SUPPORTING INFORMATION

Esterase-sensitive Sulfur Dioxide Prodrugs Inspired by

Modified Julia Olefination

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Materials and Methods

All chemicals, reagents, and solvents were purchased from commercial suppliers as reagent grade and were used without further purification. NMR spectra were recorded on a Bruker Avance NMR spectrometer at 400 MHz for ¹H spectra and 101 MHz for ¹³C spectra at room temperature. Solvent peaks were used as internal standards. Mass spectral analyses were performed by the GSU Mass Spectrometry Facilities. HPLC analyses were performed on a Shimadzu HPLC equipped with UV detector. Column: Shimadzu C18 3μ m 4.6×50 mm column.

Synthesis

General procedure of the oxidation of sulfide to sulfone:

Sulfone compounds were synthesized following similar procedure with necessary adjustments using 3-chloroperoxybenzoic acid (m-CPBA) as the oxidant.

Generally, 1 equiv. of sulfide was dissolved in dichloromethane (DCM) at a concentration of ~0.5 mmol/10 mL. The resulting solution was treated with 2.5~3 equiv. of m-CPBA in small portions under stirring at room temperature (ice bath can be applied for large scale reaction). Reaction process was monitored by thin layer chromatography (TLC). Usual reaction time is between 1~4 hours. When reaction finished, the reaction was quenched with an equal volume of sat. NaHCO₃ aqueous solution. For small scale reactions, dilution with DCM was used to facilitate separation and washing. The aqueous phase was separated with separatory funnel and back-extracted with fresh DCM twice. Combined organic phase was washed with brine and dried over Na₂SO₄. Solvent was then removed under reduced pressure. Column chromatography was used for purification. Further washing with sat. NaHCO₃ solution was used to remove residual m-CPBA/m-CBA.

2-(2-Benzothiazolylthio)ethanol

2-(Benzo[d]thiazol-2-ylsulfonyl)ethan-1-ol (1)

Synthesis followed the general procedure of oxidation of sulfide to sulfone. Product was obtained as white solid. Yield: 77%. ¹H NMR (CDCl₃) δ 8.21 (d, *J* = 7.9 Hz, 1H), 8.04 (d, *J* = 7.9 Hz, 1H), 7.72 – 7.56 (m, 2H), 4.21 (s, 2H), 3.81 – 3.73 (m, 2H), 2.97 (s, 1H) ppm. ¹³C NMR (CDCl₃) δ 166.2, 152.4, 136.6, 128.3, 127.9, 125.42, 122.4, 57.7, 56.5 ppm. IR: v_{max}/cm⁻¹ 3388 (O-H), 2929 (C-H), 1322 (S=O), 1301 (C-O), 1128 (S=O). HRMS (ESI): calcd. for C₉H₁₀NO₃S₂⁺ [M+H]⁺ 244.0097, found 244.0089.



2-(Benzo[d]thiazol-2-ylthio)-1-phenylethan-1-one (5a)

2-Mercaptobenzothiazole (1.0 g, 6.0 mmol) and 2-bromoacetophenone (1.2 g, 6.0 mmol) were dissolved in 50 mL acetone and treated with K_2CO_3 (0.83 g, 6.0 mmol). The reaction mixture was heated at reflux temperature for 0.5 h and then cooled down to room temperature. Solvent was evaporated under reduced pressure. The residue was re-dissolved in ethyl acetate (EA, ~50 mL) and

washed with brine (~50 mL). Organic phase was then dried over Na₂SO₄, and concentrated *in vacuo*. Residue was recrystallized in ethanol to yield 1.4 g of product as yellow needle crystals. Yield: 82%. ¹H NMR (CDCl₃) δ 8.13 – 8.05 (m, 2H), 7.81 (dd, *J* = 8.1, 0.4 Hz, 1H), 7.75 (dd, *J* = 8.0, 0.6 Hz, 1H), 7.68 – 7.58 (m, 1H), 7.56 – 7.47 (m, 2H), 7.43 – 7.37 (m, 1H), 7.33 – 7.27 (m, 1H), 4.98 (s, 2H) ppm. ¹³C NMR (CDCl₃) δ 193.0, 165.2, 152.9, 135.6, 133.9, 128.8, 128.6, 126.1, 124.4, 121.5, 121.1, 41.0 ppm. HRMS (ESI): calcd. for C₁₅H₁₁NOS₂Na⁺ [M+Na]⁺ 308.0174, found 308.0189.

2-(Benzo[d]thiazol-2-ylthio)-1-phenylpropan-1-one (5b)

To a solution of 2-mercaptobenzothiazole (334 mg, 2 mmol) in 4 mL DCM was added triethylamine (TEA, 0.39 mL, 2.8 mmol). The 2-bromopropiophenone (0.30 mL, 2 mmol) was added dropwise. The reaction mixture was stirred at room temperature for 1 h and then dried under vacuum. The residue was re-dissolved in 10 mL EA and washed with water (10 mL × 2). Water phase was back extracted with 15 mL EA. Combined organic phase was dried over Na₂SO₄, concentrated in vacuo, and purified by column chromatography (EA/Hex 1:20 to 1:10). Product (470 mg) was obtained as yellow oil. Yield: 78%. ¹H NMR (CDCl₃) δ 8.11 (dt, *J* = 8.5, 1.7 Hz, 2H), 7.83 (dd, *J* = 8.1, 0.4 Hz, 1H), 7.75 (dd, *J* = 8.0, 0.6 Hz, 1H), 7.64 – 7.54 (m, 1H), 7.53 – 7.45 (m, 2H), 7.42 (ddd, *J* = 8.3, 7.4, 1.2 Hz, 1H), 7.35 – 7.27 (m, 1H), 5.88 (q, *J* = 7.1 Hz, 1H), 1.77 (d, *J* = 7.1 Hz, 3H) ppm. ¹³C NMR (CDCl₃) δ 197.3, 164.7, 152.9, 135.7, 135.0, 133.6, 128.9, 128.8, 126.1, 124.5, 121.6, 121.1, 47.1, 18.3 ppm. HRMS (ESI): calcd. for C₁₆H₁₃NOS₂Na⁺ [M+Na]⁺ 322.0331, found 322.0352.

