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Supporting Information

B(C₆F₅)₃-promoted hydrogenations of N-heterocycles with ammonia borane

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1. General Information

Procedure for the synthesis of **10-1u**, please see article.^[31] Unless otherwise noted, otherwise noted, reagents were obtained from commercial suppliers and were used without further purification. ¹H NMR (400 MHz or 600 MHz) and ¹³C NMR (150 MHz) spectra were obtained on Bruker 400 M or 600 M nuclear resonance spectrometers. Chemical shifts are reported in ppm from tetramethylsilane with the solvent resonance as the internal standard. The following abbreviations were used to designate chemical shift multiplicities: s = singlet, d = doublet, t = triplet, m = multiplet. Flash column chromatography was performed using 200-300 mesh silica gel.

General Procedure for the Hydrogenations of N-Heterocycles: To a mixture of an appropriate N-heterocycles (0.5 mmol) and $B(C_6F_5)_3$ (5 mol %) in a standard Schlenk tube, 1.5 mmol of ammonia borane for quinoline derivatives and quinoxaline derivatives was added. The mixture was evacuated and backfilled with N₂ (3 times). Then, the solvent (1.5 mL) was added via syringe, and the mixture was stirred at 80 °C for 8 h. The resulting reaction mixture was concentrated under reduced pressure and purified by column chromatography using petroleum ether/ethyl acetate to afford the corresponding products.

General Procedure for the Asymmetric Hydrogenations of N-Heterocycles: To a mixture of an appropriate N-heterocycles (0.1 mmol), $B(C_6F_5)_3$ (5 mol%), PA (10 mol%) and additive (0.1 mmol) in a standard Schlenk tube, 0.3 mmol of ammonia borane for quinoline derivatives was added. The mixture was evacuated and backfilled with N₂ (3 times). Then, the solvent (1.5 mL) was added via syringe, and the mixture was stirred at 80 °C for 8 h. The resulting reaction mixture was concentrated under reduced pressure and purified by column chromatography using petroleum ether/ethyl acetate to afford the corresponding products.

Ĺ		$\begin{array}{c} B(C_6F_5)_3 \\ \underline{NH_3\cdotBH_3} \\ \text{solvents, 80} \\ \text{cat. add} \end{array}$	5 mol% 3 <u>3 eq</u> () °C, 24 h ditive		Ph
Entry	Cat. 5 mol%	Solvent	Additive 5 mol%	Yield (%)	ee (%) ^b
1	S1a	toluene	-	65	0
2	S1b	toluene	-	63	0
3	S1c	toluene	-	66	12
4	S1d	toluene	-	50	13
5	S1e	toluene	-	55	0
6	quinine	toluene	-	57	0
7	chinchonine	toluene	-	62	3
8	S1c	DCE	-	70	5
9	S1c	DCM	-	71	10
10	S1c	THF	-	66	3
11	S1c	toluene	NEt ₃	70	10
12	S1c	toluene	DMAP	65	17
13	S1c	toluene	ру	72	15
14	S1c	toluene	$PhNH_2$	65	10
15	S1c	toluene	Bn ₂ NH	60	14
16	S1c	toluene	4-morpholin pyridine	o 75	20

Table S1 Optimization studies for the asymmetric transfer hydrogenation of 10^a



^a Reaction conditions: substrates (0.1 mmol, 1 equiv.) and ammonia borane (0.3 mmol, 3 equiv.) with $B(C_6F_5)_3$ (5 mol %) in solvent (1.5 mL) at 80 °C for 8 h.^b The ee values were determined by HPLC.

		B(C ₆ F ₅); NH ₃ BH; toluene, 80° S1c add	C, 8 h		$Ar \qquad O \qquad $
Entry	$B(C_6F_5)_3 mol\%$	S1c mol%	Additive mol%	ee (%) ^b	
1	5	10	10	21	(R) N
2	5	20	20	22	S1a : Ar = 1-naphthyl 4-morpholinopyridin
3	10	20	20	23	S1b : Ar = 2-naphthyl additive
4	20	20	20	13	S1c : Ar = 9-anthracenyl
5	5	10	50	23	S1d : Ar = 9-phenanthyl
6	5	10	100	24	S1e : Ar = $3,5-(CF_3)_2-C_6H_3$

Table S2 Other effect on the asymmetric transfer hydrogenation of 10^a

^a Reaction conditions: substrates (0.1 mmol, 1 equiv.) and ammonia borane (0.3 mmol, 3 equiv.) with $B(C_6F_5)_3$, S1c in toluene (1.5 mL) at 80 °C for 8 h. ^b The ee values were determined by HPLC.

