

Fluorescent and cooperative ion pair receptor based on tolan for Na⁺ (or Li⁺) and HSO₄⁻ : logic AND gate

June-Ho Shin, Jung-Ho Hong, Min-Sung Ko, and Dong-Gyu Cho*

Department of Chemistry, Inha University, Functional Molecule Synthesis Laboratory, Incheon 402-751, Republic of Korea

dgcho@inha.ac.kr

Contents

1. General experimental and synthetic details	S2
2. Spectroscopic titrations	S3
3. DFT calculations	S24
4. Quantum Yields	S26
5. NMR Spectra	S27

1. General experimental and synthetic details

General experimental

Reagents were purchased at the highest commercial quality and used without further purification, unless otherwise stated. Yields of synthesized compounds were measured after chromatographic purification. UV-vis spectra were recorded on a UV 1800 (Shimadzu) spectrophotometer. Proton and ¹³C-NMR spectra were measured at 25 °C using Jeol 400 instruments.

Synthetic Details

2,2'-(phenylazanediy)bis(ethan-1-ol) (1)

In a round-bottom flask fitted with a reflux condenser, 2-iodoaniline (1 g, 2.56 mmol), CaCO₃ (0.9 g, 5.12 mmol), KI (90 mg, 0.26 mmol), and water (6 mL) were charged. The resulting solution was then stirred at reflux for 12 h. After cooling to room temperature, the mixture was filtered and then the solution was extracted with water and EtOAc. The combined organic layers were dried over anhydrous Na₂SO₄ and then filtered; the filtrate was concentrated under reduced pressure to obtain the residue. The residue was purified over silica gel to afford **1** (250 mg, 54%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.86 (d, *J* = 9.4 Hz, 1H), 7.39 – 7.32 (m, 1H), 7.28 (dd, *J* = 8.0, 1.7 Hz, 1H), 6.92 (ddd, *J* = 8.0, 7.2, 1.7 Hz, 1H), 3.59 (m, 4H), 3.19 (m, *J* = 9.5 Hz, 4H), 2.47 (s, 2H), ¹³C NMR (101 MHz, CHLOROFORM-*D*) δ 151.71, 139.69, 129.87, 127.92, 126.23, 102.86, 60.02, 57.94, HRMS–EI: *m/z* [M]⁺ calcd for C₁₀H₁₄NO₂I: 307.0069; found:307.0070.

13-(2-iodophenyl)-1,4,7,10-tetraoxa-13-azacyclopentadecane (2)

In a round-bottom flask fitted with a reflux condenser, compound **1** (0.5 g, 1.63 mmol) and di-tosylate (0.60 g, 1.30 mmol) were dissolved in THF (40 mL). NaH (0.31 g, 13.04 mmol) was slowly added to the mixture, which was then stirred at reflux for 3 days. After cooling to room temperature, the solution was concentrated under reduced pressure to afford a residue. The residue was extracted with water and EtOAc. The combined organic layers were dried over anhydrous Na₂SO₄ and then filtered; the filtrate was concentrated under reduced pressure to obtain the residue. The residue was purified over silica gel to afford **2** (137 mg, 20%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.82 (dd, *J* = 7.9, 1.4 Hz, 1H), 7.33 – 7.25 (m, 2H), 6.77 (ddd, *J* = 7.9, 6.7, 2.1 Hz, 1H), 3.69 (s, 4H), 3.68 – 3.58 (m, 12H), 3.38 (t, *J* = 6.2 Hz, 4H), ¹³C NMR (101 MHz, CHLOROFORM-*D*) δ 152.92, 139.99, 128.91, 125.85, 125.40, 100.83, 70.95, 70.73, 70.57, 70.45, 54.47, HRMS–EI: *m/z* [M]⁺ calcd for C₁₆H₂₄NO₄I: 421.0750; found:421.0751.

1-(2-((2-(1,4,7,10-tetraoxa-13-azacyclopentadecan-13-yl)phenyl)ethynyl)phenyl)-3-hexylurea (4)

3a (160 mg, 0.712 mmol) and tributyltin methoxide (0.056 mL, 2.13 mmol) were charged in a pressure bottle. The solution was stirred at 120 °C for 4 h. The resulting solution of **3b** was used without purification. To a well stirred solution of **2** (200 mg, 0.475 mmol) in DMF (4 mL), Pd(PPh₃)₄ (37 mg, 0.0475 mmol) and the solution of **3b** was added and then flushed with argon for 5-10 min. The solution was stirred at 70 °C for 5h. After cooling to room temperature, the solution was concentrated under reduced pressure to afford a residue. The residue was extracted with water and EtOAc. The combined organic layers were dried over anhydrous Na₂SO₄ and then filtered; the filtrate was concentrated under reduced pressure to obtain the residue. The residue was purified over silica gel to afford **4** (66 mg, 26%). ¹H NMR (400 MHz, Acetone-*d*₆) δ 8.24 (dd, *J* = 8.5, 1.2 Hz, 1H), 7.69 (s, 1H), 7.44 (ddd, *J* = 7.7, 1.6, 0.5 Hz, 1H), 7.41 (dd, *J* = 7.7, 1.7 Hz, 1H), 7.29 – 7.21 (m, 2H), 7.08 (dd, *J* = 8.5, 1.1 Hz, 1H), 6.93 (td, *J* = 7.6, 1.2 Hz, 1H), 6.83 (td, *J* = 7.4, 1.1 Hz, 1H), 6.41 (s, 1H), 3.78 – 3.63 (m, 8H), 3.60 – 3.49 (m, 12H), 3.19 (td, *J* = 7.0, 5.7 Hz, 2H), 1.57 – 1.42 (m, 2H), 1.38 – 1.21 (m, 6H), 0.85 (t, *J* = 7.0 Hz, 3H), ¹³C NMR (101 MHz, ACETONE-*D*₆) δ 155.73, 153.11, 142.23, 136.10, 132.76, 130.45, 129.89, 122.08, 120.62, 119.97, 119.67, 114.67, 112.73, 96.15, 90.26, 71.76, 71.11, 70.75, 54.07, 40.62, 40.50, 32.44, 31.03, 27.46, 23.39, 14.42, HRMS–EI: *m/z* [M]⁺ calcd for C₃₁H₄₃N₃O₅: 537.3203; found: 537.3201.

2. Spectroscopic titrations

Stock solutions of all of the compounds studied were made up in CH₃CN with the final concentrations being between 2.0×10^{-5} M. ACS grade solvents were purchased and used without purification. The stock solutions were appropriately diluted with the solvents for the ensuing studies.

Binding Constants by UV-vis titrations

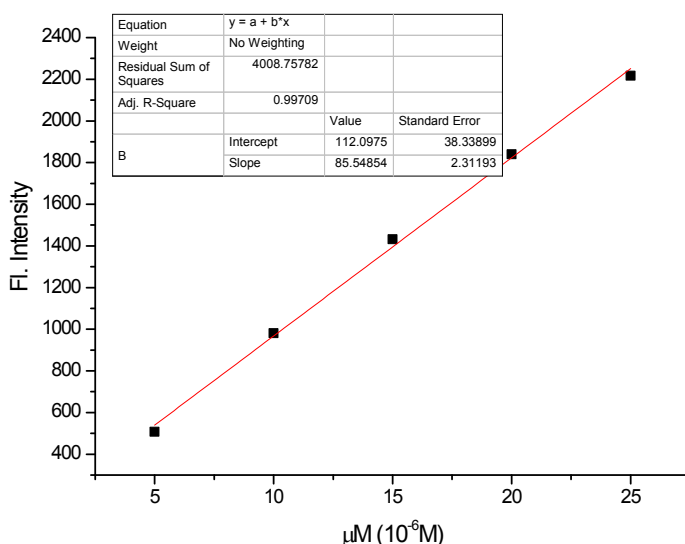
Upon addition of incremental amounts of anions to the solution of receptor **4**, absorbance change of **4** (2.0×10^{-5} M of **4** was used, unless otherwise stated) were recorded in CH₃CN. Equilibrium constants of complexes were calculated using the equation, $y = (b \times x)/(1 + x \times K)$, where $x = [G]$, $y = A - A_0$ (A is the absorbance of the solution of **4** at a certain concentration of anions and A_0 is the absorbance of the solution of **4** without anions).

Binding constants by fluorescent titrations

Upon addition of incremental amounts of anion to the solution of receptor **4**, fluorescence change of each chemosensor (2.0×10^{-5} M) were recorded in CH₃CN by irradiating the solution at $\lambda_{\text{ex}} = 340$ nm. Equilibrium constants of complexes were calculated using the equation below.

$y = (1 + b \times x \times K)/(1 + x \times K)$, where $x = [G]$, $y = F/F_0$ (F intensity at a certain concentration of anions/ F_0 intensity without anions).

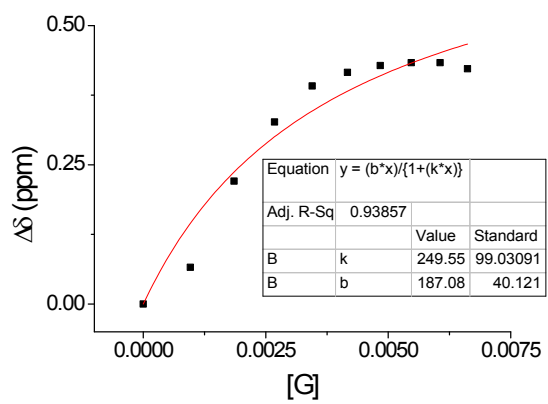
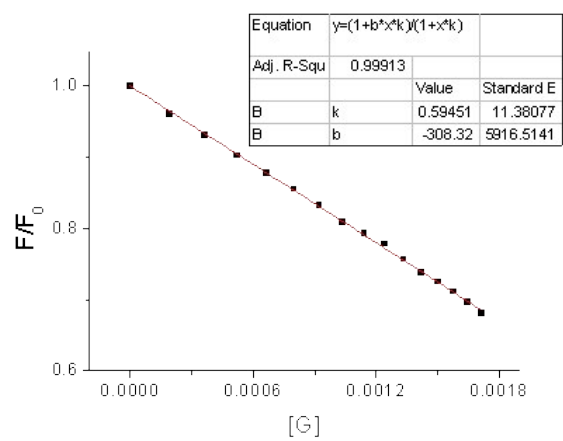
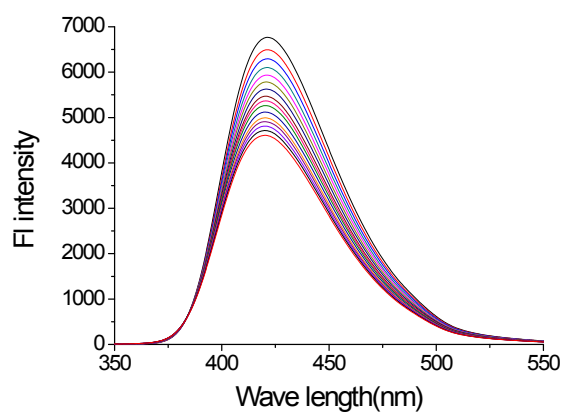
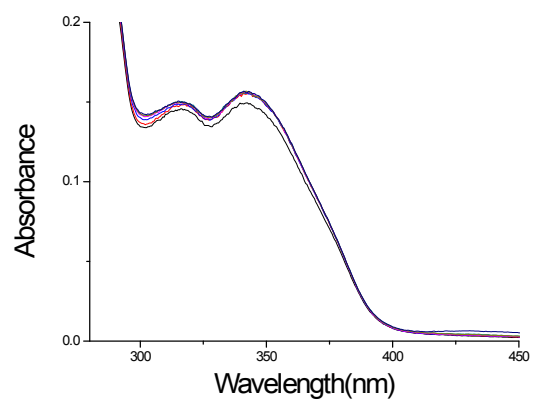
The observed fluorescent intensities were shown below at a various concentrations of **4** in CH₃CN at 420nm.



