

Supplementary Information

Beware of phosphate: evidence of specific dendrimer— phosphate interactions

Mónika Kéri^{a*}, Zoltán Nagy^a, Levente Novák^a, Edit Szarvas^a, Lajos P. Balogh^b and István

Bányai^a

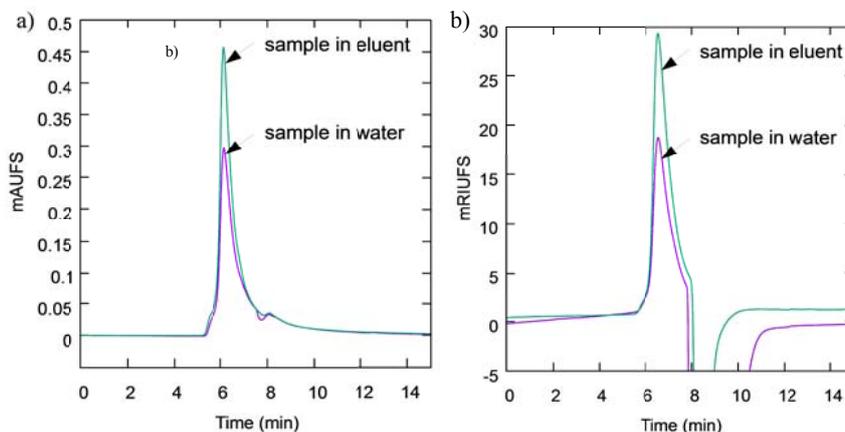
^aDepartment of Physical Chemistry, University of Debrecen, Egyetem tér 1, H-4032 Hungary

^bAA Nanomedicine & Nanotechnology, North Andover, MA 01845, USA

*E-mail: keri.monika@science.unideb.hu

Size Exclusion Chromatography results:

For dendrimer samples dissolved in water, both area and height of the chromatographic peaks were always smaller than for dendrimer samples dissolved in the chromatographic eluent (SI Fig. 1). Moreover, these effects were independent of the detector used (either UV spectrophotometer at $\lambda=210$ nm or differential refractometer, RI, shows these differences). Peak areas and heights for G5.NH₂ dissolved in water were around only 65% of the respective parameters obtained for that dissolved in the eluent.



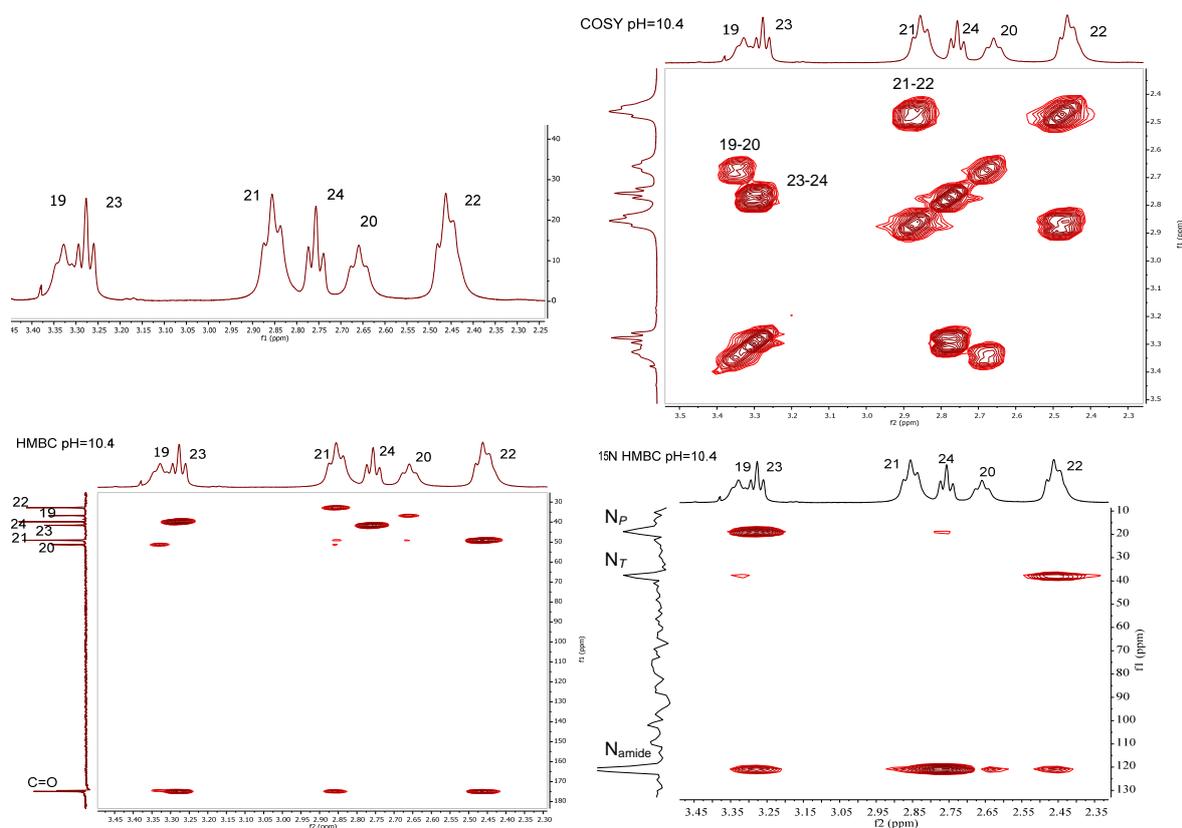
SI Figure 1. Size exclusion chromatograms of 1 g/l G5 PAMAM dissolved and stored in eluent and water respectively. Detection: UV at $\lambda=210$ nm (a) and RI (b). Eluent: 100 mM Na₂HPO₄, 100 mM NaH₂PO₄ and 10% CH₃CN in water. Flow rate: 0.5 ml/min, $T=303$ K, pH~6.8.

