

**ESI for**

**Trinuclear complexes of palladium(II) with chalcogenated *N*-heterocyclic carbenes: catalysis of selective nitrile-primary amide interconversion and Sonogashira coupling**

Pooja Dubey, Sonu Gupta, and Ajai K. Singh\*

Department of Chemistry, Indian Institute of Technology, Delhi, New Delhi 110016, India

**Content**

<b>Table S1.</b>	Crystal Data and Structural Refinement Parameters of 1-2	2
<b>Table S2.</b>	Selected Bond Lengths and Bond Angles of Complex 1-2	3
<b>Table S3.</b>	Distances [ $\text{\AA}$ ] of inter and intra-molecular interactions	4
<b>Figure S1–S68.</b>	NMR Spectra	4-38
<b>Figure S69–S72.</b>	Mass Spectrum	38-41
<b>NMR Data.</b>		41-45
<b>Figure S73-76.</b>	Mass Spectrum	45-46
<b>References.</b>		47-48

**Table S1.** Crystal data and structural refinement parameters

Compounds	1	2
Empirical formula	C40 H44 Cl6 N8 Pd3 S2	C44 H50 Cl6 N10 Pd3 Se2
Formula wt.	1232.85	1408.76
Crystal size [mm]	0.33 x 0.32 x 0.29	0.33 x 0.32 x 0.27
Crystal system	Monoclinic	Triclinic
Space group	P21/n	P-1
Unit Cell dimension	a = 11.797(3)Å b = 10.941(3)Å c = 19.404(6)Å $\alpha = 90^\circ$ $\beta = 97.804(6)^\circ$ $\gamma = 90^\circ$	a = 8.380(2)Å b = 11.721(3)Å c = 14.883(4)Å $\alpha = 67.79(2)^\circ$ $\beta = 81.88(3)^\circ$ $\gamma = 82.02(2)^\circ$
Volume [Å <sup>3</sup> ]	2481.3(12)	1334.0(6)
Z	2	1
Density (Calc.) [Mg.m <sup>-3</sup> ]	1.650	1.754
Absorption coeff. [mm <sup>-1</sup> ]	1.520	2.706
F(000)	1224.0	692.0
$\theta$ range [°]	1.91–25.00	1.485– 24.979
Index ranges	-14<=h<=14-13<=k<= 13- 23<=l<=23	-9<=h<=9-13<=k<= 13-17<=l<=17
Reflections collected	23279	12922
Independent reflections (Rint.)	4358 (0.0831)	4672 (0.1111)
Max./min. Transmission	0.646/0.610	0.495/ 0.436
Data/restraints/parameters	4353/0/271	4632/0/299
Goodnessofit on F2	1.091	0.661
Final R indices [ I > 2 $\sigma$ (I)]	R1 = 0.0632, wR2 = 0.1383	R1 = 0.0391, wR2 = 0.0730
R indices (all data)	R1 = 0.0978, wR2 = 0.1523	R1 = 0.0499, wR2 = 0.0764
Largest diff. peak/hole [e.Å <sup>-3</sup> ]	0.992/-0.651	0.909/-0.980
CCDC no	1523310	1569993

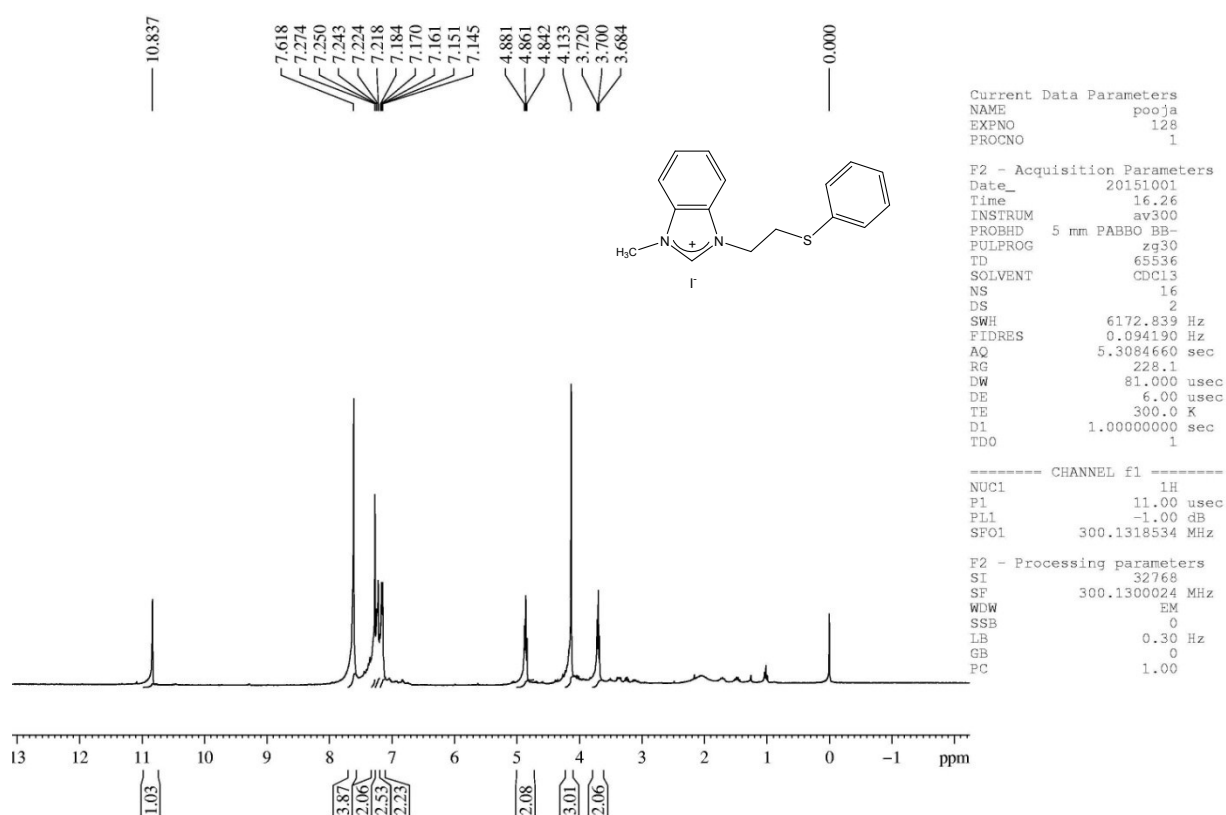
**Table S2.** Selected bond lengths [ $\text{\AA}$ ] and bond angles [ $^\circ$ ]

	Bond length [ $\text{\AA}$ ]	Bond angle [ $^\circ$ ]
<b>1</b>	Pd(1)—S(1) 2.3179(19) Pd(1)—Cl(1) 2.2784(19)  Pd(2)—C(9) 1.968(7) Pd(2)—Cl(2) 2.298(2) Pd(2)—Cl(3) 2.285(2) Pd(2)—N(3) 2.073(8)  S(1)—C(1) 1.775(8) S(1)—C(7) 1.816(7)  N(1)—C(9) 1.317(9) N(1)—C(8) 1.470(9) N(1)—C(10) 1.393(9)  N(2)—C(9) 1.329(9) N(2)—C(15) 1.387(9) N(2)—C(16) 1.461(10)	Cl(1)—Pd(1)—S(1) 94.80(7) Cl(1)—Pd(1)—S(1) 85.20(7) Cl(1)—Pd(1)—Cl(1) 180.00 S(1)—Pd(1)—S(1) 180.00 Cl(2)—Pd(2)—N(3) 90.4(2) Cl(3)—Pd(2)—N(3) 93.9(2)  C(9)—Pd(2)—N(3) 178.4(3) C(9)—Pd(2)—Cl(2) 88.0(2) C(9)—Pd(2)—Cl(3) 87.7(2)  Cl(3)—Pd(2)—Cl(2) 175.70(9) C(1)—S(1)—Pd(1) 102.5(2) C(7)—S(1)—Pd(1) 110.7(3)  N(1)—C(9)—Pd(2) 126.3(6) N(2)—C(9)—Pd(2) 123.9(6)  C(9)—N(1)—C(8) 124.8(6) C(9)—N(1)—C(10) 109.1(6) C(9)—N(2)—C(15) 108.3(7) C(9)—N(2)—C(16) 126.4(7)
<b>2</b>	Pd(1)—Se(1) 2.4312(10) Pd(1)—Cl(1) 2.2971(12)  Pd(2)—C(9) 1.952(4) Pd(2)—Cl(2) 2.2940(15) Pd(2)—Cl(3) 2.3003(15) Pd(2)—N(3) 2.079(4)  Se(1)—C(1) 1.924(4) Se(1)—C(7) 1.960(4)  N(1)—C(9) 1.341(5) N(1)—C(8) 1.462(5) N(1)—C(10) 1.394(5)  N(2)—C(9) 1.353(5) N(2)—C(15) 1.399(5) N(2)—C(16) 1.460(5)	Cl(1)—Pd(1)—Se(1) 83.50(4) Cl(1)—Pd(1)—Se(1) 96.50(4) Cl(1)—Pd(1)—Cl(1) 180.00(1) Se(1)—Pd(1)—Se(1) 180.00(2) Cl(2)—Pd(2)—N(3) 90.16(12) Cl(3)—Pd(2)—N(3) 91.20(12) Cl(3)—Pd(2)—Cl(2) 178.64(4)  C(9)—Pd(2)—N(3) 176.42(18) C(9)—Pd(2)—Cl(2) 87.67(13) C(9)—Pd(2)—Cl(3) 90.98(13)  C(1)—Se(1)—Pd(1) 97.27(13) C(7)—S(1)—Pd(1) 104.83(14)  N(1)—C(9)—Pd(2) 125.4(3) N(2)—C(9)—Pd(2) 127.4(3) C(9)—N(1)—C(8) 124.5(3) C(9)—N(1)—C(10) 110.2(3) C(9)—N(2)—C(15) 109.6(3) C(9)—N(2)—C(16) 125.2(4)

**Table S3.** Distances [Å] of inter and intra-molecular interactions for **1** and **2**

Complex 1		Complex 2	
C11-H16	2.702	C11-H20A	2.925
C11-H14	2.939	N4-H16A	2.669

**NMR Spectra of L1-L2 and 1-2**



**Figure S1.** <sup>1</sup>H NMR of benzimidazolium salt, L1

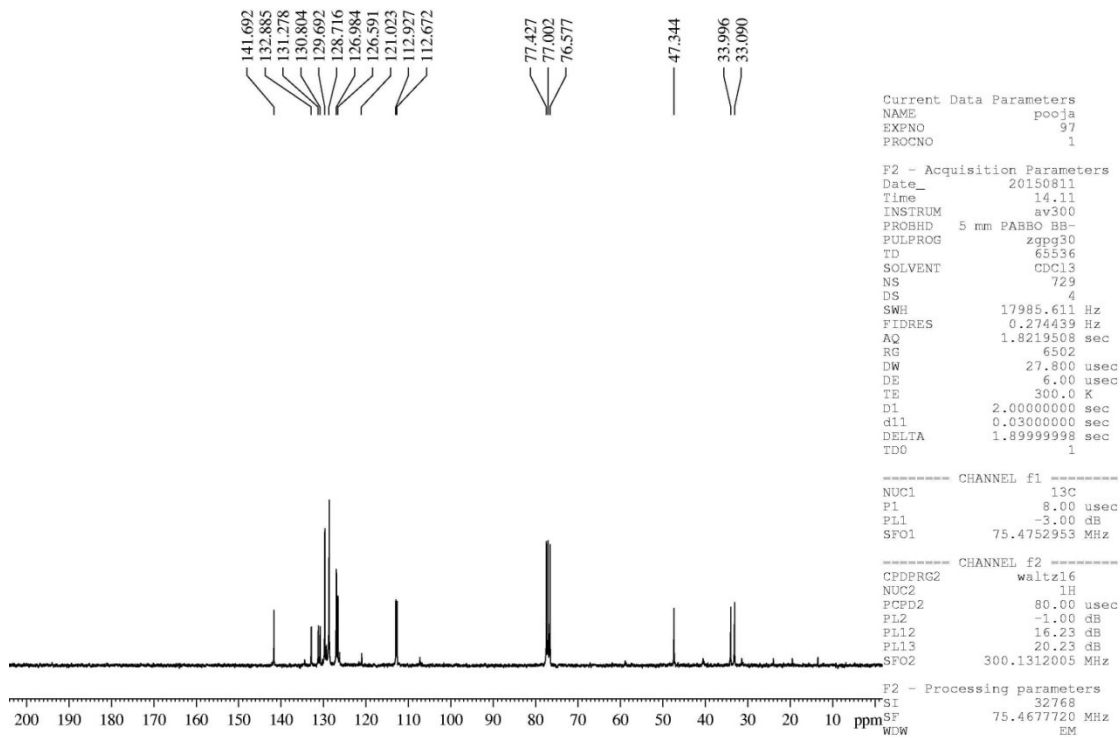


Figure S2.  $^{13}\text{C}\{^1\text{H}\}$  NMR of benzimidazolium salt, L1

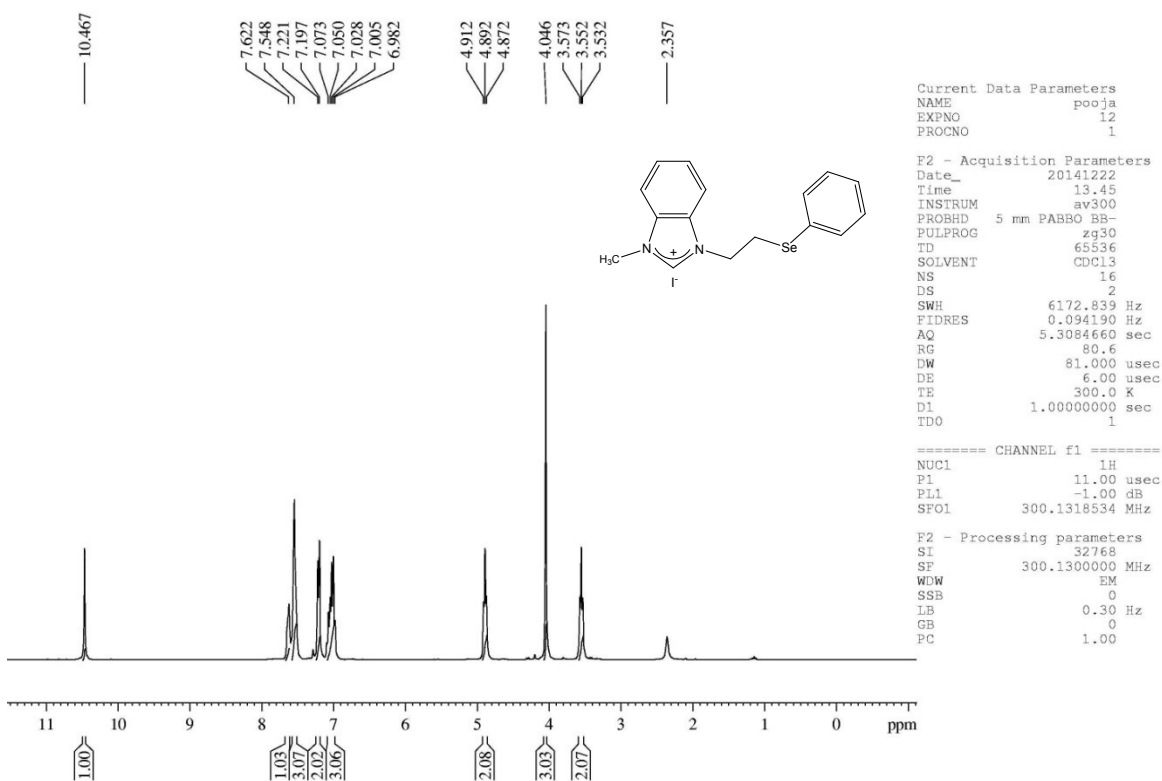


Figure S3.  $^1\text{H}$  NMR of benzimidazolium salt, L2

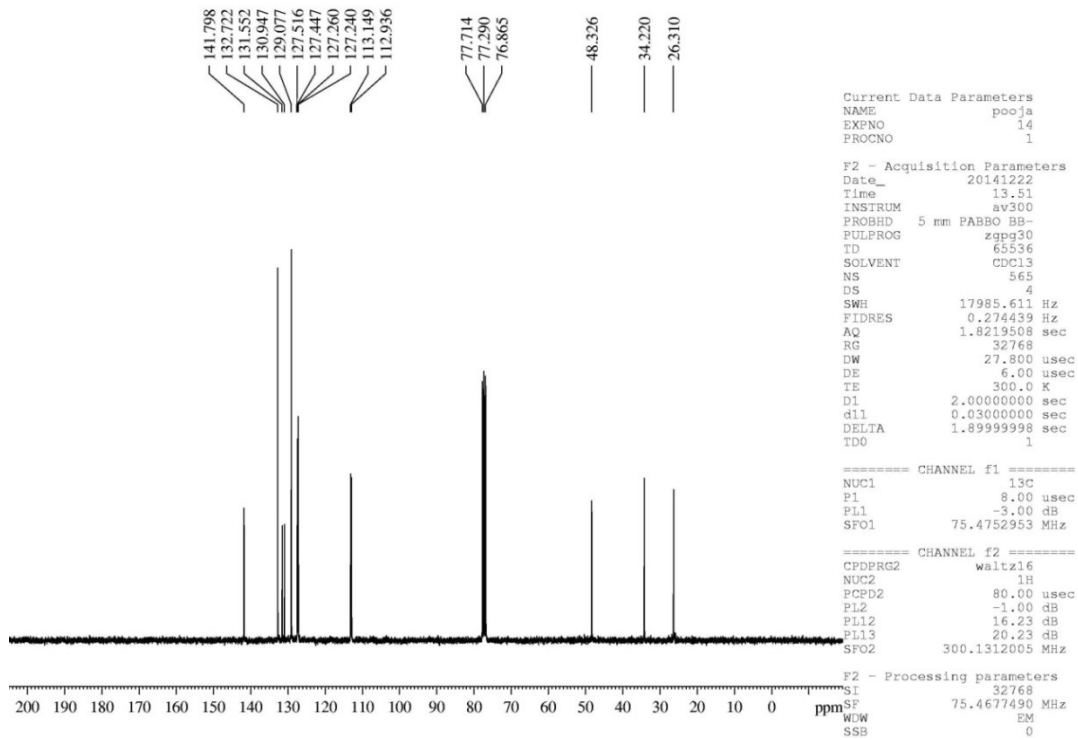


Figure S4.  $^{13}\text{C}\{^1\text{H}\}$  NMR of benzimidazolium salt, L2

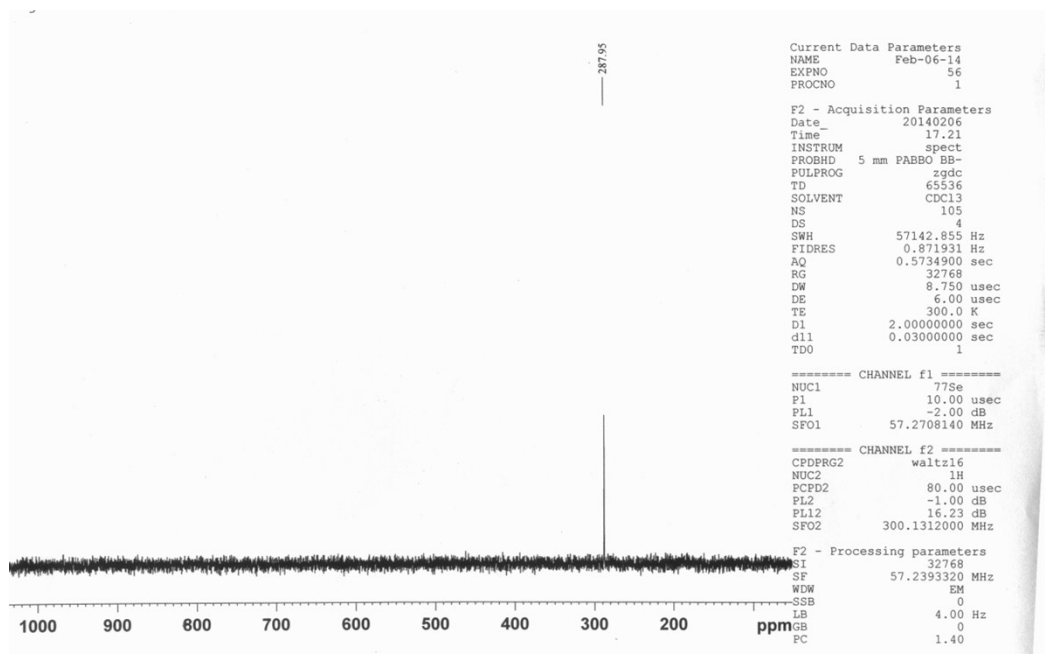


Figure S5.  $^{77}\text{Se}$  NMR of benzimidazolium salt, L2

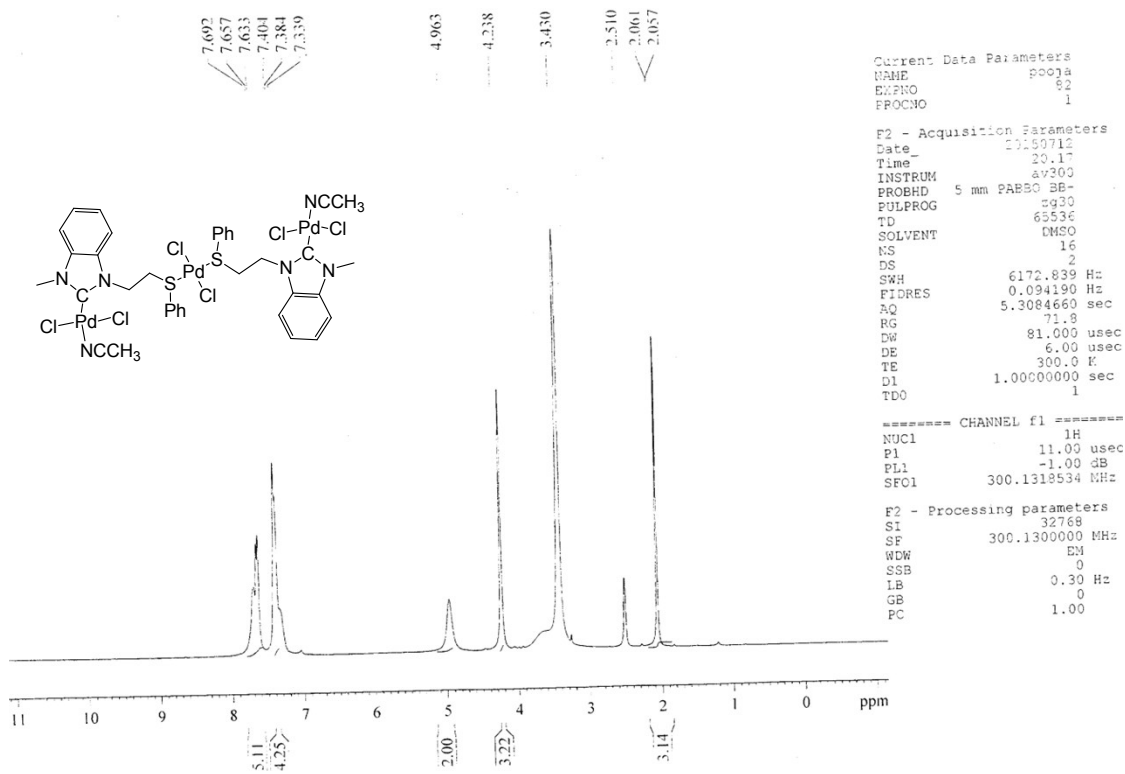


Figure S6.  $^1\text{H}$  NMR of complex 1.

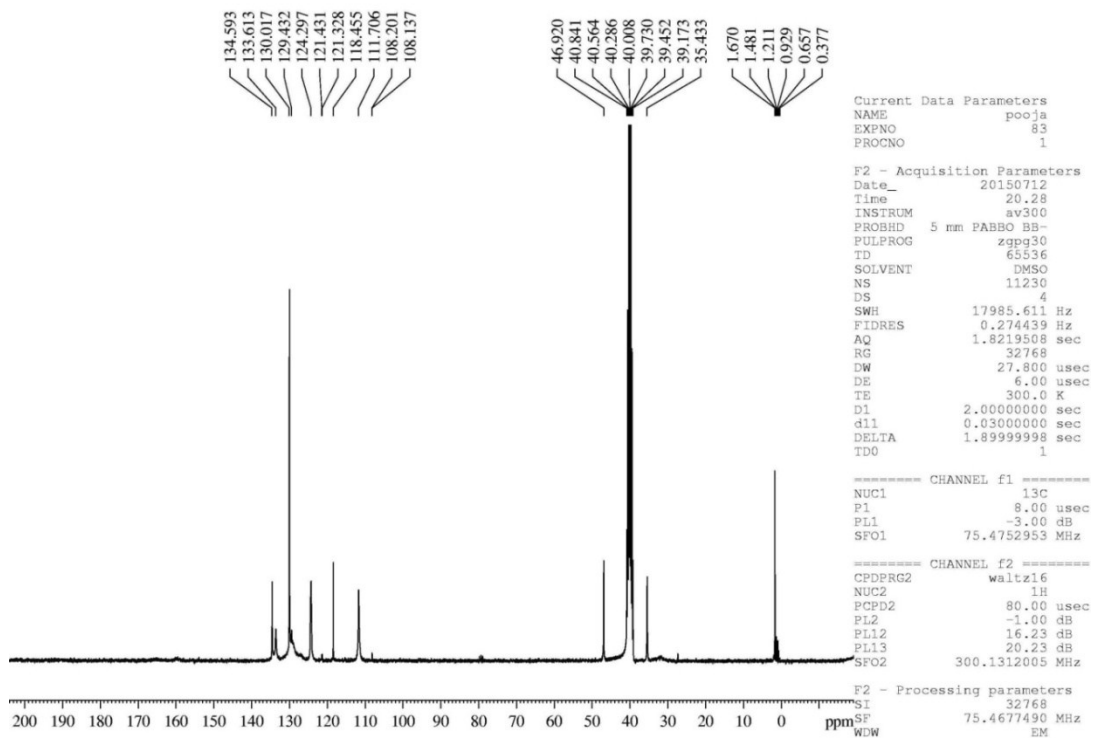


Figure S7.  $^{13}\text{C}\{^1\text{H}\}$  NMR of complex 1

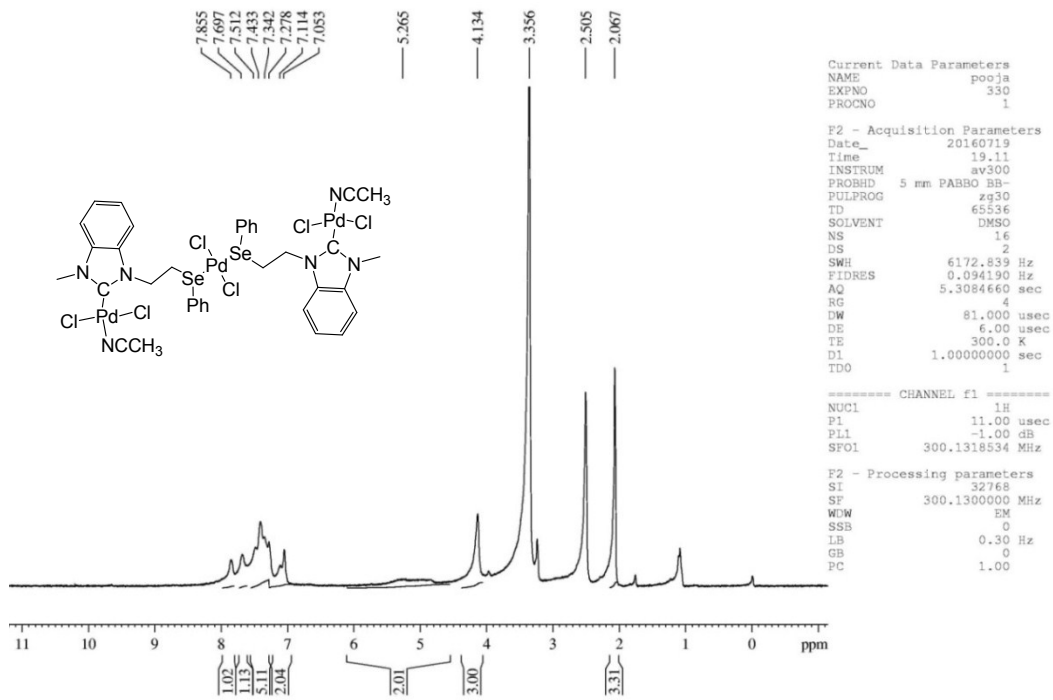


Figure S8.  $^1\text{H}$  NMR of complex 2.

