Supporting Information

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# **Supporting Information**

# A combinatorial approach towards the synthesis of non-hydrolysable triazole-iduronic acid-hybrid inhibitors of human α-L-iduronidase: Discovery of enzyme stabilizers for the potential treatment of MPSI

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#### A. General Information

All solvents and reagents were obtained commercially and used without further purification. NMR spectra were recorded on a Bruker AVANCE 600 spectrometer in deuterium solvents at ambient temperature. Chemical shifts are reported in units of parts per million (ppm,  $\delta$ ) for solutions in chloroform-d ( $\delta$  = 7.24 for <sup>1</sup>H,  $\delta$  = 77.0 for <sup>13</sup>C) and coupling constants (J) are given in Hz. The splitting patterns are reported as s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), and dd (doublet of doublets). High resolution mass spectra were obtained on a Bruker Daltonics BioTOF III spectrometer (ESI-MS). Optical rotations were measured with a Perkin-Elmer Model 341 polarimeter. Flash column chromatography was carried out using Merck Kieselgel Si60 (40-63 µm). Reactions were monitored by analytical thin-layer chromatography (TLC) in silica gel 60 F254 plates and visualized by exposure to ultraviolet light at 254 nm and/or immersion in a staining solution (p-anisaldehyde or phosphomolybdic acid) followed by heating on a hot plate. Concentration refers to rotary evaporation. Parallel synthesis was performed on Buchi SynCore 96-well synthesizer and the reaction vessels ( $16 \times 150$  mm). Multiple-functional liquid handler (Freedom EVO, TECAN) was utilized for extraction and separation. Solvent evaporation was performed on Thermo Scientific Savant Explorer SpeedVac Concentrator Explorer-220. α-L-iduronidase (glycosaminoglycan a-L-iduronohydrolase, EC 3.2.1.76) is a lysosomal hydrolase.<sup>S1</sup> The substrate 4-methylumbelliferyl  $\alpha$ -L-iduronide (4-MUI) used for activity assays was prepared from the reported papers with slight modification.<sup>S2</sup>

#### **B.** Chemistry section

#### 1. Preparation of iduronyl azides 3 and 4



**Compound 5.** D-glucuronolactone (50.0 g, 283.9 mmol) and NaOH (120 mg, 0.01 equiv) in MeOH (350 mL) was stirred at room temperature for 2 h. The reaction was neutralized with HOAc (300 µL), and then MeOH was removed *in vacuo*. The residue was added pyridine (370 mL, 16 equiv) and Ac<sub>2</sub>O (214 mL, 8 equiv) at 0 °C and stirred overnight. After evaporation *in vacuo*, the residue was purified by flash column chromatography [silica gel, hexanes/ethyl acetate 1/1 (v/v)] to give **5** (98.2 g, 92%). The beta form can be obtained from recrystallization in hot ethanol to get (45.1 g). Beta form: <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  5.69 (d, 1H, *J* = 7.8 Hz), 5.23 (dd, 1H, *J* = 9.0, 9.3 Hz), 5.14 (dd, 1H, *J* = 9.3, 9.5 Hz), 5.05 (dd, 1H, *J* = 7.8, 9.0 Hz), 4.10 (d, 1H, *J* = 9.5 Hz), 3.65 (s, 3H), 2.02 (s, 3H), 1.94 (s, 6H), 1.93 (s, 3H); <sup>13</sup>C NMR (150 MHz,CDCl<sub>3</sub>)  $\delta$  169.8, 169.3, 169.1, 168.7, 166.7, 91.3, 72.9, 71.7, 70.0, 68.8, 52.9, 20.7, 20.5, 20.4 × 2. HRMS calcd for [C<sub>15</sub>H<sub>20</sub>O<sub>11</sub>+Na]<sup>+</sup> 399.0910, found 399.0898.



**Compound 6.** A mixture of **5** (10.0 g, 26.6 mmol) and NBS (18.9 g, 4 equiv) in dry CCl<sub>4</sub> (90 mL) was stirred for 1.5 under irradiation with two 250W tungsten light blubs. The reaction was quenched with 10% aqueous Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub>. The organic layer was washed with 10% aqueous Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub>, brine, dried over anhydrous MgSO<sub>4</sub>, and concentrated. The residue was purified by flash column chromatography [silica gel, hexanes/ethyl acetate 3/1 (v/v)] to afford the title product **6** as a yellow solid (8.5 g, 70%). Beta form: <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  6.23 (d, 1H, *J* = 8.7 Hz), 5.48 (t, 1H, *J* = 9.5 Hz), 5.27 (d, 1H, *J* = 9.5 Hz), 5.19 (dd, 1H, *J* = 8.7, 9.5 Hz), 3.78 (s, 3H), 2.09 (s, 3H), 2.04 (s, 3H), 2.00 (s, 3H), 1.97 (s, 3H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  169.5, 169.0, 168.9, 168.2, 164.2, 90.8, 88.9, 70.7, 69.8, 69.1, 54.1, 20.6, 20.5, 20.4 × 2. HRMS calcd for [C<sub>15</sub>H<sub>19</sub>BrO<sub>11</sub>+Na]<sup>+</sup> 477.0001, found 477.0003.



**Compound 7.** A mixture of **6** (8.7 g, 19.2 mmol), tributyltin hydride (44 mL, 2.5 equiv) and AIBN (723 mg, 0.23 equiv) in dry THF (64 mL) was refluxed for 45 min. After cooling and evaporation *in vacuo*, and the residue was treated with EtOAc and aqueous KF for 20 min. The mixture was filtered through Celite, the filtrate was concentrated, and purified by flash column chromatography [silica gel, toluene/ethyl acetate 2/1 (v/v)] to get 7 (2.2 g, 30%) and **5** (4.3 g, 60%). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  6.26 (s, 1H), 5.14 (t, 1H, *J* = 3 Hz), 5.13 (d, 1H, *J* = 3.6 Hz), 4.84 (d, 1H, *J* = 2.4 Hz), 4.83 (s, 1H, *J* = 2.4 Hz), 3.76 (s, 3H), 2.10 (s, 3H), 2.08 (s, 3H), 2.07 (s, 3H), 2.05 (s, 3H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  169.3, 169.0, 168.5, 167.9, 167.6, 90.4, 68.2, 66.7, 66.2, 65.8, 52.7, 20.7, 20.6 × 2, 20.5.HRMS calcd for [C<sub>15</sub>H<sub>20</sub>O<sub>11</sub>+Na]<sup>+</sup> 399.0801, found 399.0898.



**Compound 3.** A mixture of 7 (300 mg, 0.9 mmol) in  $CH_2Cl_2$  (9 mL) was added tin(IV) chloride (50  $\mu$ L, 0.4 equiv) and trimethylsilyl azide (28  $\mu$ L, 2.3 equiv) at 0 °C for 4 h. The reaction was quenched with saturated aqueous NaHCO<sub>3</sub> and stirred for 30 min. The mixture was filtered through Celite, the filtrate was diluted with EtOAc (60 mL) and washed with water (3 × 20 mL). The separated organic layer was dried over anhydrous MgSO<sub>4</sub>, concentrated, and purified by column chromatography [silica gel,

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hexanes/ethyl acetate 2/1 (v/v)] to afford the title product **3** (200 mg, 75%). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  5.53 (d, 1H, *J* = 3.5 Hz, H-1), 5.14 (t, 1H, *J* = 4.5 Hz, H-3), 5.11 (dd, 1H, *J* = 4.8, 3.5 Hz, H-4), 4.90 (d, 1H, *J* = 3.4 Hz, H-5), 4.73 (t, 1H, *J* = 4.3 Hz, H-2), 3.78 (s, 3H, CO<sub>2</sub><u>CH<sub>3</sub></u>), 2.09 (s, 3H, CO<u>CH<sub>3</sub></u>), 2.07 (s, 3H, CO<u>CH<sub>3</sub></u>), 2.05 (s, 3H, CO<u>CH<sub>3</sub></u>); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  169.3, 169.2, 168.9, 167.9, 86.5, 68.5, 67.0, 66.6, 52.7, 20.7, 20.6, 20.5. HRMS calcd for [C<sub>13</sub>H<sub>17</sub>N<sub>3</sub>O<sub>9</sub>+Na]<sup>+</sup> 382.0857, found 382.0872.



