Electronic Supplementary Information

A Simple Quinolone Schiff-Base Containing CHEF Based Fluorescence 'turn-on' Chemosensor for Distinguishing Zn²⁺ and Hg²⁺ with High Sensitivity, Selectivity and Reversible

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Figure S1. IR spectra of chemosensor MQA and complexes MQA-Zn²⁺, MQA-Hg²⁺ in KBr disks.



Figure S2. ¹H NMR spectra of chemosensor MQA and complexes MQA-Zn²⁺, MQA-Hg²⁺ in d_6 -DMSO.



Figure S3. ¹³C NMR spectra of chemosensor MQA and complexes MQA-Zn²⁺, MQA-Hg²⁺ in d_6 -DMSO.



Figure S4. ESI-MS spectra of (a) MQA, (b) MQA-Z n^{2+} and (c) MQA-H g^{2+} .



Figure S5. Thermal gravimetric curve of chemosensor MQA.

Association constant (K_a) calculation

Assuming a 1:1 complex formation, the association constant was calculated on the basis of the titration curves of the chemosensor **MQA** with M^{2+} (Zn²⁺/Hg²⁺). The association constant was calculated according to the Benesi–Hildebrand equation (1):

$$\frac{1}{I - I_0} = \frac{1}{K_a (I_{max} - I_0) [M]^n} + \frac{1}{I_{max} - I_0}$$
⁽¹⁾

Where K_a is complex association constant, I_0 is the fluorescent intensity of **MQA** in the absence of M^{2+} , I is the fluorescent intensity recorded in the presence of added M^{2+} , I_{max} is the fluorescent intensity in presence of added $[M^{2+}]_{max}$, and n is the binding stoichiometry ratio between **MQA** and M^{2+} (Zn²⁺/Hg²⁺). The association constant (K_a) could be determined from the slope of the straight line of the plot of $1/(I - I_0)$ against $1/[M]^n$.



Figure S6. (a) Benesi-Hildebrand equation plot (fluorescence intensity at 565 nm) of **MQA**, assuming 1:1 stoichiometry for association between **MQA** and Zn²⁺. (b) Benesi-Hildebrand equation plot (fluorescence intensity at 530 nm) of **MQA**, assuming 1:1 stoichiometry for

association between MQA and Hg^{2+} .

Standard deviation and detection limit calculation

The detection limit was calculated based on the fluorescence titration. To determine the S/N ratio, fluorescence intensity of **MQA** without M^{2+} (Zn²⁺/Hg²⁺) was measured by five times and the standard deviation of blank measurements was determined. To gain the slop, the fluorescence intensity data at 565 nm and 530 nm for Zn²⁺ and Hg²⁺, respectively, were plotted as a concentration of M²⁺. So the detection limit was calculated with the follow equation (2):

Detection limit =
$$3\sigma/m$$
 (2)

Where σ is the standard deviation of blank measurements, and m is the slop of fluorescence versus M^{2+} concentration.

	Fluorescence intensity
Test 1	28.20
Test 2	28.68
Test 3	27.09
Test 4	30.40
Test 5	29.85
Standard Deviation (σ)	1.32

Table S1. Standard deviation calculation.

Table S2. Detection limit calculation.

	Detection Zn ²⁺	Detection Hg ²⁺
Slope (m)	117.79 μM ⁻¹	32.65 μM ⁻¹
Detection limit $(3\sigma/m)$	0.011 µM	0.040 μM



Figure S7. (a) Plot of fluorescence intensity of MQA with Zn²⁺ at 565 nm. (b) Plot of fluorescence intensity of MQA with Hg²⁺ at 530 nm.



Figure S8. Fluorescence spectra of MQA (10 μ M) with Hg²⁺ (1.0 equiv.) upon titration of Zn²⁺ (0.2, 0.4, 0.6, 0.8 and 1.0 equiv.).



Figure S9. (a) Fluorescence spectra and (b) fluorescence intensity change of Zn²⁺-bound **MQA** in DMSO/water mixture (1/99 v/v) in the presence other metal ions. (c) Fluorescence spectra and (d) fluorescence intensity change of Hg²⁺-bound **MQA** in DMSO/water mixture (1/99 v/v) in the presence other metal ions.



Figure S10. (a) Fluorescence spectra and (b) fluorescence intensity change of Zn²⁺-bound **MQA** in DMSO/water mixture (1/99 v/v) with different anions.



Figure S11. (a) Fluorescence spectra and (b) fluorescence intensity change of Hg²⁺-bound **MQA** in DMSO/water mixture (1/99 v/v) with different anions.



None Glycine Cysteine Alanine Proline Serine Tyrosine







Figure S13. Fluorescence response ($I_{530 \text{ nm}}$) of Hg²⁺ sensing by **MQA** (10 µM) with various amino acids (2.0 equiv.) in DMSO/water mixture (1/99 v/v). The black bars represent the addition 2.0 equiv. of the various amino acids to a 10 µM solution of **MQA**. The red bars represent the change of the emission that occurs upon the subsequent addition of 1.0 equiv. Hg²⁺ to the above solution.



Figure S14. Changes in fluorescence intensity of MQA (10 μ M) measured with Zn²⁺ (1.0 equiv.) as a function of pH at room temperature.



Figure S15. Changes in fluorescence intensity of MQA (10 μ M) measured with Hg²⁺ (1.0 equiv.) as a function of pH at room temperature.



Figure S16. (a) Fluorescence spectral changes of MQA (10 μ M) after the sequential addition of Hg²⁺ and EDTA in buffer solution (bis–tris, pH 7.0). (b) Reversible changes in fluorescence intensity and (c) luminescent photos of MQA (excitation under a 365 nm UV lamp) after the sequential addition of Hg²⁺ and EDTA.



