Direct diversification of unmasked quinazolin-4(3*H*)-ones through orthogonal reactivity modulation

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Supplementary Information

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I. General Information

All reagents were purchased from standard suppliers (Sigma-Aldrich, Alfa Aesar, or TCI) and were used without further purification. All reactions were performed in oven-dried Schlenk tubes (capacity, 10 mL). ¹H, ¹³C, and ¹⁹F NMR spectra were recorded in DMSO- d_6 or CDCl₃ on a Bruker Avance III HD (400 MHz for proton, 100 MHz for carbon, 376 MHz for fluorine). All chemical shifts are given on the δ -scale in ppm, and residual solvent peaks were used as references. High resolution mass spectrometry (HRMS) analyses were performed on a Thermo Scientific Q Exactive Plus Hybrid Quadrupole-Orbitrap mass spectrometer. Thin layer chromatography (TLC) was carried out on Merck Kieselgel 60 F₂₅₄ glass plates. Flash column chromatography was carried out using Merck Silica gel 60 silica.

II. Substrate Synthesis and Characterization



Substrates 1a-1p

General Procedure: Synthesis of 2-arylquinazolin-4(3H)-one (1a,1b,1d-1i,1k-1p)

2-Arylquinazolin-4(3*H*)-ones were prepared according to the previously reported method.^{S1} To a two-neck round bottom flask were added substituted 2-halobenzoic acid (8.55 mmol, 1.0 equiv) and benzamidine hydrochloride (12.77 mmol, 1.5 equiv) in DMF (20 mL). The reaction flask was evacuated, back-filled with nitrogen several times, and then vigorously stirred for 10 min. Cs_2CO_3 (17.03 mmol, 2.0 equiv) was added to the reaction mixture. After 15 min, Cul (1.70 mmol, 0.2 equiv) was subsequently added and stirred at room temperature for 12 h under a nitrogen atmosphere. After completion of the reaction, the reaction mixture was filtered. The solvent residue was concentrated *in vacuo* and the residue was purified by flash column chromatography.

Procedure for the synthesis of 2-(4-methoxyphenyl)quinazolin-4(3H)-one (1c)

2-(4-Methoxyphenyl)quinazolin-4(3*H*)-one was prepared according to the previously reported method.^{S2} To a round bottom flask were added anthranilamide (20 mmol, 1.0 equiv), 4-methoxybenzaldehyde (20 mmol, 1.0 equiv) and Cu₂O (0.6 mmol, 0.3 equiv) in DMA (60 mL), and stirred at 120 °C for 24 h under air. After completion of the reaction, the reaction mixture was filtered. The solvent residue was concentrated *in vacuo* and the residue was purified by flash column chromatography.

Procedure for the synthesis of 2-(4-fluorophenyl)quinazolin-4(3H)-one (1j)

2-(4-Fluorophenyl)quinazolin-4(3*H*)-one was prepared according to the previously reported method.^{S3} To a round bottom flask were added anthranilamide (20 mmol, 1.0 equiv), 4-fluorobenzyl alcohol (20 mmol, 1.0 equiv), and KOH (40 mmol, 2.0 equiv) in toluene (80 mL), and stirred at 90 °C for 20 h under air. After completion of the reaction, the solvent residue was concentrated *in vacuo* and the residue was purified by flash column chromatography.

*1a,^{S1} 1b,^{S4} 1c,^{S2} 1d,^{S5} 1e,^{S5} 1g,^{S6} 1h,^{S5} 1i,^{S6} 1j,^{S3} 1k,^{S5} 1l,^{S5} and 1m^{S5} have been reported.

8-Nitro-2-phenylquinazolin-4(3*H*)-one (**1f**)



Yellowish solid; **ATR-FTIR** (cm⁻¹): 3167, 3120, 3076, 1670, 1602, 1566, 1528, 1479, 1365, 1287, 1156, 1060, 966, 889, 825, 766, 692; ¹**H NMR** (400 MHz, DMSO- d_6) δ 12.95 (s, 1H), 8.38 (d, J = 7.9, 1.4 Hz, 1H), 8.33 – 8.29 (m, 1H), 8.16 (d, J = 7.6 Hz, 2H), 7.68 – 7.54 (m, 4H); ¹³**C NMR** (100 MHz, DMSO- d_6) δ 161.38, 155.05, 147.10, 140.99, 132.64, 132.38, 130.15, 129.19, 129.00, 128.67, 128.59, 126.43,

122.97; **HRMS-ESI**: m/z [M+H]⁺ calcd for C₁₄H₁₀N₃O₃: 268.0717; found: 268.0713.

