Electronic Supplementary Information (ESI)

Dinuclear Zirconium Complex Bearing 1,5-Bridged-Calix[8]arene Ligand as Effective Catalyst for the Synthesis of Macrolactones

Rosita Lapenta,^a Nicola Alessandro De Simone,^a Antonio Buonerba,^{a,b,}* Carmen Talotta,^{a*} Carmine Gaeta,^a Placido Neri,^a Alfonso Grassi^{a,b} and Stefano Milione^{a,b}

- ^b Interuniversity Consortium Chemical Reactivity and Catalysis, CIRCC, via Celso Ulpiani 27, 70126 (BA), Italy.
- * Correspondence should be addressed to: A.B.: abuonerba@unisa.it; C.T.: ctalotta@unisa.it

^a Dipartimento di Chimica e Biologia, Università degli Studi di Salerno, via Giovanni Paolo II, 84084 Fisciano (SA), Italy

Table of Contents

1. NMR Characterization	S4
Figure S1. ¹ H NMR spectrum of C (600 MHz, pyridine- <i>d</i> ₅ , 25 °C).	S4
Figure S2. ¹ H- ¹ H COSY spectrum (600 MHz, pyridine- d_5 , 90°C) of C (a) with mag the diagnostic regions (b-d).	nifications of
Figure S3. ¹ H- ¹³ C HSQC spectrum (600 MHz, pyridine- d_5 , 25°C) of the complemagnifications of the diagnostic regions (b-d).	ex C (a) with
Figure S4. ¹ H- ¹ H NOESY spectrum (600 MHz, pyridine- d_5 , 90°C) of the complemagnifications of the diagnostic regions (b-d).	ex C (a) with
Figure S5. DEPT135 (a) and ¹³ C NMR (b) spectra (600 MHz, pyridine- d_5 , 90°C) of C (Scheme 1).	f the complex S8
Figure S6. DOSY NMR spectra of the complex C (600 MHz, 25°C, benzene- d_6) (600 MHz, 25°C, benzene- d_6) in presence of the internal standard S as reference (coefficients: C = $5.6 \cdot 10^{-8} \pm 4.6 \cdot 10^{-10} \text{ m}^2 \text{s}^{-1}$; S = $6.4 \cdot 10^{-8} \pm 3.9 \cdot 10^{-10} \text{ m}^2 \text{s}^{-1}$.	(a) and of C (b). Diffusion
Figure S7. Possible configurations of the ligand L with $C_{2\nu}$ symmetry (a-d) and th the complex C, compatible with NMR information, resulting from the configuration	e structure of in bS10
Figure S8. ¹ H- ¹ H COSY spectrum (600 MHz, benzene- d_6 , 25°C) of C' (a) with mag the diagnostic regions (b-d).	nifications ofS11
Figure S9. ¹ H- ¹³ C HSQC spectrum (600 MHz, benzene- d_6 , 25°C) of the complex magnifications of the diagnostic regions (b-d).	x C' (a) with
Figure S10. ¹³ C NMR spectrum of the complex C' with diagnostic signal labelle benzene- d_6 , 25°C)	d (600 MHz,
Figure S11. DOSY NMR spectra of the complex C' (a) and of C' in presence of standard S' as reference (b). Diffusion coefficients: $C = 4.30 \cdot 10^{-10} \pm 8.56 \cdot 10^{-13} \text{ m}^2 \text{s}^{-1}$; ${}^{10}\pm 5.68 \cdot 10^{-13} \text{ m}^2 \text{s}^{-1}$.	f the internal ; $S = 4.35 \cdot 10^{-10}$
Figure S12 . ¹ H NMR spectrum of cyclic PLA synthesized by C (entry 4 of Table 1 MHz, 25°C).	; CDCl ₃ , 300
Figure S13 . ¹ H NMR spectrum of linear PLA synthesized by C' (entry 9 of Table 1 MHz, 25°C).	; CDCl ₃ , 300
2. UV-Vis Analysis	S16
Figure S14. UV-Vis spectrum of C ($5.8 \cdot 10^{-3}$ M; pyridine; $25 \circ$ C; $\varepsilon_{436} = 166$ Lmol ⁻¹ c	em ⁻¹)S16
3. ESI-MS and MALDI-MS Analyses	S17
Figure S15. ESI-MS spectrum of the complex C (toluene/methanol solvents mixture	e)S17
Figure S16. ESI-MS spectrum of the complex C' (toluene/methanol solvents mixtur	e)S17
4. Kinetic Investigations	S18

1. NMR CHARACTERIZATION



Figure S1. ¹H NMR spectrum of C (600 MHz, pyridine-*d*₅, 25 °C).



Figure S2. ¹H-¹H COSY spectrum (600 MHz, pyridine- d_5 , 90°C) of C (a) with magnifications of the diagnostic regions (b-d).



Figure S3. ¹H-¹³C HSQC spectrum (600 MHz, pyridine- d_5 , 25°C) of the complex C (a) with magnifications of the diagnostic regions (b-d).



Figure S4. ¹H-¹H NOESY spectrum (600 MHz, pyridine- d_5 , 90°C) of the complex C (a) with magnifications of the diagnostic regions (b-d).