For dendrimer samples in water, the initial contact of macromolecules with phosphate ions can only take place inside the column with a time scale of a few minutes determined by the elution time. However, for samples dissolved directly in the eluent, longer time is available for any possible interaction. The interaction might appear as an increased detector signal (SI

Fig. 1) when phosphate ions are already present in the injection vial, provided that the approximate time scale of this phenomenon is larger than the elution time.

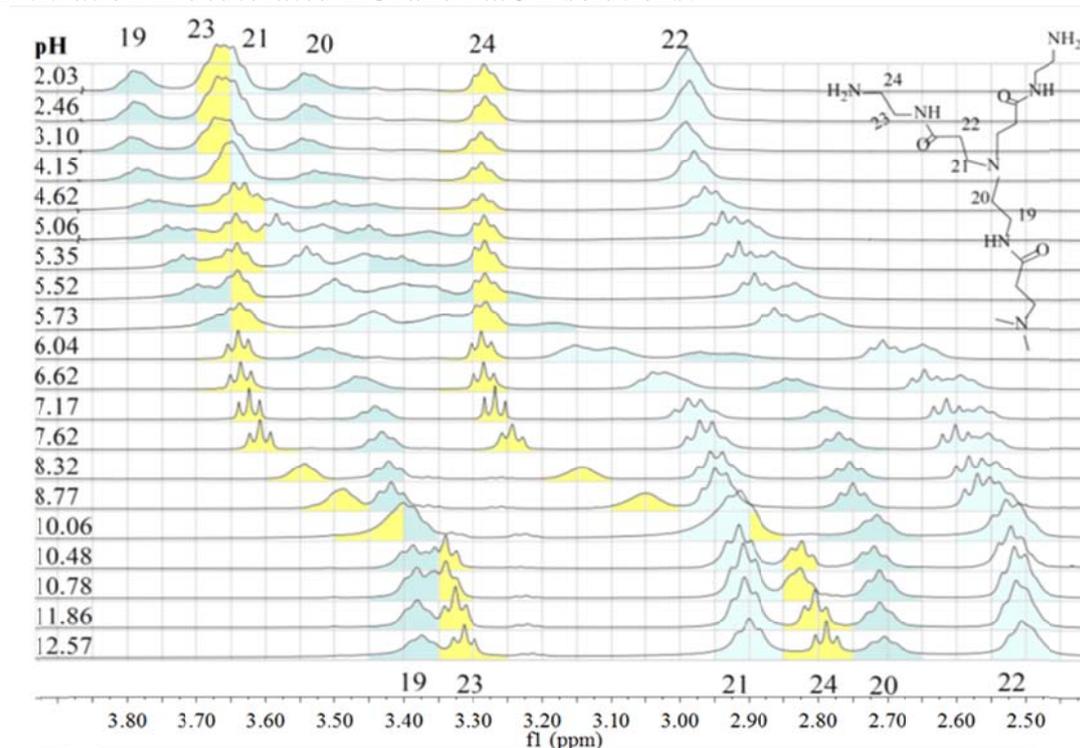
^1H NMR peak assignment:

The peak assignment was proved with COSY (Correlation Spectroscopy), showing the scalar coupling between protons, and HMBC (Heteronuclear Multiple Bond Correlation Spectroscopy) spectra, showing the same between protons and carbons. In order to determine the spatial structure of the molecule NOESY experiment was used, showing that dipolar coupling arises between protons near the tertiary nitrogens in the branching points.



