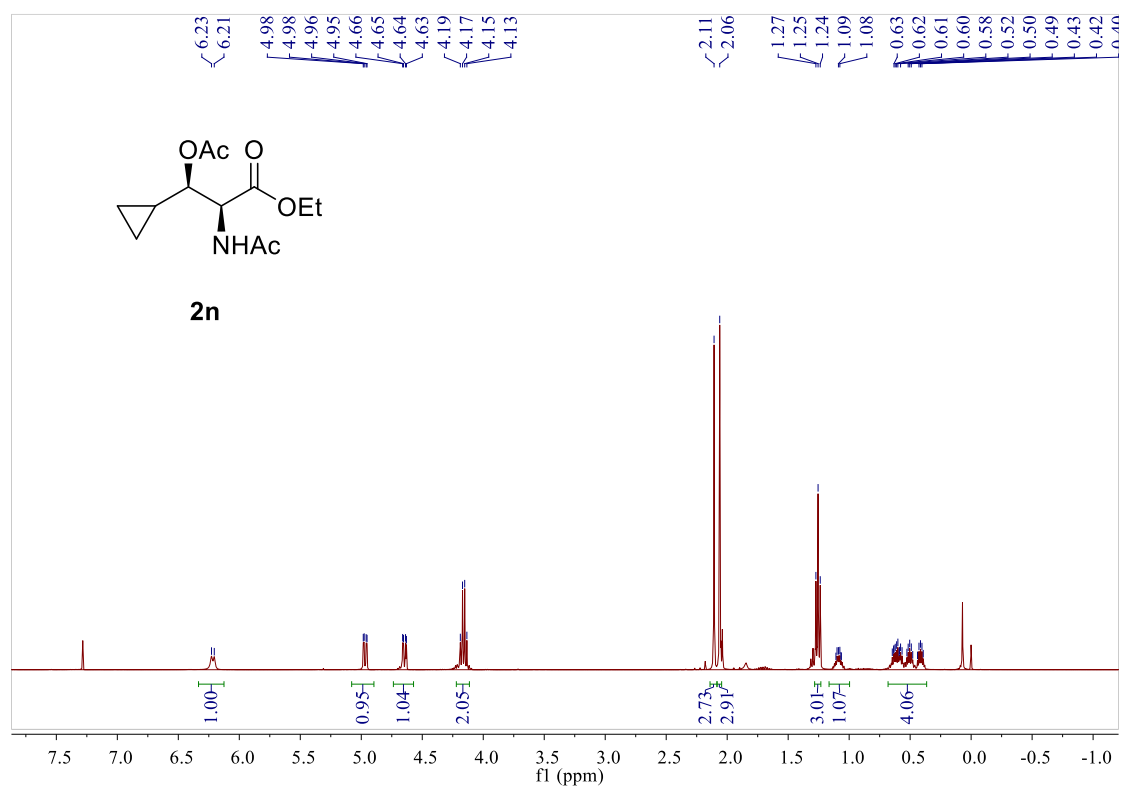


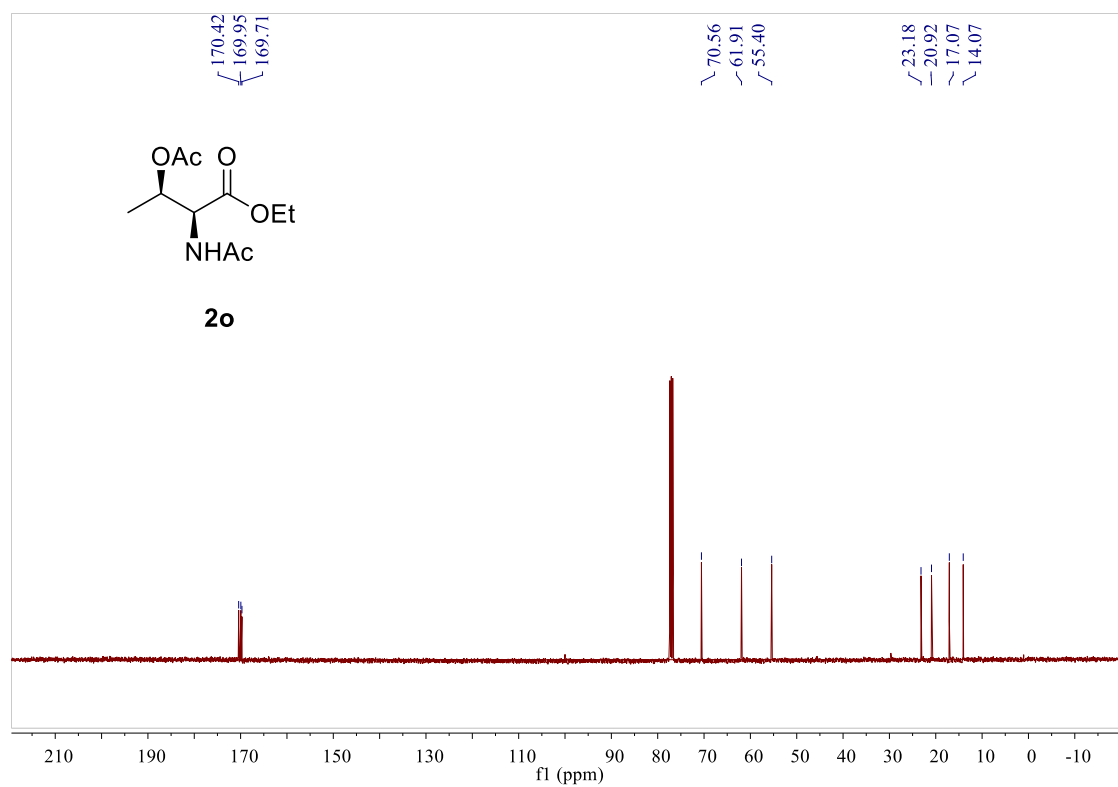
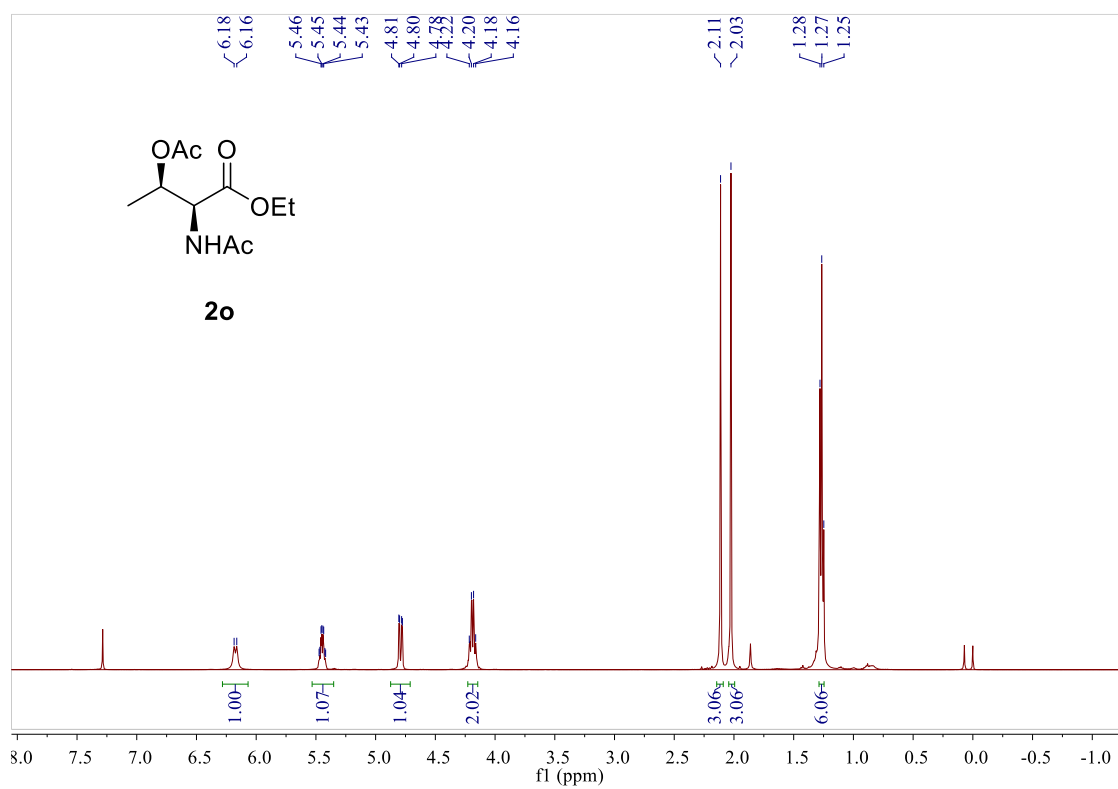


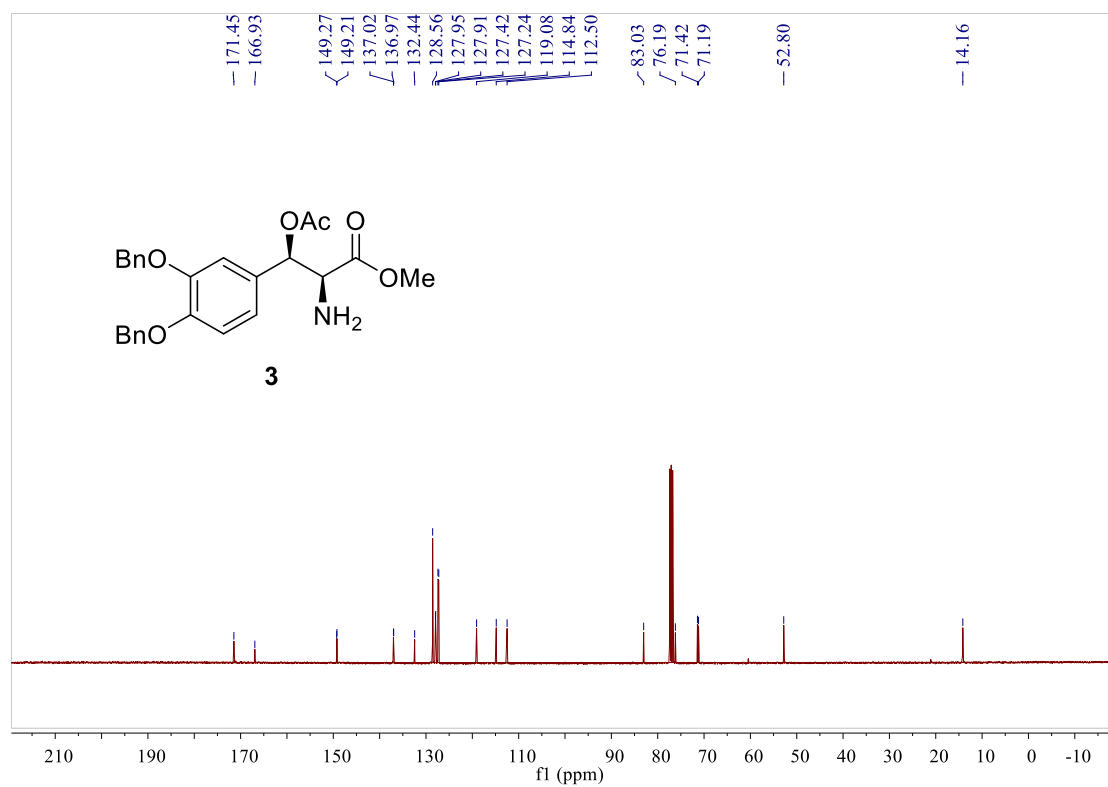
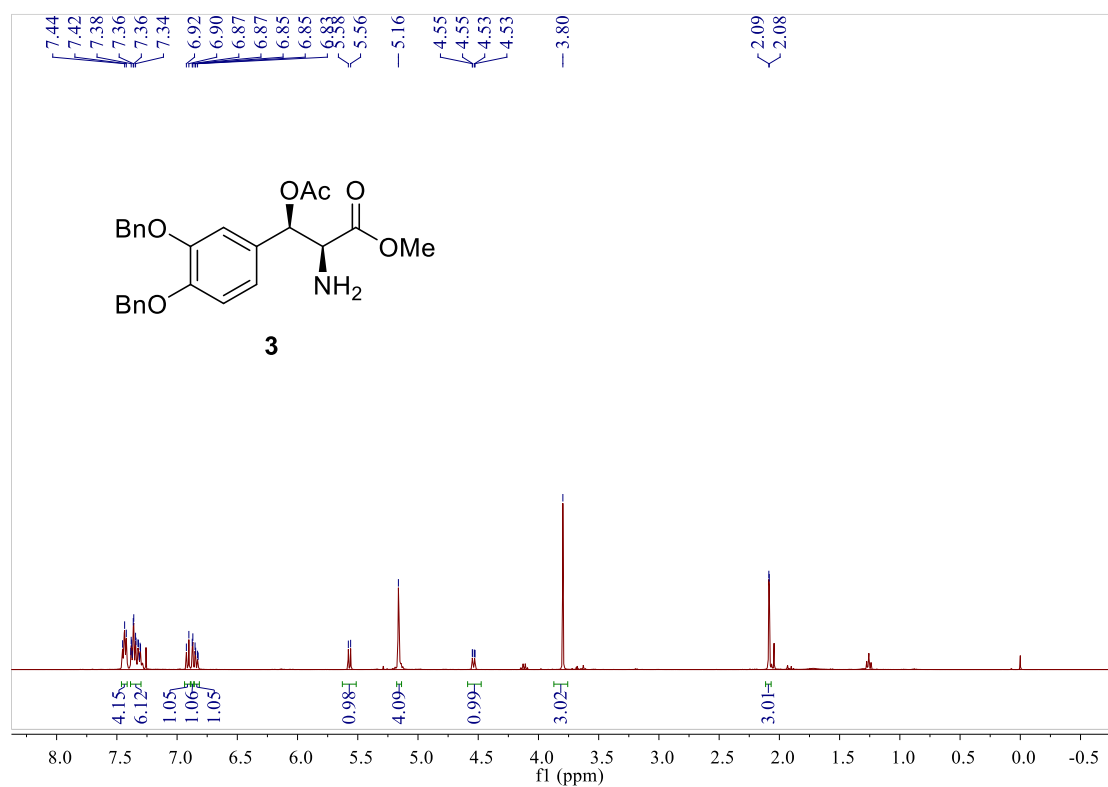