2-(Benzo[d]thiazol-2-ylthio)-2-methyl-1-phenylpropan-1-one (5c)

A solution of 2-mercaptobenzothiazole (167 mg, 1 mmol) in 2 mL DCM was added with 0.20 mL TEA (1.4 mmol). To the solution was then added 2-bromo-2-methylpropiophenone (0.17 mL, 1 mmol) dropwise. The reaction mixture was stirred at room temperature for 0.5 h and then heated to 60 °C for 4 h. After cooling to room temperature, solvent was removed by reduced pressure. Residue was re-dissolved in 10 mL EA and washed with water (10 mL × 2). Aqueous phase was back extracted with 15 mL EA. Combined organic phase was dried over Na₂SO₄, concentrated *in vacuo*, and purified by column chromatography (EA/Hex 1:20 to 1:10). 175 mg product was obtained as yellow oil. Yield: 56%. ¹H NMR (CDCl₃) δ 8.13 – 8.05 (m, 2H), 7.89 (d, *J* = 8.1 Hz, 1H), 7.71 (d, *J* = 8.0 Hz, 1H), 7.50 – 7.43 (m, 1H), 7.43 – 7.35 (m, 3H), 7.33 – 7.27 (m, 1H), 1.86 (s, 6H) ppm. ¹³C NMR (CDCl₃) δ 200.6, 161.4, 153.1, 136.7, 136.3, 131.8, 129.1, 127.9, 126.1, 125.0, 122.5, 121.0, 58.1, 27.4 ppm. HRMS (ESI): calcd. for C₁₇H₁₆NOS₂⁺ [M+H]⁺ 314.0668, found 314.0655.

1-(Benzo[d]thiazol-2-ylthio)-3-methylbutan-2-one (5d)

1-Bromo-3-methyl-2-butanone (0.12 mL, 1 mmol) and 2-mercaptobenzothiazole (167 mg, 1 mmol) were dissolved in 5 mL acetone. To the solution was added K₂CO₃ (138 mg, 1 mmol). The reaction mixture was stirred at room temperature for 1 h then dried under reduced pressure. The residue was re-dissolved in 10 mL EA and washed with water (10 mL \times 2). The aqueous phase was back extracted with EA (15mL \times 2). Combined organic phase was dried over Na₂SO₄ and concentrated over vacuum. The crude product was purified by column chromatography (EA/Hex 1:20 to 1:10). 204 mg product was obtained as white solid. Yield: 81%. ¹H NMR (CDCl₃) δ 7.81 (d, J = 8.1 Hz, 1H), 7.74 (d, J = 8.0 Hz, 1H), 7.44 – 7.36 (m, 1H), 7.32 – 7.27 (m, 1H), 4.35 (s, 2H), 2.97 (dt, J = 13.9, 6.9 Hz, 1H), 1.22 (d, J = 6.9 Hz, 6H) ppm. ¹³C NMR (CDCl₃) δ 207.3, 165.3, 152.9, 135.6, 126.1, 124.4, 121.4, 121.1, 41.4, 40.1, 18.4 ppm. HRMS (ESI): calcd. for C₁₂H₁₄NOS₂⁺ [M+H]⁺ 252.0511, found 252.0505.

2-(Benzo[d]thiazol-2-ylthio)cyclohexan-1-one (5e)

To a solution of 2-mercaptobenzothiazole (500 mg, 3.0 mmol) and trimethylamine (0.58 mL, 4.2

mmol) in 6 mL DCM was added 2-chlorocyclohexan-1-one (0.36 mL, 3.1 mmol) slowly. The solution was heated at reflux for 4.5 h, then cooled to room temperature, and stirred overnight. The reaction mixture was diluted with DCM to 15 mL and washed with water (10 mL × 2). The combined aqueous phase was back extracted with DCM (15 mL × 2). Combined organic phase was dried over Na₂SO₄, concentrated in vacuo, and purified by column chromatography (EA/Hex 1:10 to 1:5) to yield the product (454 mg) as yellow solid. Yield: 57%. ¹H NMR (CDCl₃) δ 7.89 (d, *J* = 8.2 Hz, 1H), 7.77 (dd, *J* = 8.0, 0.6 Hz, 1H), 7.50 – 7.39 (m, 1H), 7.38 – 7.30 (m, 1H), 4.93 (dd, *J* = 10.2, 5.7 Hz, 1H), 2.82 – 2.65 (m, 2H), 2.65 – 2.48 (m, 1H), 2.25 – 2.09 (m, 1H), 2.05 – 1.74 (m, 4H) ppm. ¹³C NMR (CDCl₃) δ 205.5, 164.8, 153.1, 135.5, 126.0, 124.4, 121.6, 121.0, 57.4, 41.6, 35.5, 27.6, 25.1 ppm. HRMS (ESI): calcd. for C₁₃H₁₃NOS₂Na⁺ [M+Na]⁺ 286.0331, found 286.0339.

General procedure of reducing β -ketosulfides to β -alkoxysulfides:

Compounds **6a-e** were prepared by reducing compounds **5a-e** with NaBH₄. In general, 1 equiv. of compounds **5a-e** was dissolved in MeOH (10~15 mL/mmol) and cooled to 0 °C in an ice bath. NaBH₄ (4 equiv.) was added to the solution in small portions. The reaction was then allowed to warm up to room temperature. The reaction process was monitored by TLC. After completion, the reaction was quenched with sat. NH₄Cl aqueous solution. The mixture was then extracted with EA and washed with water and brine. The organic phase was dried over Na₂SO₄ and concentrated under reduced pressure. Crude product was purified by column chromatography.

2-(Benzo[d]thiazol-2-ylthio)-1-phenylethan-1-ol (6a)

Synthesis followed the general procedure of reducing β-ketosulfides to β-alkoxysulfides from **5a**. Product was obtained as yellow solid. Yield: 99%. ¹H NMR (CDCl₃) δ 7.92 (d, J = 8.1 Hz, 1H), 7.76 (d, J = 7.9 Hz, 1H), 7.57 – 7.25 (m, 7H), 5.31 – 5.05 (m, 2H), 3.74 (dd, J = 14.3, 2.5 Hz, 1H), 3.58 (dd, J = 14.3, 7.9 Hz, 1H) ppm. ¹³C NMR (CDCl₃) δ 167.9, 152.5, 142.9, 135.5, 128.6, 127.9, 126.3, 125.9, 124.7, 121.4, 121.1, 73.6, 42.8 ppm. HRMS (ESI): calcd. for C₁₅H₁₃NOS₂Na⁺ [M+Na]⁺ 310.0331, found 310.0340.