The optimum reaction condition is as follows: substrates (0.1 mmol, 1 equiv.) and ammonia borane (0.3 mmol, 3 equiv.) with $B(C_6F_5)_3$ (5 mol%), S1c (10 mol%) and 4-morpholinopyridine (0.1 mmol, 1 equiv.) in toluene (1.5 mL) at 80 °C for 8 h.



Fig. 1 Proposed catalytic cycle for the hydrogenations of N-heterocycles.

Based on relevant reports in the literature,^{9, 17, 19} a plausible reaction mechanism for this $B(C_6F_5)_3$ -promoted hydrogenation of N-heterocycles with ammonia borane has been proposed in Fig. 1. We assume that the FLPs of quinaldine and $B(C_6F_5)_3$ can split the N–H and B–H bonds of ammonia borane to generate zwitterion species (**B**). The location of the subsequent hydride attack could occur at the 2- or 4- position of quinaldine, because both sites are electrophilic. On the one hand, attacking at the 4- position would produce a transient enamine (**C**). On the other hand, attacking by hydride at the 2-position would generate the 1,2-dihydro-quinoline which can undergo 1,3 proton shifts to regenerate an imine (**D**). As a result, either attacking at the 4- position or at the 2- position, it would give the same intermediate product (**E**). Subsequently, the FLPs of imine **E** and $B(C_6F_5)_3$ activate ammonia borane to generate zwitterion species (**F**). Finally, imines were hydrogenated giving the reductive products (**2a**) with catalyst $B(C_6F_5)_3$ released. In addition, the mechanism for the hydrogenations of indole is similar to that of quinaldine.

2. Experimental Details

2-methyl-1,2,3,4-tetrahydroquinoline (2a)^[1]



¹H NMR (400 MHz, Chloroform-*d*) δ 7.11 – 6.88 (m, 2H), 6.64 (td, *J* = 7.4, 1.1 Hz, 1H), 6.58 – 6.40 (m, 1H), 3.69 (s, 1H), 3.43 (dqd, *J* = 9.3, 6.2, 2.8 Hz, 1H), 2.98 – 2.69 (m, 2H), 1.96 (ddt, *J* = 12.5, 6.1, 3.2 Hz, 1H), 1.63 (m, 1H), 1.24 (d, *J* = 6.3 Hz, 3H).

¹³C NMR (150 MHz, CDCl₃) δ 144.8, 129.4, 126.8, 121.2, 117.1, 114.1, 47.3, 30.2, 26.7, 22.7. HPLC (OJ-H, elute: Hexanes/i-PrOH = 90/10, detector: 254 nm, flow rate: 0.5 mL/min), t_1 = 19.9 min (major), t_2 = 22.0 min (minor).

1,2,3,4-tetrahydroquinolin-8-amine (2b)^[1]



¹H NMR (600 MHz, Chloroform-*d*) δ 6.69 – 6.47 (m, 3H), 3.39 – 3.32 (m, 2H), 3.28 (s, 3H), 2.78 (t, *J* = 6.4 Hz, 2H), 1.92 (dt, *J* = 11.0, 6.3 Hz, 2H).

¹³C NMR (150 MHz, CDCl₃) δ 133.9, 133.8, 123.4, 121.2, 118.2, 114.2, 42.6, 27.1, 22.4. 1,2,3,4-tetrahydroquinoline (**2c**)^[1]



¹H NMR (600 MHz, Chloroform-*d*) δ 7.05 – 6.92 (m, 2H), 6.62 (td, *J* = 7.4, 1.2 Hz, 1H), 6.49 (d, *J* = 7.9 Hz, 1H), 3.77 (s, 1H), 3.38 – 3.26 (m, 2H), 2.78 (t, *J* = 6.5 Hz, 2H), 2.02 – 1.91 (m, 2H). ¹³C NMR (150 MHz, CDCl₃) δ 144.8, 129.6, 126.8, 121.6, 117.1, 114.3, 42.1, 27.1, 22.3. 8-bromo-1,2,3,4-tetrahydroquinoline (**2d**)^[2]



