Supplementary Information

Lipopeptidomimetics Derived from Teixobactin have Potent Antimicrobial Activity against *Staphylococcus aureus*

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General Experimental

All solvents and reagents used in the experiments were purchased from Novabiochem or Sigma Aldrich and were used without purification. All amino acids used in peptides synthesis were either L- or D-configuration (enantiopure) and were Fmoc protected. All amino acids, except Glycine had their side residues protected. DMF was dried over 4 Å molecular sieves and stored under nitrogen.

Solid phase peptide synthesis was performed manually using a polypropylene syringe tube fitted with a polypropylene frit. Solvents were evaporated under reduced pressure on Büchi vacuum rotary evaporators. Peptides were freeze-dried on a Labconco FreeZone2.5 lyophiliser.

Analytical RP-HPLC analyses were preformed on a GeminiTM 5u C18 110Å column (Phenomenex[®] Inc., Torrance, California, 150 mm × 4.6 mm, 5 µm) with a flow rate of 0.5 mL/min using a linear gradient of solvent A [water (0.1% TFA)] and solvent B [acetonitrile (0.1% TFA)]. Retention times (t_R) from analytical RP-HPLC are reported in minutes. Peptides were purified with a GeminiTM 5u C18 110Å column (Phenomenex[®] Inc., Torrance, California, 250 mm × 21 mm, 5 µm, C18) using a specified linear gradient of solvent A [water (0.1% TFA)] and solvent B [acetonitrile (0.1% TFA)], with a flow rate of 10.6 mL/min. UV detection wavelengths in analytical HPLC were 214 nm and 260 nm. UV detection wavelength in semi-preparative HPLC was 214 nm.

LCMS analysis was performed on a Waters Xevo Q ToF mass spectrometer coupled to a Water Acquity LC system with Waters Acquity UPLC BEH C18 column (2.1 x 50 mm). Solvent system was 0.1% formic acid in MeCN and 0.1% formic acid in H₂O (deionised). Gradient was 5-100% of 0.1% formic acid in MeCN at 0.6 mL/min. Mass accuracy was accomplished by using a reference lock mass scan once per second. ES cone voltage was 30V. Collision energy was 4 eV. MS acquisition rate was 10 spectra per seconds with m/z range 50-2000 Da.

General Peptide Synthesis Experimental Procedures

Loading of resin

2-Chlorotrityl chloride resin (1.51 mmol/g, 1.32 g, 2 mmol, 1 eq) was added to an oven dried glass filtration vessel. Anhydrous DCM (10 mL) was added, and the resin allowed to swell for 20 minutes. Fmoc-Ala-OH (1.87 g, 6 mmol, 3 eq) was then added, forming a suspension. DIEA (2.09 mL, 12 mmol, 6 eq) was then also added and the vessel flushed with nitrogen. The reaction was shaken overnight at room temperature. Following this, the vessel was drained and the resin washed with DMF (3 x 10 mL), MeOH (3 x 10 mL) and DCM (3 x 10 mL). An Fmoc-loading test was carried out by measuring UV absorption of the Fmoc cleavage product at 300 nm, and found to be 0.75 mmol/g (50% loading, 1 mmol).

Fmoc deprotection

To the loaded resin, 20% piperidine/DMF (10 mL) was added. The vessel was sealed and shaken at room temperature for 20 minutes, after which the solution was drained. The resin was then washed as previously with DMF (3 x 10 mL), MeOH (3 x 10 mL) and DCM (3 x 10 mL). The reaction was monitored by Kaiser Test and repeated if a negative test was observed.

Fmoc-amino acid coupling

The loaded resin was swollen with DMF (10 mL) for 15 minutes before being drained. In a vial, a solution of Fmoc-protected amino acid (3 mmol, 3 eq), HCTU (1.241 g, 3 mmol, 3 eq) and DIEA (1.05 mL, 6 mmol, 6 eq) in DMF (8 mL) was prepared and added to the resin, and shaken at room temperature for one hour, after which the solution was drained. The resin was then washed with DMF (3 x 10 mL), MeOH (3 x 10 mL) and DCM (3 x 10 mL). Reaction was monitored by Kaiser Test and repeated if a positive test was observed.

Acetyl capping

To the loaded resin, DMF (8 mL) was added, followed by acetic anhydride (113 μ L, 1.2 mmol, 1.2 eq). The vessel was shaken at room temperature for 30 minutes, before being drained and washed with DMF (3 x 8 mL), MeOH (3 x 8 mL) and DCM (3 x 8 mL).

Esterification

The peptide-bound resin was swollen in DMF (10 mL) for 15 minutes. In a vial, a solution of Fmoc-Ile-OH (1.41 g, 4 mmol, 4 eq) and DMAP (49 mg, 0.4 mmol, 0.4 eq) were dissolved in DMF (8 mL). This solution was added to the resin. DIC (625 μ L, 4 mmol, 4 eq) was then added, and the reaction shaken at room temperature for 2 hours. Reaction was monitored by cleaving a small portion of resin with 1% TFA / DCM, and analysed with LCMS.