2-(Benzo[d]thiazol-2-ylthio)-1-phenylpropan-1-ol (6b)

Synthesis followed the general procedure of reducing β -ketosulfides to β -alkoxysulfides from **5b**. Two isomers were obtained in (Ration of 35:65 between the compounds with a high and low R_f) with a total yield of 100 %. Both isomers were obtained as clear oil. Isomer with the lower R_f on TLC was characterized as below and used for the next step. ¹H NMR (CDCl₃) δ 7.95 (d, *J* = 8.1 Hz, 1H), 7.79 (d, *J* = 7.7 Hz, 1H), 7.51 – 7.29 (m, 7H), 4.88 (d, *J* = 7.2 Hz, 1H), 4.12 – 4.02 (m, 1H), 1.45 (d, *J* = 7.2 Hz, 3H) ppm. ¹³C NMR (CDCl₃) δ 166.9, 152.5, 142.5, 135.6, 128.4, 127.9, 126.7, 126.3, 124.8, 121.5, 121.1, 78.5, 52.3, 18.3 ppm. HRMS (ESI): calcd. for C₁₆H₁₆NOS₂⁺ [M+H]⁺ 302.0668, found 302.0661.

2-(Benzo[d]thiazol-2-ylthio)-2-methyl-1-phenylpropan-1-ol (6c)

Synthesis followed the general procedure of reducing β-ketosulfides to β-alkoxysulfides from **5c**. Product obtained as white solid. Yield: 87%. ¹H NMR (CDCl₃) δ 7.99 (d, J = 8.1 Hz, 1H), 7.81 (d, J = 7.6 Hz, 1H), 7.56 – 7.44 (m, 1H), 7.44 – 7.35 (m, 3H), 7.35 – 7.27 (m, 3H), 4.95 (s, 1H), 1.58 (s, 3H), 1.41 (s, 3H) ppm. ¹³C NMR (CDCl₃) δ 165.3, 152.6, 141.0, 136.1, 128.1, 127.7, 127.7, 126.5, 125.2, 121.9, 121.1, 81.7, 60.1, 27.1, 24.1 ppm. HRMS (ESI): calcd. for C₁₇H₁₇NOS₂Na⁺ [M+Na]⁺ 338.0644, found 338.0663.

1-(Benzo[d]thiazol-2-ylthio)-3-methylbutan-2-ol (6d)

Synthesis followed the general procedure of reducing β -ketosulfides to β -alkoxysulfides from **5d**. Product was obtained as clear oil. Yield: 90%. ¹H NMR (CDCl₃) δ 7.84 (d, *J* = 8.1 Hz, 1H), 7.74 (d,

J = 8.0 Hz, 1H), 7.48 – 7.37 (m, 1H), 7.37 – 7.27 (m, 1H), 4.37 (s, br, 1H), 3.76 (ddd, J = 8.2, 6.0, 2.5 Hz, 1H), 3.52 (dd, J = 14.4, 2.5 Hz, 1H), 3.38 (dd, J = 14.4, 7.9 Hz, 1H), 1.88 (dq, J = 13.4, 6.7 Hz, 1H), 1.05 (d, J = 6.8 Hz, 3H), 1.00 (d, J = 6.8 Hz, 3H) ppm. ¹³C NMR (CDCl₃) δ 168.0, 152.5, 135.4, 126.2, 124.6, 121.3, 121.0, 76.6, 38.8, 33.6, 18.7, 17.8 ppm. HRMS (ESI): calcd. for C₁₂H₁₆NOS₂⁺ [M+H]⁺ 254.0668, found 254.0660.

Cis-2-(benzo[d]thiazol-2-ylthio)cyclohexan-1-ol (6e)

Synthesis followed the general procedure of reducing β-ketosulfides to β-alkoxysulfides from **5e**. Both *cis*- and *trans*-products were obtained in about 5:4 ratio (according to crude NMR). Total yield: 91%. *cis*-product was characterized as below and used for further synthesis. ¹H NMR (CDCl₃) δ 7.84 (d, *J* = 8.1 Hz, 1H), 7.73 (d, *J* = 7.9 Hz, 1H), 7.47 – 7.35 (m, 1H), 7.35 – 7.27 (m, 1H), 4.38 – 4.23 (m, 1H), 4.23 – 4.08 (m, 1H), 2.15 – 2.01 (m, 1H), 1.92 (dtd, *J* = 10.3, 6.4, 4.1 Hz, 1H), 1.85 – 1.67 (m, 3H), 1.64 – 1.49 (m, 2H), 1.49 – 1.33 (m, 1H). ¹³C NMR (CDCl₃) δ 167.4, 152.7, 135.4, 126.2, 124.5, 121.4, 121.0, 70.2, 53.9, 31.5, 29.2, 23.9, 21.8 ppm. HRMS (ESI): calcd. for C₁₃H₁₅NOS₂Na⁺ [M+Na]⁺ 288.0487, found 288.0497.

Trans-product characterization: ¹H NMR (CDCl₃) δ 7.86 (d, J = 8.1 Hz, 1H), 7.74 (d, J = 7.9 Hz, 1H), 7.48 – 7.35 (m, 1H), 7.35 – 7.27 (m, 1H), 4.67 (s, 1H), 3.73 – 3.56 (m, 2H), 2.33 – 2.14 (m, 2H), 1.84 – 1.72 (m, 2H), 1.66 – 1.29 (m, 4H) ppm. ¹³C NMR (CDCl₃) δ 167.4, 152.5, 135.6, 126.3, 124.7, 121.6, 121.0, 75.0, 55.6, 35.6, 32.2, 26.1, 24.1 ppm. HRMS (ESI): calcd. for C₁₃H₁₆NOS₂⁺ [M+H]⁺ 266.0668, found 266.0672.

2-(Benzo[d]thiazol-2-ylsulfonyl)-1-phenylethan-1-ol (7a)

Synthesis followed the general procedure of the oxidation of sulfide to sulfone from **6a**. Product was obtained as white solid. Yield: 78%. ¹H NMR (CDCl₃) δ 8.28 – 8.17 (m, 1H), 8.09 – 7.97 (m, 1H), 7.75 – 7.54 (m, 2H), 7.42 – 7.27 (m, 5H), 5.51 (dd, J = 9.7, 1.8 Hz, 1H), 3.94 (dd, J = 14.9, 9.7 Hz, 1H), 3.82 (dd, J = 14.9, 2.1 Hz, 1H), 3.59 (s, 1H) ppm. ¹³C NMR (CDCl₃) δ 166.3, 152.4, 140.3, 136.7, 128. 9, 128.6, 128.3, 127.9, 125.7, 125.5, 122.4, 68.7, 63.0 ppm. IR: v_{max}/cm^{-1} 3347 (O-H), 2930 (C-H), 1334 (S=O) and 1141 (S=O). HRMS (ESI): calcd. for C₁₅H₁₃NO₃S₂⁺ [M+H]⁺ 320.0410, found 320.0400.