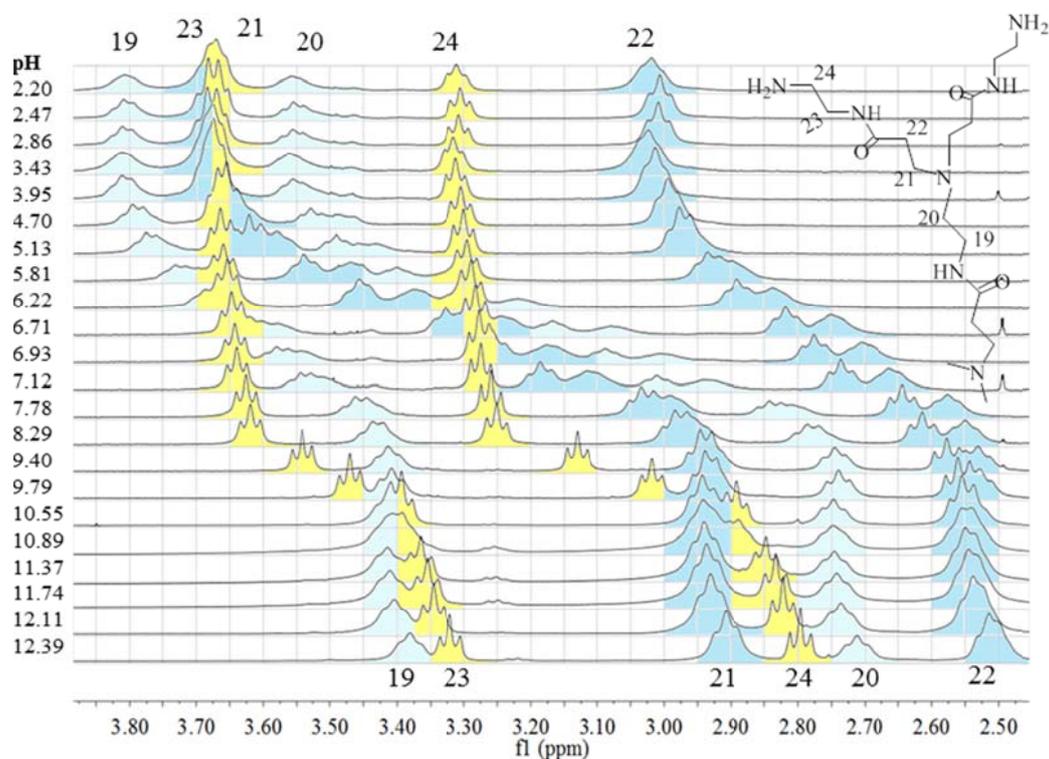
SI Figure 2. ^1H , COSY, ^{13}C and ^{15}N HMBC NMR spectra of G5.NH₂ (10 mg/ml G5.NH₂, D₂O, pH = 10.4, $T = 298$ K).

NMR titration in deuterated HCl and NaOH solutions:

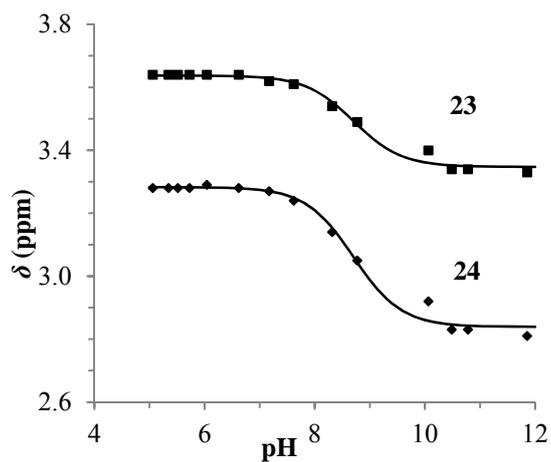


SI Figure 3. ^1H NMR spectra of G5.NH₂ dendrimer as a function of pH (blue for N_T, yellow for N_P protons, 10 mg/ml G5.NH₂, D₂O, $T = 298$ K).