¹H NMR (400 MHz, Chloroform-*d*) δ 7.24 (d, *J* = 7.8 Hz, 1H), 6.90 (d, *J* = 7.4 Hz, 1H), 6.46 (t, *J* = 7.7 Hz, 1H), 4.44 (s, 1H), 3.40 (t, *J* = 5.6 Hz, 2H), 2.79 (t, *J* = 6.4 Hz, 2H), 2.08 – 1.85 (m, 2H). ¹³C NMR (150 MHz, CDCl₃) δ 141.8, 130.1, 128.5, 122.9, 117.0, 108.8, 42.1, 27.5, 21.8. 5,8-dibromo-1,2,3,4-tetrahydroquinoline (**2e**)^[1]



¹H NMR (400 MHz, Chloroform-*d*) δ 3.43 – 3.29 (m, 1H), 2.78 (t, *J* = 6.5 Hz, 1H), 2.05 – 1.89 (m, 1H).

¹³C NMR (150 MHz, CDCl₃) δ 143.3, 130.7, 124.6, 122.0, 120.6, 107.6, 77.4, 41.6, 28.4, 21.9. 6-methoxy-1,2,3,4-tetrahydroquinoline (**2f**)^[1]



¹H NMR (600 MHz, Chloroform-*d*) δ 6.61 (dd, *J* = 8.6, 3.0 Hz, 1H), 6.57 (d, *J* = 2.7 Hz, 1H), 6.48 (d, *J* = 8.6 Hz, 1H), 3.74 (d, *J* = 1.0 Hz, 3H), 3.73 – 3.69 (m, 1H), 3.31 – 3.19 (m, 2H), 2.76 (t, *J* = 6.5 Hz, 2H), 2.02 – 1.86 (m, 2H).

¹³C NMR (150 MHz, CDCl₃) δ 152.0, 138.7, 123.1, 115.8, 115.0, 113.0, 55.9, 42.5, 27.2, 22.5. 5,7-dibromo-1,2,3,4-tetrahydroquinolin-8-ol (**2g**)^[3]



¹H NMR (600 MHz, Chloroform-*d*) δ 6.96 (s, 1H), 5.17 (s, 1H), 4.44 (s, 1H), 3.31 (t, *J* = 5.5 Hz, 2H), 2.71 (t, *J* = 6.5 Hz, 2H), 1.96 (dd, *J* = 6.6, 4.9 Hz, 2H).

¹³C NMR (150 MHz, CDCl₃) δ 138.0, 135.5, 120.4, 120.4, 116.3, 106.7, 40.9, 27.6, 21.8. 3-methyl-1,2,3,4-tetrahydroquinoline (**2h**)^[1]



¹H NMR (400 MHz, Chloroform-*d*) δ 6.95 (t, *J* = 6.9 Hz, 2H), 6.62 (t, *J* = 7.4 Hz, 1H), 6.52 (d, *J* = 8.0 Hz, 1H), 3.28 (d, *J* = 11.2 Hz, 1H), 2.90 (t, *J* = 10.4 Hz, 1H), 2.78 (dd, *J* = 16.1, 4.8 Hz, 1H), 2.43 (dd, *J* = 16.2, 10.2 Hz, 1H), 2.07 (s, 1H), 1.26 (s, 1H), 1.05 (dd, *J* = 6.6, 1.9 Hz, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 144.0, 129.7, 126.9, 121.6, 117.4, 114.3, 49.0, 35.6, 27.3, 19.2.

1,2,3,4-tetrahydroquinoxaline (2i)^[1]



¹H NMR (400 MHz, Chloroform-*d*) δ 6.59 (dd, *J* = 5.7, 3.5 Hz, 2H), 6.50 (dd, *J* = 5.6, 3.6 Hz, 2H), 3.50 (s, 2H), 3.42 (s, 4H).

¹³C NMR (150 MHz, CDCl₃) δ 133.5, 119.1, 115.1, 41.4.

5-methyl-1,2,3,4-tetrahydroquinoxaline (2j)^[1]



¹H NMR (400 MHz, Chloroform-*d*) δ 6.65 – 6.51 (m, 2H), 6.43 (dd, J = 6.7, 2.6 Hz, 1H), 3.63 – 3.47 (m, 4H), 3.46 – 3.39 (m, 2H), 2.12 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 133.2, 131.7, 122.2, 120.4, 118.0, 112.9, 41.8, 41.2, 17.0. 6-chloro-1,2,3,4-tetrahydroquinoxaline (**2k**)^[4]



¹H NMR (400 MHz, Chloroform-*d*) δ 6.51 (dd, *J* = 8.3, 2.3 Hz, 1H), 6.44 (d, *J* = 2.3 Hz, 1H), 6.38 (d, *J* = 8.2 Hz, 1H), 3.58 (s, 2H), 3.38 (s, 4H).