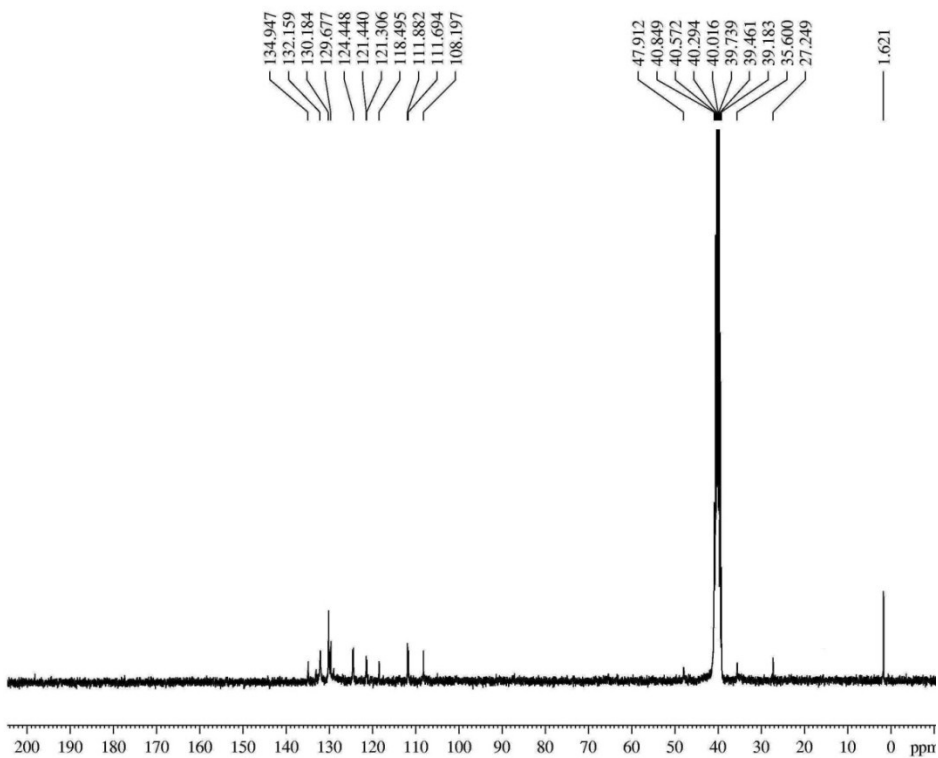


Figure S9.  $^{13}\text{C}\{^1\text{H}\}$  NMR of complex 2 [ $^{13}\text{C}$  NMR (75 MHz, solvent  $\text{d}_6$ -DMSO)]



