

ELECTRONIC SUPPLEMENTARY INFORMATION

Direct surface-enhanced Raman scattering (SERS) spectroscopy of nucleic acids: from fundamental studies to real-life applications

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Table S1. Summary of the reviewed articles listed according to the silver colloids exploited as SERS substrates: citrate-reduced silver nanoparticles (AgCT), hydroxylamine-reduced silver nanoparticles (AgHX), iodide-modified citrate silver nanoparticles (AgI), and spermine-coated silver nanoparticles (AgSp). The (potential) aggregating agent employed, as well as the description of the targeted nucleic acids and major findings, were also reported.

COLLOIDS	AGGR. AGENT	NUCLEIC ACIDS	MAJOR FINDINGS	REF
AgCT		• 24-nt tetra-end-linked ssDNA	• discrimination of G-quadruplex structures	1
AgHX	MgSO ₄	• 23-nt and 25-nt ssDNA • 12-nt polyAC • 30-bp dsDNA • 27-nt ssDNA	• detection of single-base mismatch	2
			• quantification of base composition	3
			• discrimination ss. vs dsDNA	3
			• detection of chemically modified nucleobases	4
AgI	MgSO ₄	• 12-nt polyAC • 55-bp dsDNA • < 9-nt polyCT	• quantification of base composition	5
			• discrimination ss. vs dsDNA	5
			• identification of i-motif	6
Highly concentrated AgCT	/	• genomic dsDNA • genomic dsDNA	• detection of dehydration-induced conformational changes	7
			• detection of ROS-induced lesions	8
AgSp	/	• 21-bp dsDNA • 21-nt ssDNA, 21-bp dsDNA • 21-nt ssRNA, 21-bp dsRNA • 21-nt ssRNA, 21-bp dsRNA, 42-nt ssRNA • genomic dsDNA • 21-nt ssDNA, 21-bp dsDNA, genomic dsDNA • 141-nt ssDNA • ca. 200-bp dsDNA	• quantification of hybridization events	9
			• detection of chemically modified nucleobases	10
			• discrimination ss. vs dsRNA; detection of chemically modified nucleobases	11
			• discrimination of RNA conformations	11
			• interaction with exogenous agents	12
			• quantification of base composition	13
			• discrimination of point mutations	14
			• discrimination of strands	15

References

1. G. Rusciano, A. C. De Luca, G. Pesce, A. Sasso, G. Oliviero, J. Amato, N. Borbone, S. D'Errico, V. Piccialli, G. Piccialli and L. Mayol, *Anal. Chem.*, 2011, **83**, 6849-6855.
2. E. Papadopoulou and S. E. J. Bell, *Angew. Chem.-Int. Edit.*, 2011, **50**, 9058-9061.
3. E. Papadopoulou and S. E. J. Bell, *Chem.-Eur. J.*, 2012, **18**, 5394-5400.
4. S. Dick, S. E. J. Bell, K. J. Alexander, I. A. O'Neil and R. Cosstick, *Chem.-Eur. J.*, 2017, **23**, 10663-10669.
5. L.-J. Xu, Z.-C. Lei, J. Li, C. Zong, C. J. Yang and B. Ren, *J. Am. Chem. Soc.*, 2015, **137**, 5149-5154.
6. Y. Li, X. Han, Y. Yan, Y. Cao, X. Xiang, S. Wang, B. Zhao and X. Guo, *Anal. Chem.*, 2018, **90**, 2996-3000.
7. S. R. Panikkanvalappil, M. A. Mackey and M. A. El-Sayed, *J. Am. Chem. Soc.*, 2013, **135**, 4815-4821.
8. S. R. Panikkanvalappil, M. A. Mahmoud, M. A. Mackey and M. A. El-Sayed, *ACS Nano*, 2013, **7**, 7524-7533.
9. L. Guerrini, Ž. Krpetić, D. van Lierop, R. A. Alvarez-Puebla and D. Graham, *Angew. Chem.-Int. Edit.*, 2015, **54**, 1144-1148.
10. J. Morla-Folch, H.-n. Xie, P. Gisbert-Quilis, S. Gómez-de Pedro, N. Pazos-Perez, R. A. Alvarez-Puebla and L. Guerrini, *Angew. Chem.-Int. Edit.*, 2015, **54**, 13650-13654.
11. J. Morla-Folch, H.-n. Xie, R. A. Alvarez-Puebla and L. Guerrini, *ACS Nano*, 2016, **10**, 2834-2842.
12. M. Masetti, H.-n. Xie, Ž. Krpetić, M. Recanatini, R. A. Alvarez-Puebla and L. Guerrini, *J. Am. Chem. Soc.*, 2015, **137**, 469-476.
13. J. Morla-Folch, R. A. Alvarez-Puebla and L. Guerrini, *J. Phys. Chem. Lett.*, 2016, **7**, 3037-3041.
14. J. Morla-Folch, P. Gisbert-Quilis, M. Masetti, E. Garcia-Rico, R. A. Alvarez-Puebla and L. Guerrini, *Angew. Chem.-Int. Edit.*, 2017, **56**, 2381-2385.
15. J. Wang, K. M. Koo, E. J. H. Wee, Y. Wang and M. Trau, *Nanoscale*, 2017, **9**, 3496-3503.