Supporting information

Synthesis and application of a water-soluble phosphorescent iridium complex as turn-on sensing material for human serum albumin

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The NMR spectrum of "Ir"

Fig. S1.H¹-NMR spectra ofIr

H¹-NMR (400 MHz, DMSO-d6), δ(ppm): 8.90 (d, J = 4.0 Hz, 1H), 8.66 (d, J = 8.0 Hz, 2H), 8.58 (d, J = 7.9 Hz, 1H), 8.45 (s, 2H), 8.32 – 8.22 (m, 4H), 8.16 (d, J = 8.2 Hz, 2H), 8.09 (td, J = 7.8, 1.7 Hz, 2H), 7.86 – 7.82 (m, 2H), 7.77 (ddt, J = 4.9, 3.5, 1.4 Hz, 2H), 7.71 – 7.67 (m, 2H), 7.58 – 7.52 (m, 4H), 7.37 – 7.23 (m, 6H).



Fig. S2.C¹³-NMR spectra ofIr

C¹³-NMR (101 MHz, DMSO), δ(ppm):167.68, 160.38, 156.08, 155.82, 155.63, 152.41, 150.24, 148.90, 148.78, 148.42, 137.94, 137.79, 137.53, 134.99, 134.84, 130.21, 129.97, 129.94, 129.83, 129.79, 128.99, 127.99, 127.78, 127.75, 127.57, 127.22, 127.03, 126.96, 126.45, 125.82, 125.45, 123.93, 123.49, 119.12.

Density functional theory (DFT) calculation



Fig.S3.The HOMO level and LUMO level of **Ir**in basic solution (a) and in acidic solution (b).

pKa of Ir

We can use the following equilibrium from the literature¹ to describe the relationship between the pH value and the phosphorescence intensity at λ_{ex} =338 nm quantitatively.

$$pKa = pH - log \left[\frac{I_{max} - I_x}{I_x - I_{min}} \right]$$
(3)

In the equilibrium, I_{min} and I_{max} represent the phosphorescence intensity of the probe in its acid and conjugate base, respectively. In Fig.S4, In wedescribed the linear regression relationship equilibrium (3) and pH value with the function of I =0.16473 pH-0.2857 and R² = 0.9941. According to the equation in the literature^{2, 3}, we calculate that pKa1 and pKa2 is 2.51 and 3.23 which stems from carboxylic acid. Similarly, pKa3 and pKa4 is calculated to be 4.60 and 6.02 which belongs to ammonium ion.



Fig. S4 (a) PL intensityat 634 nmversus pH, λ_{ex} =338nm. (b) The linear relationship of the phosphorescence intensity with pH value.

pH response



Fig. S5. The pH effect on the phosphorescence intensity ratio of d $I_{Ir \cdot HSA}/I_{Ir}$ in BR

solution.

Phosphorescence turn-on detection of Ir • HSA



Fig. S6. The pH effect on the phosphorescence intensity ratio of d $I_{Ir \cdot HSA}/I_{Ir}$ in BR solution.



Fig. S7. Job's plots of phosphorescence intensities as functions of the molar fraction of HSA in BR solution at pH 2.0. The total concentration c(HSA) + c(Ir) iskept constant at 10 μ M. The inflection points of job's plot give a value of 0.67, which correspond to the stoichiometric ratioof n(Ir : HSA) = 2:1. Excitation is performed at 338 nm.



Fig.S8. The linear relationship of the phosphorescence intensity with HSA

concentration of 0-280nM (Error bars, SD, n=3), Conditions: in BR buffer (pH=2.0 0.04M), λ_{ex} =338 nm.

The interference study

Fig. S9. Phosphorescence responses of Ir to HSA (blank, 10 μ M) in the presence of (a) different metal ions: Na⁺, K⁺, Zn²⁺, Mg²⁺, Ca²⁺, Ba²⁺, Al³⁺, Cl⁻, NO₃⁻, SO₄²⁻, CO₃²⁻(10mM), Ag⁺, Fe³⁺, Cu²⁺(1mM) and (b) amino acids (10 mM). Amino acids including Ser, Arg, His, Ala, Trp, Pro, Leu, Gln, Ile, Asp,Cys, Tyr, Val, GSH, GSSG. $\lambda ex = 338$ nm (Error bars, SD, n=3).