2-(Benzo[d]thiazol-2-ylsulfonyl)-1-phenylpropan-1-ol (7b)

Synthesis followed the general procedure of the oxidation of sulfide to sulfone from 6b.

Product was obtained as white solid. Yield: 23% ¹H NMR (CDCl₃) δ 8.33 – 8.18 (m, 1H), 8.09 – 7.95 (m, 1H), 7.73 – 7.63 (m, 1H), 7.63 – 7.57 (m, 1H), 7.40 – 7.28 (m, 5H), 5.18 (d, *J* = 9.1 Hz, 1H), 4.08 – 3.94 (m, 1H), 3.59 (d, *J* = 1.9 Hz, 1H), 1.13 (d, *J* = 7.2 Hz, 3H) ppm. ¹³C NMR (CDCl₃) δ 166.0, 152.7, 139.5, 136.9, 128.9, 128.8, 128.1, 127.7, 127.1, 125.6, 122.3, 74.0, 66.2, 11.9 ppm. IR: ν_{max}/cm^{-1} 3484 (O-H), 2851 (C-H), 1313 (S=O), and 1137(S=O). HRMS (ESI): calcd. for C₁₆H₁₅NO₃S₂Na⁺ [M+Na]⁺ 356.0386, found 356.0381.

2-(Benzo[d]thiazol-2-ylsulfonyl)-2-methyl-1-phenylpropan-1-ol (7c)

Synthesis followed the general procedure of the oxidation of sulfide to sulfone from **6c**. Product was obtained as white solid. Yield: 35%. ¹H NMR (CDCl₃) δ 8.33 – 8.22 (m, 1H), 8.09 – 7.99 (m, 1H), 7.72 – 7.56 (m, 2H), 7.39 – 7.27 (m, 5H), 5.54 (s, 1H), 1.54 (s, 3H), 1.22 (s, 3H) ppm. ¹³C NMR (CDCl₃) δ 164.9, 152.8, 138.0, 137.3, 128.5, 128.2, 128.1, 128.0, 127.8, 125.7, 122.2, 74.8, 70.4, 20.8, 14.4 ppm. IR: ν_{max}/cm^{-1} 3335 (O-H), 1314 (S=O), and 1158 (S=O). HRMS (ESI): calcd. for C₁₇H₁₈NO₃S₂⁺ [M+H]⁺ 348.0723, found 348.0718.

1-(Benzo[d]thiazol-2-ylsulfonyl)-3-methylbutan-2-ol (7d)

Synthesis followed the general procedure of the oxidation of sulfide to sulfone from 6d. Product

was obtained as white solid. Yield: 70%. ¹H NMR (CDCl₃) δ 8.21 (dd, J = 7.5, 1.3 Hz, 1H), 8.02 (dd, J = 7.1, 1.1 Hz, 1H), 7.71 – 7.51 (m, 2H), 4.19 (ddd, J = 9.0, 5.1, 2.1 Hz, 1H), 3.68 (dd, J = 14.7, 2.1 Hz, 1H), 3.61 (dd, J = 14.7, 9.0 Hz, 1H), 1.91 – 1.73 (m, 1H), 0.96 (t, J = 6.7 Hz, 6H) ppm. ¹³C NMR (CDCl₃) δ 166.4, 152.4, 136.6, 128.2, 127.8, 125.4, 122.4, 70.5, 59.5, 33.5, 18.1, 17.1 ppm. IR: ν_{max}/cm^{-1} 3540 (O-H), 2969 (C-H), 1304 (S=O), and 1126(S=O). HRMS (ESI): calcd. for C₁₂H₁₆NO₃S₂⁺ [M+H]⁺ 286.0566, found 286.0561.

Cis-2-(benzo[d]thiazol-2-ylsulfonyl)cyclohexan-1-ol (7e)

Synthesis followed the general procedure of the oxidation of sulfide to sulfone from 6e.

Product was obtained as white solid. Yield: 79%. ¹H NMR (CDCl₃) δ 8.29 – 8.13 (m, 1H), 8.09 – 7.93 (m, 1H), 7.72 – 7.51 (m, 2H), 4.59 (s, 1H), 3.59 – 3.43 (m, 1H), 3.29 (s, 1H), 2.20 – 1.85 (m, 4H), 1.85 – 1.63 (m, 1H), 1.55 – 1.20 (m, 3H). ¹³C NMR (CDCl₃) δ 165.5, 152.7, 136.9, 128.2, 127.8, 125.5, 122.4, 67.2, 63.8, 32.5, 25.1, 19.5, 18.6 ppm. IR: ν_{max}/cm^{-1} 3548 (OH) 2925 (C-H), 1318 (S=O) and 1141 (S=O). HRMS (ESI): calcd. for C₁₃H₁₆NO₃S₂⁺ [M+H]⁺ 298.0566, found 298.0565.



2-(Benzo[d]thiazol-2-ylthio)-1-phenylpropyl acetate (8b)

A solution of **6b** (100 mg, 0.33 mmol) in 2 mL DCM was treated with Ac₂O (0.09 mL, 1 mmol), TEA (0.14 mL, 1 mmol), and *N*,*N*-dimethylaminopyridine (DMAP, 4 mg, 0.03 mmol) under stirring at room temperature. After 1 hour, the reaction mixture was directly dried under reduced pressure and purified by column chromatography (EA/Hex 1:20). Product was obtained as clear oil. Yield: 98%. ¹H NMR (CDCl₃) δ 7.89 (d, *J* = 8.1 Hz, 1H), 7.75 (dd, *J* = 7.9, 0.5 Hz, 1H), 7.42 (dd, *J* = 11.2, 4.1 Hz, 3H), 7.38 – 7.27 (m, 4H), 5.98 (d, *J* = 7.1 Hz, 1H), 4.50 (p, *J* = 7.1 Hz, 1H), 2.03 (s, 3H), 1.42 (d, *J* = 7.0 Hz, 3H) ppm. ¹³C NMR (CDCl₃) δ 169.8, 165.6, 153.2, 137.5, 135.4, 128.6, 128.4, 127.3, 126.1, 124.4, 121.7, 121.0, 77.8, 48.3, 21.0, 18.1 ppm. HRMS (ESI): calcd. for C₁₈H₁₈NO₂S₂⁺ [M+H]⁺ 344.0773, found 344.0768.