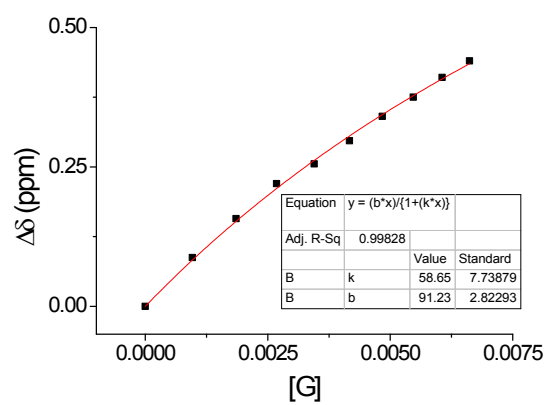
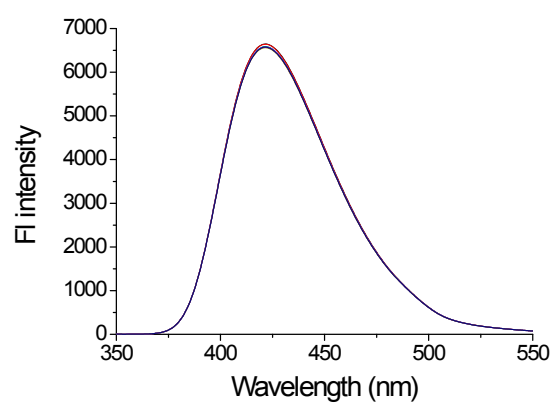
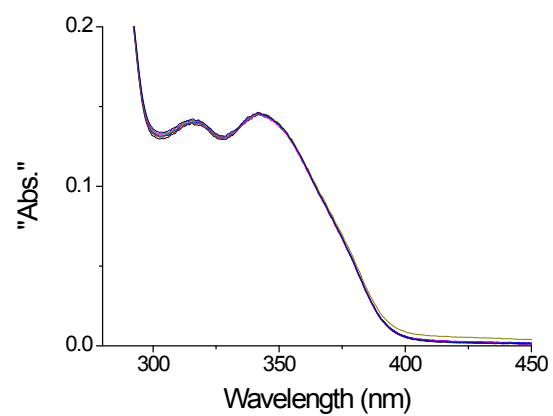
Binding Constants by ¹H NMR titrations

Upon addition of incremental amounts of anions to the solution of receptor **4**, chemical shift change of **4** (4 mM of **4** was used, unless otherwise stated) were recorded in CD₃CN. Equilibrium constants of complexes were calculated using the equation, $y = (b \times x)/(1 + x \times K)$, where $x = [G]$, $y = \delta - \delta_0$ (δ is a chemical shift of **4** at a certain concentration of anions and δ_0 is a chemical shift of **4** without anions).

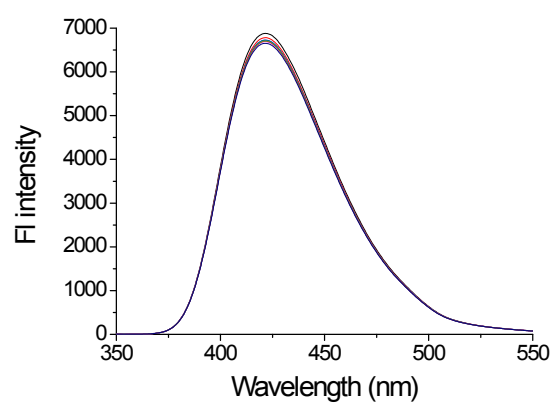
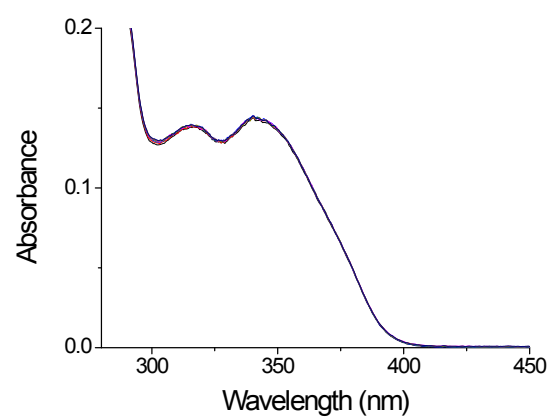
F⁻



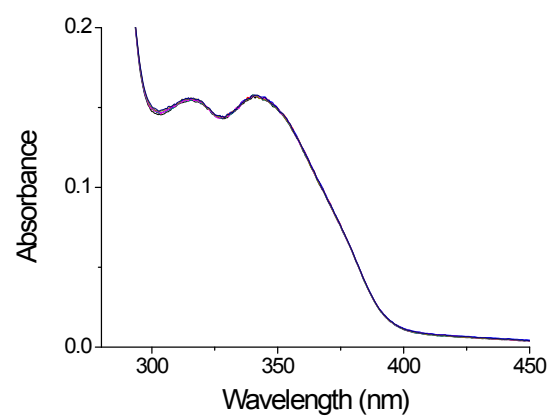
Cl⁻

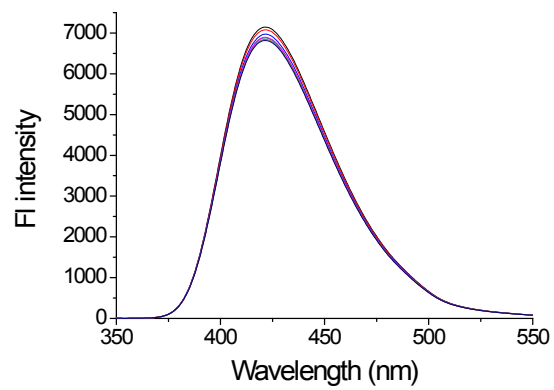


Br⁻

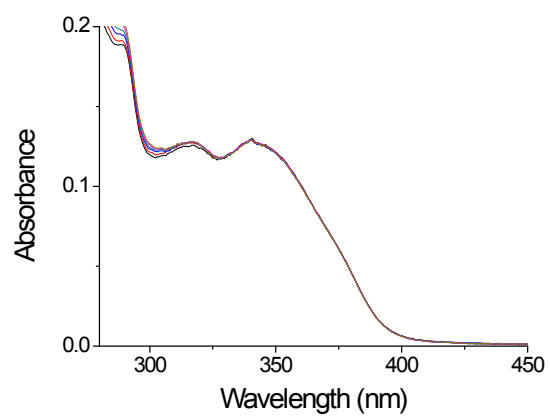


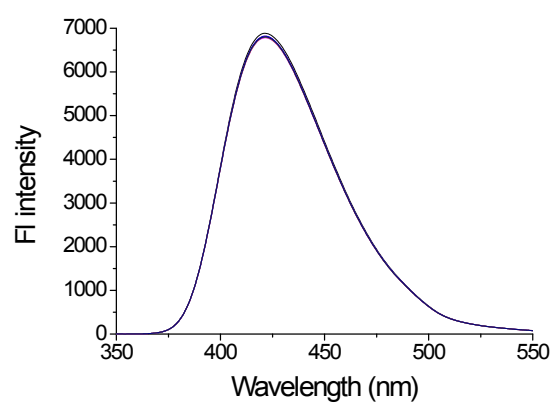
I⁻



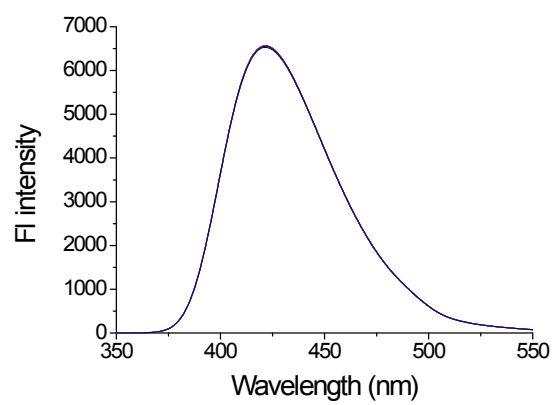
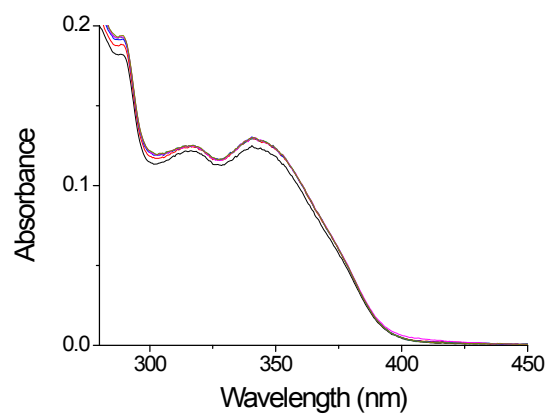


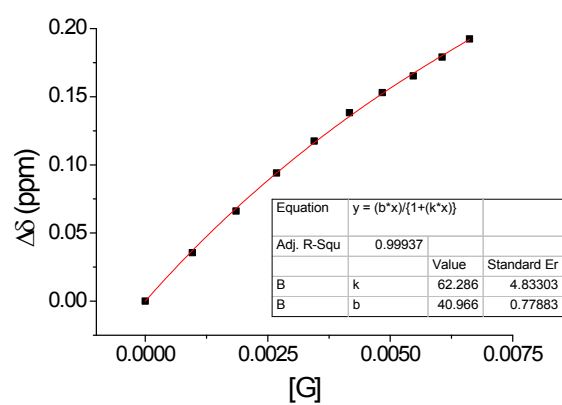
BzO⁻



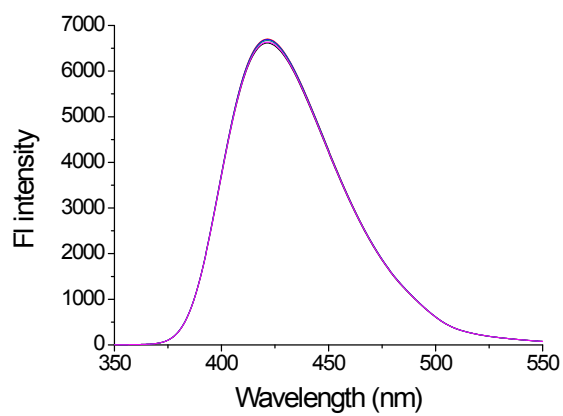
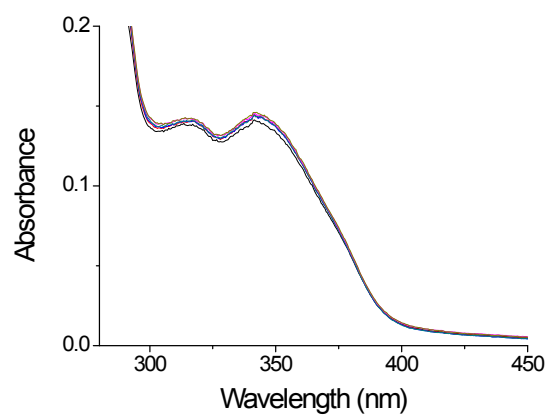


