# **Supporting Materials**

# Effect of Ligand Structures on the Size Control of Mono- and Bi-Thiolate-Protected Silver Nanoclusters

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#### **EXPERIMENTAL SECTION**

#### 1. Materials

Ultrapure water (18.2 M $\Omega$ ) was used throughout the study. All glassware and magnetic stir bars were washed with diluted nitric acid and *aqua regia*, rinsed with abundant ethanol and water, and dried in an oven before use. All chemicals were commercially available and used as received: 3-mercaptopropionic acid (MPA), 4-mercaptobutyric acid (MTBA), 6mercaptohexanoic acid (MHA), sodium borohydride (NaBH<sub>4</sub>), *para*-mercaptobenzoic acid (*p*-MBA), and glutathione (GSH) were purchased from Sigma-Aldrich; silver nitrate (AgNO<sub>3</sub>) and sodium hydroxide (NaOH) were purchased from Merck; ethanol was purchased from Fisher.

#### 2. Instruments

UV-vis absorption spectra were recorded by a Shimadzu UV-1800 spectrometer. Native polyacrylamide gel electrophoresis (PAGE) was carried out on a Bio-Rad Mini-Protean Tetra Cell system using discontinuous gels ( $1.0 \times 83 \times 73$  mm). 20 wt% and 4 wt% acrylamide monomers were prepared for resolving and stacking gels, respectively. Electrospray ionization mass spectra (ESI-MS) of Ag nanoclusters (NCs) were acquired on a Bruker microTOF-Q system.

#### 3. Synthesis of alkane-thiol-protected Ag<sub>25</sub>NCs

Aqueous solutions of AgNO<sub>3</sub> (20 mM) and alkane-thiol ligands (10 mM) were freshly prepared with ultrapure water. In a typical synthesis of  $Ag_{25}(MHA)_{18}$  NCs, aqueous solutions of AgNO<sub>3</sub> (20 mM, 0.125 mL; as silver precursor) and MHA (10 mM, 0.5 mL; as protecting ligand) were mixed in water (4.225 mL) or mixed solvents of ethanol/water (up to 40 vol% of ethanol) to form Ag(I)-MHA complexes with an opalescent color under stirring condition at 50 °C. Next, NaOH solution (1 M) was added to the system to adjust the pH to ~11-12. Afterwards, fresh NaBH<sub>4</sub> solution (0.1 mL, prepared by dissolving 43 mg of NaBH<sub>4</sub> powder in 10 mL of 0.2 M NaOH solution) was added dropwise to reduce the complexes. The  $Ag_{25}(MHA)_{18}$  NCs were collected

after 30 min for further characterization. By using the same synthetic conditions, MTBA- and MPA-protected Ag<sub>25</sub> NCs can also be synthesized.

## 4. Synthesis of *p*-MBA-protected Ag<sub>44</sub> NCs

In a typical synthesis of  $Ag_{44}(p-MBA)_{30}$  NCs, an aqueous solution of  $AgNO_3$  (20 mM, 0.125 mL; as silver precursor) and an ethanolic solution of *p*-MBA (50 mM, 0.1 mL; as protecting ligand) were added to a mixed solvent of ethanol/water (4.625 mL, from 0 vol% to 40 vol% of ethanol) to form light-yellow Ag(I)-*p*-MBA complexes under stirring condition at 50 °C. Next, NaOH solution (1 M) was added to the system to adjust the pH to 12. Afterwards, fresh NaBH<sub>4</sub> solution (0.1 mL) was added dropwise to reduce the complexes. The  $Ag_{44}(p-MBA)_{30}$  NCs were collected after 2 h.

#### 5. Synthesis of GSH-protected Ag<sub>9-15</sub> NCs

In a typical synthesis, freshly prepared aqueous solutions of  $AgNO_3$  (20 mM, 0.125 mL; as silver precursor) and GSH (10 mM, 0.5 mL; as protecting ligand) were mixed in water (4.225 mL) to form opalescent Ag(I)-GSH complexes under stirring condition at 50 °C. Next, NaOH solution (1 M) was added to the system to adjust the pH to ~11-12. Afterwards, fresh NaBH<sub>4</sub> solution (0.1 mL) was added dropwise to reduce the complexes. The  $Ag_{9-15}(GSH)_{5-10}$  NCs were collected after 30 min.