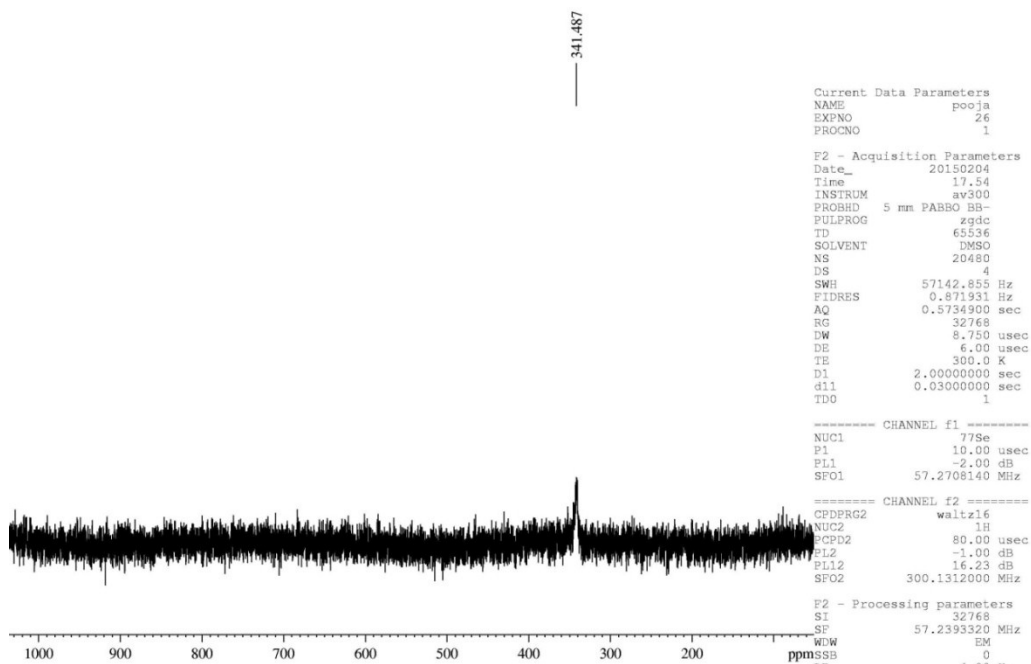


Figure S10.  $^{77}\text{Se}\{^1\text{H}\}$  NMR of complex 2

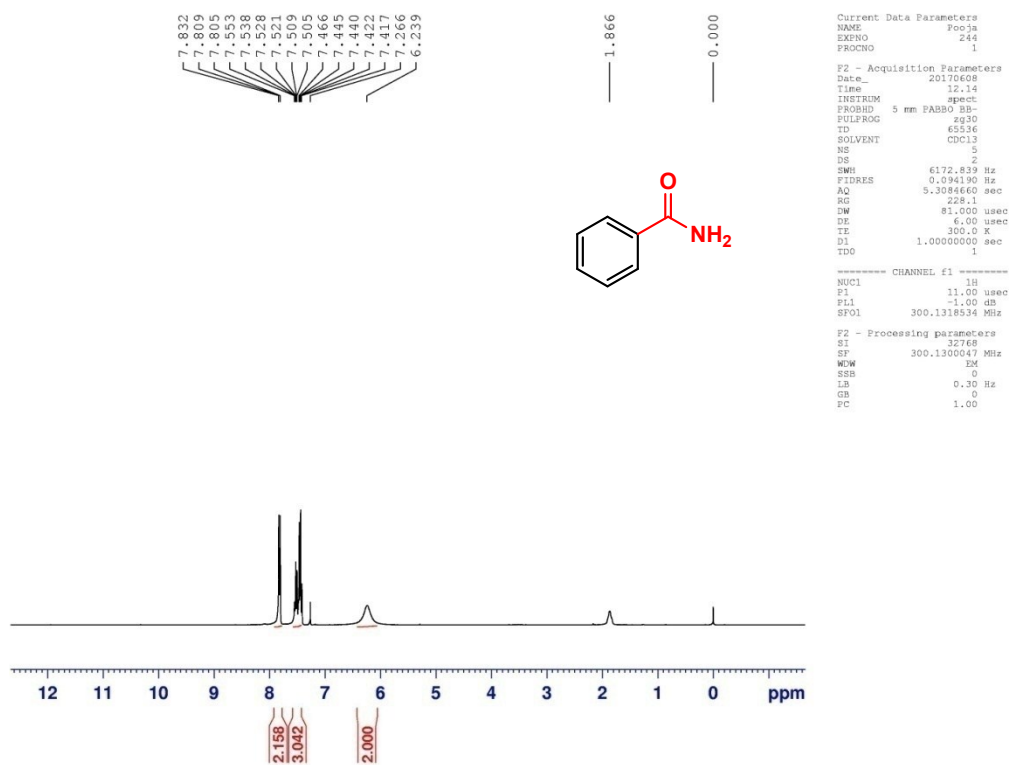


Figure S11.  $^1\text{H}$  NMR of 4a

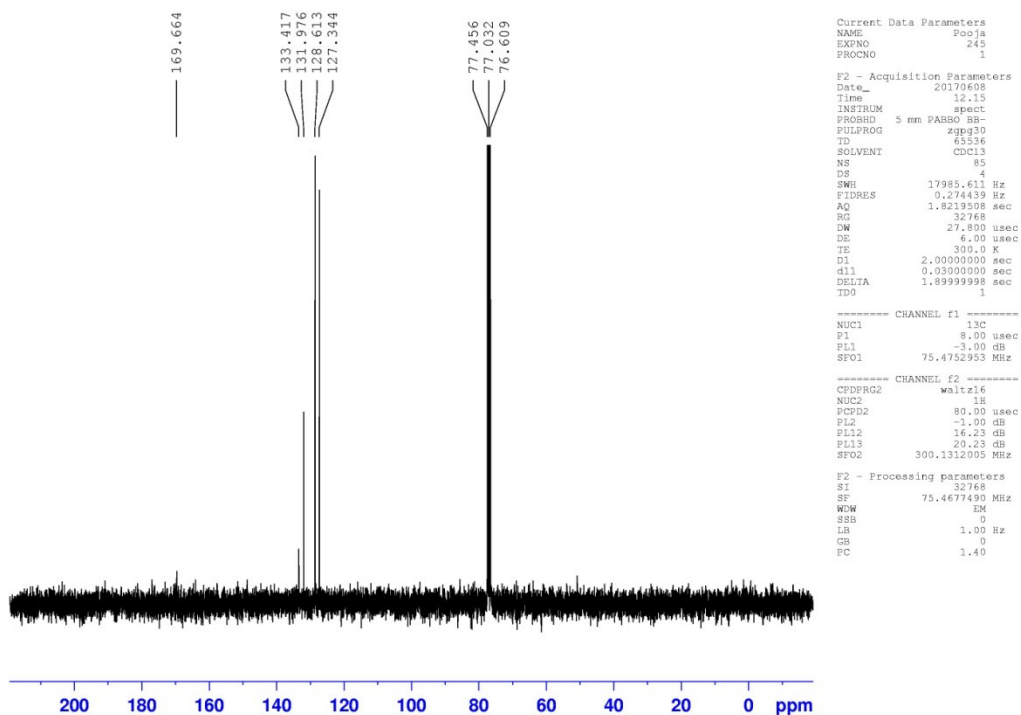


Figure S12. <sup>13</sup>C NMR of 4a

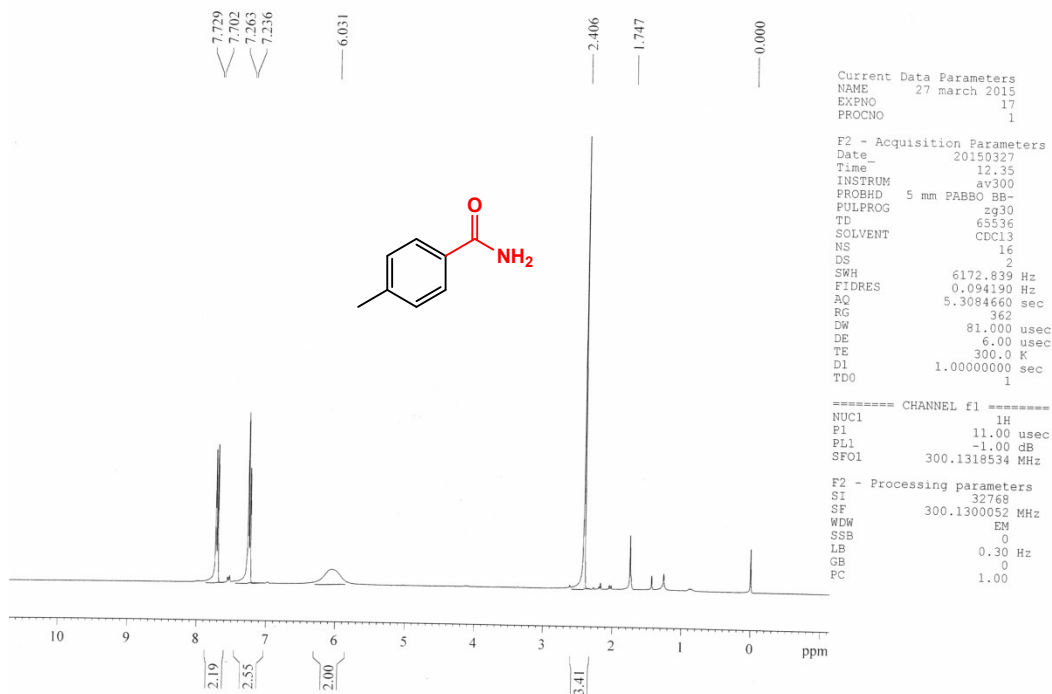


Figure S13. <sup>1</sup>H NMR of 4b

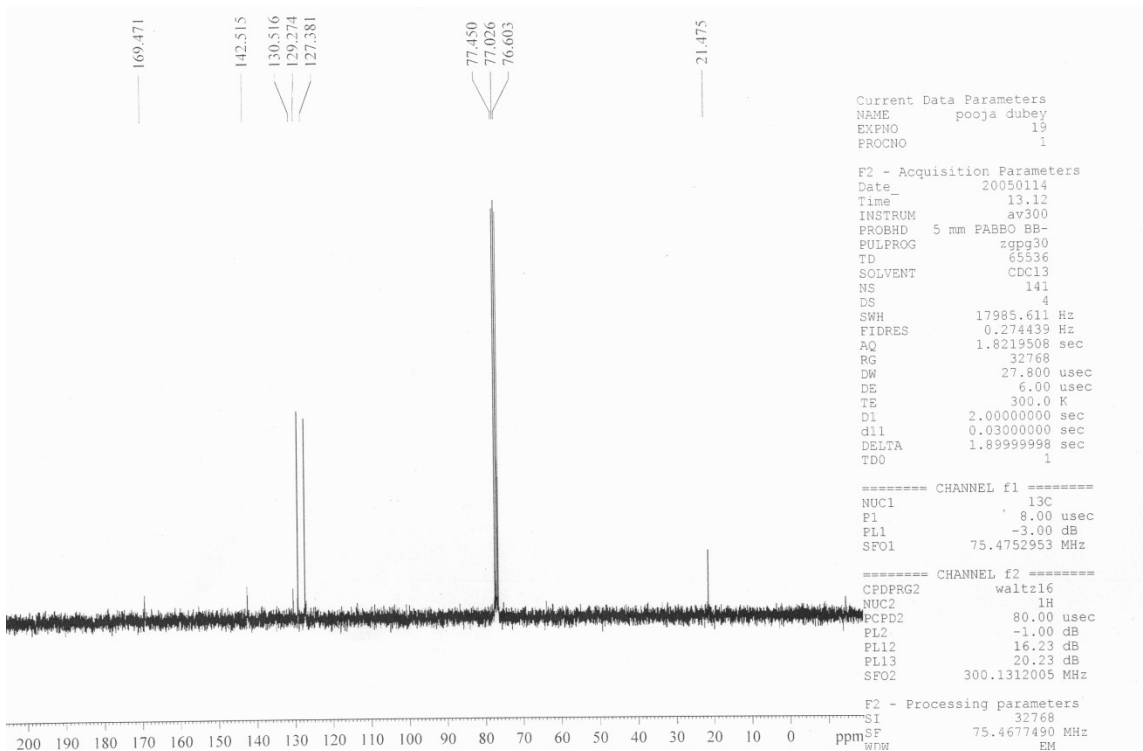


Figure S14. <sup>13</sup>C NMR of 4b

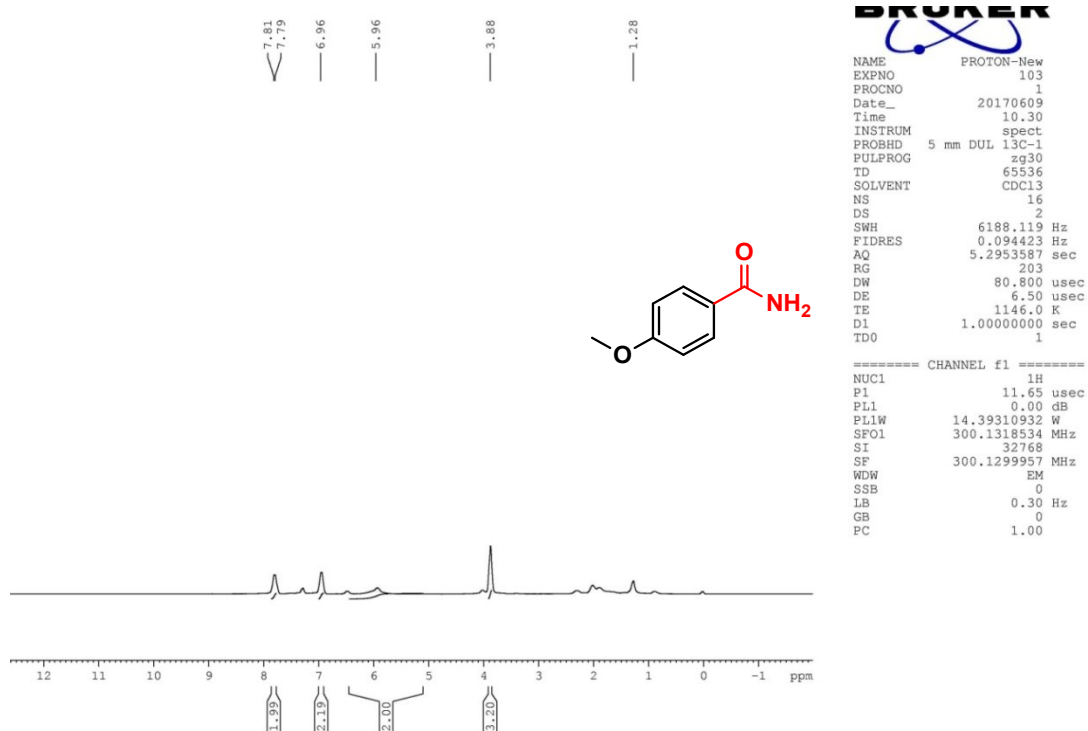


Figure S15. <sup>1</sup>H NMR of 4c

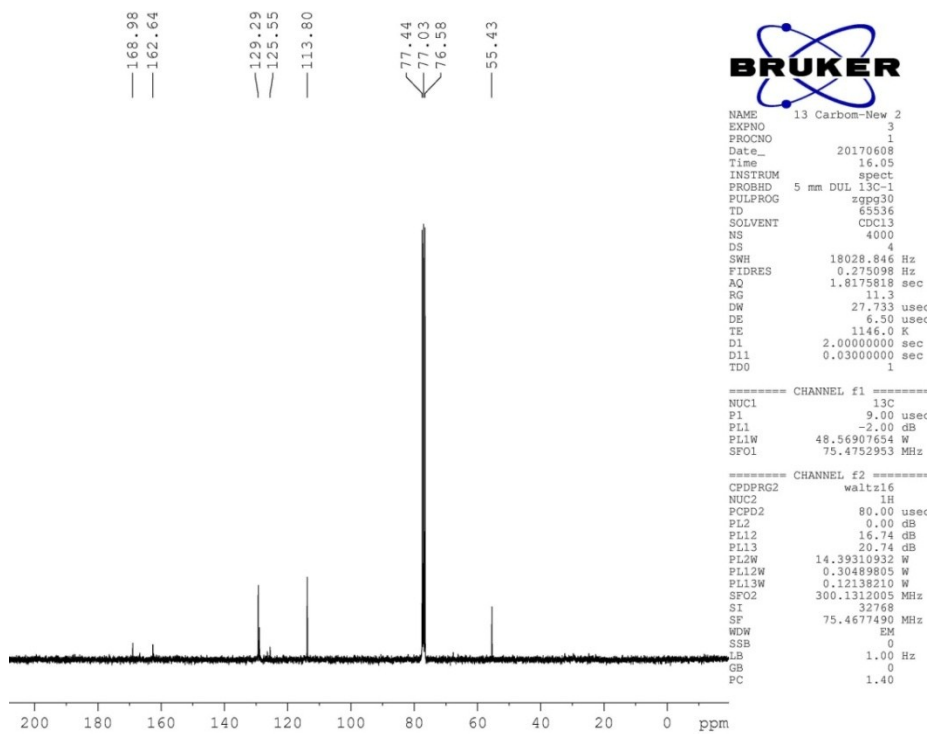


Figure S16. <sup>13</sup>C NMR of 4c

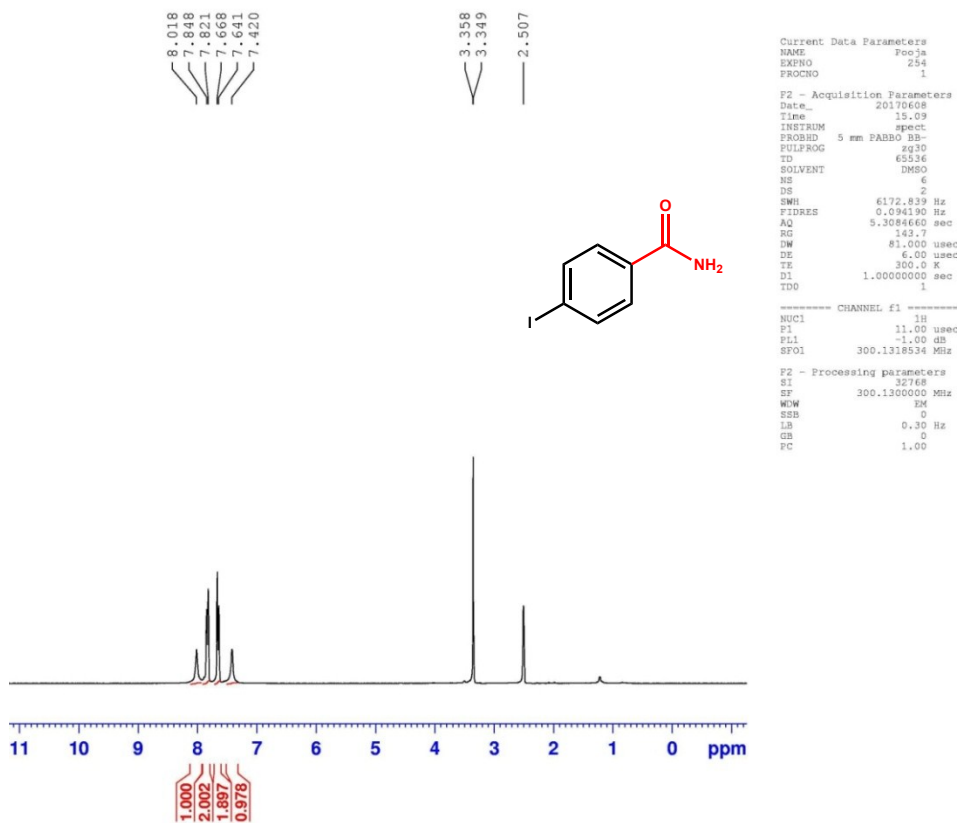


Figure S17. <sup>1</sup>H NMR of 4d

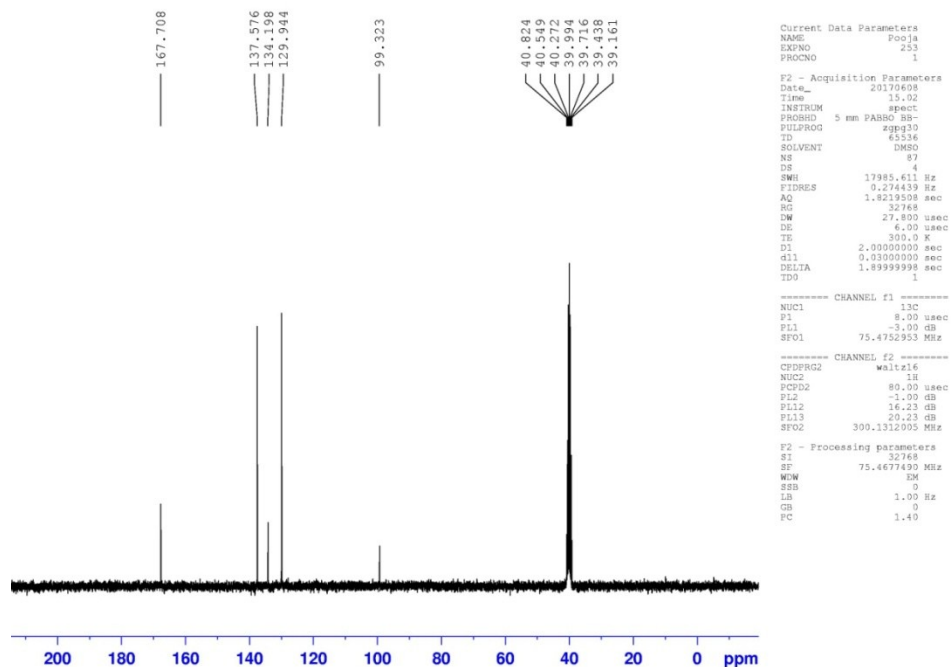


Figure S18. <sup>13</sup>C NMR of 4d

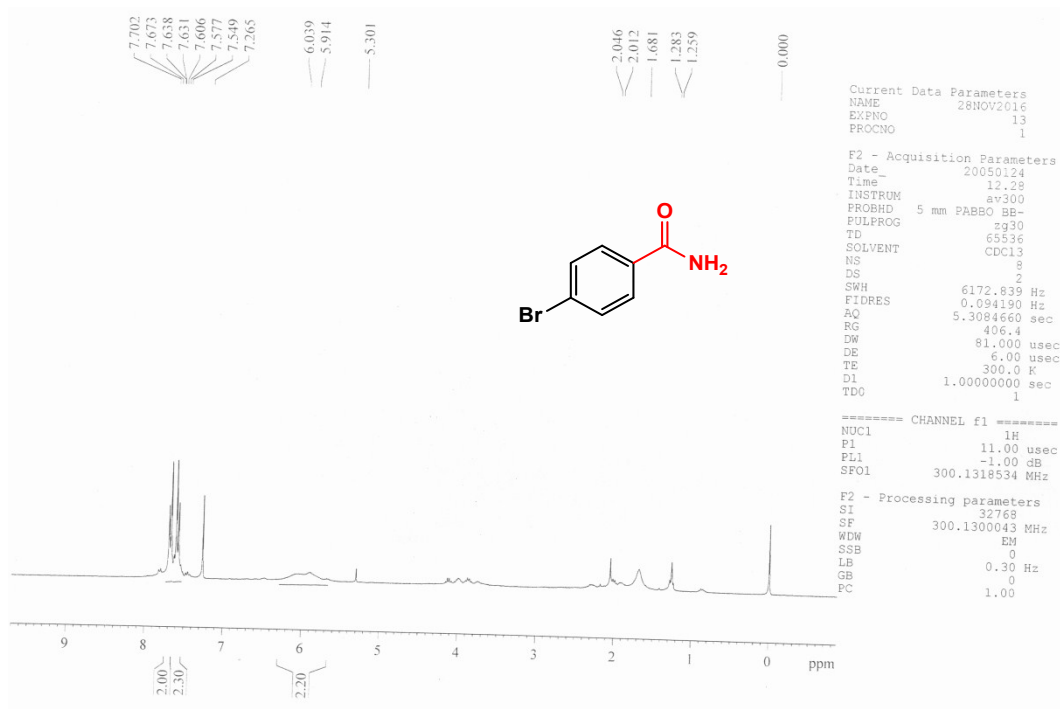


Figure S19. <sup>1</sup>H NMR of 4e

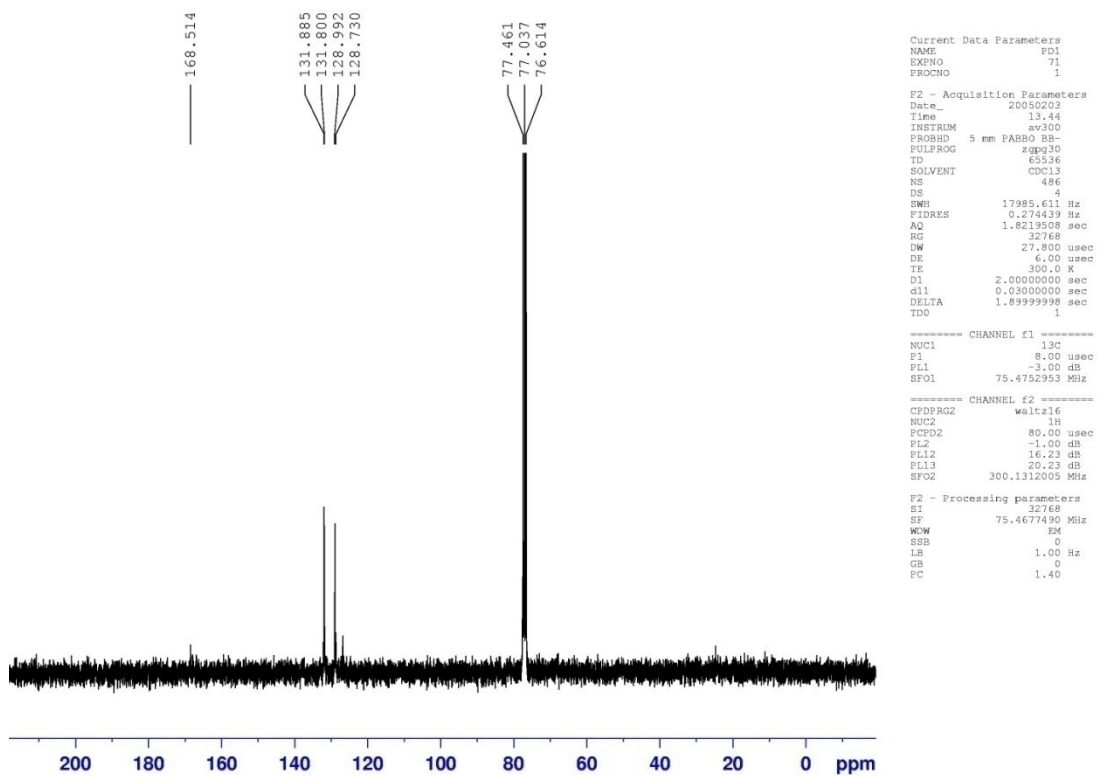


Figure S20. <sup>13</sup>C NMR of 4e

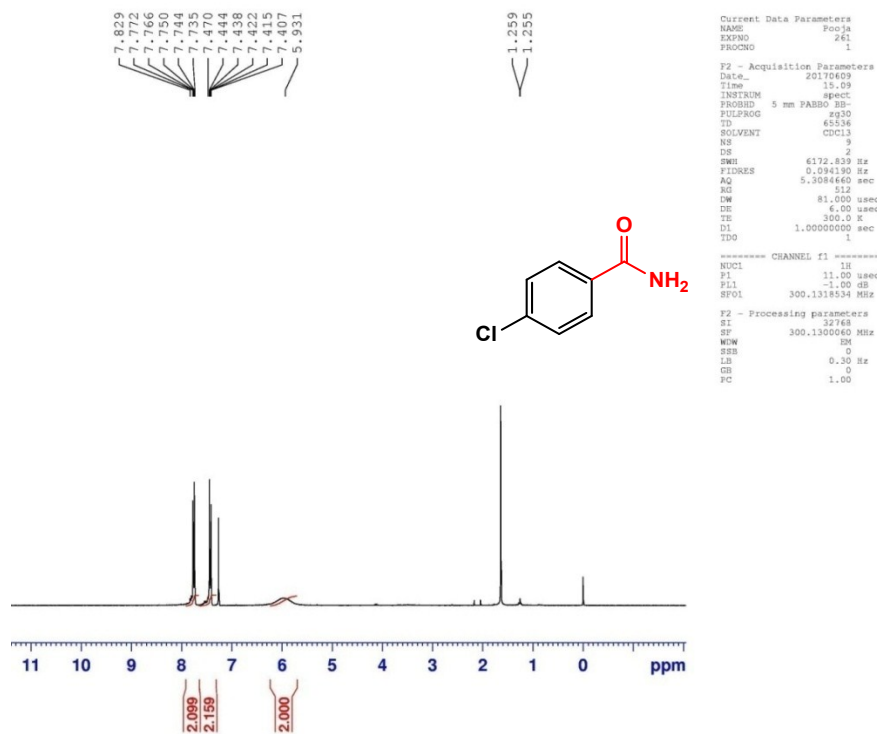


Figure S21. <sup>1</sup>H NMR of 4f

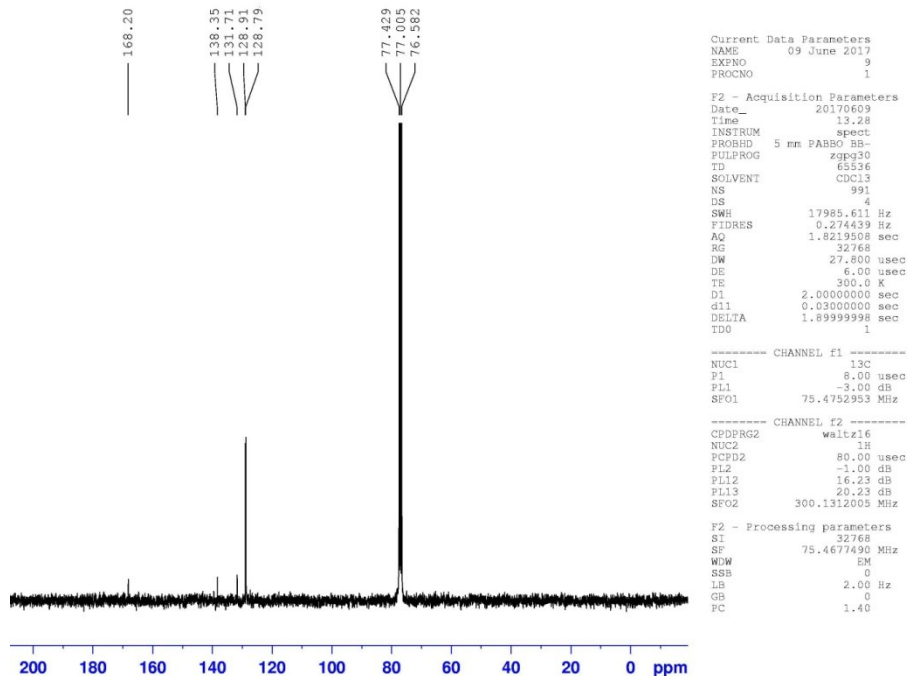


Figure S22. <sup>13</sup>C NMR of 4f

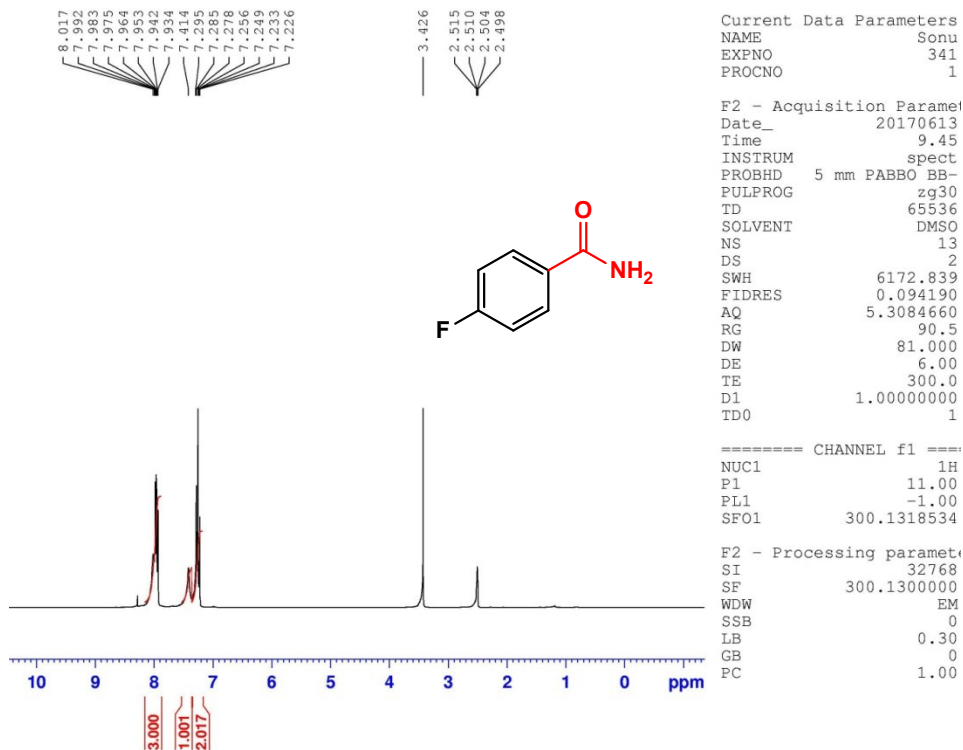


Figure S23. <sup>1</sup>H NMR of 4g

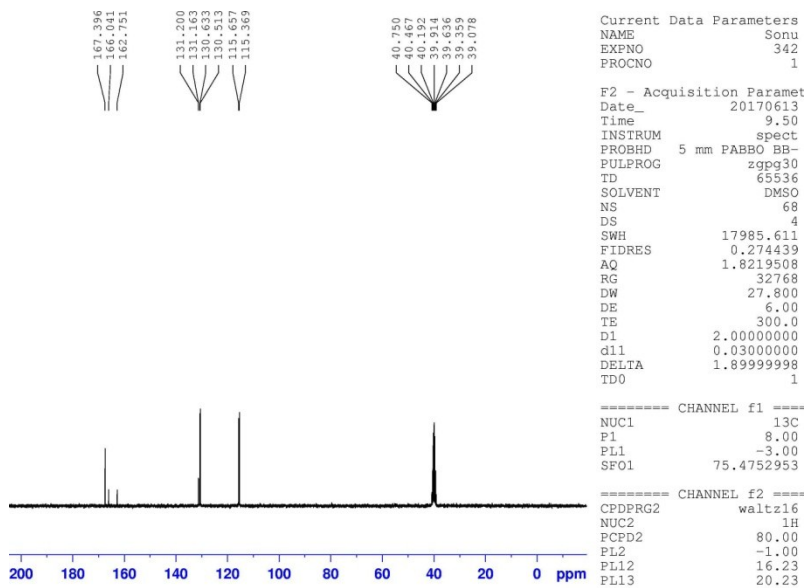


Figure S24. <sup>13</sup>C NMR of 4g

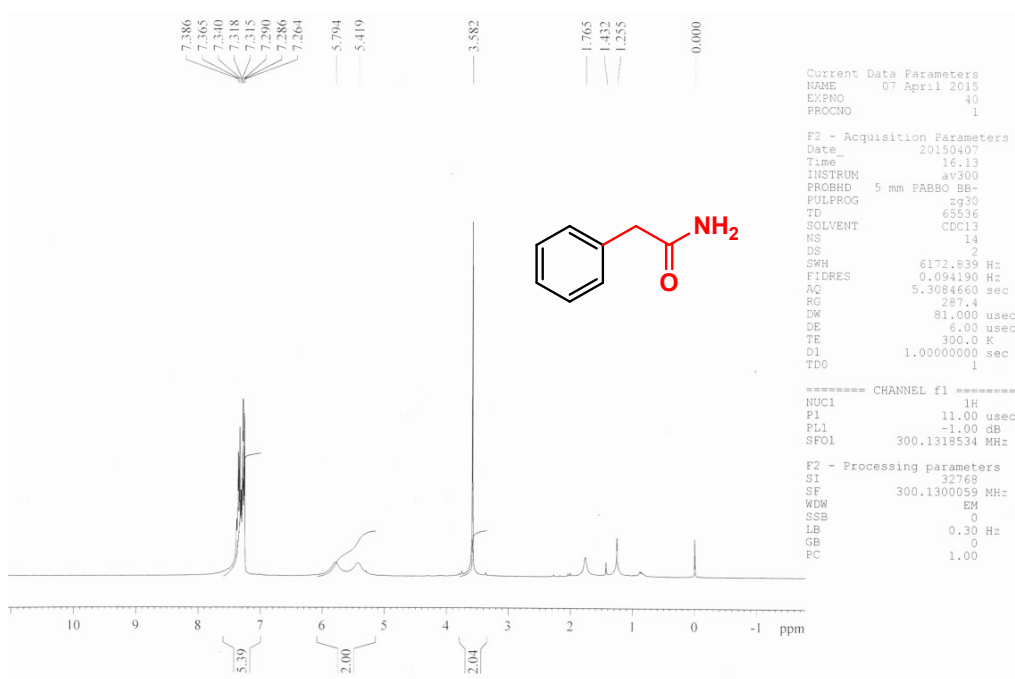


Figure S25. <sup>1</sup>H NMR of 4h



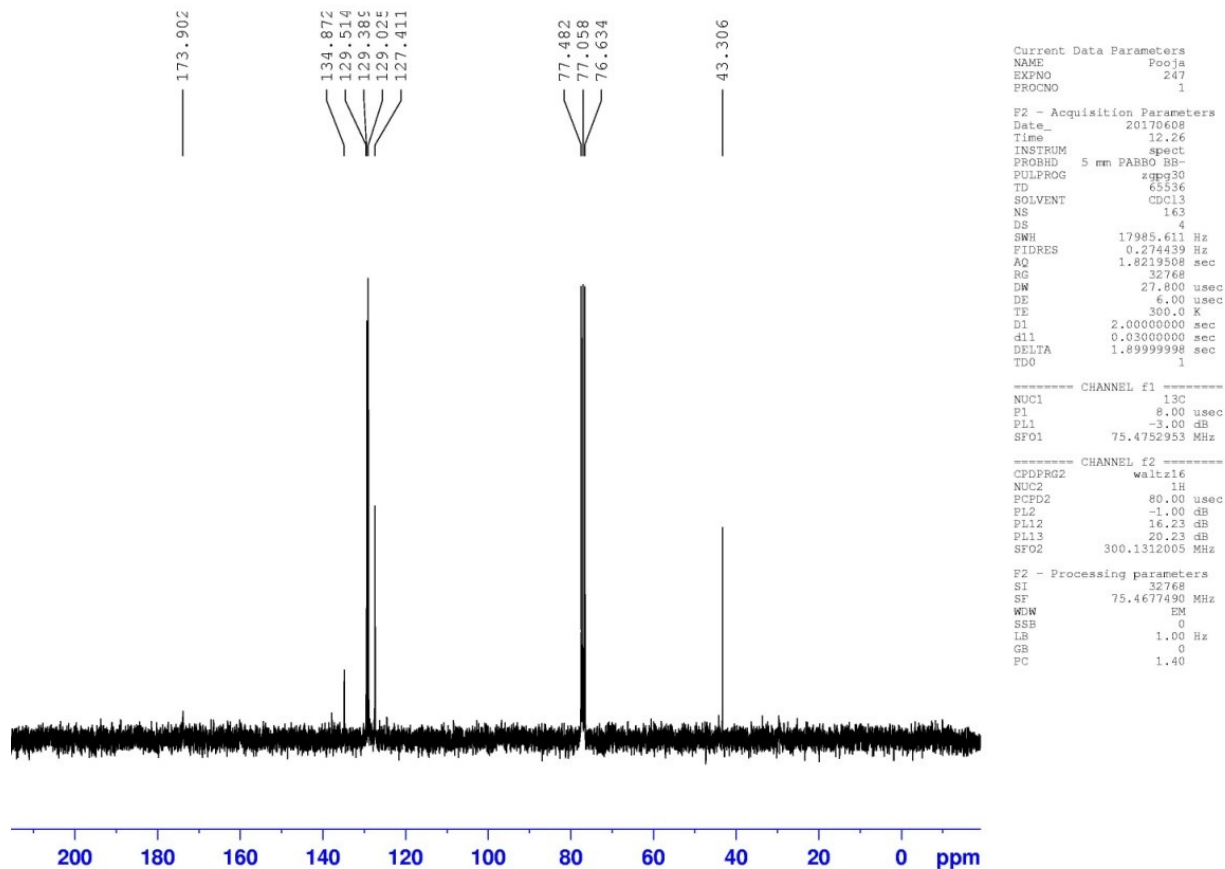


Figure S26. <sup>13</sup>C NMR of 4h

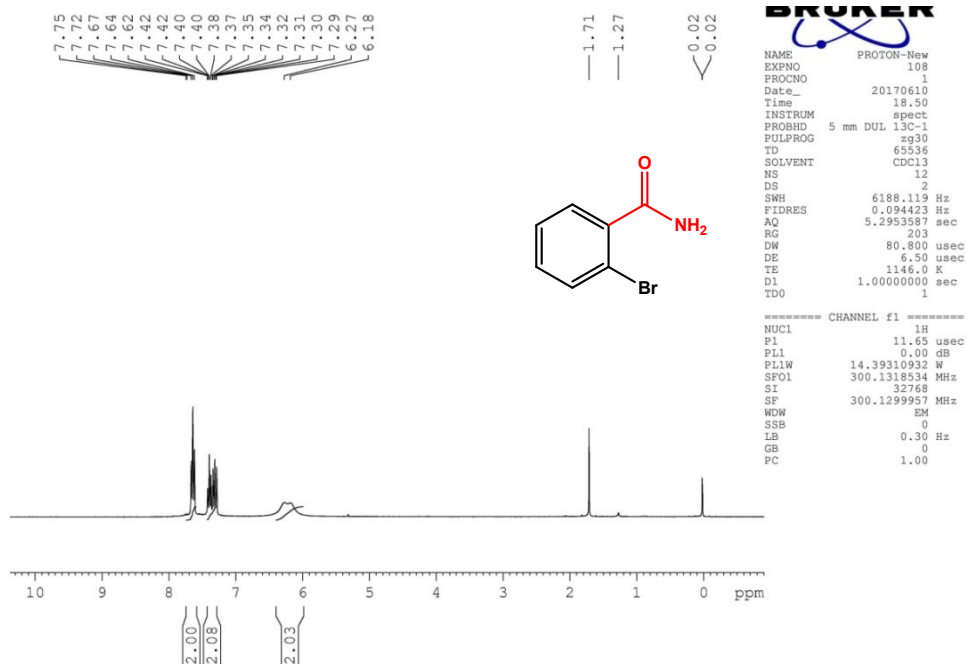


Figure S27. <sup>1</sup>H NMR of 4i

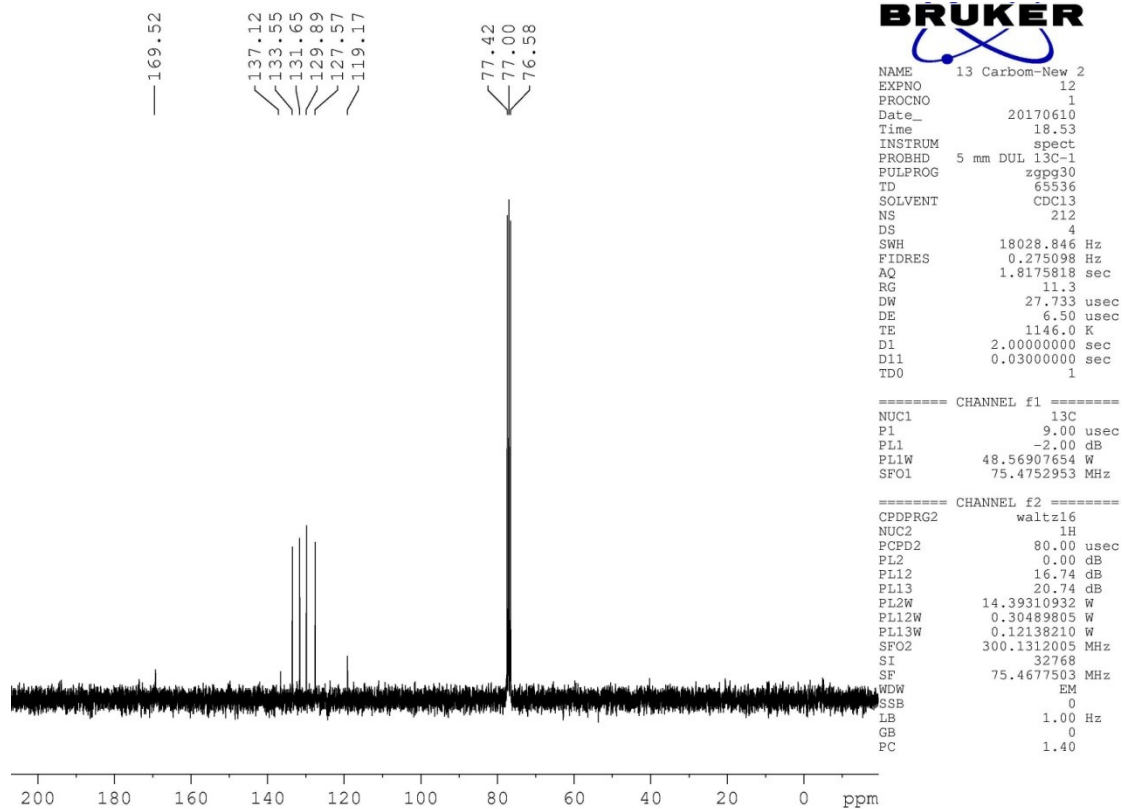


Figure S28. <sup>13</sup>C NMR of 4i

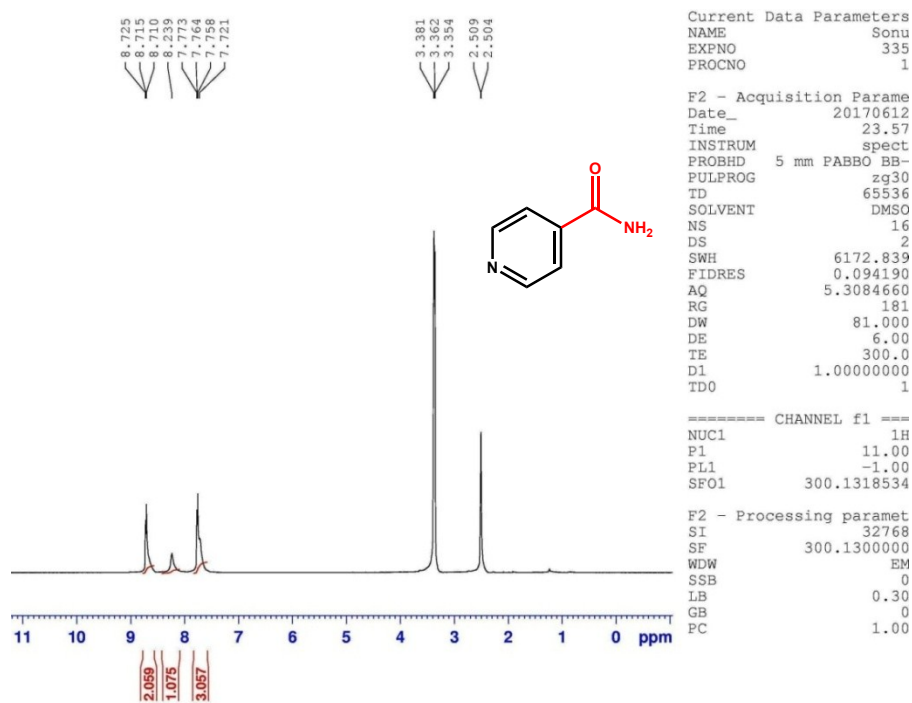
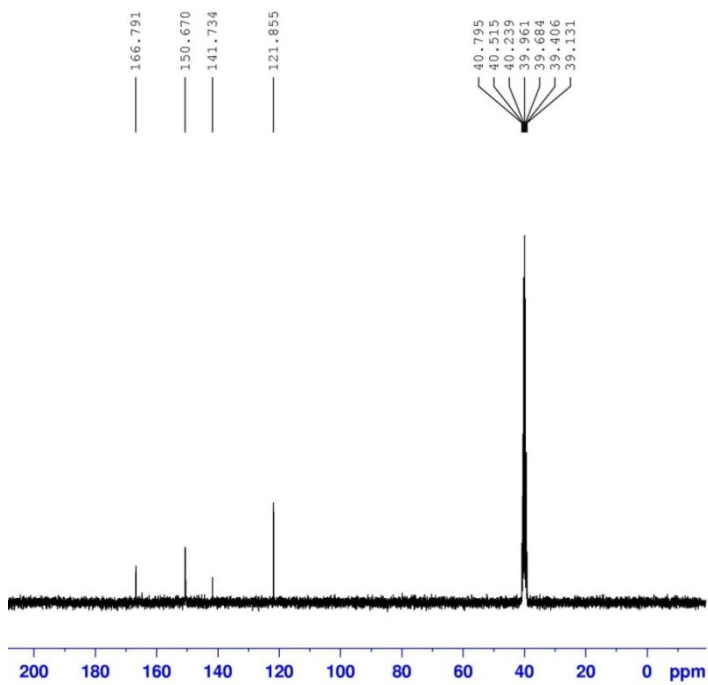