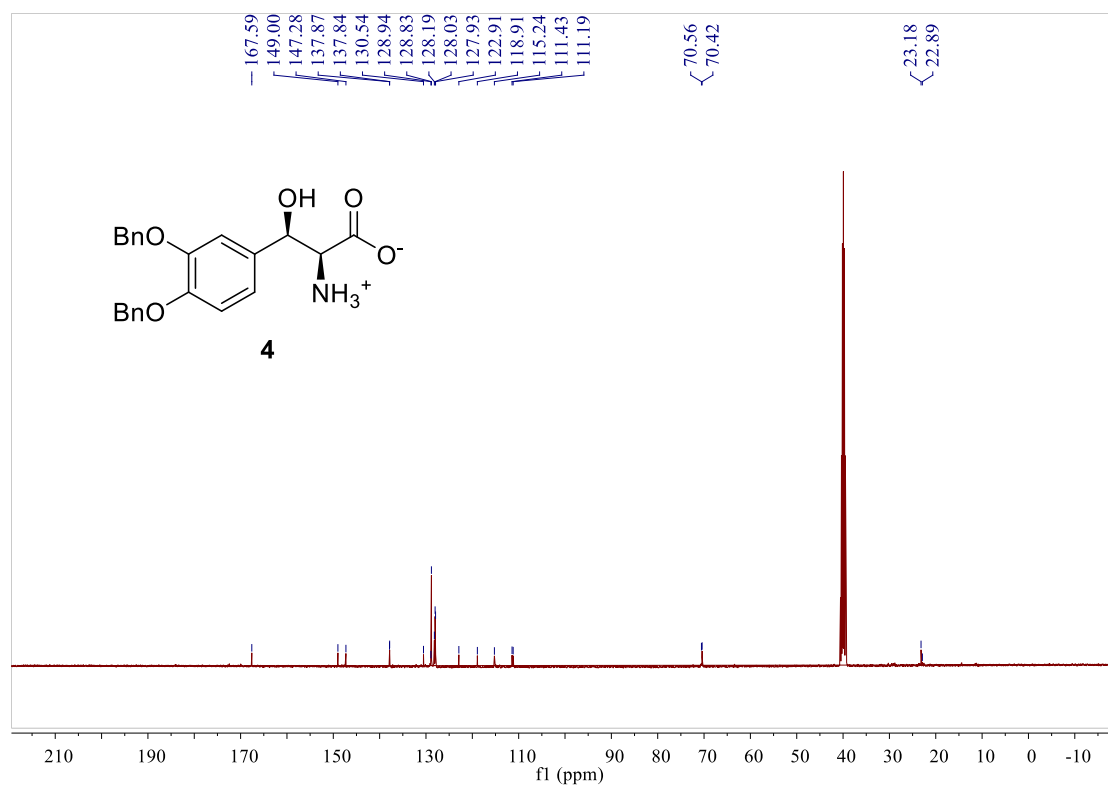
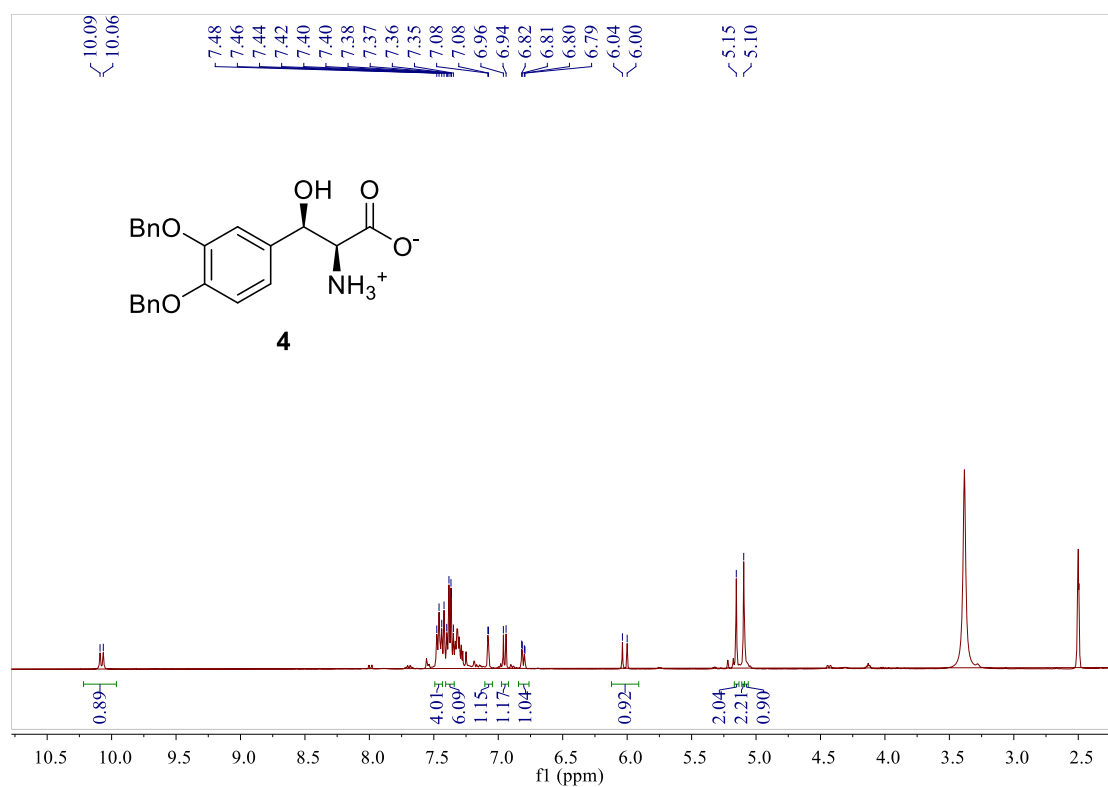










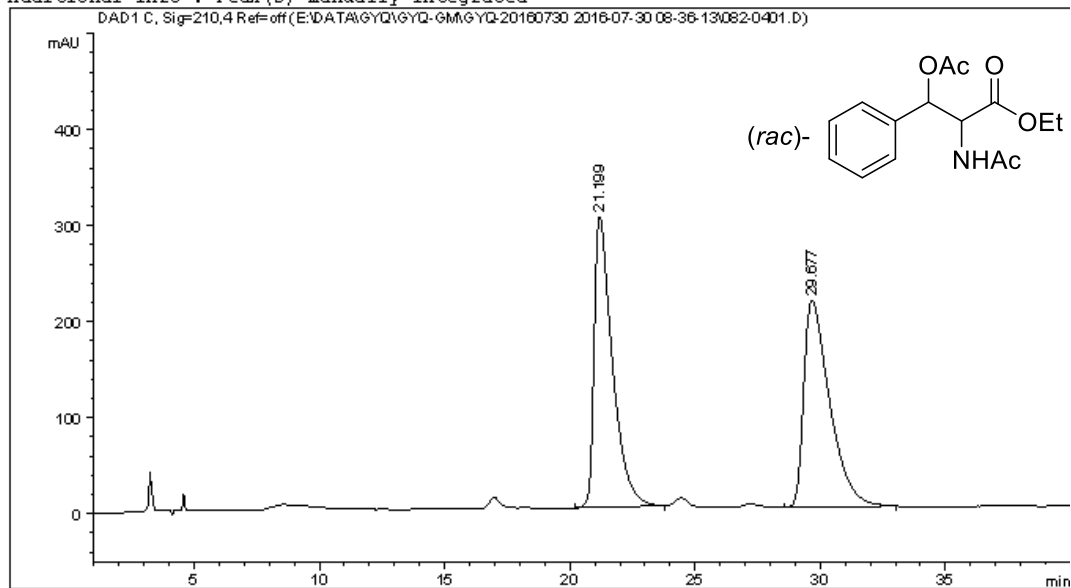


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Sample Name: Ph-rac

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Injection Date	: 7/30/2016 9:59:02 AM	Inj	: 1
		Inj Volume	: 5.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\GYQ-20160730 2016-07-30 08-36-13\DAD-0J(1-6)-95-5-1. OML-5-210NM-50MIN.M		
Last changed	: 7/30/2016 9:58:07 AM by SYSTEM		
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Last changed	: 6/10/2017 8:57:32 PM by SYSTEM (modified after loading)		

Additional Info : Peak(s) manually integrated



Area Percent Report

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 C, Sig=210,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	21.199	BB	0.7367	1.53169e4	302.53458	49.7010
2	29.677	BB	1.0322	1.55012e4	215.25539	50.2990

Totals : 3.08182e4 517.78996

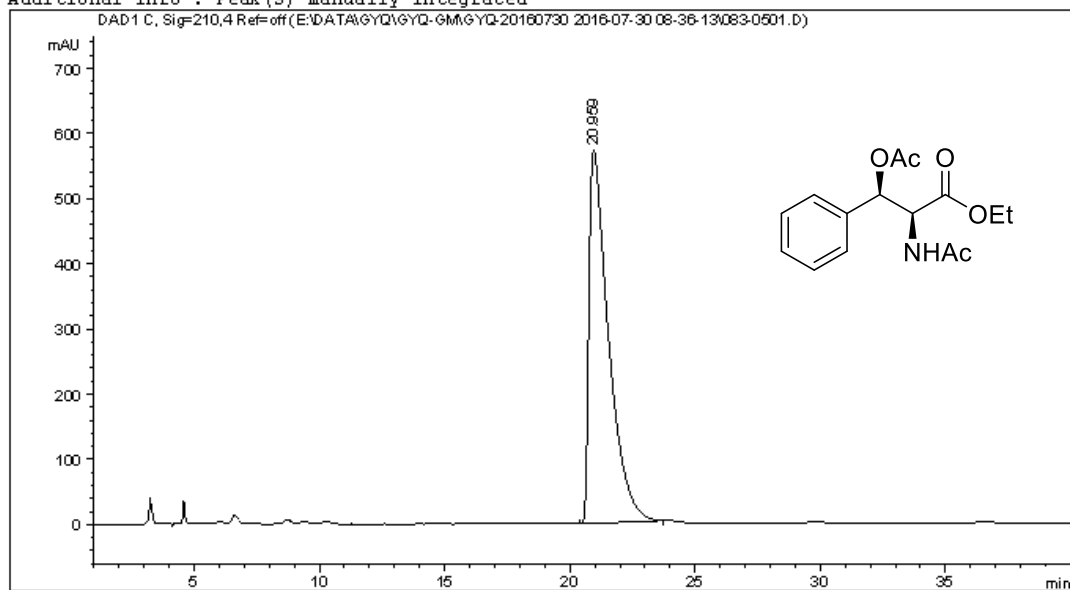
*** End of Report ***

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Sample Name: 30 bar

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Injection Date	: 7/30/2016 10:47:56 AM	Inj	: 1
		Inj Volume	: 5.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\GYQ-20160730 2016-07-30 08-36-13\DAD-0J(1-6)-95-5-1. OML-5-210NM-50MIN.M		
Last changed	: 7/30/2016 9:58:07 AM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\GYQ-20160730 2016-07-30 08-36-13\DAD-0J(1-6)-95-5-1. OML-5-210NM-50MIN.M (Sequence Method)		
Last changed	: 6/10/2017 9:01:32 PM by SYSTEM (modified after loading)		

Additional Info : Peak(s) manually integrated



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Area Percent Report
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Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 C, Sig=210,4 Ref=off

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Totals : 3.09458e4 572.71411

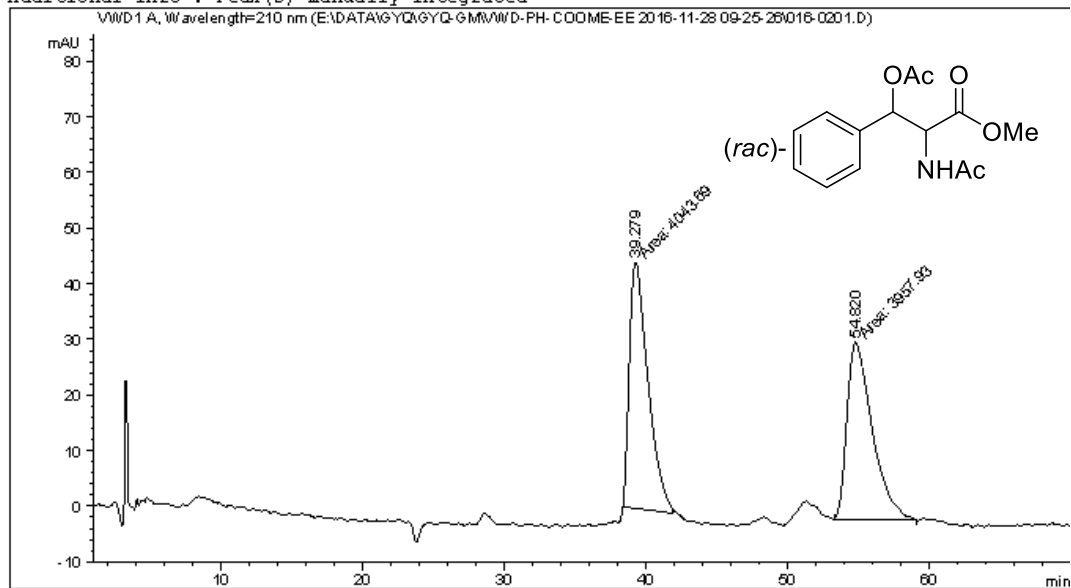
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*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\VWD-PH-COOME-EE 2016-11-28 09-25-26\016-0201.D
Sample Name: PH-COOME-RAC