**Compound 8.** A mixture of **6** (1.5 g, 3.3 mmol) and silver(I) fluoride (800 mg, 2 equiv) in acetonitrile (16 mL) was stirred at room temperature in the dark (the flask was coated by a layer of foil) for 8 h. The reaction mixture was concentrated, and purified by column chromatography [silica gel, hexanes/ethyl acetate 4/1 (v/v)] to afford the title product **8** as a yellow gel (900 mg, 75%). Alpha form: <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  6.16 (dd, 1H, J = 1.5, 2.5 Hz), 5.63 (dd, 1H, J = 12.0, 9.2 Hz), 5.42 (dd, 1H, J = 6.2, 9.2Hz), 5.16(dd, 1H, J = 2.8, 6.2Hz); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  169.5, 168.5, 167.9, 163.6, 163.4, 106.3, 92.0, 72.3, 69.6, 69.5, 53.5, 20.8, 20.6, 20.5, 20.4. HRMS calcd for [C<sub>15</sub>H<sub>19</sub>FO<sub>11</sub>+Na]<sup>+</sup> 417.0805, found 417.0804.

$$\begin{array}{c} \begin{array}{c} OAc \\ MeO_2C \\ \hline \\ OAc \\ 7 \end{array} \begin{array}{c} OAc \\ \hline \\ 7 \end{array} \begin{array}{c} 1.33\% \ HBr_{(HOAc)}, \\ DCM, \ 0 \ ^{\circ}C \ to \ rt, \ 6 \ h \\ \hline \\ 2. \ NaN_3, \ TBAHS, \\ NaHCO_3, \ DCM, \ H_2O, \\ \hline \\ 1.33\% \ HBr_{(HOAc)}, \\ OAc \\ \hline \\ OAc \\ \hline \\ 0Ac \\ OAc \\ \hline \\ 0Ac \\ \hline \\ 0Ac \\ OAc \\ \hline \\ 0Ac \\ \hline 0Ac \\ \hline \\ 0Ac \\ \hline \hline 0Ac \\ \hline \\ 0Ac \\ \hline \hline 0Ac \\ \hline \\ 0Ac \\ \hline \hline 0Ac \\ \hline \hline 0Ac \\ \hline 0Ac \\ \hline \hline 0A$$

**Compound 4.** A mixture of 7 (300 mg, 0.9 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (2 mL) was added acetic anhydride (300  $\mu$ L, 0.3 equiv) and 30% HBr<sub>(HOAc)</sub> (1.2 mL, 4.8 mmol) at 0 °C for 6 h. The reaction was warmed up to room temperature. The mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub> (40 mL) and washed with saturated aqueous NaHCO<sub>3</sub> (2 × 20 mL). The organic layer was dried over anhydrous MgSO<sub>4</sub> and concentrated to get the crude iduronyl bromide as a yellow gel. A mixture of the crude iduronyl bromide (0.9 mmol), NaN<sub>3</sub> (100 mg, 1.9 equiv), and TBAHS (40 mg, 0.1 equiv) in dry CH<sub>2</sub>Cl<sub>2</sub> (4 mL) and NaHCO<sub>3</sub> (4 mL, 1M aqueous solution) was stirred vigorously at room temperature for 2 h. The mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub> (60 mL), and washed with water (3 × 20 mL). The organic layer was dried over anhydrous MgSO<sub>4</sub>, concentrated, and purified by column chromatography [silica gel, hexanes/ethyl acetate 2/1 (v/v)] to afford 4 (140 mg, 45%). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  5.18 (t, 1H, *J* = 3.1 Hz, H-3), 5.08 (brs, 1H, H-4), 5.00 (d, 1H, *J* = 1.5 Hz, H-1), 4.90 (brs, 1H, H-2), 4.60 (d, 1H, *J* = 2.0 Hz, H-5), 3.77 (s, 3H, CO<sub>2</sub>CH<sub>3</sub>), 2.12 (s, 6H, CO<u>CH<sub>3</sub></u>), 2.07 (s, 3H, CO<u>CH<sub>3</sub></u>); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  169.2, 169.1, 167.8, 166.8, 85.0, 73.7, 66.5, 65.9, 65.8, 52.7, 20.7, 20.6, 20.5. HRMS calcd for [C<sub>13</sub>H<sub>17</sub>N<sub>3</sub>O<sub>9</sub>+Na]<sup>+</sup> 382.0857, found 382.0872.

#### 2. Preparation of iduronyl triazoles

## General procedure 1 for CuAAC click reaction

A mixture of an azide (1 mmol), alkyne (2 equiv), CuSO<sub>4</sub>-5H<sub>2</sub>O (1M aqueous solution, 0.01 equiv, 10  $\mu$ L), and sodium ascorbate (1M aqueous solution, 0.1 equiv, 100  $\mu$ L) in DMF (400  $\mu$ L) was irradiated in a microwave reactor reaction until the reaction was complete (TLC). The reaction was cooled to room temperature, then diluted with EtOAc (60 mL), and washed with water (3 × 20 mL). The organic layer was dried over anhydrous MgSO<sub>4</sub>, concentrated, and purified by column chromatography [silica gel, hexanes/ethyl acetate 2/1 (v/v)] to afford iduronyl triazole.

#### General procedure 2 for CuAAC click reaction

A mixture of an azide (1 mmol), alkyne (2 equiv), CuSO<sub>4</sub>-5H<sub>2</sub>O (1M aqueous solution, 0.3 equiv, 300  $\mu$ L), and sodium ascorbate (1M aqueous solution, 0.3 equiv, 300  $\mu$ L) in *t*-BuOH/H<sub>2</sub>O (400  $\mu$ L) was stirred at room temperature for 48 h. Workup of the reaction mixture was carried out in the automated liquid-liquid extraction module. The mixture was diluted with H<sub>2</sub>O (60 mL) and washed with EtOAc (3 × 20 mL). The aqueous layer was concentrated and purified by column chromatography [silica gel, ethyl acetate/methanol/water 7/2/1 (v/v)].

#### General procedure 3 for deprotection of esters

A mixture of an ester (1 mmol) in THF (5 mL) was added LiOH (2M aqeous solution, 500  $\mu$ L, 1 equiv) at 0 °C and was stirred for 2 h. The reaction was adjusted pH value to approximately 7 by adding DOWEX 50WX8-200 ion-exchange resin. The mixture was filtered, the filtrate was concentrated and purified by column chromatography [silica gel, ethyl acetate/methanol/water 7/2/1 (v/v)].