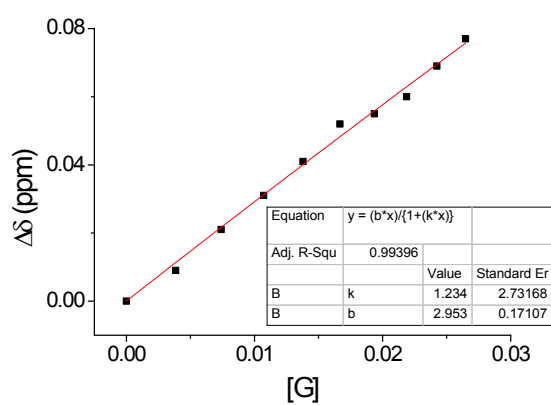
H_2PO_4^-



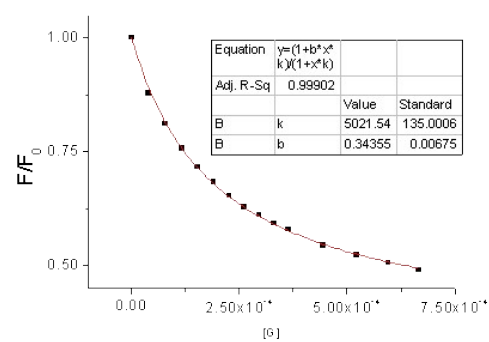
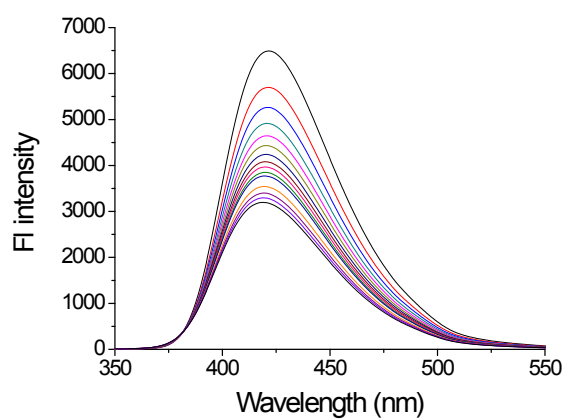
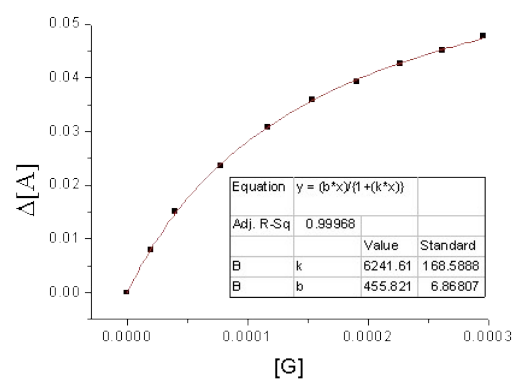
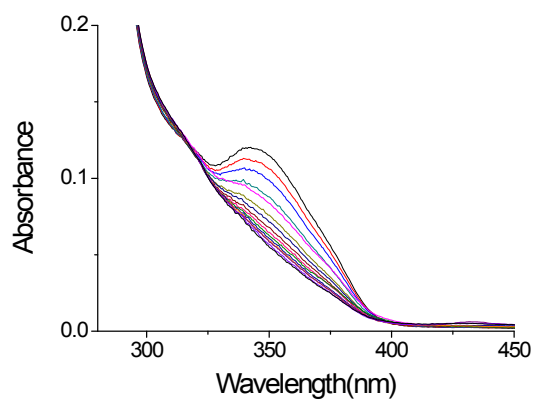


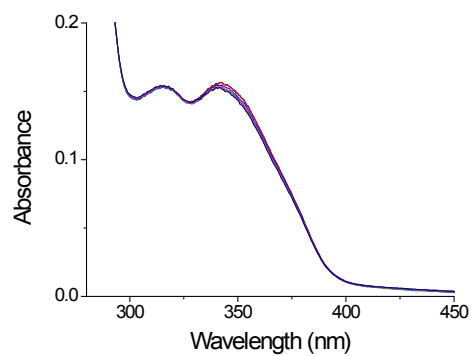
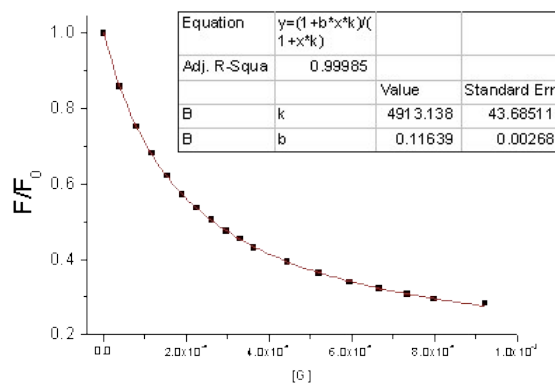
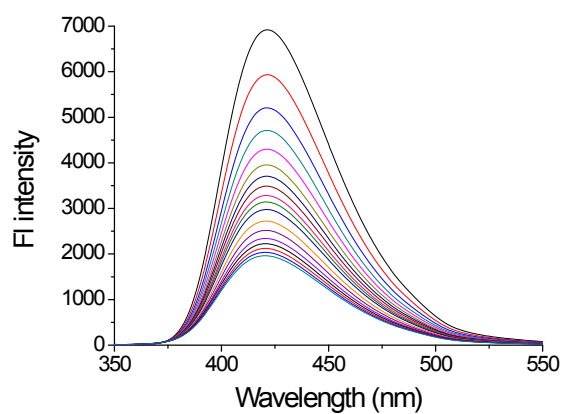
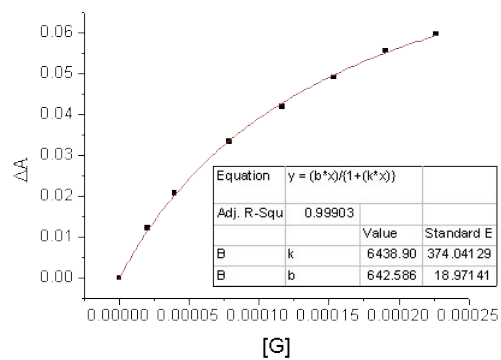
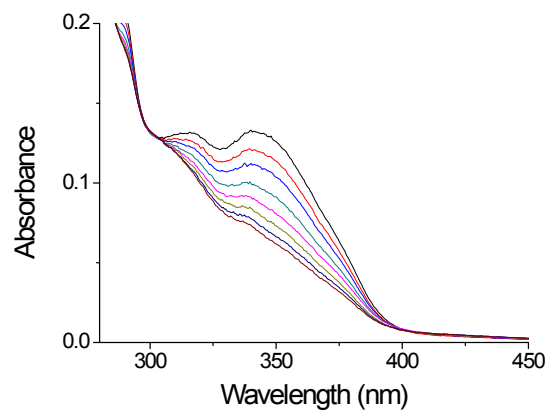
HSO_4^-

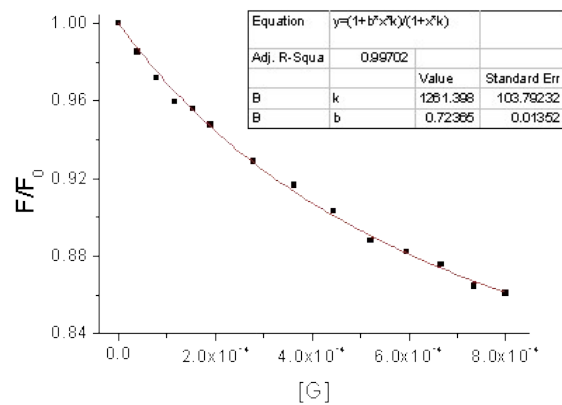
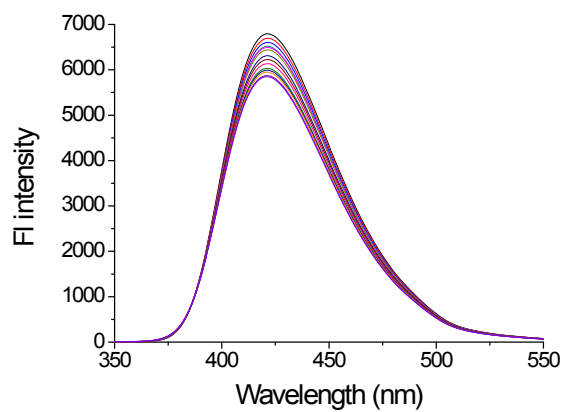




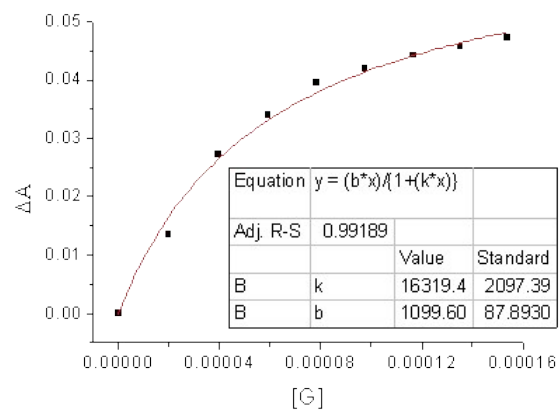
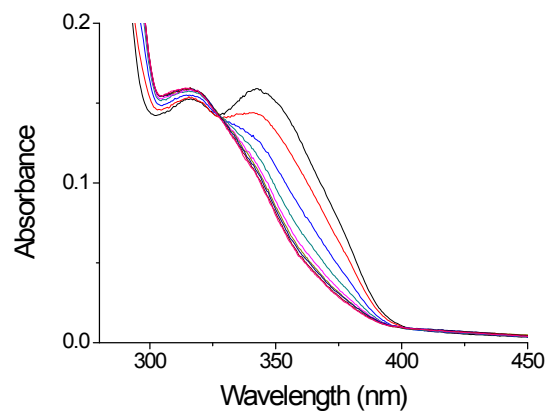
$\text{Li}^+\text{ClO}_4^-$

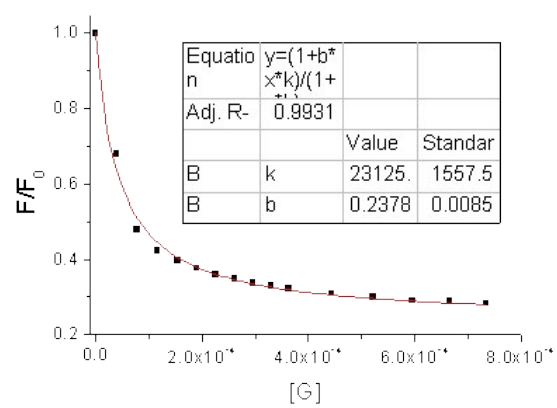
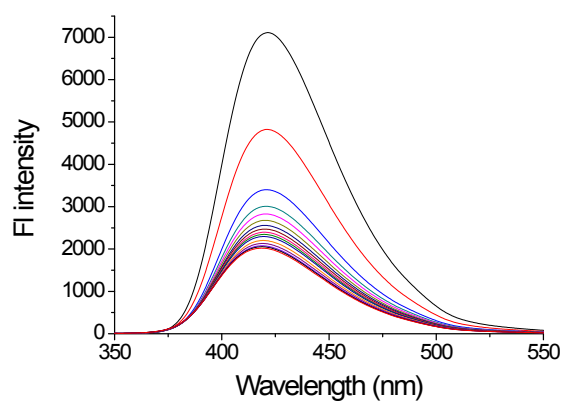




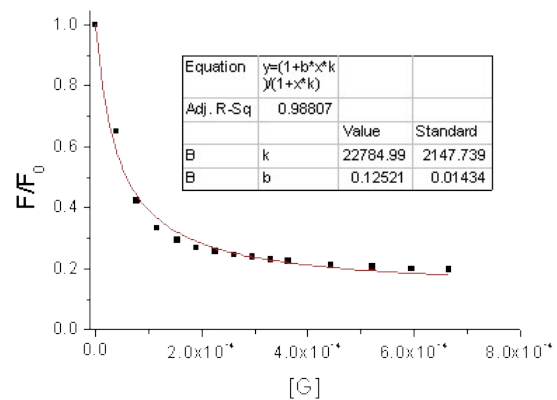
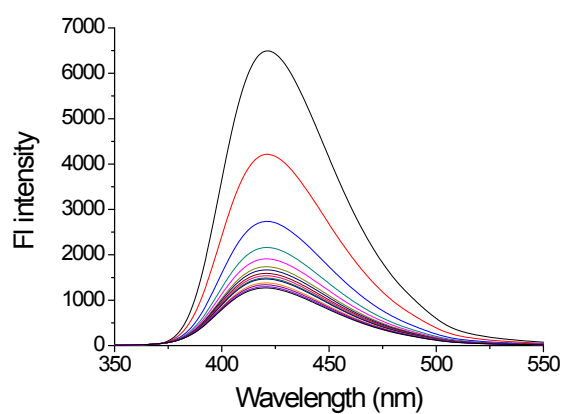
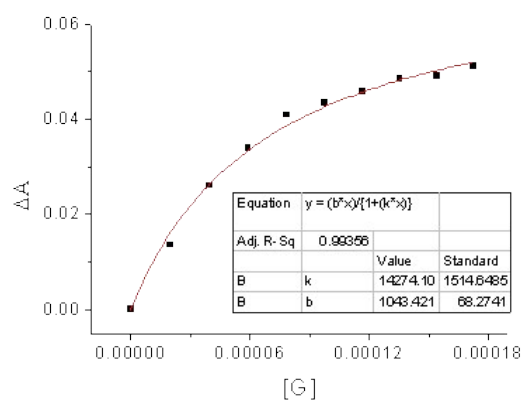
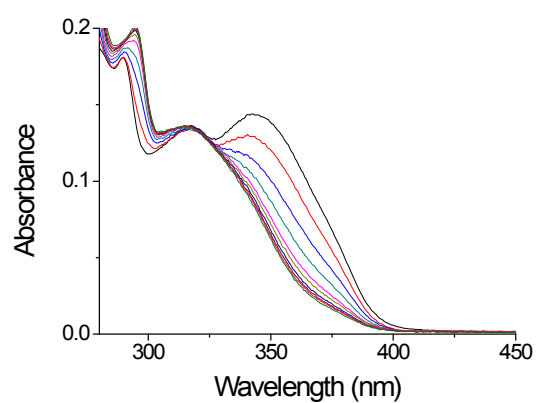