Figure S17. Experimental (top) and simulated (bottom) PXRD patterns of complexes MQA-Zn²⁺ (a) and MQA-Hg²⁺ (b).



Figure S18. Depiction of the dihedral angle between the phenyl ring and quinolone ring in complexes MQA-Zn²⁺ (a) and MQA-Hg²⁺ (b).

Chemosensor	No. of steps for synthesis	Detection Limit (µM)	Binding constant (M ⁻¹)	Percent of water in solution	Reference
	1	2.5	$1.9 \times 10^5 (Zn1)$	100%	[S1]
			1.5×10^4 (Zn2)		
			6.1×10^3 (Zn3)		
N N					
	5	0.0719	1.05×10^{6}	0 (100% methanol)	[S2]
	4	2.2	4×104	0 (5% acatonitrila)	[\$2]
	4	5.2	$4 \times 10^{\circ}$	0(3% acetomume)	[33]
0	3	0.066	$1.4 imes 10^{6}$	0 (100% acetonitrile)	[S4]
	1	65.41	$4.8 imes 10^5$	50%	[S5]
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 Table S3. Comparison of a few aspects of some recently published Zn-sensors.



Chemosensor	No. of steps for synthesis	Detection Limit (µM)	Binding constant (M ⁻¹ )	Percent of water in solution	Reference
	1	3.1	No data	0 (100% ethanol)	[S11]
N- PF6	2	0.0718	No data	100%	[S12]
HC=N-	1	0.05595	$9.35 \times 10^4$	0 (100% DMSO)	[S13]
N N N N N N N N N N N S	3	No data	1.02 × 10 ⁶	0 (40% ethanol)	[S14]

Table S4. Comparison of	a few aspects of some	recently published Hg	-sensors.
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	MQA-Zn ²⁺	MQA-Hg ²⁺
CCDC No.	1460774	1460775
formula	$C_{17}H_{14}Cl_2N_2OZn$	C ₁₇ H ₁₄ Cl ₂ N ₂ OHg
Mr	398.59	533.79
A cryst syst	Monoclinic	Triclinic
space group	$P2_1/c$	Р
a [Å]	7.492(18)	8.108(12)
b [Å]	15.272(4)	8.616(12)
	15.500(4)	12.299(18)
α ^[°]	90	77.039(4)
β[°]	112.715(11)	87.533(4)
γ [°]	90	82.928(4)
Volume [Å ³ ]	1636.0(7)	830.9(2)
Z	4	2
$D_{\rm c} [\rm g \cdot \rm cm^{-3}]$	1.618	2.134
$\mu \text{[mm}^{-1}\text{]}$	1.832	9.588
F(000)	808	504
Θ range [°]	1.95 - 25.00	1.70 - 25.00
<i>h</i> range	$-8 \le h \le 8$	$-9 \le h \le 9$
k range	$-16 \le k \le 18$	$-10 \le k \le 8$
<i>l</i> range	$-18 \le l \le 18$	$-14 \le l \le 14$
data/restraints/params	2829 / 0 / 208	2920 / 0 / 209
GOF	0.844	0.909
$R_1, wR_2[I > 2\sigma(I)]^a$	0.0549, 0.1011	0.0452, 0.0766
$R_1$ , $wR_2$ [all data] ^a	0.1639, 0.1268	0.0664, 0.0840
$\Delta \rho_{\rm max},  \Delta \rho_{\rm min}  [{\rm e} \cdot {\rm \AA}^{-3}]$	0.523, -0.670	0.930, -1.280

**Table S5.** Crystallographic and structural determination data for complexes  $MQA-Zn^{2+}$  and  $MQA-Hg^{2+}$ .

 $\overline{a_{R_1} = \sum ||F_o| - |F_c|| / \sum |F_o|}; wR_2 = \left[\sum [w (F_o^2 - F_c^2)^2] / \sum [w (F_o^2)^2]\right]^{1/2}.$ 

Table S6. Selected bond distances (Å) and angles (°) for  $MQA-Zn^{2+}$  and  $MQA-Hg^{2+}$ .

Parameter	MQA-Zn ²⁺	MQA-Hg ²⁺
M(1)-N(1)	2.135(5)	2.397(7)
M(1)-N(2)	2.073(5)	2.359(7)
M(1)-Cl(1)	2.226(2)	2.384(3)
M(1)-Cl(2)	2.218(2)	2.419(3)
M(1)-O(1)	2.393(4)	2.668(6)
N(2)-C(10)	1.296(8)	1.261(11)
N(2)-M(1)-N(1)	78.6(2)	71.7(2)
N(1)-M(1)-Cl(1)	108.40(15)	111.12(18)
N(2)-M(1)-Cl(1)	110.51(15)	131.60(18)
N(1)-M(1)-Cl(2)	104.57(15)	107.20(18)
N(2)-M(1)-Cl(2)	128.33(16)	101.94(18)
Cl(2)-M(1)-Cl(1)	116.56(8)	121.09(10)
N(1)-M(1)-O(1)	147.08(19)	129.2(2)
N(2)-M(1)-O(1)	70.08(18)	62.8(2)
Cl(1)-M(1)-O(1)	92.32(13)	85.32(16)
Cl(2)-M(1)-O(1)	87.57(12)	103.11(16)
C(12)-O(1)-M(1)	114.1(4)	115.9(5)

C(17)-O(1)-M(1)	125.3(4)	123.4(5)
C(1)-N(1)-M(1)	112.2(4)	112.7(6)
C(5)-N(1)-M(1)	129.8(5)	126.3(6)
C(10)-N(2)-M(1)	115.2(4)	114.3(6)
C(11)-N(2)-M(1)	122.8(4)	122.7(6)

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