**Compound 10.** The preparation was accorded on General procedures 1 and 3. Compound **3** and phenylacetylene were used. Yield was 58%. <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.57 (s, 1H), 7.82–7.83 (m, 2H), 7.47–7.54 (m, 3H), 6.12 (d, 1H, *J* = 9.0 Hz), 4.61 (d, 1H, *J* = 6.0 Hz), 4.12 (dd, 1H, *J* = 9.0, 9.2 Hz), 4.00 (dd, 1H, *J* = 6.0, 9.3 Hz), 3.78 (dd, 1H, *J* = 9.2, 9.3 Hz); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.3, 147.6, 129.1 × 3, 128.9, 125.7 × 2, 121.5, 85.4, 74.3, 73.8, 72.1, 70.5. HRMS calcd for [C<sub>14</sub>H<sub>15</sub>N<sub>3</sub>O<sub>6</sub>-H]<sup>-</sup> 320.0891, found 320.0877.



**Compound 11.** The preparation was accorded on General procedures 1 and 3. Compound 4 and phenylacetylene were used. Yield was 53%. <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.78 (s, 1H), 7.88–7.89 (m, 2H), 7.54–7.57 (m, 2H), 7.47–7.49 (m, 1H), 6.29 (s, 1H), 4.66 (s, 1H), 4.31 (dd, 1H, *J* = 2.7, 3.1 Hz), 4.15 (d, 1H, *J* = 1.3 Hz), 4.07 (d, 1H, *J* = 1.3 Hz); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.1, 147.0, 129.5, 129.2 × 2, 128.8, 125.7 × 2, 121.8, 85.3, 77.2, 69.1, 69.0, 68.9. HRMS calcd for [C<sub>14</sub>H<sub>15</sub>N<sub>3</sub>O<sub>6</sub>+H]<sup>+</sup> 322.1044, found 322.1034.



**Compound 12.** The preparation was accorded on General procedure 3. Yield was 83%. <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  5.13 (d, 1H, *J* = 6.3 Hz), 4.48 (d, 1H, *J* = 4,7 Hz), 3.83 (dd, 1H, *J* = 7.3, 4.7 Hz), 3.67 (dd, 1H, *J* = 7.5, 14.0 Hz), 3.39 (dd, 1H, *J* = 6.3, 14.0 Hz); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.6, 88.6, 72.2, 72.2, 71.0, 70.3. HRMS calcd for [C<sub>6</sub>H<sub>9</sub>N<sub>3</sub>O<sub>6</sub>-H]<sup>-</sup> 218.0422, found 218.0408.



**Compound S1.** A mixture of **5** (3.1 g, 8.35 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (80 mL) was added tin(IV) chloride (500  $\mu$ L, 0.5 equiv) and trimethylsilyl azide (500  $\mu$ L, 2.4 equiv) at 0 °C for 4 h. The reaction was quenched with saturated aqueous NaHCO<sub>3</sub> and stirred for 30 min. The mixture was filtered through Celite, the filtrate was diluted with EtOAc (150 mL) and washed with water (3 × 20 mL). The separated organic layer was dried over anhydrous MgSO<sub>4</sub>, concentrated, and purified by column chromatography [silica gel, hexanes/ethyl acetate 2/1 (v/v)] to afford the title product **S1** (2.5 g, 85%). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  5.23 (t, 1H, *J* = 9.4 Hz, H-3), 5.19 (t, 1H, *J* = 9.5 Hz, H-4), 4.92 (t, 1H, *J* = 9.0 Hz, H-2), 4.68 (d, 1H, *J* = 8.7 Hz, H-1), 4.08 (d, 1H, *J* = 9.6 Hz, H-5), 3.73 (s, 1H, CO<sub>2</sub>CH<sub>3</sub>), 2.03 (s, 3H, CO<u>CH<sub>3</sub></u>), 1.99 (s, 3H, <u>CH<sub>3</sub></u>), 1.98 (s, 3H, <u>CH<sub>3</sub></u>); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  169.9, 169.2, 169.0, 166.4, 88.0, 74.1, 71.7, 70.3, 68.9, 53.0, 20.4 × 2, 20.3. HRMS calcd for [C<sub>13</sub>H<sub>17</sub>N<sub>3</sub>+Na]<sup>+</sup> 382.0865, found 382.0857.

**Compound 13.** The preparation was accorded on General procedure 3. Yield was 80%. <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  3.82 (d, 1H, J = 9.1 Hz), 3.52–3.57 (m, 2H), 3.32 (t, 1H, J = 8.7 Hz); <sup>13</sup>C NMR (150 MHz,

 $D_2O$ )  $\delta$  175.3, 89.9, 77.6, 75.5, 72.6, 71.4. HRMS calcd for  $[C_6H_9N_3O_6-H]^-$  218.0422, found 218.0408.



**Compound 14a.** The preparation was accorded on General procedure 2. 2,2-Dimethyl-1-butyne was used. Yield was 63%. <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.08 (s, 1H), 6.02 (d, 1H, *J* = 9.1 Hz), 4.56 (d, 1H, *J* = 6.0 Hz), 4.04 (dd, 1H, *J* = 9.1, 9.3 Hz), 3.95 (dd, 1H, *J* = 6.0, 9.4 Hz), 3.74 (dd, 1H, *J* = 9.3, 9.4 Hz); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.3, 158.2, 120.5 , 85.1, 74.2, 73.8, 72.1, 70.4, 30.1, 29.3 × 3. HRMS calcd for [C<sub>12</sub>H<sub>19</sub>N<sub>3</sub>O<sub>6</sub>-H]<sup>-</sup> 300.1204, found 300.1190.



**Compound 14b.** The preparation was accorded on General procedure 2. 1-Ethynyl-4-fluorobenzene was used. Yield was 63%. <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.52 (s, 1H), 7.77–7.79 (m, 2H), 7.21–7.24 (m, 2H), 6.11 (d, 1H, *J* = 9.0 Hz), 4.61 (d, 1H, *J* = 6.0 Hz), 4.11 (dd, 1H, *J* = 9.0, 9.2 Hz), 3.99 (dd, 1H, *J* = 6.0, 9.3 Hz), 3.78 (dd, 1H, *J* = 9.2, 9.3 Hz); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.3, 162.7, 146.9, 127.7 × 2, 125.4, 121.3, 115.9 × 2, 85.4, 74.3, 73.8, 72.1, 70.5. HRMS calcd for [C<sub>14</sub> H<sub>14</sub>FN<sub>3</sub>O<sub>6</sub>-H]<sup>-</sup> 338.0795, found 338.0783



**Compound 14c.** The preparation was accorded on General procedure 2. Propargyl alcohol was used. Yield was 51%. <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.26 (s, 1H), 6.06 (d, 1H, *J* = 9.2 Hz), 4.59 (d, 1H, *J* = 6.2 Hz), 4.08 (t, 1H, *J* = 9.2 Hz), 3.97 (dd, 1H, *J* = 6.2, 9.2 Hz), 3.75 (t, 1H, *J* = 9.2 Hz); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.2, 147.0, 123.6, 85.3, 74.2, 73.8, 72.1, 70.4. 54.5. HRMS calcd for [C<sub>9</sub>H<sub>13</sub>N<sub>3</sub>O<sub>7</sub>-H]<sup>-</sup> 274.0745, found 274.0670.