LiHSO_4

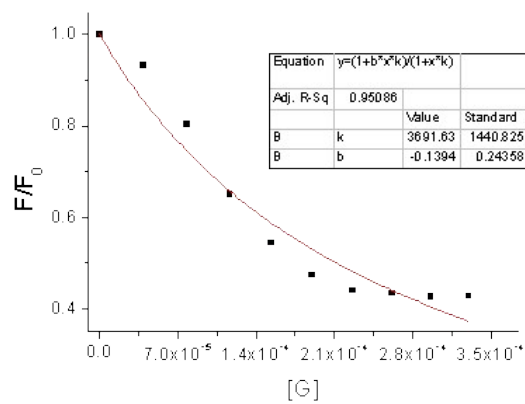
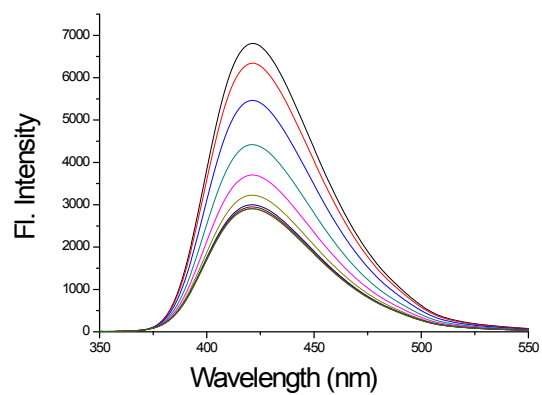
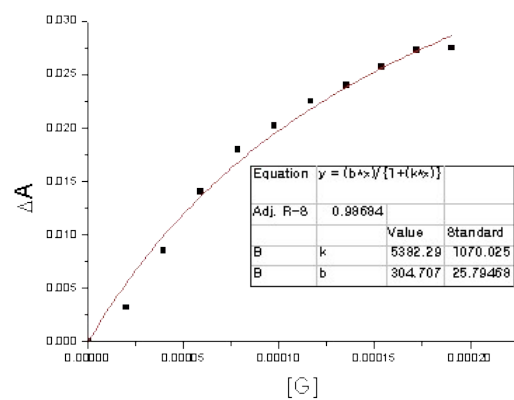
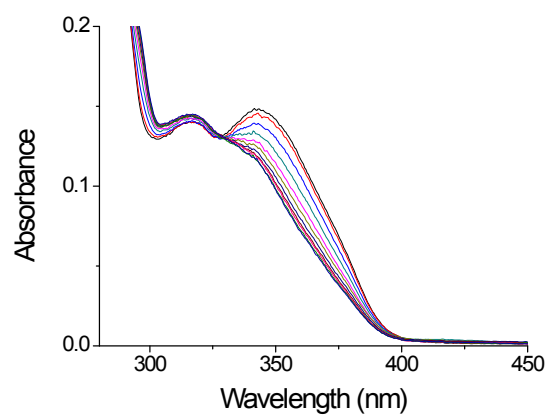




NaHSO₄



KHSO₄



Job Plot for $4 \cdot \text{M}^+ \text{HSO}_4^-$

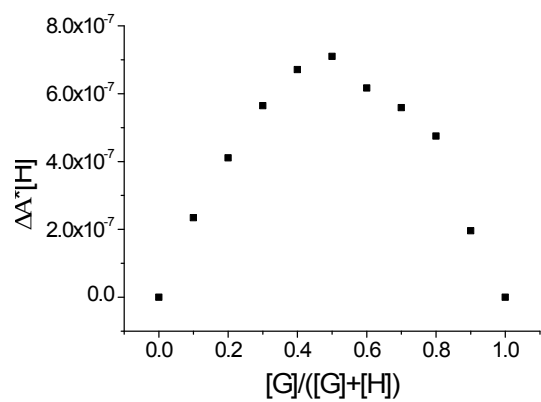


Figure S1. Job plots for LiHSO_4 obtained by UV-vis titrations

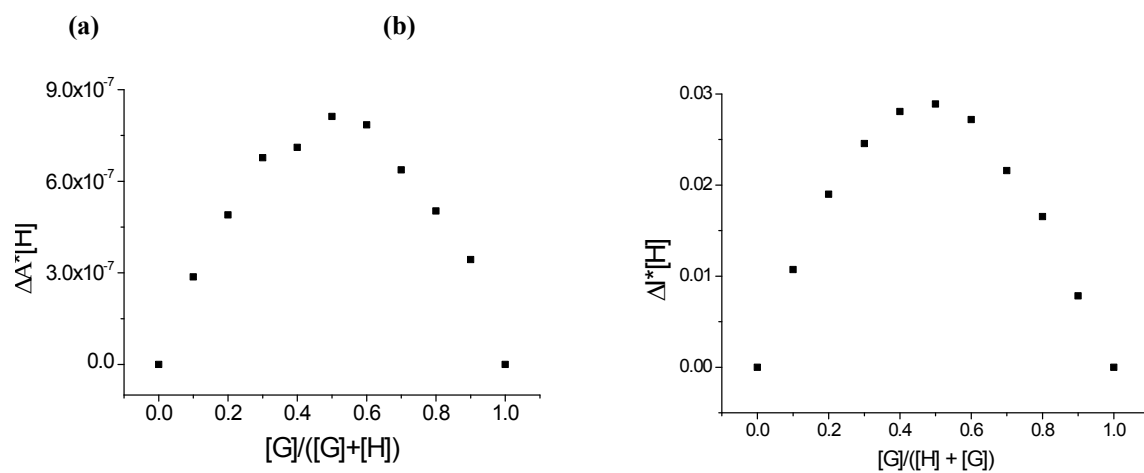


Figure S2. Job plots for NaHSO_4 obtained by a) UV-vis titrations b) fluorescent titrations

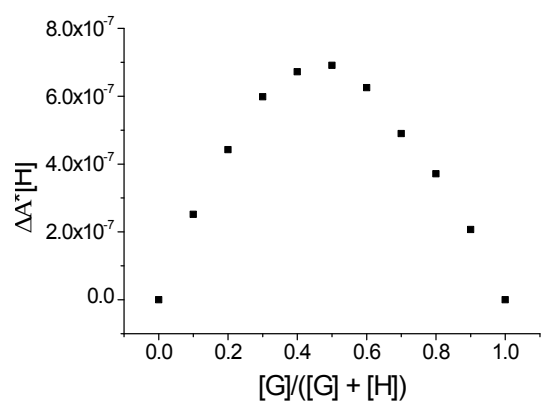


Figure S3. Job plots for KHSO₄ obtained by UV-vis titrations

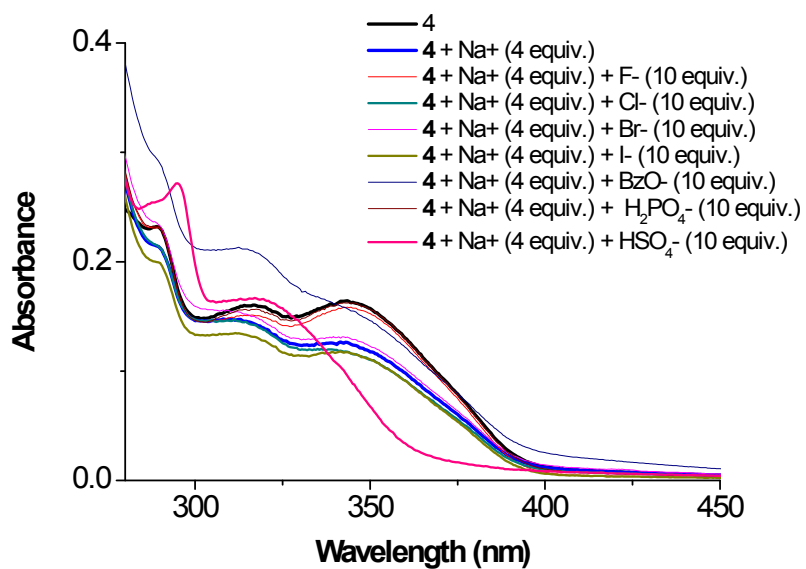


Figure S4. UV-vis spectra changes of **4** (20 μ M in CH₃CN) were recorded upon the addition of various anions (10 equiv.) in the presence of Na⁺ (4equiv.) at 25 °C.

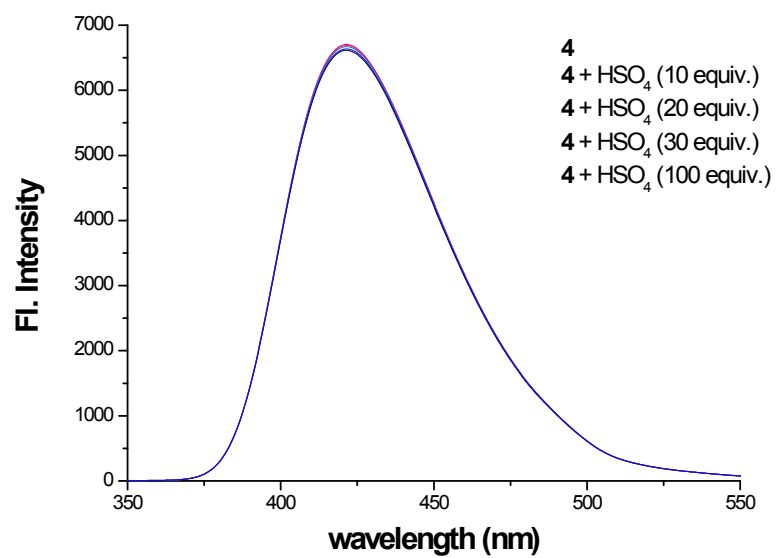


Figure S5. Fluorescent changes of **4** (20 μM in CH₃CN) Upon the addition of various amount of HSO₄⁻ at 25 °C.

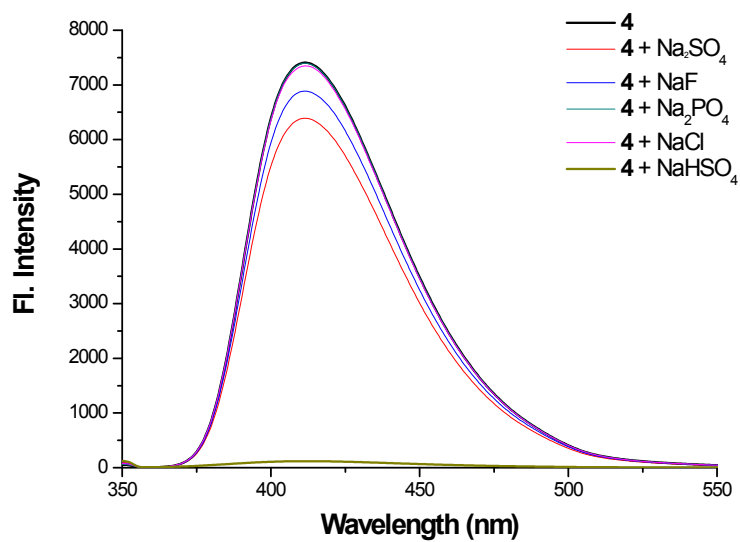


Figure S6. Fluorescent changes of **4** (20 μM in CH_2Cl_2) after the extraction of various inorganic salts (excess amount of solid salts) with the solution at 25 $^\circ\text{C}$.