Figure S29. <sup>1</sup>H NMR of 4j



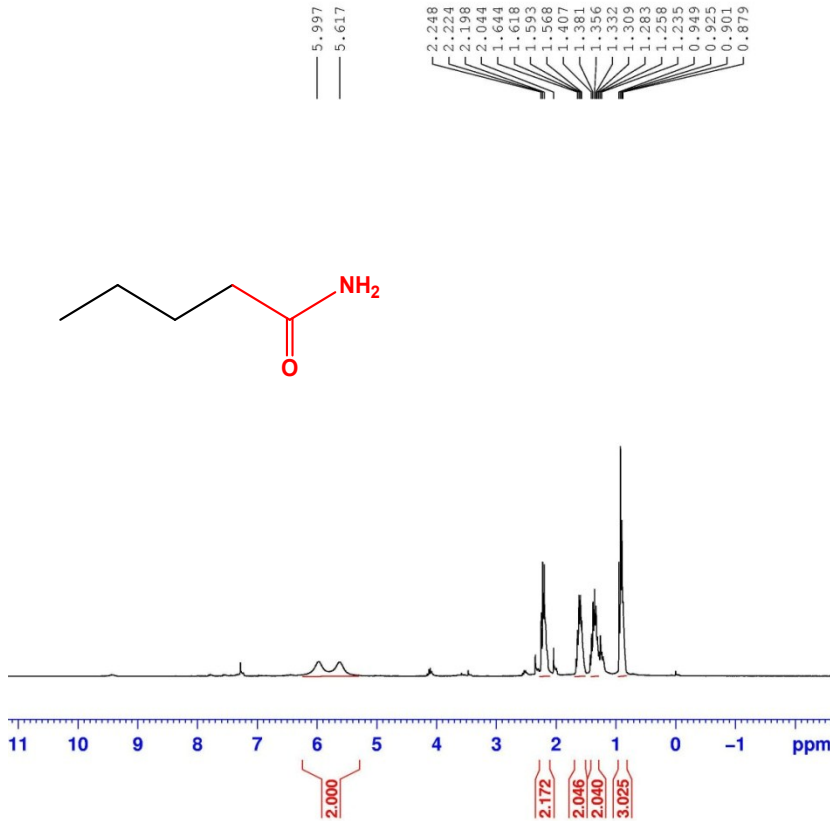
Current Data Parameters  
 NAME Sonu  
 EXPNO 336  
 PROCNO 1

F2 - Acquisition Paramet  
 Date\_ 20170613  
 Time\_ 0.00  
 INSTRUM spect  
 PROBHD 5 mm PABBO BB-  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT DMSO  
 NS 177  
 DS 4  
 SWH 17985.611  
 FIDRES 0.274439  
 AQ 1.8219508  
 RG 32768  
 DW 27.800  
 DE 6.00  
 TE 300.0  
 D1 2.00000000  
 d11 0.03000000  
 DELTA 1.89999999  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 13C  
 P1 8.00  
 PL1 -3.00  
 SFO1 75.4752953

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 80.00  
 PL2 -1.00  
 PL12 16.23  
 PL13 20.23  
 SFO2 300.1312005

Figure S30. <sup>13</sup>C NMR of 4j



Current Data Parameters  
 NAME Fooja  
 EXPNO 251  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20170608  
 Time\_ 13.00  
 INSTRUM spect  
 PROBHD 5 mm PABBO BB-  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 5  
 DS 2  
 SWH 6172.839 Hz  
 FIDRES 0.094190 Hz  
 AQ 5.3084660 sec  
 RG 128  
 DW 81.000 usec  
 DE 6.00 usec  
 TE 300.0 K  
 D1 1.00000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 11.00 usec  
 PL1 -1.00 dB  
 SFO1 300.1318534 MHz

F2 - Processing parameters  
 SI 32768  
 SF 300.1300000 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00

Figure S31. <sup>1</sup>H NMR of 4k

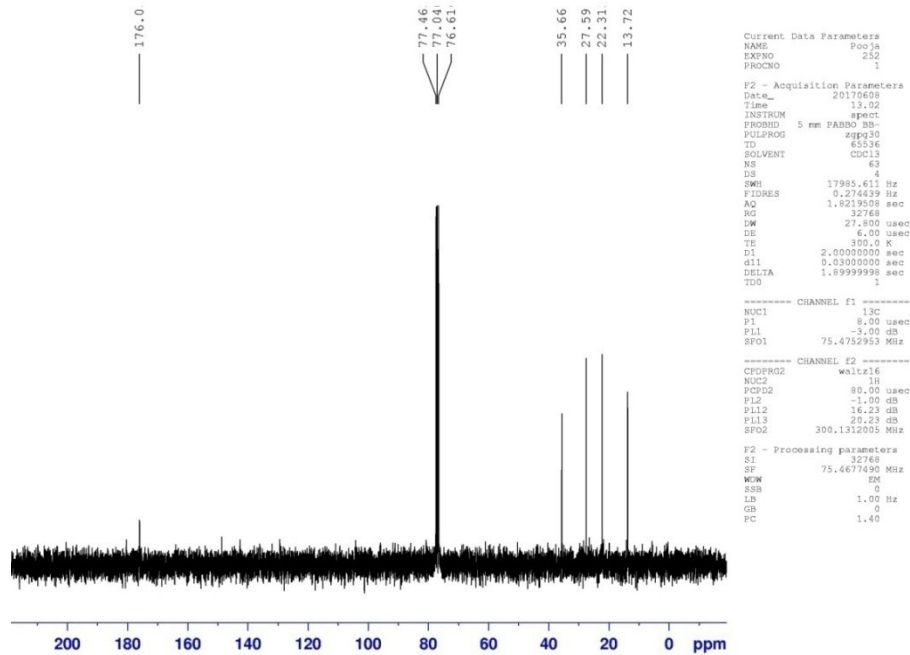


Figure S32. <sup>13</sup>C NMR of 4k

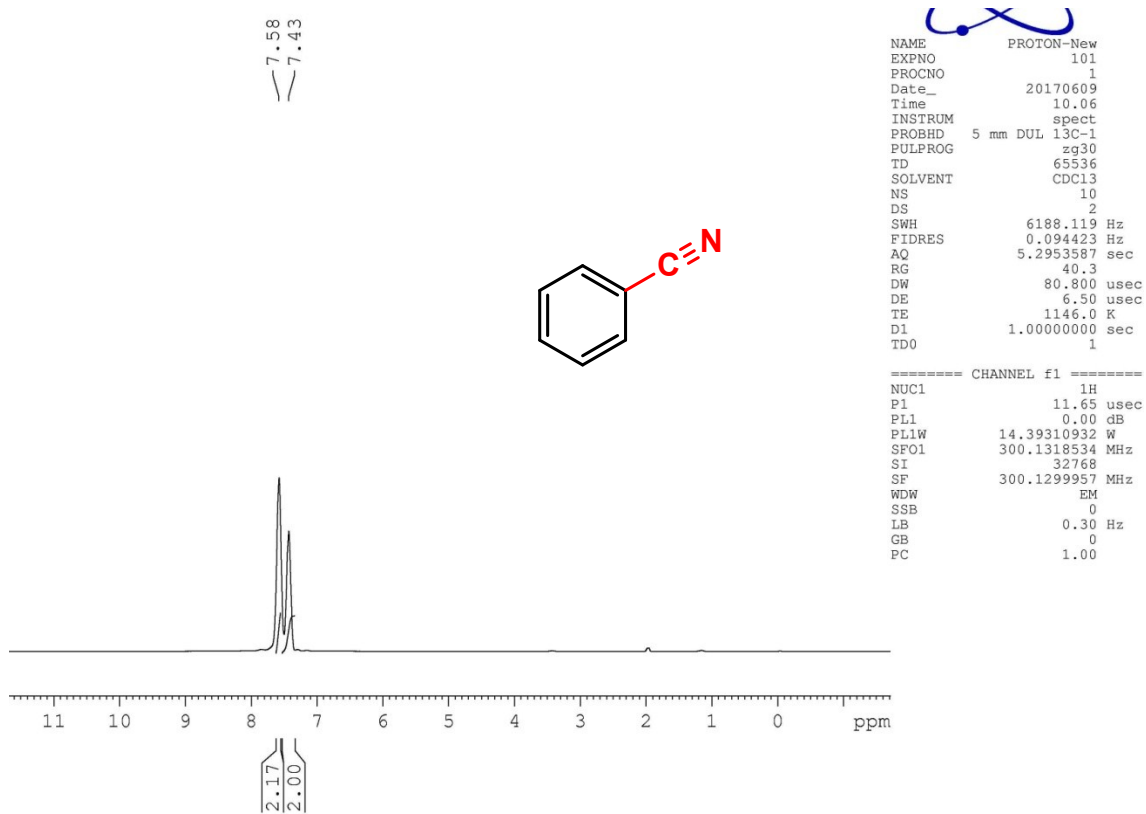


Figure S33. <sup>1</sup>H NMR of 6a

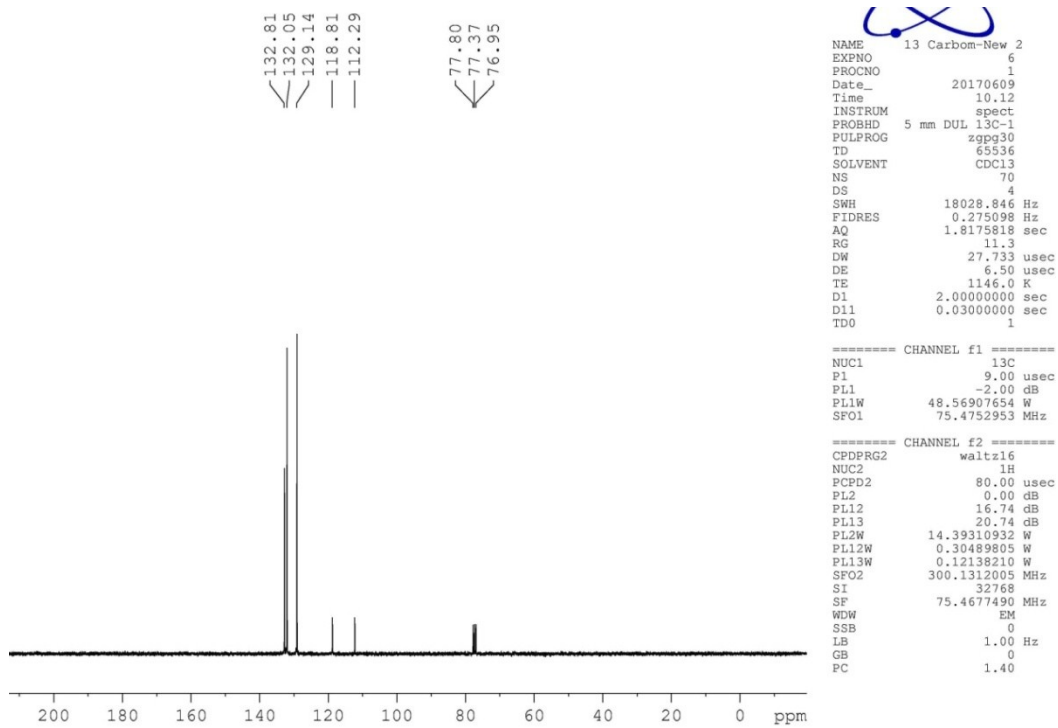


Figure S34. <sup>13</sup>C NMR of 6a

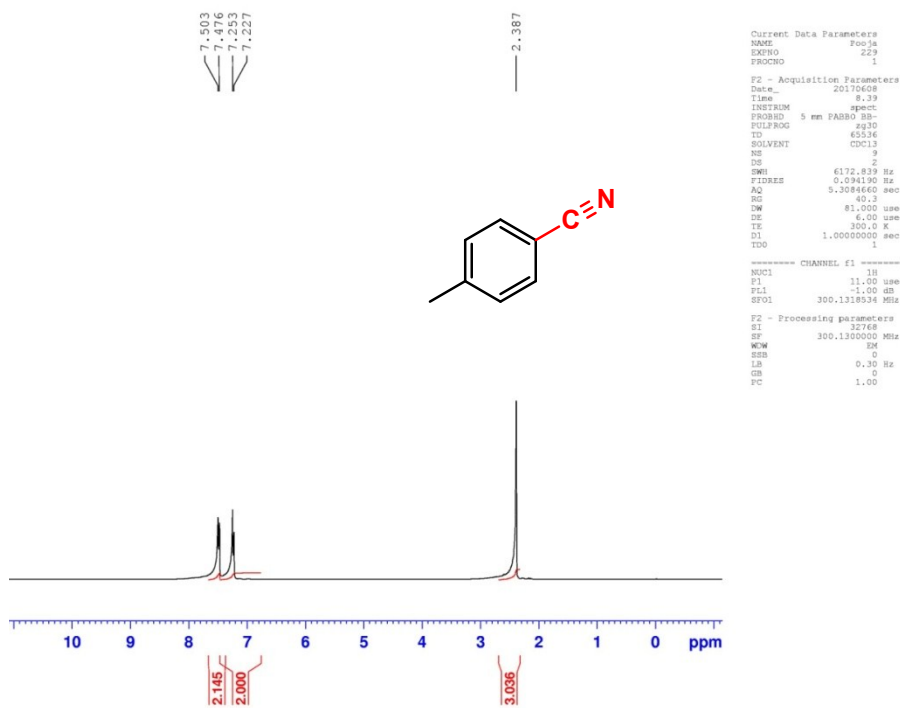
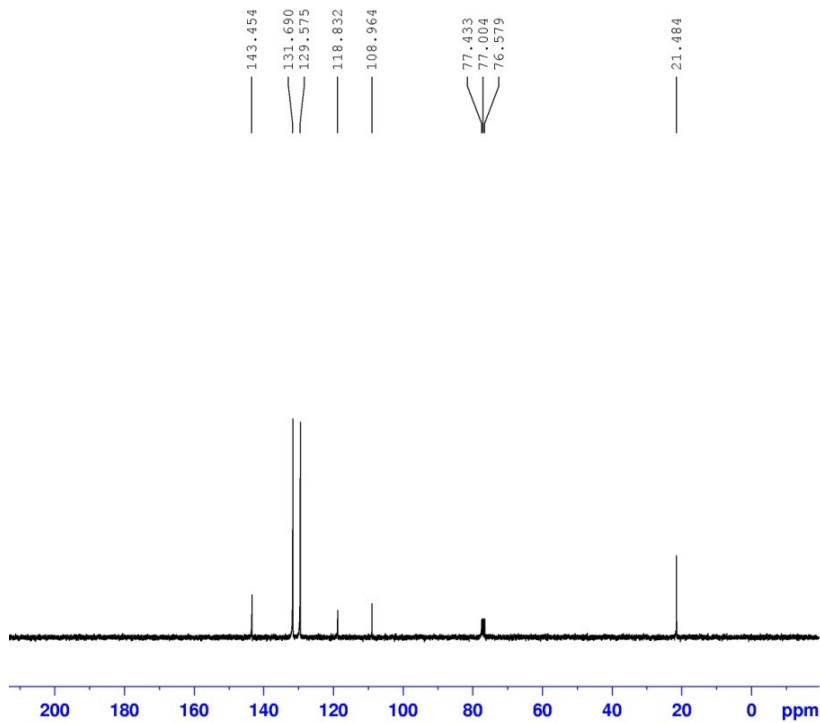


Figure S35. <sup>1</sup>H NMR of 6b



```

Current Data Parameters
NAME      Poo3a
EXPNO    230
PROCNO    1

F2 - Acquisition Parameters
Date_     20170608
Time      8.42
INSTRUM   spect
PROBHD    5 mm PABBO BB-
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         29
DS         4
SWH        17985.611 Hz
FIDRES     0.274439 Hz
AQ         1.8219508 sec
RG         32768
DW         27.800 usec
DE         6.00 usec
TE         300.0 K
D1         2.0000000 sec
d11        0.0300000 sec
DELTA     1.89999998 sec
TD0        1
  
```

```

===== CHANNEL f1 =====
NUC1      13C
P1        8.00 usec
PL1       -1.00 dB
SFO1      75.4752953 MHz
  
```

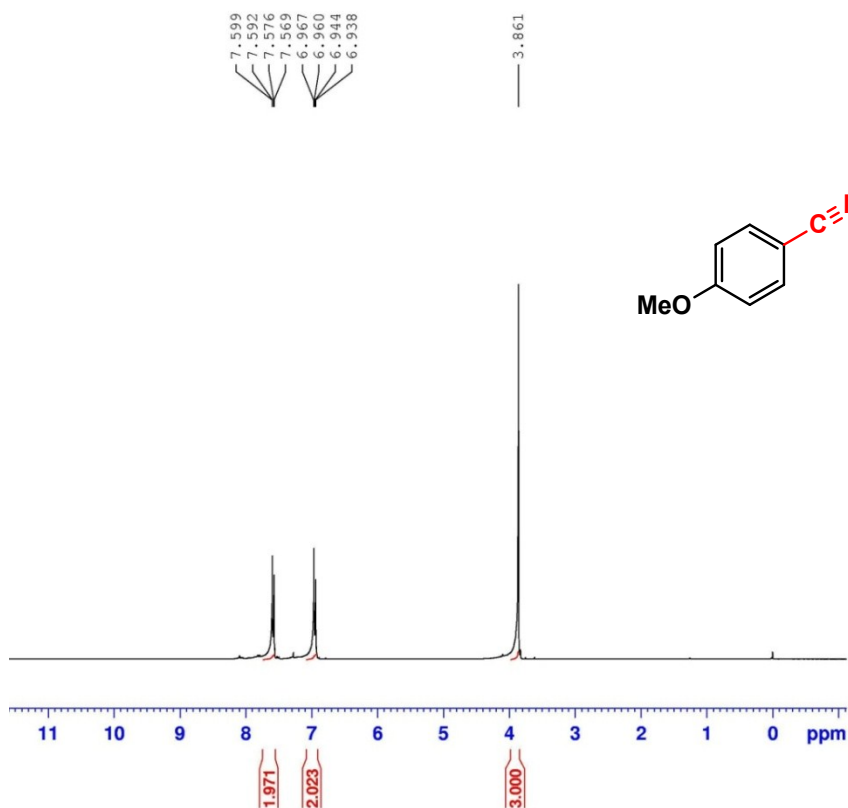
```

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2      1H
PCPD2     80.00 usec
PL2       -1.00 dB
PL12      16.23 dB
PL13      0.23 dB
SFO2      300.1312005 MHz
  
```

```

F2 - Processing parameters
SI         32768
SF         75.4677697 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40
  
```

Figure S36. <sup>13</sup>C NMR of 6b



```

Current Data Parameters
NAME      PDI
EXPNO    27
PROCNO    1

F2 - Acquisition Parameters
Date_     20050119
Time      13.34
INSTRUM   av300
PROBHD    5 mm PABBO BB-
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         16
DS         2
SWH        6172.839 Hz
FIDRES     0.094190 Hz
AQ         5.3084660 sec
RG         114
DW         81.000 usec
DE         6.00 usec
TE         300.0 K
D1         1.00000000 sec
TD0        1
  
```

```

===== CHANNEL f1 =====
NUC1      1H
P1        11.00 usec
PL1       -1.00 dB
SFO1      300.1318534 MHz

F2 - Processing parameters
SI         32768
SF         300.1300000 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
  
```

Figure S37. <sup>1</sup>H NMR of 6c

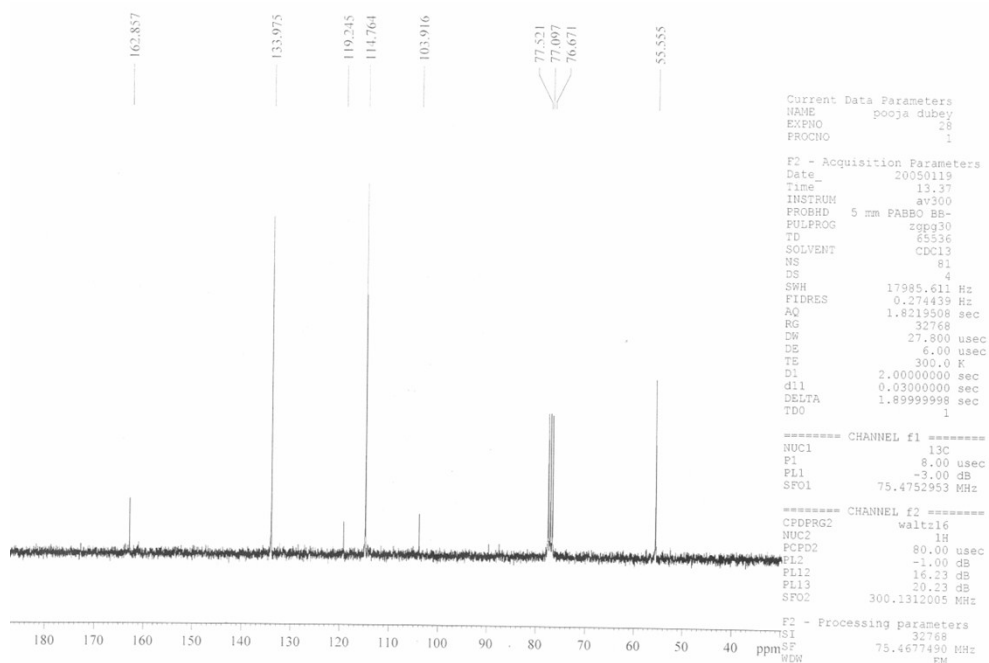


Figure S38. <sup>13</sup>C NMR of 6c

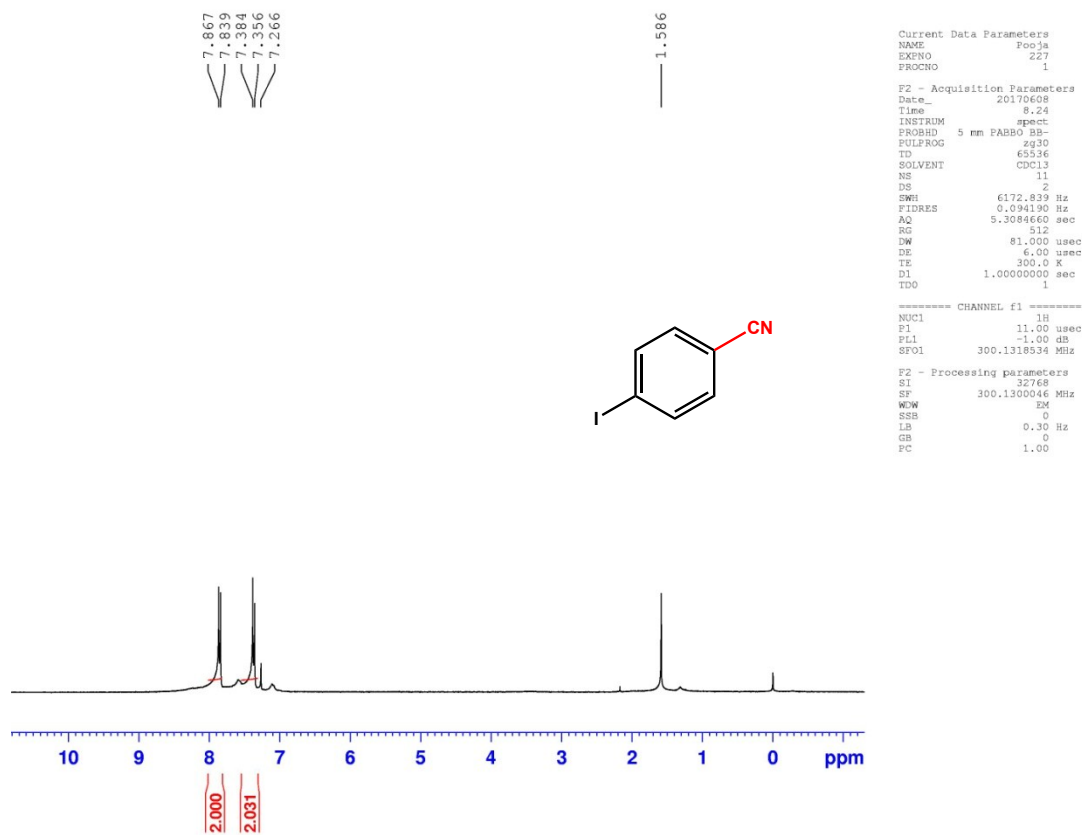
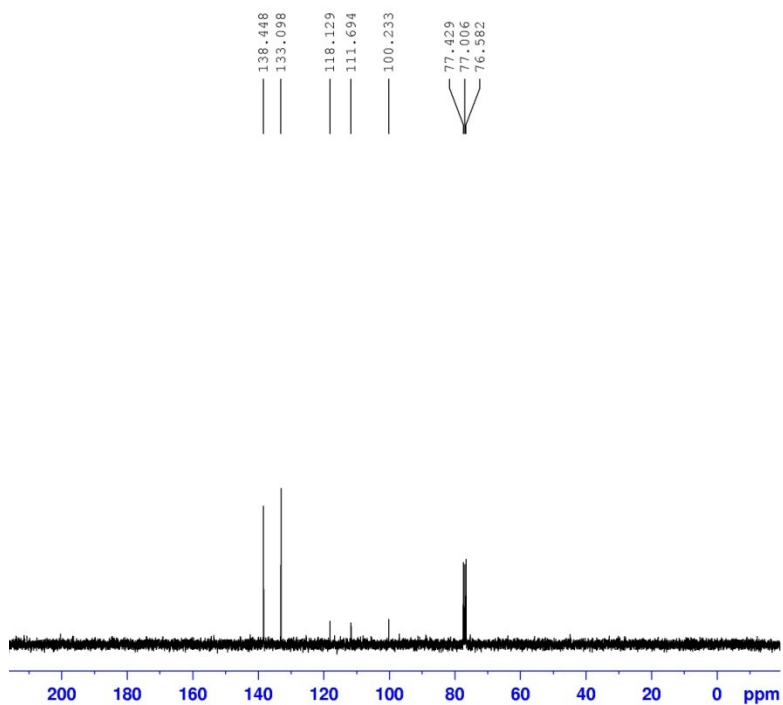