**Compound 14d.** The preparation was accorded on General procedure 2. 1-Nonyne was used. Yield was 60%. <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.02 (s, 1H), 6.01 (d, 1H, *J* = 9.1 Hz), 4.54 (d, 1H, *J* = 6.0 Hz), 4.05 (dd, 1H, *J* = 9.1, 9.3 Hz), 3.95 (dd, 1H, *J* = 6.0, 9.4 Hz), 3.74 (dd, 1H, *J* = 9.3, 9.4 Hz), 2.74–2.76 (m, 2H), 1.67–1.69 (m, 2H), 1.28–1.33 (m, 8H), 0.86–0.88 (m, 3H); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.3, 149.0, 122.5, 85.1, 74.2, 73.8, 72.1, 70.4, 30.9, 28.3, 28.0, 27.9, 24.3, 21.9, 13.3. HRMS calcd for [C<sub>15</sub>H<sub>25</sub>N<sub>3</sub>O<sub>6</sub>-H]<sup>-</sup> 342.1680, found 342.1660.



**Compound 15a.** The preparation was accorded on General procedure 2. 2,2-Dimethyl-1-butyne was used. Yield was 56%. <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.09 (s, 1H), 5.74 (d, 1H, *J* = 9.2 Hz), 4.01–4.04 (m, 2H), 3.70–3.77 (m, 2H), 1.35 (s, 1H); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  174.9, 158.3, 120.2, , 87.1, 78.6, 75.7, 72.1, 71.4, 30.1, 29.2 × 3. HRMS calcd for [C<sub>12</sub>H<sub>19</sub>N<sub>3</sub>O<sub>6</sub>-H]<sup>-</sup> 300.1202, found 300.1190.



**Compound 15b.** The preparation was accorded on General procedure 2. 1-Ethynyl-4-fluorobenzene was used. Yield was 61%. <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.46 (s, 1H), 7.69–7.71 (m, 2H), 7.17–7.20 (m, 2H), 5.78 (d, 1H, J = 9.2 Hz), 4.05–4.09 (m, 2H), 3.74–3.80 (m, 2H); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  174.9, 162.5, 146.9, 127.6, 125.3 × 2, 120.9, 115.9 × 2, 87.2, 78.8, 75.7, 72.2, 71.4. HRMS calcd for [C<sub>14</sub>H<sub>14</sub>FN<sub>3</sub>O<sub>6</sub>-H]<sup>-</sup> 338.0795, found 338.0783.



## 3. Parallel synthesis of an N-acyl propargylamine-based library

To a mixture of propargylamine (128  $\mu$ L, 2 mmol), a carboxylic acid (2 mmol), and EDCI (460 mg, 1.2 equiv) in dry CH<sub>2</sub>Cl<sub>2</sub> (5 mL) was added DIPEA (418  $\mu$ L, 1.2 equiv). The mixture was stirred at room temperature for 12 h. After dilution with CH<sub>2</sub>Cl<sub>2</sub> (30 mL), the mixture was washed with 1N HCl<sub>(aq)</sub> (3 × 10 mL) by assistance with a liquid handler (TECAN). The organic layer was dried over anhydrous MgSO<sub>4</sub>, concentrated to get *N*-acyl propargylamine by assistance with a concentrator (Speed-Vac).

### 4. Parallel synthesis of a 48-membered library



Supporting Information



Figure S1. Alkyne library A1–A6 (for compound L17) and acid library A6–D12 (for compound L16) for preparation of *N*-acyl propargylamine-based library

	1	2	3	4	5	6	7	8	9	10	11	12
 А	82 <sup>a</sup> (87)	<sup>b</sup> 87(96)	75(94)	75(93)	78(80)	84(81)	66(86)	89(87)	83(99)	64(85)	61(98)	62(88)
В	75(93)	84(90)	66(81)	86(97)	88(84)	64(95)	66(88)	80(88)	75(94)	68(83)	62(95)	60(96)
С	64(98)	69(86)	80(94)	85(82)	56(94)	83(90)	70(88)	65(80)	65(93)	66(94)	65(99)	64(91)
D	62(95)	83(96)	86(95)	76(88)	66(86)	80(98)	85(94)	69(83)	84(90)	56(94)	63(90)	81(98)

Table S1. Yields and purities of iduronyl triazole-based library

<sup>*a*</sup> Yield was given after solid-liquid extraction. <sup>*b*</sup> Determined by HPLC.

**Compound L17-A1.** <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.59 (s, 1H), 7.53–7.61 (m, 2H), 7.48–7.51 (m, 1H), 7.16–7.19 (m, 1H), 6.12 (d, 1H, J = 9.0 Hz), 4.61 (d, 1H, J = 6.0 Hz), 4.12 (dd, 1H, J = 9.2, 9.3 Hz), 4.00 (dd, 1H, J = 6.0, 9.2 Hz), 3.78 (dd, 1H, J = 9.0, 9.3 Hz); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.2, 162.8, 146.6, 131.2, 130.9, 122.0, 121.6, 115.5, 112.5, 85.5, 74.3, 73.8, 72.1, 70.5. HRMS calcd for [C<sub>14</sub>H<sub>14</sub>FN<sub>3</sub>O<sub>6</sub>-H]<sup>-</sup> 338.0759, found 338.0783.

**Compound L17-A2.** NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.58 (s, 1H), 7.42–7.99 (m, 2H), 7.30–7.33 (m, 1H), 7.24–7.28 (m, 1H), 6.14 (d, 1H, J = 9.0 Hz), 4.62 (d, 1H, J = 6.0 Hz), 4.15 (dd, 1H, J = 9.0, 9.3 Hz), 4.00 (dd, 1H, J = 6.0, 9.2 Hz), 3.79 (dd, 1H, J = 9.0, 9.3 Hz); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.2, 159.1, 141.4, 130.5, 127.5, 124.7, 123.9, 116.9, 115.9, 85.5, 74.3, 73.8, 72.1, 70.5. HRMS calcd for [C<sub>14</sub>H<sub>14</sub>FN<sub>3</sub>O<sub>6</sub>-H]<sup>-</sup> 338.0759, found 338.0783.

**Compound L17-A3.** <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.52 (s, 1H), 7.67–7.69 (m, 2H), 7.43–7.44 (m, 2H), 6.12 (d, 1H, J = 9.0 Hz), 4.62 (d, 1H, J = 5.9 Hz), 4.12 (t, 1H, J = 9.0 Hz), 4.00 (dd, 1H, J = 6.0, 9.0 Hz), 3.78 (dd, 1H, J = 5.0, 9.0 Hz); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.2, 146.6, 133.9, 129.0 × 2, 127.7 × 2, 127.0, 121.6, 85.5, 74.4, 73.7, 72.1, 70.5. HRMS calcd for  $[C_{14}H_{14}CIN_3O_6-H]^-$  354.0466, found 354.0487.

**Compound L17-A4.** <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.49 (s, 1H), 7.67 (s, 1H), 7.59–7.60 (m, 1H), 7.34–7.37 (m, 1H), 6.10 (d, 1H, J = 9.0 Hz), 4.60 (d, 1H, J = 6.0 Hz), 4.11 (dd, 1H, J = 9.1, 9.3 Hz), 3.98 (dd, 1H, J = 6.0, 9.1 Hz), 3.78 (dd, 1H, J = 9.0, 9.3 Hz); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.2, 146.3, 134.2, 130.8, 130.4, 128.5, 125.4, 123.9, 121.9, 85.5, 74.4, 73.8, 72.1, 70.5. HRMS calcd for [C<sub>14</sub>H<sub>14</sub>ClN<sub>3</sub>O<sub>6</sub>-H]<sup>-</sup> 354.0466, found 354.0487.