Splitting of material for analogue synthesis

After deprotection of the Fmoc-lle residue with piperidine, the resin was split into 0.01 mmol portions for the synthesis of analogues with the various enduracididine substitutions. Each of these portions were handled as a slurry of DCM/DMF and transferred to a plastic SPF tube (5 mL) plugged with a frit. The appropriate Fmoc amino acid was then coupled as described above.

Acid Cleavage

After the completion of all Fmoc-amino acid couplings and the final Fmoc-deprotection, a solution of 1% TFA/DCM (8 mL) was added to the resin. This reaction was shaken at room temperature for 20 mins, before the acidic peptide solution was filtered into a Falcon tube (30 mL). The resin was washed with further additions of DCM (3 x 8 mL), and evaporated under reduced pressure. The residual solution was diluted with H₂O/MeCN (5 mL) and lyophilised to afford the TFA salt of the crude product as an off-white solid.

Macrocyclisation

The crude linear branched peptide Ac-C(Trt)t(IK(Boc)-NH₂)I-OH TFA salt (70 mg, 0.076 mmol, 1 eq) was dissolved in DMF (10 mL, 7.6 mM) and DIEA (80 μ L, 0.456 mmol, 6 eq) was added. Separately, in a round bottomed flask (RBF), HATU (58 mg, 0.152 mmol, 2 eq) was dissolved in DMF (20 mL, 7.5 mM) and stirred and room temperature. The peptide/base solution was added dropwise to the HATU solution using a syringe pump over 1 hour. After addition, the reaction was stirred for a further 30 minutes, before evaporating under reduced pressure. The concentrated crude solution was dissolved in MeCN/ H₂O (2 mL), and purified by semi-preparative RP-HPLC (5-100 % solvent B over a 30 minute gradient), to give the purified cyclic peptide as the TFA salt (9.3 mg, 0.010 mmol, 25% yield).

Removal of cysteine trityl protection

A freshly prepared solution of TFA/EDT/TES/DCM (90: 2.5: 2.5: 5) was added to the cyclic peptide and the solution stirred at room temperature for 30 minutes. After this time, the solution was concentrated with a stream of nitrogen and the peptide was then precipitated with ice cold diethyl ether (10 mL).

Prenylation of cysteine thiol

A solution of $Zn(OAc)_2.2H_2O$ (9 mg, 0.04 mmol, 4 eq) in DMF (1 mL) was prepared and added to the free-thiol containing cyclic peptide (1 eq). TFA (5 μ L) was then added, and effervescence observed. Finally, geranyl bromide (8 uL, 0.04 mmol, 4 eq) was added and the reaction shaken at room temperature for 30 minutes. After this time, the reaction was concentrated *in vacuo*, diluted with water and lyophilised to dryness. The prenylated cyclic peptide was purified by semi-preparative RP-HPLC (5-100 % solvent B over a 30 minute gradient), to give the final product as the TFA salt.

Conversion to HCI salt

The prenylated cyclic peptide was dissolved in aqueous HCI (5mM) and lyophilised to dryness. This was repeated a further two times. The product was analysed by ¹⁹F NMR to determine the conversion of TFA to HCI.

Determination of peptide content

Due to the lack of chromophore present in the peptide analogues, the peptide content was determined by ¹H NMR. A known volume of peptide in D_2O (or D_2O /MeCN depending on solubility) was added to a known quantity of *para*-nitrophenol in D_2O . The peptide content was calculated by comparing relative integrations of peaks in the 1H NMR spectrum from both compounds respectively.

Figure S1: Structures of cyclic tetradepsipeptides 15 – 17



Table S1: Yields	of purified	Teixobactin	analogues
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Analogue	Yield (mg)	Yield (%)	Purity (%)	Calc mass	Obs mass
Arg ₁₀ -farnesylbactin (1)	4.3	5	95%	791.4853	791.4873
Lys ₁₀ -farnesylbactin (2)	4.0	5	>99%	763.4792	763.4813
Orn ₁₀ -farnesylbactin (3)	3.0	7	>99%	749.4635	749.4639
Cit ₁₀ -farnesylbactin (4)	4.5	5	98%	792.4694	792.4731
His ₁₀ -farnesylbactin (5)	1.9	4	>99%	772.4431	772.4462
Ala ₁₀ -farnesylbactin (6)	2.6	7	>99%	706.4213	706.4241
Glu ₁₀ -farnesylbactin (7)	3.3	9	>99%	764.4268	764.4266
Arg ₁₀ -geranylbactin (8)	5.2	12	>99%	723.4227	723.4250
Lys ₁₀ -geranylbactin (9)	3.0	7	>99%	695.4166	695.4200
Orn ₁₀ -geranylbactin (10)	4.7	12	>99%	681.4009	681.4003
Cit ₁₀ -geranylbactin (11)	2.8	7	>99%	724.4068	724.4099
His ₁₀ -geranylbactin (12)	1.3	3	>99%	704.3805	704.3825
Ala ₁₀ -geranylbactin (13)	2.4	8	>99%	638.3608	638.3587
Glu ₁₀ -geranybactin (14)	3.0	9	86%	696.3642	696.3663
Cyclic Ac-tARI (15)	3.5	14	>99%	484.2884	484.2900
Cyclic Ac-tAKI (16)	0.5	2	>99%	456.2822	456.2834
Cyclic Ac-tAHI (17)	0.6	2	>99%	465.2462	465.2462