### 6. Synthesis of bi-ligand-protected Ag NCs

# 6.1. Synthesis of bi-thiolate MHA\_MPA- and MHA\_GSH-protected Ag<sub>25</sub> NCs

The bi-thiolate MHA\_MPA- and MHA\_GSH-protected  $Ag_{25}$  NCs were synthesized by using the same synthetic condition for  $Ag_{25}(MHA)_{18}$  NCs except for the introduction of bi-thiolate ligands into the reaction system. Typically, aqueous solutions of MHA and MPA or GSH (10 mM) with different feeding ratios such as 4 : 1 (0.4 mL MHA and 0.1 mL MPA or GSH), or 2 : 1 (0.333 mL MHA and 0.167 mL MPA or GSH), or 1 : 1 (0.25 mL MHA and 0.25 mL MPA or GSH) were mixed with AgNO<sub>3</sub> (20 mM, 0.125 mL) in water (4.225 mL) to form Ag(I)-MHA\_MPA or Ag(I)-MHA\_GSH complexes, Next, NaOH solution (1 M) was added to the system to adjust the pH to ~11-12. Afterwards, fresh NaBH<sub>4</sub> solution (0.1 mL) was added dropwise to reduce the

complexes. The bi-thiolate MHA\_MPA- or MHA\_GSH-protected  $Ag_{25}$  NCs were collected after 30 min for further characterization.

# 6.2. Synthesis of bi-thiolate MHA\_p-MBA-protected Ag<sub>25</sub> NCs

In a typical synthesis of bi-thiolate MHA\_*p*-MBA-protected Ag NCs, an aqueous solution of MHA (50 mM) and an ethanolic solution of *p*-MBA (50 mM) with different feeding ratios such as 4 : 1 (0.08 mL MHA and 0.02 mL *p*-MBA), or 2 : 1 (0.067 mL MHA and 0.033 mL *p*-MBA), or 1 : 1 (0.05 mL MHA and 0.05 mL *p*-MBA) were mixed with AgNO<sub>3</sub> solution (20 mM, 0.125 mL) in a mixed solvent of ethanol/water (4.625 mL, 0–40 vol% of ethanol) to form Ag(I)-MHA\_*p*-MBA complexes, Next, NaOH solution (1 M) was added to the system to adjust the pH to 12. Afterwards, fresh NaBH<sub>4</sub> solution (0.1 mL) was added dropwise to reduce the complexes. The bi-thiolate MHA\_*p*-MBA-protected Ag<sub>25</sub> NCs were collected after 2 h for further characterization.



**Fig. S1** UV-vis absorption spectra of (a)  $Ag_{25}(MHA)_{18}$  NCs and (b)  $Ag_{44}(p-MBA)_{30}$  NCs synthesized at pH 12 in mixed solvents of ethanol/water: 40 vol% ethanol (black curves), 20 vol% ethanol (blue curves) and 0 vol% ethanol (red curves).

**Note:** As shown in Fig. S1a, the synthesis of  $Ag_{25}(MHA)_{18}$  NCs was tolerant with solvent proportions. The absorption features of  $Ag_{25}(SR)_{18}$  NCs were observed for the MHA-protected Ag NCs synthesized in 40 vol% ethanol, 20 vol% ethanol, and 0 vol% ethanol, which offers a good opportunity in synthesizing bi-thiolate MHA\_p-MBA-protected Ag NCs. Similarly, Fig. S1b also showed that  $Ag_{44}(p-MBA)_{30}$  NCs could be synthesized in the same reaction conditions as that for  $Ag_{25}(MHA)_{18}$  NCs, although 40 vol% of ethanol was the optimized condition for the synthesis of high-quality *p*-MBA-protected  $Ag_{44}$  NCs.



**Fig. S2** ESI mass spectra of  $Ag_{25}(MHA)_{18}$  NCs. Broad spectrum of  $Ag_{25}(MHA)_{18}$  NCs (top panel), zoom-in spectrum of  $[Ag_{25}(MHA)_{18} - 4H]^{5-}$  (middle panel), and isotope patterns acquired experimentally (black curve) and theoretically (red curve) of  $[Ag_{25}(MHA)_{18} - 4H]^{5-}$  (bottom panel). In the middle panel, peak 0 corresponds to  $[Ag_{25}(MHA)_{18} - 4H]^{5-}$ , and other peaks (#1–12) are from the successive coordination of [+ Na – H] of peak 0.