6,7-Dimethyl-2-phenylquinazolin-4(3H)-one (1n)



White solid; **ATR-FTIR** (cm⁻¹): 3048, 2969, 2917, 1666, 1623, 1602, 1566, 1490, 1456, 1349, 1306, 1292, 1193, 1163, 1027, 998, 940, 851, 837, 776, 694; ¹**H NMR** (400 MHz, DMSO- d_6) δ 12.36 (s, 1H), 8.15 (dt, J = 8.2, 1.8 Hz, 2H), 7.90 (s, 1H), 7.55 (dd, J = 5.2, 2.8 Hz, 4H), 2.39 (s, 3H), 2.37 (s, 3H); ¹³**C NMR** (100 MHz, DMSO- d_6) δ 162.49, 151.96, 147.65, 144.92, 136.32, 133.31, 131.63,

 $129.05,\ 128.03,\ 126.02,\ 119.21,\ 20.39,\ 19.77;\ \textbf{HRMS-ESI}:\ m/z\ [M+H]^+\ calcd\ for\ C_{16}H_{15}N_2O:\ 251.1179;\ found:\ 251.1174.$

7-Chloro-5-methyl-2-phenylquinazolin-4(3*H*)-one (**1o**)



White solid; **ATR-FTIR** (cm⁻¹): 3196, 3166, 3114, 2960, 2923, 2359, 1660, 1590, 1610, 1555, 1513, 1478, 1451, 1427, 1299, 1210, 1194, 1164, 1135, 1089, 1043, 986, 934, 890, 869, 849, 819, 777, 715, 693;¹**H NMR** (400 MHz, DMSO- d_6) δ 12.44 (s, 1H), 8.19 – 8.13 (m, 2H), 7.57 (dt, *J* = 15.9, 6.8 Hz, 4H), 7.35 (s, 1H), 2.80 (s, 3H); ¹³**C NMR** (100 MHz, DMSO- d_6) δ 162.92, 153.75, 151.87, 143.13,

138.37, 132.59, 132.12, 129.08, 128.89, 128.28, 125.08, 118.63, 22.68; **HRMS-ESI**: $m/z [M+H]^+$ calcd for $C_{15}H_{12}CIN_2O$: 271.0633; found: 271.0629.

7-Fluoro-5-methyl-2-phenylquinazolin-4(3*H*)-one (1p)



White solid; **ATR-FTIR** (cm⁻¹): 3167, 3093, 3066, 2962, 2924, 2360, 2341, 1656, 1596, 1565, 1482, 1451, 1409, 1384, 1343, 1306, 1194, 1155, 1123, 1101, 1042, 1001, 979, 930, 862, 835, 780, 695;¹**H NMR** (400 MHz, DMSO- d_6) δ 12.40 (s, 1H), 8.21 – 8.12 (m, 2H), 7.62 – 7.51 (m, 3H), 7.31 (dd, J = 9.6, 2.5 Hz, 1H), 7.18 (d, J = 9.8 Hz, 1H), 2.82 (s, 3H); ¹³**C NMR** (100 MHz, DMSO- d_6) δ 164.91 (d, J =

250.4 Hz), 162.77, 153.63, 152.98 (d, J = 13.4 Hz), 144.51 (d, J = 10.8 Hz), 132.60, 132.08, 129.07, 128.27, 117.20 (d, J = 22.9 Hz), 116.87, 110.94 (d, J = 20.8 Hz), 23.07; ¹⁹**F NMR** (376 MHz, DMSO- d_6) δ -105.88; **HRMS-ESI**: m/z [M+H]⁺ calcd for C₁₅H₁₂FN₂O: 255.0928; found: 255.0923.