Numbered Compound Structures and NMR Data Tables





Orn₁₀-geranylbactin (10)



Cit₁₀-geranylbactin (11)

11



His₁₀-geranylbactin (12)



Ala₁₀-geranylbactin (13)



Glu₁₀-geranybactin (14)



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Compound		7		4	5	٩		8	۰ 	P	
Arg ₁₀ -farnesylbactin (1)	2.13 (3H s)	4.65 (1H t, <i>J=1.5</i>)	3.35-2.98 (1H m) & 2.98-2.90 (1H m)	3.50-3.42 (1H m) and 3.38-3.32 (1H m)	5.29 (1H t, <i>J=1</i> .5)	1.75 (3H s)	2.15-2.10 (2H m)	2.02-1.97 (2H m)	7(m HI) 60.2-01.2	1.69 (3H s)	
Lys ₁₀ -farnesylbactin (2)	2.06 (3H s)	4.76-4.75 (1H m) ⁷	2.90-2.80 (2H m)	3.33 (2H qd, <i>J</i> =13, 7.5)	5.24 (1H t, <i>J</i> =8)	1.70 (3H s)	2.13-2.05 (2H m)	2.08-2.02 (2H m)	5.14-5.07 (1H m) ³	1.67 (3H s)	
Orn ₁₀ -farnesylbactin (3)	not assigned	4.52-4.45 (1H m) ⁶	3.04-2.89 (2H m)	3.49-3.39 (1H m) & 3.37-3.30 (1H m)	5.32-5.25 (1H, m)	1.75 (3H s)	2.12-2.04 (2H m)	2.01-1.95 (2H m)	5.16-5.09 (1H m) ³	1.68 (3H s)	
Cit ₁₀ -farnesylbactin (4)	1.98 (3H s)	4.46 (1H t, J=7.5)	2.83 (2H d, J=7.5)	3.26 (2H d, <i>J</i> =8)	5.21 (1H t, <i>J</i> =8)	1.66 (3H s)	2.10-2.04 (2H m)	2.05-2.00 (2H m)	5.12-5.05 (1H m) ³	1.64 (3H s)	
His ₁₀ -farnesylbactin (5)		_	_	_	/			_	_		
Ala ₁₀ -farnesvlbactin (6)	not assigned	4.35 (1H t, <i>J</i> =8)	3.66-3.50 (1H br m) & 2.85-2.74 (1H m)	3.26-3.15 (1H m) & 3.12-3.03 (1H m)	5.26-5.13 (1H br m)	1.61 3H s)	2.08-2.01 (2H m)	2.00-1.94 (2H m)	5.23-5.14 (1H m) ³	1.53 (3H s)	
Glu ₁₀ -famesylbactin (7)	not assigned	4.58 (1H t, <i>J</i> =7.5)	2.98 (2H br d, <i>J</i> =7.5)	3.45-3.36 (2H m)	5.36 (1H t, J=7.5)	1.80 (3H s)	2.22-2.15 (2H m)	2.15-2.05 (2H m)	5.27-5.19 (1H m) ³	1.79 (3H s)	
Arg., -geranvlbactin (8)	2.14 (3H s)	4.61 (1H t, <i>J</i> =8)	3.04-2.95 (2H m)	3.40 (2H ad. J=8. 5)	5.34 (1H t, <i>J</i> =8)	1.78 (3H. s)	2.24-2.19 (2H m)	2.19-2.15 (2H m)	5.21 (1H t, <i>J</i> =7)	1.77 (3H s)	
Lvs.a-geranvlbactin (9)	2.09 (3H. s)	4.58 (1H t, <i>J</i> =7.5)	3.01 (2H t. <i>J</i> =6.5)	3.36-3.31 (2H m)	5.29 (1H t. J=8.5)	1.71 (3H s)	2.18-2.14 (2H m)	2.14-2.09 (2H m)	5.16 (1H t, <i>J</i> =6.5)	1.69 (3H s)	
Orngeranvlhactin (10)	2 13 (3H c)	4 61 (1H t /=7 5)	3 03-2 03 (2H m)	3 45-3 36 (2H m)	5 38-5 37 (1H m)	1 78 (3H c)	2 23-2 19 (2H m)	2 19-2 15 (2H m)	5 21 (1H1 /=65)	1 77 (3H c)	
Cit zoumihoatin (11)	le 110) 07.2	A 74 6 64 (1 11)2			EAE (111 + 1-0)		(III 117) CT 7- CZ 7	(11117) CT-2-CT-2	C:0-111+ 1-6)	(5116) / / / 7	
Cit ₁₀ -geranyibacun (11)	riot assigned	4.7.4-0.04 (III)	(III UZ) /0.5-41.5	3.30-3.47 (ZH III)	10-11 LT 10-10	(SHS)06.1	(III IIZ) UE.Z-CE.Z	(111 LIZ) 07:7-05:7	10-1111 7C.C	(5 1 5) 50'T	
HIS ₁₀ -geranyIbactin (12)		/									
Ala ₁₀ -geranylbactin (13)	not assigned	4.43 (1H t, J=7.5)	2.83 (2H d, J=7)	3.26 (2H qd J=7.5, 2.5)	5.21 (1H t, J=7)	1.66 (3H s)	2.10-2.04 (2H m)	2.04-2.00 (2H m)	5.07 (1H t, J=5.5)	1.64 (3H s)	