¹³C NMR (150 MHz, CDCl₃) δ 134.8, 132.0, 123.3, 118.0, 115.4, 114.0, 41.1. 2-methyl-1,2,3,4-tetrahydroquinoxaline (**2I**)^[1]



¹H NMR (600 MHz, Chloroform-*d*) δ 6.60 (dd, J = 5.7, 3.4 Hz, 2H), 6.52 (dt, J = 6.0, 3.1 Hz, 2H), 3.68 (s, 2H), 3.50 (d, J = 8.4 Hz, 1H), 3.39 – 3.25 (m, 1H), 3.03 (t, J = 9.6 Hz, 1H), 1.20 (d, J = 6.3 Hz, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 133.5, 133.1, 119.0, 119.0, 114.8, 114.7, 48.3, 45.9, 19.9. 9,10-dihydroacridine (**2m**)^[1]



¹H NMR (400 MHz, Chloroform-*d*) δ 7.19 – 7.03 (m, 4H), 6.86 (t, *J* = 7.4 Hz, 2H), 6.67 (d, *J* = 7.8 Hz, 2H), 5.95 (s, 1H), 4.07 (s, 2H).

¹³C NMR (150 MHz, CDCl₃) δ 140.2, 128.7, 127.1, 120.8, 120.1, 113.6, 31.5. 1,2,3,4-tetrahydro-1,10-phenanthroline (**2n**)^[1]



¹H NMR (400 MHz, Chloroform-*d*) δ 8.68 (dd, *J* = 4.2, 1.6 Hz, 1H), 8.01 (dd, *J* = 8.3, 1.6 Hz, 1H), 7.29 (dd, *J* = 8.2, 4.2 Hz, 1H), 7.16 (d, *J* = 8.2 Hz, 1H), 6.97 (d, *J* = 8.2 Hz, 1H), 5.93 (s, 1H), 3.53 (t, *J* = 5.5 Hz, 2H), 2.92 (t, *J* = 6.3 Hz, 2H), 2.15 – 2.00 (m, 2H).

¹³C NMR (150 MHz, CDCl₃) δ 147.0, 140.8, 137.6, 136.1, 129.2, 127.5, 120.7, 116.8, 113.2, 41.4, 27.2, 21.9.

Indoline (4a)^[5]



¹H NMR (600 MHz, Chloroform-*d*) δ 7.13 (dd, J = 7.3, 1.3 Hz, 1H), 7.03 (td, J = 7.6, 1.2 Hz, 1H), 6.72 (td, J = 7.4, 1.0 Hz, 1H), 6.67 (d, J = 7.7 Hz, 1H), 3.79 – 3.62 (m, 1H), 3.56 (t, J = 8.4 Hz, 2H), 3.04 (t, J = 8.4 Hz, 2H).

¹³C NMR (150 MHz, CDCl₃) δ 151.2, 129.6, 127.4, 124.8, 119.1, 109.9, 47.4, 29.9.
5-fluoroindoline (4b)^[5]



¹H NMR (600 MHz, Chloroform-*d*) δ 6.84 (ddt, *J* = 8.4, 2.4, 1.1 Hz, 1H), 6.71 (m, 1H), 6.54 (dd, *J* = 8.4, 4.3 Hz, 1H), 3.56 (t, *J* = 8.4 Hz, 2H), 3.47 (s, 1H), 3.01 (td, *J* = 8.4, 1.1 Hz, 2H). ¹³C NMR (150 MHz, CDCl₃) δ 158.0, 156.4, 147.5, 131.3, 131.3, 113.3, 113.2, 112.2, 112.1, 109.8, 109.7, 48.1, 30.3, 30.3.

5-bromoindoline (4c)^[6]



¹H NMR (600 MHz, Chloroform-*d*) δ 7.19 (dt, *J* = 2.2, 1.2 Hz, 1H), 7.12 – 7.07 (m, 1H), 6.49 (d, *J* = 8.3 Hz, 1H), 3.73 – 3.61 (m, 1H), 3.56 (t, *J* = 8.4 Hz, 2H), 3.01 (tt, *J* = 8.3, 1.0 Hz, 2H). ¹³C NMR (150 MHz, CDCl₃) δ 150.7, 131.9, 129.9, 127.7, 110.6, 110.2, 47.7, 29.8. 6-methoxyindoline (**4d**)^[7]



¹H NMR (600 MHz, Chloroform-*d*) δ 7.00 (dt, *J* = 8.4, 1.2 Hz, 1H), 6.25 (d, *J* = 7.2 Hz, 2H), 3.76 (s, 3H), 3.56 (t, *J* = 8.3 Hz, 2H), 2.96 (td, *J* = 8.3, 1.1 Hz, 2H).