III. Products Synthesis and Characterization

III.1 Optimization for the Synthesis of 3a





Entry	Pd catalyst variations	Yield ⁱ	° (%)
		3a	3a'
1	-	37	-
2	$(PPh_3)_2PdCl_2$ instead of $Pd(OAc)_2$	-	-
3	PEPPSI-IPr instead of Pd(OAc) ₂	13	-
4	$Pd(TFA)_2$ instead of $Pd(OAc)_2$	17	-
5	PEPPSI-SIPr instead of Pd(OAc) ₂	28	-
6	30 mol% Pd(OAc) ₂ instead of 10 mol% Pd(OAc) ₂	49	0
7	40 mol% Pd(OAc) ₂ instead of 10 mol% Pd(OAc) ₂	73	3
8	50 mol% Pd(OAc) ₂ instead of 10 mol% Pd(OAc) ₂	82	5
9	70 mol% Pd(OAc) ₂ instead of 10 mol% Pd(OAc) ₂	77	8

^a The initial conditions: **1a** (0.09 mmol, 1.0 equiv), Ph₂IOTf (0.09 mmol, 1.0 equiv), Pd(OAc)₂ (10 mol%), KOAc (0.27 mmol, 3.0 equiv), DMF (2.0 mL), 130 °C, 12 h, air. ^b Yield determined by GC analysis using *n*-dodecane as an internal standard.

Table S2 Base screening^a



Entry	Base variations	Yield	^b (%)
	-	3a	3a'
1		82	5
2	CsOAc instead of KOAc	75	5
3	NaOAc instead of KOAc	68	8
4	K ₂ CO ₃ instead of KOAc	67	20
5	2.0 equiv KOAc instead of 3.0 equiv KOAc	70	6
6	2.5 equiv KOAc instead of 3.0 equiv KOAc	74	4

^a The initial conditions: **1a** (0.09 mmol, 1.0 equiv), Ph₂IOTf (0.09 mmol, 1.0 equiv), Pd(OAc)₂ (50 mol%), KOAc (0.27 mmol, 3.0 equiv), DMF (2.0 mL), 130 °C, 12 h, air. ^b
Yield determined by GC analysis using *n*-dodecane as an internal standard.

Table S3 Solvent and temperature screening^a



Entry	Solvent and temperature variations	Yield	d⁵ (%)
		3a	3a'
1		82	5
2	Toluene instead of DMF	37	2
3	DMSO instead of DMF	72	2
4	DMA instead of DMF	82	6
5	MeCN at 100 °C instead of DMF at 130 °C	17	-
6	DMC at 100 $^\circ\text{C}$ instead of DMF at 130 $^\circ\text{C}$	23	2
7	THF at 100 °C instead of DMF at 130 °C	20	1
8	Dioxane at 100 °C instead of DMF at 130 °C	-	-

 ^a The initial conditions: **1a** (0.09 mmol, 1.0 equiv), Ph₂IOTf (0.09 mmol, 1.0 equiv), Pd(OAc)₂ (50 mol%), KOAc (0.27 mmol, 3.0 equiv), DMF (2.0 mL), 130 °C, 12 h, air. ^b
Yield determined by GC analysis using *n*-dodecane as an internal standard.

Table S4 Oxidant screening^a

Ia	20 mol% Pd(OAc) ₂ Ph ₂ IOTf (1.0 equiv) Oxidant (1.0 equiv) DMF, 130 °C, 15 h, air 'initial conditions' 3a	O N Ph ^o 3a'	
Entry	Oxidant variations	Yiel	d ^b (%)
		3a	3a'
1°		82	5
2	Ag(OAc) ₂	31	-
3	Agl	17	-
4	AgCO ₃	12	-
5	AgNO ₃	10	-
6	Ag ₂ O	38	-
7	AgSbF ₆	13	-
8	AgTFA	74	1

^a The initial conditions: **1a** (0.09 mmol, 1.0 equiv), Ph₂IOTf (0.09 mmol, 1.0 equiv), Pd(OAc)₂ (20 mol%), oxidant (0.09 mmol, 1.0 equiv), KOAc (0.27 mmol, 3.0 equiv), DMF (2.0 mL), 130 °C, 15 h, air. ^b Yield determined by GC analysis using *n*-dodecane as an internal standard. ^c 50 mol% Pd(OAc)₂ for 12 h instead of 20 mol% Pd(OAc)₂ for 15 h.

