## Effect of ligand binding on a protein with a complex folding landscape

Hisham Mazal, Haim Aviram, Inbal Riven and Gilad Haran\*

Department of Chemical Physics, Weizmann Institute of Science, Rehovot 76100, Israel

\*Send correspondence to: Gilad Haran (gilad.haran@weizmann.ac.il)

**Supplementary Information** 



Figure S1| Comparison between bulk chemical denaturation and smFRET denaturation curves. (A) Bulk chemical denaturation of AKE at 0 M (green) and 200  $\mu$ M (red) Ap<sub>5</sub>A. (B) smFRET denaturation of AKE at 0 M (blue) and 200  $\mu$ M (orange) Ap<sub>5</sub>A. Free energies of unfolding at 0 M and 200  $\mu$ M Ap<sub>5</sub>A, obtained from a fitting to a two-state model were in a good agreement between the two methods (see Table S1). (C) Full denaturation curve of AKE at 0 M Ap<sub>5</sub>A constructed from smFRET experiment. The shaded area indicates the range of denaturant concentrations over which the MST measurements of Fig. 4A were conducted, showing that the protein is still folded within this range.



Figure S2| Urea denaturation experiments using smFRET in the presence of  $Ap_5A$ . Free energy versus denaturant concentration at different  $Ap_5A$  concentrations (given in the legend in  $\mu$ M).



**Figure S3 AMP binding to AKE measured by MST.** The fraction of bound AKE as a function of AMP concentration is plotted. Data was fitted to obtain a K<sub>d</sub> value of 286.0  $\pm$  56.9  $\mu$ M, close to the literature value of 210  $\mu$ M<sup>1</sup>. The error values shown in the figure were calculated from three repeats of the experiment.



Figure S4| Effect of AMPNP binding on AKE stability measured using urea denaturation. (A) MST measurement of AMPPNP binding to native single-labeled AKE. The fraction of bound AKE as a function of AMPPNP concentration is plotted. From a fit to the data we obtained a K<sub>d</sub> of 83.7 ± 21.2  $\mu$ M, close to the literature value of 50  $\mu$ M<sup>1</sup>. (B) The change in AKE stability with AMPPNP concentration was measured. No significant stabilization was registered. The error values shown A were calculated from three repeats of the experiment.



Figure S5| Bulk thermal denaturation of AKE in the presence of  $Ap_5A$ . (A) Thermal unfolding of AKE at different substrate concentrations (given in the legend in  $\mu$ M) was monitored by CD at 222 nm ( $\theta_{222}$ ). The graph shown was obtained by using the "Van't Hoff with no baseline" method <sup>2</sup>, and shows the derivative of the CD signal with temperature. Curves were fitted to a two-state denaturation model to obtain the transition enthalpy and the melting temperature, from which the free energy of binding was calculated (equation 2 of the main text). (B) Effect of  $Ap_5A$  concentration on the free energy of binding. A K<sub>d</sub> value of  $10.9 \pm 1.2 \mu$ M was obtained from a fit of these results, which is much higher than the value measured by MST, as well as literature value <sup>1</sup>, but in good agreement with the result obtained from the smFRET chemical denaturation curves. The error values for the data in (B) are very small and are not shown. They were calculated from two repeats of the experiment.



Figure S61 Human carbonic anhydrase stability in the presence of the inhibitor ligand trifluoromethane sulfonamide (TFMSA). (A) Bulk chemical denaturation at 0 and 10  $\mu$ M of the inhibitor. (B) Free energy versus denaturant concentration calculated from the results in A. Ligand binding increases protein stability significantly. The calculated K<sub>d</sub> obtained using equation 1 of the main text is 33.7 ± 6.9 nM, close to the literature value of 163 nM <sup>3</sup>.

**Table 1**. Comparison of between the free energies of unfolding of AKE obtained from bulk and smFRET experiments.

$^{Ap_{5}A}$ concentration [ $^{\mu}$ M]	Bulk [Kcal/mol]	smFRET [Kcal/mol]
0	$4.6 \pm 0.3$	$4.8\pm0.4$
200	$5.2 \pm 0.4$	$5.5 \pm 0.4$

## References

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