Fig. S3 Photograph of native PAGE result of the as-synthesized  $Ag_{25}(MHA)_{18}$  NCs.



Fig. S4 (a-b) Time evolution UV-vis absorption spectra of the reaction solution during the synthesis of  $Ag_{25}(MHA)_{18}$  NCs. (c) ESI mass spectrum of the reaction solution (for the synthesis of  $Ag_{25}(MHA)_{18}$  NCs) after 3 min reaction.



**Fig. S5** Detailed ESI mass spectra of the as-synthesized  $Ag_{9-15}$  NCs protected by GSH ligand. Isotope patterns acquired experimentally (black curve) and theoretically (red curve) of (a)  $[Ag_9(GSH)_5 - 5H + 2Na]^{3-}$ , (b)  $[Ag_{12}(GSH)_8 - 7H + 3Na]^{4-}$ , (c)  $[Ag_{15}(GSH)_8 - 13H + 9Na]^{4-}$ , (d)  $[Ag_{11}(GSH)_7 - 9H + 6Na]^{3-}$ , (e)  $[Ag_{15}(GSH)_{10} - 9H + 5Na]^{4-}$ , and (f)  $[Ag_{12}(GSH)_8 - 10H + 7Na]^{3-}$ .



**Fig. S6** Isotope patterns of  $[Ag_{44}(p-MBA)_{30}]^{4-}$  species acquired experimentally (black curve) and theoretically (red curve).



**Fig. S7** (a) UV-vis absorption and (b) ESI mass spectra of the as-synthesized  $Ag_{25}(MTBA)_{18}$ NCs. The molecular structure of MTBA is shown in the inset of Fig. S7a. In Fig. S7b, a broad spectrum of  $Ag_{25}(MTBA)_{18}$  (top panel), zoom-in spectrum of  $[Ag_{25}(MTBA)_{18} - 4H]^{5-}$  (middle panel), and isotope patterns of  $[Ag_{25}(MTBA)_{18} - 6H + 2Na]^{5-}$  (bottom panel) acquired experimentally (black curve) and theoretically (red curve), were included. In the middle panel, peak 1 corresponds to  $[Ag_{25}(MTBA)_{18} - 5H + Na]^{5-}$ , and other peaks (#2–11) are from the successive coordination of [+ Na - H] of peak 1.



**Fig. S8** (a) UV-vis absorption and (b) ESI mass spectrum of the as-synthesized  $Ag_{25}(MPA)_{18}$  NCs. The molecular structure of MPA is shown in the inset of Fig. S8a. The well-defined UV-vis absorption feature shown in Fig. S8a confirmed the formation of high-quality  $Ag_{25}(MPA)_{18}$  NCs. However, as shown in Fig. S8b, no peak corresponding to  $Ag_{25}(MPA)_{18}$  species was observed in the ESI mass spectrum, which suggests that the  $Ag_{25}(MPA)_{18}$  NCs were very fragile and could be decomposed during the ionization process of ESI analysis, indicating the poor stability of  $Ag_{25}(MPA)_{18}$  NCs.



**Fig. S9** UV-vis absorption spectra of (a)  $Ag_{25}(MHA)_x(MPA)_{18-x}$  NCs, (b)  $Ag_{25}(MHA)_x(GSH)_{18-x}$  NCs, and (c)  $Ag_{25}(MHA)_x(p-MBA)_{18-x}$  NCs synthesized with a feeding ratio of MHA-to-MPA (or GSH, or *p*-MBA) of 4:1 (black curves), 2:1 (blue curves) and 1:1 (red curves).



**Fig. S10** Zoom-in ESI mass spectra of  $Ag_{25}(MHA)_x(MPA)_{18-x}$  NCs (top panels) and corresponding isotope patterns acquired experimentally (black curves) and theoretically (red curves) of  $Ag_{25}(MHA)_x(MPA)_{18-x}$  NCs (bottom panels) synthesized with a feeding MHA-to-MPA ratio of (a) 4:1, (b) 2:1, and (c) 1:1.