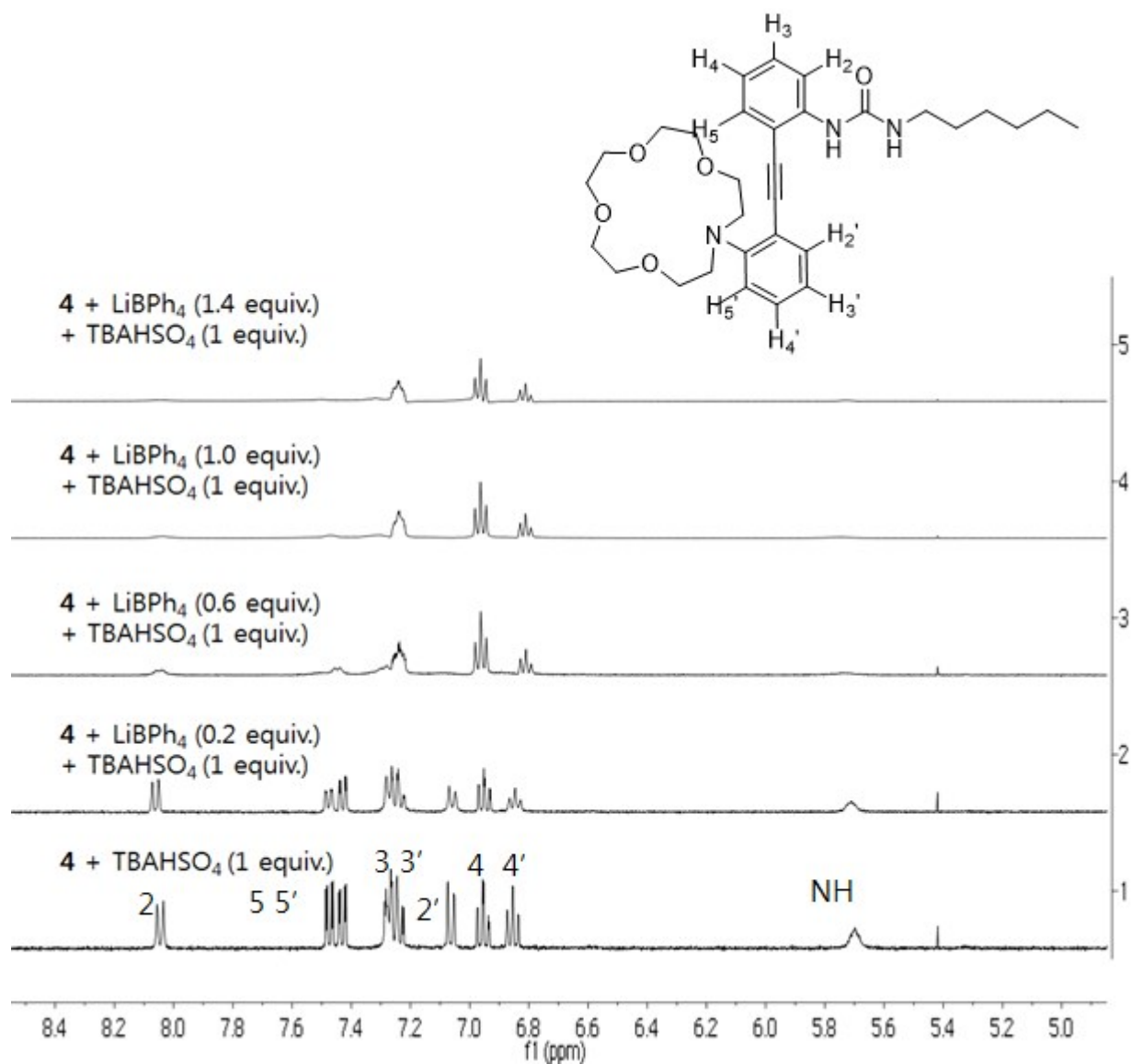


Figure S7. Partial ¹H-NMR spectra recorded during the titration of **4** ([H] = 4.0 × 10⁻³ M in CD₃CN) with LiBPh₄ in the presence of 1 equiv. of TBA·HSO₄

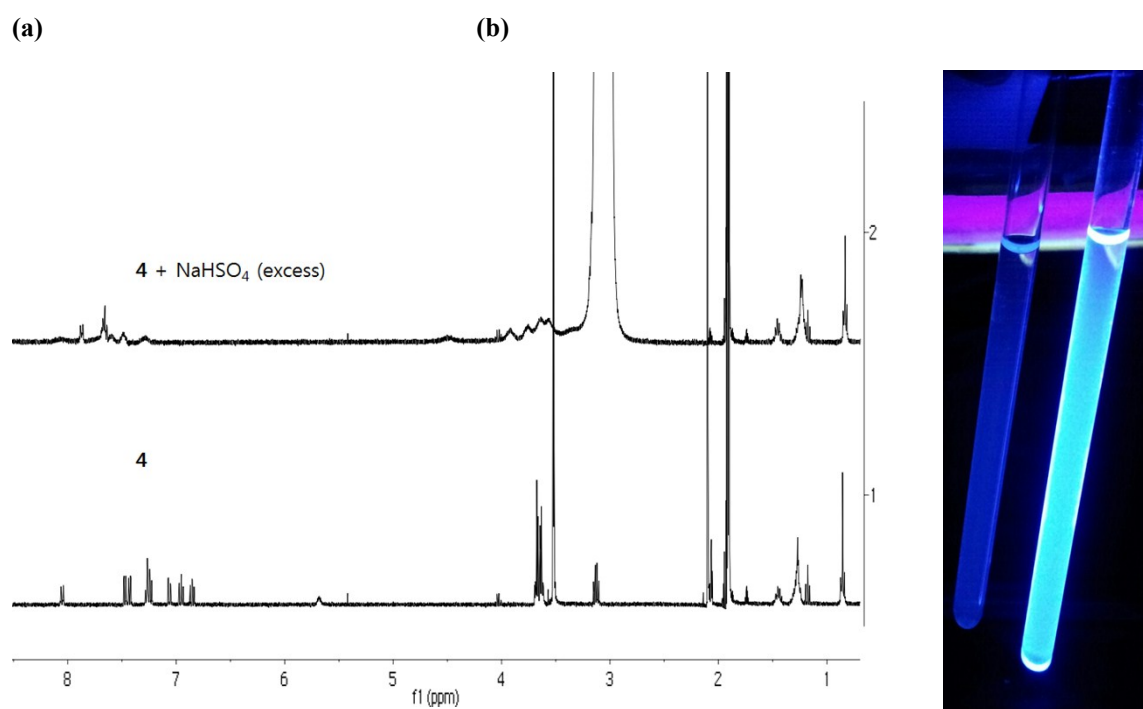


Figure S8. (a) ¹H-NMR spectra of **4** and **4** containing NaHSO₄ after the extraction of solid NaHSO₄ with **4** (4 mM in CD₃CN) (b) left (**4** + NaHSO₄) and right (only **4**) under a laboratory hand-held UV light.

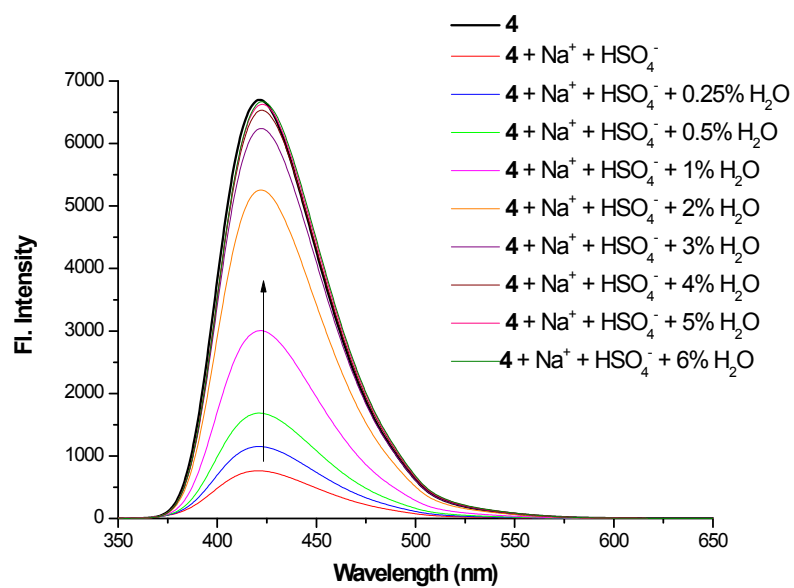


Figure S9. Evolution of the fluorescent spectrum of **4** (2.0×10^{-5} M in CH_3CN containing 4 equiv. of Na^+ and HSO_4^-) during the titration with water (from 0 – 6%)

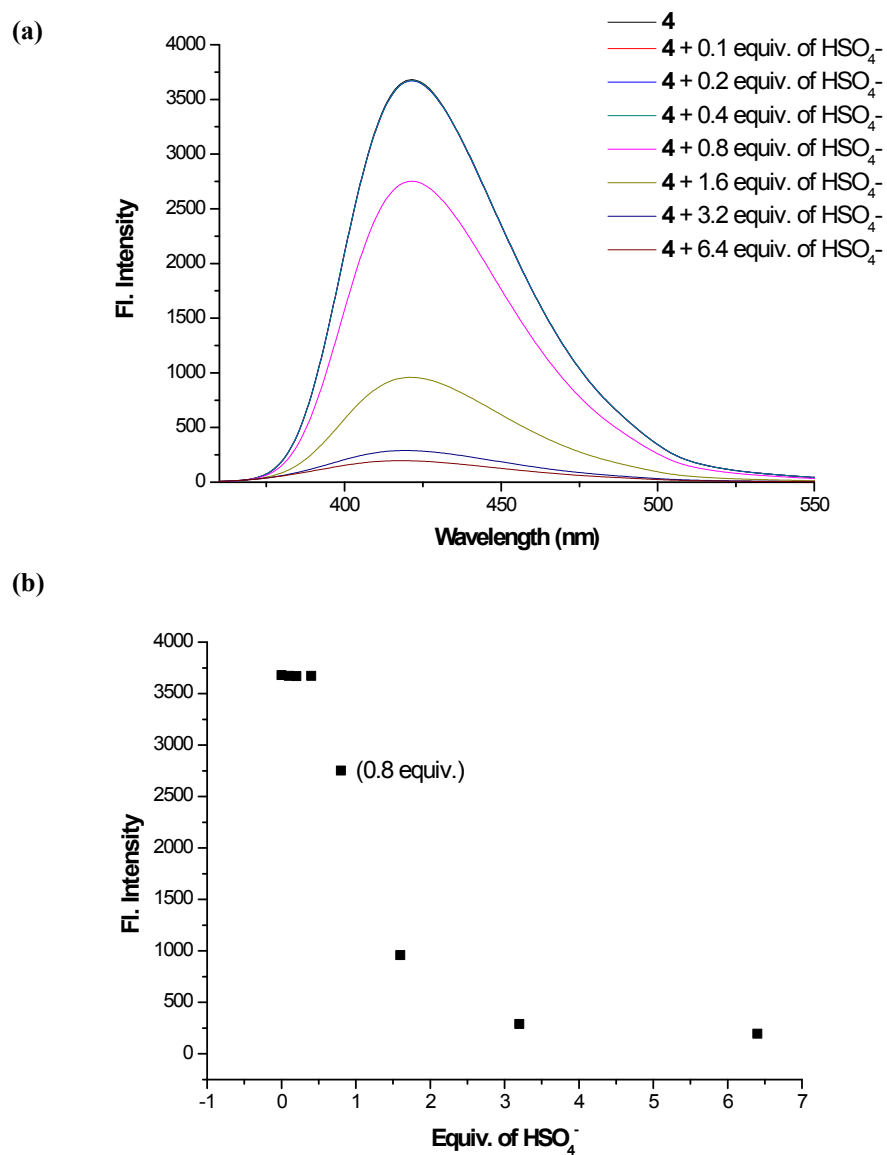


Figure S10. (a) Evolution of the fluorescent spectrum of **4** (2.0×10^{-5} M in CH_3CN containing 1% water and 32 equiv. of Na^+) during the titration with tetrabutylammonium HSO_4^- ($\text{TBA} \cdot \text{HSO}_4$; 0 – 6.4 equiv.) (b) The fluorescent intensities of **4** at 422 nm vs equiv. of HSO_4^- obtained from Figure S10a.