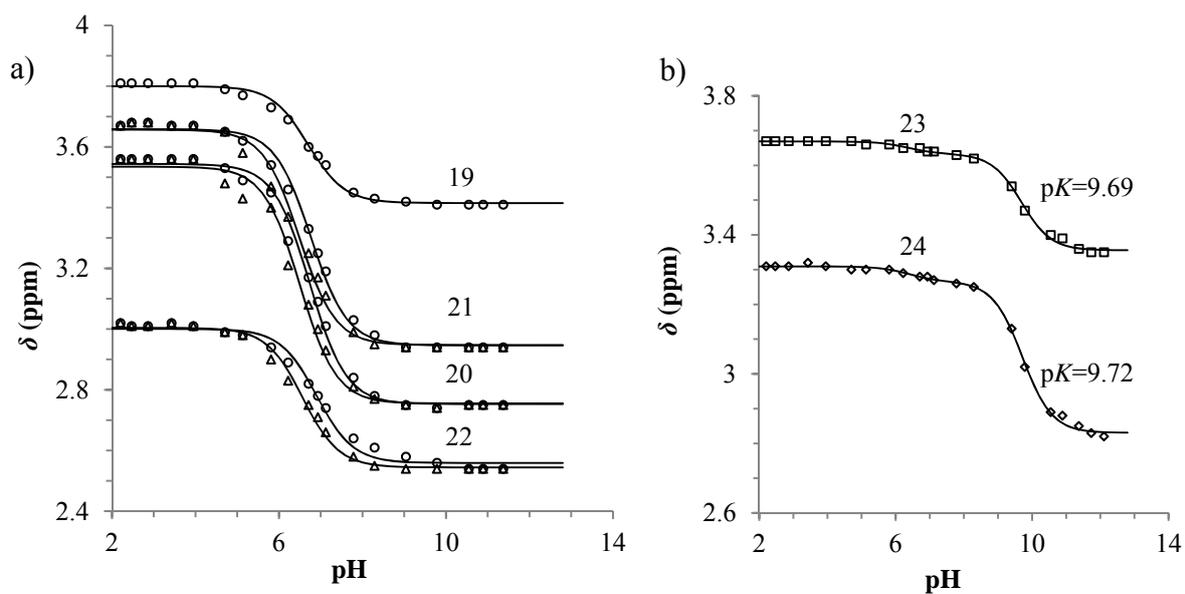
NMR titration in PBS:



SI Figure 4. ^1H NMR spectra of G5.NH₂ dendrimer as a function of pH set with PBS (10 mg/ml G5.NH₂, D₂O, $c_{\text{H}_2\text{PO}_4} = 0.1$ M, $T = 298$ K).

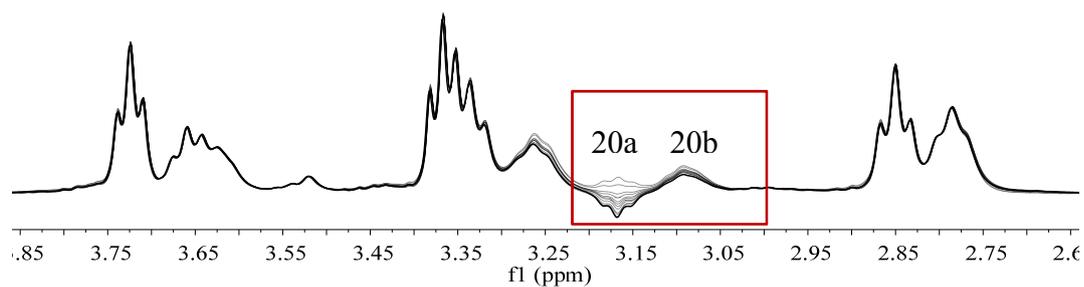


SI Figure 5. The pH dependence of the chemical shifts of CH₂-peaks 23-24 in the absence of phosphate ions (10 mg/ml G5.NH₂, D₂O, c_{H_xPO₄} = 0.1 M T = 298 K).