Glu ₁₀ -geranybactin (14)	not assigned	(4.54 (1H T, J=8)	3.04-2.96 (2H m)	3.45-3.35 (2H m)	(C./=/,1H1,)=.c	1.78 (3H s)	2.24-2.16 (2H m)	2.24-2.16 (2H m)	(C.0=L,1HI) 12.C	1.77 (3H S)	
Compound	11	12	13	14	15	16	17	18	19	20	
Arg ₁₀ -farnesylbactin (1)	2.12-2.05 (3H m)	not assigned	4.26-4.21 (1H m)	1.56 (3H d, <i>J=</i> 7)	4.49 (1H t, J=7.5)	2.01-1.94 (1H m) and 1.76-1.66 (1H m)	3.33-3.26 (2H m)	not assigned	not assigned	not assigned	
Lys ₁₀ -farnesylbactin (2)	not assigned	4.73 (1H d, <i>J</i> =6)	4.09 (1H q, <i>J=</i> 7.5)	1.45 (3H d, <i>J</i> =7)	4.36 (1H dd, <i>J</i> =10, 6) ⁷		2.91 (2H t, <i>J</i> =7.5)	not assigned	not assigned	_	
Orn ₁₀ -farnesylbactin (3)	not assigned	4.71-4.68 (1H m) ²	4.24 (1H q, <i>J=</i> 7.5)	1.57 (3H d, <i>J</i> =7)	4.26-4.19 (1H m)	1.88-1.80 (1H m) and 1.85-1.76 (1H m)	3.07 (2H t, <i>J</i> =6.5)		_	_	
Cit ₁₀ -farnesylbactin (4)	not assigned	4.72-4.63 (1H m)	4.09 (1H q, J=7)	1.41 (3H d, <i>J</i> =7.5)	4.30 (1H dd, J=9.5, 6.5)	1.55-1.49 (1H m) and 1.50-1.43 (1H m)	3.05 (2H t, <i>J</i> =7)	1.83-1.72 (2H m)	_		
His10-farnesylbactin (5)	_	/		1	/		1	/	_	/	
Ala ₁₀ -farnesylbactin (6)	not assigned	4.76-4.55 (1H br m)	4.13-4.02 (1H m) ²	1.35 (3H d, <i>J</i> =7)	4.32 (1H q, <i>J</i> =7.5)	1.35 (3H d, <i>J=</i> 7)	1	/	_	/	
Glu ₁₀ -famesylbactin (7)	not assigned	4.90-4.75 (1H br m)	4.27-4.23 (1H m) ²	1.54 (3H d, <i>J</i> =7.5)	4.49 (1H dd, J=9.5, 6.5)	not assigned	2.48 (2H br t, J=7.5)	_	_	/	
Arg ₁₀ -geranylbactin (8)	1.70 (3H s)	4.87 (1H m) ²	4.24 (1H q, J=7.5)	1.55 (3H d, <i>J</i> =7)	4.48 (1H dd, J=9.5, 7.5)	1.95-1.85 (1H, m) and 1.79-1.74 (1H, m)	3.34-3.25 (2H m)	not assigned	8.01 (2H br s) ¹	8.01 (2H br s) ¹	
Lys10-geranylbactin (9)	1.63 (3H s)	4.75 (1H m) ²	4.18 (1H q, <i>J=</i> 7.5)	1.49 (3H d, <i>J</i> =7.5)	4.43 (1H dd, J=9.5, 7)	1.95-1.89 (1H m) & 1.90-1.84 (1H m)	3.01 (2H t, J=7.5)	not assigned	not assigned	/	
Orn ₁₀ -geranylbactin (10)	1.70 (3H s)	4.89 (1H m) ²	4.24 (1H q, <i>J</i> =7.5)	1.55 (3H d, <i>J</i> =7.5)	4.49 (1H dd, J=9.5, 6)	2.04-1.96 (1H m) & 1.87-1.80 (1H m)	3.09 (2H t, <i>J</i> =7.5)	not assigned	_	/	
Cit ₁₀ -geranylbactin (11)	1.89 (3H s)	not assigned	4.36 (1H q, <i>J</i> =7.5)	1.66 (3H d, <i>J</i> =7.5)	4.55 (1H dd, J=9.5, 6.5)	2.08-2.01 (1H m) & 1.81-1.73 (1H m)	3.31 (2H t, <i>J</i> =7)	/	_	/	
His10-geranylbactin (12)	/	1		1	/		1	/	/	/	
Ala ₁₀ -geranylbactin (13)	1.57 (3H s)	5.71-4.60 (1H m)	4.16-4.09 (1H m) ⁵	1.38 (3H d, <i>J</i> =7.5)	4.41-4.34 (1H m)	1.39 (3H d, <i>J=</i> 7.5)	1	_	_	/	
Glu ₁₀ -geranybactin (14)	1.70 (3H s)	4.75 (1H m) ²	4.25 (1H q, J=7.5)	1.54 (3H d, <i>J</i> =7.5)	4.48 (1H dd, J=9.5, 6.5)	2.22-2.12 (2H m)	2.50 (2H t, <i>J</i> =7.5)	1	. /		
Compound	21	22	23	24	25	26	27	A	8	J	٩
Arg ₁₀ -farnesylbactin (1)	4.28 (1H d, J=9)	1.96-1.89 (1H m)	0.99-0.92 (3H m) [*]	1.59-1.52 (1H m) and 1.28-1.22 (1H m)	0.99-0.92 (3H m) [*]	5.67-5.60 (1H m)	1.38 (3H d, J=6)	2.10-2.05 (2H m)	5.16-5.09 (1H m) ³	1.61 (3H s) 1	1.61 (3H s)