2-(Benzo[d]thiazol-2-ylthio)-2-methyl-1-phenylpropyl acetate (8c)

A solution of **6c** (63 mg, 0.20 mmol) in 2 mL DCM was treated with Ac₂O (0.06 mL, 0.60 mmol), TEA (0.08 mL, 0.6 mmol), and DMAP (2 mg, 0.02 mmol) under stirring at room temperature. After 1 hour, the reaction mixture was directly dried under reduced pressure and purified by column chromatography (EA/Hex 1:20). Product was obtained as clear oil. Yield: 90%. ¹H NMR (CDCl₃) δ 8.03 (d, *J* = 7.9 Hz, 1H), 7.80 (dd, *J* = 8.0, 0.5 Hz, 1H), 7.50 – 7.44 (m, 1H), 7.44 – 7.27 (m, 6H), 6.36 (s, 1H), 2.08 (s, 3H), 1.60 (s, 3H), 1.51 (s, 3H) ppm. ¹³C NMR (CDCl₃) δ 169.6, 162. 6, 153.7, 136.9, 136.4, 128.3, 128.2, 127.9, 126.2, 125.1, 122.8, 120.9, 79.7, 56.4, 25.7, 24.3, 21.1 ppm. HRMS (ESI): calcd. for C₁₉H₁₉NO₂S₂Na⁺ [M+Na]⁺ 380.0794, found 380.0761.

2-(Benzo[d]thiazol-2-ylsulfonyl)-1-phenylpropyl acetate (9)

Synthesis procedure followed the general procedure of the oxidation of sulfide to sulfone from **8b**. Product obtained as pale white oil. Yield: 83%. ¹H NMR (CDCl₃) δ 8.27 (dd, *J* = 8.1, 0.5 Hz, 1H), 8.03 (dd, *J* = 8.0, 0.6 Hz, 1H), 7.72 – 7.63 (m, 1H), 7.63 – 7.55 (m, 1H), 7.39 – 7.28 (m, 5H), 6.05 (d, *J* = 9.7 Hz, 1H), 4.35 (dq, *J* = 9.7, 7.3 Hz, 1H), 1.53 (s, 3H), 1.26 (d, *J* = 7.3 Hz, 3H) ppm. ¹³C NMR (CDCl₃) δ 168. 7, 166.9, 152. 8, 136.7, 136.6, 129.2, 128.9, 128.1, 127.8, 127.6, 125.5, 122.27, 74.9, 62.8, 20. 6, 10.6 ppm. IR: v_{max}/cm⁻¹ 2941 (C-H), 1745 (C=O), 1317 (S=O), 1221 (C-O), and

1145 (S=O). HRMS (ESI): calcd. for C₁₈H₁₈NO₄S₂⁺ [M+H]⁺ 376.0672, found 376.0666.

2-(Benzo[d]thiazol-2-ylsulfonyl)-2-methyl-1-phenylpropyl acetate (10)

Synthesis followed the general procedure of the oxidation of sulfide to sulfone from **8c**. Product was obtained as white solids. Yield: 85%. ¹H NMR (CDCl₃) δ 8.27 (d, *J* = 7.7 Hz, 1H), 8.02 (d, *J* = 7.6 Hz, 1H), 7.69 – 7.62 (m, 1H), 7.60 (td, *J* = 7.7, 1.3 Hz, 1H), 7.36 – 7.24 (m, 5H), 6.29 (s, 1H), 1.76 (s, 3H), 1.66 (s, 3H), 1.31 (s, 3H) ppm. ¹³C NMR (CDCl₃) δ 168.7, 165.3, 153.1, 137.3, 135.8, 128.8, 128.4, 128.1, 127.8, 127.7, 125.7, 122.2, 75.7, 69.1, 20.7, 20.3, 16.5 ppm. IR: v_{max}/cm⁻¹ 2992 (C-H), 1748 (C=O), 1312 (S=O) 1225 (C-O), 1153 (S=O). HRMS (ESI): calcd. for C₁₉H₁₉NO₄S₂Na⁺ [M+Na]⁺ 412.0648, found 412.0651.



2-(Benzo[d]thiazol-2-ylthio)-2-methyl-1-phenylpropyl propionate (15)

A solution of **6c** (150 mg, 0.48 mmol) was dissolved in 1 mL DCM. To the solution was added DMAP (6 mg, 0.05 mmol), TEA (0.13 mL, 0.96 mmol), and propionyl chloride (0.05 mL, 0.58 mmol) sequentially under stirring. The reaction mixture was heated to 60 °C for 8 hours and then stirred at room temperature overnight. The reaction mixture was directly dried over reduced pressure and purified by column chromatography (EA/Hex 1:20). Product was obtained as clear oil. Yield: 52%. ¹H NMR (CDCl₃) δ 8.04 (d, *J* = 7.8 Hz, 1H), 7.80 (dd, *J* = 8.0, 0.5 Hz, 1H), 7.53 – 7.27 (m, 7H), 6.38 (s, 1H), 2.48 – 2.27 (m, 2H), 1.61 (s, 3H), 1.52 (s, 3H), 1.15 (t, *J* = 7.6 Hz, 3H) ppm. ¹³C NMR (CDCl₃) δ 172.9, 162.6, 153.7, 137.1, 136.4, 128.2, 128.2, 127.9, 126.2, 125.1, 122.8, 120.9, 79.4, 56.5, 27. 8, 25.6, 24.4, 9.1 ppm. HRMS (ESI): calcd. for C₂₀H₂₁NO₂S₂Na⁺ [M+Na]⁺ 394.0906, found 394.0905.

2-(Benzo[d]thiazol-2-ylsulfonyl)-2-methyl-1-phenylpropyl propionate (11)

Synthesis followed the general procedure of the oxidation of sulfide to sulfone from **11**. Product was obtained as clear oil. Yield: 81%. ¹H NMR (CDCl₃) δ 8.26 (d, *J* = 7.7 Hz, 1H), 8.01 (d, *J* = 7.6 Hz, 1H), 7.70 – 7.61 (m, 1H), 7.61 – 7.54 (m, 1H), 7.36 – 7.23 (m, 5H), 6.31 (s, 1H), 2.02 – 1.79 (m, 2H), 1.76 (s, 3H), 1.30 (s, 3H), 0.89 (t, *J* = 7.5 Hz, 3H) ppm. ¹³C NMR (CDCl₃) δ 172.0, 165.3, 153.1, 137.2, 135.9, 128.8, 128.4, 128.1, 127.8, 127.7, 125.7, 122.1, 75.4, 69.1, 27.3, 20.4, 16.5, 8.5 ppm. IR: v_{max}/cm⁻¹ 2985 (C-H), 1746 (C=O), 1319 (S=O), 1151 (S=O). HRMS (ESI): calcd. for C₂₀H₂₁NO₄S₂Na⁺ [M+Na]⁺ 426.0804, found 426.0821.