¹³C NMR (150 MHz, CDCl₃) δ 159.9, 153.0, 124.8, 121.7, 103.4, 96.4, 55.5, 55.5, 48.1, 29.1. 7-methylindoline (**4e**)^[8]



¹H NMR (600 MHz, Chloroform-*d*) δ 7.05 – 6.99 (m, 1H), 6.94 – 6.86 (m, 1H), 6.69 (t, *J* = 7.4 Hz, 1H), 3.59 (t, *J* = 8.4 Hz, 2H), 3.08 (td, *J* = 8.4, 0.9 Hz, 2H), 2.17 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 150.2, 128.7, 128.2, 122.2, 119.0, 118.9, 47.3, 30.2, 17.0. 2-methylindoline (**4f**)^[5]



¹H NMR (400 MHz, Chloroform-*d*) δ 7.08 (dd, J = 7.3, 1.3 Hz, 1H), 7.01 (td, J = 7.6, 1.2 Hz, 1H), 6.69 (td, J = 7.4, 1.0 Hz, 1H), 6.61 (d, J = 7.7 Hz, 1H), 4.07 – 3.91 (m, 1H), 3.52 (s, 1H), 3.15 (dd, J = 15.4, 8.5 Hz, 1H), 2.64 (dd, J = 15.4, 7.8 Hz, 1H), 1.30 (d, J = 6.2 Hz, 3H).

¹³C NMR (150 MHz, CDCl₃) δ 150.9, 129.1, 127.4, 124.8, 118.7, 109.4, 55.4, 37.8, 22.4.
2-phenylindoline (4g)^[9]



¹H NMR (400 MHz, Chloroform-*d*) δ 7.51 – 7.41 (m, 2H), 7.40 – 7.33 (m, 2H), 7.32 – 7.26 (m, 1H), 7.16 – 7.03 (m, 2H), 6.76 (td, *J* = 7.4, 1.0 Hz, 1H), 6.69 (d, *J* = 7.7 Hz, 1H), 4.97 (t, *J* = 9.0 Hz, 1H), 4.03 (s, 1H), 3.46 (dd, *J* = 15.6, 9.2 Hz, 1H), 3.01 (ddt, *J* = 15.6, 8.7, 1.1 Hz, 1H). ¹³C NMR (150 MHz, CDCl₃) δ 150.5, 144.3, 129.1, 128.7, 128.4, 127.7, 127.6, 126.5, 125.3, 124.7, 119.3, 109.4, 63.6, 39.6.

3-methylindoline (4h)^[6]



¹H NMR (400 MHz, Chloroform-*d*) δ 7.14 – 6.98 (m, 2H), 6.75 (t, *J* = 7.4 Hz, 1H), 6.66 (d, *J* = 7.8 Hz, 1H), 3.71 (t, *J* = 8.7 Hz, 1H), 3.38 (q, *J* = 7.7 Hz, 1H), 3.12 (t, *J* = 8.6 Hz, 2H), 1.33 (d, *J* = 6.7 Hz, 3H).

¹³C NMR (150 MHz, CDCl₃) δ 151.2, 134.5, 127.4, 123.5, 118.9, 109.7, 55.5, 36.7, 18.7. 2,3-dihydrobenzo[d]thiazole (**4i**)^[1]



¹H NMR (600 MHz, Chloroform-*d*) δ 7.38 – 6.80 (m, 2H), 6.67 – 6.24 (m, 2H), 4.93 – 4.71 (m, 1H), 2.70 (d, *J* = 3.8 Hz, 2H).

¹³C NMR (150 MHz, CDCl₃) δ 150.4, 137.1, 132.2, 118.5, 116.3, 109.9, 30.5.