3. DFT calculation

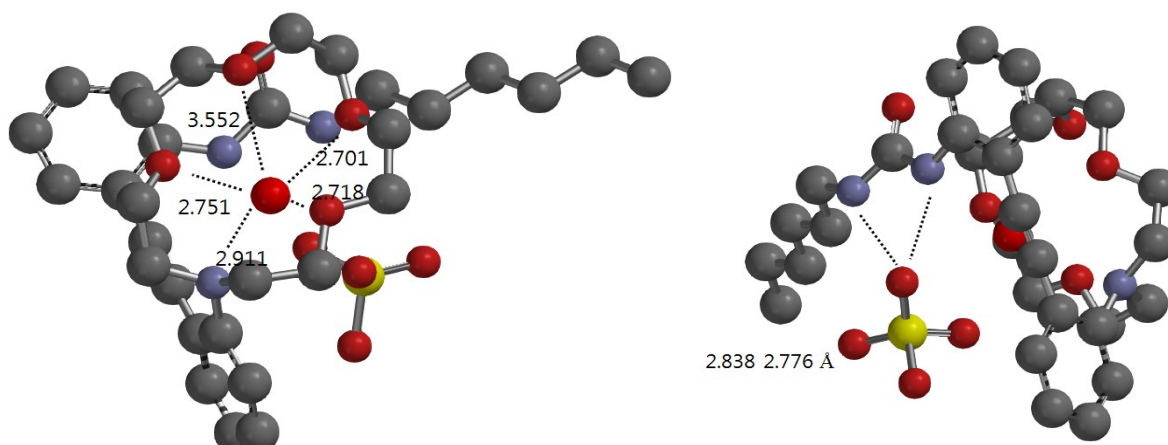


Figure S11. Optimized structures of 4·KHSO₄ based on DFT methods at the EDF2/6–31G* level of theory. Bond distances were denoted and all of hydrogen atoms are omitted for clarity.

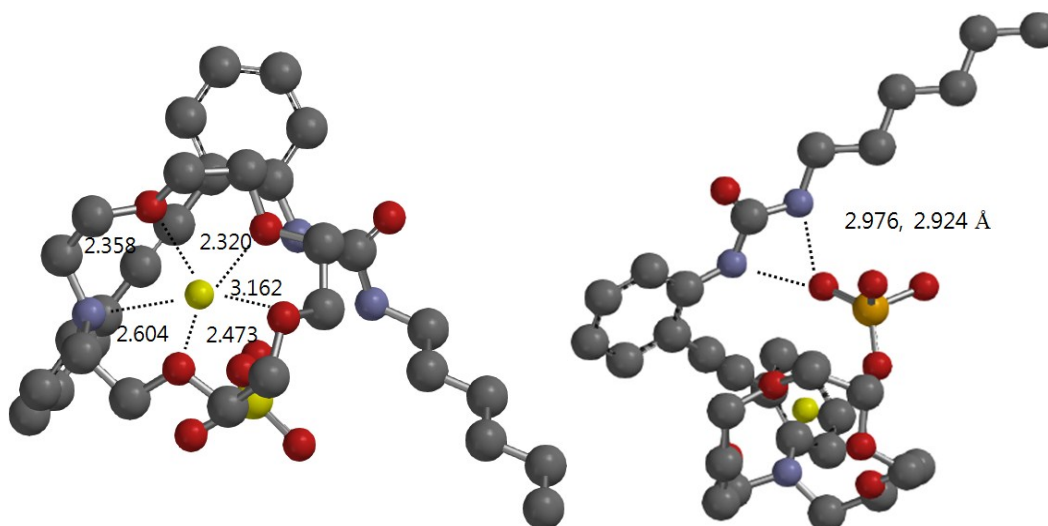


Figure S12. Optimized structures of 4·NaHSO₄ based on DFT methods at the EDF2/6–31G* level of theory. Bond distances were denoted and all of hydrogen atoms are omitted for clarity.

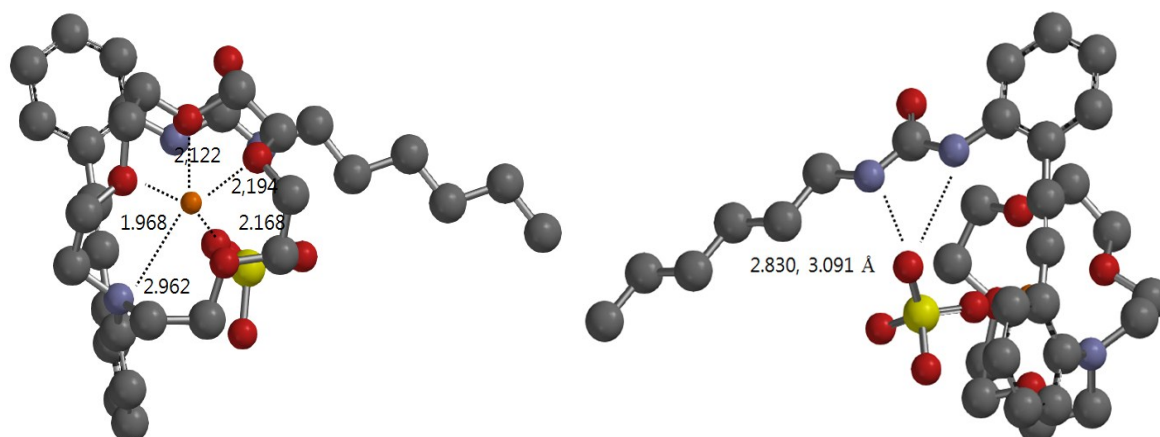


Figure S13. Optimized structures of 4·LiHSO₄ based on DFT methods at the EDF2/6–31G* level of theory. Bond distances were denoted and all of hydrogen atoms are omitted for clarity.

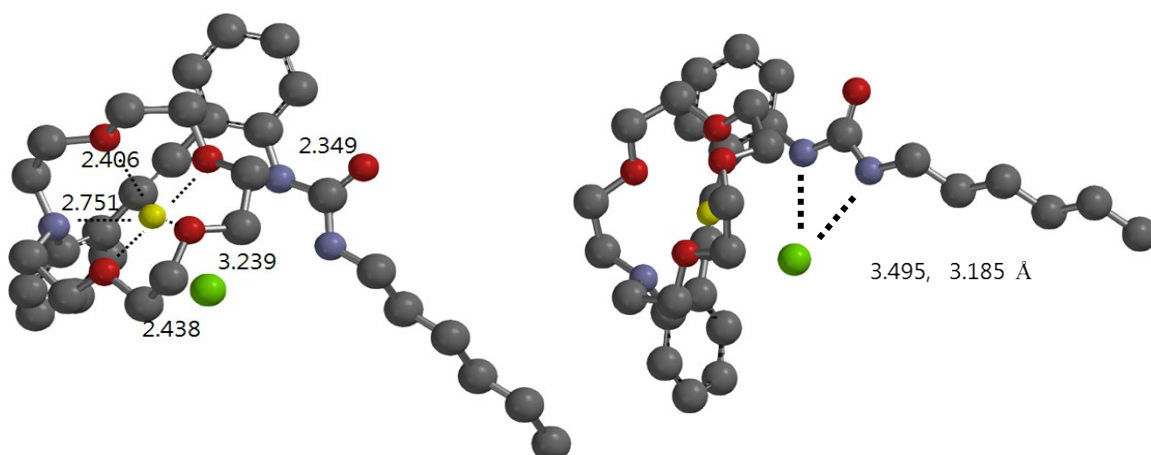


Figure S14. Optimized structures of $4 \cdot \text{NaCl}$ based on DFT methods at the EDF2/6–31G* level of theory. Bond distances were denoted and all of hydrogen atoms are omitted for clarity.

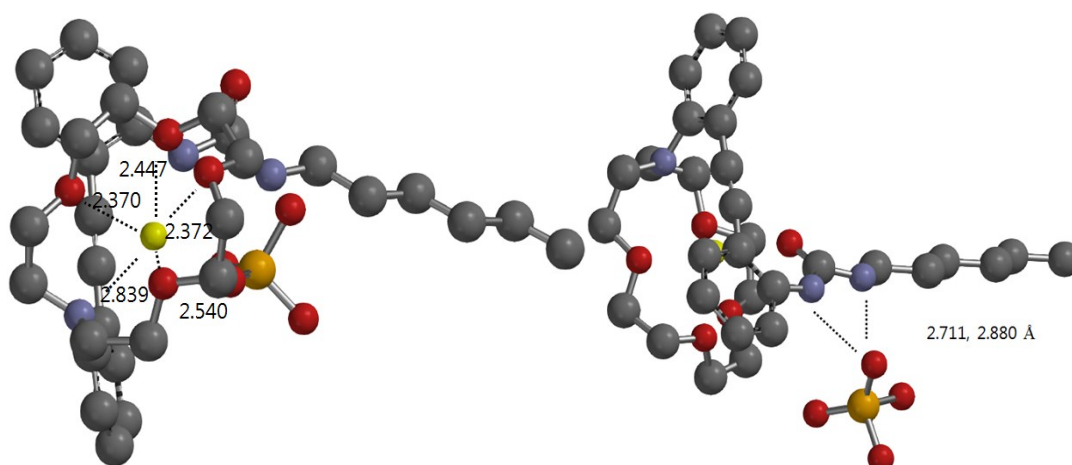


Figure S15. Optimized structures of $4 \cdot \text{H}_2\text{PO}_4$ based on DFT methods at the EDF2/6–31G* level of theory. Bond distances were denoted and all of hydrogen atoms are omitted for clarity.

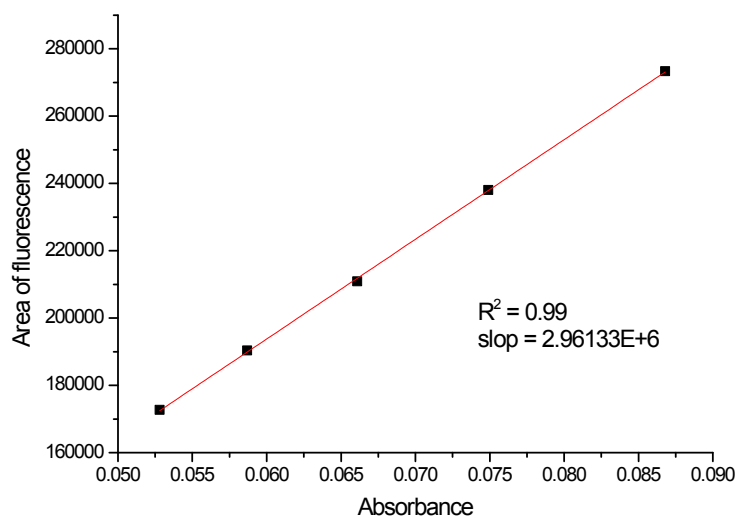
4. Quantum Yields

Table S1. Summarized slops and the relative quantum yields of **4** from those of coumarin 120.