Figure S5. DEPT135 (a) and ¹³C NMR (b) spectra (600 MHz, pyridine- d_5 , 90°C) of the complex C (Scheme 1).



Figure S6. DOSY NMR spectra of the complex C (600 MHz, 25°C, benzene-*d*₆) (a) and of C (600 MHz, 25°C, benzene-*d*₆) in presence of the internal standard S as reference (b). Diffusion coefficients: $C = 5.6 \cdot 10^{-8} \pm 4.6 \cdot 10^{-10} \text{ m}^2 \text{s}^{-1}$; $S = 6.4 \cdot 10^{-8} \pm 3.9 \cdot 10^{-10} \text{ m}^2 \text{s}^{-1}$.



Figure S7. Possible configurations of the ligand L with $C_{2\nu}$ symmetry (a-d) and the structure of the complex C, compatible with NMR information, resulting from the configuration in b.



Figure S8. ¹H-¹H COSY spectrum (600 MHz, benzene- d_6 , 25°C) of C' (a) with magnifications of the diagnostic regions (b-d).



Figure S9. ¹H-¹³C HSQC spectrum (600 MHz, benzene- d_6 , 25°C) of the complex C' (a) with magnifications of the diagnostic regions (b-d).



Figure S10. ¹³C NMR spectrum of the complex C' with diagnostic signal labelled (600 MHz, benzene- d_6 , 25°C).



Figure S11. DOSY NMR spectra of the complex C' (a) and of C' in presence of the internal standard S' as reference (b). Diffusion coefficients: $C = 4.30 \cdot 10^{-10} \pm 8.56 \cdot 10^{-13} \text{ m}^2 \text{s}^{-1}$; $S = 4.35 \cdot 10^{-10} \pm 5.68 \cdot 10^{-13} \text{ m}^2 \text{s}^{-1}$.



Figure S12. ¹H NMR spectrum of cyclic PLA synthesized by C (entry **4** of Table 1; CDCl₃, 300 MHz, 25°C).



Figure S13. ¹H NMR spectrum of linear PLA synthesized by **C'** (entry **9** of Table 1; CDCl₃, 300 MHz, 25°C).

2. UV-VIS ANALYSIS



Figure S14. UV-Vis spectrum of C (5.8·10⁻³ M; pyridine; 25 °C; $\epsilon_{436} = 166 \text{ Lmol}^{-1}\text{cm}^{-1}$).

3. ESI-MS AND MALDI-MS ANALYSES



Figure S15. ESI-MS spectrum of the complex C (toluene/methanol solvents mixture).



Figure S16. ESI-MS spectrum of the complex C' (toluene/methanol solvents mixture).

4. Kinetic Investigations



Figure S17. Polymerization of *rac*-LA catalyzed by C (a) with the corresponding plot of $ln([LA]_0/[LA])$ versus time.



Figure S18. Plot of number-averaged molecular weights $M_{n(exp)}$ (square) vs monomer to initiator ratio with theoretical molecular weights $M_{n(th)}$ (dots) for LA polymerization catalyzed by C (reaction conditions in Table 1).



Figure S19. Polymerization of *rac*-LA catalyzed by C' (a) with the corresponding plot of $ln([LA]_0/[LA])$ versus time.

Entry	[C]	[<i>rac</i> -LA]/[C]	r
	(M)	(molar ratio)	(M s ⁻¹)
S1	1.42.10-2	50	2.7·10 ⁻³ ±1.5·10 ⁻⁴
S2	7.07.10-3	100	$1.7 \cdot 10^{-3} \pm 1.3 \cdot 10^{-4}$
S3	$4.72 \cdot 10^{-3}$	150	$1.2 \cdot 10^{-3} \pm 6.3 \cdot 10^{-3}$
S4	3.53.10-3	200	7.3·10 ⁻⁴ ±3.7·10 ⁻⁵

Table S1. Reaction rate as a function of catalyst concentration for ROP of LA with C.

Reaction conditions: [rac-LA] = 0.706 M, TCE- $d_2 = 0.6$ mL, T = 80 °C; reactions carried out inside NMR tubes and monitored with interval of one minute.



5. Gel Permeation Chromatography.

Figure S20. Gel permeation chromatogram of the polymer sample from entry 1 of Table 1.



Figure S21. Gel permeation chromatogram of the polymer sample from entry 2 of Table 1.



Figure S22. Gel permeation chromatogram of the polymer sample from entry 3 of Table 1.



Figure S23. Gel permeation chromatogram of the polymer sample from entry 4 of Table 1.



Figure S24. Gel permeation chromatogram of the polymer sample from entry 5 of Table 1.



Figure S25. Gel permeation chromatogram of the polymer sample from entry 6 of Table 1.



Figure S26. Gel permeation chromatogram of the polymer sample from entry 7 of Table 1.



Figure S27. Gel permeation chromatogram of the polymer sample from entry 8 of Table 1.



Figure S28. Gel permeation chromatogram of the polymer sample from entry 9 of Table 1.



Figure S29. Gel permeation chromatogram of the polymer sample from entry 10 of Table 1.



Figure S30. Gel permeation chromatogram of the polymer sample from entry 11 of Table 1.