Figure S39. <sup>1</sup>H NMR of 6d



```

Current Data Parameters
NAME      Pooja
EXPNO    241
PROCNO   1

F2 - Acquisition Parameters
Date_    20170608
Time     9.29
INSTRUM  spect
PROBHD   5 mm PABBO BB-
PULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       18
DS       4
SWH      17985.611 Hz
FIDRES   0.274439 Hz
AQ       1.8219508 sec
RG       32768
DW       27.800 usec
DE       6.00 usec
TE       300.0 K
D1       2.00000000 sec
d11      0.03000000 sec
DELTA    1.89999998 sec
TDO      1
  
```

```

----- CHANNEL f1 -----
NUC1     13C
P1       8.00 usec
PL1      -3.00 dB
SFO1     75.4752993 MHz
  
```

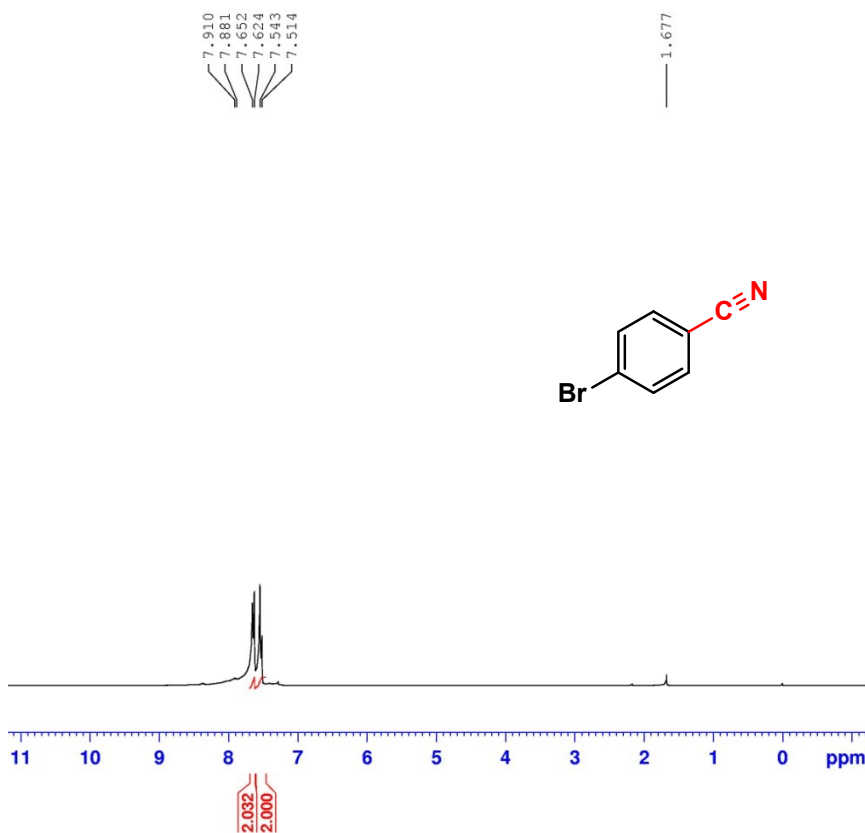
```

----- CHANNEL F2 -----
CPDPRG2  waltz16
NUC2     1H
PCPD2    80.00 usec
PL2      -1.00 dB
PL12     16.23 dB
PL13     20.23 dB
SFO2     300.1312005 MHz
  
```

```

F2 - Processing parameters
SI       32768
SF       75.4677547 MHz
WEN      EM
SSB      0
LB       1.00 Hz
GB       0
PC       1.40
  
```

Figure S40. <sup>13</sup>C NMR of 6d



```

Current Data Parameters
NAME      Pooja
EXPNO    237
PROCNO   1
  
```

```

F2 - Acquisition Parameters
Date_    20170608
Time     9.13
INSTRUM  spect
PROBHD   5 mm PABBO BB-
PULPROG  zg30
TD       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      6172.839 Hz
FIDRES   0.094190 Hz
AQ       5.3084660 sec
RG       142.7
DW       81.000 usec
DE       6.00 usec
TE       300.0 K
D1       1.00000000 sec
TDO      1
  
```

```

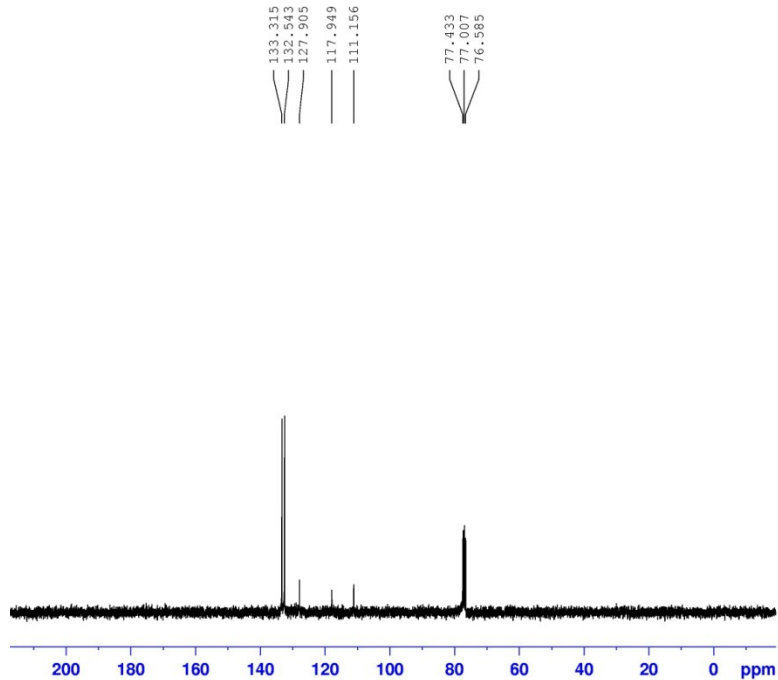
----- CHANNEL f1 -----
NUC1     1H
P1       11.00 usec
PL1      -1.00 dB
SFO1     300.1318534 MHz
  
```

```

F2 - Processing parameters
SI       32768
SF       300.1300000 MHz
WEN      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00
  
```

Figure S41. <sup>1</sup>H NMR of 6e





```

Current Data Parameters
NAME      Foo3a
EXPNO     238
PROCNO    1

F2 - Acquisition Parameters
Date_     20170608
Time      9.15
INSTRUM   spect
PROBHD    5 mm PABBO BB-
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         50
DS         4
SWH        17985.611 Hz
FIDRES     0.274439 Hz
AQ         1.8219508 sec
RG         32768
DW         27.800 usec
DE         6.00 usec
TE         300.0 K
D1         2.00000000 sec
sH1        0.03000000 sec
DELTA     1.89999998 sec
TD0        1
  
```

```

===== CHANNEL f1 =====
NUC1      13c
P1         8.00 usec
PL1        -3.00 dB
SFO1      75.4752953 MHz
  
```

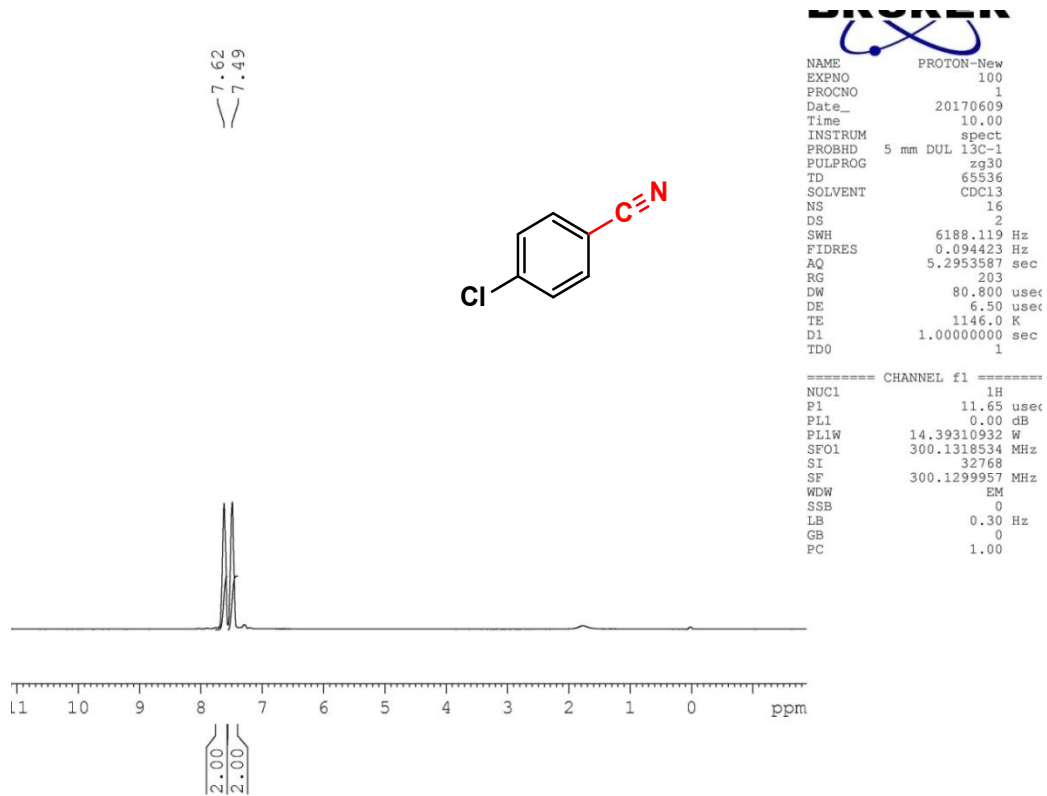
```

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2       1H
PCPD2     80.00 usec
PL2        -1.00 dB
PL12       16.23 dB
PL13       20.23 dB
SFO2      300.1312005 MHz
  
```

```

F2 - Processing parameters
SI         32768
SF         75.4677566 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
EC         1.40
  
```

Figure S42. <sup>13</sup>C NMR of 6e



PRON-NEW

```

NAME      PRON-NEW
EXPNO     100
PROCNO    1
Date_     20170609
Time      10.00
INSTRUM   spect
PROBHD    5 mm DUL 13C-1
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         16
DS         2
SWH        6188.119 Hz
FIDRES     0.094423 Hz
AQ         5.2953587 sec
RG         203
DW         80.800 usec
DE         6.50 usec
TE         1146.0 K
D1         1.00000000 sec
TD0        1
  
```

```

===== CHANNEL f1 =====
NUC1      1H
P1         11.65 usec
PL1        0.00 dB
PL1W      14.39310932 W
SFO1      300.1318534 MHz
SI         32768
SF         300.1299957 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
EC         1.00
  
