

## Synthesis and shift-reagent-assisted full NMR assignment of the bacterial (*Z*<sub>8</sub>,*E*<sub>2</sub>, $\omega$ )-undecaprenol

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### Electronic Supplementary Information

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## Experimental Section

**Synthesis.** Synthesis of compound **3** from *E,E*-farnesol was published previously by our laboratory.<sup>1</sup> Undecaprenol **2** was purchased from American Radiolabeled Chemicals, Inc.

**Compound 7.** Phosphorus tribromide (34  $\mu$ L, 0.36 mmol) in dry ether (0.8 mL) was added dropwise to a mixture of heptaprenol **3**<sup>2</sup> (0.4 g, 0.8 mmol) and pyridine (6  $\mu$ L) in dry ether (2 mL) at ice-water temperature. The resulting solution was stirred for 2 h at the same temperature. The brown mixture was poured into a mixture of ice and water and then washed with hexanes. The separated organic layer was washed with a series of aqueous solutions (water, saturated NaHCO<sub>3</sub>, and brine), dried over anhydrous MgSO<sub>4</sub>, filtered, and concentrated under reduced pressure. The residue, the resultant bromide intermediate, was purified by column chromatography on silica gel. This intermediate (0.3 g, 0.5 mmol) was dissolved in acetonitrile:DMF:ether (1:1:1, 3 mL) and the solution was added dropwise to a suspension of sodium *p*-toluenesulfinate (0.18 g, 1.0 mmol) in DMF (5 mL) in an ice-water bath. The mixture was brought to room temperature over a few minutes and was allowed to stir for 6 h. The mixture was diluted with ethyl acetate and was washed sequentially with water and brine, dried over anhydrous MgSO<sub>4</sub>, filtered, and the filtrate was concentrated under reduced pressure. Compound **7** was purified by column chromatography on silica gel (0.3 g, 59% calculated from **3**). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  1.60, 1.61, 1.65, 1.68, 1.73 (8  $\times$  s, 24H), 1.75 - 2.14 (m, 24H), 2.44 (s, 3H, CH<sub>3</sub>C<sub>6</sub>H<sub>4</sub>-), 3.78 (d, *J* = 8.0 Hz, 2H, -SO<sub>2</sub>CH<sub>2</sub>-), 4.95 (t, *J* = 6.7 Hz, 1H), 5.06-5.16 (m, 5H), 5.20 (t, *J* = 7.6 Hz, 1H), 7.33 (d, *J* = 8.0 Hz, 2H), 7.75 (d, *J* = 8.2 Hz, 2H); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  16.19 (q), 16.20 (q), 17.9 (q), 21.8 (q, CH<sub>3</sub>C<sub>6</sub>H<sub>4</sub>-), 23.5 (q), 23.64 (q), 23.67 (q), 23.75 (q), 25.9 (q), 26.0 (t), 26.5 (t), 26.6 (t), 26.8 (t), 26.9 (t), 32.18 (t), 32.26 (t), 32.32 (t), 32.41 (t), 39.92 (t), 39.94 (t), 56.2 (t, -SO<sub>2</sub>CH<sub>2</sub>-), 111.2 (d, -SO<sub>2</sub>CH<sub>2</sub>CH=), 124.2 (d, -CH<sub>2</sub>CH=),

124.3 (d, -CH<sub>2</sub>CH=), 124.4 (d, -CH<sub>2</sub>CH=), 124.6 (d, -CH<sub>2</sub>CH=), 125.0 (d, -CH<sub>2</sub>CH=), 125.1 (d, -CH<sub>2</sub>CH=), 128.6 (d), 129.8 (d), 131.5 (s), 135.1 (s), 135.4 (s), 135.57 (s), 135.61 (s), 136.1 (s), 136.2 (s), 144.6 (s), 146.0 (s); HRMS (ESI/Q-TOF) *m/z* [M+Na]<sup>+</sup> Calcd 655.4519 for C<sub>42</sub>H<sub>64</sub>O<sub>2</sub>SNa; found 655.4528.

**Compound 8.** The sulfone **7** (300 mg, 470 μmol) was dissolved in a mixture of anhydrous THF and hexamethylphosphoramide (HMPA) (4:1, 5 mL) and the resulting solution was cooled to -78 °C. *n*-Butyllithium (0.4 mL, 640 μmol, 1.6 M in hexane) was added dropwise to this solution, which was stirring for 1 h at the same temperature. A solution of bromide **5**<sup>2</sup> (430 mg, 700 μmol) in anhydrous THF-HMPA (4:1, 2 mL) was added dropwise to the sulfone solution and the mixture was allowed to stir for an additional 3 h at -78 °C. After warming the mixture to iced-water temperature, it was poured into a mixture of ice and water and the mixture was washed with hexanes-ether (1:1). The separated organic layer was washed with water and brine, dried over anhydrous MgSO<sub>4</sub>, filtered, and the filtrate was concentrated under reduced pressure. The residue was purified by column chromatography on silica gel to afford compound **8** (470 mg, 85%). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 1.57 - 1.78 (m, 38H), 1.91 - 2.10 (m, 30H), 2.35 - 2.70 (m, 4H), 2.41, 2.44 (2 × s, 6H, -SO<sub>2</sub>C<sub>6</sub>H<sub>4</sub>CH<sub>3</sub>), 3.79 - 3.87 (m, 2H, -CHSO<sub>2</sub>C<sub>6</sub>H<sub>4</sub>CH<sub>3</sub>), 3.98 (d, *J* = 6.6 Hz, 2H, -CH<sub>2</sub>OBn), 4.49 (s, 2H, -OCH<sub>2</sub>Ph), 4.83 - 5.02 (m, 4H), 5.05 - 5.19 (m, 6H), 5.41 (t, *J* = 6.8 Hz, 1H, -CHCH<sub>2</sub>OBn), 7.26 - 7.36 (m, 9H), 7.67 - 7.74 (m, 4H); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 16.2 (q), 17.9 (q), 21.80, 21.84 (2 × q, CH<sub>3</sub>C<sub>6</sub>H<sub>4</sub>), 23.48 (q), 23.50 (q), 23.52 (q), 23.61 (q), 23.65 (q), 23.68 (q), 23.77 (q), 23.79 (q), 25.8 (t), 25.9 (q), 26.0 (t), 26.4 (t), 26.5 (t), 26.75 (t), 26.78 (t), 26.81 (t), 26.9 (t), 30.2 (t), 30.5 (t), 32.12 (t), 32.17 (t), 32.27 (t), 32.33 (t), 32.4 (t), 39.91 (t), 39.93 (t), 63.3, 63.4 (2 × d, CHTs), 66.5 (t, CH<sub>2</sub>OBn), 72.3 (t, OCH<sub>2</sub>Ph), 117.9 (d, -CH<sub>2</sub>CH=), 118.0 (d, -CH<sub>2</sub>CH=), 122.4 (d, BnOCH<sub>2</sub>CH=), 124.25 (d, -

CH<sub>2</sub>CH=), 124.27 (d, -CH<sub>2</sub>CH=), 124.4 (d, -CH<sub>2</sub>CH=), 124.5 (d, -CH<sub>2</sub>CH=), 124.96 (d, -CH<sub>2</sub>CH=), 125.01 (d, -CH<sub>2</sub>CH=), 127.7 (d), 127.9 (d, -CH<sub>2</sub>CH=), 128.0 (d), 128.3 (d, -CH<sub>2</sub>CH=), 128.5 (d), 129.3 (d), 129.4 (d), 129.53 (d), 129.61 (d, ArH), 130.63 (s), 130.9 (s), 131.4 (s), 135.05 (s), 135.14 (s), 135.4 (s), 135.57 (s), 135.64 (s), 135.9 (s), 138.7 (s), 140.2 (s), 144.5 (s), 144.55 (s), 144.62 (s), 144.8 (s), 145.1 (s); HRMS (ESI/Q-TOF) *m/z* [M+Na]<sup>+</sup> Calcd for C<sub>76</sub>H<sub>108</sub>O<sub>5</sub>S<sub>2</sub>Na 1187.7530; found 1187.7547.