Table S1 The binding constants for Ir ·HSA through nonlinear finance	tting.

Compound	Stoichiometry	Temp (K)	K ₁ (10 ⁴ M ⁻¹)	K ₂ (10 ⁴ M ⁻¹)
		283	32.5	2.5
Ir ∙HSA	2:1	293	29.9	2.2
		313	20.9	1.1

Table S2. Thermodynamic parameters of Ir ·HSA at 283K, 293K, 313K.

compound	Temp (K)	∆G1(kJ/mol)	ΔH ₁ (kJ/mol)	ΔS ₁ (J/K mol)	∆G₂(kJ/mol)	ΔH ₂ (kJ/mol)	ΔS ₂ (J/K mol)
	283	-24.3	-16.5	27.6	-23.8	-10.5	47.0
Ir ∙HSA	293	-25.3	-15.1	34.8	-24.4	-10.5	47.4
	313	-28.0	-10.2	56.9	-24.2	-12.6	37.1

^aIn K =-ΔH/RT + constant; Δ G=-RT InK; Δ G= Δ H-T Δ S.

Real sample analysis

Real Sample	This method		The clinical data	
neuroumpie	Found (mg ml ⁻¹)	RSD (%) ^b	Found(mg ml ⁻¹) ^c	RSD(%) ^b
Serum 1	23.7	2.3	23.5	2.3
Serum 1	40.3	2.0	40.1	2.0
Serum 1	50.9	1.8	51.3	1.8
Urine 1	10.5ª	1.6	10.0ª	1.7
Urine 2	53.0ª	2.4	52.5ª	2.0
Urine 3	218.1ª	2.8	216 .6ª	2.3

Table S3. Determination of HSA in real blood serum samples and urine samples.

a: μ g ml⁻¹; b: relative standard deviation (n=5); c: The data got from hospital.

methods	Linear range	Detection limit	Reference
Albumin-probe and host molecule	10-30 μM	0.5 μΜ	4
CulnZnS quantum dots- Co ²⁺	0.075 -100μM	45nM	5
Zero Current Potentiometry	30-363 nM	30nM	6
Dye Nanoparticles Fluorescence Sensor	3 -3000 nM	3 nM	7
Organic fluorescent probe DH1	0-3μΜ	83nM	8
Organic fluorescent probe AL-1	0-1 μM	6 nM	9
lridium complex Phosphorescent probe	1-280nM	0.8nM	Present work

Table S4.Comparison of different methods for HSA detection.

References:

- 1. X. Zhang, G.-J. Song, X.-J. Cao, J.-T. Liu, M.-Y. Chen, X.-Q. Cao and B.-X. Zhao, *Rsc Advances*, 2015, 5, 89827-89832.
- 2. M. Parambath, Q. S. Hanley, F. J. Martin-Martinez, T. Giesa, M. J. Buehler and C. C. Perry, *Physical Chemistry Chemical Physics*, 2016, 18, 5938-5948.
- 3. A. Lobnik, I. Oehme, I. Murkovic and O. S. Wolfbeis, *Analytica Chimica Acta*, 1998, 367, 159-165.
- 4. D. Patra, *Biosensors and Bioelectronics*, 2010, 25, 1149-1154.
- 5. W. Y. Gui, X. Q. Chen and Q. Ma, *Analytical and Bioanalytical Chemistry*, 2017, 409, 3871-3876.
- 6. H. Wang, Y. Wu and J.-F. Song, *Biosensors and Bioelectronics*, 2015, 72, 225-229.
- 7. P. Anees, S. Sreejith and A. Ajayaghosh, *Journal of the American Chemical Society*, 2014, 136, 13233-13239.
- 8. J. Fan, W. Sun, Z. Wang, X. Peng, Y. Li and J. Cao, *Chemical Communications*, 2014, 50, 9573-9576.
- 9. Y.-Y. Wu, W.-T. Yu, T.-C. Hou, T.-K. Liu, C.-L. Huang, I. C. Chen and K.-T. Tan, *Chemical Communications*, 2014, 50, 11507-11510.