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Acq. Operator	: SYSTEM	Seq. Line	: 2
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Injection Date	: 11/28/2016 9:46:59 AM	Inj	: 1
		Inj Volume	: 5.000 µl
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Analysis Method	: E:\DATA\GYQ\GYQ-GM\VWD-PH-COOME-EE 2016-11-28 09-25-26\VWD-0J(1-6)-95-5-1.0ML-210NM-75MIN.M (Sequence Method)		
Last changed	: 6/10/2017 9:04:41 PM by SYSTEM		
	(modified after loading)		
Additional Info : Peak(s) manually integrated			

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Area Percent Report
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Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: VWD1 A, Wavelength=210 nm

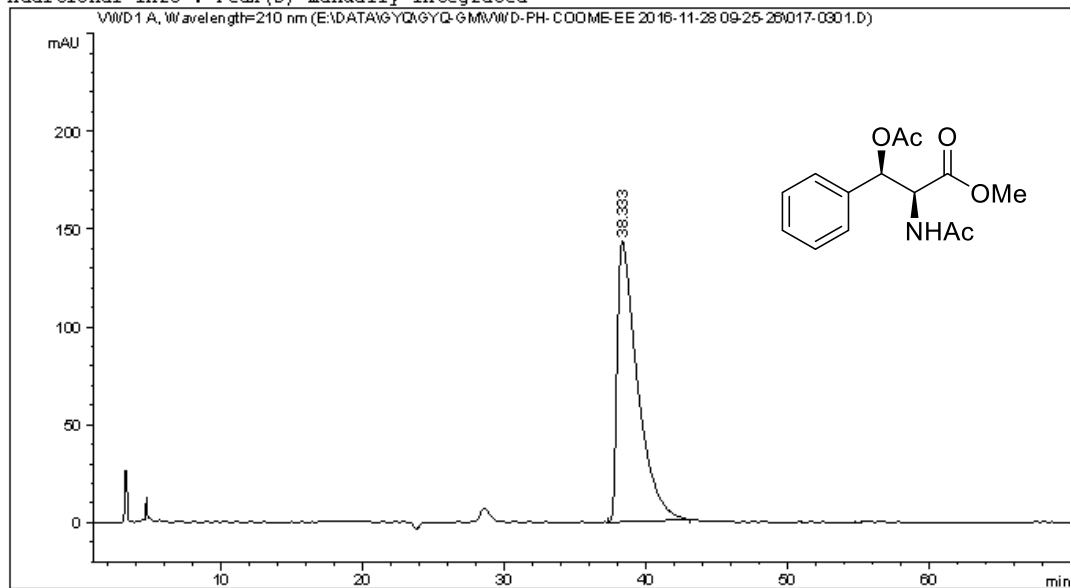
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
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2	54.820	MM	2.0652	3957.92822	31.94175	49.4641

Totals : 8001.61426 76.24330

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*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\VWD-PH-COOME-EE 2016-11-28 09-25-26\017-0301.D
Sample Name: PH-COOME-EE