**Compound 1.** Ethylamine (50 mL) was added to a flask containing lithium (100 mg, 8.6 mmol) at -78 °C and ether (10 mL) was added. After stirring for 10 min at -78 °C, an ethereal solution of compound **8** (470 mg, 400 μmol in 10 mL) was added dropwise to the blue solution over 20 min. After stirring for 40 min at -78 °C, the reaction was quenched by the addition of isoprene (5 mL) and MeOH (10 mL). After addition of saturated NH<sub>4</sub>Cl, the reaction mixture was washed with ethyl acetate. The combined ethyl acetate layer was washed with brine, dried over anhydrous MgSO<sub>4</sub>, filtered through a layer of silica gel, and the filtrate was concentrated under reduced pressure. The residue was purified by column chromatography on silica gel to afford the desired product **1** (190 mg, 61%). The complete assignment of <sup>1</sup>H and <sup>13</sup>C NMR is given in Table S1. HRMS (ESI/Q-TOF) *m/z* [M+Na]<sup>+</sup> Calcd 789.6884 for C<sub>55</sub>H<sub>90</sub>ONa; found 789.6842.

**NMR Spectroscopy.** The structures of undecaprenol **1** and **2** [10 mg of each was dissolved in 600 μL C<sub>6</sub>D<sub>6</sub> (100%, 99.96 atom % D) containing 10 mg Eu(hfc)<sub>3</sub>] were determined by interpretation of the 2D homonuclear DQF-COSY, TOCSY, ROESY and heteronuclear <sup>1</sup>H-<sup>13</sup>C HSQC, HSQC-TOCSY, HMBC NMR spectra. All NMR spectra were recorded at 25 °C on a four-channel Bruker AVANCE II spectrometer at field strength of 18.79 T using a 5-mm inverse triple-resonance (TCI) <sup>1</sup>H/<sup>13</sup>C/<sup>15</sup>N, z-axis PFG cryoprobe, and running the TopSpin 3.2, pl6

software. The above spectra were measured by employing ordinary pulse sequences in the Bruker pulse sequence library. To enhance spectral resolution, linear prediction and zero filling were applied to the time domain data. Also, the squared shifted sine weighting window functions were used before 2D Fourier transformation. The homonuclear spectra and the  $^1\text{H}$  dimension in heteronuclear spectra were referenced to the residual solvent signal ( $\text{C}_6\text{D}_6$ ,  $\delta_{\text{H}}$  7.15 ppm). The  $^{13}\text{C}$  dimension in the heteronuclear spectra was referenced indirectly.<sup>2</sup> Resonance signals in the measured spectra exhibited slight downfield shifts as the samples stayed in the magnet longer. Consequently, Table S1 contains values of the proton and carbon chemical shifts for carbons with directly attached protons corresponding to the positions of the crosspeaks in the HSQC spectra. The chemical-shift values for the quaternary carbons then correspond to the positions of the crosspeaks in the HMBC spectra.

### References

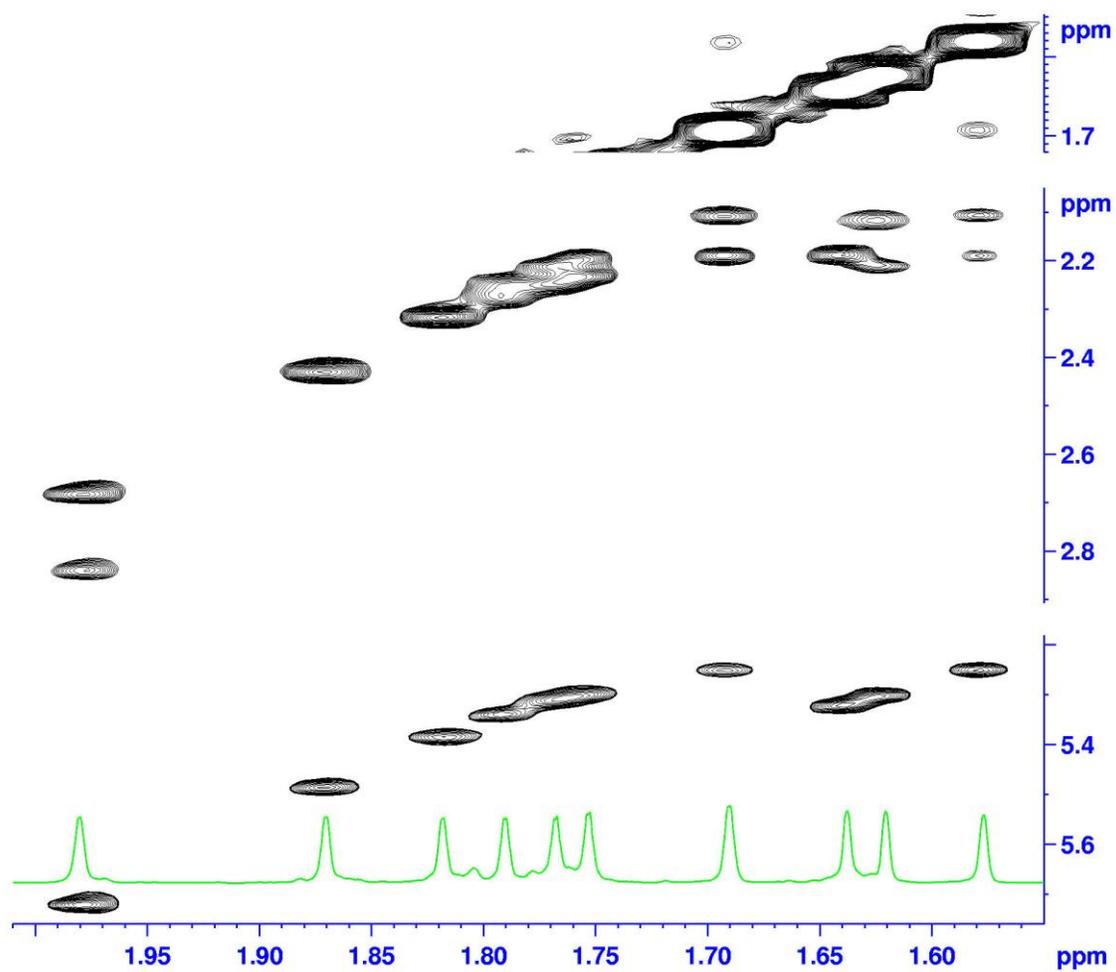
- (1) Hesk, D.; Lee, M.; Zajíček, J.; Fisher, J. F.; Mobashery, S. *J. Am. Chem. Soc.* **2012**, *134*, 13881-13888.
- (2) Wishart, D. S.; Bigam, C. G.; Yao, J.; Abildgaard, F.; Dyson, H. J.; Oldfield, E.; Markley, J. L.; Sykes, B. D. *J. Biomol. NMR* **1995**, *6*, 135-140.

**Table S1.** NMR spectroscopic data for **1** (synthetic sample) and **2** (from *Magnolin kobus*) in C<sub>6</sub>D<sub>6</sub> at 298 K (800.13 MHz).<sup>a,b</sup>