1,2-diphenylhydrazine (5)^[10]



¹H NMR (600 MHz, Chloroform-*d*) δ 7.29 – 7.16 (m, 5H), 6.88 – 6.84 (m, 5H), 5.62 (s, 2H). ¹³C NMR (150 MHz, CDCl₃) δ 149.0, 129.5, 120.0, 112.4. 2-phenyl-1,2,3,4-tetrahydroquinoline (**2o**)^[31]



¹H NMR (400 MHz, Chloroform-d): δ 7.42 – 7.35 (m, 4H), 7.32 – 7.30 (m, 1H), 7.05 – 7.01 (m, 2H), 6.69 – 6.65 (m, 1H), 6.57 – 6.55 (m, 1H), 4.47 – 4.44 (dd, *J* = 9.2, 3.2 Hz, 1H), 4.06 (s, 1H), 2.97 – 2.90 (m, 1H), 2.79 – 2.72 (m, 1H), 2.16 – 2.11 (m, 1H), 2.06 – 1.99 (m, 1H);

¹³C NMR (150 MHz, CDCl₃): δ 145.0, 144.9, 129.4, 128.7, 127.6, 127.0, 126.7, 121.0, 117.3, 114.1, 56.4, 31.1, 26.5.

HPLC (OD-H, elute: Hexanes/i-PrOH = 80/20, detector: 254 nm, flow rate: 1.0 mL/min), $t_1 = 7.5$ min (minor), $t_2 = 9.5$ min (major).

2-([1,1'-biphenyl]-4-yl)-1,2,3,4-tetrahydroquinoline (**2p**)^[31]



¹H NMR (400 MHz, Chloroform-d): δ 7.64 – 7.61 (m, 4H), 7.58 – 7.46 (m, 4H), 7.40 – 7.36 (m, 1H), 7.08 – 7.04 (m, 2H), 6.70 (t, *J* = 7.2 Hz, 1H), 6.60 (d, *J* = 8.4 Hz, 1H), 4.52 (dd, *J* = 9.2, 3.2 Hz, 1H), 4.16 (s, 1H), 3.02 – 2.94 (m, 1H), 2.83 – 2.77 (m, 1H), 2.23 – 2.16 (m, 1H), 2.12 – 2.02 (m, 1H); ¹³C NMR (150 MHz, CDCl₃): δ 144.8, 144.0, 141.0, 140.6, 129.5, 128.9, 127.5, 127.4, 127.2, 127.1, 127.1, 121.0, 117.4, 114.2, 56.1, 31.1, 26.5.

HPLC (OD-H, elute: Hexanes/i-PrOH = 80/20, detector: 254 nm, flow rate: 1.0 mL/min), $t_1 = 13.4$ min (minor), $t_2 = 18.7$ min (major).

6-chloro-3-(4-chlorophenyl)-3,4-dihydro-2H-benzo[b][1,4]oxazine (2q)^[31]



¹H NMR (400 MHz, Chloroform-d): δ 7.33 (dd, *J* = 21.2, 8 Hz, 4H), 6.76 – 6.74 (m, 1H), 6.65 – 6.64 (m, 2H), 4.48 (d, *J* = 8.4 Hz, 1H), 4.24 (d, *J* = 10.4 Hz, 1H), 3.95 – 3.90 (m, 2H). ¹³C NMR (150 MHz, CDCl₃): δ 142.1, 137.4, 134.6, 134.4, 129.2, 128.6, 126.4, 118.8, 117.7, 115.0, 70.7, 53.5. HPLC (OD-H, elute: Hexanes/i-PrOH = 80/20, detector: 254 nm, flow rate: 1.0 mL/min), $t_1 = 9.0$ min (minor), $t_2 = 15.3$ min (major).

3-(3,4-dichlorophenyl)-3,4-dihydro-2H-benzo[b][1,4]oxazine (2r)^[31]



¹H NMR (400 MHz, Chloroform-d): δ 7.51 (d, J = 2.0 Hz, 1H), 7.45 (d, J = 8.0 Hz, 1H), 7.24 (dd, J = 8.0, 2.0 Hz, 1H), 6.86 – 6.81 (m, 2H), 6.75 – 6.68 (m, 2H), 4.49 (dd, J = 8.0, 2.8 Hz, 1H), 4.26 (dd, J = 10.4, 2.8 Hz, 1H), 3.99 – 3.93 (m, 2H);

¹³C NMR (150 MHz, CDCl₃): δ 143.6, 139.8, 133.3, 133.2, 132.5, 130.9, 129.3, 126.6, 121.9, 119.5, 116.9, 115.7, 70.6, 53.4.

HPLC (OD-H, elute: Hexanes/i-PrOH = 80/20, detector: 254 nm, flow rate: 1.0 mL/min), t₁ = 13.0 min (minor), t₂ = 27.5 min (major).