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Injection Date  : 11/28/2016 11:02:44 AM      Inj       :    1
                                           Inj Volume: 5.000 µl
Acq. Method     : E:\DATA\GYQ\GYQ-GM\VWD-PH-COOME-EE 2016-11-28 09-25-26\VWD-0J(1-6)-95-5-
                                           1.0ML-210NM-75MIN.M
Last changed    : 11/28/2016 9:25:26 AM by SYSTEM
Analysis Method : E:\DATA\GYQ\GYQ-GM\VWD-PH-COOME-EE 2016-11-28 09-25-26\VWD-0J(1-6)-95-5-
                                           1.0ML-210NM-75MIN.M (Sequence Method)
Last changed    : 6/10/2017 9:08:02 PM by SYSTEM
                                           (modified after loading)
Additional Info  : Peak(s) manually integrated
=====
```



=====
Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: VWD1 A, Wavelength=210 nm

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	38.333	BB	1.4611	1.43183e4	143.24849	100.0000

Totals : 1.43183e4 143.24849

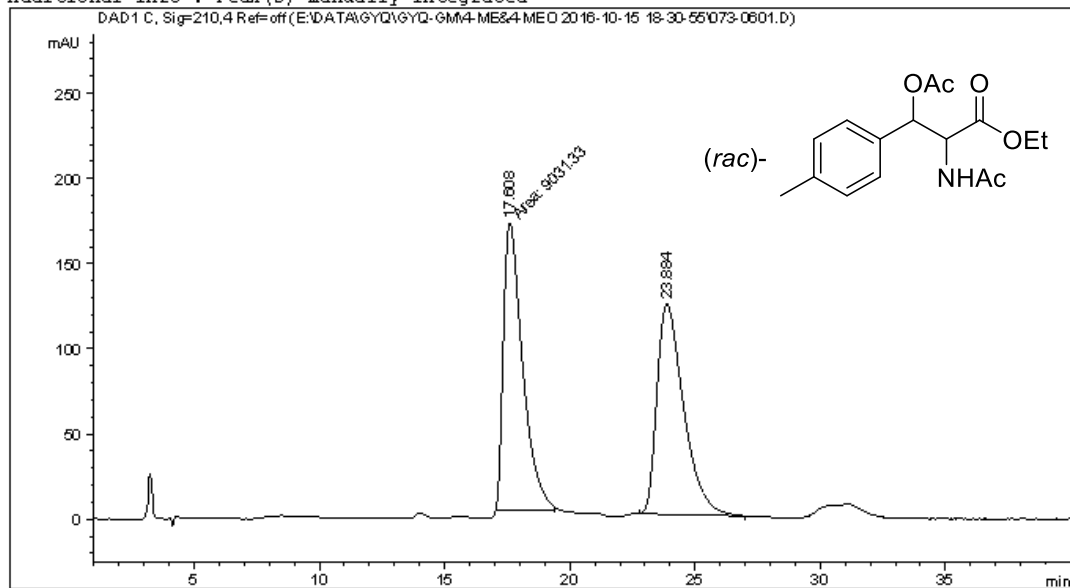
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*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\4-ME&4-MEO 2016-10-15 18-30-55\073-0601.D
Sample Name: 4-Me-rac

=====

Acq. Operator	: SYSTEM	Seq. Line	: 6
Acq. Instrument	: 1260HPLC-DAD	Location	: Vial 73
Injection Date	: 10/15/2016 10:33:42 PM	Inj	: 1
		Inj Volume	: 5.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\4-ME&4-MEO 2016-10-15 18-30-55\DAD-0J(1-6)-95-5-1.OML-5-210NM-50MIN.M		
Last changed	: 10/15/2016 6:30:56 PM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\4-ME&4-MEO 2016-10-15 18-30-55\DAD-0J(1-6)-95-5-1.OML-5-210NM-50MIN.M (Sequence Method)		
Last changed	: 6/10/2017 9:10:36 PM by SYSTEM (modified after loading)		

Additional Info : Peak(s) manually integrated



=====
Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 C, Sig=210,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.608	MM	0.8916	9031.32617	168.82851	49.6320
2	23.884	BB	0.9673	9165.26660	123.57682	50.3680

Totals : 1.81966e4 292.40533

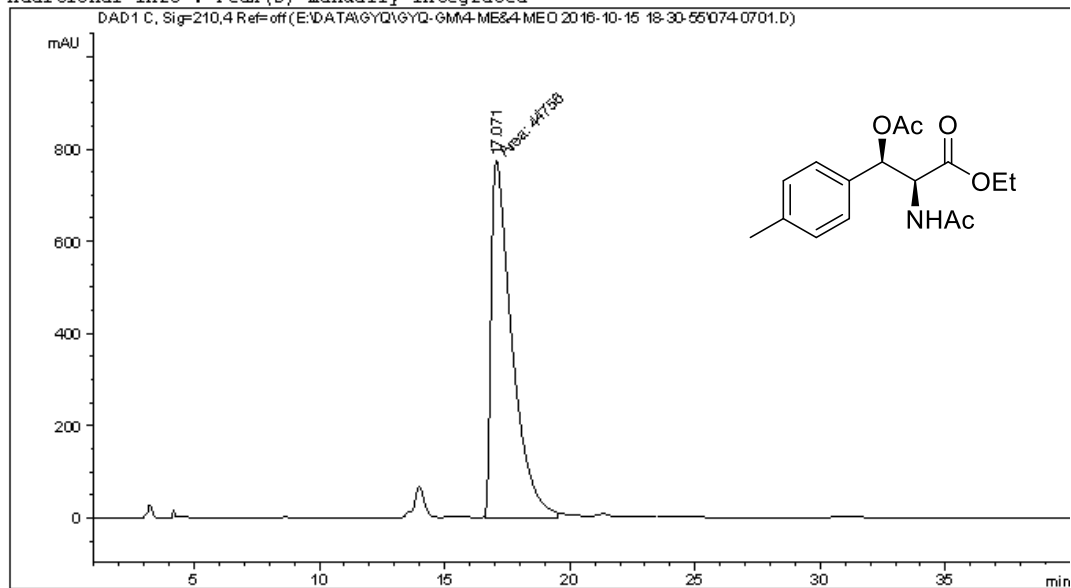
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Data File E:\DATA\GYQ\GYQ-GM\4-ME&4-MEO 2016-10-15 18-30-55\074-0701.D
Sample Name: 4-Me-ee

=====

Acq. Operator	: SYSTEM	Seq. Line	: 7
Acq. Instrument	: 1260HPLC-DAD	Location	: Vial 74
Injection Date	: 10/15/2016 11:24:41 PM	Inj	: 1
		Inj Volume	: 5.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\4-ME&4-MEO 2016-10-15 18-30-55\DAD-0J(1-6)-95-5-1.OML-5-210NM-50MIN.M		
Last changed	: 10/15/2016 6:30:56 PM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\4-ME&4-MEO 2016-10-15 18-30-55\DAD-0J(1-6)-95-5-1.OML-5-210NM-50MIN.M (Sequence Method)		
Last changed	: 6/10/2017 9:12:41 PM by SYSTEM (modified after loading)		

Additional Info : Peak(s) manually integrated



=====
Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 C, Sig=210,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.071	MM	0.9642	4.47560e4	773.66003	100.0000

Totals : 4.47560e4 773.66003

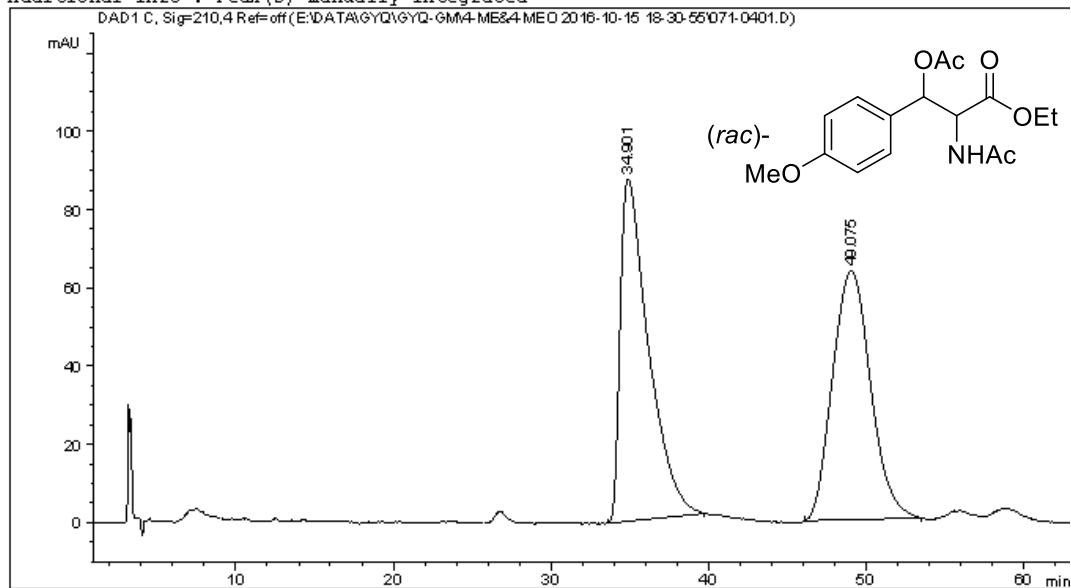
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*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\4-ME&4-MEO 2016-10-15 18-30-55\071-0401.D
Sample Name: 4-MeO-rac

=====

Acq. Operator	: SYSTEM	Seq. Line	: 4
Acq. Instrument	: 1260HPLC-DAD	Location	: Vial 71
Injection Date	: 10/15/2016 9:06:45 PM	Inj	: 1
		Inj Volume	: 10.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\4-ME&4-MEO 2016-10-15 18-30-55\DAD-0J(1-6)-94-6-1.OML -5-210NM-65MIN.M		
Last changed	: 10/15/2016 6:30:55 PM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\4-ME&4-MEO 2016-10-15 18-30-55\DAD-0J(1-6)-94-6-1.OML -5-210NM-65MIN.M (Sequence Method)		
Last changed	: 6/10/2017 9:16:27 PM by SYSTEM (modified after loading)		

Additional Info : Peak(s) manually integrated



=====
Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 C, Sig=210,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	34.901	BB	1.4598	1.09251e4	87.55154	50.5613
2	49.075	BB	1.9706	1.06826e4	63.63345	49.4387

Totals : 2.16077e4 151.18500

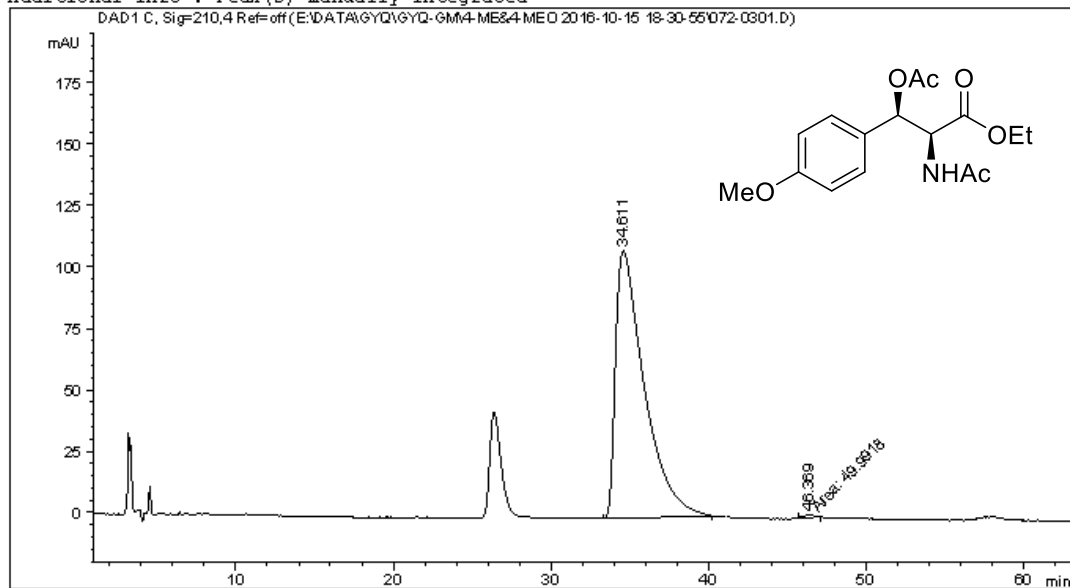
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*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\4-ME&4-MEO 2016-10-15 18-30-55\072-0301.D
Sample Name: 4-MeO-ee

=====

Acq. Operator	: SYSTEM	Seq. Line	: 3
Acq. Instrument	: 1260HPLC-DAD	Location	: Vial 72
Injection Date	: 10/15/2016 8:00:47 PM	Inj	: 1
		Inj Volume	: 10.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\4-ME&4-MEO 2016-10-15 18-30-55\DAD-0J(1-6)-94-6-1.OML-5-210NM-65MIN.M		
Last changed	: 10/15/2016 6:30:55 PM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\4-ME&4-MEO 2016-10-15 18-30-55\DAD-0J(1-6)-94-6-1.OML-5-210NM-65MIN.M (Sequence Method)		
Last changed	: 6/10/2017 9:17:51 PM by SYSTEM (modified after loading)		

Additional Info : Peak(s) manually integrated



=====
Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 C, Sig=210,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	34.611	BB	1.4914	1.37987e4	108.81527	99.6390
2	46.369	MM	0.8820	49.99180	9.44638e-1	0.3610

Totals : 1.38487e4 109.75991

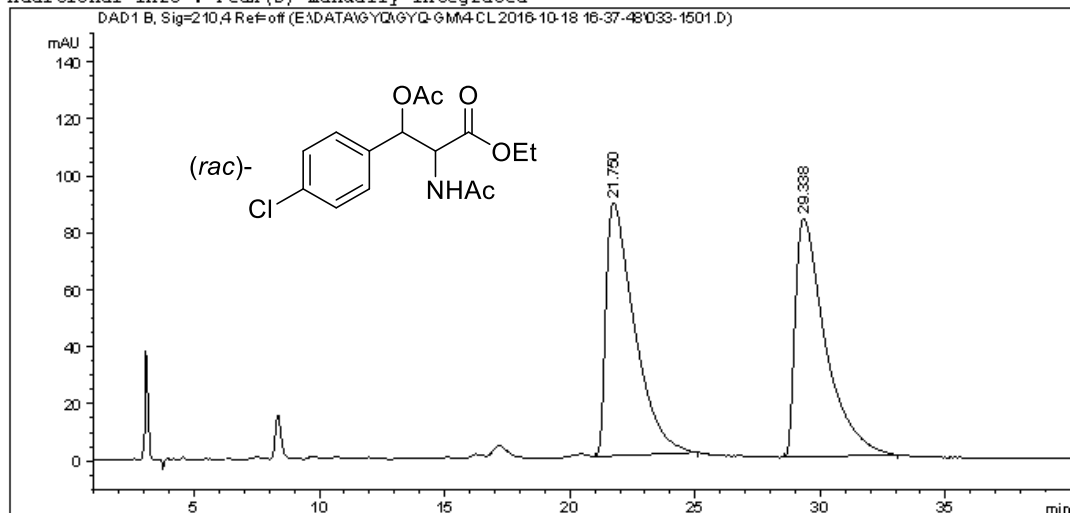
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*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\4-CL 2016-10-18 16-37-48\033-1501.D
Sample Name: 4-Cl-rac

=====

Acq. Operator	: SYSTEM	Seq. Line	: 15
Acq. Instrument	: 1260HPLC-DAD	Location	: Vial 33
Injection Date	: 10/19/2016 12:32:34 AM	Inj	: 1
		Inj Volume	: 5.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\4-CL 2016-10-18 16-37-48\DAD-0D(1-2)-95-5-1ML-50MIN-210.M		
Last changed	: 10/18/2016 4:37:50 PM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\4-CL 2016-10-18 16-37-48\DAD-0D(1-2)-95-5-1ML-50MIN-210.M (Sequence Method)		
Last changed	: 6/10/2017 9:20:10 PM by SYSTEM (modified after loading)		

Additional Info : Peak(s) manually integrated



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Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 B, Sig=210,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	21.750	BB	0.9250	6998.43799	88.84822	49.9765
2	29.338	BB	1.0107	7005.03320	83.84902	50.0235

Totals : 1.40035e4 172.69724

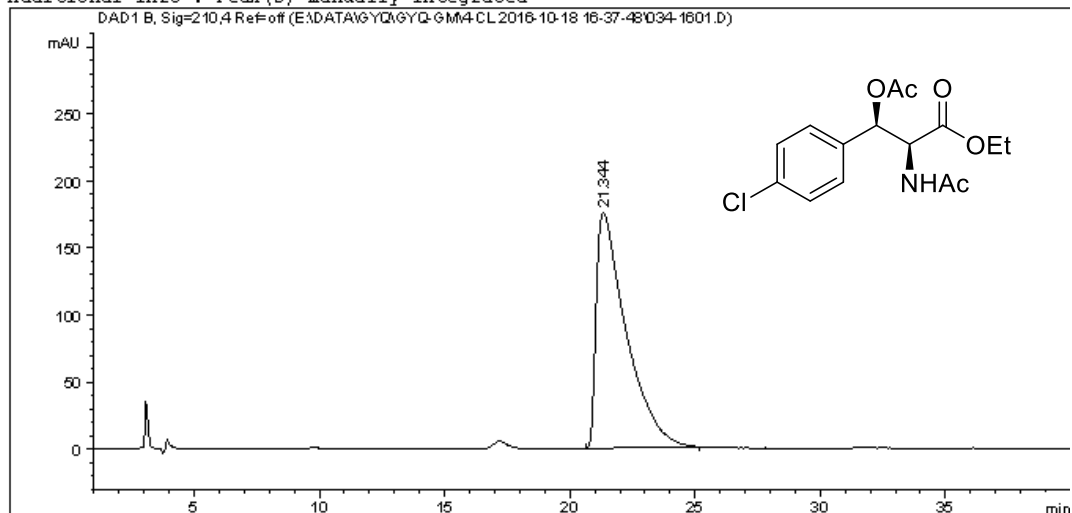
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*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\4-CL 2016-10-18 16-37-48\034-1601.D
Sample Name: 4-CL-ee

=====

Acq. Operator	: SYSTEM	Seq. Line	: 16
Acq. Instrument	: 1260HPLC-DAD	Location	: Vial 34
Injection Date	: 10/19/2016 1:23:28 AM	Inj	: 1
		Inj Volume	: 5.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\4-CL 2016-10-18 16-37-48\DAD-0D(1-2)-95-5-1ML-50MIN-210.M		
Last changed	: 10/18/2016 4:37:50 PM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\4-CL 2016-10-18 16-37-48\DAD-0D(1-2)-95-5-1ML-50MIN-210.M (Sequence Method)		
Last changed	: 6/10/2017 9:22:13 PM by SYSTEM		
	(modified after loading)		
Additional Info : Peak(s) manually integrated			

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Area Percent Report
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Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 B, Sig=210,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	21.344	BB	1.0863	1.46163e4	175.73535	100.0000

Totals : 1.46163e4 175.73535

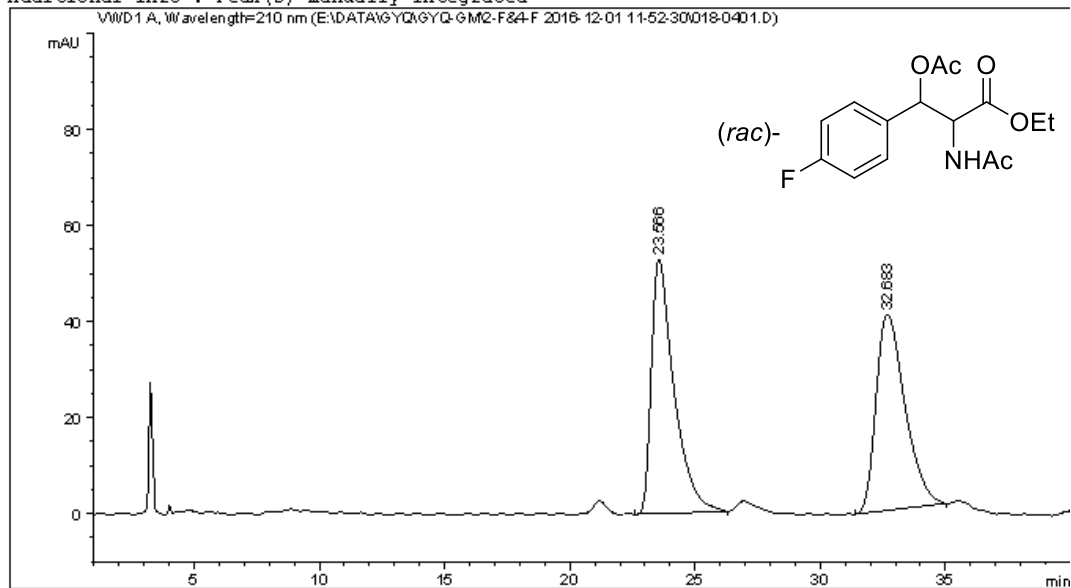
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*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\2-F&4-F 2016-12-01 11-52-30\018-0401.D
Sample Name: 4-F-RAC

=====

Acq. Operator	: SYSTEM	Seq. Line	: 4
Acq. Instrument	: 1260HPLC-VWD	Location	: Vial 18
Injection Date	: 12/1/2016 1:56:34 PM	Inj	: 1
		Inj Volume	: 5.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\2-F&4-F 2016-12-01 11-52-30\VWD-0J(1-2)-95-5-1.0ML-5UL-210NM-40MIN.M		
Last changed	: 12/1/2016 11:52:30 AM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\2-F&4-F 2016-12-01 11-52-30\VWD-0J(1-2)-95-5-1.0ML-5UL-210NM-40MIN.M (Sequence Method)		
Last changed	: 6/10/2017 9:28:49 PM by SYSTEM (modified after loading)		

Additional Info : Peak(s) manually integrated