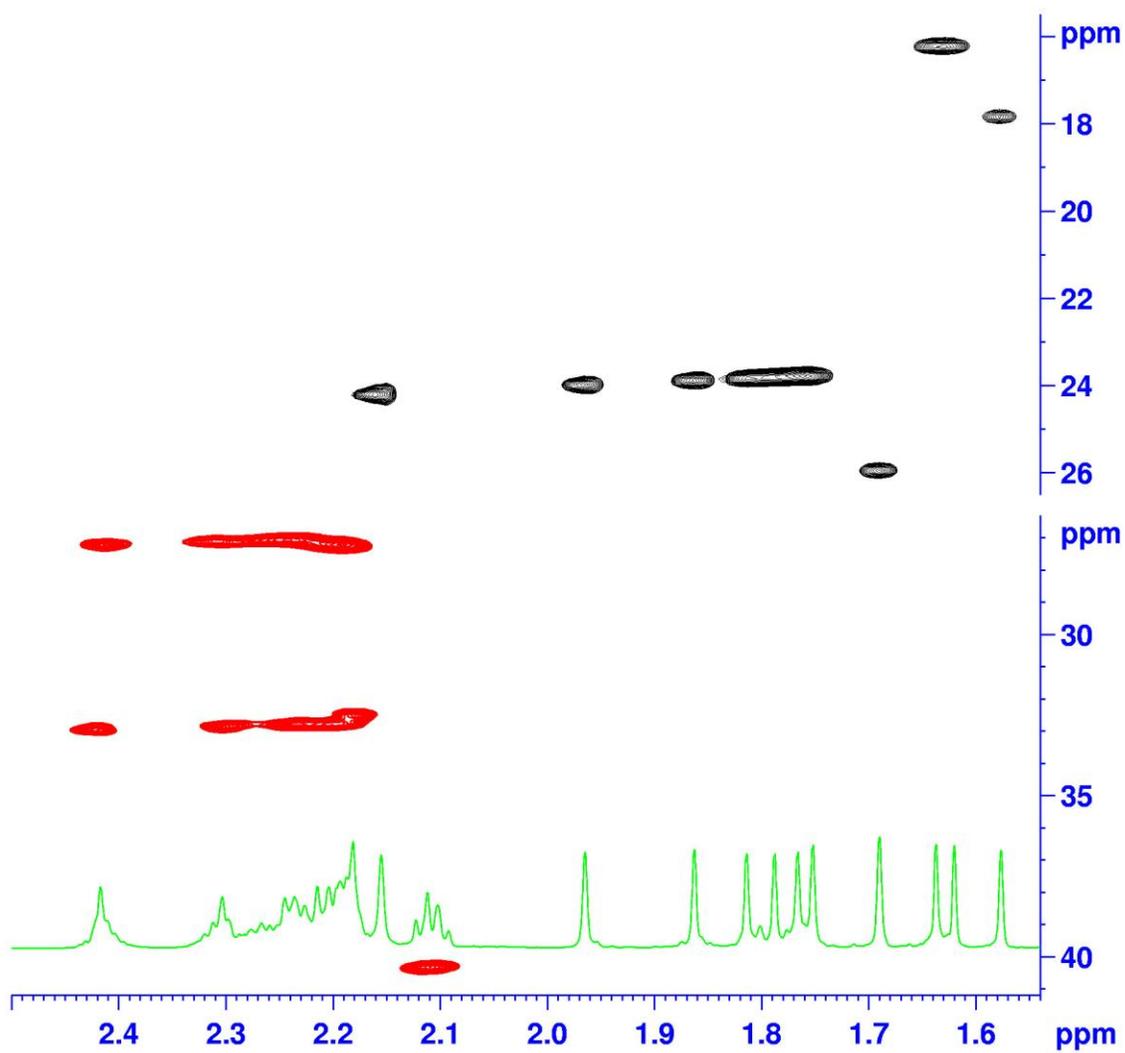
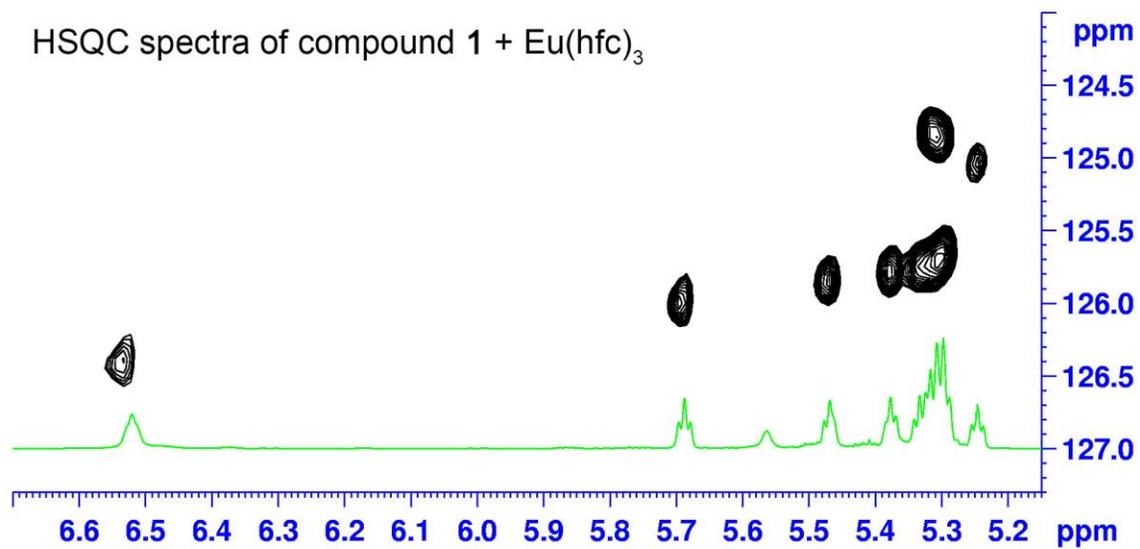
<b>1 + Eu(hfc)<sub>3</sub></b>					<b>2 + Eu(hfc)<sub>3</sub></b>				
unit	<sup>13</sup> C δ [ppm] <sup>c,d</sup>	<sup>1</sup> H δ [ppm] <sup>c</sup>	HMBC connectivities		unit	<sup>13</sup> C δ [ppm] <sup>c,d</sup>	<sup>1</sup> H δ [ppm] <sup>c</sup>	HMBC connectivities	
CH <sub>2</sub> -1		14.37			CH <sub>2</sub> -1		18.02		
CH-2	Z	129.09	11.50		CH-2	Z	129.51	10.35	
C-3		143.62			C-3		142.98		
CH <sub>2</sub> -4		35.42	4.83	2, 3, 5, 6, 55	CH <sub>2</sub> -4		34.90	4.01	2, 3, 5, 6, 55
CH <sub>2</sub> -5		28.33	3.77	3, 4, 6, 7	CH <sub>2</sub> -5		28.21	3.48	3, 4, 6, 7
CH-6	Z	126.54	6.70	4, 5, 7, 8, 54	CH-6	Z	126.33	6.41	4, 5, 8, 54
C-7		137.08			C-7		137.14		
CH <sub>2</sub> -8		33.35	2.88	6, 7, 9, 10, 54	CH <sub>2</sub> -8		33.27	2.74	6, 7, 9, 10, 54
CH <sub>2</sub> -9		27.41	2.70	7, 8, 10, 11	CH <sub>2</sub> -9		27.34	2.61	7, 8, 10, 11
CH-10	Z	126.02	5.71	8, 9, 11, 12, 53	CH-10	Z	125.93	5.65	8, 9, 11, 12, 53
C-11		135.83			C-11		136.02		
CH <sub>2</sub> -12		32.97	2.45	10, 11, 13, 14, 53	CH <sub>2</sub> -12		32.90	2.40	10, 11, 13, 53
CH <sub>2</sub> -13		27.19	2.41	11, 12, 14, 15	CH <sub>2</sub> -13		27.14	2.40	11, 12, 14, 15
CH-14	Z	125.83	5.47	12, 13, 15, 16, 52	CH-14	Z	125.82	5.45	12, 13, 52
C-15		135.58			C-15		135.83		
CH <sub>2</sub> -16		32.85	2.33	14, 15, 17, 18, 52	CH <sub>2</sub> -16		32.83	2.29	14, 17, 18, 52
CH <sub>2</sub> -17		27.09	2.32		CH <sub>2</sub> -17		27.10	2.30	15, 16, 18, 19
CH-18	Z	125.79	5.39	16, 17, 19, 20, 51	CH-18	Z	125.75	5.37	16, 17, 19, 20, 51
C-19		135.46			C-19		135.73		
CH <sub>2</sub> -20		32.77	2.25	21, 51	CH <sub>2</sub> -20		32.76	2.23	<sup>e</sup>
CH <sub>2</sub> -21		27.07	2.26	19, 20, 22, 23	CH <sub>2</sub> -21		27.07	2.25	<sup>e</sup>
CH-22	Z	125.73	5.34	20, 21, 23, 24, 50	CH-22	Z	125.72	5.33	20, 21, 24, 50
C-23		135.45			C-23		135.69		
CH <sub>2</sub> -24		32.72	2.23	25, 50	CH <sub>2</sub> -24		32.75	2.23	<sup>e</sup>
CH <sub>2</sub> -25		27.03	2.23	24	CH <sub>2</sub> -25		27.04	2.24	<sup>e</sup>
CH-26	Z	125.72	5.32	24, 25, 27, 28, 49	CH-26	Z	125.65	5.31	24, 25, 28, 49
C-27		135.43			C-27		135.76		
CH <sub>2</sub> -28		32.74	2.30	<sup>e</sup>	<b>CH<sub>2</sub>-28</b>		<b>32.45</b>	<b>2.19</b>	<sup>e</sup>
CH <sub>2</sub> -29		27.03	2.21	<sup>e</sup>	CH <sub>2</sub> -29		27.14	2.20	<sup>e</sup>
CH-30	Z	125.60	5.29	28, 29, 31, 32, 48	<b>CH-30</b>	<i>E</i>	<b>124.76</b>	5.33	28, 29, 32, 48
C-31		135.49			C-31		135.69		
CH <sub>2</sub> -32		32.46	2.18	<sup>e</sup>	<b>CH<sub>2</sub>-32</b>		<b>40.30</b>	<b>2.12</b>	<sup>e</sup>
CH <sub>2</sub> -33		27.14	2.19	<sup>e</sup>	CH <sub>2</sub> -33		27.21	2.20	<sup>e</sup>
CH-34	<i>E</i>	124.78	5.32	32, 33, 35, 36, 47	CH-34	<i>E</i>	124.85	5.31	32, 33, 36, 47
C-35		135.38			C-35		135.38		
CH <sub>2</sub> -36		40.30	2.12	34, 35, 37, 47	CH <sub>2</sub> -36		40.30	2.12	<sup>e</sup>
CH <sub>2</sub> -37		27.19	2.21		CH <sub>2</sub> -37		27.21	2.21	<sup>e</sup>
CH-38	<i>E</i>	124.87	5.29	36, 37, 39, 40, 46	CH-38	<i>E</i>	124.90	5.30	36, 37, 40, 46
C-39		135.12			C-39		135.25		
CH <sub>2</sub> -40		40.30	2.10	38, 39, 41, 42, 46	CH <sub>2</sub> -40		40.30	2.11	<sup>e</sup>
CH <sub>2</sub> -41		27.32	2.19		CH <sub>2</sub> -41		27.30	2.19	<sup>e</sup>
CH-42	<i>ω</i>	125.02	5.25	40, 41, 43, 44, 45	CH-42	<i>ω</i>	125.03	5.25	40, 41, 44, 45
C-43		131.21			C-43		131.46		
CH <sub>3</sub> -44	<i>ω</i>	17.82	1.58	42, 43, 45	CH <sub>3</sub> -44	<i>ω</i>	25.91	1.69	42, 43, 45
CH <sub>3</sub> -45	<i>ω</i>	25.92	1.69	42, 43, 44	CH <sub>3</sub> -45	<i>ω</i>	17.82	1.58	42, 43, 44
CH <sub>3</sub> -46	<i>E</i>	16.20	1.62	38, 39, 40	CH <sub>3</sub> -46	<i>E</i>	16.20	1.62	38, 39, 40
CH <sub>3</sub> -47	<i>E</i>	16.19	1.64	34, 35, 36	CH <sub>3</sub> -47	<i>E</i>	16.20	1.63	34, 35, 36
CH <sub>3</sub> -48	Z	23.73	1.75	30, 31, 32	<b>CH<sub>3</sub>-48</b>	<i>E</i>	<b>16.20</b>	<b>1.65</b>	30, 31, 32
CH <sub>3</sub> -49	Z	23.76	1.77	26, 27, 28	CH <sub>3</sub> -49	Z	23.76	1.76	26, 27, 28
CH <sub>3</sub> -50	Z	23.80	1.79	22, 23, 24	CH <sub>3</sub> -50	Z	23.78	1.78	22, 23, 24
CH <sub>3</sub> -51	Z	23.82	1.82	18, 19, 20	CH <sub>3</sub> -51	Z	23.81	1.81	18, 19, 20
CH <sub>3</sub> -52	Z	23.87	1.86	14, 15, 16	CH <sub>3</sub> -52	Z	23.85	1.86	14, 15, 16
CH <sub>3</sub> -53	Z	23.98	1.98	10, 11, 12	CH <sub>3</sub> -53	Z	23.92	1.95	10, 11, 12
CH <sub>3</sub> -54	Z	24.19	2.19	6, 7, 8	CH <sub>3</sub> -54	Z	24.12	2.11	6, 7, 8
CH <sub>3</sub> -55	Z	25.73	3.11	2, 3, 4	CH <sub>3</sub> -55	Z	25.35	2.82	2, 3, 4