Lys ₁₀ -farnesylbactin (2)	4.14 (1H d, <i>J</i> =9)	1.93-1.85 (1H m)	0.89 (3H d, <i>J=</i> 7)	1.55-1.47 (1H m) and 1.25-1.15 (1H m)	0.87 (3H t, <i>J=</i> 7)	5.45 (1H qd), <i>J</i> =6.5, 2	1.23 (3H d, <i>J</i> =6.5)	not assigned	5.14-5.07 (1H m) ³	1.60 (3H s) 1	1.60 (3H s)
Orn10-farnesylbactin (3)	4.27 (1H d, J=9)	1.82-1.74 (1H m)	1.00-0.92 (3H m) ¹¹	1.63-1.57 (1H m) & 1.28-1.22 (1H m)	1.00-0.92 (3H m) ¹¹	5.64 (1H qd, <i>J=7</i> , 2)	1.38 (3H d, <i>J</i> =6)	not assigned	5.16-5.09 (1H m) ³	1.61 (3H s) 1	1.61 (3H s)
Cit ₁₀ -farnesylbactin (4)	4.15 (1H d, J=9)	1.83-1.74 (1H m)	0.86 (3H d, <i>J</i> =7)	1.51-1.42 (1H m) and 1.18-1.11 (1H m)	0.86 (3H t, <i>J=</i> 7)	5.49 (1H qd, <i>J</i> =6.5, 2.5)	1.24 (3H d, J=6.5)	not assigned			
His10-farnesylbactin (5)	/	/	_	1	/		/	/	/	/	_
Ala ₁₀ -farnesylbactin (6)	4.13-4.05 (1H m) ²	1.75-1.63 (1H m)	0.83-0.77 (3H m)	1.48-1.36 (1H m) & 1.12-1.01 (1H m)	0.86-0.79 (3H m)	5.51-5.41 (1H m)	1.21 (3H d, J=5.5)	not assigned	not assigned	not assigned r	not assigned
Glu ₁₀ -famesylbactin (7)	4.31 (1H d, <i>J</i> =9)	1.94-1.85 (1H m)	1.04-0.97 (3H m) ⁴	1.64-1.56 (1H m) & 1.33-1.23 (1H m)	1.04-0.97 (3H m) ⁴	5.65 (1H qd, <i>J</i> =6.5, 2)	1.40 (3H d, J=6.5)	not assigned	5.27-5.19 (1H m) ³	1.72 (3H s) 1	1.72 (3H s)
Arg ₁₀ -geranylbactin (8)	4.30 (1H d, J=8.5)	1.95-1.85 (1H, m)	0.98 (3H d, J=7.5)	1.61-1.52 (1H m) and 1.30-1.23 (1H m)	0.99 (3H t, J=6.5)	5.65 (1H qd, <i>J</i> =6.5, 2.5)	1.41 (3H d, J=6.5)	/	1	/	
Lys10-geranylbactin (9)	4.23 (1H d, <i>J</i> =9)	1.89-1.84 (1H m)	0.92 (3H d, <i>J=</i> 7)	1.55-1.50 (1H m) & 1.24-1.17 (1H m)	0.90 (3H t, J=7.5)	5.60 (1H qd, <i>J</i> =6.5, 2.5)	1.35 (3H d, J=6.5)	_	_	/	
Orn ₁₀ -geranylbactin (10)	4.30 (1H d, J=9)	1.97-1.87 (1H m)	0.99 (3H d, <i>J=</i> 7)	1.62-1.52 (1H m) & 1.30-1.23 (1H m)	0.98 (3H t, J=7.5)	5.64 (1H qd, <i>J</i> =6.5, 2)	1.41 (3H d, J=6.5)	_	_	/	
Cit ₁₀ -geranylbactin (11)	4.42 (1H d, J=8.5)	2.01.193 (1H m)	1.13-1.08 (3H m)	1.72-1.64 (1H m) & 1.42 -1.34 (1H m)	1.13-1.08 (3H m)	5.77 (1H qd, <i>J</i> =6.5, 2.5)	1.52 (3H d, J=6.5)	_	_	/	
His ₁₀ -geranylbactin (12)	_	_		1	1		_	/	_	/	
Ala10-geranylbactin (13)	4.16-4.10 (1H m) ⁵	1.79-1.72 (1H m)	0.90-0.82 (3H m)	1.52-1.42 (1H m) & 1.15-1.10 (1H m)	0.90-0.82 (3H m) ⁴	5.49 (1H qd, <i>J</i> =6.5, 2.5)	1.25 (3H d, J=6.5)	_	_	/	
Glu10-geranybactin (14)	4.32 (1H d, J=8.5)	1.90-1.82 (1H m)	1.01-0.96 (3H m) ⁴	1.60-1.53 (1H m)and 1.31-1.22 (1H m)	1.01-0.96 (3H m) ⁴	5.66 (1H qd, <i>J</i> =6.5, 2.5)	1.41 (3H d, J=6.5)	/	/		
1 Pure to exchange 10 and 20 -	dece and at second		2 Out on the D	E Overlan with 31 and 13	C bas Ct dtim ashaw 2						
2 Obscured by water peak	appear as one peak		ט UVeriap א מוט ש 4 Overlap between 23 and 25	כב טווש בז חזוא veriap כב טווש כב טווש 6 Overlap 2 and 15	/ Overiap witiו ⊥ב מווט ב						