**Compound L17-A5.** <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.55 (s, 1H), 7.39–7.44 (m, 2H), 7.34 (s, 1H), 7.00–7.01 (m, 1H), 6.12 (d, 1H, J = 9.0 Hz), 4.60 (d, 1H, J = 6.0 Hz), 4.11 (dd, 1H, J = 9.0, 9.3 Hz), 3.99 (dd, 1H, J = 6.0, 9.3 Hz), 3.87 (s, 1H), 3.78 (t, 1H, J = 9.3 Hz); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.3, 159.2, S11

147.3, 130.4 × 2, 121.7,118.5, 114.5, 111.0, 85.5, 74.3, 73.8, 72.1, 70.5, 55.3. HRMS calcd for  $[C_{15}H_{17}N_{3}O_{7}-H]^{-}$  350.0961, found 350.0983.

**Compound L17-A6.** <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O) δ 8.60 (s, 1H), 7.97–7.98 (m, 1H), 7.44–7.46 (m, 1H), 7.13–7.18 (m, 2H), 6.12 (d, 1H, J = 9.0 Hz), 4.60 (d, 1H, J = 6.0 Hz), 4.15 (dd, 1H, J = 9.0, 9.3 Hz), 3.99 (dd, 1H, J = 6.0, 9.3 Hz), 3.94 (s, 1H), 3.78 (t, 1H, J = 9.3 Hz); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.3, 155.7, 143.1, 130.2, 127.3, 124.2, 121.1, 117.7, 111.9, 85.4, 74.3, 73.8, 72.1, 70.5, 55.4. HRMS calcd for  $[C_{15}H_{17}N_{3}O_{7}-H]^{-}$  350.0959, found 350.0983.

**Compound L16-A7.**  $[\alpha]_D^{25}$  –26.4 (c 1.0 in H<sub>2</sub>O); <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.00 (s, 1H), 7.24–7.20 (m, 1H), 6.91–6.86 (m, 2H), 5.91 (d, 1H, J = 9.1 Hz), 4.46 (d, 1H, J = 6.1 Hz), 4.42 (s, 2H), 3.91 (t, 1H, J = 9.1 Hz), 3.84 (dd, 1H, J = 9.1, 6.1 Hz), 3.62 (t, 1H, J = 9.1 Hz), 3.57 (s, 2H); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O) δ 175.2, 173.6, 163.0, 161.7, 161.3, 160.0, 132.4, 132.3, 117.7, 111.4, 103.7, 85.3, 73.7, 72.1, 70.4, 35.1, 34.6. HRMS calcd for  $[C_{17}H_{18}F_2N_4O_7-H]^-$  427.1060, found 427.1056.

**Compound L16-A10.** [α]<sub>D</sub><sup>25</sup> –31.7 (*c* 1.0 in H<sub>2</sub>O); <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O) δ 7.91 (s, 1H), 6.77 (d, 1H, J = 7.9 Hz), 6.73 (s, 1H), 6.69 (d, 1H, J = 7.9 Hz), 5.90 (d, 1H, J = 9.1 Hz), 5.87 (s, 2H), 4.44 (d, 1H, J = 5.9 Hz), 4.39 (s, 2H), 3.87 (t, 1H, J = 9.1 Hz), 3.83–3.81 (m, 1H), 3.61 (t, 1H, J = 9.1 Hz), 3.45 (s, 2H); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.2, 174.8, 147.3 × 2, 146.2, 128.4, 123.0, 122.5, 109.5, 108.6, 101.1, 85.3, 74.2, 73.7, 72.2, 70.4, 41.8, 34.6. HRMS calcd for  $[C_{18}H_{20}N_4O_9-H]^-$  435.1147, found 435.1138.

**Compound L16-A11.**  $[\alpha]_{D}^{25}$  -29.7 (c 1.0 in H<sub>2</sub>O); <sup>1</sup>H NMR(600 MHz, D<sub>2</sub>O)  $\delta$  8.18 (s, 1H), 7.54–7.55 (m, 2H), 7.45 (d, 1H, 15.7Hz), 6.99–7.00 (m, 2H), 6.46 (d, 1H, 15.7 Hz), 6.03 (d, 1H, J = 9.0 Hz), 4.60 (s, 2H), 4.55 (s, 1H), 4.04 (t, 1H, J = 9.0 Hz), 3.94 (s, 1H), 3.84 (s, 2H), 3.72 (dd, 1H, J = 9.2, 9.3 Hz); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$ 175.2, 168.9, 160.5, 144.9, 141.2, 129.7 × 2, 127.3, 123.3, 117.3, 114.3 × 2, 85.3, 74.2, 73.7, 72.1, 70.4, 55.3, 34.6. HRMS calcd for [C<sub>19</sub>H<sub>22</sub>N<sub>4</sub>O<sub>8</sub>-H]<sup>-</sup> 433.1354, found 433.1353.

**Compound L16-A12.**  $[\alpha]_D^{25}$  -40.4 (c 1.0 in H<sub>2</sub>O); <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  7.95 (s, 1H), 7.21 (br s, 2H), 6.90 (br s, 2H), 6.01–6.00 (m, 1H), 4.56 (br s, 1H), 4.51 (s, 2H), 3.98–3.95 (m, 2H), 3.74–3.73 (m, 1H), 3.57 (s, 2H);  $^{13}$ C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.3, 175.2, 154.6, 145.0, 130.5 × 2, 126.7, 122.9, 115.7  $\times$  2, 85.3, 74.2, 73.7, 72.2, 70.4, 41.4, 34.6. HRMS calcd for  $[C_{17}H_{20}N_4O_8-H]^-$  407.1197, found 407.1193.

**Compound L16-B1.**  $[\alpha]_D^{25}$  –35.8 (c 1.0 in H<sub>2</sub>O); <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.09 (s, 1H), 7.28 (d, 1H, J = 15.8 Hz), 7.21 (dd, 1H, J = 7.8, 8.0 Hz), 7.04 (d, 1H, J = 7.8 Hz), 6.95 (s, 1H), 6.85 (d, 1H, J = 8.0 Hz), 6.39 (d, 1H, J = 15.8 Hz), 5.94 (d, 1H, J = 9.1 Hz), 4.47 (s, 2H), 4.44 (d, 1H, J = 6.1 Hz), 3.94 (dd, 1H, J = 9.1, 9.3 Hz), 3.83 (dd, 1H, J = 6.1, 9.3 Hz), 3.69 (s, 3H), 3.62 (t, 1H, J = 9.3 Hz); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.2, 168.4, 159.0, 141.1, 135.7, 130.1 × 2, 120.9 × 2, 119.9, 115.8 × 2, 112.9, 85.3, 73.7, 72.1, 70.4, 55.2, 34.5. HRMS calcd for [C<sub>19</sub>H<sub>22</sub>N<sub>4</sub>O<sub>8</sub>-H]<sup>-</sup> 433.1354, found 433.1347.

**Compound L16-B5.**  $[\alpha]_D^{25}$  –35.1 (*c* 1.0 in H<sub>2</sub>O); <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.04 (s, 1H), 5.93 (d, 1H, *J* = 9.1 Hz), 4.48 (d, 1H, *J* = 6.1 Hz), 4.43 (s, 2H), 3.96 (t, 1H, *J* = 9.1 Hz), 3.87 (dd, 1H, *J* = 9.1, 6.1 Hz), 3.64 (t, 1H, *J* = 9.1 Hz), 1.45 (q, 2H, *J* = 7.5 Hz), 1.07 (s, 6H), 0.66 (t, 3H, *J* = 7.5 Hz); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  181.8, 175.3, 145.4, 123.3, 85.3, 74.3, 73.7, 72.1, 70.5, 42.3, 34.5, 33.3, 24.0, 8.4. HRMS calcd for [C<sub>15</sub>H<sub>24</sub>N<sub>4</sub>O<sub>7</sub>-H]<sup>-</sup> 371.1561, found 371.1572.