Table S5 Ligand screening^a



Entry	Ligand variations	Yield ^b (%)
1°		74
2	2,2'-Bipyridyl	2
3	Bathophenanthroline	1
4	XPhos	21
5	<i>t</i> BuXPhos	4
6	XantPhos	35
7	CyJohnPhos	43
8	BrettPhos	>99
9	Tricyclohexylphosphine	85
10	Triphenylphosphine	72

^a The initial conditions: **1a** (0.09 mmol, 1.0 equiv), Ph₂IOTf (0.11 mmol, 1.2 equiv), Pd(OAc)₂ (10 mol%), AgTFA (0.09 mmol, 1.0 equiv), ligand (40 mol%), KOAc (0.27 mmol, 3.0 equiv), DMF (2.0 mL), 130 °C, 24 h, air. ^b Yield determined by GC analysis using *n*-dodecane as an internal standard. ^c 20 mol% Pd(OAc)₂ and 1.0 equiv Ph₂IOTf are used for 15 h.

Table S6 Selected optimization conditions under an oxygen atmosphere^a



Entry	Reaction conditions	Yield ^b (%)
1	•	76
2	20 mol% Pd(OAc) ₂	78
3	20 mol% BrettPhos	63
4	20 mol% Pd(OAc) ₂ , 30 mol% BrettPhos, 1.1 equiv Ph ₂ IOTf and 4.0 equiv KOAc	97

^e The initial conditions: **1a** (0.09 mmol, 1.0 equiv), Ph₂IOTf (0.11 mmol, 1.2 equiv), Pd(OAc)₂ (10 mol%), BrettPhos (40 mol%), AgTFA (0.09 mmol, 1.0 equiv), KOAc (0.27 mmol, 3.0 equiv), DMF (2.0 mL), 130 °C, 24 h, O₂. ^{*b*} Isoated yield.

III.2 Procedure for the *N*-arylation (2a)

To a Schlenk tube were added 2-phenylquinazolin-4(3*H*)-one **1a** (0.09 mmol, 1.0 equiv), Ph_2IOTf (0.135 mmol, 1.5 equiv), Cul (0.009 mmol, 10 mol%), Na_2CO_3 (0.18 mmol, 2.0 equiv), Li-*t*-Yu-butyl-quinoline **L7** (0.036, 40 mol%) and DMF (2.0 mL). The resulting mixture was stirred at 130 °C for 24 h. The reaction was monitored by TLC or GC analysis. After completion of the reaction, the reaction mixture was concentrated *in vacuo* and purified by flash column chromatohraphy.

*2a^{S7} has been reported.

III.3 Procedure for the Annuative π -Extension (3)

To a Schlenk tube were added 2-arylquinazolin-4(3*H*)-one **1** (0.09 mmol, 1.0 equiv), Ar_2IOTf (0.10 mmol, 1.1 equiv), $Pd(OAc)_2$ (0.018 mmol, 20 mol%), BrettPhos (0.027 mmol, 30 mol%), KOAc (0.36 mmol, 4.0 equiv), AgTFA (0.09 mmol, 1.0 equiv) and DMF (2.0 mL). The resulting mixture was bubbled with oxygen for 5 min, and then stirred at 130 °C for 24 h under an O₂ atmosphere. The reaction was monitored by TLC or GC analysis. After completion of the reaction, the reaction mixture was concentrated *in vacuo* and purified by flash column chromatohraphy.



Scheme S1 Proposed mechanism of annulative π-extension: alternative Pd(II)/Pd(IV) manifold in cycle II.

*3a, S8 3c, S8 3d, S9 3e, S8 3h, S9 3j, S8 3l, S8 3q, S10 3r, S10 and 3s S10 have been reported.

