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Supporting Information For

A cerium-based metal-organic framework having inherent oxidase-like activity

applicable for colorimetric sensing of biothiols and aerobic oxidation of thiols

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Compound	<i>a</i> (Å)	$V(Å^3)$
as-synthesized 1	23.111(6)	12343.6(54)
[Zr ₆ O ₄ (OH) ₄ (DMTDC) ₆]·4.8DMF·10H ₂ O ^[1]	23.0917(6)	12313.1(3)
Zr-DMTDC ^[2]	23.12	12358.43

Table S1. Refined unit cell parameters for as-synthesized 1 and isostructural Zr(IV)-based compound having cubic unit cells.



Figure S1. XPS spectrum of the as-synthesized form of 1 (Ce 3d core level).



Figure S2. FT-IR spectra of the as-synthesized (black) and thermally activated (red) forms of **1**.



Figure S3. TG curves of as-synthesized (red) and thermally activated (black) forms of **1** measured under air atmosphere in the range 25-700 $^{\circ}$ C at a heating rate of 5 $^{\circ}$ C min⁻¹.



Figure S4. XRPD patterns of 1' after treatment with (a) water, (b) 1M HCl and (c) acetic acid.





Figure S5. FE-SEM images of activated 1'.



Figure S6. Effect of pH on the oxidase-like catalytic activity of 1' towards TMB.



Figure S7. UV-vis absorption spectra of AzBTS in the absence (black) and presence (red) of **1**'. Inset: digital photographs of vials containing AzBTS in the absence (a) and presence (b) of **1**'.



Figure S8. Recyclability of the oxidase-like catalytic activity of 1' towards TMB.



Figure S9. XRPD patterns of 1' before and after oxidase-like catalytic activity towards TMB.



Figure S10. Steady-state kinetic assay of 1' with TMB at pH = 4. Inset: Lineweaver–Burk plot of the double reciprocal of the Michaelis–Menten equation.

Sl. No.	Nano-enzyme	$K_{\rm m}$ (mM)	$V_{\rm max}$ (M s ⁻¹)	Ref.
1.	Ce-DMTDC (1')	0.088	0.11 × 10 ⁻⁶	This work
2.	HRP	0.43	10.0 × 10 ⁻⁸	[3]
3.	isPNC (nanoceria)	3.8	0.7 × 10 ⁻⁶	[4]
4.	swPNC (nanoceria)	1.9	0.6 × 10 ⁻⁶	
5.	isDNC (nanoceria)	1.8	0.5 × 10 ⁻⁶	
6.	swDNC (nanoceria)	0.8	0.3 × 10 ⁻⁶	
7.	MIL-53(Fe)	1.08	8.78 × 10 ⁻⁸	[5]
8.	JFSNs	3.05	4.16 × 10 ⁻⁶	[6]
9.	Fe ₃ O ₄ @C	0.38	73.99 × 10 ⁻⁸	[7]
10.	Ce-MOF (MVCM)	0.37	5.5 × 10 ⁻⁶	[8]
11.	CuNPs@C	1.65	12.05 × 10 ⁻⁸	[9]
12.	Fe-MIL-88NH ₂	0.284	10.47 × 10 ⁻⁸	[10]
13.	hemin@MIL-53(Al)-	0.068	6.07 × 10 ⁻⁸	[11]

Table S2. Comparison of the kinetic parameters of different nano-enzymes that mimic peroxidase/oxidase at pH = 4.

	NH ₂			
14.	MIL-53(Fe) by MW	0.28	4.48×10^{-8}	[12]
15.	Fe-MIL-101-FA	0.164	1.55 × 10 ⁻⁸	[13]



Figure S11. Absorbance spectra of ox-TMB in presence of glutathione at varying concentration.



Figure S12. Absorbance spectra of ox-TMB in presence of homocysteine at varying concentration.



Figure S13. Change in the absorbance of ox-TMB in presence of different biothiols.



Figure S14. UV-vis absorption spectra of ox-TMB in presence of various concentrations of alanine.



Figure S15. UV-vis absorption spectra of ox-TMB in presence of various concentrations of arginine.



Figure S16. UV-vis absorption spectra of ox-TMB in presence of various concentrations of aspartic acid.



Figure S17. UV-vis absorption spectra of ox-TMB in presence of various concentrations of glycine.



Figure S18. UV-vis absorption spectra of ox-TMB in presence of various concentrations of histidine.



Figure S19. UV-vis absorption spectra of ox-TMB in presence of various concentrations of isoleucine.



Figure S20. UV-vis absorption spectra of ox-TMB in presence of various concentrations of leucine.



Figure S21. UV-vis absorption spectra of ox-TMB in presence of various concentrations of lysine.