```

Figure S43. <sup>1</sup>H NMR of 6f

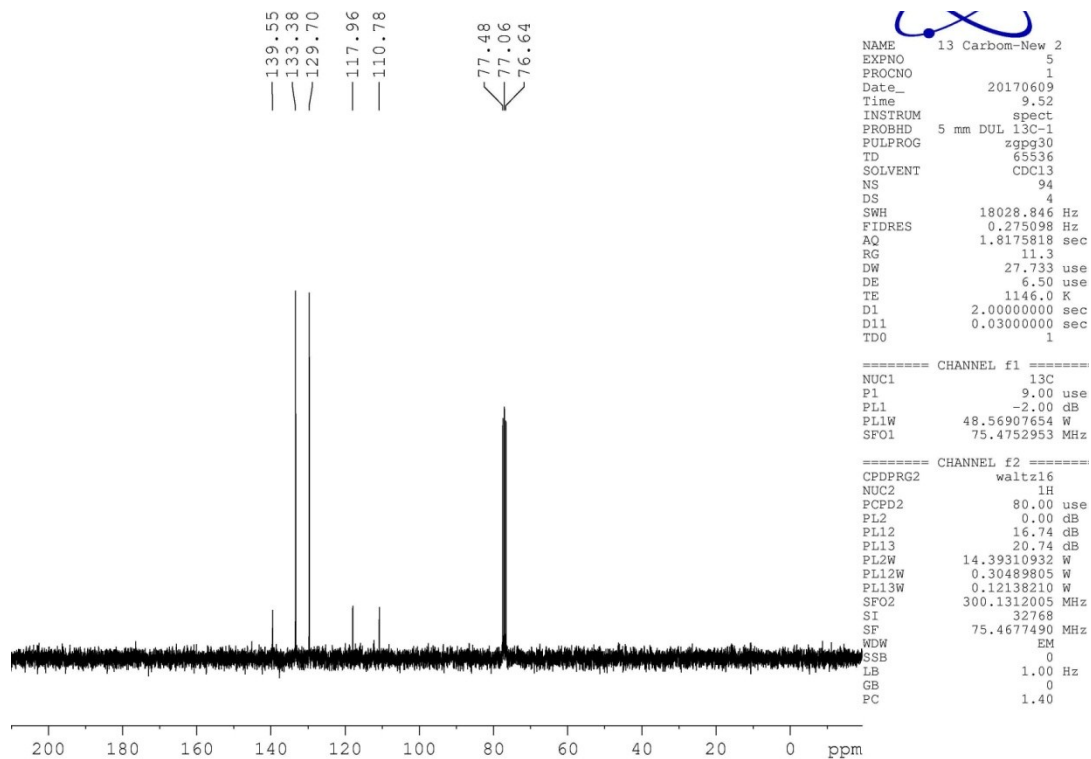


Figure S44.  $^{13}\text{C}$  NMR of 6f

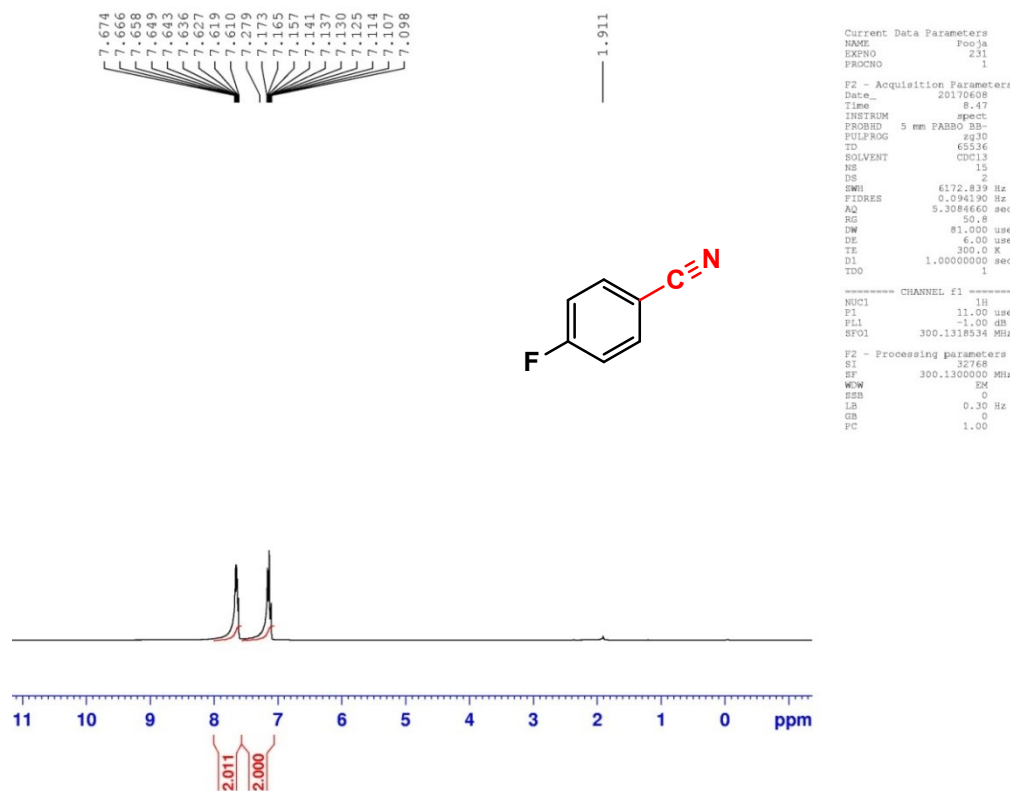


Figure S45.  $^1\text{H}$  NMR of 6g

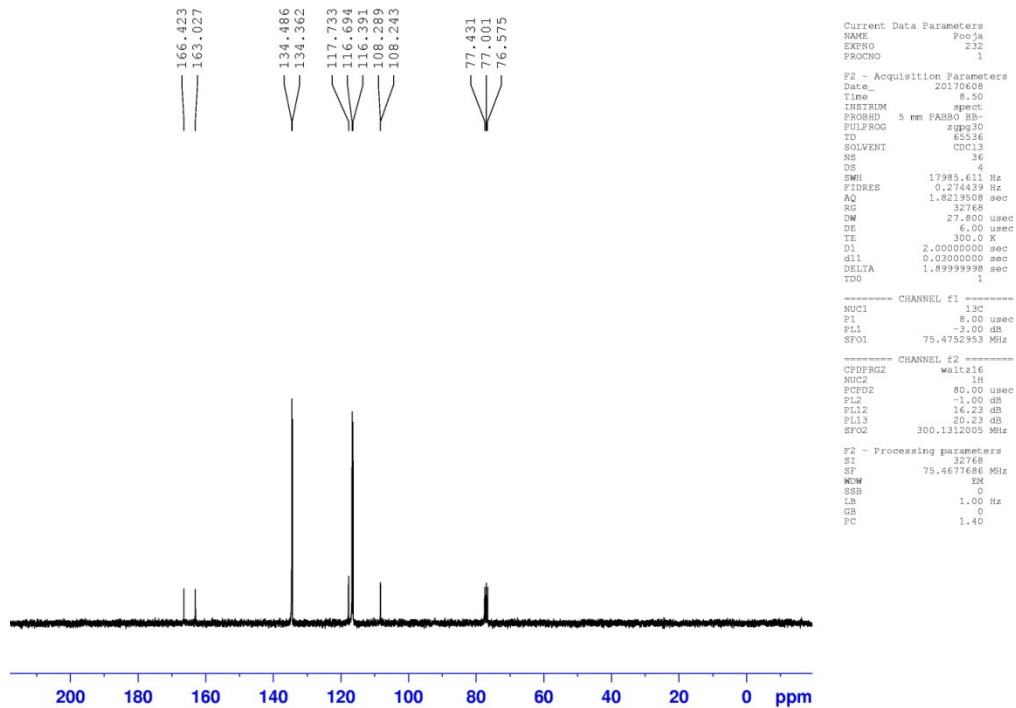


Figure S46. <sup>13</sup>C NMR of 6g

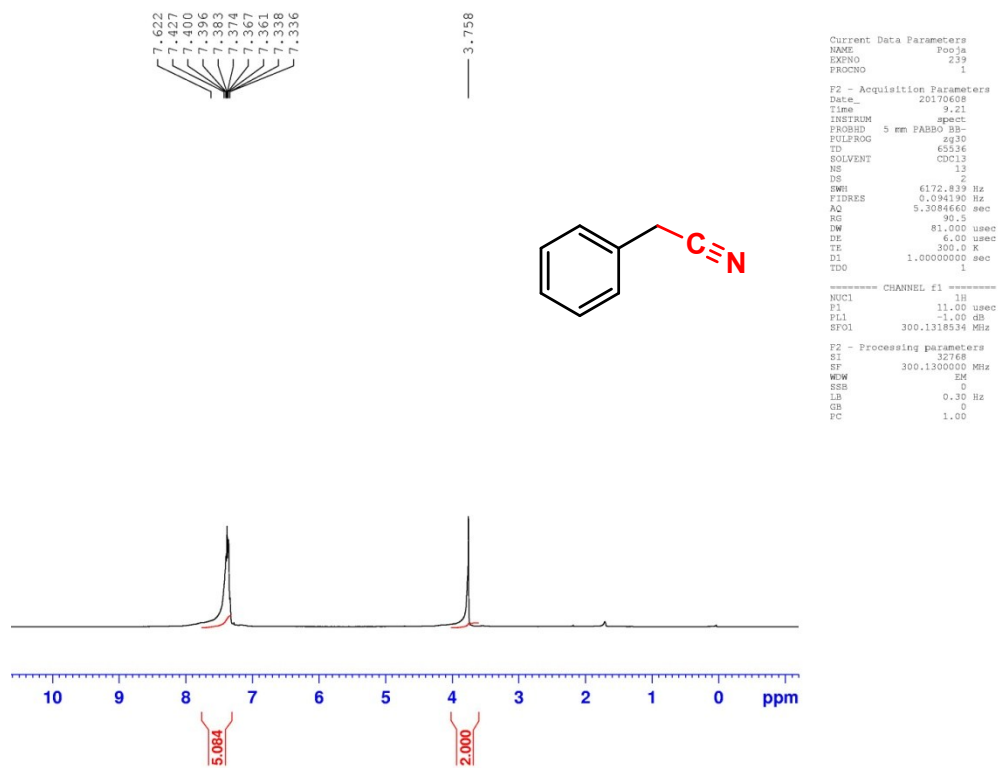


Figure S47. <sup>1</sup>H NMR of 6h

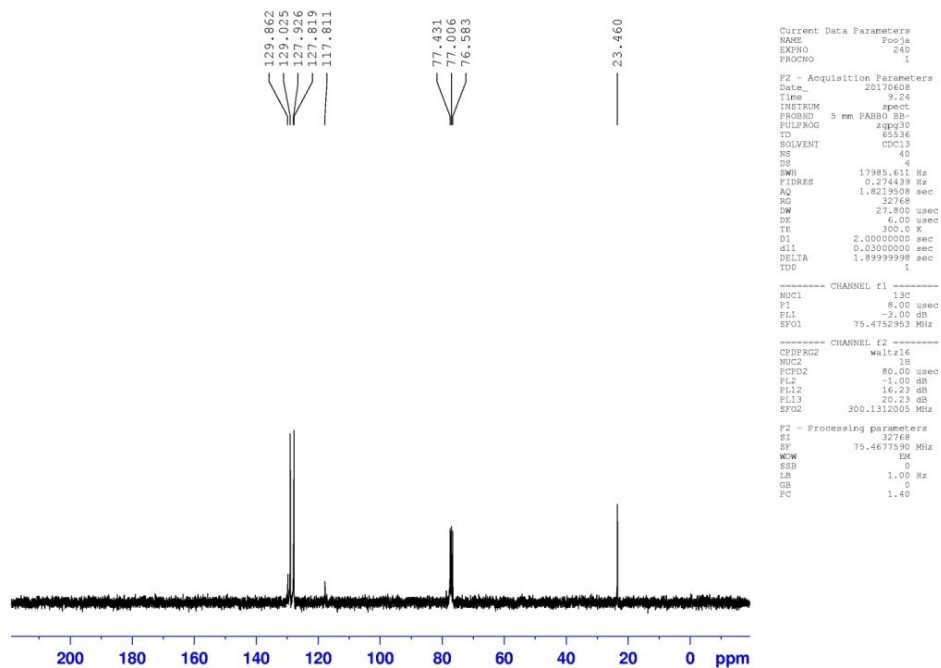


Figure S48. <sup>13</sup>C NMR of 6h

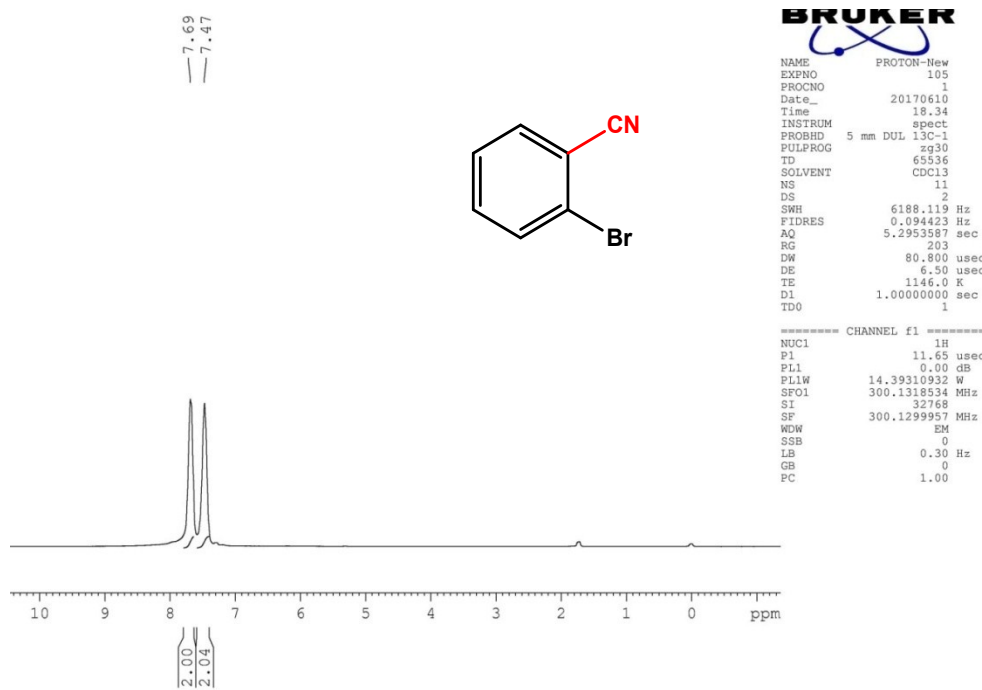


Figure S49. <sup>1</sup>H NMR of 6i

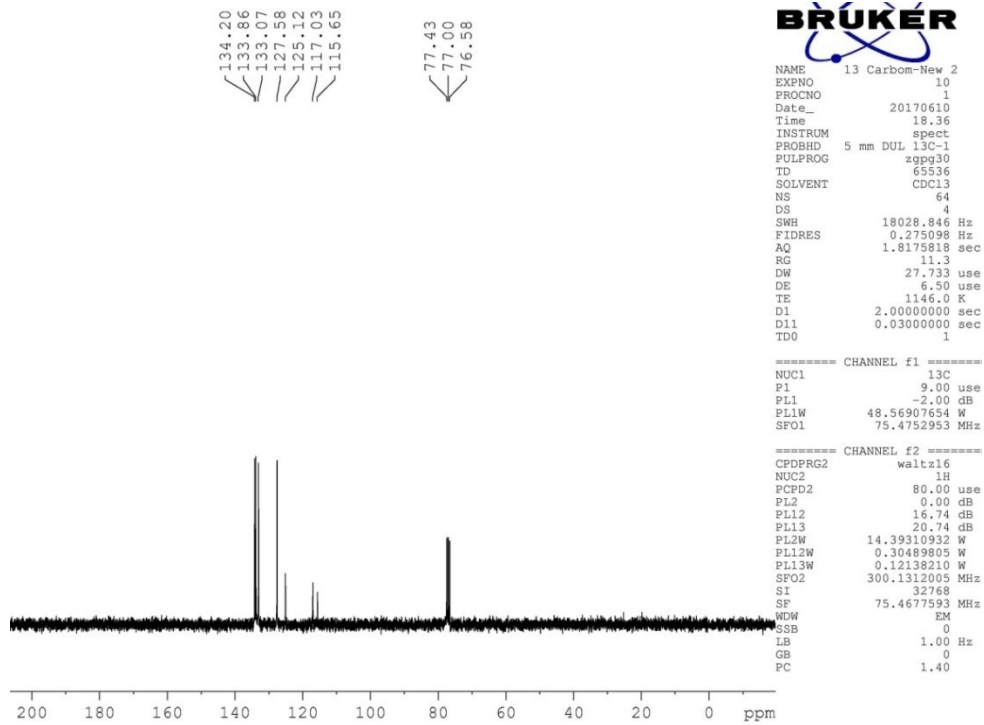


Figure S50. <sup>13</sup>C NMR of 6i

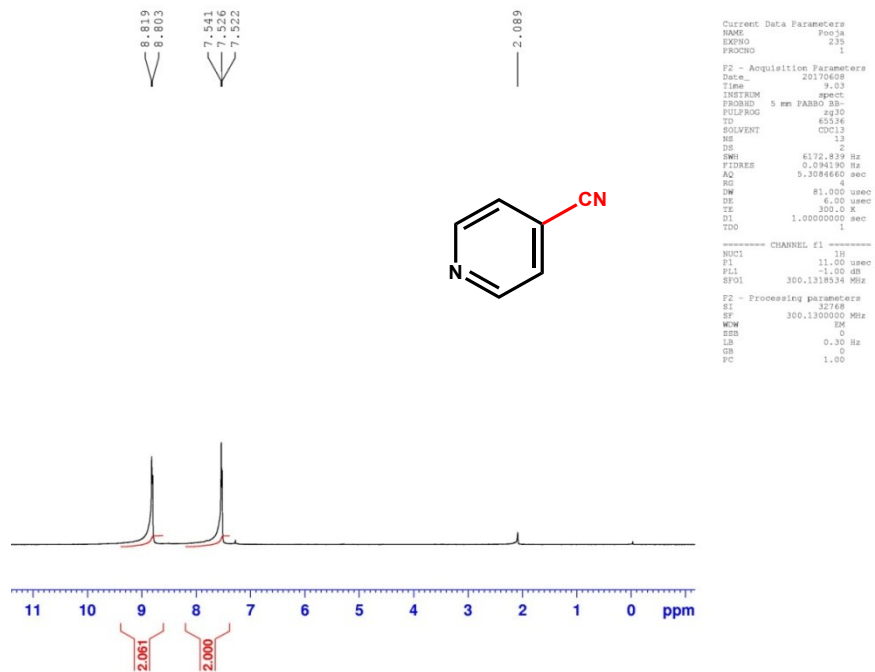
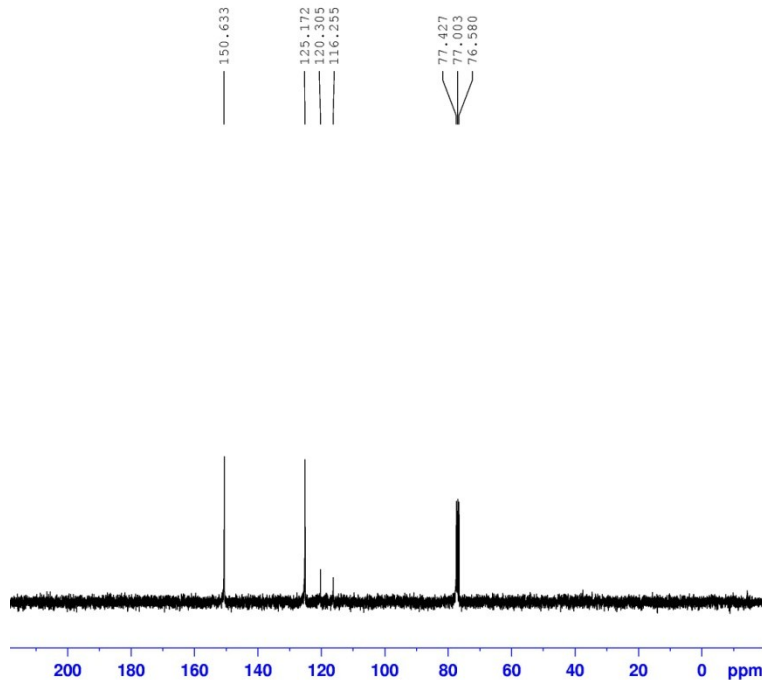


Figure S51. <sup>1</sup>H NMR of 6j



```

Current Data Parameters
NAME      Fooja
EXPNO    236
PROCNO   1

F2 - Acquisition Parameters
Date_    20170608
Time     9.06
INSTRUM  spect
PROBHD   5 mm PABBO BB-
PULPROG  zgpg30
TD        65536
SOLVENT  cdcl3
NS        51
DS        4
SWH       17985.611 Hz
FIDRES   0.274439 Hz
AQ        1.8219508 sec
RG         32768
DW        27.800 usec
DE         6.00 usec
TE        300.0 K
D1        2.0000000 sec
d11       0.0300000 sec
DELTA    1.89999998 sec
TDO       1

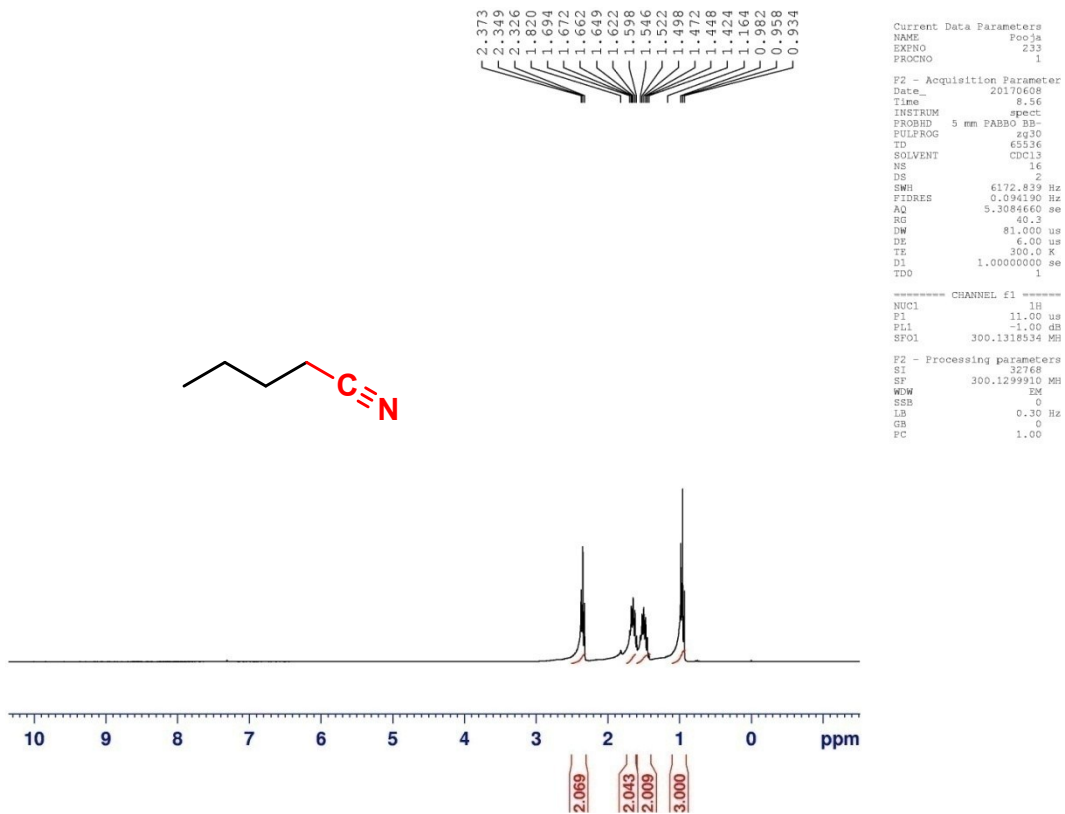
----- CHANNEL f1 -----
NUC1      13C
P1        8.00 usec
PL1       -3.00 dB
SFO1      75.4752953 MHz

----- CHANNEL f2 -----
CPDPRG2  waltz16
NUC2      1H
PCPD2    80.00 usec
PL2       -1.00 dB
PL12     16.23 dB
PL13     20.23 dB
SFO2     300.1312003 MHz

F2 - Processing parameters
SI        32768
SF        75.4677573 MHz
WDW       EM
SSB       0
LB        1.00 Hz
GB         0
PC        1.40

```

Figure S52. <sup>13</sup>C NMR of 6j



```

Current Data Parameters
NAME      Fooja
EXPNO    233
PROCNO   1

F2 - Acquisition Parameter
Date_    20170608
Time     8.56
INSTRUM  spect
PROBHD   5 mm PABBO BB-
PULPROG  zg30
TD        65536
SOLVENT  CDCl3
NS        16
DS        2
SWH       6172.839 Hz
FIDRES   0.094190 Hz
AQ        5.3084660 se
RG         40.3
DW        81.000 us
DE         6.00 us
TE        300.0 K
D1        1.00000000 se
TDO       1

----- CHANNEL f1 -----
NUC1      1H
P1        11.00 us
PL1       -1.00 dB
SFO1      300.1318534 MHz

F2 - Processing parameters
SI        32768
SF        300.1299910 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB         0
PC        1.00

```

Figure S53. <sup>1</sup>H NMR of 6k

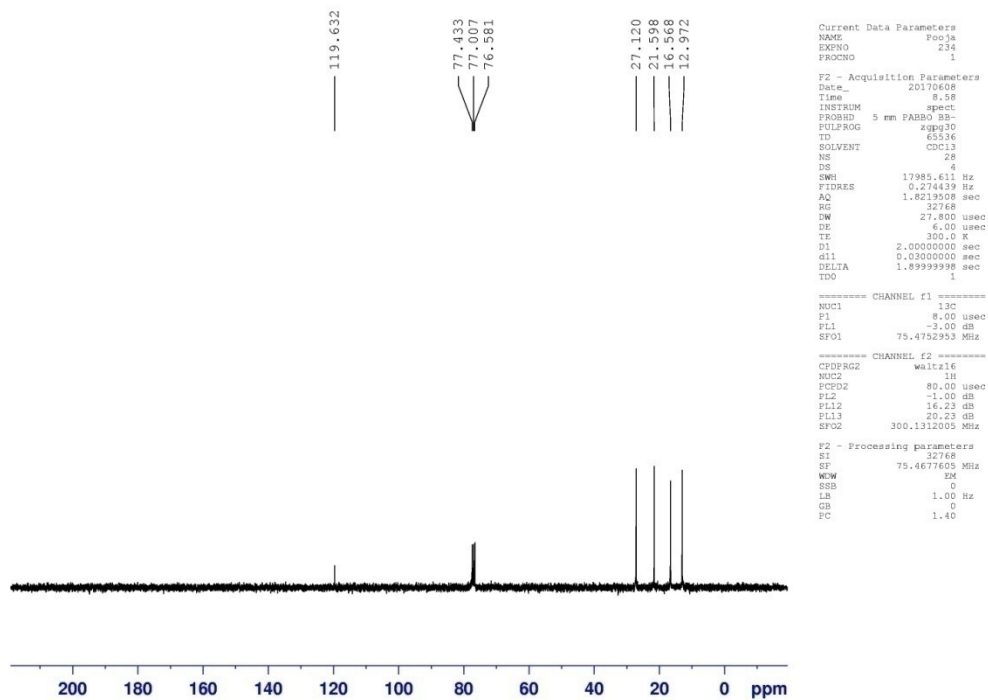


Figure S54. <sup>13</sup>C NMR of 6k

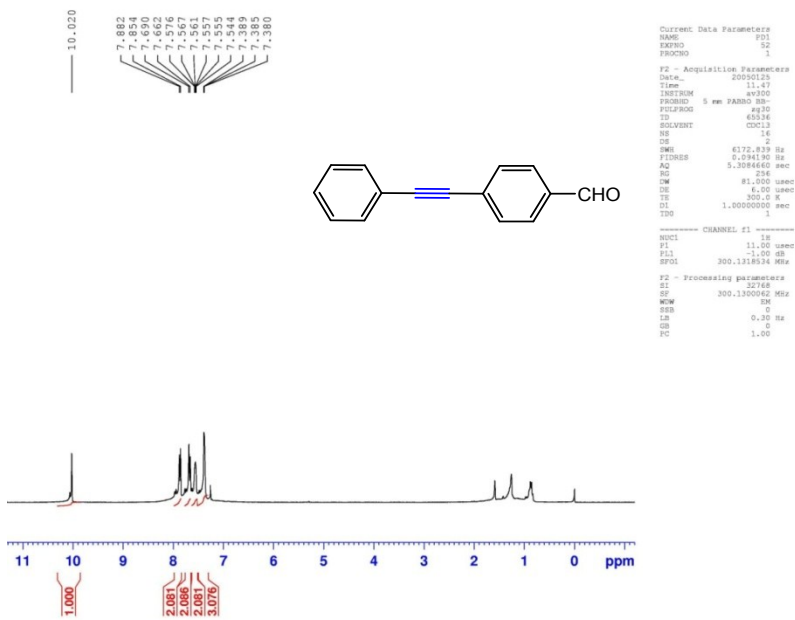
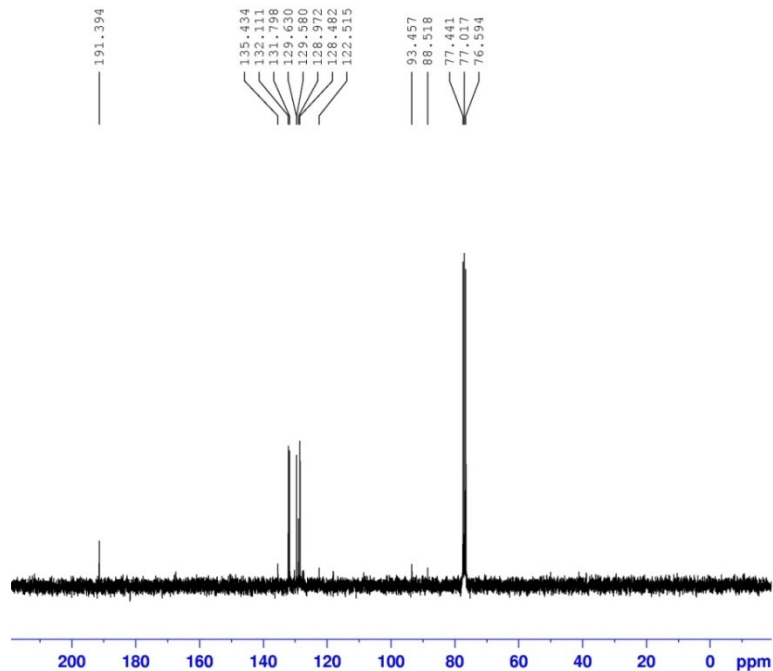


Figure S55. <sup>1</sup>H NMR of 9a



```

Current Data Parameters
NAME          F01
EXPNO         55
PROCNO        1

F2 - Acquisition Parameters
Date_         20050125
Time          13.47
INSTRUM       av300
PROBHD        5 mm PABBO BB-
PULPROG       zg30
TD            65536
SOLVENT       CDCl3
NS            175
DS            4
SWH           17985.611 Hz
FIDRES        0.274439 Hz
AQ            1.8219508 sec
RG            32768
DM            27.800 usec
DE            6.00 usec
TE            300.0 K
D1            2.00000000 sec
d11           0.03000000 sec
DELTA         1.89999998 sec
TDO           1

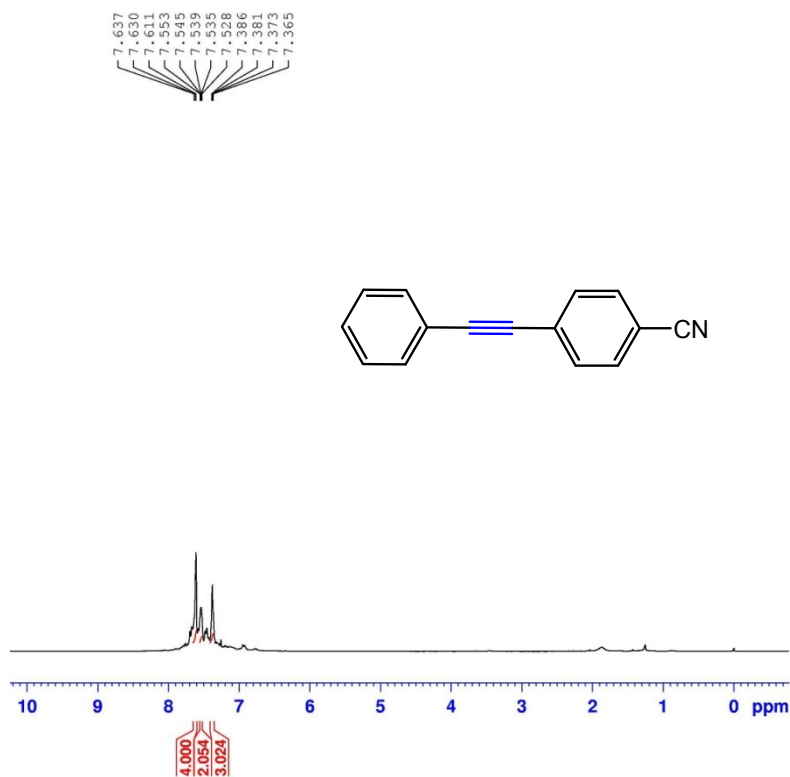
===== CHANNEL f1 =====
NUC1          13C
P1            8.00 usec
PL1           -3.00 dB
SFO1          75.4752933 MHz

===== CHANNEL f2 =====
CPDPRG2       waltz16
NUC2          1H
F2RES         80.00 usec
PL2           -1.00 dB
PL12          16.23 dB
PL13          20.23 dB
SFO2          300.1312005 MHz

F2 - Processing parameters
SI            32768
SF            75.4677490 MHz
KW           EM
SSB           0
LB            1.00 Hz
GB            0
PC            1.40

```

Figure S56. <sup>13</sup>C NMR of 9a



```

Current Data Parameters
NAME          9a
EXPNO         259
PROCNO        1

F2 - Acquisition Parameters
Date_         20170609
Time          12.21
INSTRUM       spect
PROBHD        5 mm PABBO BB-
PULPROG       zg30
TD            65536
SOLVENT       CDCl3
NS            15
DS            2
SWH           6172.839 Hz
FIDRES        0.094190 Hz
AQ            5.3084660 sec
RG            161.3
DM            81.000 usec
DE            6.00 usec
TE            300.0 K
D1            1.00000000 sec
TDO           1

===== CHANNEL f1 =====
NUC1          1H
P1            11.00 usec
PL1           -1.00 dB
SFO1          300.1318524 MHz

F2 - Processing parameters
SI            32768
SF            300.1300071 MHz
KW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            1.00

```

Figure S57. <sup>1</sup>H NMR of 9b



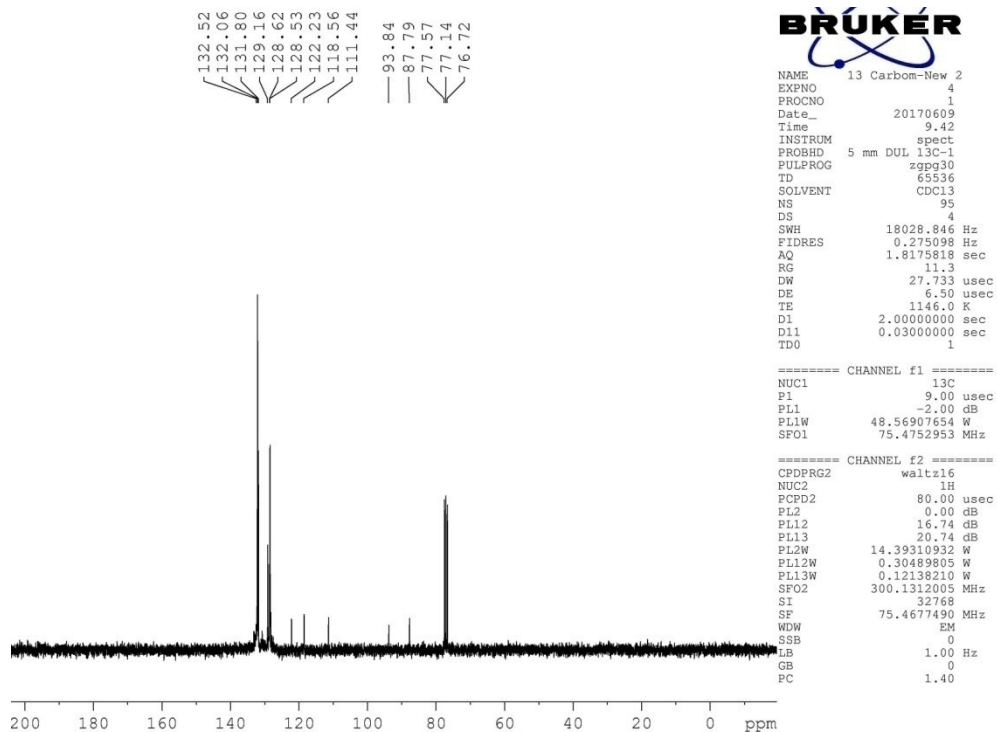


Figure S58. <sup>13</sup>C NMR of 9b

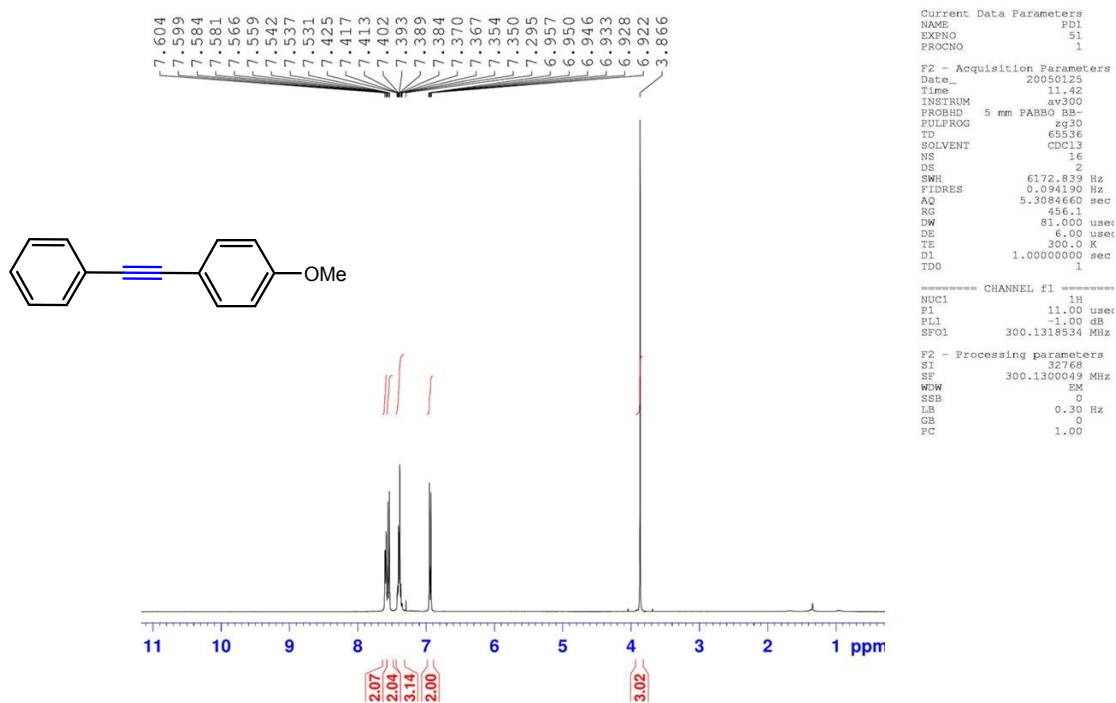


Figure S59. <sup>1</sup>H NMR of 9c



Figure S60.  $^{13}\text{C}$  NMR of 9c

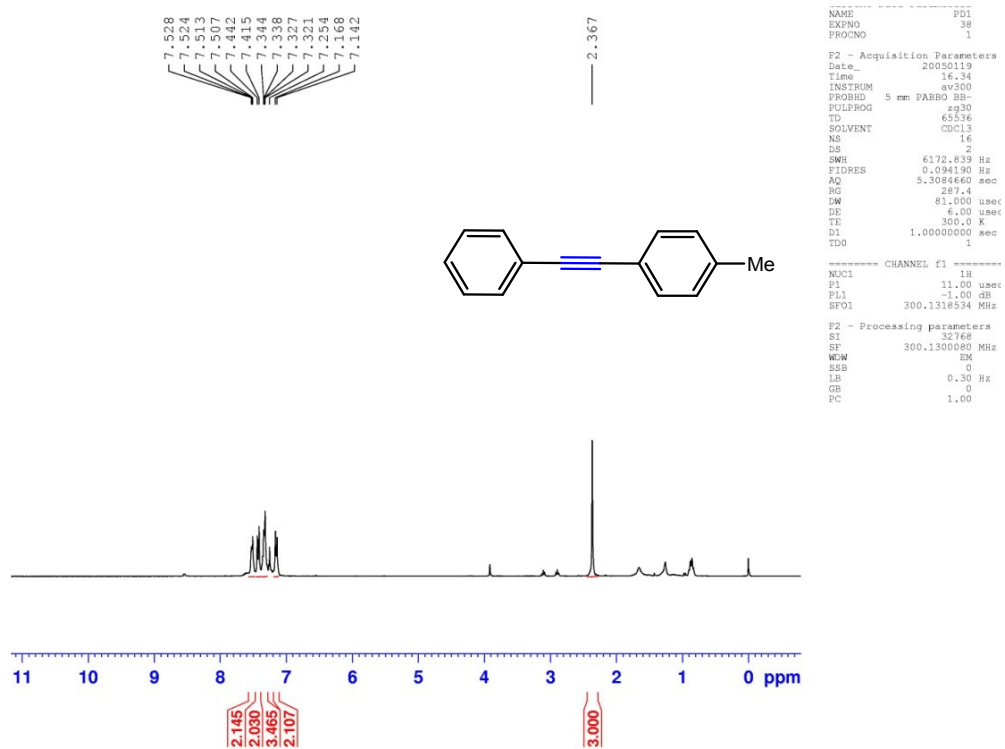
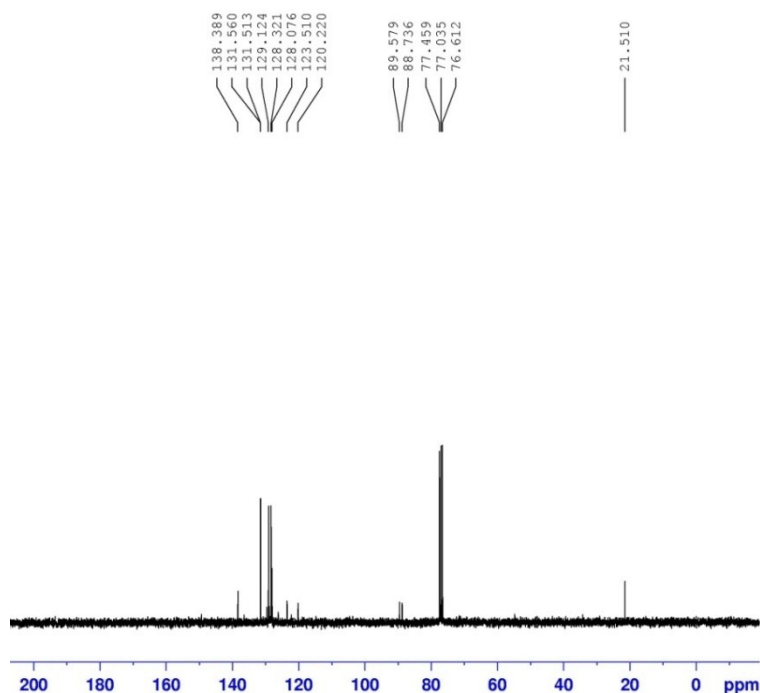