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Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: VWD1 A, Wavelength=210 nm

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	23.566	BB	0.9158	3328.55859	53.06331	50.3793
2	32.683	BB	1.2065	3278.43726	40.71347	49.6207

Totals : 6606.99585 93.77678

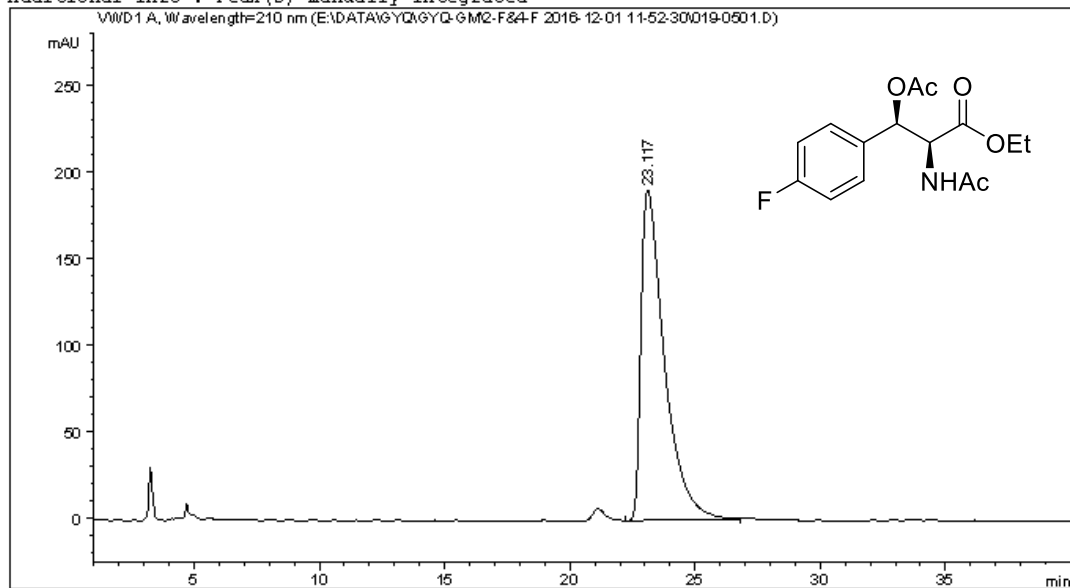
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*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\2-F&4-F 2016-12-01 11-52-30\019-0501.D
Sample Name: 4-F-EE

=====

Acq. Operator	: SYSTEM	Seq. Line	: 5
Acq. Instrument	: 1260HPLC-VWD	Location	: Vial 19
Injection Date	: 12/1/2016 2:37:21 PM	Inj	: 1
		Inj Volume	: 5.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\2-F&4-F 2016-12-01 11-52-30\VWD-0J(1-2)-95-5-1.0ML-5UL-210NM-40MIN.M		
Last changed	: 12/1/2016 11:52:30 AM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\2-F&4-F 2016-12-01 11-52-30\VWD-0J(1-2)-95-5-1.0ML-5UL-210NM-40MIN.M (Sequence Method)		
Last changed	: 6/10/2017 9:30:30 PM by SYSTEM		
	(modified after loading)		
Additional Info : Peak(s) manually integrated			

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Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: VWD1 A, Wavelength=210 nm

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	23.117	BB	0.9716	1.23525e4	190.71989	100.0000

Totals : 1.23525e4 190.71989

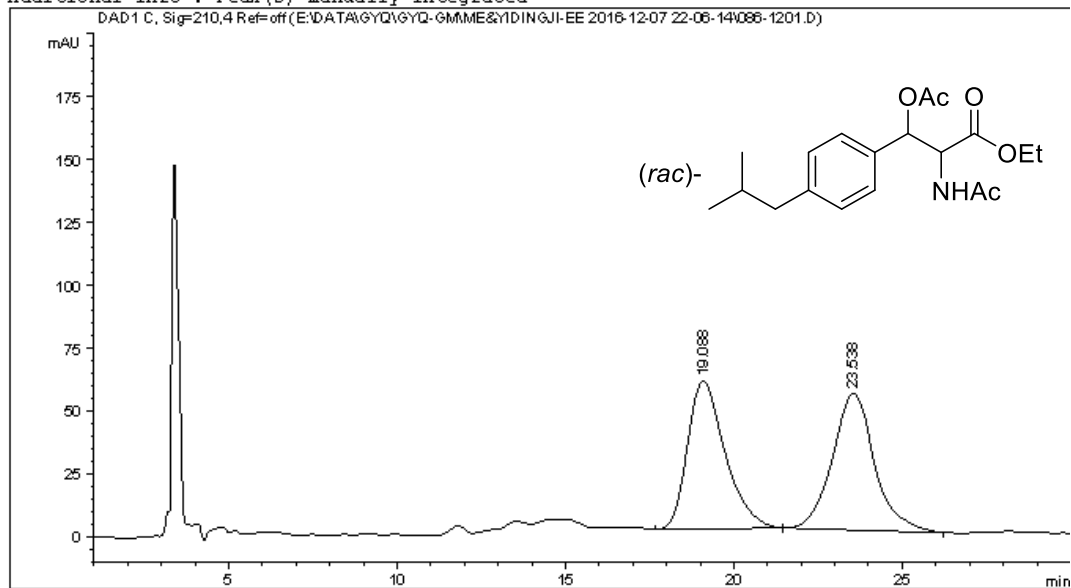
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*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\ME&YIDINGJI-EE 2016-12-07 22-06-14\086-1201.D
Sample Name: yidingji-rac

=====

Acq. Operator	: SYSTEM	Seq. Line	: 12
Acq. Instrument	: 1260HPLC-DAD	Location	: Vial 86
Injection Date	: 12/8/2016 4:19:51 AM	Inj	: 1
		Inj Volume	: 10.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\ME&YIDINGJI-EE 2016-12-07 22-06-14\DAD-0J(1-6)-97-3-1		
	.OML-10-210NM-75MIN.M		
Last changed	: 12/7/2016 10:06:16 PM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\ME&YIDINGJI-EE 2016-12-07 22-06-14\DAD-0J(1-6)-97-3-1		
	.OML-10-210NM-75MIN.M (Sequence Method)		
Last changed	: 6/10/2017 9:45:48 PM by SYSTEM		
	(modified after loading)		
Additional Info	: Peak(s) manually integrated		

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Area Percent Report
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Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 C, Sig=210,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.088	BB	1.1227	4571.60693	58.95211	49.1674
2	23.538	BB	1.1568	4726.43604	54.33568	50.8326

Totals : 9298.04297 113.28780

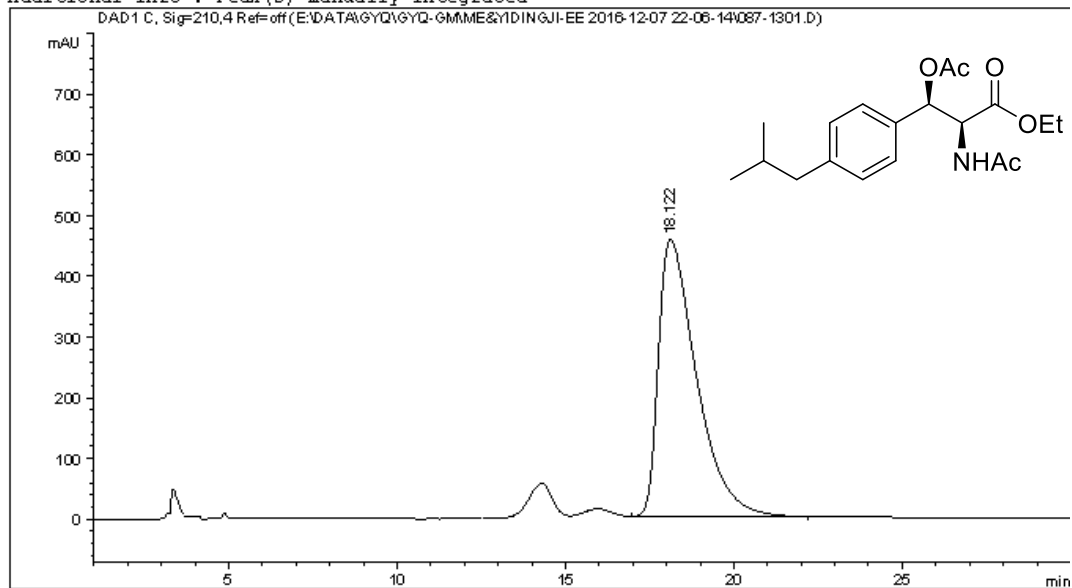
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*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\ME&YIDINGJI-EE 2016-12-07 22-06-14\087-1301.D
Sample Name: yidingji-ee

=====

Acq. Operator	: SYSTEM	Seq. Line	: 13
Acq. Instrument	: 1260HPLC-DAD	Location	: Vial 87
Injection Date	: 12/8/2016 5:35:51 AM	Inj	: 1
		Inj Volume	: 10.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\ME&YIDINGJI-EE 2016-12-07 22-06-14\DAD-0J(1-6)-97-3-1		
	.OML-10-210NM-75MIN.M		
Last changed	: 12/7/2016 10:06:16 PM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\ME&YIDINGJI-EE 2016-12-07 22-06-14\DAD-0J(1-6)-97-3-1		
	.OML-10-210NM-75MIN.M (Sequence Method)		
Last changed	: 6/10/2017 9:47:02 PM by SYSTEM		
	(modified after loading)		
Additional Info : Peak(s) manually integrated			

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Area Percent Report

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Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 C, Sig=210,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	18.122	BB	1.1764	3.58620e4	456.55618	100.0000

Totals : 3.58620e4 456.55618

=====

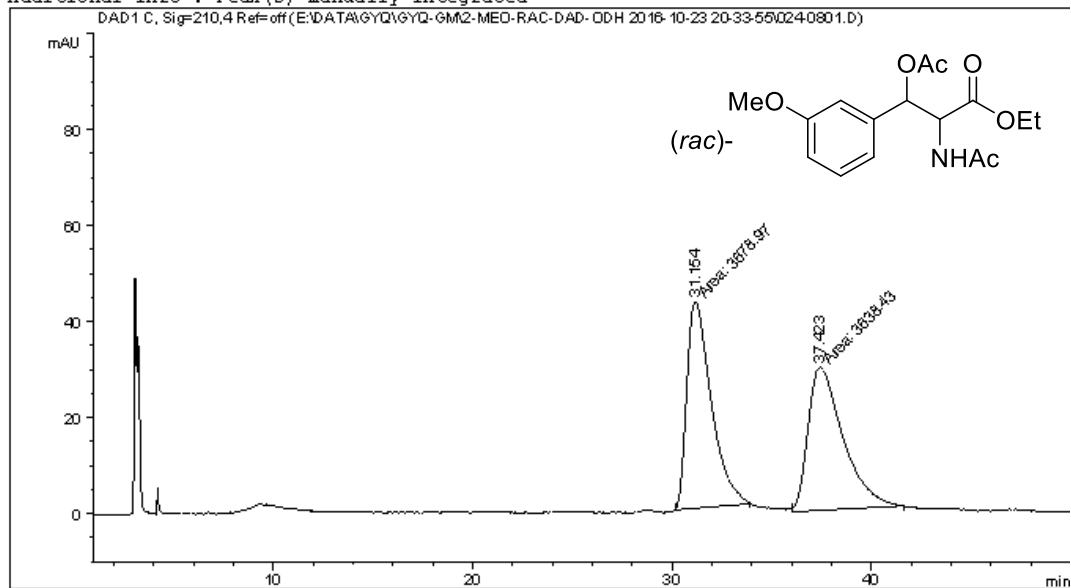
*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\2-MEO-RAC-DAD-ODH 2016-10-23 20-33-55\024-0801.D
Sample Name: 3-MeO-rac

=====

Acq. Operator	: SYSTEM	Seq. Line	: 8
Acq. Instrument	: 1260HPLC-DAD	Location	: Vial 24
Injection Date	: 10/24/2016 12:38:22 AM	Inj	: 1
		Inj Volume	: 5.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\2-MEO-RAC-DAD-ODH 2016-10-23 20-33-55\DAD-0J(1-6)-95-5-1.OML-5-210NM-50MIN.M		
Last changed	: 10/23/2016 10:10:49 PM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\2-MEO-RAC-DAD-ODH 2016-10-23 20-33-55\DAD-0J(1-6)-95-5-1.OML-5-210NM-50MIN.M (Sequence Method)		
Last changed	: 6/10/2017 9:49:21 PM by SYSTEM		
	(modified after loading)		
Additional Info : Peak(s) manually integrated			

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Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 C, Sig=210,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	31.154	MM	1.4238	3678.96729	43.06536	50.2770
2	37.423	MM	2.0392	3638.43311	29.73787	49.7230

Totals : 7317.40039 72.80323

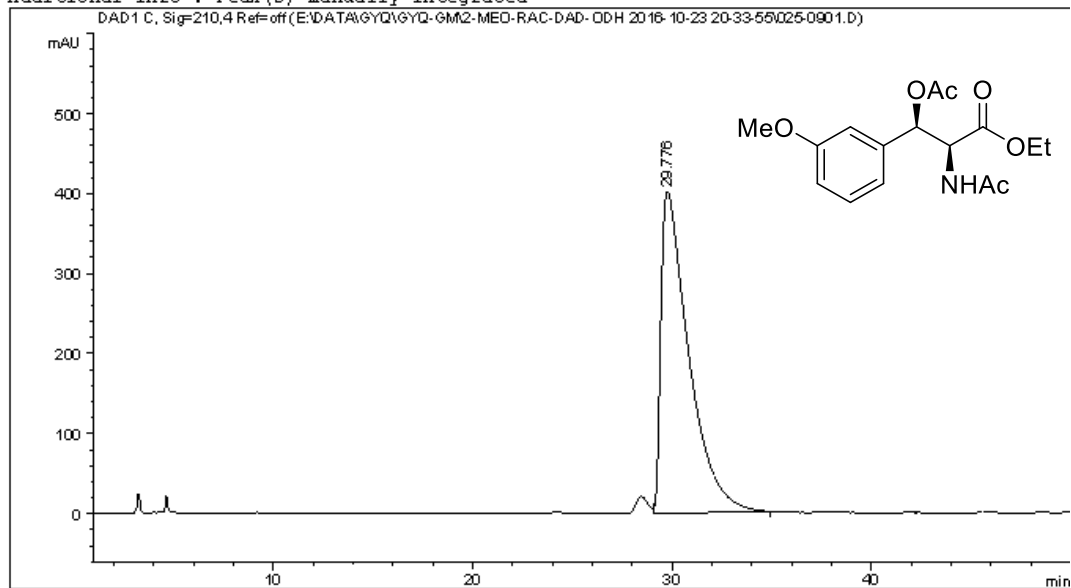
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*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\2-MEO-RAC-DAD-ODH 2016-10-23 20-33-55\025-0901.D
Sample Name: 3-MeO-ee

=====

Acq. Operator	: SYSTEM	Seq. Line	: 9
Acq. Instrument	: 1260HPLC-DAD	Location	: Vial 25
Injection Date	: 10/24/2016 1:29:17 AM	Inj	: 1
		Inj Volume	: 5.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\2-MEO-RAC-DAD-ODH 2016-10-23 20-33-55\DAD-0J(1-6)-95-5-1.OML-5-210NM-50MIN.M		
Last changed	: 10/23/2016 10:10:49 PM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\2-MEO-RAC-DAD-ODH 2016-10-23 20-33-55\DAD-0J(1-6)-95-5-1.OML-5-210NM-50MIN.M (Sequence Method)		
Last changed	: 6/10/2017 9:50:49 PM by SYSTEM		
	(modified after loading)		
Additional Info : Peak(s) manually integrated			

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Area Percent Report

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Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 C, Sig=210,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	29.776	VB	1.2468	3.88495e4	401.51578	100.0000

Totals : 3.88495e4 401.51578

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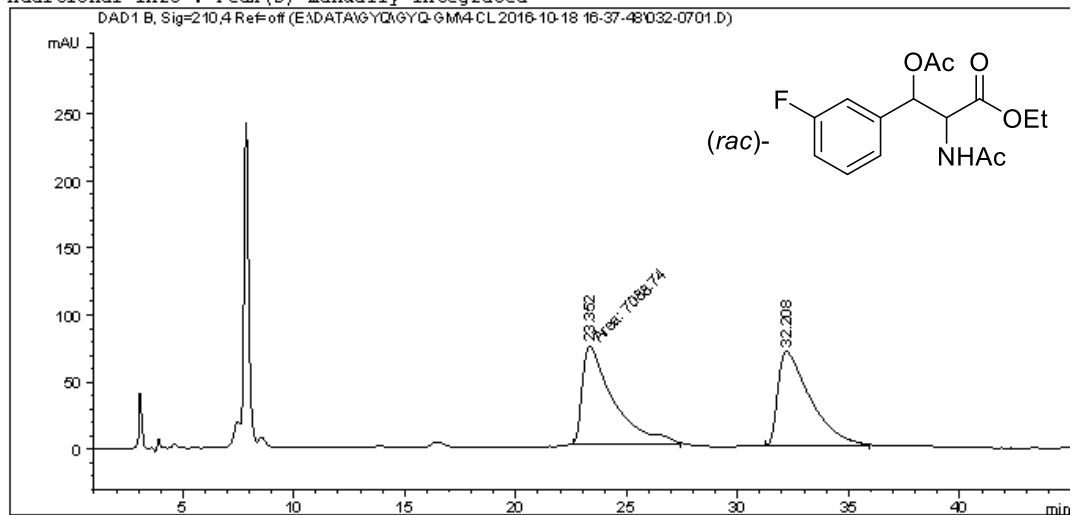
*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\4-CL 2016-10-18 16-37-48\032-0701.D
Sample Name: 3-F-rac

=====

Acq. Operator	: SYSTEM	Seq. Line	: 7
Acq. Instrument	: 1260HPLC-DAD	Location	: Vial 32
Injection Date	: 10/18/2016 8:14:45 PM	Inj	: 1
		Inj Volume	: 5.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\4-CL 2016-10-18 16-37-48\DAD-0D(1-2)-95-5-1ML-50MIN-210.M		
Last changed	: 10/18/2016 4:37:50 PM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\4-CL 2016-10-18 16-37-48\DAD-0D(1-2)-95-5-1ML-50MIN-210.M (Sequence Method)		
Last changed	: 6/10/2017 9:55:48 PM by SYSTEM		
	(modified after loading)		
Additional Info : Peak(s) manually integrated			

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Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 B, Sig=210,4 Ref=off

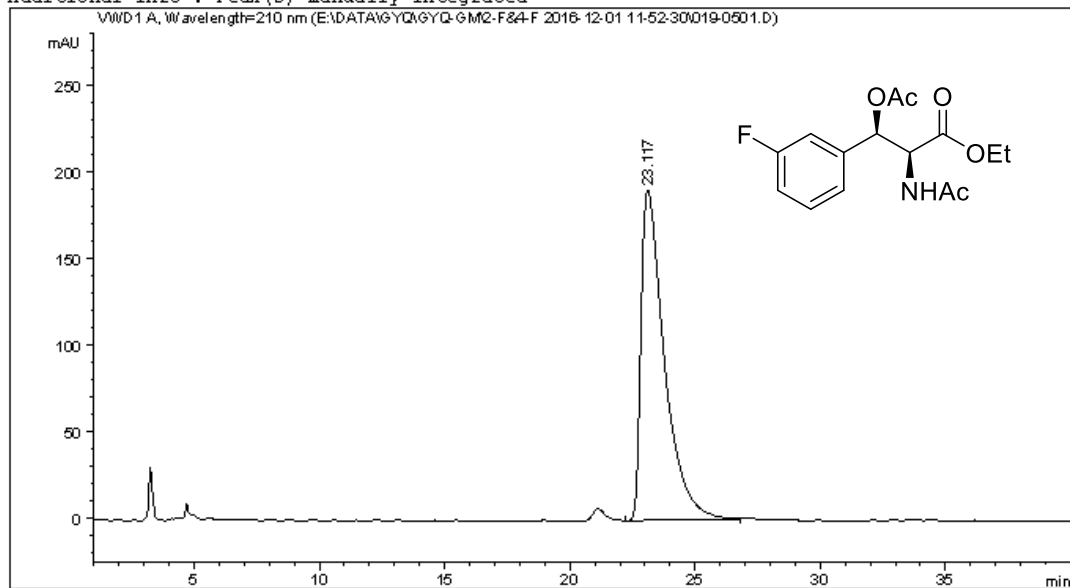
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	23.352	MM	1.6241	7088.74121	72.74491	50.2422
2	32.208	BB	1.1787	7020.39209	69.84410	49.7578

Totals : 1.41091e4 142.58901

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*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\2-F&4-F 2016-12-01 11-52-30\019-0501.D
Sample Name: 4-F-EE

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=====
Acq. Operator   : SYSTEM                      Seq. Line :    5
Acq. Instrument : 1260HPLC-VWD                Location  : Vial 19
Injection Date  : 12/1/2016 2:37:21 PM         Inj       :    1
                                           Inj Volume: 5.000 µl
Acq. Method     : E:\DATA\GYQ\GYQ-GM\2-F&4-F 2016-12-01 11-52-30\VWD-0J(1-2)-95-5-1.0ML-
                                           SUL-210NM-40MIN.M
Last changed    : 12/1/2016 11:52:30 AM by SYSTEM
Analysis Method : E:\DATA\GYQ\GYQ-GM\2-F&4-F 2016-12-01 11-52-30\VWD-0J(1-2)-95-5-1.0ML-
                                           SUL-210NM-40MIN.M (Sequence Method)
Last changed    : 6/10/2017 9:30:30 PM by SYSTEM
                                           (modified after loading)
Additional Info  : Peak(s) manually integrated
=====
```