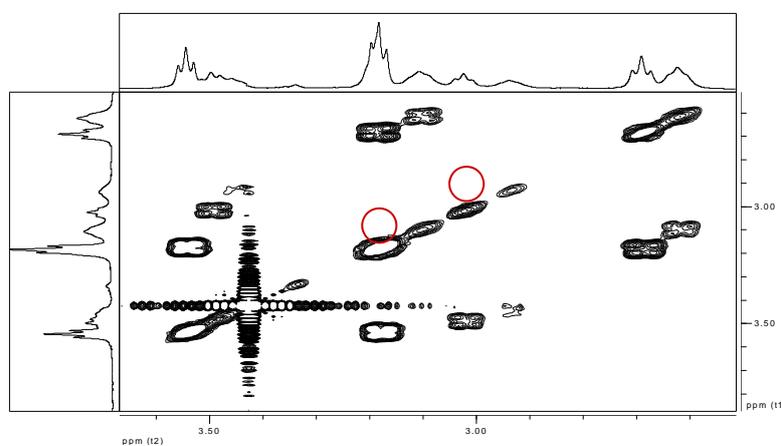


SI Figure 6. The pH dependence of the chemical shifts of CH₂-peaks 19-22 (a) and 23-24 (b) in PBS: in the case of duplicated peaks \circ is for peak 'a' and Δ is for peak 'b' (10 mg/ml G5.NH₂, D₂O, c_{H_xPO₄} = 0.1 M T = 298 K).

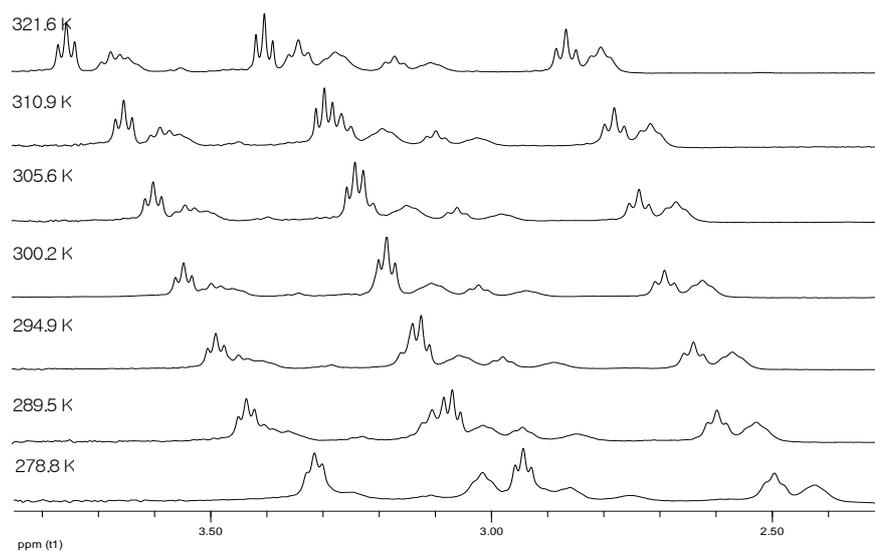
Study of peak duplication:



SI Figure 7. Selective excitation of peak 20a (10 mg/g G5.NH₂, c_{H₂PO₄}=0.023M, D₂O, pH=6.4, T = 298 K)