**Compound L16-B6.**  $[\alpha]_D^{25}$  –31.5 (*c* 1.0 in H<sub>2</sub>O); <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.18 (s, 1H), 7.32–7.31 (m, 1H), 7.06 (s, 1H), 7.02 (s, 1H), 6.89 (s, 1H), 6.37–6.35 (m, 1H), 6.02 (s, 1H), 4.56 (s, 2H), 4.53 (s, 1H), 4.021–4.018 (m, 1H), 3.91 (s, 1H), 3.78 (s, 6H), 3.70 (s, 1H); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.3, 168.8, 149.8, 148.0, 144.9, 141.4, 127.4, 123.3, 122.6, 117.4, 111.4, 110.0, 85.3, 74.2, 73.7, 72.1, 70.4, 55.5, 55.4, 34.6. HRMS calcd for  $[C_{20}H_{24}N_4O_9-H]^-$  463.1460, found 463.1476.

**Compound L16-B8.**  $[\alpha]_D^{25}$  –39.0 (*c* 1.0 in H<sub>2</sub>O); <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.12 (s, 1H), 5.96 (d, 1H, *J* = 9.1 Hz), 4.49 (s, 3H), 3.98 (t, 1H, *J* = 9.1 Hz), 3.90–3.87 (m, 1H), 3.66 (t, 1H, *J* = 9.1 Hz), 3.22 (s, 2H), 3.70 (br s, 1H); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.3, 172.7, 144.7, 123.4, 85.4, 74.3, 73.8, 72.1, 70.5, 36.8, 34.7, 14.9. HRMS calcd for  $[C_{12}H_{18}N_4O_7S-H]^-$  361.0812, found 361.0821.

**Compound L16-C3 (18).**  $[\alpha]_D^{25}$  -27.1 (*c* 1.0 in H<sub>2</sub>O); <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.17 (s, 1H), 7.36–744 (m, 4H), 7.34 (s, 1H), 7.24 (s, 1H), 6.03 (d, 1H, *J*= 9.2 Hz), 4.59 (s, 2H), 4.53 (d, 1H, *J* = 6.1 Hz), 4.03 (t, 1H, *J* = 9.2 Hz), 3.92 (dd, 1H, *J* = 6.1, 9.4 Hz), 3.71 (dd, 1H, *J* = 9.2, 9.4 Hz), 2.02 (s, 3H); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.2, 172.9, 135.6, 134.6, 131.3, 129.3 × 2, 128.5 × 2, 128.2, 123.2, 85.3, 74.2, 73.7, 72.1, 70.4, 34.8, 13.4.HRMS calcd for [C<sub>19</sub>H<sub>22</sub>N<sub>4</sub>O<sub>7</sub>-H]<sup>-</sup> 417.1405, found 417.1397.

**Compound L16-C5.**  $[\alpha]_D^{25}$  –21.8 (*c* 1.0 in H<sub>2</sub>O); <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.10 (s, 1H), 7.54 (s, 1H), 6.93 (s, 1H), 6.85 (s, 2H), 6.47 (s, 1H), 5.94 (s, 1H), 4.50 (s, 2H), 4.45 (s, 1H), 3.93 (m, 1H), 3.83 (s, 1H), 3.70 (s, 3H), 3.67 (s, 3H), 3.61 (m, 1H); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.3, 168.8, 152.8, 152.4, 144.9, 136.1, 123.7, 123.3, 120.6, 117.0, 113.5, 113.2, 85.3, 74.2, 73.7, 72.1, 70.4, 56.3, 55.7, 34.6. HRMS calcd for  $[C_{20}H_{24}N_4O_9-H]^-$  463.1460, found 463.1450.

**Compound L16-C7.**  $[\alpha]_D^{25}$  –38.5 (*c* 1.0 in H<sub>2</sub>O); <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.09 (s, 1H), 7.69 (m, 1H), 7.44–7.47 (m, 1H), 7.34 (d, 1H, *J* = 15.7 Hz), 7.21–7.23 (m, 1H), 6.50 (d, 1H, *J* = 15.7 Hz), 5.91 (d, 1H, S12)

J = 9.2 Hz), 4.52 (s, 2H), 4.44 (d, 1H, J = 6.1 Hz), 3.93 (t, 1H, J = 9.2 Hz), 3.82 (dd, 1H, J = 6.1, 9.4 Hz), 3.60 (dd, 1H, J = 9.2, 9.4 Hz); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.2, 168.2, 144.7, 140.0, 136.4, 132.6, 130.5, 130.4, 126.7, 123.3, 122.2, 120.9, 85.2, 74.2, 73.7, 72.1, 70.4, 34.6. HRMS calcd for  $[C_{18}H_{19}BrN_4O_7-H]^-$  481.0353, found 481.0370.

**Compound L16-C8.**  $[\alpha]_D^{25}$  –24.8 (*c* 1.0 in H<sub>2</sub>O); <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  7.89 (s, 1H), 7.14 (d, 2H, *J* = 8.1 Hz), 7.10 (d, 2H, *J* = 8.1 Hz), 5.88 (d, 1H, *J* = 9.2 Hz), 4.45 (d, 1H, *J* = 6.1 Hz), 4.34 (s, 2H), 3.87 (t, 1H, *J* = 9.2 Hz), 3.82 (dd, 1H, *J* = 9.2, 6.1 Hz), 3.61 (t, 1H, *J* = 9.2 Hz), 3.49 (s, 2H), 2.22 (s, 3H); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.2, 174.9, 144.9, 137.4, 131.7, 129.5 × 2, 129.0 × 2, 123.0, 85.3, 74.2, 73.7, 72.1, 70.4, 41.8, 34.5, 20.1 .HRMS calcd for [C<sub>18</sub>H<sub>22</sub>N<sub>4</sub>O<sub>7</sub>-H]<sup>-</sup> 405.1405, found 405.1420.

**Compound L16-C9.**  $[\alpha]_D^{25}$  –20.0 (*c* 1.0 in H<sub>2</sub>O); <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  7.83 (s, 1H), 7.36 (s, 1H), 6.97 (d, 1H, *J* = 8.0 Hz), 6.86 (d, 1H, *J* = 8.0 Hz), 6.09 (d, 1H, *J* = 8.7 Hz), 4.63 (d, 1H, *J* = 6.0 Hz), 4.37 (s, 2H), 3.97 (dd, 1H, *J* = 6.0, 7.2 Hz), 3.91 (dd, 1H, *J* = 8.7, 8.8 Hz), 3.77 (t, 1H, *J* = 8.8 Hz), 3.42 (s, 2H); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  174.5, 173.0, 150.4, 130.3 × 2, 128.8 × 2, 127.9, 122.7, 120.2, 116.8, 85.4, 72.7, 71.7, 69.7, 41.0, 34.5.HRMS calcd for [C<sub>17</sub>H<sub>19</sub>ClN<sub>4</sub>O<sub>8</sub>-H]<sup>-</sup> 441.0808, found 441.0824.