13-Methoxy-14H-quinazolino[3,2-f]phenanthridin-14-one (3b)



White solid (92%); **ATR-FTIR** (cm⁻¹): 3009, 2922, 2852, 2239, 1683, 1593, 1547, 1470, 1346, 1261, 1101, 1085, 1024, 914, 860, 810, 779, 747, 717; ¹**H NMR** (400 MHz, CDCl₃) δ 8.95 (dd, *J* = 8.1, 1.4 Hz, 1H), 8.91 – 8.86 (m, 1H), 8.26 – 8.17 (m, 2H), 7.70 (td, *J* = 8.2, 7.6, 6.3 Hz, 2H), 7.58 (ddd, *J* = 8.2, 7.1, 1.2 Hz, 1H), 7.45 (tdt, *J* = 8.2, 5.6, 2.9 Hz, 2H), 7.41 – 7.36 (m, 1H), 6.91 (dd, *J* = 8.3, 0.9 Hz, 1H),

4.07 (s, 3H); ¹³**C NMR** (100 MHz, CDCl₃) δ 160.11, 159.09, 147.71, 145.71, 133.74, 131.68, 131.18, 130.54, 127.41, 126.99, 126.92, 125.91, 125.13, 121.98, 121.93, 121.03, 120.74, 118.07, 109.67, 106.13, 55.38; **HRMS-ESI**: m/z [M+H]⁺ calcd for C₂₁H₁₅N₂O₂: 327.1128; found: 327.1123.

10-Nitro-14H-quinazolino[3,2-f]phenanthridin-14-one (3f)



Yellowish solid (89%); **ATR-FTIR** (cm⁻¹): 3081, 2959, 2923, 2854, 1732, 1693, 1597, 1550, 1523, 1447, 1362, 1335, 1258, 11522, 1018, 869, 799, 760, 738, 714; ¹**H NMR** (400 MHz, $CDCl_3$) δ 9.11 – 9.06 (m, 1H), 9.02 (dd, *J* = 8.2, 1.4 Hz, 1H), 8.65 (dd, *J* = 8.1, 1.5 Hz, 1H), 8.36 – 8.22 (m, 3H), 7.81 (ddd, *J* = 8.3,

7.1, 1.5 Hz, 1H), 7.65 (ddd, J = 8.2, 7.2, 1.2 Hz, 1H), 7.60 – 7.52 (m, 3H); ¹³**C NMR** (100 MHz, CDCl₃) δ 160.39, 147.52, 144.56, 138.05, 132.33, 131.33, 131.09, 130.76, 128.27, 128.05, 127.77, 127.41, 126.18, 125.46, 123.70, 122.27, 122.23, 121.32, 121.11, 120.77; **HRMS-ESI**: m/z [M+H]⁺ calcd for C₂₀H₁₂N₃O₃: 342.0873; found: 342.0868.

11-Fluoro-14H-quinazolino[3,2-f]phenanthridin-14-one (3g)



White solid (91%); ATR-FTIR (cm⁻¹): 2922, 2852, 1686, 1597, 1552, 1474, 1451, 1335, 1281, 1149, 1089, 1041, 971, 858, 774, 764, 746, 715; ¹H NMR (400 MHz, CDCl₃) δ 9.08 (dd, *J* = 8.0, 1.8 Hz, 1H), 8.97 (dd, *J* = 8.1, 1.3 Hz, 1H), 8.41 (dd, *J* = 8.9, 6.1 Hz, 1H), 8.31 – 8.18 (m, 3H), 7.80 – 7.70 (m, 1H), 7.67 – 7.57 (m, 1H), 7.56 – 7.46 (m, 2H), 7.21 (td, *J* = 8.5, 2.5 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ

166.74 (d, J = 254.4 Hz), 162.22, 148.22 (d, J = 13.5 Hz), 147.54, 132.86, 132.52, 131.51, 130.22 (d, J = 10.8 Hz), 128.60, 128.35, 128.20, 126.83, 126.59, 123.09, 123.02, 122.17, 121.81, 117.50, 115.10 (d, J = 23.7 Hz), 111.90 (d, J = 21.7 Hz); ¹⁹**F NMR** (376 MHz, CDCl₃) δ -103.10; **HRMS-ESI**: m/z [M+H]⁺ calcd for C₂₀H₁₂FN₂O: 315.0928; found: 315.0926.