<sup>a</sup>C, CH-, CH<sub>2</sub>-, and CH<sub>3</sub>- are colored in white, blue, pink, and gray. <sup>b</sup>The numbers for **2** that are significantly different from those for **1** are in bold. <sup>c</sup>The proton and carbon chemical shift values for carbons with directly attached protons represent positions of the corresponding crosspeaks in the HSQC spectra. <sup>d</sup>The chemical shift values of quaternary carbons represent positions of the corresponding crosspeaks in the HMBC spectra. <sup>e</sup>The connectivities could not be unambiguously established due to strong signal overlaps.

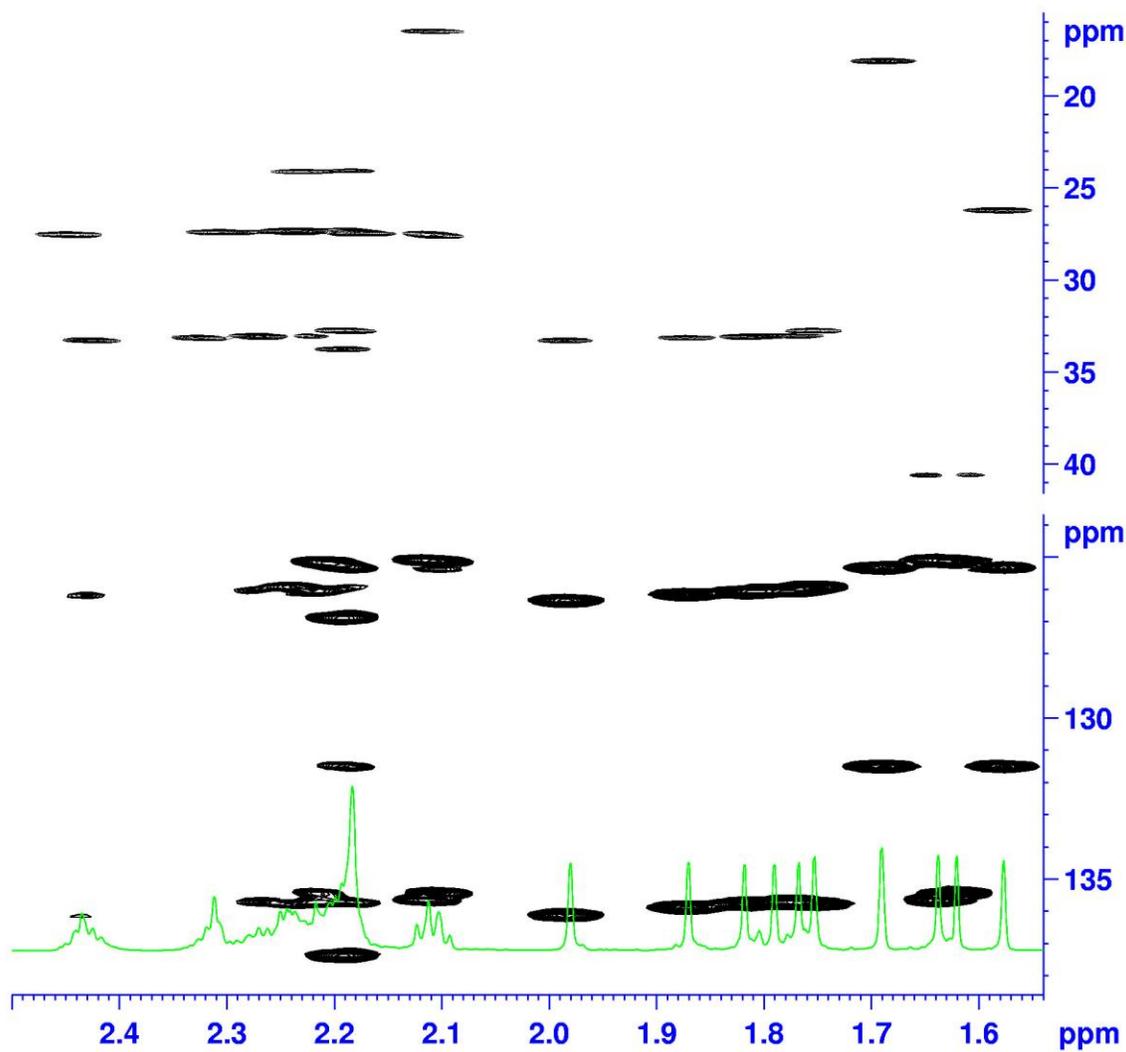
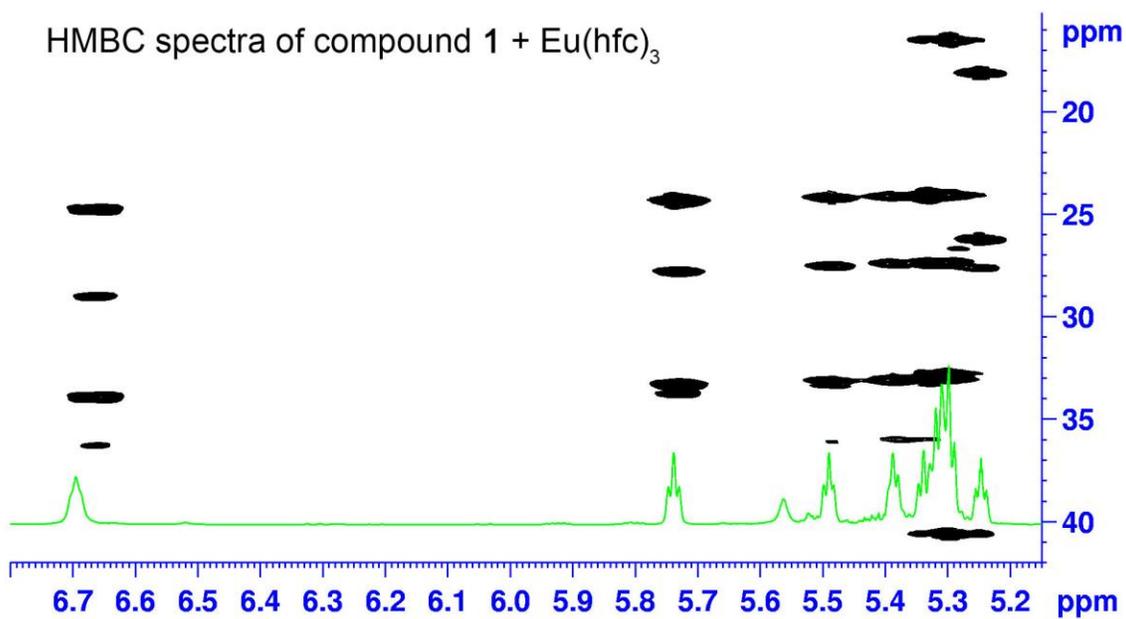
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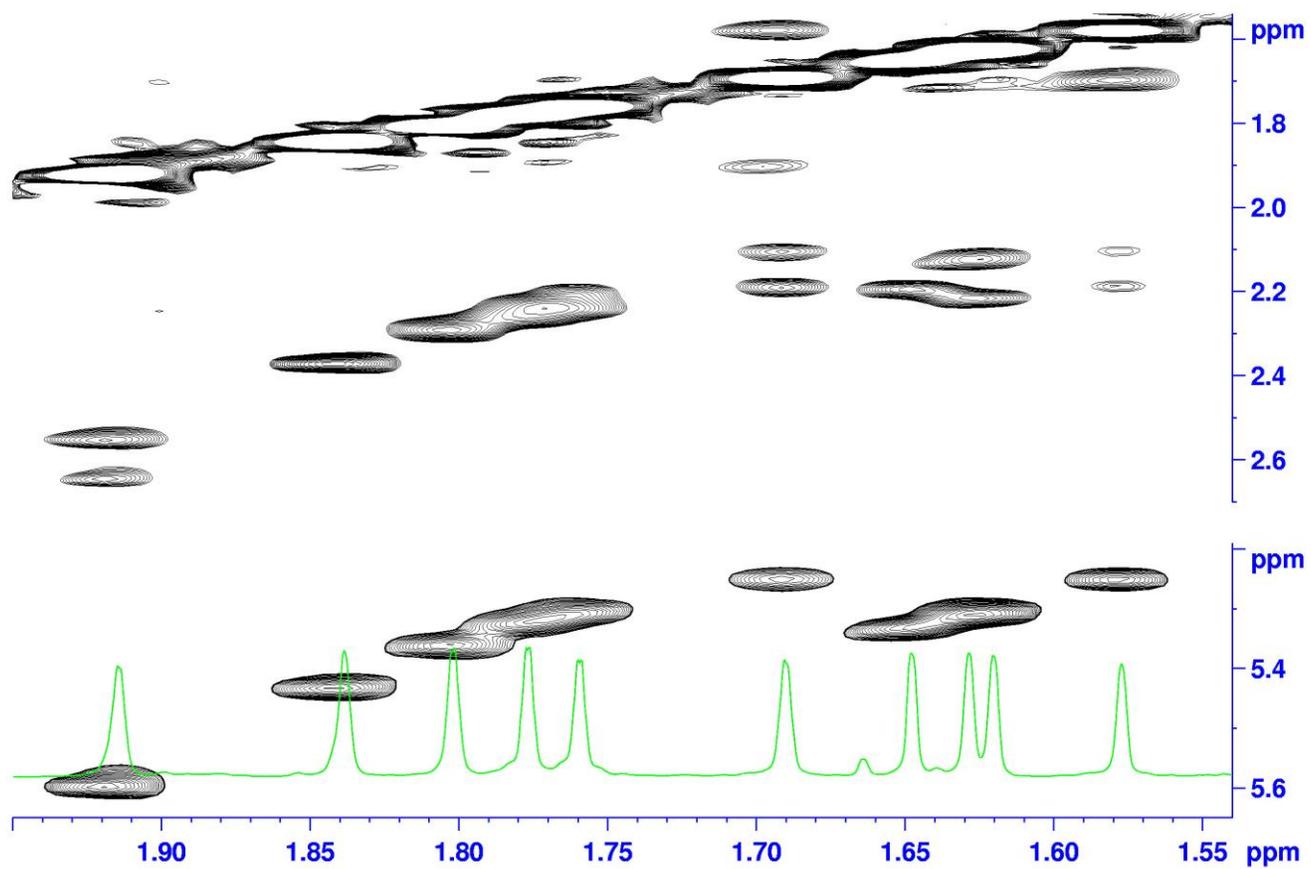
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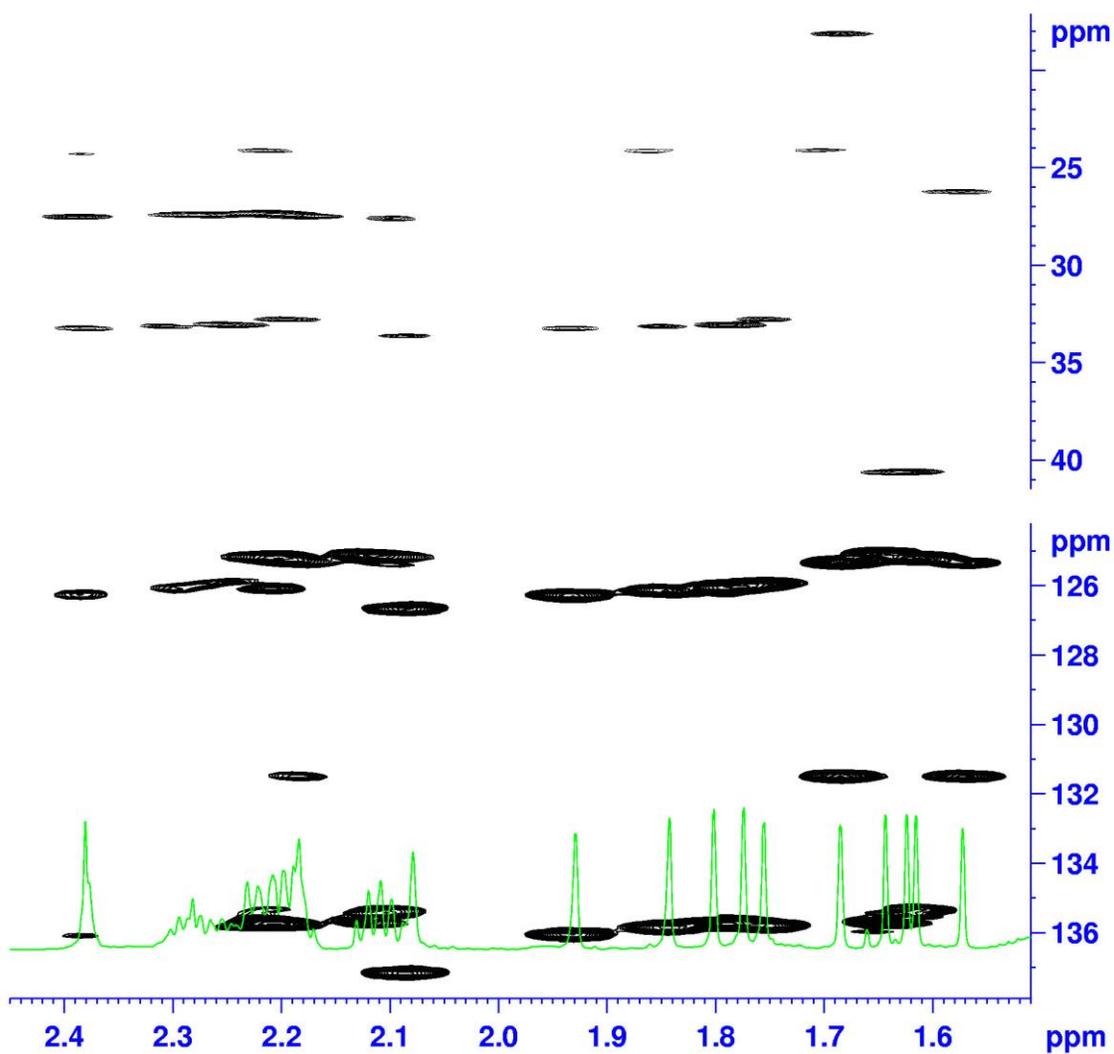
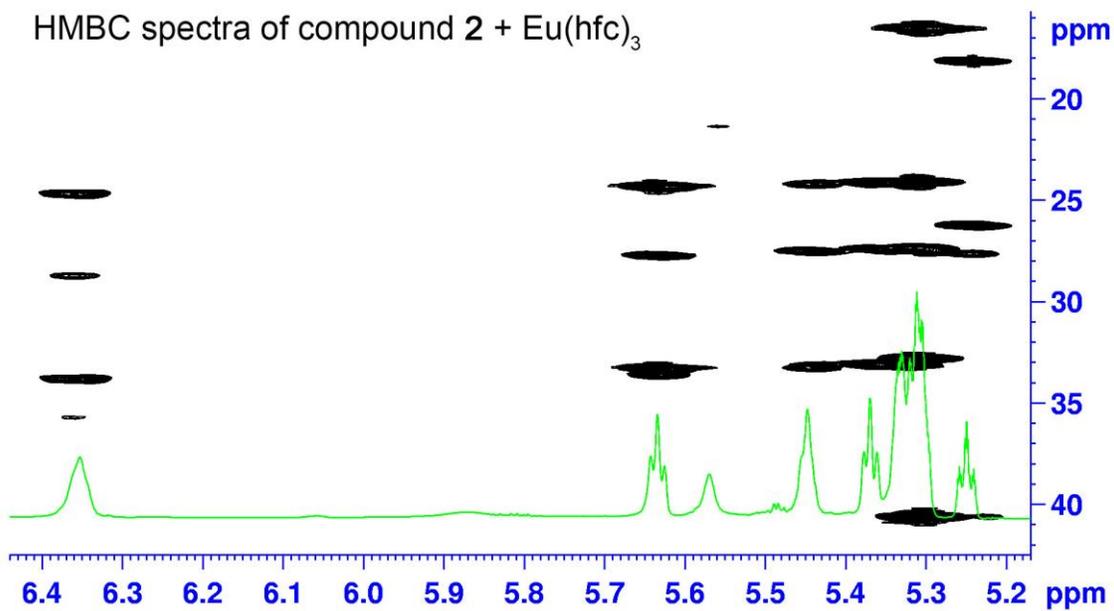