RP-HPLC, MS and NMR spectra of Teixobactin lipopeptidomimetic Analogues

Arg₁₀-farnesylbactin (1)



No.	Ret.Time	Peak Name	Height	Area	Rel.Area	Amount	Туре
	min		mAU	mAU*min	%		
1	22.41	n.a.	7.996	2.021	4.83	n.a.	BMB
2	23.26	n.a.	100.728	39.822	95.17	n.a.	BMB
Total:			108.724	41.843	100.00	0.000	



Monoisotopic Mass, Even Electron Ions 901 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass) Elements Used: C: 39-39 H: 0-180 N: 0-30 O: 0-30 S: 1-1 graf-515-61/32to34 Mus158 236 (2.113) Cm (236:241-192:198)







Lys₁₀-farnesylbactin (2)



No.	Ret.Time	Peak Name	Height	Area	Rel.Area	Amount	Туре
	min		mAU	mAU*min	%		
1	22.66	n.a.	112.547	34.597	100.00	n.a.	BMB
Total:			112.547	34.597	100.00	0.000	



Monoisotopic Mass, Even Electron Ions 880 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass) Elements Used: C: 39-39 H: 0-180 N: 0-30 O: 0-30 S: 1-1 gcg4-513-60

ML9167	7 235 (2.105) C	Cm (235:240	-201:211)							1: TOF MS ES+ 1.18e+004
100									763	3.4813	
%	30	305.1 3.1582	306.1757	401.2137	421.7246	539.2109 559	2920	663.3465679.466	4 751.4084	764.4875 785.40	634 4699 847.4319
0	250	300	350	400	450	500 550	600	650 7	700 750	0 800	850
Minim Maxim	um: um:			5.0	5.0	-1.5 80.0					
Mass		Calc	. Mass	mDa	PPM	DBE	i-FI1	i-FIT	(Norm) F	ormula	
763.4	813	763.	4792	2.1	2.8	9.5	96.0	0.0	С	39 Н67	N6 07 S



Orn₁₀-farnesylbactin (3)