Figure S61.  $^1\text{H}$  NMR of 9d



```
Current Data Parameters
NAME      PDI
EXPNO    39
PROCNO   1

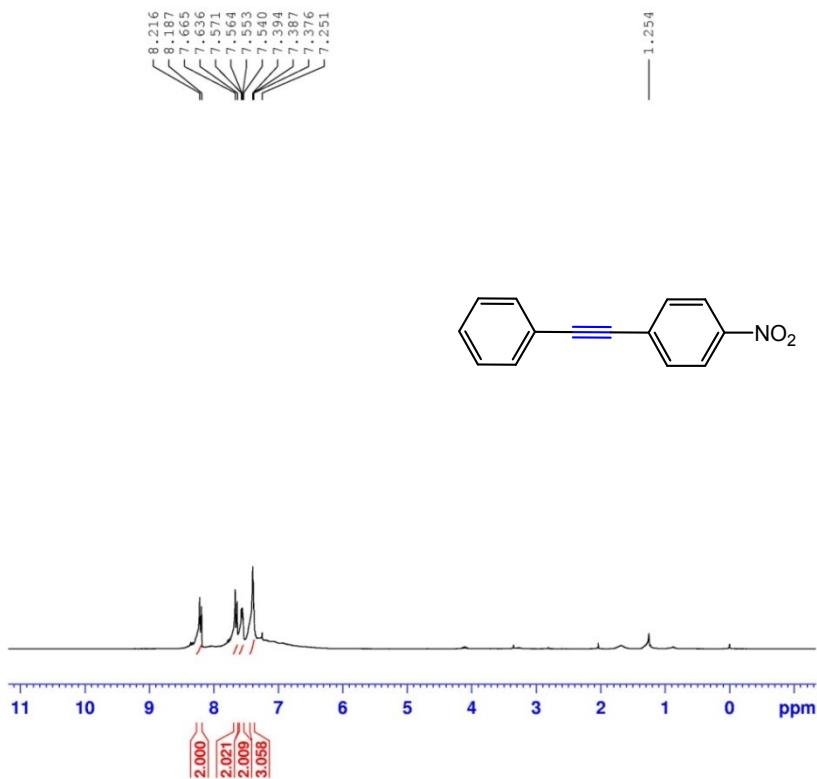
F2 - Acquisition Parameters
Date_    20050120
Time     16.39
INSTRUM  av300
PROBHD   5 mm FAPB0 B1-
PULPROG  zgpg30
TD        65536
SOLVENT  CDCl3
NS        92
DS        4
SWH       17985.611 Hz
FIDRES    0.274439 Hz
AQ        1.8219508 sec
RG        32768
DM        27.800 usec
DE        6.00 usec
TE        300.0 K
D1        2.00000000 sec
d11       0.02000000 sec
DELTA    1.89999998 sec
TD0       1
```

```
----- CHANNEL f1 -----
NUC1      13C
P1         8.00 usec
PL1        -3.00 dB
SFO1      75.4752953 MHz

----- CHANNEL f2 -----
CFDPRG2   waitz16
NUC2       1H
PCPD2     80.00 usec
PL2        -1.00 dB
PL12       16.23 dB
PL13       20.23 dB
SFO2      300.1312005 MHz
```

```
F2 - Processing parameters
SI         32768
SF         75.4677490 MHz
WVW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40
```

Figure S62. <sup>13</sup>C NMR of 9d



```
Current Data Parameters
NAME      POOJA DUBEY
EXPNO    339
PROCNO   1
```

```
F2 - Acquisition Parameters
Date_    20160722
Time     12.39
INSTRUM  av300
PROBHD   5 mm FAPB0 B1-
PULPROG  zg30
TD        65536
SOLVENT  CDCl3
NS        13
DS        2
SWH       6172.839 Hz
FIDRES    0.094190 Hz
AQ        5.3084660 sec
RG        128
DM        81.000 usec
DE        6.00 usec
TE        300.0 K
D1        1.00000000 sec
TD0       1
```

```
----- CHANNEL f1 -----
NUC1      1H
P1         11.00 usec
PL1        -1.00 dB
SFO1      300.1316534 MHz
```

```
F2 - Processing parameters
SI         32768
SF         300.1300086 MHz
WVW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
```

Figure S63. <sup>1</sup>H NMR of 9e

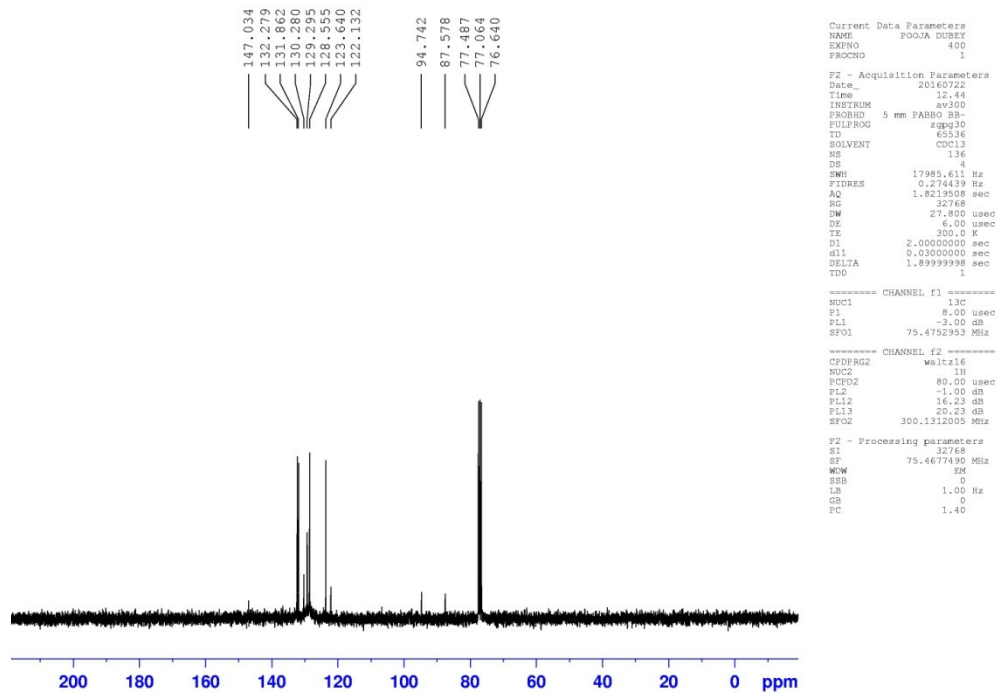


Figure S64. <sup>13</sup>C NMR of 9e

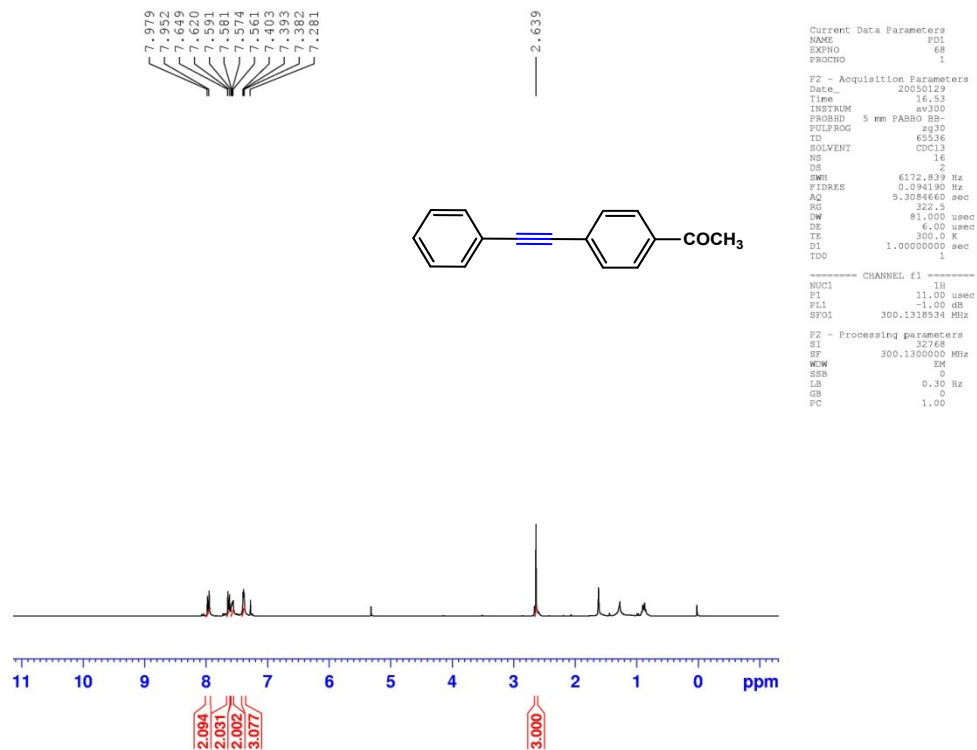


Figure S65. <sup>1</sup>H NMR of 9f

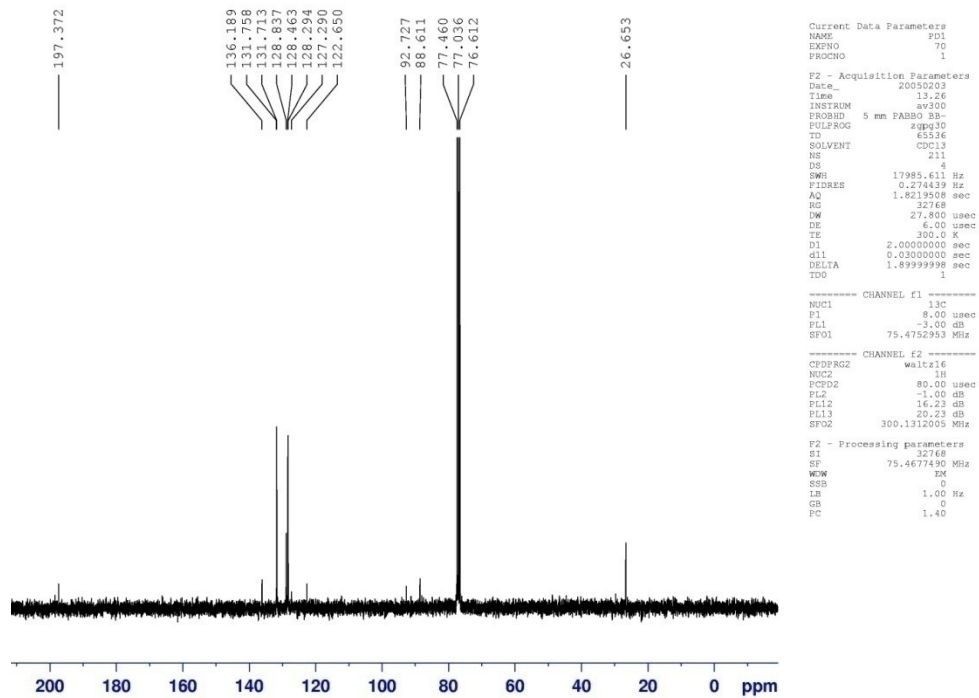


Figure S66. <sup>13</sup>C NMR of 9f

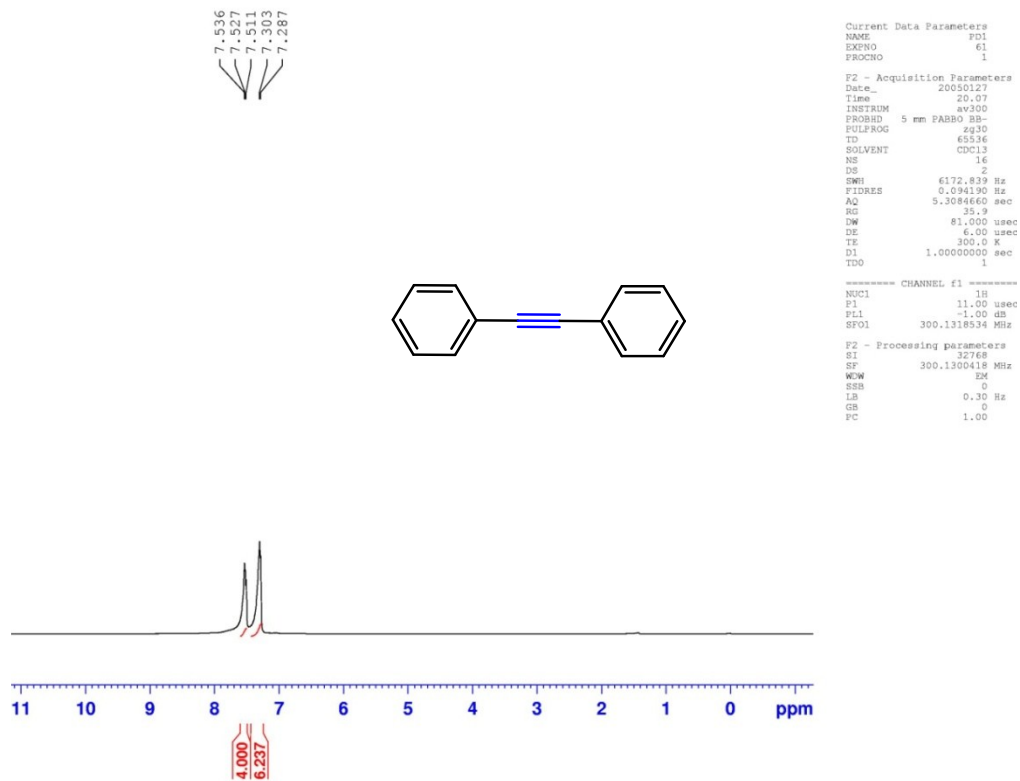


Figure S67. <sup>1</sup>H NMR of 9g

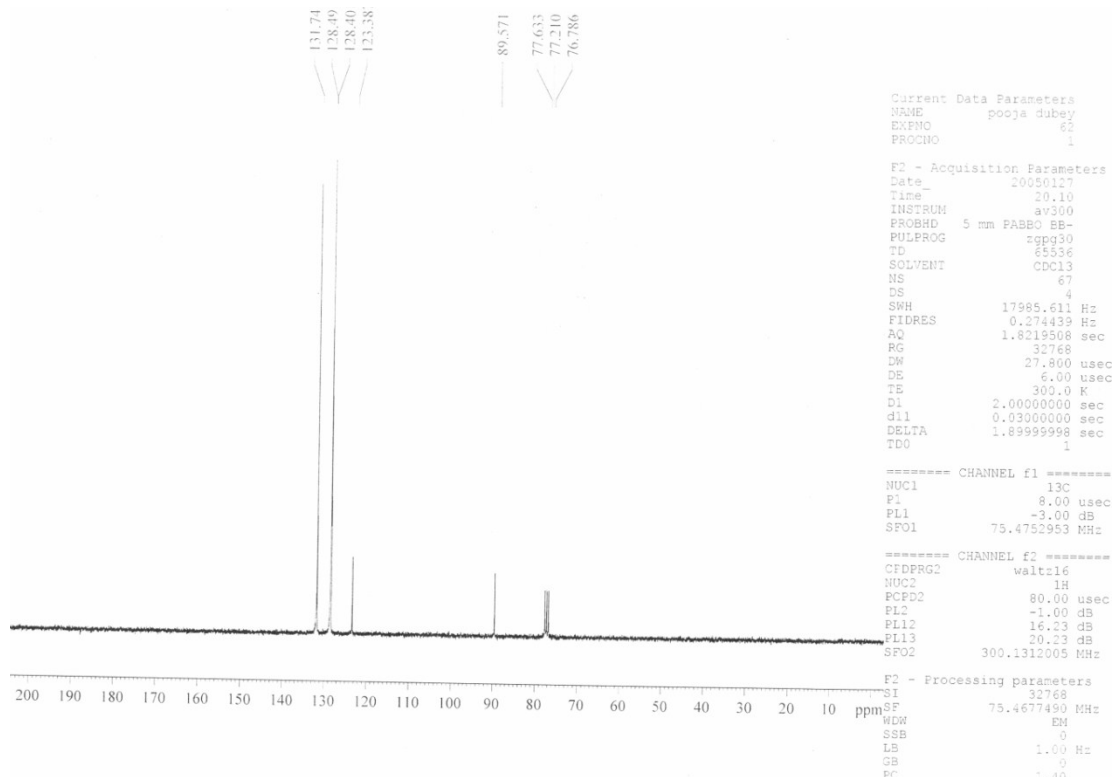


Figure S68. <sup>13</sup>C NMR of 9g

HR-MS Spectra of L1-L2 and Complexes 1-2

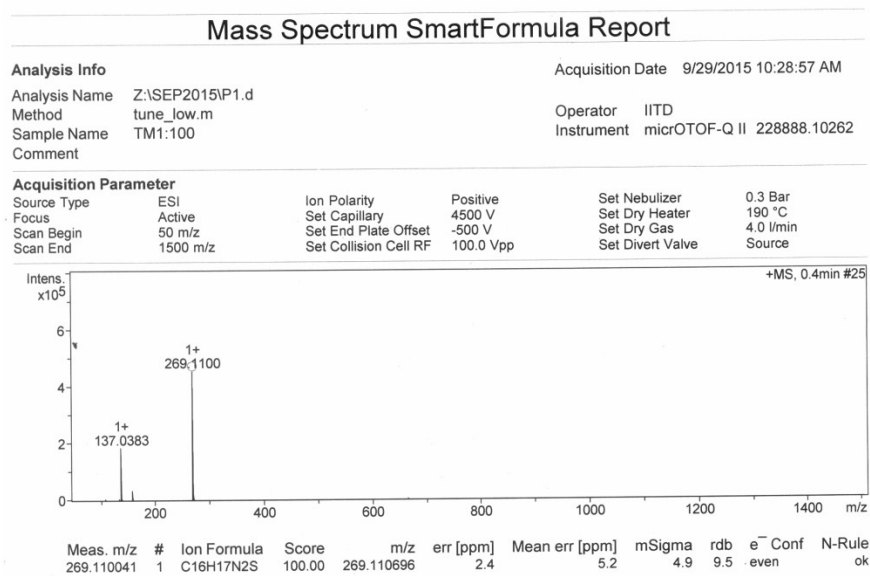
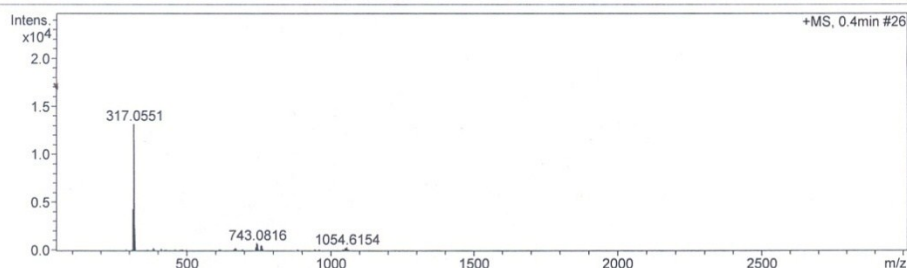


Figure S69. Mass Spectra of L1

## Mass Spectrum SmartFormula Report

<b>Analysis Info</b>		Acquisition Date	2/13/2014 11:30:23 AM
Analysis Name	D:\Data\Feb_2014\PUJA2.d	Operator	Sharma/Singh
Method	tune_wide.m	Instrument / Ser#	micrOTOF-Q II 10262
Sample Name	TM 1:100		
Comment			

<b>Acquisition Parameter</b>					
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.3 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	180 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set Collision Cell RF	600.0 Vpp	Set Divert Valve	Source



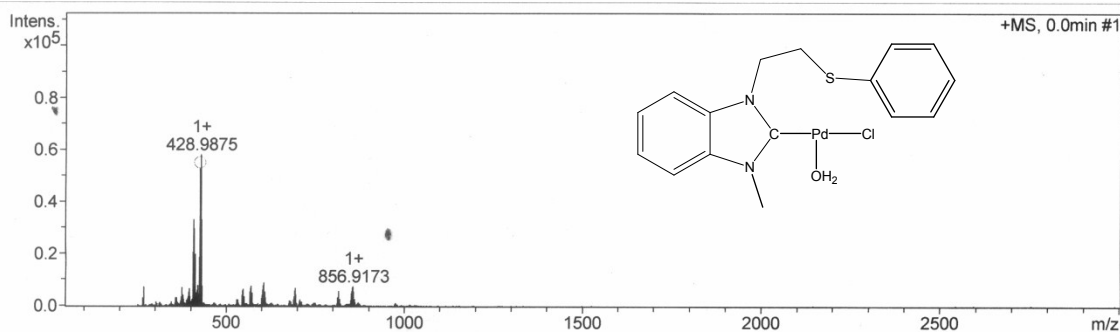
Meas. m/z	#	Formula	m/z	err [ppm]	Mean err [ppm]	rdB	N-Rule	e <sup>-</sup> Conf	mSigma	Std I	Std Me an m/z	Std I Var Nor m	Std Diff	Std Comb Dev
317.0551	1	C <sub>16</sub> H <sub>17</sub> N <sub>2</sub> Se	317.0552	0.4	3.2	9.5	ok	even	76.2	110.7	1.4	25.1	1.5	842.7

**Figure S70. Mass Spectra of L2**

## Mass Spectrum SmartFormula Report

<b>Analysis Info</b>		Acquisition Date	7/20/2016 12:04:01 PM
Analysis Name	Z:\JULY2016\laks p2.d	Operator	IITD
Method	tune_wide.m	Instrument	micrOTOF-Q II 228888.10262
Sample Name	Ayesha_5		
Comment			

<b>Acquisition Parameter</b>					
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.3 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	180 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set Collision Cell RF	500.0 Vpp	Set Divert Valve	Source



Meas. m/z	#	Ion Formula	m/z	err [ppm]	Mean err [ppm]	rdB	N-Rule	e <sup>-</sup> Conf	mSigma	Std I	Std Me an m/z	Std I Var Nor m	Std Diff	Std Comb Dev
426.986313	1	C <sub>16</sub> H <sub>18</sub> CIN <sub>2</sub> OPdS	426.986065	0.6	-3.8	8.5	ok	even	19.2	16.1	2.2	3.6	1.6	842.7

# Display Report

## Analysis Info

Analysis Name Z:\JULY2016\aks p2.d  
Method tune\_wide.m  
Sample Name Ayesha\_5  
Comment

Acquisition Date 7/20/2016 12:04:01 PM

Operator IITD  
Instrument micrOTOF-Q II 228888.10262

## Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.3 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	180 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set Collision Cell RF	500.0 Vpp	Set Divert Valve	Source

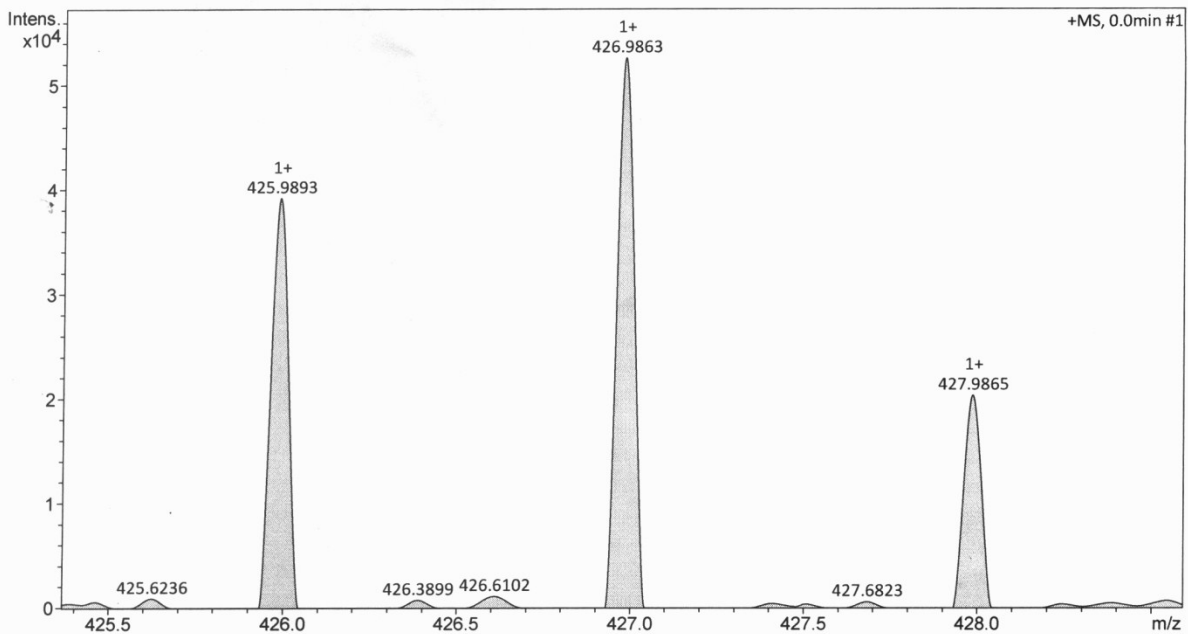
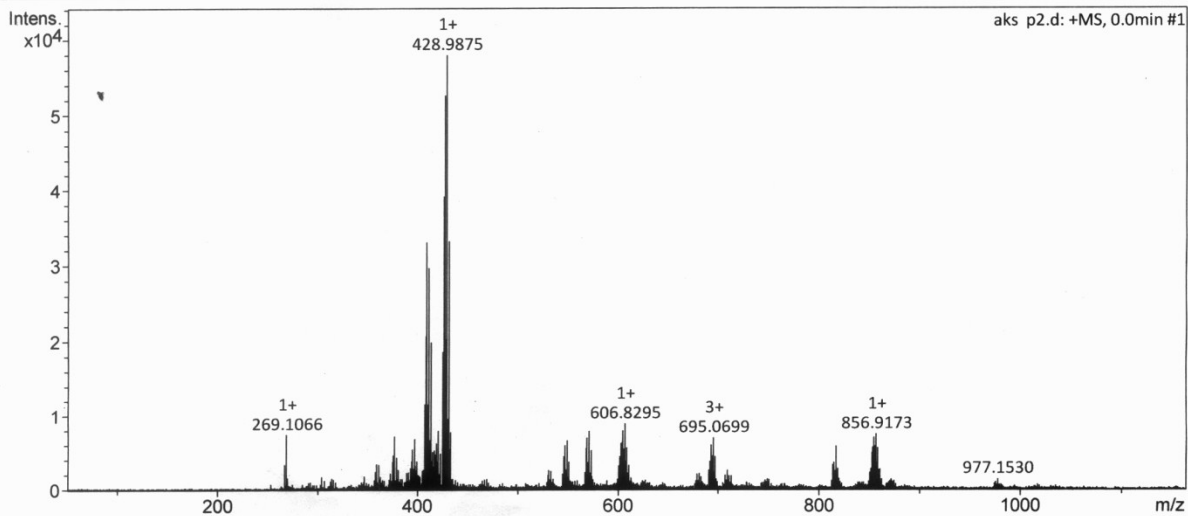