13-Fluoro-14H-quinazolino[3,2-f]phenanthridin-14-one (3i)



White solid (64%); **ATR-FTIR** (cm⁻¹): 3076, 2926, 2853, 2360, 1674, 1603, 1553, 1482, 1453, 1359, 1291, 1242, 1152, 1108, 947, 833, 782, 743, 714; ¹**H NMR** (400 MHz, CDCl₃) δ 8.97 – 8.89 (m, 2H), 8.21 (dd, *J* = 9.9, 7.7 Hz, 2H), 7.77 – 7.68 (m, 2H), 7.58 (dt, *J* = 7.6, 3.5 Hz, 2H), 7.48 (ddd, *J* = 6.8, 4.5, 1.8 Hz, 2H), 7.12 (dd, *J* = 10.7, 8.1 Hz, 1H); ¹³**C NMR** (100 MHz, CDCl₃) δ 161.38 (d, *J* = 266.2 Hz),

148.19, 147.15, 134.74 (d, J = 10.7 Hz), 132.52, 132.43, 131.56, 131.26, 128.55, 128.21, 128.16, 128.12, 127.55, 126.66, 126.58, 123.09, 122.96, 122.76 (d, J = 4.2 Hz), 121.89 (d, J = 14.6 Hz), 112.45 (d, J = 21.0 Hz); ¹⁹**F NMR** (376 MHz, CDCl₃) δ -111.48; **HRMS-ESI**: m/z [M+H]⁺ calcd for C₂₀H₁₂FN₂O: 315.0928; found: 315.0925.

13-Chloro-14*H*-quinazolino[3,2-*f*]phenanthridin-14-one (3k)



Yellowish solid (66%); **ATR-FTIR** (cm⁻¹): 3062, 2957, 2920, 2851, 1736, 1691, 1571, 1540, 1452, 1342, 1238, 1148, 1118, 1017, 965, 896, 862, 809, 769, 748, 718; ¹**H NMR** (400 MHz, CDCl₃) δ 8.97 (dd, *J* = 8.1, 1.4 Hz, 1H), 8.86 – 8.80 (m, 1H), 8.26 (ddd, *J* = 7.7, 5.0, 2.0 Hz, 2H), 7.79 – 7.71 (m, 2H), 7.68 –

7.60 (m, 2H), 7.55 – 7.48 (m, 3H); ¹³**C NMR** (100 MHz, CDCl₃) δ 160.11, 147.64, 145.88, 133.56, 133.03, 132.78, 131.56, 131.40, 130.61, 127.61, 127.57, 127.19, 127.06, 125.58, 125.07, 122.14, 122.01, 120.88, 120.81, 116.93; **HRMS-ESI**: m/z [M+H]⁺ calcd for C₂₀H₁₂CIN₂O: 331.0633; found: 331.0628.

12-Bromo-14*H*-quinazolino[3,2-*f*]phenanthridin-14-one (**3m**)



Yellowish solid (78%); **ATR-FTIR** (cm⁻¹): 2921, 2852, 1734, 1679, 1599, 1552, 1469, 1449, 1341, 1288, 1259, 1147, 1093, 1017, 881, 761, 742, 714; ¹**H NMR** (400 MHz, CDCl₃) δ 9.11 (dd, *J* = 8.1, 1.7 Hz, 1H), 9.03 (dt, *J* = 8.2, 0.9 Hz, 1H), 8.43 (dd, *J* = 8.0, 1.1 Hz, 1H), 8.30 – 8.21 (m, 1H), 7.85 – 7.82 (m, 2H), 7.79 – 7.71 (m, 1H), 7.66 – 7.59 (m, 1H), 7.51 (ddd, *J* = 9.2, 6.0, 2.0 Hz, 3H); ¹³**C NMR** (100

MHz, CDCl₃) δ 162.99, 146.14, 134.55, 133.51, 133.08, 132.22, 131.35, 128.98, 128.56, 128.24, 128.14, 127.41, 127.17, 126.94, 126.49, 126.25, 123.12, 123.10, 122.18, 121.82; **HRMS-ESI**: m/z [M+H]⁺ calcd for C₂₀H₁₂BrN₂O: 375.0128; found: 375.0128.