	Slop in CH ₂ Cl ₂	Slop in CH ₃ CN	Q* in CH ₂ Cl ₂	Q* in CH ₃ CN
4	2.96133E+6	3.00872E+6	0.558 (56%)	0.392 (39%)
Coumarin 120	4.35014E+6	4.68023E+6	0.82 (82%)	0.61 (61%)

Q* denotes quantum yields.

a)



b)

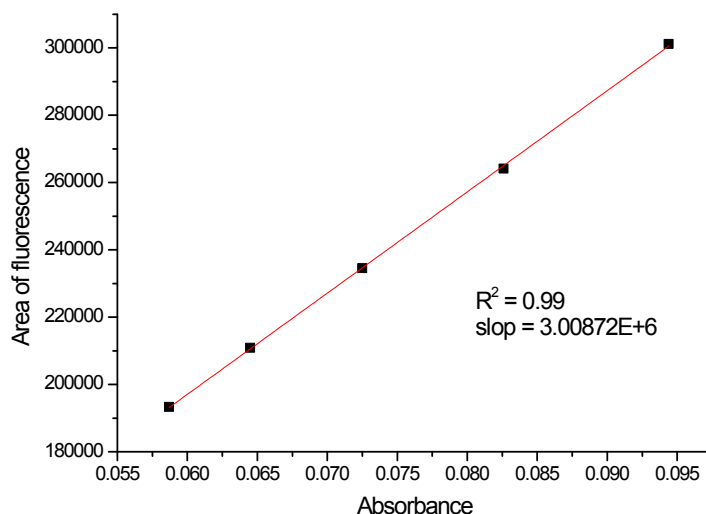
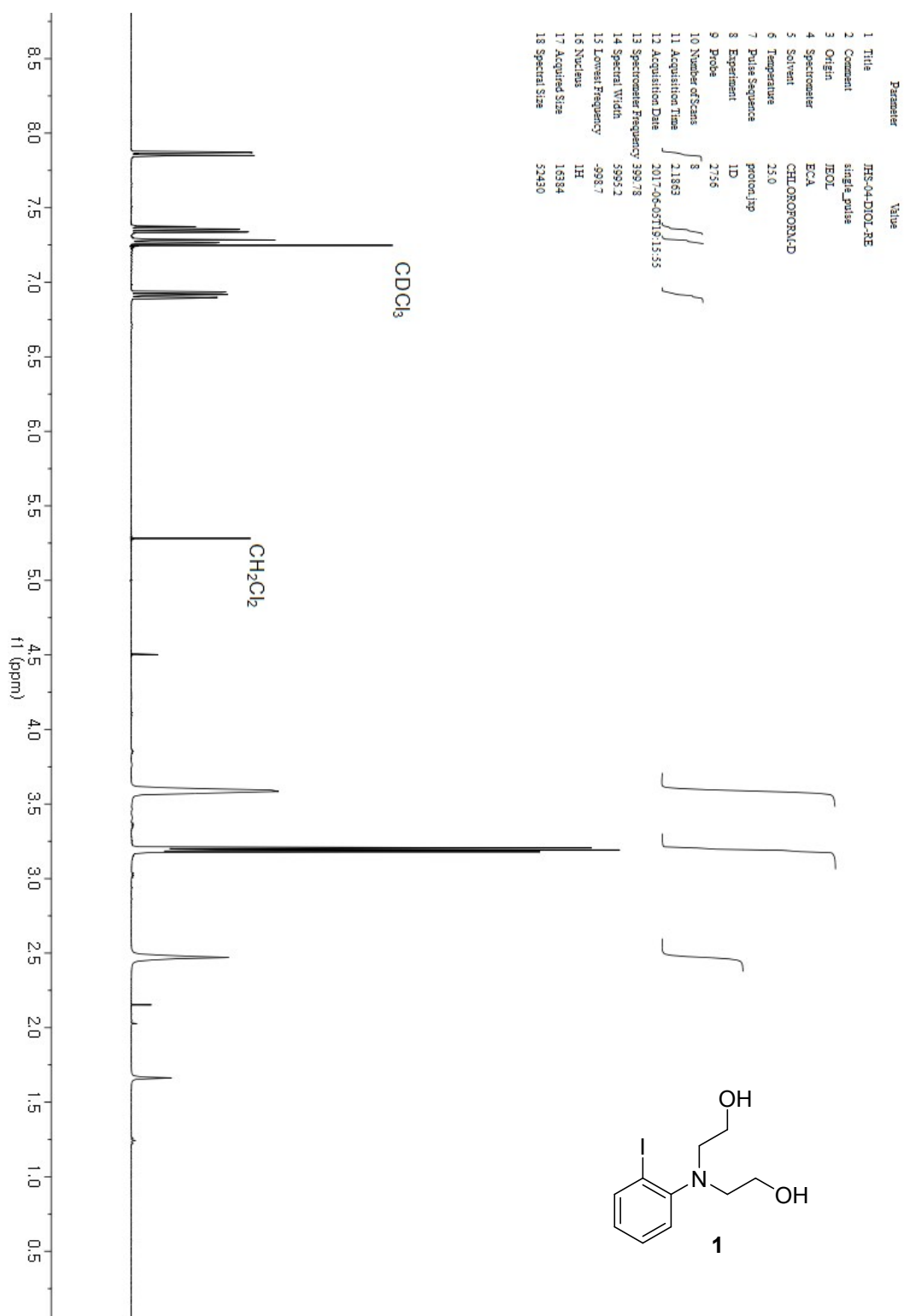


Figure S16. Plots of absorbance against the area under the fluorescence curve of **4**. a) In CH₂Cl₂. b) In CH₃CN.

5. NMR Spectra



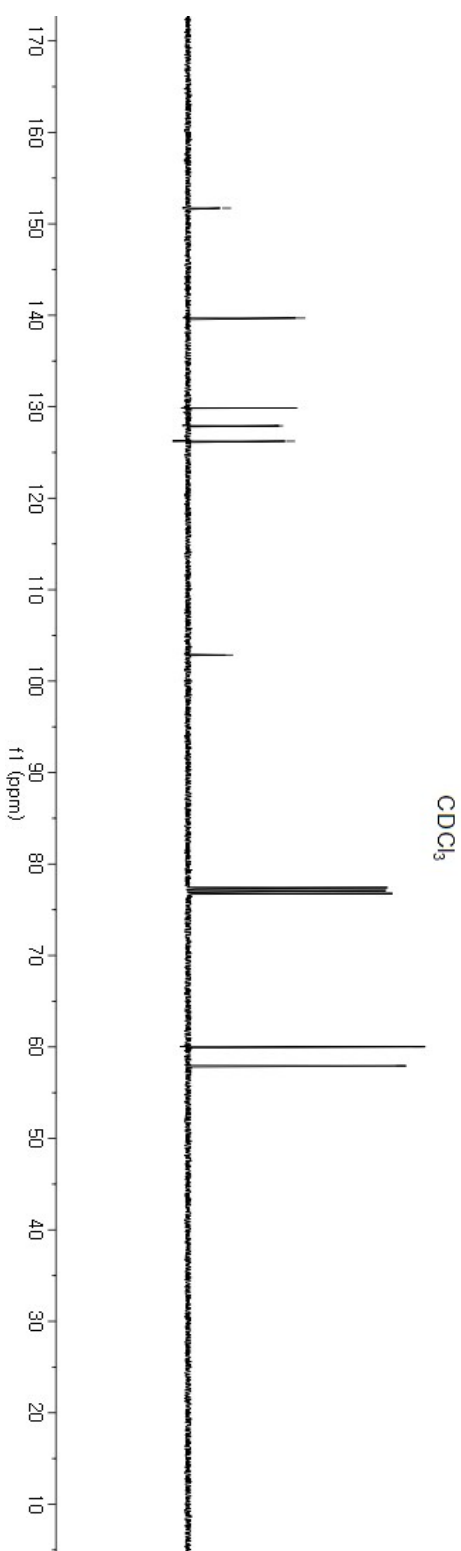
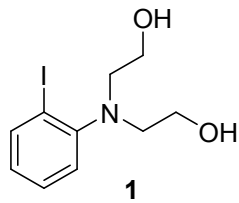
¹H NMR spectrum of **1** recorded in CDCl₃

Parameter	Value
1 Title	46
2 Comment	512.4 DIOL-RE
3 Origin	JEOL
4 Spectrometer	ECX
5 Solvent	CHLOROFORM-D
6 Temperature	25.0
7 Pulse Sequence	carbon,pp
8 Experiment	ID
9 Probe	2156
10 Number of Scans	376
11 Acquisition Time	1.0381
12 Acquisition Date	2017-06-05T19:18:37
13 Spectrometer Frequency	100.53
14 Spectral Width	25232.5
15 Lower Frequency	-2573.7
16 Nucleus	¹³ C
17 Acquired Size	32768
18 Spectral Size	52430

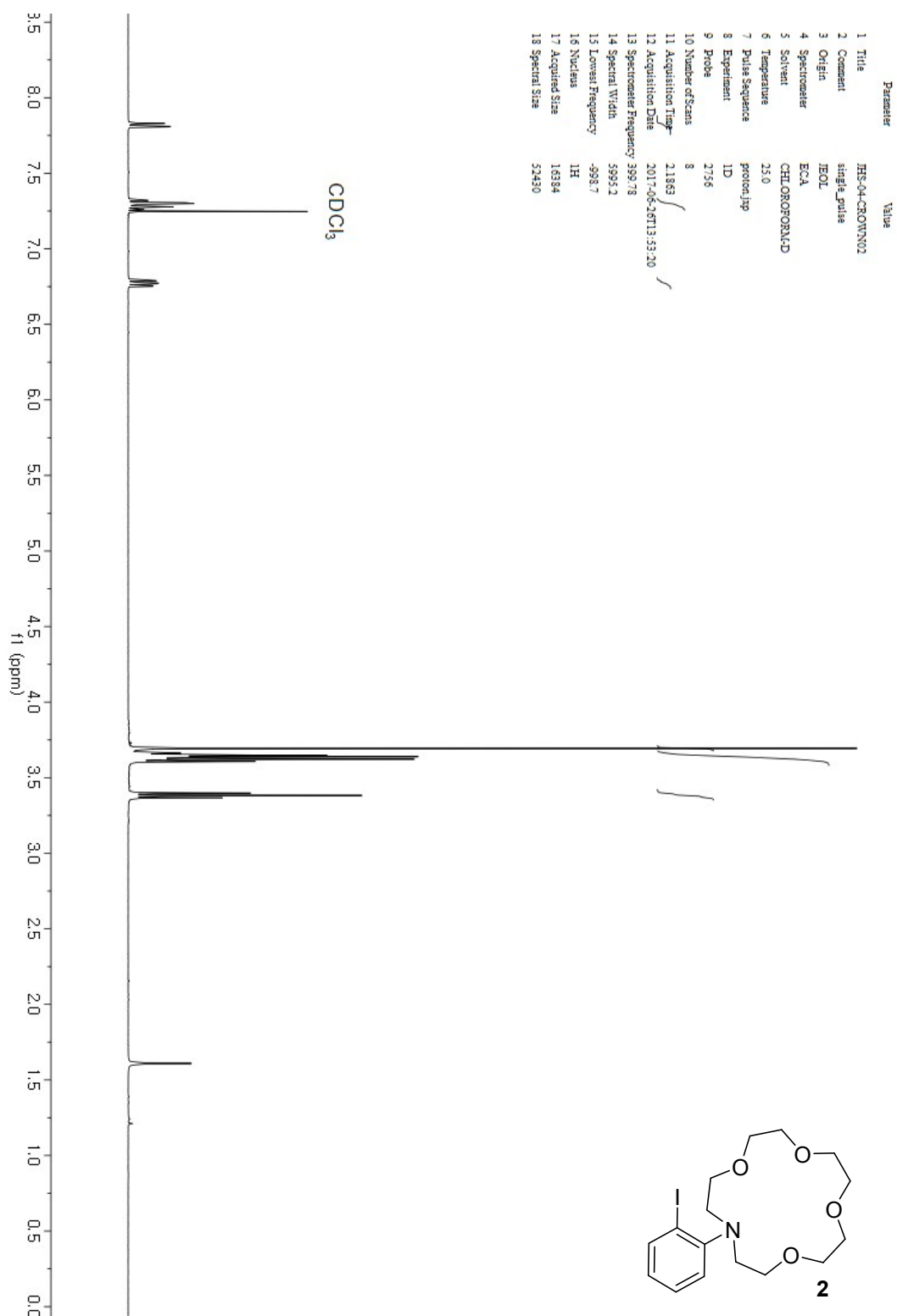
129.6906
129.8679
127.9172
126.2323

102.8595

60.0174
57.9371



¹³C NMR spectrum of **1** recorded in CDCl₃



¹H NMR spectrum of **2** recorded in CDCl₃

Parameter	Value
1 Title	HS-04-CROWN-1
2 Comment	single pulse excitation
3 Origin	IEOL
4 Spectrometer	ECA
5 Solvent	CHLOROFORM-D
6 Temperature	25.0
7 Pulse Sequence	carbon,mp
8 Experiment	ID
9 Probe	7566
10 Number of Scans	141
11 Acquisition Time	1.0381
12 Acquisition Date	2011-06-14T17:08:07
13 Spectrometer Frequency	100.53
14 Spectral Width	25252.5
15 Lowest Frequency	-25712.7
16 Nucleus	¹³ C
17 Acquired Size	37368
18 Spectral Size	52450