Figure S71. Mass Spectra of complex 1



## Mass Spectrum SmartFormula Report

### Analysis Info

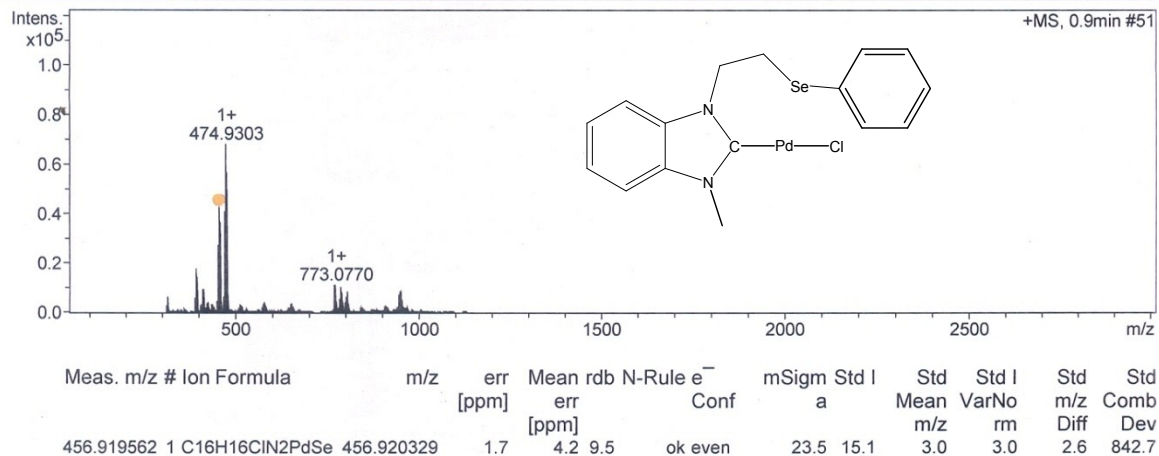
Analysis Name Z:\Aug\_2014\PD6.d  
 Method tune\_wide.m  
 Sample Name TM1:100  
 Comment

Acquisition Date 8/5/2014 10:20:55 AM

Operator Sharma/Singh  
 Instrument micrOTOF-Q II 228888.10262

### Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.3 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	180 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set Collision Cell RF	600.0 Vpp	Set Divert Valve	Source



**Figure S72. Mass Spectra of complex 2**

The NMR spectral data of compounds found as reported in literature.<sup>1-8</sup>

**Benzamide**:<sup>1,7</sup> White solid. (Table 3, entry 4a), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 7.80-7.82 (d, *J* = 6 Hz, 2H), 7.55-7.44 (m, 3H), 6.23 (bs, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 169.6, 133.4, 131.9, 128.6, 127.3.

**4-Methylbenzamide**:<sup>1,7</sup> White solid. (Table 3, entry 4b), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 7.72-7.70 (d, *J* = 6 Hz, 2H), 7.26-7.23 (d, *J* = 9 Hz, 2H), 6.03 (bs, 2H), 2.40 (s, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 169.4, 142.5, 130.5, 129.2, 127.3, 21.4.

**4-Methoxybenzamide**:<sup>1</sup> White solid. (Table 3, entry 4c), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 7.81-7.79 (d, *J* = 6 Hz, 2H), 6.96 (d, *J* = 8.7 Hz, 2H), 5.96 (bs, 2H), 3.88 (s, 3H). <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 168.9, 162.6, 129.2, 125.5, 113.8, 55.4.

**4-Iodobenzamide.**<sup>10a</sup> White solid. (Table 3, entry 4d), <sup>1</sup>H NMR (300 MHz, d<sub>6</sub>-DMSO): δ 8.01 (b, 1H), 7.84-7.82 (d, *J* = 6 Hz, 2H), 7.66-7.64 (d, *J* = 6 Hz, 2H), 7.42 (b, 1H). <sup>13</sup>C NMR (75 MHz, d<sub>6</sub>-DMSO): δ 167.7, 137.5, 134.1, 129.9, 99.3.

**4-Bromobenzamide:**<sup>1</sup> White solid. (Table 3, entry 4e), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 7.70-7.63 (d, *J* = 12 Hz, 2H), 7.60-7.54 (d, *J* = 9, 2H), 5.91 (bs, 2H). <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 168.5, 131.8, 131.8, 128.9, 128.7.

**4-Chlorobenzamide:**<sup>1</sup> White solid. (Table 3, entry 4f), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 7.82-7.73 (d, *J* = 3 Hz, 2H), 7.47-7.40 (d, *J* = 3, 2H), 5.93 (bs, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 168.2, 138.3, 131.7, 128.9, 128.7.

**4-Fluorobenzamide:**<sup>1</sup> White solid. (Table 3, entry 4g), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 7.86-7.81 (m, 2H), 7.15-7.10 (t, *J* = 8.4, 2H), 5.93 (bs, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 168.3, 166.7, 129.8, 129.6, 115.8.

**2-Phenylacetamide:**<sup>7</sup> White powder. (Table 3, entry 4h), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 7.38 – 7.28 (m, 5H), 5.79-5.41 (m, 2H), 3.58 (s, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 173.9, 134.8, 129.3, 129.0, 43.0

**2-Bromobenzamide:**<sup>1,4</sup> White solid. . (Table 3, entry 4i), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 7.49 (s, 1H), 7.75-7.62 (m, 2H), 7.42-7.29 (m, 2H), 6.27-6.18 (bs, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 169.5, 137.1, 133.5, 131.6, 129.8, 127.5, 119.1.

**4-Pyridine carboxamide:**<sup>10b</sup> White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): 8.37-8.20 (2H, d, *J* 12), 8.05 (1H, s) δ 7.74 (2H, d *J* 6).

**Pentanamide:**<sup>1</sup> White solid. (Table 3, entry 6k), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 0.97-0.92 (t, *J* = 7.2, 3H), 1.25-1.40 (m, 2H), 1.56-1.64 (t, *J* = 7.5 2H), 2.20-2.24 (m, 2H), 5.99 (bs, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 176.5, 35.6, 27.5, 22.3, 13.7.

**Benzonitrile:**<sup>2,8</sup> Colourless oil. (Table 4, entry 6a), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 7.58 (m, 3H), 7.43 (m, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 132.8, 132.0, 129.1, 118.8, 112.9.

**4-Methylbenzonitrile**:<sup>2,8</sup> Colourless oil. (Table 4, entry 6b), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 7.47-7.50 (d, *J* = 9 Hz, 2H), 7.22-7.25 (d, *J* = 9 Hz, 2H), 2.38 (s, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 143.4, 131.6, 129.5, 118.8, 108.9, 21.4.

**4-Methoxybenzonitrile**:<sup>2,8</sup> White solid. (Table 4, entry 6c), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 7.59-7.56 (d, *J* = 3 Hz, 2H), 6.96-6.93 (d, *J* = 3 Hz, 2H), 3.86 (s, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 162.6, 133.2, 119.2, 114.7, 103.9, 55.5.

**4-Iodobenzonitrile**:<sup>10c</sup> White solid. (Table 4, entry 6d), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 7.86-7.83 (m, 2H), 7.38-7.26 (m, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 138.4, 133.0, 118.1, 111.6, 100.2.

**4-Bromobenzonitrile**:<sup>2,8</sup> White solid. (Table 4, entry 6e), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 7.65-7.62 (m, 2H), 7.54-7.51 (m, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 133.3, 132.5, 127.9, 117.9, 111.1.

**4-Chlorobenzonitrile**:<sup>3,8</sup> White solid. (Table 4, entry 6f), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 7.62 (m, 2H), 7.49 (m, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 139.5, 133.3, 129.7, 117.9, 110.7.

**4-Fluorobenzonitrile**:<sup>4,8</sup> White solid. (Table 4, entry 6g), <sup>1</sup>H NMR (300 MHz, DMSO-d<sub>6</sub>, 25 °C vs Me<sub>4</sub>Si): δ 8.01-7.93 (m, 3H), 7.41 (s, 1H), 7.29-7.23 (m, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, DMSO-d<sub>6</sub>, 25 °C vs Me<sub>4</sub>Si): δ 167.3, 166.0, 162.7, 131.2, 131.1, 130.6, 130.5, 115.6, 115.3.

**2-Phenylacetonitrile**:<sup>8</sup> Colourless oil. (Table 4, entry 6h), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 7.42 – 7.35 (m, 5H), 3.75 (s, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 129.8, 129.0, 127.9, 127.8, 117.8, 23.4.

**2-Bromobenzonitrile**:<sup>4,5</sup> White solid. (Table 4, entry 6i), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 7.69 (s, 2H), 7.47 (s, 2H), 6.49 (s, 1H); <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 134.0, 133.8, 133.0, 127.5, 125.1, 117.0, 115.6.

**4-Cyanopyridine**:<sup>10d</sup> White solid; (Table 4, entry 6j) <sup>1</sup>H NMR (300 MHz; DMSO-d<sub>6</sub>) δ: 8.72-8.71 (m, 2H), 8.23 (s, 1H); 7.73-7.72 (m, 3H); <sup>13</sup>C-NMR (75 MHz, DMSO-d<sub>6</sub>) δ: 166.7, 150.6, 141.7, 121.8.

**Pentanenitrile:**<sup>6</sup> Colourless oil. (Table 4, entry 6k), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 2.37-2.32 (t, *J* = 6, 2H), 1.69-1.62 (m, 2H), 1.59-1.42 (m, 2H), 0.93-0.98 (t, *J* = 7.2, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>, 25 °C vs Me<sub>4</sub>Si): δ 119.6, 27.1, 21.5, 16.5, 12.9.

Spectroscopic data of coupled products of Sonogashira cross coupling reactions<sup>9</sup>

**4-(Phenylethynyl) benzaldehyde:** Light yellow solid. (Table 6, entry 9a), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C, TMS); δ (ppm): 7.38 (m, 3H), 7.54-7.57 (m, 2H), 7.66-7.69 (d, 2H, *J* = 9 Hz), 7.85-7.88 (d, 2H, *J* = 9 Hz), 10.02 (s, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>, 25 °C, TMS); δ (ppm): 88.5, 93.4, 122.5, 128.4, 128.9, 129.5, 129.6, 131.7, 132.1, 135.4, 191.3.

**4-(Phenylethynyl)benzonitrile:** Yellow solid. (Table 2, entry 3), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C, TMS); δ (ppm): 7.36–7.38 (m, 3H), 7.38–7.54 (m, 2H), 7.55–7.63 (m, 4H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>, 25 °C, TMS); δ (ppm): 87.9, 93.8, 111.4, 118.5, 122.2, 128.5, 128.6, 129.1, 131.8, 132.0, 132.5.

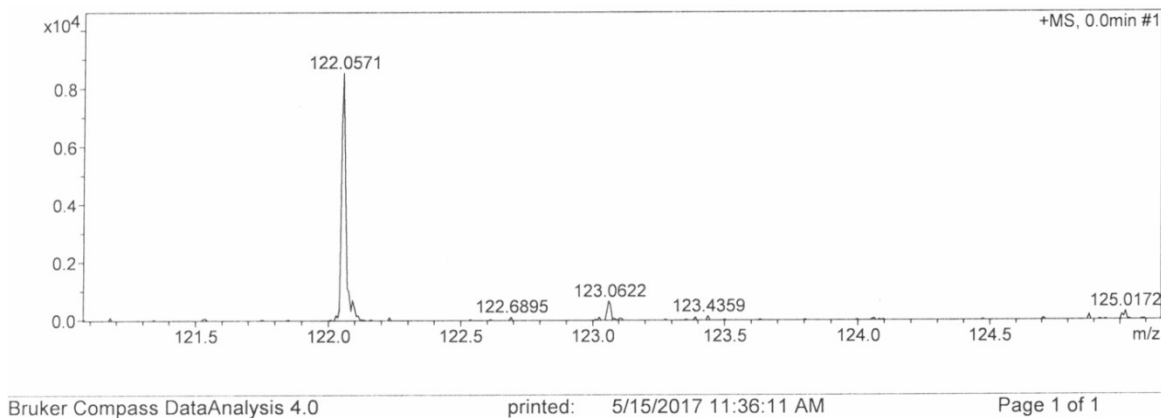
**1-Methoxy-4-(phenylethynyl)benzene:** Light-yellow solid. (Table 6, entry 9c), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C, TMS); δ (ppm): 3.87 (s, 3H), 6.96-6.92 (m, 3H), 7.43-7.35 (m, 2H), 7.60-7.53 (m, 2H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>, 25 °C, TMS); δ (ppm): 55.3, 88.2, 89.5, 114.1, 115.4, 123.6, 128.0, 128.3, 133.1, 131.5, 159.6.

**1-Methyl-4-(phenylethynyl)benzene:** White solid. (Table 6, entry 9d), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C, TMS); δ (ppm): 2.36 (s, 3H), 7.14-7.16 (d, *J* = 6 Hz, 2H), 7.32-7.34 (m, 3H), 7.41-7.44 (d, 2H, *J* = 9 Hz), 7.50-7.52 (m, 2H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>, 25 °C, TMS); δ (ppm): 21.5, 88.7, 89.5, 120.2, 123.5, 128.0, 128.3, 129.1, 131.5, 131.5, 138.3.

**1-Nitro-4-(phenylethynyl)benzene:** Light-yellow solid. (Table 6, entry 9e) <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C, TMS); δ (ppm): 7.25-7.39 (m, 3H), 7.54-7.56 (m, 2H), 7.63-7.66 (d, *J* = 9, 2H), 8.18-8.21 (d, 2H, *J* = 9 Hz); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>, 25 °C, TMS); δ (ppm): 87.5, 94.7, 121.1, 123.6, 128.5, 129.2, 130.2, 131.8, 132.2, 147.0.

**4-acetyl-diphenylacetylene:** White Solid. (Table 6, entry 9f), <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C, TMS) δ = 2.63 (s, 3H), δ = 7.38-7.40 (m, 3H), 7.56-7.59 (m, 2H), 7.62-7.64 (d, *J* = 6 Hz, 2H), 7.95-7.97 (d, *J* = 6 Hz, 2H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>, 25 °C, TMS); δ (ppm): 26.6, 88.6, 92.7, 122.6, 127.2, 128.2, 128.4, 128.8, 131.7, 131.7, 136.1, 197.3.

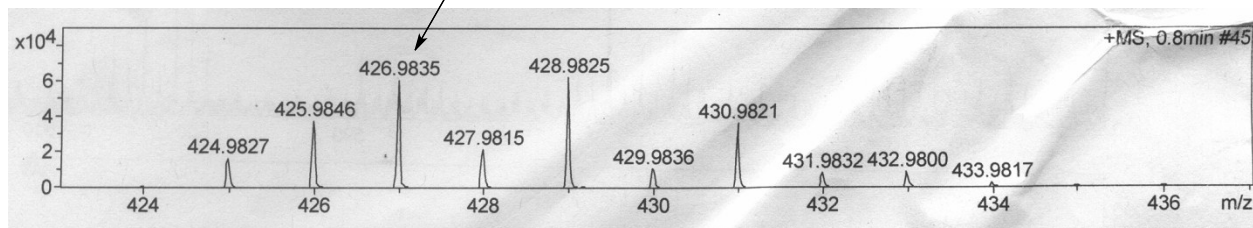
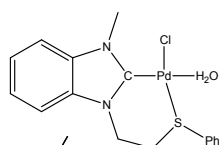
**1,2 Diphenylethyne:** white solid. (Table 2, entry 7),  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ , 25 °C, TMS) :  $\delta$  7.28–7.30 (m, 6H), 7.52–7.53 (d, 4H).  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ , 25 °C, TMS);  $\delta$  (ppm): 89.5, 123.3, 128.4, 128.4, 131.7.



**Figure S73. Mass spectra of reaction mixture**

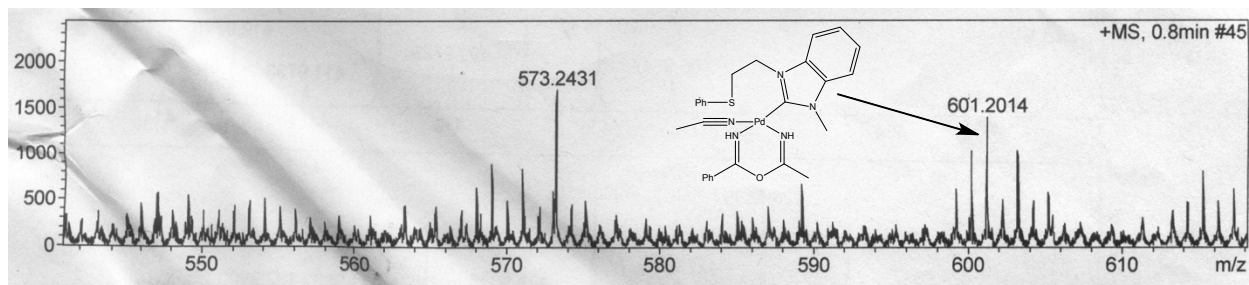
**ESI-MS Data**

Molecular formula	Exact mass	Mass found
$\text{C}_{16}\text{H}_{18}\text{ClN}_2\text{OPdS}$	426.98	426.98



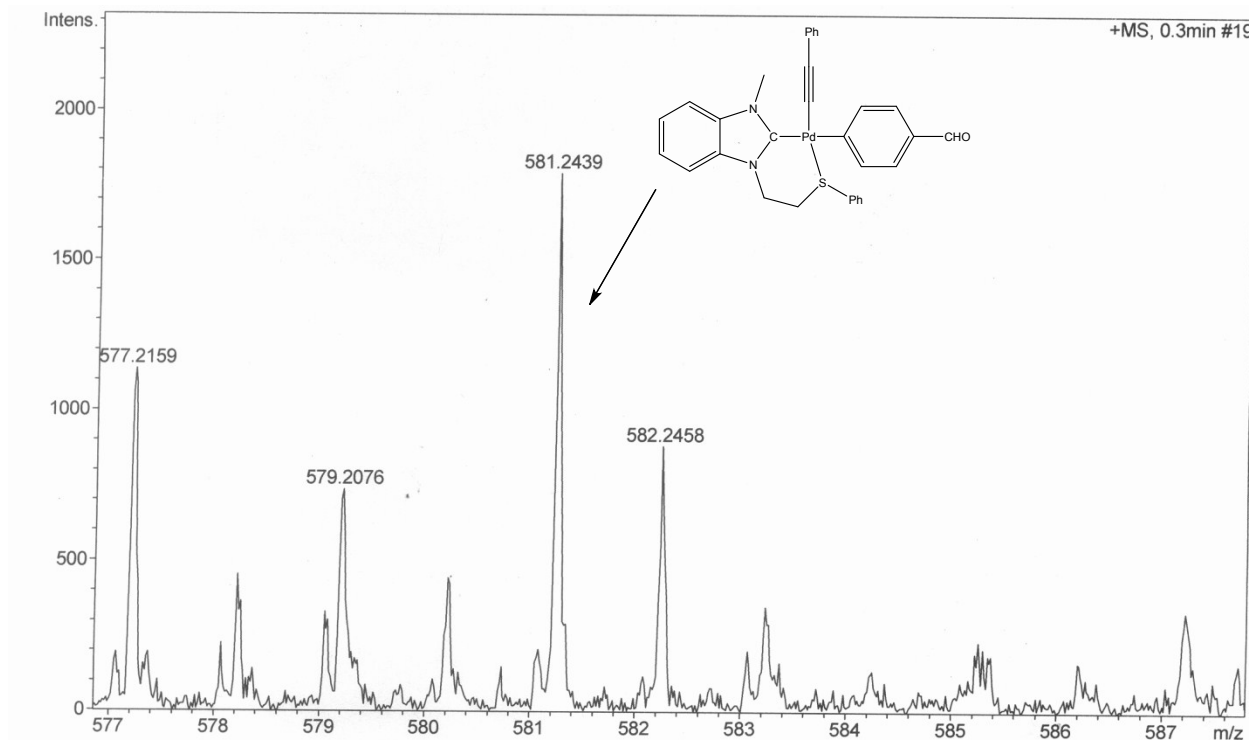
**Figure S74. ESI-MS of  $[\text{Pd}(\text{II})(\text{Cl})(\text{L})(\text{H}_2\text{O})]^+$**

Molecular formula	Exact mass	Mass found
$\text{C}_{27}\text{H}_{30}\text{N}_5\text{ONaPdS}$	601.11	601.20



**Figure S75. ESI-MS of [Intermediate F]+Na+H]<sup>2+</sup>**

Molecular formula	Exact mass	Mass found
C <sub>31</sub> H <sub>27</sub> N <sub>2</sub> OPdS	581.08	581.24



**Figure S76. ESI-MS of Intermediate [Intermediate H]+H]<sup>+</sup>**

## Two-phase test

A mixture of 4-bromobenzoic acid-immobilized silica (0.20 g) (prepared by reported methods),<sup>11</sup> phenyl acetylene (0.224 g, 2.2 mmol), 4-bromobenzaldehyde (0.185 g, 1.0 mmol), and K<sub>2</sub>CO<sub>3</sub> (0.441 g, 3.0 mmol) were heated in an inert atmosphere at 90 °C for 8 h in dry DMSO (1 mL) in the presence of 1 mol% of **1**. After completion of the reaction, the mixture was cooled and filtered through a G-4 crucible. The residue left in the crucible was washed with 20 mL of water followed by diethyl ether (2×20 mL). The filtrate and washings were collected together. The resulting mixture was extracted with 30 mL of diethyl ether. The solvent of the extract was evaporated and the residue was analyzed with <sup>1</sup>H-NMR. The yield of the cross-coupled product, 4-(phenylethynyl)benzaldehyde was ~92%. The residue in G-4 crucible was hydrolysed with KOH (1.68 g dissolved in 10 mL of EtOH + 5 mL of H<sub>2</sub>O) at 90 °C for 3 days. The hydrolysed solution was neutralized with aqueous 20% (v/v) HCl and, extracted with dichloromethane (30 mL) followed by ethyl acetate (40 mL). The organic phases were combined together and its solvent was evaporated off. The hydrolyzed products were analyzed by <sup>1</sup>H NMR spectroscopy. Of the immobilized 4-bromobenzoic acid (as amide), nearly 77% (by <sup>1</sup>H NMR) was converted into the cross-coupled product, 4-(phenylethynyl)benzoic acid.

## Reference

1. Joshi, H.; Sharma, K. N.; Sharma, A. K.; Prakash, O.; Kumar A.; Singh, A. K.; *Dalton Trans.* **2014**, *43*, 12365.
2. Yin W.; Wang C.; Huang Y.; *Org. Lett.* **2013**, *15*, 1850.
3. Rokade, B. V.; Prabhu K. R.; *J. Org. Chem.* **2012**, *77*, 5364.
4. Dornan, L. M.; Cao, Q.; Flanagan, J. C. A.; Crawford, J. J.; Cook, M. J.; Muldoon, M. J.; *Chem. Comm.* **2013**, *49*, 6030.
5. Molla, R. A.; Tuhina, K. K.; Islam, S. M.; *New J. Chem.* **2015**, *39*, 921.
6. Duret, P. G.; Blanchard, N.; *Org. Chem. Front.* **2014**, *1*, 825.
7. Matsuoka, A.; Isogawa, T.; Morioka, Y.; Knappett, B. R.; Wheatley, A. E.; Saito, H. S.; Naka, H.; *RSC Adv.*, **2015**, *5*, 12152.
8. Dighe, S. U.; Chowdhury, D.; Batra. S.; **2014**, 356, 18.

9. (a) Elangovan, A.; Wang, Y-H.; Ho, T-I.; *Org. Lett.* **2003**, 5, 1841. (b) Kumar, S.; Saleem, F.; Singh, A. K.; *Dalton Trans.* **2016**, 45, 11445. (c) Singh, V.V.; Singh, A. K.; *Dalton Trans.* **2015**, 44, 725.
10. (a) Haonan Chen, Wujie Dai, a Yi Chen, a Qing Xu, Jianhui Chen, Lei Yu, Yajuan Zhao, Mingde Yea and Yuanjiang Pan. *Green Chem.*, 2014, 16, 2136-2141. (b) Claudio Battilocchio, Joel M. Hawkins, and Steven V. Ley *Org. Lett.* 2014, 16, 1060–1063. (c) Valizadeh<sup>1</sup>, H.; Noorshargh<sup>1</sup>, S.; Shomali, A. *Synthetic Communications* DOI: 10.1080/00397911.2014.994129. (d) Tanuwidjaja, J.; Peltier, H. M.; Lewis, J. C.; Schenkel, L. B.; Ellman, J. A. *Microwave-Promoted Synthesis of Nitriles*, **2007**, 21 3385-3389.
11. Webb, J. D.; MacQuarrie, S.; McEleney, K.; Crudden, C. M. *J. Catal.*, **2007**, 252, 97.