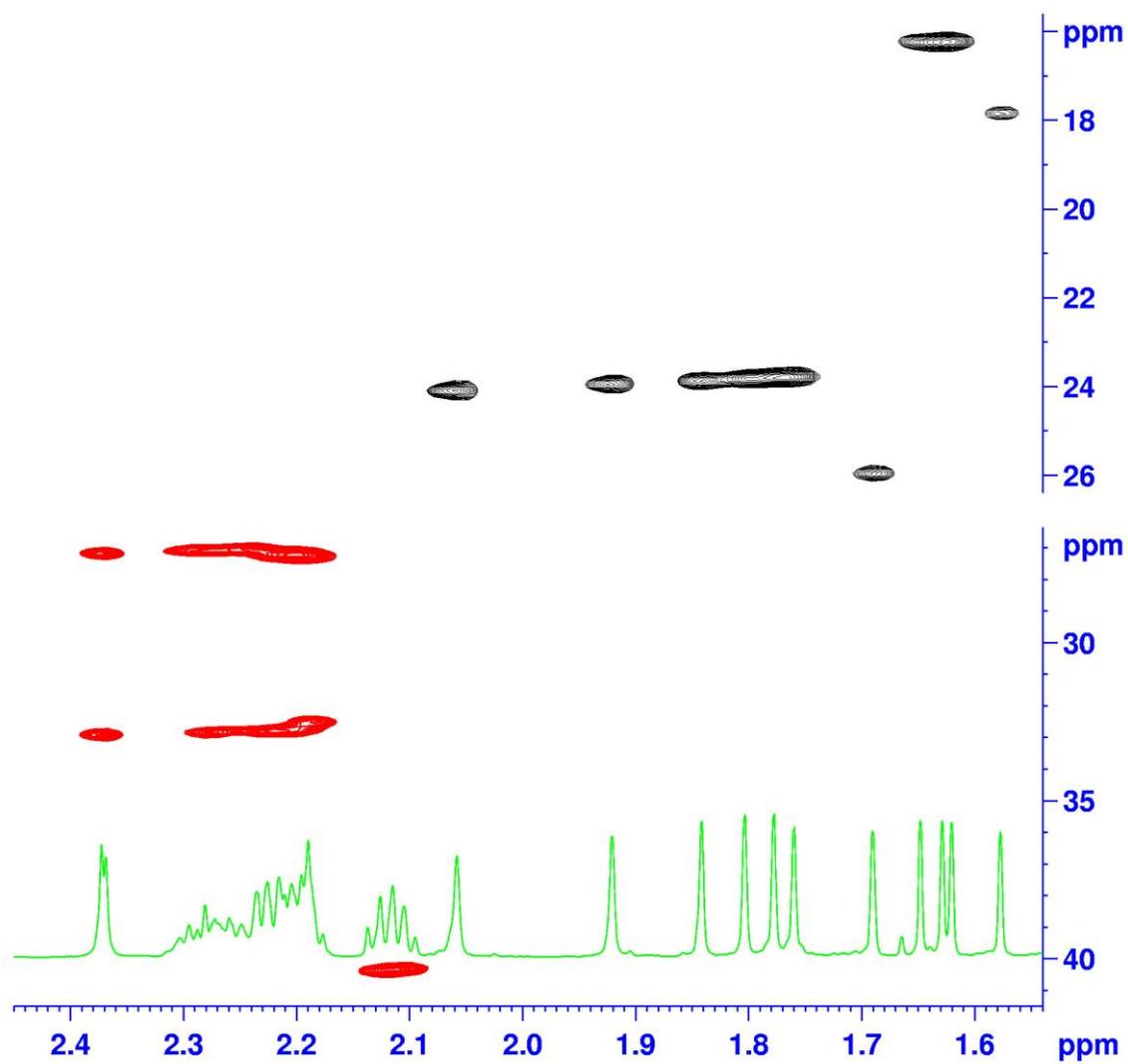
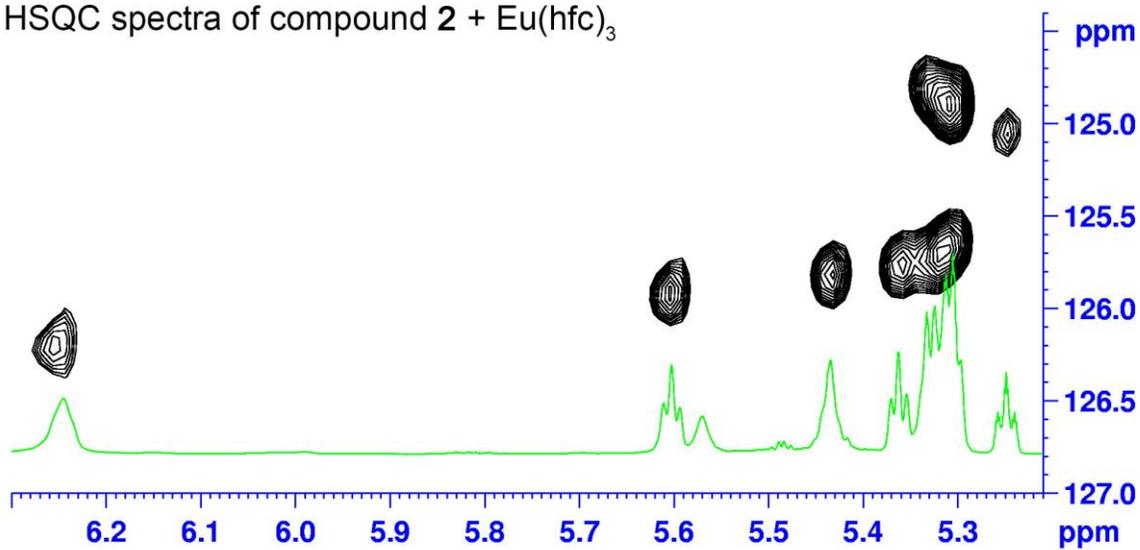
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HMBC spectra of compound 2 + Eu(hfc)<sub>3</sub>