No.	Ret.Time	Peak Name	Height	Area	Rel.Area	Amount	Туре
	min		mAU	mAU*min	%		
1	22.76	n.a.	86.629	28.680	100.00	n.a.	BMB
Total:			86.629	28.680	100.00	0.000	







Cit₁₀-farnesylbactin (4)



No.	Ret.Time	Peak Name	Height	Area	Rel.Area	Amount	Туре
	min		mAU	mAU*min	%		
1	24.63	n.a.	11.829	1.907	1.92	n.a.	BMb
2	24.99	n.a.	364.246	97.555	98.08	n.a.	bMB
Total:			376.075	99.462	100.00	0.000	



Monoisotopic Mass, Even Electron lons 902 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass) Elements Used: C: 39-39 H: 0-180 N: 0-30 O: 0-30 S: 1-1

gcg4-514-67 ML9168 271 (2.428) Cm (271-240:243)







His₁₀-farnesylbactin (5)



No.	Ret.Time	Peak Name	Height	Area	Rel.Area	Amount	Туре
	min		mAU	mAU*min	%		
1	22.41	n.a.	7.996	2.021	4.83	n.a.	BMB
2	23.26	n.a.	100.728	39.822	95.17	n.a.	BMB
Total:			108.724	41.843	100.00	0.000	



Monoisotopic Mass, Even Electron lons 888 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass)

Elements Used: C: 39-39 H: 0-180 N: 0-30 O: 0-30 S: 1-1 gcg4-516F-64 ML9159 244 (2.191) Cm (244:245-230:231) 1: TOF MS ES+ 2.50e+003 772.4462 100 773.4518 % 794.4312 832.6559 446.7263 483.2014 795.4258 332.2217 405.6943 568.2609 619.3123 ^{647.7856689.0459}751.3778 0 850 500 550 600 650 700 750 800 400 450 300 350 Minimum: -1.5 5.0 5.0 80.0 Maximum: PPM DBE i-FIT i-FIT (Norm) Formula Mass Calc. Mass mDa C39 H62 N7 O7 S 772.4462 772.4431 4.0 12.5 63.7 0.0 3.1

Ala₁₀-farnesylbactin (6)



No.	Ret.Time	Peak Name	Height	Area	Rel.Area	Amount	Туре
	min		mAU	mAU*min	%		
1	32.36	n.a.	475.432	257.147	100.00	n.a.	BMB
Total:			475.432	257.147	100.00	0.000	



Monoisotopic Mass, Even Electron lons 826 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass) Elements Used: C: 36-36 H: 0-180 N: 0-30 O: 0-30 S: 1-1 gcg4-517F-62to64 ML9169 289 (2.589) Cm (289-250:259)







Glu₁₀-farnesylbactin (7)



No.	Ret.Time	Peak Name	Height	Area	Rel.Area	Amount	Туре
	min		mAU	mAU*min	%		
1	25.31	n.a.	476.311	130.641	100.00	n.a.	BMB
Total:			476.311	130.641	100.00	0.000	



Monoisotopic Mass, Even Electron lons 881 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass) Elements Used:

C: 38-38 H: 0-180 N: 0-30 O: 0-30 S: 1-1

gcg4-518F-69 ML9171 275 (2.461) Cm (275-267:271)







Arg₁₀-geranylbactin (8)







Lys₁₀-geranylbactin (9)



No.	Ret.Time min	Peak Nam	e Hei m.	ght AU m	Area AU*min	Rel.Area %	Amount	Туре
1	20.29	n.a.	43	3.006	56.436	100.00	n.a	. BMB
otal:			43	3.006	56.436	100.00	0.000	
L9133	100		1.81 695.42				1:	TOF MS ES+ BPI 2.01e4
	8- 0.11 167.01			2.07 102.03	2.41 102.03	3.31 124.0	1 3.67 09 124.09	3.88 124.09
Monois 316 for Elemer C: 34-3	otopic Mass, Eve mula(e) evaluate hts Used: 34 H: 0-180	0.50 1.00 en Electron Ions d with 1 results withir N: 0-30 O: 0-30	1.50 n limits (all resul S: 1-1	2.00 ts (up to 10	2.50 000) for eac	3.00 ch mass)	3.50	4.00
JCg4-48 VIL9133 100	37-52,53 3 202 (1.819)					695.4200		1: TOF MS ES 1.75e+0
%-						696.4197		
0	102.0325 107. 107. 208 100 208	0135 8.0410 280.9907 367 00 300	.1818 408.2064 400	487.8908 5	59.2982 600	697.4272 718.40 700	081 785.3887 800	915.3399 983.3064 900 1000
linimu Iaximu	: m: : m:	5.0	10.0 8	1.5 0.0				
lass	Calc. M	ass mDa	PPM D	BE	i-FIT	i-FIT (No	orm) Formu	la
COE 42	200 695.416	6 3.4	4.9 8	. 5	107.7	0.0	C34	H59 N6 07 S