—139.9880

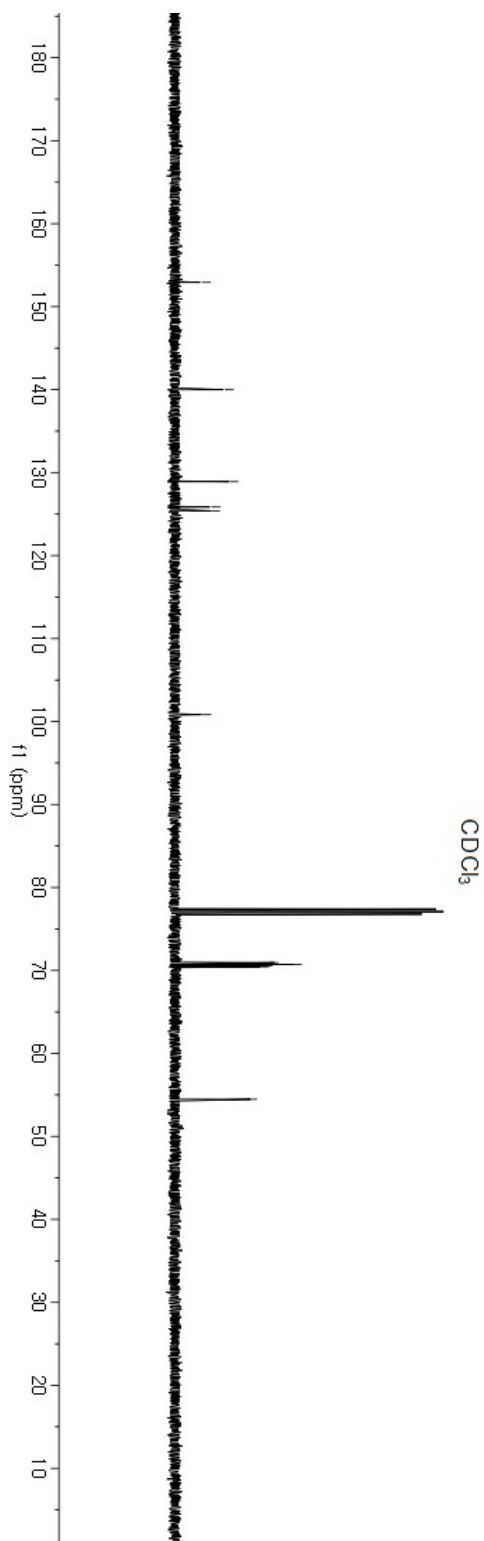
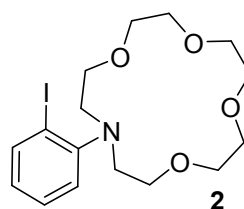
✓ 128.9060

✓ 125.8536

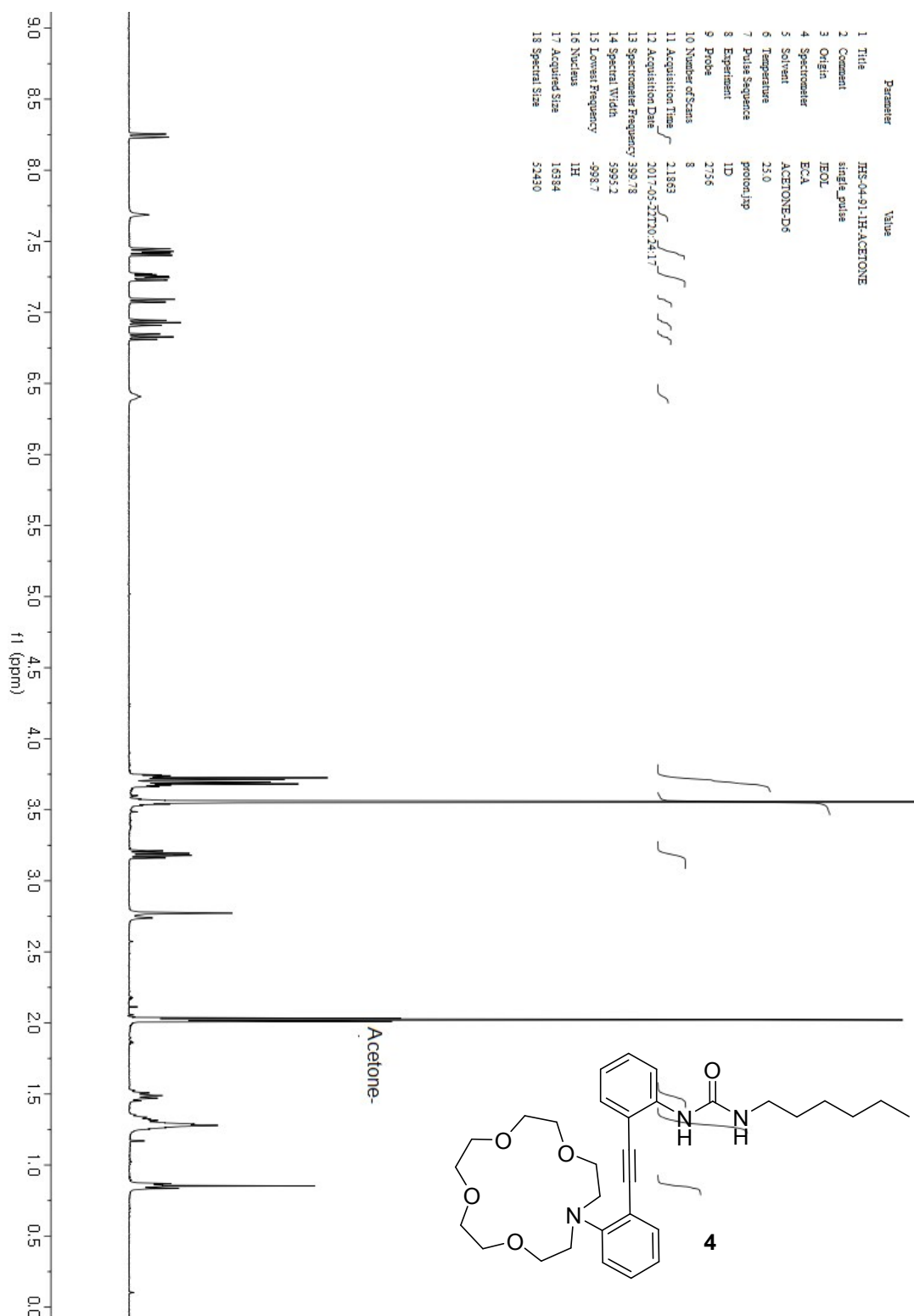
✓ 125.3996

-100.8285
$$\left\{ \begin{array}{l} 70.9514 \\ 70.7253 \\ 70.5735 \\ 70.4454 \end{array} \right.$$

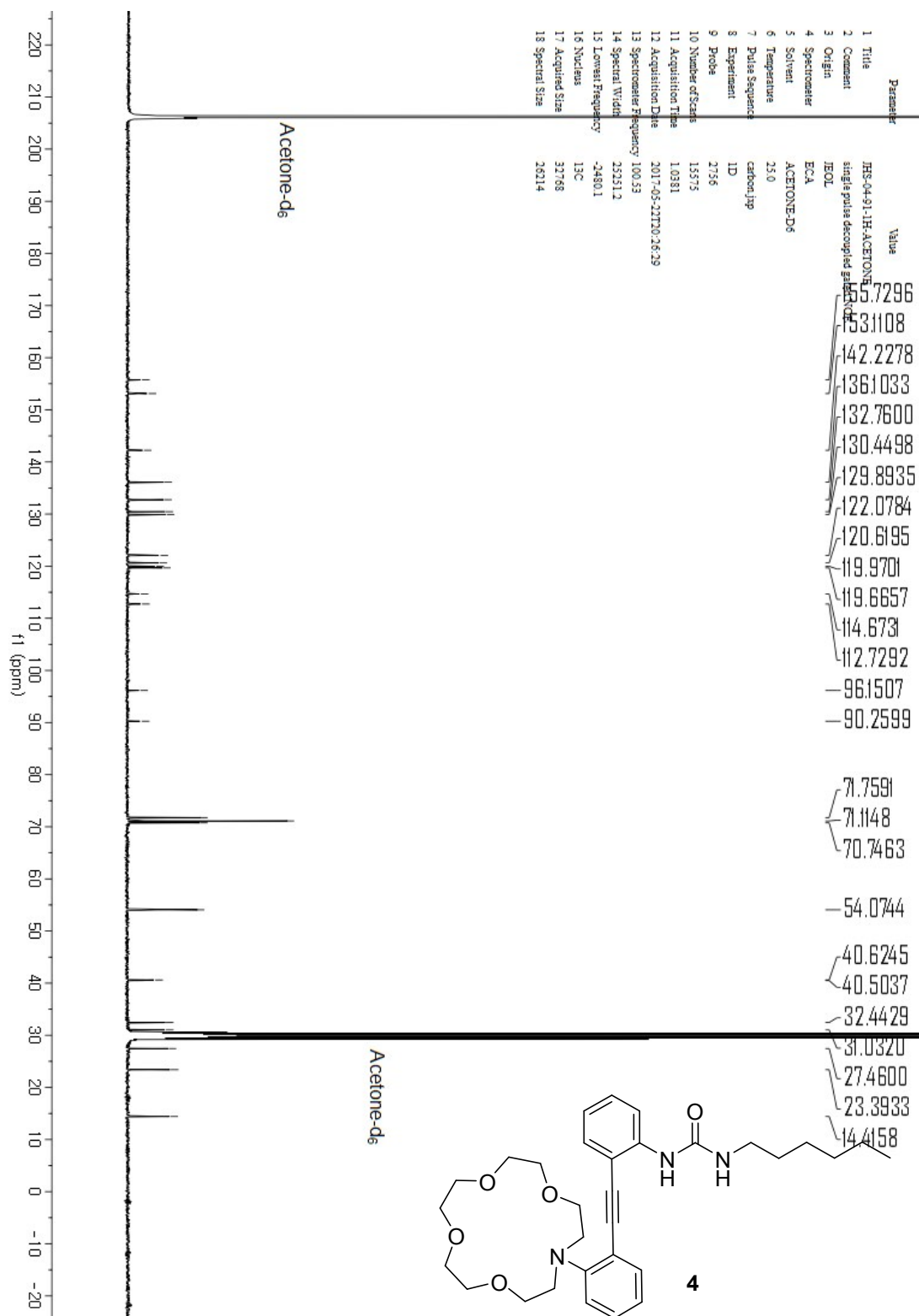
—54.4746



^{13}C NMR spectrum of **2** recorded in CDCl_3



^1H NMR spectrum of **4** recorded in Acetone- d_6



¹³C NMR spectrum of **4** recorded in Acetone-d₆