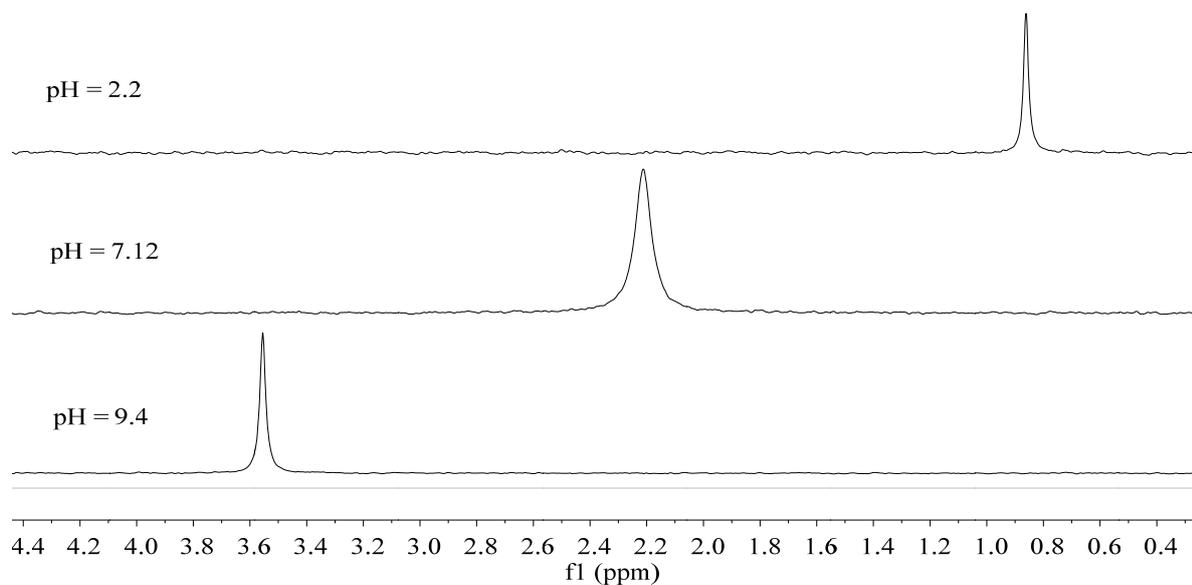


SI Figure 8. COSY spectrum of G5.NH₂ with phosphate (10 mg/g G5.NH₂, c_{H₂PO₄}=0.023M, D₂O, pH=6.4, T = 298 K).

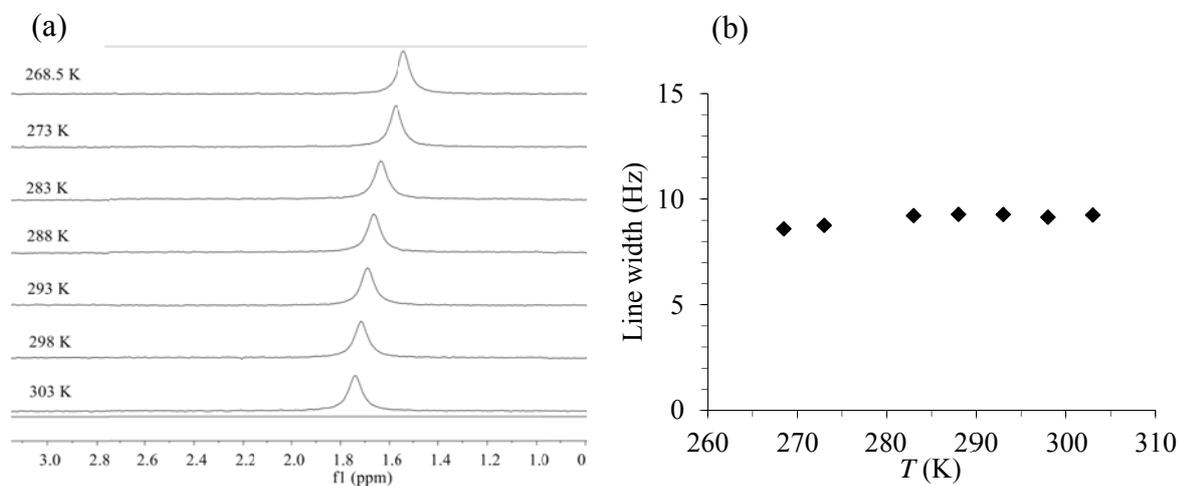


SI Figure 9. Effect of temperature on the ¹H spectrum of G5.NH₂ with phosphate (10 mg/g G5.NH₂, c_{H₂PO₄} = 0.023 M, D₂O, pH = 6.4, T = 279-322 K).

³¹P spectra:

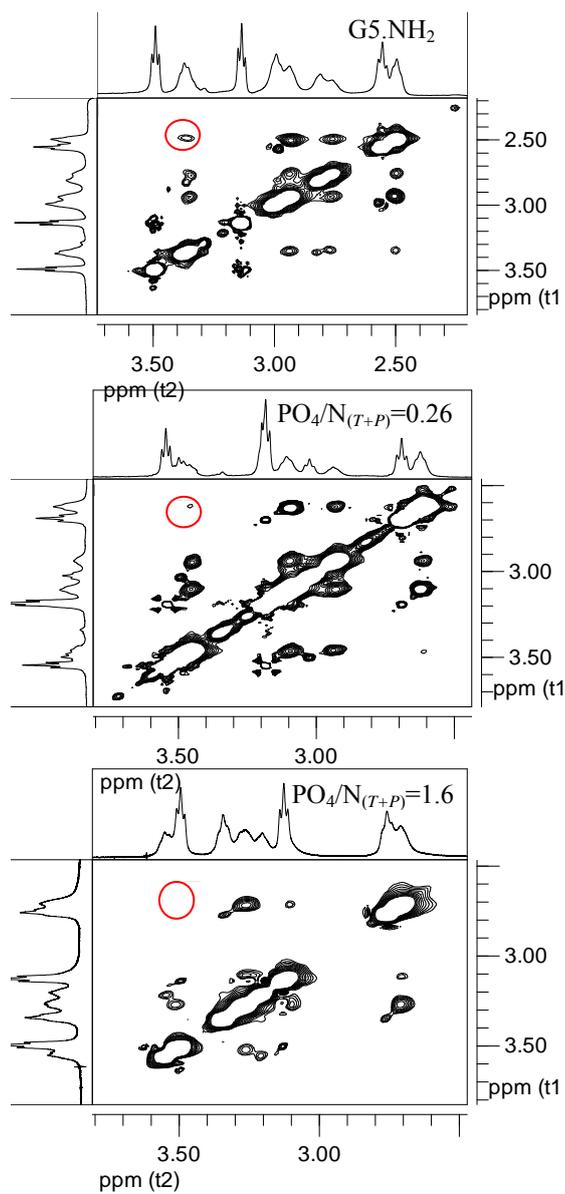


SI Figure 10. ³¹P line width in the presence of G5.NH₂ at different pH. (10 mg/ml G5.NH₂, c_{H_xPO₄} = 0.1 M, T = 298 K).

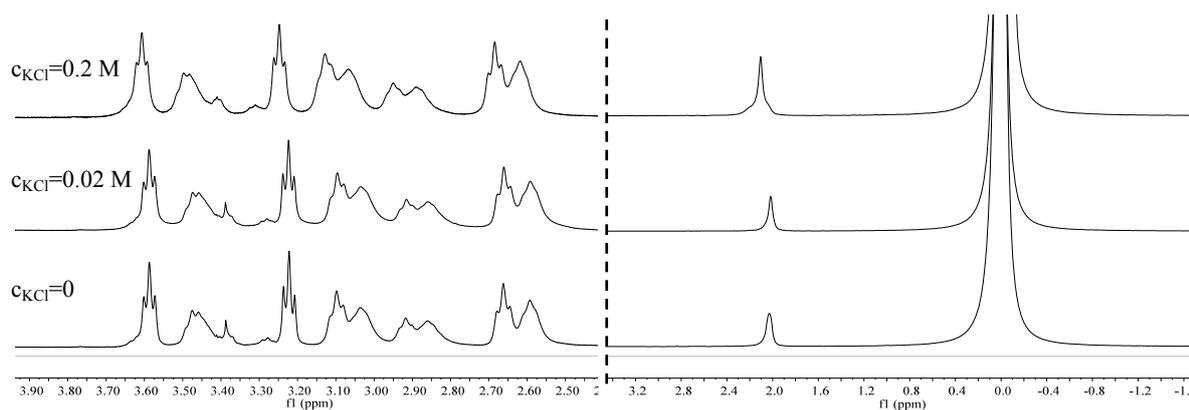


SI Figure 11. ³¹P spectra (a) and line width (b) as a function of temperature (10 mg/ml G5.NH₂, c_{H_xPO₄} = 0.1 M, pH = 6.71, T = 268-303 K).