**Compound L16-D1.**  $[\alpha]_D^{25}$  –25.4 (*c* 1.0 in H<sub>2</sub>O); <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  7 .85 (s, 1H), 7.51–7.53 (m, 1H), 6.89–6.94 (m, 2H), 5.91 (dd, 1H, *J* = 9.0, 9.1 Hz), 4.46 (d, 1H, *J* = 6.1 Hz), 4.38 (d, 1H, *J* = 15.4 Hz), 4.29 (dd, 1H, *J* = 15.4, 15.5 Hz), 3.82–3.92 (m, 1H), 3.83–3.85 (m, 1H), 3.76–3.78 (m, 3H), 3.67–3.72 (m, 1H), 3.61–3.66 (m, 1H), 2.82–2.87 (m, 1H), 2.66–2.69 (m, 1H), 2.52–2.56 (m, 1H), 2.44–2.49 (m, 1H); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  208.87, 175.2, 173.8, 165.8, 161.3, 144.5, 128.8, 125.4, 123.1, 116.2, 109.1, 85.2, 74.2, 73.7, 72.1, 70.4, 55.8, 42.5, 40.1, 35.0, 34.1. HRMS calcd for  $[C_{21}H_{24}N_4O_9-H]^-$  475.1460, found 475.1479.

**Compound L16-D3.**  $[\alpha]_D^{25}$  –34.6 (*c* 1.0 in H<sub>2</sub>O); <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.04 (s, 1H), 5.92 (d, 1H, *J* = 9.1 Hz), 4.44 (d, 1H, *J* = 6.2Hz), 4.39 (s, 2H), 3.92 (dd, 1H, *J* = 9.1, 9.2 Hz), 3.83 (dd, 1H, *J* = 6.2, 9.4 Hz), 3.61 (dd, 1H, *J* = 9.3, 9.4 Hz), 2.18–2.20 (m, 2H), 1.56–1.61 (m, 3H), 1.45–1.51 (m, 4H), 1.35–1.37 (m, 2H), 0.93–0.96 (m, 2H); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  177.4, 175.2, 123.2, 85.3, 74.2, 74.1, 73.7, 72.0, 70.4, 38.9, 34.9, 34.2, 31.8 × 2, 31.5, 24.6×2. HRMS calcd for  $[C_{17}H_{26}N_4O_7-H]^-$  397.1718, found 397.1718.

**Compound L16-D7.**  $[\alpha]_D^{25}$  –37.8 (*c* 1.0 in H<sub>2</sub>O); <sup>1</sup>H NMR(600 MHz, D<sub>2</sub>O)  $\delta$  8.11 (s, 1H), 7.95 (d, 1H, *J* = 8.4 Hz), 7.77 (d, 1H, *J* = 15.7 Hz), 7.6–7.61 (m, 2H), 7.46–7.49 (m, 1H), 6.43 (d, 1H, *J* = 15.7 Hz), 5.93 (d, 1H, *J* = 9.2 Hz), 4.53 (s, 2H), 4.44 (d, 1H, *J* = 6.2 Hz), 3.94 (t, 1H, *J* = 9.2 Hz), 3.83 (dd, 1H, *J* = 6.2, S14

9.4Hz), 3.61 (t, 1H, J = 9.4Hz); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.2, 167.6, 147.6, 144.6, 137.2, 134.1, 130.3, 130.2, 129.1, 124.8, 124.2, 123.4, 85.3, 74.2, 73.7, 72.1, 70.4, 34.5. HRMS calcd for  $[C_{18}H_{19}N_5O_9-H]^-$  448.1099, found 448.1115.

**Compound L16-D8.**  $[\alpha]_D^{25}$  –24.1 (*c* 1.0 in H<sub>2</sub>O); <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  7.93 (s, 1H), 7.39 (d, 1H, *J* = 8.3 Hz), 7.35 (d, 1H, *J* = 1.3 Hz), 7.08 (dd, 1H, *J* = 1.3, 8.3 Hz), 5.91 (d, 1H, *J* = 9.1 Hz), 4.44 (d, 1H, *J* = 6.0 Hz), 4.40 (s, 2H), 3.82–3.88 (m, 2H), 3.61 (t, 1H, *J* = 9.3 Hz), 3.51 (s, 2H); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.1, 173.6, 135.1, 131.7, 130.8 × 2, 130.5 × 2, 128.9 × 2, 123.0, 85.3, 73.6, 72.1, 70.3, 41.1, 34.5. HRMS calcd for  $[C_{17}H_{18}Cl_2N_4O_7-H]^-$  459.0469, found 459.0490.

**Compound L16-D9.**  $[\alpha]_D^{25}$  –30.0 (*c* 1.0 in H<sub>2</sub>O); <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  7.87 (s, 1H), 7.23–7.25 (m, 1H), 6.83–6.85 (m, 2H), 6.80–6.81 (m, 1H), 5.88 (d, 1H, *J* = 9.1 Hz), 4.44 (d, 1H, *J* = 6.1 Hz), 4.39 (s, 2H), 3.82 (dd, 1H, *J* = 6.1, 9.2 Hz), 3.80–3.81 (m, 1H), 3.71 (s, 3H), 3.60 (t, 1H, *J* = 9.2 Hz), 3.52 (s, 2H); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.2, 174.4, 159.0,136.4, 130.1, 122.9, 121.8, 114.4, 113.0, 112.0, 85.2, 74.2, 73.6, 72.1, 70.4, 55.2, 42.2, 34.5. HRMS calcd for [C<sub>18</sub>H<sub>22</sub>N<sub>4</sub>O<sub>8</sub>-H]<sup>-</sup> 421.1354, found 421.1344.

**Compound L16-D10.**  $[\alpha]_D^{25}$  –31.5 (*c* 1.0 in H<sub>2</sub>O); <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  7.87 (s, 1H), 7.13 (d, 2H, J = 8.8 Hz), 6.88 (d, 2H, J = 8.8 Hz), 5.88 (d, 1H, J = 9.2 Hz), 4.43 (d, 1H, J = 6.1 Hz), 4.38 (s, 2H), 3.85 (d, 1H, J = 9.2, 9.3 Hz), 3.80–3.83 (m, 2H), 3.72 (s, 3H), 3.60 (t, 1H, J = 9.3 Hz), 3.47 (s, 2H); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.2, 175.0, 157.9, 144.9, 130.3, 127.3, 122.9, 114.3,85.2, 74.1, 73.6, 72.1, 70.4, 55.3, 41.3, 34.5. HRMS calcd for  $[C_{18}H_{22}N_4O_8-H]^-$  421.1354, found 421.1367.

**Compound L16-D11.**  $[\alpha]_D^{25}$  –34.0 (*c* 1.0 in H<sub>2</sub>O); <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  7.91 (s, 1H), 7.18–7.20 (m, 2H), 7.00–7.03 (m, 2H), 5.89 (d, 1H, *J* = 9.2 Hz), 4.44 (d, 1H, *J* = 6.1 Hz), 4.39 (s, 2H), 3.87 (t, 1H, *J* = 9.2 Hz), 3.83 (dd, 1H, *J* = 6.1, 9.4 Hz), 3.60 (dd, 1H, *J* = 9.2, 9.4 Hz), 3.52 (s, 2H); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  175.2, 174.6, 162.6, 160.9, 144.8, 130.8, 130.7, 123.0, 115.5, 115.4, 85.2, 74.2, 73.6, 72.1, 70.4, 41.3, 34.5. HRMS calcd for  $[C_{17}H_{19}FN_4O_7-H]^-$  409.1154, found 409.1167.