=====
Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: VWD1 A, Wavelength=210 nm

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	23.117	BB	0.9716	1.23525e4	190.71989	100.0000

Totals : 1.23525e4 190.71989

=====
*** End of Report ***

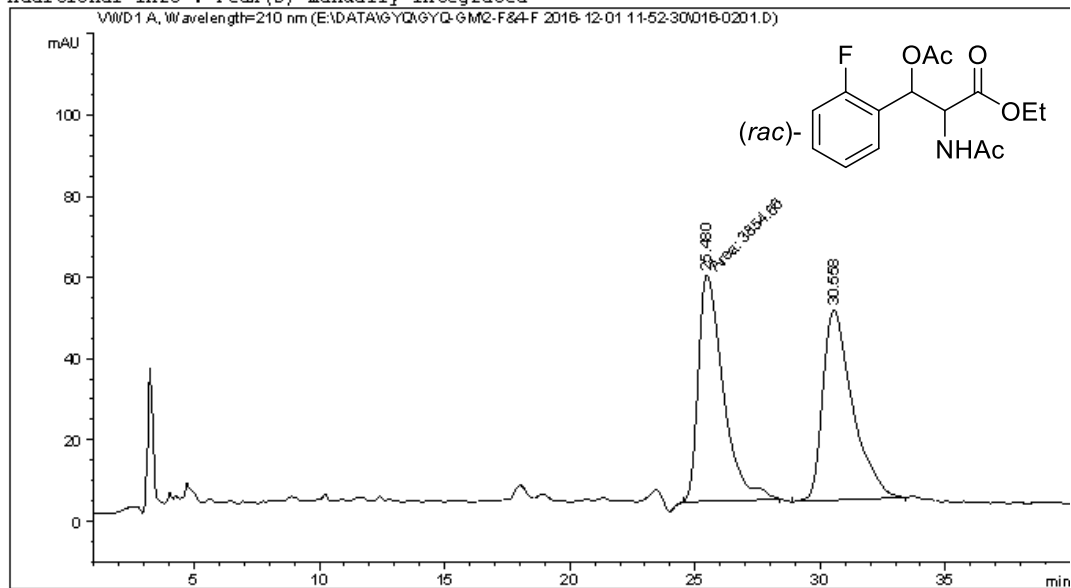
Data File E:\DATA\GYQ\GYQ-GM\2-F&4-F 2016-12-01 11-52-30\016-0201.D
Sample Name: 2-F-RAC

=====

Acq. Operator	: SYSTEM	Seq. Line	: 2
Acq. Instrument	: 1260HPLC-VWD	Location	: Vial 16
Injection Date	: 12/1/2016 12:15:02 PM	Inj	: 1
		Inj Volume	: 5.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\2-F&4-F 2016-12-01 11-52-30\VWD-0J(1-2)-95-5-1.0ML-3UL-210NM-60MIN.M		
Last changed	: 12/1/2016 11:52:30 AM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\2-F&4-F 2016-12-01 11-52-30\VWD-0J(1-2)-95-5-1.0ML-3UL-210NM-60MIN.M (Sequence Method)		
Last changed	: 6/10/2017 9:33:13 PM by SYSTEM (modified after loading)		

Additional Info : Peak(s) manually integrated

=====



=====
Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: VWD1 A, Wavelength=210 nm

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	25.480	MM	1.1541	3854.65820	55.66693	49.8791
2	30.558	BB	1.2099	3873.35205	46.73795	50.1209

Totals : 7728.01025 102.40487

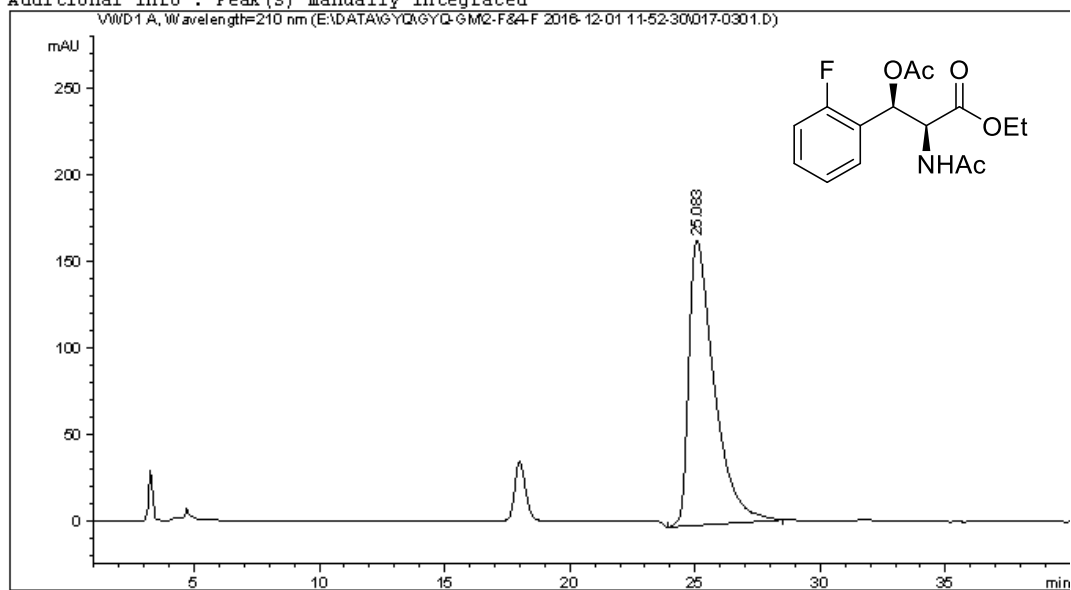
=====
*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\2-F&4-F 2016-12-01 11-52-30\017-0301.D
Sample Name: 2-F-EE

=====

Acq. Operator	: SYSTEM	Seq. Line	: 3
Acq. Instrument	: 1260HPLC-VWD	Location	: Vial 17
Injection Date	: 12/1/2016 1:15:49 PM	Inj	: 1
		Inj Volume	: 5.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\2-F&4-F 2016-12-01 11-52-30\VWD-0J(1-2)-95-5-1.0ML-5UL-210NM-40MIN.M		
Last changed	: 12/1/2016 11:52:30 AM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\2-F&4-F 2016-12-01 11-52-30\VWD-0J(1-2)-95-5-1.0ML-5UL-210NM-40MIN.M (Sequence Method)		
Last changed	: 6/10/2017 9:32:28 PM by SYSTEM		
Additional Info	: Peak(s) manually integrated		

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=====
Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: VWD1 A, Wavelength=210 nm

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	25.083	BB	1.0454	1.16220e4	164.27574	100.0000

Totals : 1.16220e4 164.27574

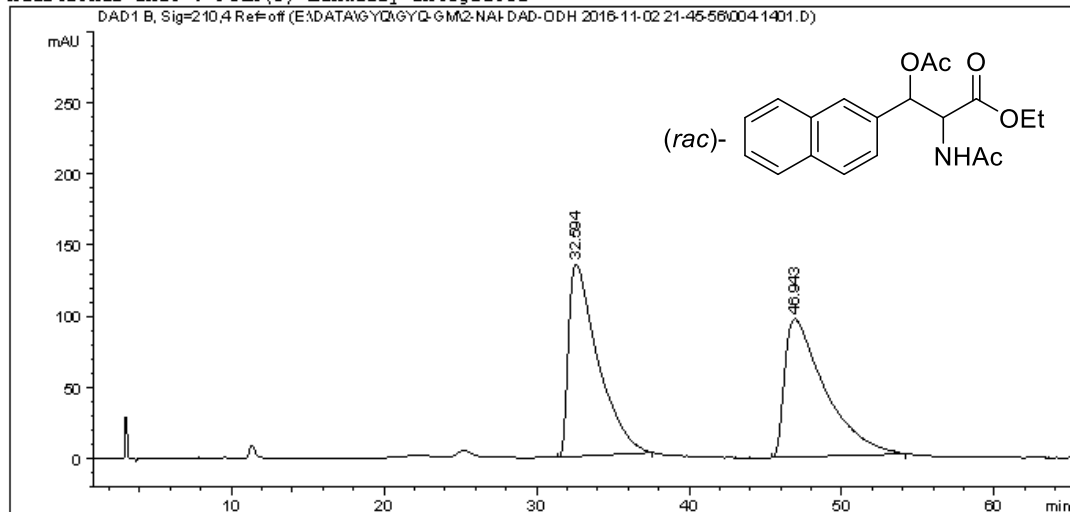
=====
*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\2-NAI-DAD-ODH 2016-11-02 21-45-56\004-1401.D
Sample Name: 2-nai-rac