3-(naphthalen-2-yl)-3,4-dihydro-2H-benzo[b][1,4]oxazine (2s)^[31]



¹H NMR (400 MHz, Chloroform-d): δ 7.89 – 7.86 (m, 4H), 7.53 – 7.51 (m, 3H), 6.91 – 6.84 (m, 2H), 6.77 – 6.72 (m, 2H), 4.68 (dd, J = 8.4, 2.4 Hz, 1H, CH2), 4.37 (dd, J = 10.4, 2.4 Hz, 1H), 4.12 – 4.07 (m, 2H).

¹³C NMR (150 MHz, CDCl₃): δ 143.6, 136.6, 133.9, 133.4, 133.4, 128.7, 128.0, 127.8, 126.4, 126.3, 126.2, 125.0, 121.6, 119.0, 116.7, 115.5, 71.0, 54.4.

HPLC (OD-H, elute: Hexanes/i-PrOH = 80/20, detector: 254 nm, flow rate: 1.0 mL/min), t₁ = 16.9 min (minor), t₂ = 32.0 min (major).

2-(2-(benzo[d][1,3]dioxol-5-yl)ethyl)-1,2,3,4-tetrahydroquinoline (2t)^[31]



¹H NMR (400 MHz, Chloroform-d): δ 6.97 – 6.95 (m, 2H), 6.76 – 6.71 (m, 2H), 6.67 – 6.62 (m, 2H), 6.48 (d, J = 8.0 Hz, 1H), 5.93 (s, 2H), 3.82 (br, 1H), 3.30 – 3.28 (m, 1H), 2.86 – 2.65 (m, 4H), 2.01 – 1.98 (m, 1H), 1.82 – 1.77 (m, 2H), 1.72 – 1.62 (m, 1H).

¹³C NMR (150 MHz, CDCl₃) δ 147.8, 145.8, 144.5, 135.7, 129.4, 126.9, 121.5, 121.2, 117.3, 114.4, 108.9, 108.4, 100.9, 51.1, 38.6, 32.0, 28.0, 26.3.

HPLC (OD-H, elute: Hexanes/i-PrOH = 80/20, detector: 254 nm, flow rate: 1.0 mL/min), t₁ = 11.1 min (minor), t₂ = 14.2 min (major).

2-pentyl-1,2,3,4-tetrahydroquinoline (2u)^[31]



¹H NMR (400 MHz, Chloroform-d): δ 6.96 – 6.95 (m, 2H), 6.62 – 6.59 (m, 1H), 6.50 – 6.48 (m, 1H), 3.25 – 3.24 (m, 1H), 2.82 – 2.71 (m, 2H), 2.05 – 1.95 (m, 1H), 1.61 – 1.50 (m, 1H), 1.49 – 1.47 (m, 2H), 1.41 – 1.27 (m, 7H), 0.92 – 0.90 (m, 3H);

¹³C NMR (150 MHz, CDCl₃): δ 144.8, 129.4, 126.8, 121.6, 117.1, 114.2, 51.8, 36.8, 32.1, 28.2, 26.6, 25.5, 22.8, 14.2.

HPLC (OD-H, elute: Hexanes/i-PrOH = 90/10, detector: 254 nm, flow rate: 0.5 mL/min), $t_1 = 12.4$ min (major), $t_2 = 13.5$ min (minor).







50 145 140 135 130 125 120 115 110 105 100 95 90 85 80 75 70 65 60 55 50 45 40 35 30 25 20 15 10 5 0 f1 (ppm)



































4 References

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HPLC trace of racemic 2-methyl-1,2,3,4-tetrahydroquinoline (**2a**)

HPLC trace of enantioenriched 2-methyl-1,2,3,4-tetrahydroquinoline (2a)

0.4175 4.53837e4 1652.17798 38.3836

2 22.236 BB

HPLC trace of racemic 2-phenyl-1,2,3,4-tetrahydroquinoline (20)

峰	保留时间 类型	峰宽	峰面积	峰高	峰面积
#	[min]	[min]	[mAU*s]	[mAU]	%
	.				
1	7.514 MM	0. 2098	3.87299e4	3076.91602	48.3355
2	9.551 MM	0. 2557	4.13973e4	2698.10034	51.6645

HPLC trace of enantioenriched 2-phenyl-1,2,3,4-tetrahydroquinoline (20)

峰	保留时间	类型	峰宽	峰面积	峰高	峰面积
#	[min]		[min]	[mAU*s]	[mAU]	%
1	7.704	BB	0. 1590	6429.29492	624.83246	37.8103
2	9.799	BB	0. 2110	1.05748e4	775.82117	62.1897