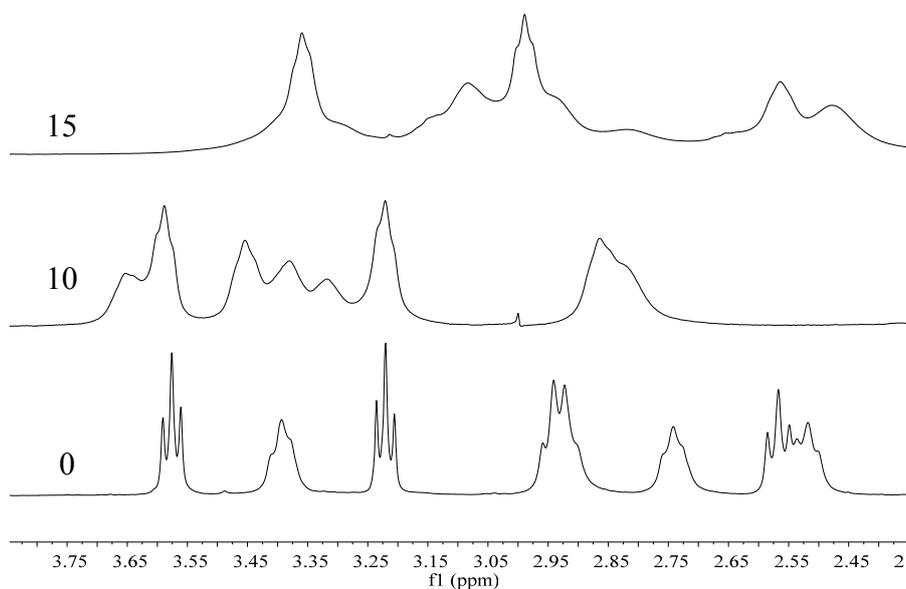
PBS concentration dependence:



SI Figure 12. NOESY spectra of G5.NH₂ by increasing phosphate concentration (10 mg/ml G5.NH₂, c_{H₂PO₄}=0.02 and 0.14 M, pH=6.4, mixing time: 150 ms).

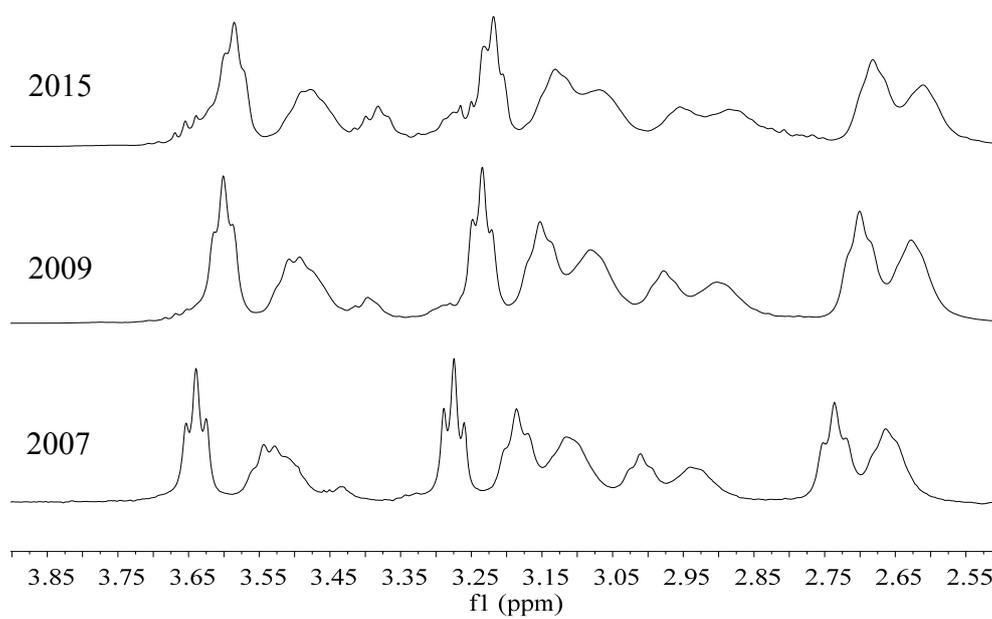


SI Figure 13. ^1H (left) and ^{31}P (right) NMR spectra (with H_3PO_4 as reference material at 0 ppm) of G5.NH_2 in PBS by increasing KCl concentration, as labelled (10 mg/ml G5.NH_2 , $\text{PO}_4/\text{N}_{(T+P)} = 0.5$, $\text{pH}=7.2$, $T=298\text{ K}$).



SI Figure 14. ^1H NMR spectra of G5.NH_2 by excess PBS concentrations (10 mg/ml G5.NH_2 , $\text{PO}_4/\text{N}_{(T+P)} = 0, 10$ and 15 , as labelled, $\text{pH}=7.2$, $T=298\text{ K}$)

Time dependence:



SI Figure 15. ¹H NMR spectra of the same G5.NH₂ sample in PBS, recorded in three different years (10 mg/g G5.NH₂, D₂O, c_{H₂PO₄}=0.1 M, pH=7.12).