Compound **6c** (200 mg, 0.63 mmol) and DMAP (7 mg, 0.06 mmol) were dissolved in 2 mL DCM. To this solution was added TEA (0.18 mL, 1.26 mmol) and pivaloyl chloride (0.09 mL, 0.73 mmol) slowly under stirring. The reaction mixture was heated at 60 °C for 8 hours, then cooled to room temperature, and stirred overnight. The reaction mixture was diluted with 10 mL DCM, and washed with 8 mL water. The aqueous phase was back extracted with DCM (10 mL \times 2). Combined organic phase was dried over Na₂SO₄, concentrated in vacuo, and purified by column chromatography (DCM/Hex 1:2 to 1:1 to pure DCM). Product was obtained as clear oil. Yield: 63%. ¹H NMR (CDCl₃)

δ 8.03 (d, J = 7.8 Hz, 1H), 7.80 (dd, J = 8.0, 0.5 Hz, 1H), 7.52 – 7.28 (m, 7H), 6.35 (s, 1H), 1.61 (s, 3H), 1.52 (s, 3H), 1.26 (s, 9H) ppm. ¹³C NMR (CDCl₃) δ 176.6, 162.6, 153.8, 137.1, 136.3, 128.2, 128.1, 127.8, 126.1, 125.0, 122.8, 120.9, 79.09, 56.7, 39.0, 27.2, 25.5, 24.4 ppm. HRMS (ESI): calcd. for C₂₂H₂₆NO₂S₂⁺ [M+H]⁺ 400.1399, found 400.1384.

2-(Benzo[d]thiazol-2-ylsulfonyl)-2-methyl-1-phenylpropyl pivalate (12)

Synthesis followed the general procedure of the oxidation of sulfide to sulfone from **15**. Product yielded as white solid. Yield: 86%. ¹H NMR (CDCl₃) δ 8.32 – 8.20 (m, 1H), 8.09 – 7.93 (m, 1H), 7.72 – 7.53 (m, 2H), 7.37 – 7.27 (m, 5H), 6.33 (s, 1H), 1.77 (s, 3H), 1.23 (s, 3H), 1.15 (s, 9H) ppm. ¹³C NMR (CDCl₃) δ 176.1, 164.6, 153.2, 137.3, 136.0, 128.8, 128.4, 128.1, 127.9, 127.6, 125.8, 122.2, 74.9, 68.6, 38.8, 26.9, 21.6, 16.4 ppm. IR: v_{max} /cm⁻¹ 2975 (C-H), 1734 (C=O), 1321 (S=O), 1139 (S=O). HRMS (ESI): calcd. for C₂₂H₂₆NO₄S₂⁺ [M+H]⁺ 432.1298, found 432.1284.



2-(Benzo[d]thiazol-2-ylthio)-2-methyl-1-phenylpropyl 2-chloroacetate (17)

Compound **6c** (140 mg, 0.44 mmol) was dissolved in 5 mL DCM. To this solution was added TEA (0.11 mL, 0.80 mmol). The solution was cooled to 0 °C in ice bath. 2-Chloroacetyl chloride (0.04 mL, 0.50 mmol) was then added slowly to the solution. The reaction mixture was stirred at 0 °C for 90 min. Then 5 mL water was added into the reaction mixture. The aqueous layer was extracted with DCM (10 mL × 3). Combined organic phase was washed with brine, dried over Na₂SO₄, concentrated, and purified by column chromatography (EA/Hex 1:40 to 1:20). Product was obtained as clear oil. Yield: 50%. ¹H NMR (CDCl₃) δ 8.03 (dd, *J* = 8.1, 0.4 Hz, 1H), 7.81 (dd, *J* = 8.0, 0.6 Hz, 1H), 7.53 – 7.45 (m, 1H), 7.45 – 7.28 (m, 6H), 6.54 (s, 1H), 4.10 (s, 2H), 1.61 (s, 3H), 1.51 (s, 3H) ppm. ¹³C NMR (CDCl₃) δ 165.8, 162.3, 153.7, 136.2, 136.1, 128.6, 128.1, 128.0, 126.2, 125.2, 122.8, 121.0, 81.2, 56.1, 41.0, 25.4, 24.1 ppm. HRMS (ESI): calcd. for C₁₉H₁₈CINO₂S₂Na⁺ [M+Na]⁺ 414.0360, found 414.0383.

2-(Benzo[d]thiazol-2-ylsulfonyl)-2-methyl-1-phenylpropyl 2-chloroacetate (18)

Synthesis followed the general procedure of the oxidation of sulfide to sulfone from **16**. Product was obtained as white solid. Yield: 88%. ¹H NMR (CDCl₃) δ 8.35 – 8.17 (m, 1H), 8.10 – 7.91 (m, 1H), 7.76 – 7.56 (m, 2H), 7.42 – 7.27 (m, 5H), 6.41 (s, 1H), 3.88 – 3.68 (m, 2H), 1.73 (s, 3H), 1.31 (s, 3H) ppm. ¹³C NMR (CDCl₃) δ 165.2, 164.7, 153.0, 137.3, 134.8, 129.2, 128.6, 128.3, 127.9, 127.8, 125.7, 122.3, 77.1, 69.0, 40.8, 20.4, 16.3 ppm. IR: v_{max}/cm^{-1} 2989 (C-H), 1743 (C=O), 1313 (S=O), 1152 (S=O). HRMS (ESI): calcd. for C₁₉H₁₈ClNO₄S₂Na⁺ [M+Na]⁺ 446.0258, found 446.0271.