HPLC trace of racemic 2-([1,1'-biphenyl]-4-yl)-1,2,3,4-tetrahydroquinoline (**2p**)

 1
 13.429 VB R
 0.5370 4719.20020
 129.55571
 50.2877

 2
 18.699 BB
 0.5448 4665.21143
 132.80237
 49.7123

HPLC trace of enantioenriched 2-([1,1'-biphenyl]-4-yl)-1,2,3,4-tetrahydroquinoline(**2p**)

0.5434 4508.97314 128.50739 53.7673

2 18.728 BB

HPLC trace of racemic 6-chloro-3-(4-chlorophenyl)-3,4-dihydro-2H-benzo [b][1,4]oxazine (**2q**)

1	8.914 BB	0.2238 4367.47803	301. 92303	50.0061
2	14.988 BB	0.4071 4366.41650	166. 98267	49.9939

HPLC trace of enantioenriched 6-chloro-3-(4-chlorophenyl)-3,4-dihydro-2H-benzo[b][1,4]oxazine (**2q**)

HPLC trace of racemic 3-(3,4-dichlorophenyl)-3,4-dihydro-2H-benzo[b] [1,4]oxazine (**2r**)

峰	保留时间	类型	峰宽	峰面积	峰高	峰面积
#	[min]		[min]	[mAU*s]	[mAU]	%
1	12.986	BB	0.3450	2820. 04761	127.02346	50.0846
2	27.964	BB	0.7913	2810. 52612	55.08429	49.9154

HPLC trace of enantioenriched 3-(3,4-dichlorophenyl)-3,4-dihydro-2H-benzo[b] [1,4]oxazine (**2r**)

峰	保留时间	类型	峰宽	峰面积	峰高	峰面积
#	[min]		[min]	[mAU*s]	[mAU]	%
1	12.295 I	BB	0.3159	3368.27759	165.90019	38.3180
2	25.952 I	BBA	0.7306	5422.05322	115. 10542	61.6820

HPLC trace of racemic 3-(naphthalen-2-yl)-3,4-dihydro-2H-benzo[b] [1,4]oxazine (**2s**)

峰	保留时间	类型 峰等	宽 峰口	面积 峭	峰高 峰面	积
#	[min]	[mi	n] [mAU	J*s] [m	AU] %	
	-					
1	16.883 E	3B 0.4	1350 6621.	44922 236	. 26204 49.87	799
2	32.031 E	3B 0.8	3574 6653.	34717 120	. 63855 50. 12	201

HPLC trace of enantioenriched 3-(naphthalen-2-yl)-3,4-dihydro-2H-benzo [b][1,4]oxazine (**2s**)

2	20 225 BI		6961 16015	12/ 58812	58 03/0
Ζ.	30.333 DI	0.0037	0904.40040	134. JOOIZ	00.9040

HPLC trace of racemic 2-(2-(benzo[d][1,3]dioxol-5-yl) ethyl)-1,2,3,4-tetrahydroquinoline (**2t**)

峰	保留时间	类型 峰宽	峰面积	峰高	峰面积
#	[min]	[min]	[mAU*s]	[mAU]	%
	• •		·		
1	11.299 Bl	B 0.2828	3791.93921	207.93303	47.8923
2	14.505 BI	B 0.3687	4125.70020	173.95863	52.1077

HPLC trace of enantioenriched 2-(2-(benzo[d][1,3]dioxol-5-yl) ethyl)-1,2,3,4-tetrahydroquinoline (**2t**)

峰 伊 # 	R留时间 [min]	类型 ·	峰宽 [min]	峰面积 [mAU*s] 	峰高 [mAU] 	峰面积 %
1	11.111	BB	0. 2798	1.03722e4	574.06506	35.5270
2	14.203	BB	0.3689	1.88231e4	793.17834	64.4730

HPLC trace of racemic 2-pentyl-1,2,3,4-tetrahydroquinoline (2u)

峰	保留时间	类型	峰宽	峰面积	峰高	峰面积
#	[min]		[min]	[mAU*s]	[mAU]	%
		-				
1	12.282	BB	0. 2126	2.66411e4	1947. 52844	50.0071
2	2 13.364	BB	0. 2232	2.66335e4	1847.40735	49.9929

HPLC trace of enantioenriched 2-pentyl-1,2,3,4-tetrahydroquinoline (2u)