HSQC spectra of compound 2 + Eu(hfc)<sub>3</sub>











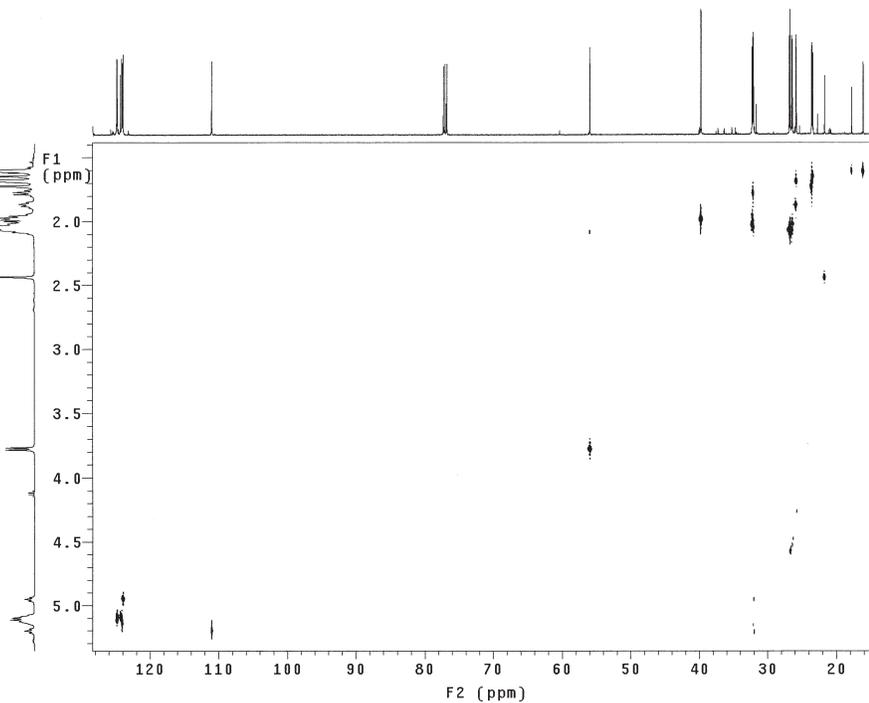
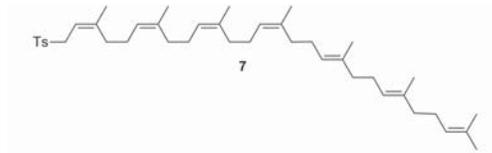