**Fig. S11** Zoom-in ESI mass spectra (top panel) and isotope patterns (bottom panel) acquired experimentally (black curves) and theoretically (red curves) of bi-thiolate MHA\_GSH-protected Ag<sub>25</sub> NCs: (a)  $[Ag_{25}(MHA)_{17}(GSH) - 7H + 2Na]^{6-}$  species, (b)  $[Ag_{25}(MHA)_{16}(GSH)_2 - 7H + 2Na]^{6-}$  species, (c)  $[Ag_{25}(MHA)_{15}(GSH)_3 - 8H + 3Na]^{6-}$  species. The peak number labeled in top panels represents the number of [+ Na - H].



**Fig. S12** (a) Zoom-in ESI mass spectra (the peak number represents the number of [+ Na - H]) of Ag<sub>25</sub>(MHA)<sub>18</sub> (top panel) and Ag<sub>25</sub>(MHA)<sub>x</sub>(*p*-MBA)<sub>18-x</sub> (bottom panel, with a feeding MHA-to-*p*-MBA ratio of 2:1). The dotted red lines marked the right shift from  $[Ag_{25}(MHA)_{18} - 9H + 4Na]^{6-}$  to  $[Ag_{25}(MHA)_{11}(p-MBA)_7 - 9H + 4Na]^{6-}$ . (b) Isotope patterns acquired experimentally (black curves) and theoretically (red curves) of  $[Ag_{25}(MHA)_{11}(p-MBA)_7 - 9H + 4Na]^{6-}$ . (c) Isotope patterns acquired experimentally (black curves) and theoretically (color curves) of  $[Ag_{25}(MHA)_{11-17}(p-MBA)_{7-1} - 5H]^6$  (the number labeled in the spectra shows the number of MHA). (d) Zoom-in ESI mass spectra (the peak number represents the number of [+ Na - H]) of  $Ag_{25}(MHA)_{18}$  (top panel) and  $Ag_{25}(MHA)_x(p-MBA)_{18-x}$  (bottom panel, feeding ratio of MHA-to-*p*-MBA is 1:1). The dotted red lines marked the right shift from  $[Ag_{25}(MHA)_{18} - 9H + 4Na]^{6-}$  to  $[Ag_{25}(MHA)_{14}(p-MBA)_4 - 9H + 4Na]^{6-}$ . (e) Isotope patterns acquired experimentally (black curves) and theoretically (color curves) of  $[Ag_{25}(MHA)_{14}(p-MBA)_4 - 9H + 4Na]^{6-}$ . (f) Isotope patterns acquired experimentally (black curves) and theoretically (color curves) of  $[Ag_{25}(MHA)_{14}(p-MBA)_4 - 9H + 4Na]^{6-}$ . (f) Isotope patterns acquired experimentally (black curves) and theoretically (color curves) of  $[Ag_{25}(MHA)_{14}(p-MBA)_4 - 9H + 4Na]^{6-}$ . (f) Isotope patterns acquired experimentally (black curves) and theoretically (color curves) of  $[Ag_{25}(MHA)_{14-17}(p-MBA)_{4-1} - 5H]^6$  (the number labeled in the spectra shows the number of  $[Ag_{25}(MHA)_{14-17}(p-MBA)_{4-1} - 5H]^6$  (the number labeled in the spectra shows the number of MHA).

**Notes:** The slight difference between MHA and *p*-MBA in molecular weights leads to the peak overlaps in the ESI mass spectrum of  $Ag_{25}(MHA)_x(p-MBA)_{18-x}$  NCs, thereby it is difficult to identify the exact formula of the species of  $Ag_{25}(MHA)_x(p-MBA)_{18-x}$  NCs. To determine the maximum number of *p*-MBA in  $Ag_{25}(MHA)_x(p-MBA)_{18-x}$  NCs, the shift of the highest peak of  $Ag_{25}(MHA)_{18}$  NCs (i.e.,  $[Ag_{25}(MHA)_{18} - 9H + 4Na]^{6-}$ ) to that of  $Ag_{25}(MHA)_x(p-MBA)_{18-x}$  NCs was compared. Based on the difference of the *m/z* values of the highest peaks, the formula  $[Ag_{25}(MHA)_x(p-MBA)_{18-x}]^{6-}$  was determined accordingly.