2-(Benzo[d]thiazol-2-ylsulfonyl)-2-methyl-1-phenylpropyl dimethylglycinate (13)

Compound 17 (100 mg, 0.24 mmol) was dissolved in 5 mL acetone and added with K₂CO₃ (66 mg, 0.48 mmol). Dimethylamine (2.0 M in methanol, 0.24 mL, 0.48 mmol) was added into this solution and then the reaction mixture was stirred overnight. Subsequently, the reaction mixture was filtered. The filtrate was concentrated under reduced pressure. Residue was purified by column chromatography (EA/Hex 1:4 to 1:1). Product was obtained as yellowish solid. Yield: 62%. ¹H NMR (CDCl₃) δ 8.27 (d, *J* = 7.8 Hz, 1H), 8.01 (d, *J* = 7.5 Hz, 1H), 7.72 – 7.62 (m, 1H), 7.62 – 7.50 (m, 1H), 7.36 – 7.26 (m, 5H), 6.33 (s, 1H), 2.86 (d, *J* = 16.9 Hz, 1H), 2.71 (d, *J* = 16.9 Hz, 1H), 2.14 (s, 6H), 1.76 (s, 3H), 1.30 (s, 3H) ppm. ¹³C NMR (CDCl₃) δ 168.5, 165.2, 153.1, 137.2, 135.6, 128.9

128.4 128.1, 127.85, 127.7 125.8 122.2, 75.5, 69.0, 59.6, 44.9, 20.5, 16.4 ppm. IR: v_{max}/cm^{-1} 2934 (C-H), 1742 (C=O), 1320 (S=O), 1110 (S=O). HRMS (ESI): calcd. for $C_{21}H_{25}N_2O_4S_2^+$ [M+H]⁺ 433.1250, found 433.1244.

Confirmation of SO2 release by DTNB test

DTNB test was adopted from a published method.¹ **7b** and **7c** were prepared as 500-µM DMSO stock solutions. DTNB was prepared as 15-mM EtOH stock solution. NaHSO₃ as positive control was prepared as a 1-mM PBS (pH 7.4) stock solution and stored on ice before use. 2-OHBT and 2-methyl-1-phenyl-propene as negative controls were prepared as 500-µM DMSO stock solutions respectively.

Each group (200 μ L total volume) was prepared by adding 20 μ L stock solution into the wells of a 96-well plate followed by addition of 180 μ L PBS. Each final solution contained 10% DMSO. The plate was incubated at 37 °C for 0.5 h with gentle shaking on a Barnstead shaker. Then 20 μ L DTNB was added into each well. After incubating at room temperature for another 15 min, the UV absorption was read by a PerkinElmer multiplate reader at 405 nm (n=3).



Figure S1. DTNB test results. **: *p* < 0.01.

Kinetics study of compound 7a-e by HPLC

Compounds **7a-d** were prepared as 500- μ M DMSO stock solution at room temperature. Test solution (50 μ M) was prepared in 8 mL glass vial by adding 200 μ L of a DMSO stock solution into 1800 μ L PBS and the resulting solution was incubated immediately in a water bath at 37 °C. 100 μ L of samples for analysis were taken at random time points (n = 10) and were mixed with 300 μ L ACN in a 1.5-mL eppendorf tube and stored at -80 °C. Frozen samples were thawed and centrifuged with benchtop microcentrifuge shortly to allow salt precipitation. Supernatant was subjected to HPLC analysis. Standards of 2-OHBT, styrene, and substituted styrenes were purchased from

commercial vendors. k_{obs} was calculated using 2-OHBT peak areas. All kinetic runs were in triplicates. Curve fitting was conducted using the SigmaPlot 10 software (Figure S2).

HPLC conditions:

Binary solvent system: Solvent A: H₂O with 0.05% TFA; Solvent B: ACN

Gradient elution: Solvent B 15% to 60%

UV detector wavelength: 254 nm

Injection volume: 20 µL



Figure S2 Kinetics studies of compounds **7a-e** by measuring 2-OHBT peak areas using HPLC. (A) An example of HPLC chromatogram obtained by HPLC. Chromatogram showing 50 μM **7a** in 10% DMSO/PBS (pH 7.4) at 37 °C at the 120-min time point. Peak at 3.992 min: 2-OHBT; peak at 7.652 min: compound **7a**; peak at 9.086 min: styrene. (B) Examples of first-order reaction monitored by measuring 2-OHBT peak areas. Upper left: **7a**; upper right: **7b**; lower left: **7c**; lower right: **7d**.

Compound 7e was prepared as a 20-mM DMSO stock solution at room temperature. Test solution

 $(200 \ \mu\text{M})$ was prepared in 1.5 mL eppendorf tube by adding 10 μ L of the DMSO stock solution into 990 μ L PBS followed by incubation in a water bath at 37 °C. Samples for HPLC analyses were taken at random time points and subjected to HPLC analysis directly. 2-OHBT standard was pretested for retention time (~3.8 min). Kinetic runs were in triplicates (Figure S3).

HPLC conditions:

Binary solvent system: Solvent A: H₂O with 0.05% TFA; Solvent B: ACN

Gradient elution: Solvent B 15% to 40%

UV detector wavelength: 254 nm

Injection volume: 20 µL

0 min:



Overlay (0~196 min):



Figure S3. An example of reaction of 7e monitored by HPLC. Peaks at 8.87~8.88 min: compound 7e.

Stability study of compound 9, 10, and 13 by HPLC

Compound **9** was prepared as a 5-mM DMSO stock solution at room temperature. Test solution (50 μ M) was prepared in a 1.5 mL eppendorf tube by adding 10 μ L of the DMSO stock solution into 990 μ L PBS followed by incubation at 37 °C in a water bath. Samples for HPLC analysis were taken at random time points (n = 10) and subjected to HPLC analysis directly. Stability tests were in duplicates (Figure S4).

HPLC conditions:

Binary solvent system: Solvent A: H₂O with 0.05% TFA; Solvent B: ACN

Gradient elution: Solvent B 15% to 60%

UV detector wavelength: 254 nm

Injection volume: 20 µL



Figure S4 An example of stability tests of compound 9 monitored by HPLC. Peaks at 10.07~10.08 min: compound 9; shoulder peaks and peaks at 10.87~10.89 min: elimination product.

Compound **10** was prepared as 100- μ M DMSO stock solution at room temperature. Test solution (10 μ M) was prepared in an 8-mL glass vial by adding 500 μ L of the DMSO stock solution into 4.5 mL PBS followed by incubation at 37 °C in a water bath with stirring. Samples for HPLC analysis were taken at random time points (n = 5) and subjected to HPLC analysis directly. Stability tests were in duplicates (Figure S5).