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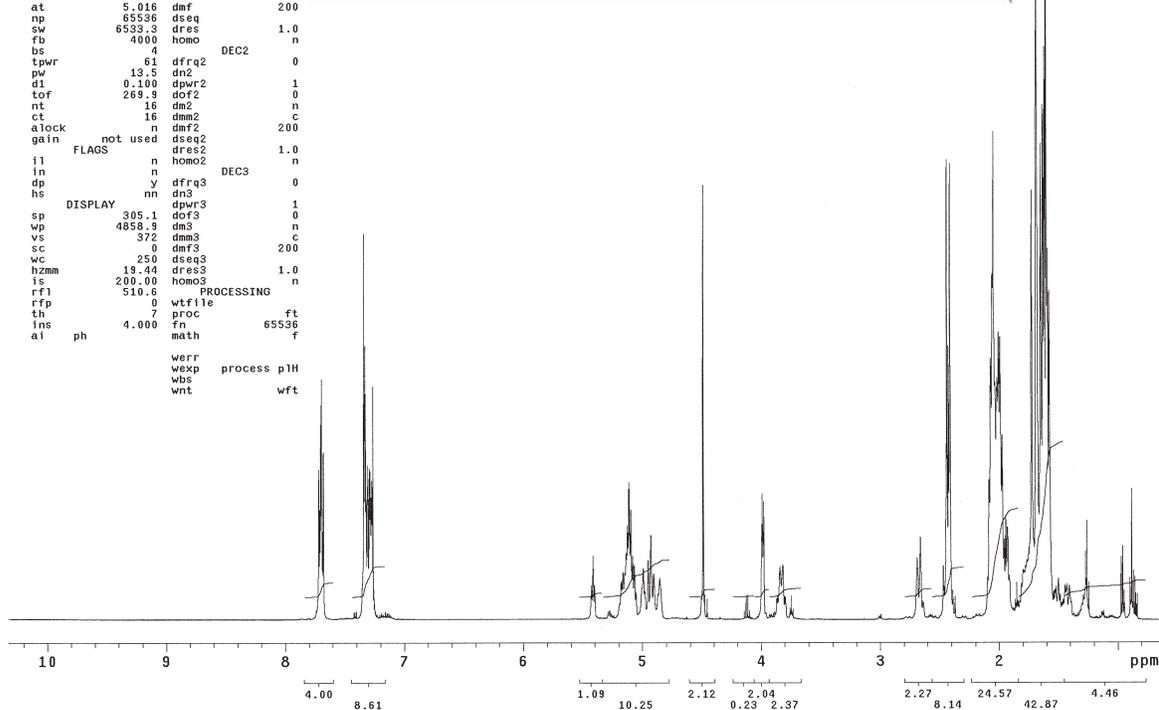
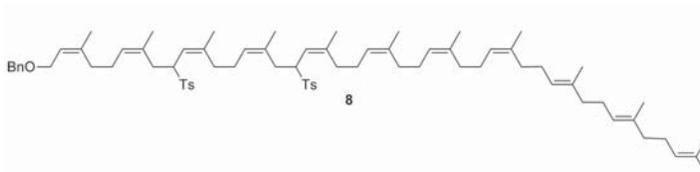
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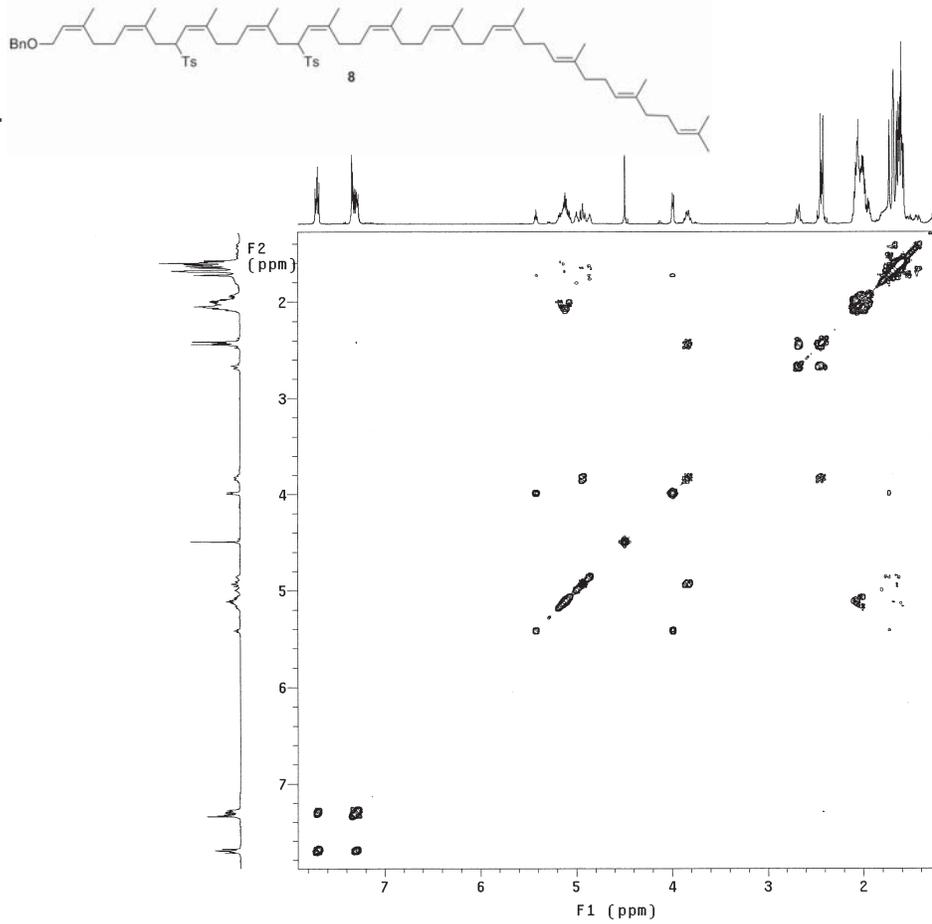
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ACQUISITION			
sfrq	499.864	dm	nm
tn	H1	dmm	c
at	5.016	dmf	200
np	65536	dseq	1.0
sw	6533.3	dres	n
fb	4000	homo	n
bs	4	DEC2	0
tpwr	61	dfrq2	0
pw	13.5	dn2	1
d1	0.100	dpwr2	0
tof	269.9	dof2	n
nt	16	dm2	n
ct	16	dmm2	c
alock	n	dmf2	200
gain	not used	dseq2	1.0
FLAGS			
il	n	dres2	n
ln	n	homo2	n
dp	y	dfrq3	0
hs	nn	dn3	0
DISPLAY			
sp	305.1	dpwr3	1
vp	4858.9	dof3	0
vs	372	dmm3	n
sc	0	dmf3	c
sc	0	dmf3	200
wc	250	dseq3	n
hzam	19.44	dres3	1.0
is	200.00	homo3	n
rfl	510.6	PROCESSING	
rfp	0	wf11e	ft
th	7	proc	65536
ins	4.000	fn	f
al	ph	math	f
werr			
wexp			
wbs			
wnt			
process p1H			
wft			



UPH-14

Pulse Sequence: relayh  
Solvent: CDCl3  
Ambient temperature  
INOVA-500 "nmr2a.chem.nd.edu"

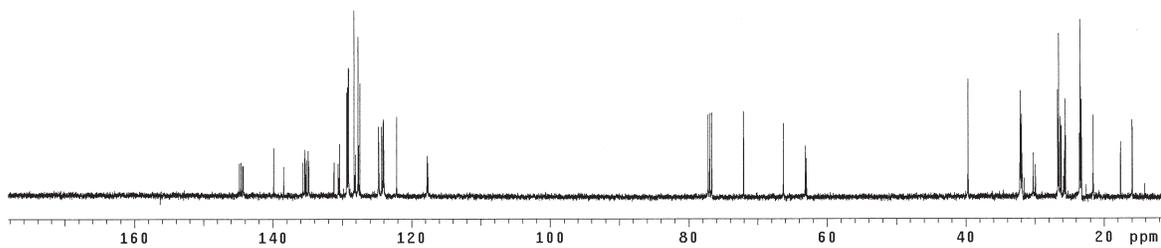
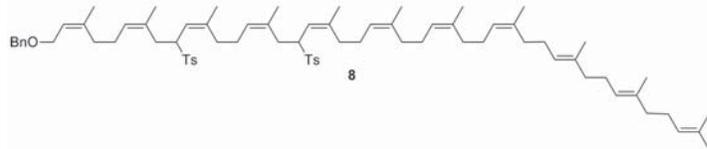
Relax. delay 1.300 sec  
CDSY 40-90  
Acq. time 0.137 sec  
Width 3734.3 Hz  
2D Width 3734.3 Hz  
8 repetitions  
256 increments  
OBSERVE H1, 499.8611751 MHz  
DATA PROCESSING  
Sine bell 0.089 sec  
F1 DATA PROCESSING  
Sine bell 0.034 sec  
FT size 1024 x 1024  
Total time 50 min, 45 sec



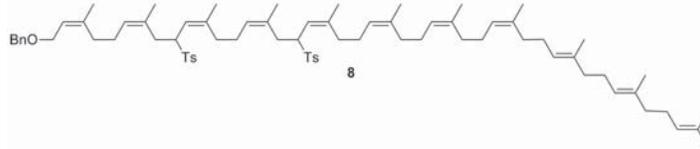
UPH-14

exp7 s2pu1

SAMPLE		DEC. & VT	
date	Jun 20 2013	dfrq	499.864
solvent	CDCl3	dn	H1
file	exp	dpwr	40
ACQUISITION			
sfrq	125.702	dof	0
tn	C13	dmm	yy
at	1.215	dmf	8787.35
np	65536	dseq	
sw	26983.3	dres	1.0
fb	15000	homo	n
bs	4		
tpwr	52	dfrq2	0
pw	15.0	dn2	
d1	1.800	dpwr2	1
tor	144.5	dof2	0
nt	12000	dm2	n
ct	360	dmm2	c
alock	n	dmf2	10000
gain	not used	dseq2	1.0
flags	n	dres2	n
il	n	homo2	n
in	n		
dp	y	dfrq3	0
hs	nn	dn3	
DISPLAY			
sp	1432.7	dpwr3	1
wp	26974.9	dof3	0
vs	365	dm3	n
sc	0	dmm3	c
wc	250	dmf3	10000
hzmm	83.90	dseq3	1.0
is	500.00	dres3	n
rfl	1437.4	homo3	n
PROCESSING			
rfp	0	lb	1.00
th	6	wf11e	
ins	100.000	proc	ft
ai	cdc ph	fn	131072
		math	f
		werr	
		wexp	
		wbs	
		wnt	



UPH-14  
Pulse Sequence: dept



CH3 carbons



CH2 carbons



CH carbons



all protonated carbons



140 130 120 110 100 90 80 70 60 50 40 30 20 ppm

UPH-14  
Pulse Sequence: hetcor  
Solvent: CDCl3  
Ambient temperature  
User: 1-14-87  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 3682.9 Hz  
16 repetitions  
256 increments  
OBSERVE C13, 125.6902015 MHz  
DECOUPLE H1, 499.8634332 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 1 hr, 54 min, 7 sec