Figure S22. UV-vis absorption spectra of ox-TMB in presence of various concentrations of methionine.



Figure S23. UV-vis absorption spectra of ox-TMB in presence of various concentrations of phenyl alanine.



Figure S24. UV-vis absorption spectra of ox-TMB in presence of various concentrations of proline.



Figure S25. UV-vis absorption spectra of ox-TMB in presence of various concentrations of serine.



Figure S26. UV-vis absorption spectra of ox-TMB in presence of various concentrations of threonine.



Figure S27. UV-vis absorption spectra of ox-TMB in presence of various concentrations of tryptophan.



Figure S28. UV-vis absorption spectra of ox-TMB in presence of various concentrations of valine.



Figure S29. Change in the UV-vis absorption spectra of ox-TMB in presence of alanine, followed by the addition of cysteine.



Figure S30. Change in the UV-vis absorption spectra of ox-TMB in presence of arginine, followed by the addition of cysteine.



Figure S31. Change in the UV-vis absorption spectra of ox-TMB in presence of aspartic acid, followed by the addition of cysteine.



Figure S32. Change in the UV-vis absorption spectra of ox-TMB in presence of glycine, followed by the addition of cysteine.



Figure S33. Change in the UV-vis absorption spectra of ox-TMB in presence of histidine, followed by the addition of cysteine.



Figure S34. Change in the UV-vis absorption spectra of ox-TMB in presence of isoleucine, followed by the addition of cysteine.



Figure S35. Change in the UV-vis absorption spectra of ox-TMB in presence of leucine, followed by the addition of cysteine.



Figure S36. Change in the UV-vis absorption spectra of ox-TMB in presence of lysine, followed by the addition of cysteine.



Figure S37. Change in the UV-vis absorption spectra of ox-TMB in presence of methionine, followed by the addition of cysteine.



Figure S38. Change in the UV-vis absorption spectra of ox-TMB in presence of phenyl alanine, followed by the addition of cysteine.



Figure S39. Change in the UV-vis absorption spectra of ox-TMB in presence of proline, followed by the addition of cysteine.



Figure S40. Change in the UV-vis absorption spectra of ox-TMB in presence of serine, followed by the addition of cysteine.



Figure S41. Change in the UV-vis absorption spectra of ox-TMB in presence of threonine, followed by the addition of cysteine.



Figure S42. Change in the UV-vis absorption spectra of ox-TMB in presence of tryptophan, followed by the addition of cysteine.



Figure S43. Change in the UV-vis absorption spectra of ox-TMB in presence of valine, followed by the addition of cysteine.



Figure S44. Change in the absorbance (monitored at $\lambda_{max} = 652$ nm) of ox-TMB as a function of cysteine concentration.



Figure S45. Change in the absorbance (monitored at $\lambda_{max} = 652$ nm) of ox-TMB as a function of glutathione concentration.



Figure S46. Change in the absorbance (monitored at $\lambda_{max} = 652$ nm) of ox-TMB as a function of homocysteine concentration.

Sl. No.	Nano-enzyme	Analytes	Dynamic range (µM)	Detection limit (µM)	Ref.
1.	Ce-DMTDC (1')	Cys		0.150	This work
		Нсу	0-1.0	0.125	
		GSH		0.132	
2.	Fe-MIL-88-NH ₂	Cys	1.0-80.0	0.390	[14]
		Нсу	1.0-80.0	0.400	
		GSH	1.0-100.0	0.450	
3.	Ce-MOF (MVCM)	Cys	0-40.0	0.139	[8]
		Нсу		0.143	
		GSH		0.129	
4.	photocatalytic UiO-66-NH ₂	Cys	5.0-120.0	0.306	[15]
		Нсу		0.330	
		GSH]	0.310	

Table S3. Comparison of the detection limits of different nano-enzymes (which mimic peroxidase/oxidase at pH = 4) towards biothiols.



Figure S47. Reusability of 1' for the aerobic oxidation of thiophenol under the optimized reaction conditions.



Figure S48. XRPD patterns of 1' before and after catalytic oxidation of thiophenol.



Figure S49. GC trace for the oxidation of thiophenol.



Figure S50. GC trace for the oxidation of 2-aminothiophenol.



Figure S51. GC trace for the oxidation of 4-methylthiophenol.



Figure S52. GC trace for the oxidation of 2-fluorothiophenol.



Figure S53. GC trace for the oxidation of 4-bromothiophenol.



Figure S54. GC trace for the oxidation of 3-chlorothiophenol.



Figure S55. GC trace for the oxidation of cyclohexanethiol.



Figure S56. GC trace for the oxidation of 4-methoxythiophenol.

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