11,12-Dimethyl-14*H*-quinazolino[3,2-*f*]phenanthridin-14-one (**3n**)



White solid (98%); **ATR-FTIR** (cm⁻¹): 2961, 2920, 2852, 2593, 2226, 1950, 1730, 1677, 1598, 1551, 1473, 1432, 1326, 1286, 1259, 1148, 1088, 1020, 870, 799, 766, 746, 716; ¹**H NMR** (400 MHz, CDCl₃) δ 9.11 (dd, *J* = 7.5, 1.2 Hz, 1H), 8.98 (ddd, *J* = 8.1, 1.5, 0.5 Hz, 1H), 8.26 – 8.20 (m, 2H), 8.16 (d, *J* = 1.0 Hz, 1H), 7.75 – 7.69 (m, 1H), 7.62 – 7.56 (m, 2H), 7.54 – 7.44 (m, 2H), 2.46 (s, 3H), 2.45

 $(s, 3H); \ensuremath{\,^{13}\text{C}}\ensuremath{\,\text{NMR}}\xspace(100\ensuremath{\,\text{MHz}}\xspace, \text{CDCI}_3)\ensuremath{\,^{\circ}}\begin{subarray}{c} 161.88, 144.91, 143.95, 143.56, 134.99, 132.25, 130.89, 130.16, 127.46, 127.03, 127.01, 126.45, 126.07, 125.27, 122.08, 122.03, 121.20, 120.97, 120.76, 117.67, 19.50, 18.83; \ensuremath{\,\text{HRMS-ESI:}}\ensuremath{\,\text{m/z}}\xspace[M+H]^+\ensuremath{\,\text{calcd}}\ensuremath{\,\text{for}}\ensuremath{\,^{\circ}}\xspace{1}\ens$

11-Chloro-13-methyl-14H-quinazolino[3,2-f]phenanthridin-14-one (30)



White solid (83%); **ATR-FTIR** (cm⁻¹): 3084, 2969, 2924, 1691, 1586,1573, 1547, 1443, 1341, 1322, 1281, 1149, 1029, 896, 857, 753, 744, 719; ¹**H NMR** (400 MHz, CDCl₃) δ 8.91 (dd, *J* = 8.1, 1.4 Hz, 1H), 8.84 – 8.79 (m, 1H), 8.28 – 8.18 (m, 2H), 7.73 (ddd, *J* = 8.2, 7.1, 1.4 Hz, 1H), 7.64 – 7.56 (m, 2H), 7.53 – 7.43 (m, 2H), 7.21 (dq, *J* = 1.8, 0.9 Hz, 1H), 2.92 (s, 3H); ¹³**C NMR** (100 MHz, CDCl₃) δ

161.89, 147.56, 146.03, 142.00, 138.40, 131.53, 131.40, 130.51, 127.74, 127.55, 127.04, 126.98, 125.73, 125.39, 123.40, 122.15, 122.06, 120.78, 120.74, 116.81, 21.98; **HRMS-ESI**: m/z [M+H]⁺ calcd for $C_{21}H_{14}CIN_2O$: 345.0789; found: 345.0782.

11-Fluoro-13-methyl-14H-quinazolino[3,2-f]phenanthridin-14-one (3p)



White solid (99%); **ATR-FTIR** (cm⁻¹): 3077, 2960, 2926, 2852, 1734, 1694, 1594, 1555, 1451, 1333, 1290, 1258, 1143, 1113, 1030, 1004, 986, 855, 766, 715; ¹**H NMR** (400 MHz, CDCl₃) δ 8.94 (ddd, J = 8.1, 1.5, 0.5 Hz, 1H), 8.87 – 8.80 (m, 1H), 8.28 – 8.20 (m, 2H), 7.74 (ddd, J = 8.2, 7.2, 1.5 Hz, 1H), 7.60 (ddd, J = 8.2, 7.2, 1.2 Hz, 1H), 7.53 – 7.44 (m, 2H), 7.31 – 7.27 (m, 1H), 6.99 (ddg, J = 9.5, 2.6,

0.9 Hz, 1H), 2.95 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 164.37 (d, J = 253.5 Hz), 161.75, 148.93, 148.79, 146.03, 143.72 (d, J = 10.6 Hz), 131.57, 131.40, 130.56, 127.55, 127.02 (d, J = 8.0 Hz), 125.72, 125.33, 122.15, 122.05, 120.80, 116.04 (d, J = 23.0 Hz), 115.22, 115.20, 109.00 (d, J = 21.1 Hz), 22.38 (d, J = 1.5 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ -104.72; **HRMS-ESI**: m/z [M+H]⁺ calcd for C₂₁H₁₄FN₂O: 329.1085; found: 329.1081.