**Compound L16-D12 (19).** <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  7.34 (s, 1H), 7.26–7.21 (m, 3H), 7.12–7.10 (m, 2H), 8.90 (dd, 1H, J = 9.1, 13.2 Hz), 4.50 (t, 1H, J = 5.4 Hz), 4.38 (dd, 1H, J = 11.5, 15.8 Hz), 4.13 (t, 1H, J = 15.4 Hz), 3.93 (t, 1H, J = 9.2 Hz), 3.89 (td, 1H, J = 4.1, 9.1 Hz), 3.68 (t, 1H, J = 9.4 Hz), 1.15–1.13 (m, 3H); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)  $\delta$  179.0, 175.2, 144.9, 139.6, 128.9, 128.6, 126.6, 123.0, 85.3, 74.4, 73.8, 72.2, 70.5, 43.0, 39.7, 34.1, 17.1. HRMS calcd for [C<sub>19</sub>H<sub>24</sub>N<sub>4</sub>O<sub>7</sub>-H]<sup>-</sup> 419.1561, found 419.1556.

5. Attempts to reduce the protected azido iduronic acid intermediate and amidation



- 1. After hydrogenation, the reaction resulted in decomposition of starting material.
- 2. The amide bond formation at C1 using the glycosyl azide and acyl chloride by Staudinger ligation.

## C. Biology section

### Inhibition assay

Compounds were mixed with 4-methylberiilyl  $\alpha$ -L-iduronic acid as substrate and rh- $\alpha$ -IDUA in pH 3.5 (citric phosphate) buffer, then assay was carried out at 37 °C for 15 min. Stop solution (0.4 mol/L K<sub>2</sub>CO<sub>3</sub>, pH 10.6) was then added and fluorescence was determined at 355 nm excitation and 460 nm emission. Inhibition was performed as relative enzyme activity to control. The active compounds were selected and further tested at lower concentration to determine their IC<sub>50</sub> values. The assays performed in 96-wells of the microtiter plate contained citric phosphate buffer.



Figure S2. The inhibition activity of 48-membered iduronyl triazoles at 100  $\mu$ M



Figure S3. The IC<sub>50</sub> plots of (a) compound 18 and (b) 2 versus rh- $\alpha$ -IDUA.

## In vitro stabilization of rh-α-IDUA<sup>S3</sup>

An assessment of the ability of small molecules to stabilize rh- $\alpha$ -IDUA against denaturation was performed by using Aldurazyme. Enzyme aliquots (20  $\lambda$ , pH 3.5) were incubated with 0, 100, 250 or 500  $\mu$ M of small molecule on ice for 10 min. The samples were heated at 48 °C as a function of time in an

#### Supporting Information

attempt to heat-inactivate (denature) enzyme, and then the samples were diluted into twenty-fold volume of 0.1 M citric phosphate buffer (pH 3.5). The enzyme was immediately incubated with substrate (0.5 mM 4-methyllumbelliferyl  $\alpha$ -L-iduronic acid) for 15 min at 37 °C before quenching with glycine buffer. Liberated 4-methylumbelliferone was measured (excitation 355 nm, emission 460 nm). Enzyme activity was reported relative to unheated enzyme.

## Thermal stability shift assay<sup>S4</sup>

The stability of rh- $\alpha$ -IDUA was assessed using a modified fluorescence thermal stability assay on a Rotor-Gene system in acidic buffer (citric phosphate, pH 3.5). Briefly, rh- $\alpha$ -IDUA (4 µg) was combined with SYPRO Orange and various concentrations of small molecules in a final reaction volume of 20  $\lambda$ . A thermal gradient was applied to the plate at a rate of 1 °C per minute, during which time the fluorescence of SYPRO Orange was continuously monitored. The fluorescence intensity at each temperature was normalized to the maximum fluorescence after complete thermal denaturation.

**TableS2.** Thermal shift study of glycerol toward rh- $\alpha$ -IDUA.

Concentration/conditions	Tm (°C)
0 mM (pH 3.5)	56.15
40 mM (pH 3.5)	56.32



**Figure S4.** Thermal stability scans of rh- $\alpha$ -IDUA in the absence and presence of various concentrations of small molecules. All experiments were performed at pH 3.5, with the exception of Apo (pH 3.5, black). Unfolding of IDUA was monitored by changes in the fluorescence SYPRO Orange as a function of temperature.

## D. Copies of <sup>1</sup>H and <sup>13</sup>C NMR spectra

 $^1\mathrm{H}$  NMR and  $^{13}\mathrm{C}$  NMR of compound 5 in CDCl\_3



## <sup>1</sup>H NMR and <sup>13</sup>C NMR of compound 6 in CDCl<sub>3</sub>





Supporting Information



Supporting Information

Page S23

240 220 200 180 160 140 120 100 80 60 40 20 0 ppm

S23



C14N3-Alpha @ 2D NOESY noesygpph











S25

## $^{1}$ H NMR and $^{13}$ C NMR of compound **10** in D<sub>2</sub>O







## $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of compound 11 in D2O



# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of compound $\boldsymbol{12}$ in $D_2\text{O}$

Supporting Information





## Supporting Information <sup>1</sup>H NMR and <sup>13</sup>C NMR of compound **S1** in CDCl<sub>3</sub>





0 ppm

S32



240 220 200 180 160 140 120 100 80 60 40 20 0 ppm



S34



Supporting Information



0 ppm





# $^1\mathrm{H}$ NMR and $^{13}\mathrm{C}$ NMR of compound 15b in $D_2\mathrm{O}$

Supporting Information



# Supporting Information $^{1}$ H NMR and $^{13}$ C NMR of compound **18** in D<sub>2</sub>O



## $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of compound 19 in $D_2\text{O}$



# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of compound L17-A1 in $D_2\text{O}$



## <sup>1</sup>H NMR and <sup>13</sup>C NMR of compound L17-A2 in $D_2O$





Supporting Information





Supporting Information



## $^{1}$ H NMR and $^{13}$ C NMR of compound L17-A5 in D<sub>2</sub>O









S47

# $^1\mathrm{H}$ NMR and $^{13}\mathrm{C}$ NMR of compound L16-A10 in D2O





240 220 200 180 160 140 120 100 80 60 40 20 0 ppm



240 220 200 180 160 140 120 100 80 60 40 20 0 ppm









190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 ppm



240 220 200 180 160 140 120 100 80 60 40 20 0 ppm



190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 ppm

S54



# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of compound L16-C7 in $D_2\text{O}$



240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 ppm



<sup>240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20</sup> ppm









240 220 200 180 160 140 120 100 80 60 40 20 0 ppm





240 220 200 180 160 140 120 100 80 60 40 20 0 ppm

# $^{1}$ H NMR and $^{13}$ C NMR of compound L16-D9 in D<sub>2</sub>O







<sup>240 220 200 180 160 140 120 100 80 60 40 20 0</sup> ppm

## **References and Notes:**

- S1. C. E. Nieman, A. W. Wong, S. He, L. Clarke, J. J. Hopwood and S. G. Withers, Biochemistry, 2003, 42, 8054-8065.
- S2. A. W. Wong, S. He and S. G. Withers, Can. J. Chem., 2001, 79, 510–518.
- S3. W.-C. Cheng, J.-H. Wang, H.-Y. Li, S.-J. Lu, J.-M. Hu, W.-Y. Yun, C.-H. Chiu, W.-B. Yang, Y.-H. Chien, W.-L. Hwu, *Eur. J. Med. Chem.*, 2001, **123**, 14–20.
- S4. T. Takai, K. Higaki, M. Aguilar-Moncayo, T. Mena-Barragán, Y. Hirano, K. Yura, L. Yu, H. Ninomiya, I.M. García-Moreno, Y. Sakakibara, *Mol. Ther.*, 2013, 21, 526–532.