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=====
Acq. Operator   : SYSTEM                      Seq. Line :   14
Acq. Instrument : 1260HPLC-DAD                Location  : Vial 4
Injection Date  : 11/3/2016 2:58:41 PM        Inj       :    1
                                           Inj Volume: 5.000 µl
Acq. Method     : E:\DATA\GYQ\GYQ-GM\2-NAI-DAD-ODH 2016-11-02 21-45-56\DAD-OD(1-2)-95-5-
                                           1ML-75MIN-210-254.M
Last changed    : 11/3/2016 4:09:32 PM by SYSTEM
                                           (modified after loading)
Analysis Method : E:\DATA\GYQ\GYQ-GM\2-NAI-DAD-ODH 2016-11-02 21-45-56\DAD-OD(1-2)-95-5-
                                           1ML-75MIN-210-254.M (Sequence Method)
Last changed    : 6/10/2017 10:03:39 PM by SYSTEM
                                           (modified after loading)
Additional Info : Peak(s) manually integrated
=====

```



Area Percent Report

```

Sorted By       : Signal
Multiplier      : 1.0000
Dilution        : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

```

Signal 1: DAD1 B, Sig=210,4 Ref=off

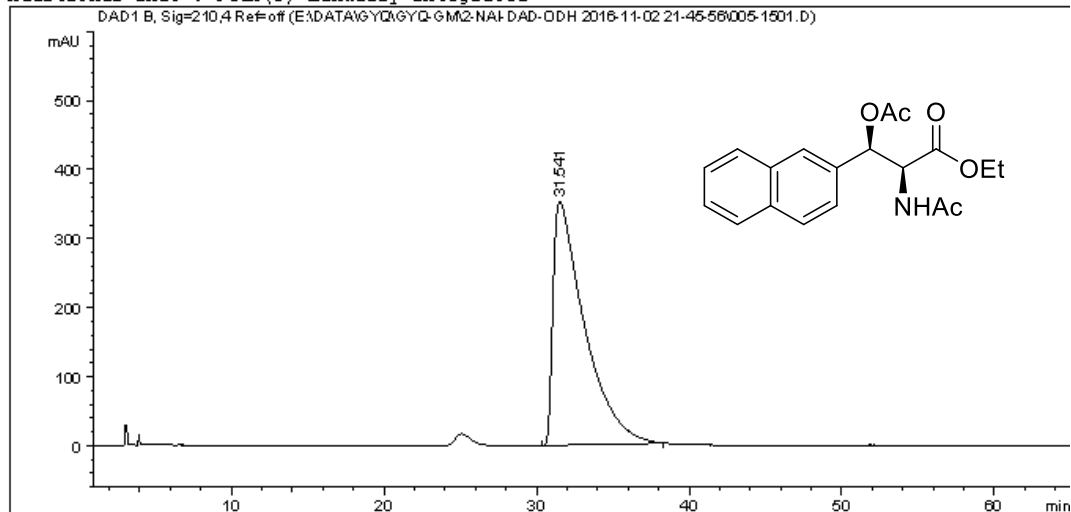
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	32.594	BB	1.5638	1.80568e4	135.23212	50.3901
2	46.943	BB	2.1466	1.77773e4	96.89047	49.6099

Totals : 3.58341e4 232.12259

*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\2-NAI-DAD-ODH 2016-11-02 21-45-56\005-1501.D
Sample Name: 2-nai-ee

```
=====
Acq. Operator   : SYSTEM                      Seq. Line :   15
Acq. Instrument : 1260HPLC-DAD                Location  : Vial 5
Injection Date  : 11/3/2016 4:10:27 PM        Inj       :    1
                                           Inj Volume: 5.000 µl
Acq. Method     : E:\DATA\GYQ\GYQ-GM\2-NAI-DAD-ODH 2016-11-02 21-45-56\
DAD-OD(1-2)-95-5-1ML-75MIN-210-254.M
Last changed    : 11/3/2016 5:15:06 PM by SYSTEM
                  (modified after loading)
Analysis Method : E:\DATA\GYQ\GYQ-GM\2-NAI-DAD-ODH 2016-11-02 21-45-56\
DAD-OD(1-2)-95-5-1ML-75MIN-210-254.M (Sequence Method)
Last changed    : 6/10/2017 10:04:50 PM by SYSTEM
                  (modified after loading)
Additional Info : Peak(s) manually integrated
=====
```



=====
Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 B, Sig=210,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	31.541	BB	1.6780	5.03139e4	352.76291	100.0000

Totals : 5.03139e4 352.76291

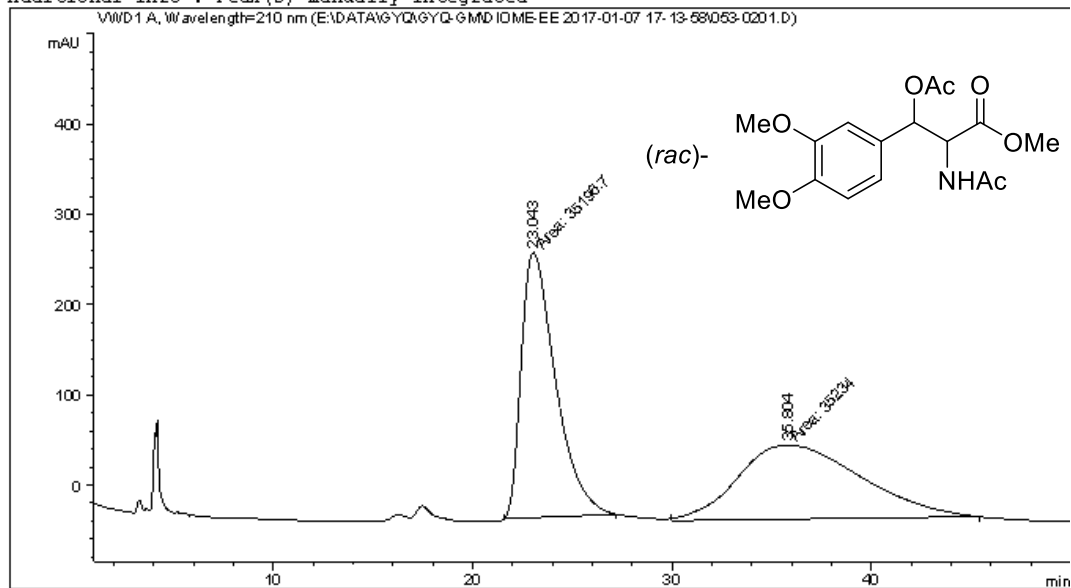
=====
*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\DIOME-EE 2017-01-07 17-13-58\053-0201.D
Sample Name: GYQ-DiOMe-rac

=====

Acq. Operator	: SYSTEM	Seq. Line	: 2
Acq. Instrument	: 1260HPLC-VWD	Location	: Vial 53
Injection Date	: 1/7/2017 5:30:36 PM	Inj	: 1
		Inj Volume	: 5.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\DIOME-EE 2017-01-07 17-13-58\VWD-0J(1-6)-80-20-1.OML-210NM-50MIN.M		
Last changed	: 1/7/2017 5:13:58 PM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\DIOME-EE 2017-01-07 17-13-58\VWD-0J(1-6)-80-20-1.OML-210NM-50MIN.M (Sequence Method)		
Last changed	: 6/10/2017 10:06:45 PM by SYSTEM (modified after loading)		

Additional Info : Peak(s) manually integrated



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Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: VWD1 A, Wavelength=210 nm

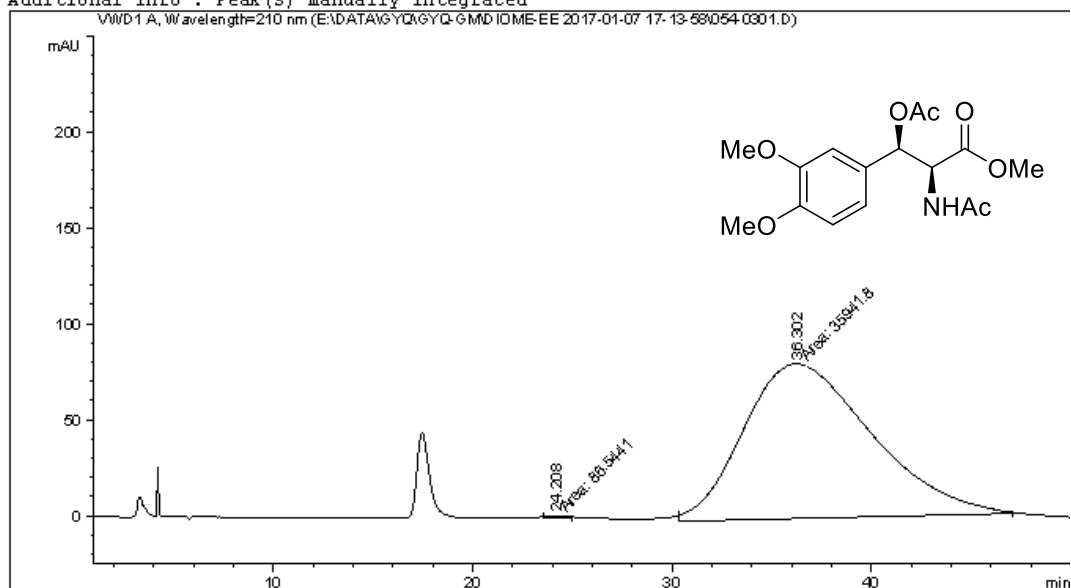
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	23.043	MM	1.9981	3.51967e4	293.58615	49.9735
2	35.804	MM	7.1639	3.52340e4	81.97152	50.0265

Totals : 7.04308e4 375.55767

=====
*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\DIOME-EE 2017-01-07 17-13-58\054-0301.D
Sample Name: GYQ-DiOMe-ee

```
=====
Acq. Operator   : SYSTEM                      Seq. Line :    3
Acq. Instrument : 1260HPLC-VWD                Location  : Vial 54
Injection Date  : 1/7/2017 6:21:24 PM          Inj       :    1
                                           Inj Volume: 5.000 µl
Acq. Method     : E:\DATA\GYQ\GYQ-GM\DIOME-EE 2017-01-07 17-13-58\VWD-0J(1-6)-80-20-1.OML-
210NM-50MIN.M
Last changed    : 1/7/2017 5:13:58 PM by SYSTEM
Analysis Method : E:\DATA\GYQ\GYQ-GM\DIOME-EE 2017-01-07 17-13-58\VWD-0J(1-6)-80-20-1.OML-
210NM-50MIN.M (Sequence Method)
Last changed    : 6/10/2017 10:11:50 PM by SYSTEM
Additional Info : Peak(s) manually integrated
=====
```



Area Percent Report