III.4 Procedure for the C–H Fluorination (4a)

To a Schlenk tube were added 2-phenylquinazolin-4(3*H*)-one **1a** (0.09 mmol, 1.0 equiv), $Pd(OAc)_2$ (10 mol%), NFSI (0.27 mmol, 3.0 equiv) and DMF (2.0 mL). The resulting mixture was stirred at 150 °C for 12 h. The reaction was monitored by TLC or GC analysis. After the completion of the reaction, the reaction mixture was concentrated *in vacuo*, and purified by flash column chromatohraphy.

*4a^{S11} has been reported.

IV. Detection of Hg²⁺ Using FET Sensors

IV.1 Methods

Fabrication of Chemically-Modified Graphene FET

Photoresist was patterned on Si substrates with 300 nm thick SiO₂, and Cr/Au (3 nm/100 nm) electrodes were made by thermal evaporation. Then, graphene was transferred onto the Cr/Au electrodes and patterned by photolithography. Graphene film was dry-etched using oxygen reactive ion etching (RIE) plasma for channel. Graphene channel was surface modified by dip coating of **3a** (300 ppm) in MeOH/DCM (9:1, v/v) via the π - π stacking interaction.

Fabrication of Graphene-AgNW Hybrid Field-Effect Sensors and Integration with µ-Fluidic Channel

For making the µ-fluidic channel, SU-8 3050 negative photoresist was spin coated onto a bare Si wafer, followed by patterning by photolithography process. A mixture of Sylgard 184 silicone elastomer and curing agent (10:1 wt.%) was poured onto the Si wafer with the thickness of 5 mm and cured at 60 °C for 5 h. The cured PDMS was peeled from the master, and then input, output, and gate terminals were punched out of the PDMS with steel tubes. The Ag/AgCl reference electrode was inserted into the PDMS channel through the gate terminal.

Real-time Sensing of Hg²⁺ by Chemically-Modified Graphene-FET

The flow rate of all solutions in the μ -fluidic channel was 1.0 mL h⁻¹. For the real-time sensing, Hg(CN)₂ solution (100 ppm) and imidazole solution (aqueous, 20 mM) were consecutively injected in the μ -fluidic channel. A solution-gate characterization was conducted by sweeping Ag/AgCl reference electrode using a probe station (Keithley 4200-SCS semiconductor parametric analyzer). All devices were characterized at 0.1 V of drain bias.



Scheme S2 Reversible and real-time sensing of Hg²⁺ using chemically-modified graphene-FET.



IV.2 Electronic Characteristics of FET for Other Metal lons

Fig. S1 Transfer characteristics of solution-gated graphene field-effect sensors with **3a**-functionalization after flowing (a) Cu^{2+} solution, (b) Na^+ solution, (c) K^+ solution (V_D = 0.1 V). (d) Optical microscope image of fabricated graphene field-effect sensor. Scale bar, 500 µm. Concentration the solution: $Cu(OAc)_2$, NaCl, or KCl; 100 ppm.



Fig. S2 Transfer characteristics of solution-gated graphene field-effect sensors without **3a**-functionalization after flowing (a) Hg²⁺ solution, (b) Cu^{2+} solution, (c) Na⁺ solution, (d) K⁺ solution (V_D = 0.1 V).

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VI. NMR Spectra



Fig. S3 1 H NMR (400 MHz, DMSO-*d*₆) of the compound 1f.



Fig. S4 $\,^{\rm 13}{\rm C}$ NMR (100 MHz, DMSO- $d_6)$ of the compound 1f.







Fig. S6 13 C NMR (100 MHz, DMSO- d_6) of the compound 1n.







Fig. S8 $\,^{13}\text{C}$ NMR (100 MHz, DMSO- $d_6)$ of the compound 10.











S17































Fig. S32 ¹³F NMR (376 MHz, CDCl₃) of the compound **3p**.