HPLC conditions:

Binary solvent system: Solvent A: H₂O with 0.05% TFA; Solvent B: ACN

Gradient elution: Solvent B 15% to 60%

UV detector wavelength: 254 nm

Injection volume: 20 μ L



Overlay (0~240 min):



Figure S5 An example of stability tests of compound 10 monitored by HPLC. Peaks at 10.60~10.61 min: compound 10

A 5-mM stock solution of Compound 13 was prepared at room temperature in DMSO. Test solution (50 µM) was prepared in an 8-mL glass vial by adding 20 µL of the DMSO stock solution into 1980 µL of PBS followed by incubation at 37 °C in a water bath. Samples for HPLC analysis were taken at random time points (n = 5) and subjected to HPLC analysis directly. Stability tests were in duplicates (Figure S6).

HPLC conditions:

Binary solvent system: Solvent A: H₂O with 0.05% TFA; Solvent B: ACN

Gradient elution: Solvent B 15% to 60%

UV detector wavelength: 254 nm

Injection volume: 20 µL

0 h:



97 h:



Figure S6 An example of stability studies of compound 13 monitored by HPLC. Peaks at 3.94~3.95min: 2-OHBT; peaks at 7.07~7.17 min: compound 13; peaks at 11.62~11.63 min: 2-methyl-1-phenyl-propene.

Kinetics study of esterase-triggered SO2 release of compound 10-13 by HPLC

Stock solutions (500 μ M) of compounds **10-13** were prepared in DMSO at room temperature. Porcine liver esterase (PLE, 18 unit/mg) was prepared as 0.1 unit/ μ L PBS stock solution. Test solution (50 μ M with 5 unit/mL PLE) was prepared in an 8-mL glass vial by first adding 200 μ L of the DMSO stock solution into 1700 μ L of PBS and thoroughly mixed. Then to the solution was added 100 μ L of the PLE stock solution followed by incubation at 37 °C in a water bath. 100 μ L samples for analysis were taken every 10 minutes (n=12) and mixed with 300 μ L ACN in a 1.5-mL eppendorf tube and stored at -80 °C. Frozen samples were thawed and centrifuged with benchtop micro-centrifuge at 14.5×10³ rpm for 4 min to allow esterase and salt to precipitate. Supernatant was subjected to HPLC analysis. 2-OHBT, styrene, and substituted styrenes were used as standard and pre-tested for retention time. All kinetic runs were in triplicates. Microsoft Excel was used for plotting. Kinetics studies of the esterase-triggered SO₂ release from compound **13** in 1% DMSO/PBS with 1 unit/mL PLE used the same method. Stock solutions were prepared accordingly (Figure 1). <u>HPLC conditions:</u>

Binary solvent system: Solvent A: H₂O with 0.05% TFA; Solvent B: ACN

Gradient elution: Solvent B 15% to 60%

UV detector wavelength: 254 nm

Injection volume: 20 µL

<u>Kinetics studies of esterase-triggered SO₂ release from compound 10-13 as examined with</u> <u>fluorescent probe 14</u>

Standard curve:

500 μ M probe 14 DMSO stock solution was prepared at room temperature. NaHSO₃ solutions of different concentrations were prepared in PBS. The test solution was prepared in a 4 mL (10 mm×10 mm) quartz cuvette by adding 300 μ L probe DMSO stock solution into 2700 μ L NaHSO₃ solution. Final solution contains 50 μ M probe 14 and 10% DMSO. The test solution was sealed with a cap, thoroughly mixed, and incubated at room temperature for 2 min. Fluorescent emission at 465 nm was then recorded on a Shimadzu RF- 5301PC fluorimeter. (λ_{ex} =400 nm; slit width: ex: 5nm, em: 3 nm) Experiments were conducted in triplicates. Standard curves were fitted with Microsoft Excel 2016 (Figure S7(A)).

Studies of reaction kinetics:

Stock solutions (1 mM) of compounds **10-13** were prepared in DMSO at room temperature. PLE (18 unit/mg) was prepared as 0.1 unit/ μ L PBS stock solution. Probe **14** was prepared as 1 mM DMSO stock solution. Test solution (50 μ M prodrug with 50 μ M probe and 5 unit/mL PLE) was prepared by adding 150 μ L of the prodrug DMSO stock solution, 150 μ L of the probe **14** DMSO stock solution, 2550 μ L of PBS, and 150 μ L PLE stock solution sequentially into a 4 mL (10 mm×10 mm) quartz cuvette. The test solution was sealed with a cap, and thoroughly mixed; and fluorescent emission at 465 nm was recorded every 10 minutes (n=13) (λ_{ex} =400 nm; slit width: ex: 5nm, em: 3 nm). Fluorescence intensities obtained were converted to NaHSO₃ concentrations using the standard curve described above. All runs were in triplicates (Figure S7(B)).



Figure S7 Kinetics study of esterase-triggered SO2 release from compound 10-13 by fluorescent probe 14. (A) Standard curve. (B) Kinetics of esterase-triggered SO2 release from compounds 10-13 monitored with fluorescent probe 14.

Cell imaging study

Cell culture:

HeLa cells were maintained in Dulbecco's Modified Eagle's Medium (DMEM) supplemented with

10 % fetal bovine serum and 1% penicillin-streptomycin at 37 °C with 5 % CO₂. The media was changed every other day. Experiment was done within 10 passages of Hela cells. <u>Cell imaging:</u>

HeLa cells were pre-seeded onto coverslips in 6-well plate a day before experiment. A 2-mM DMSO stock solution of probe **14** was prepared. Compound **13** was prepared as 20-mM DMSO stock solution. Cells were first treated with compound **13** to make a final concentration of 100 μ M (0.5% DMSO). 400 μ M NaHSO₃ was used as a positive control. A mixture containing 100 μ M 2-OHBT and 100 μ M 2-methyl-1-phenyl-propene was used as the negative control. Cell only group was also used as negative controls. All final media contained 0.5% DMSO. The cells were incubated at 37 °C for 2 hours and washed with PBS once. 2 mL of fresh DMEM media was added into each well. The cells were then treated with probe **14** to give a final concentration of 10 μ M. Cells were incubated for another 0.5 h at 37 °C. After washing twice with PBS, the cells were fixed with 4% paraformaldehyde for 30 min at room temperature. The cells were then washed once with PBS. Then 25 mM glycine in PBS was added to quench the extra formaldehyde. After storing at 4 °C overnight. The glycine solution was discarded. All coverslips were immersed in DI water and mounted onto glass slides using DAPI-free mounting media (ProLong Live Antifade Reagent; P36974). Fluorescent imaging was performed on a Zeiss fluorescent microscope using the DAPI channel (λ_{ex} : 358 nm, λ_{em} : 461 nm).

References

1. Li, Y.; Zhao, M., Simple methods for rapid determination of sulfite in food products. *Food Control* **2006**, *17* (12), 975-980.