```
Sorted By      : Signal
Multiplier     : 1.0000
Dilution      : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs
```

Signal 1: VWD1 A, Wavelength=210 nm

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	24.208	MM	1.1890	86.54407	1.21313	0.2402
2	36.302	MM	7.4595	3.59418e4	80.30405	99.7598

Totals : 3.60284e4 81.51717

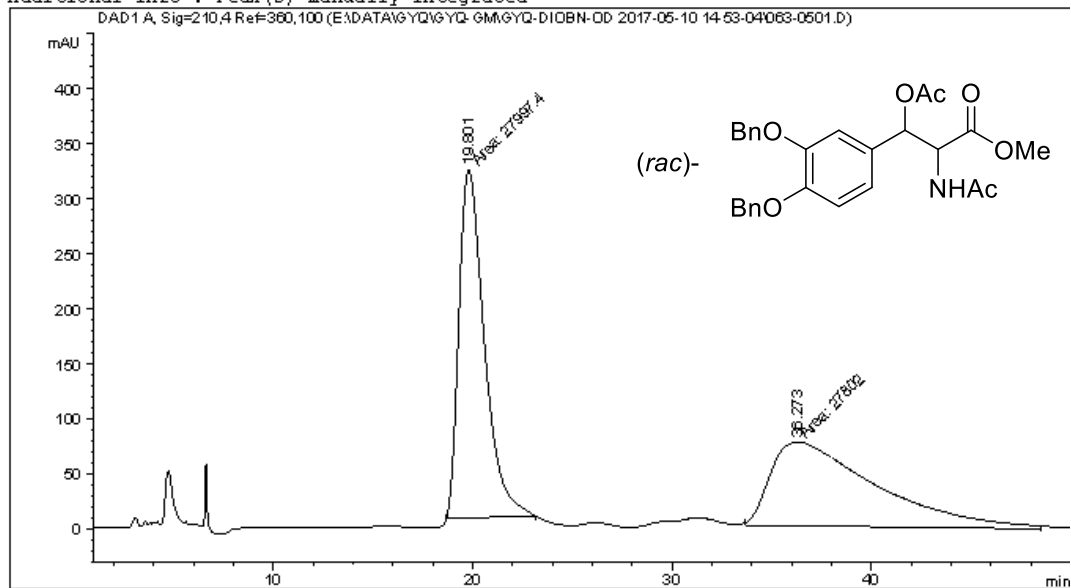
*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\GYQ-DIOBN-OD 2017-05-10 14-53-04\063-0501.D
Sample Name: GYQ-DIOBN-RAC

=====

Acq. Operator	: SYSTEM	Seq. Line	: 5
Acq. Instrument	: 1260HPLC-DAD	Location	: Vial 63
Injection Date	: 5/10/2017 5:47:55 PM	Inj	: 1
		Inj Volume	: 5.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\GYQ-DIOBN-OD 2017-05-10 14-53-04\DAD-OD (1-2)-80-20-1ML-5UL- ALL-65MIN.M		
Last changed	: 5/10/2017 5:08:19 PM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\GYQ-DIOBN-OD 2017-05-10 14-53-04\DAD-OD (1-2)-80-20-1ML-5UL- ALL-65MIN.M (Sequence Method)		
Last changed	: 6/11/2017 8:08:23 AM by SYSTEM		
	(modified after loading)		

Additional Info : Peak(s) manually integrated



=====
Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=210,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.801	MM	1.4744	2.79974e4	316.48984	50.1751
2	36.273	MM	6.0796	2.78020e4	76.21697	49.8249

Totals : 5.57993e4 392.70681

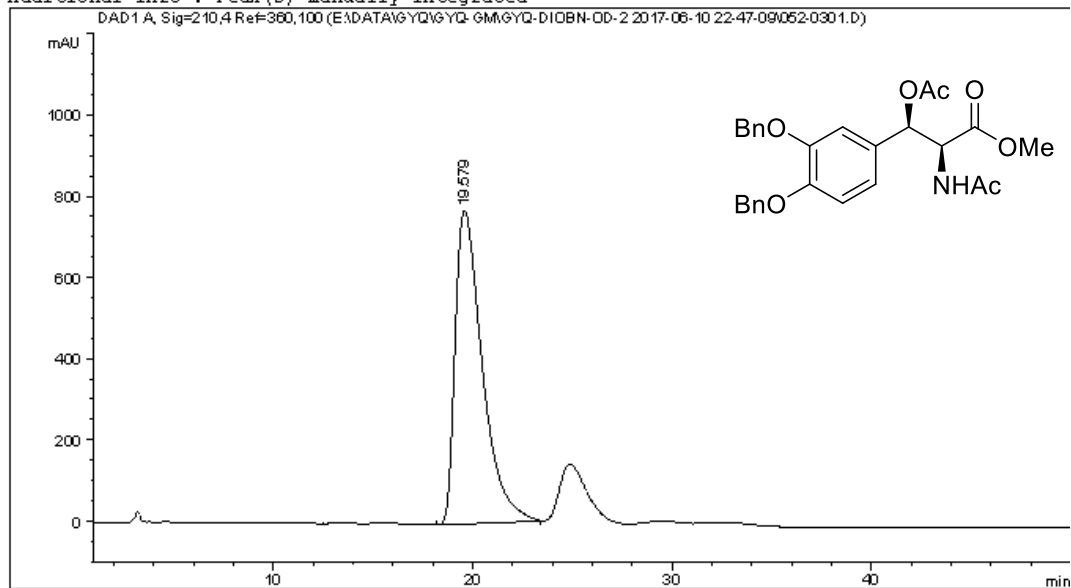
=====
*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\GYQ-DIOBN-OD-2 2017-06-10 22-47-09\052-0301.D
Sample Name: GYQ-DIOBN-EE

=====

Acq. Operator	: SYSTEM	Seq. Line	: 3
Acq. Instrument	: 1260HPLC-DAD	Location	: Vial 52
Injection Date	: 6/11/2017 12:04:17 AM	Inj	: 1
		Inj Volume	: 5.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\GYQ-DIOBN-OD-2 2017-06-10 22-47-09\DAD-OD(1-2)-80-20-1ML-5UL- ALL-55MIN.M		
Last changed	: 6/10/2017 10:47:10 PM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\GYQ-DIOBN-OD-2 2017-06-10 22-47-09\DAD-OD(1-2)-80-20-1ML-5UL- ALL-55MIN.M (Sequence Method)		
Last changed	: 6/11/2017 8:03:37 AM by SYSTEM (modified after loading)		

Additional Info : Peak(s) manually integrated



=====
Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=210,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.579	BV	1.1123	7.29634e4	770.80939	100.0000

Totals : 7.29634e4 770.80939

=====
*** End of Report ***



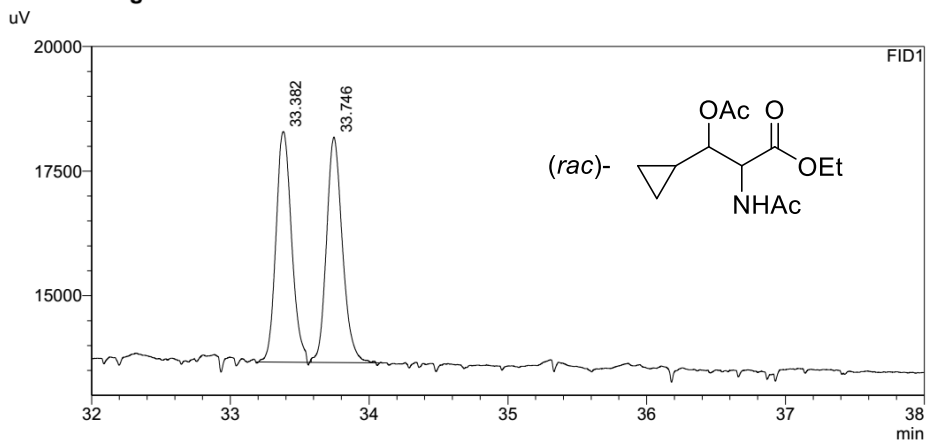
Analysis Report

<Sample Information>

Sample Name : gyq-cyclo-rac
 Sample ID : L/RhRu,2/2,0.15,i-Pr/MeOH
 Data Filename : gyq-cyclo-rac-1.gcd
 Method Filename : gamma dex-225-250-100(0)-2-160(20)-260-50min.gcm
 Batch Filename : gyq-cyclo.gcb
 Vial # : 1
 Injection Volume : 1 uL
 Date Acquired : 2016-11-1 16:55:03
 Date Processed : 2016-11-1 19:32:58

Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	33.382	36228	4620	50.079		M	
2	33.746	36114	4519	49.921		V M	
Total		72342	9139				

D:\DATA FILE\gyq\data\gyq-cyclo\gyq-cyclo-rac-1.gcd



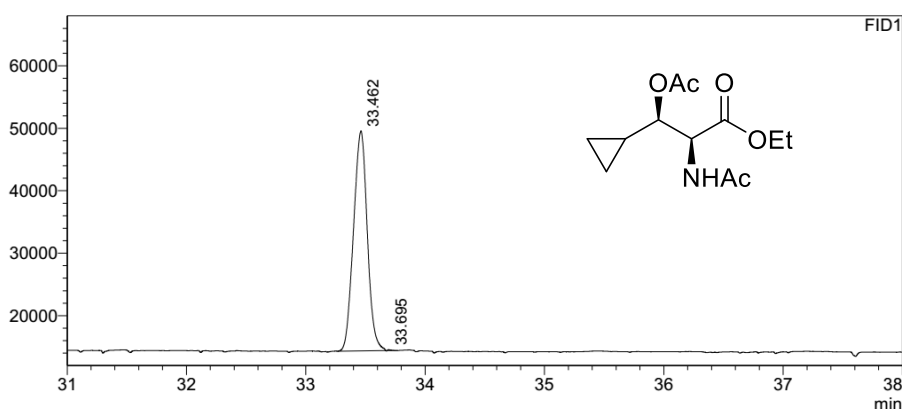
Analysis Report

<Sample Information>

Sample Name : gyq-cyclo-ee
Sample ID : L/Rh,2/2,0.15,i-Pr/TFE
Data Filename : gyq-cyclo-ee.gcd
Method Filename : gamma dex-225-250-100(0)-2-160(20)-260-50min.gcm
Batch Filename : gyq-cyclo-ee.gcb
Vial # : 2
Injection Volume : 1 uL
Date Acquired : 2016-11-2 16:47:55
Date Processed : 2016-11-3 11:26:04
Sample Type : Unknown
Acquired by : System Administrator
Processed by : System Administrator

<Chromatogram>

uV



<Peak Table>

FID1

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	33.462	282372	35203	99.940		M	
2	33.695	170	125	0.060		M	
Total		282542	35327				

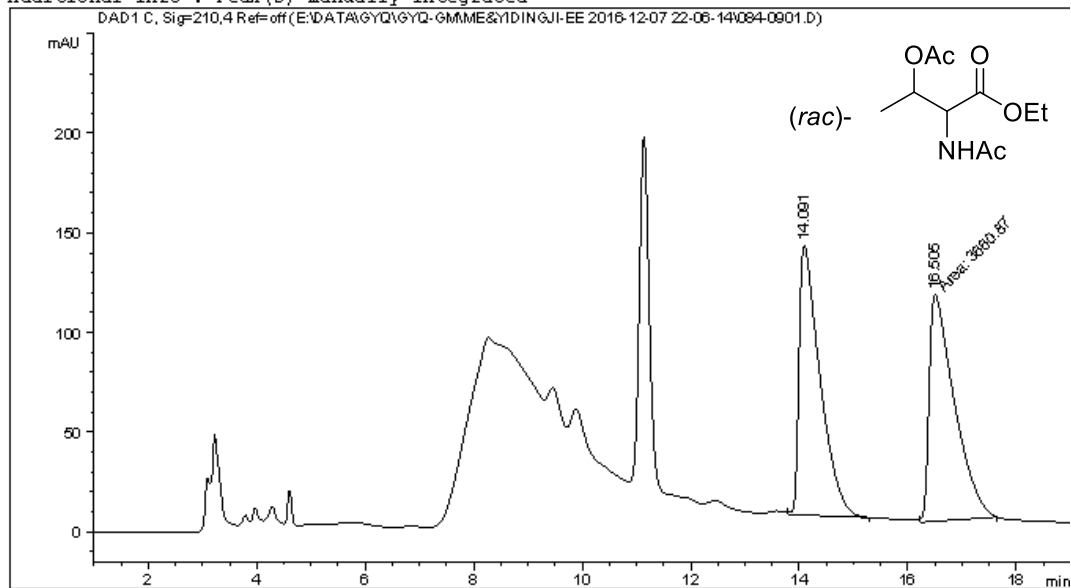
D:\DATA FILE\gyq\data\gyq-cyclo\gyq-cyclo-ee.gcd

Data File E:\DATA\GYQ\GYQ-GM\ME&YIDINGJI-EE 2016-12-07 22-06-14\084-0901.D
Sample Name: Me-rac

=====

Acq. Operator	: SYSTEM	Seq. Line	: 9
Acq. Instrument	: 1260HPLC-DAD	Location	: Vial 84
Injection Date	: 12/8/2016 2:16:41 AM	Inj	: 1
		Inj Volume	: 5.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\ME&YIDINGJI-EE 2016-12-07 22-06-14\DAD-0J(1-6)-95-5-1		
	.OML-5-210NM-50MIN.M		
Last changed	: 12/7/2016 10:06:16 PM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\ME&YIDINGJI-EE 2016-12-07 22-06-14\DAD-0J(1-6)-95-5-1		
	.OML-5-210NM-50MIN.M (Sequence Method)		
Last changed	: 6/10/2017 9:41:25 PM by SYSTEM		
	(modified after loading)		
Additional Info	: Peak(s) manually integrated		

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Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 C, Sig=210,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.091	VB	0.3868	3606.98047	135.41585	49.6293
2	16.505	MM	0.5364	3660.87134	113.74592	50.3707

Totals : 7267.85181 249.16177

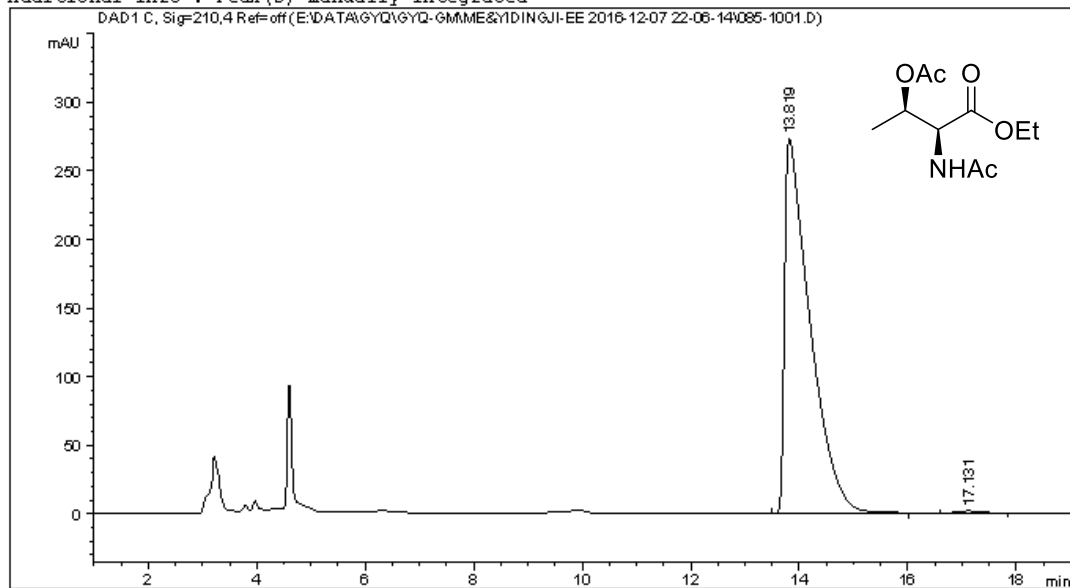
=====
*** End of Report ***

Data File E:\DATA\GYQ\GYQ-GM\ME&YIDINGJI-EE 2016-12-07 22-06-14\085-1001.D
Sample Name: Me-EE

=====

Acq. Operator	: SYSTEM	Seq. Line	: 10
Acq. Instrument	: 1260HPLC-DAD	Location	: Vial 85
Injection Date	: 12/8/2016 3:07:45 AM	Inj	: 1
		Inj Volume	: 5.000 µl
Acq. Method	: E:\DATA\GYQ\GYQ-GM\ME&YIDINGJI-EE 2016-12-07 22-06-14\DAD-0J(1-6)-95-5-1		
	.OML-5-210NM-50MIN.M		
Last changed	: 12/7/2016 10:06:16 PM by SYSTEM		
Analysis Method	: E:\DATA\GYQ\GYQ-GM\ME&YIDINGJI-EE 2016-12-07 22-06-14\DAD-0J(1-6)-95-5-1		
	.OML-5-210NM-50MIN.M (Sequence Method)		
Last changed	: 6/10/2017 9:43:20 PM by SYSTEM		
	(modified after loading)		
Additional Info : Peak(s) manually integrated			

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=====
Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 C, Sig=210,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.819	BB	0.4619	8835.72461	272.76071	99.5346
2	17.131	BB	0.3341	41.31002	1.49612	0.4654

Totals : 8877.03463 274.25683

=====
*** End of Report ***