**Fig. S13** UV-vis absorption spectra of (a)  $Ag_{25}(MHA)_x(MPA)_{18-x}$  NCs and (b)  $Ag_{25}(MHA)_x(GSH)_{18-x}$  NCs (feeding MHA-to-MPA or -GSH ratio of 1:1) after reaction of 10 min (black curves), 20 min (blue curves) and 30 min (red curves). (c) UV-vis absorption spectra of  $Ag_{25}(MHA)_x(p-MBA)_{18-x}$  NCs (feeding MHA-to-*p*-MBA ratio of 1:1) after reaction of 1 h (black curves), 1.5 h (blue curves) and 2 h (red curves).



**Fig. S14** Zoom-in ESI mass spectra (top panel) and isotope patterns (bottom panel) acquired experimentally (black curves) and theoretically (red curves) of bi-thiolate MHA\_GSH-protected Ag<sub>25</sub> NCs: (a)  $[Ag_{25}(MHA)_{14}(GSH)_4 - 9H + 4Na]^{6-}$  species, (b)  $[Ag_{25}(MHA)_{13}(GSH)_5 - 9H + 4Na]^{6-}$  species. The peak number labeled in top panels represents the number of [+ Na – H].



**Fig. S15** (a) Zoom-in ESI mass spectra (the peak number represents the number of [+ Na - H]) of Ag<sub>25</sub>(MHA)<sub>18</sub> (top panel) and Ag<sub>25</sub>(MHA)<sub>x</sub>(*p*-MBA)<sub>18-x</sub> (bottom panel, feeding MHA-to-*p*-MBA ratio is 1:1 and the reaction time is 1 h). The dotted red lines marked the red shift from  $[Ag_{25}(MHA)_{18} - 9H + 4Na]^{6-}$  to  $[Ag_{25}(MHA)_{10}(p-MBA)_8 - 9H + 4Na]^{6-}$ . (b) Isotope patterns acquired experimentally (black curves) and theoretically (red curves) of  $[Ag_{25}(MHA)_{10}(p-MBA)_8 - 9H + 4Na]^{6-}$ . (c) Isotope patterns acquired experimentally (black curves) and theoretically (color curves) of  $[Ag_{25}(MHA)_{10-15}(p-MBA)_{8-3} - 5H]^6$  (the number shown in the spectra represents the number of MHA). (d) Zoom-in ESI mass spectra (the peak number represents the number of [+ Na - H]) of  $Ag_{25}(MHA)_{18}$  (top panel) and  $Ag_{25}(MHA)_x(p-MBA)_{18-x}$  (bottom panel, feeding MHA-to-*p*-MBA ratio is 1:1 and the reaction time is 1.5 h). The dotted red lines marked the red shift from  $[Ag_{25}(MHA)_{18} - 9H + 4Na]^{6-}$  to  $[Ag_{25}(MHA)_{12}(p-MBA)_6 - 9H + 4Na]^{6-}$ . (e) Isotope patterns acquired experimentally (black curves) and theoretically (cellow curves) of  $[Ag_{25}(MHA)_{18} - 9H + 4Na]^{6-}$  to  $[Ag_{25}(MHA)_{12}(p-MBA)_6 - 9H + 4Na]^{6-}$ . (f) Isotope patterns acquired experimentally (black curves) and theoretically (clack curves) of  $[Ag_{25}(MHA)_{12}(p-MBA)_6 - 9H + 4Na]^{6-}$ . (f) Isotope patterns acquired experimentally (black curves) and theoretically (clack curves) of  $[Ag_{25}(MHA)_{12}(p-MBA)_6 - 9H + 4Na]^{6-}$ . (f) Isotope patterns acquired experimentally (black curves) and theoretically (black curves) of  $[Ag_{25}(MHA)_{12}(p-MBA)_6 - 9H + 4Na]^{6-}$ . (f) Isotope patterns acquired experimentally (black curves) and theoretically (color curves) of  $[Ag_{25}(MHA)_{12-18}(p-MBA)_{6-0} - 5H]^6$  (the number shown in the spectra represents the number of MHA).