Orn₁₀-geranylbactin (10)







Cit₁₀-geranylbactin (11)



No.	Ret.Time	Pe	Peak Name		Height mAU i		Rel.	Area	Amount	Туре
	min						1	%		
1	21.78	n.a.		- 23	3.878	7.154	100	.00	n.a	i. BMB*
Fotal:				23	3.878	7.154	100	.00	0.000)
L9134	100			2.06 746.39					1: TOF MS E 1.2	ES+ BPI 20e4
	8 0.07 167.01 0. 167	52 7.01	1.23 1.53 167.01 167.01	1.77 167.01	2.27 102.03	2.49 1 102.03	3.17 24.09 1	3.48 24.09 12	3.67 3.86 24.09 124.09	
	1 mm									
44 formu lements : 34-34 : 34-34	ula(e) evaluated v Used: H: 0-180 N 57,58 29 (2.055) Cm (229	with 1 result : 0-30 O: -208:213)	s within limits (0-30 S: 1-1	all results (u	ip to 1000) for each m	ass)			1: TOF MS E
00								724.	746.3918 4099	1.20e+0
%-									747.3955	
0	306.1780 373.6	381.6760 880 382.	1868	57.1702 523	3.8971 ₅₈₈	.2788 644.2	2023 707	.3870	748.3896	814.3907
250	300 35	60 400	450	500	550	600 6	650	700	750	800
inimum: aximum:										
	:	5.0	0 10.0	-1.5 80.0						
ass	: Calc. Mas	5.0 s mDa	0 10.0 a PPM	-1.5 80.0 DBE	i-	FIT	i-FIT	(Norm)	Formula	





His₁₀-geranylbactin (12)













Glu₁₀-geranybactin (14)



No.	Ret.Time	Peak Name	Height	Area	Rel.Area	Amount	Туре
	min		mAU	mAU*min	%		
1	21.73	n.a.	22.725	4.892	9.50	n.a.	BMB
2	22.06	n.a.	15.595	1.877	3.64	n.a.	BMb
3	22.31	n.a.	190.040	44.735	86.86	n.a.	bMB
Total:			228.360	51.503	100.00	0.000	



Monoisotopic Mass, Even Electron Ions 814 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass) Elements Used: C: 33-33 H: 0-180 N: 0-30 O: 0-30 S: 1-1 gcq4-518G-38,39 ML9164 241 (2.166) Cm (241:242-230:231)







Cyclic Ac-tARI (15)





Cyclic Ac-tAKI (16)





Monoisotopic Mass, Even Electron lons 420 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass) Elements Used:

C: 21-21 H: 0-250 N: 0-20 O: 0-30 gcg4-387-2-37,38 ML8878 110 (0.995) Cm (110:111-84:91)



Cyclic Ac-tAHI (17)



Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal Concentration (MBC) Assays

Bacterial strains

Two laboratory reference strains were used for the antimicrobial susceptibility testing. *Staphylococcus aureus* ATCC 25923 and *Escherichia coli* ATCC 25922 strains were retrieved from the -80°C frozen stock in the microbial genetics laboratory (University Of Leicester).

Microdilution method for susceptibility testing to antimicrobials

Initial preparation of Mueller–Hinton broth (MHB) medium containing double concentration of the maximum concentration of the antimicrobial compound to be tested was made (128 μ g/ml was made to start with 64 μ g/ml). 200 μ l was aliquoted into the first column of a 96 well plate and two-fold serial dilutions of the compound were performed.

MIC and MBC Assays

The Minimum inhibitory concentration (MIC) of Teixobactin and its analogues was performed as described in the Clinical and Laboratory Standards Institute (CLSI) guidelines using the broth microdilution method.¹ Briefly, each strain was streaked for a single colony on Mueller–Hinton agar plates (Oxoid Ltd., Basingstoke, UK) and incubated at 37°C for 24 hours. Next day, a starting inocula of 1×10^5 CFU/ml of *Staphylococcus aureus* ATCC 25923 and *Escherichia coli* ATCC 25922 strains were aliquoted into 96-well plates containing serial dilutions of the Teixobactin or analogues in the range 64 to 0.125 µg/ml using Mueller-Hinton broth (Oxoid Ltd., Basingstoke, UK). Results were read with a Jenway (6705UV/Vis.) spectrophotometer after 18 hours of incubation at 37 °C. Minimum bactericidal concentration (MBC) was determined by subculturing 10 µl from each well without visible bacterial growth onto Mueller–Hinton agar plates (Oxoid Ltd., Basingstoke, UK). After 24 h of incubation at 37°C, the dilution yielding three colonies or less was scored as the MBC.

Reference: CLSI. Methods for Dilution Antimicrobial Susceptibility Testing For Bacteria That Grow Aerobically. Approved Standard M7-A10. Wayne, PA: Clinical and Laboratory Standards Institute; 2009