# Effects of titanium oxide nanoparticles on tetracycline accumulation and toxicity in *Oryza* sativa (L.)

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## S1. Adsorption isothermal experiment

One and a half mg of TiO<sub>2</sub> NPs were weighed into 8 mL glass tubes containing 8 mL 1/2x Hoagland's solution (pH 5.7) amended with different concentrations of TC (0, 0.5, 1.0, 2.5, 5, 7.5, 10.0, 12.5, 15, 20 mg/L). The glass tubes were covered with aluminum-foil lined caps and shaken end-to-end in a rotator at 25 °C. The suspension was sampled at day 3 (as determined by the kinetic experiment) and filtered with 0.22  $\mu$ m filter. The TC concentrations were determined using HPLC (Waters 1525) equipped with a UV-detector (Waters 2487) at 355 nm with a C18 column. The mobile phase was a mixture of acetonitrile and oxalic acid (15/8, *v/v*). The mobile phase was adjusted to pH of 2 and flow rate was 1 mL/min. The triplicate was set in the experiment. The adsorption amount was calculated as the followed equation.

$$q = \frac{\left[(C_0 - C_t) \times V\right] - q_c}{W}$$

where q (mg/g) is the amount of TC adsorbed onto the NPs;  $C_0$  and  $C_t$  is the initial and that at time t (mg/L) concentrations of TC, respectively (calculated based on the standard curve of TC);  $q_c$  (mg/g) is the amount of TC in control without sorbents (TiO<sub>2</sub> NPs); V (L) is the initial volume of the solution; W (g) is the weight of the NPs.

# S2. Analysis for protein content and antioxidant enzyme activities

For protein content determination, plant fine powder was vigorously mixed with 10 mM Tris-HCl (pH 7.2) at a ratio of 1:10 (w/v). The mixture was centrifuged at 4000 rpm, 4 °C for 20 min. One hundred  $\mu$ L of supernatant and 1900  $\mu$ L of Bradford reagent were reacted in a 2 mL centrifuge tube for 15 min at ambient temperature. The absorbance of each sample was measured at 595 nm. Detailed information for antioxidant enzyme extraction buffer, reaction buffer, reaction time, as well as wavelength, are shown in **Table S3**.

### S4. TiO<sub>2</sub> NPs characterization as affected by TC

The hydrodynamic diameter and zeta potential of TiO<sub>2</sub> NPs in single analyte and co-exposure treatments were determined in 1/2x Hoagland's solution (**Figure S5** and **Table S2**). In TiO<sub>2</sub> NPs alone treatments, the hydrodynamic diameters decreased with increasing concentration of NPs in 1/2x Hoagland's solution. One of the possible explanations was that large aggregates might settle down before measurements. The surfaces of TiO<sub>2</sub> NPs were all negatively charged. Interestingly, the addition of different concentrations of TC notably decreased the hydrodynamic diameter of TiO<sub>2</sub> NPs regardless of the doses of TiO<sub>2</sub> NPs. The values of zeta potential also suggested that the presence of TC caused positive charges on the surface of TiO<sub>2</sub> NPs, which could be ascribed to that *pKa* value of TC is positive (3.3 - 9.7) at 25 °C. However, such alteration was only evident in 1000 and 2000 mg/L TiO<sub>2</sub> NPs amended 1/2x Hoagland's solution.

#### S5. The total protein contents in rice seedlings treated with $TiO_2 NPs \times TC$

As shown in **Figure S11**, the presence of TiO<sub>2</sub> NPs and TC significantly altered the total protein contents in rice shoots and roots. For rice shoots, 2000 mg/L TiO<sub>2</sub> NPs alone and 20 mg/L TC alone significantly increased the total protein content by approximately 30% relative to the control. In the co-exposure scenarios, elevation of the total protein content was evident as compared to 5 mg/L and 10 mg/L TC alone treatment. However, in the 20 mg/L TC treatment, the presence of TiO<sub>2</sub> NPs did not further increase the protein content, which were all decreased by 18.1% relative to its TC alone treatment; these values were still significantly higher than the control. Similar to the shoots, the protein contents in rice roots were significantly increased upon exposure to TiO<sub>2</sub> NPs alone or TC alone, regardless of the dose. It is worth mentioning that coexposure of  $TiO_2$  NPs and TC resulted in decreases of the root proteins as compared to the control or the single contaminant treatment. For example, in the 10 and 20 mg/L TC treatments with 2000 mg/L TiO<sub>2</sub> NPs, the total protein levels were only 0.5-fold of the control, or 0.2- and 0.12-fold of the respective TC alone treatment.



**Figure S1**. Phenotypic images of rice seedlings treated with different concentrations of TC in 1/2x Hoagland's solution. (A) and (B) represent images of whole seedlings and rice roots, respectively, in response to different concentrations of TC.



**Figure S2**. Seedling length and fresh biomass of different concentrations of TC treated rice. (**A**) shows root length and shoot height; (**B**) represents fresh biomass of rice shoots and roots.



**Figure S3**. Phenotypic images of rice seedlings treated with different concentrations of  $TiO_2$  NPs in 1/2x Hoagland's solution. (A) and (B) represent images of whole seedlings and rice roots, respectively, in response to different concentrations of  $TiO_2$  NPs.



**Figure S4**. Seedling length and fresh biomass of different concentrations of  $TiO_2$  NPs treated rice. (A) shows root length and shoot height; (B) represents fresh biomass of rice shoots and roots in response to  $TiO_2$  NPs exposure.

TiO <sub>2</sub> NPs TC (mg/L)	A1 (500)	A2 (1000)	A3 (2000)
<b>B1 (5)</b>	A1B1	A2B1	A3B1
<b>B2</b> (10)	A1B2	A2B2	A3B2
<b>B3</b> (20)	A1B3	A2B3	A3B3

**Table S1.** Interactions between  $TiO_2$  NPs and TC in the hydroponic system

TC (mg/L)	TiO <sub>2</sub> NPs (mg/L)	Hydrodynamic diameter (nm)	Zeta potential (mV)
	500	$521.4\pm22.1$	$-15.10\pm0.97$
0	1000	$268.0 \pm 10.9$	$-2.75 \pm 9.23$
	2000	$64.5 \pm 12.7$	$-16.08 \pm 5.92$
	500	$235.0\pm17.7$	-11.41 ±2.07
5	1000	$144.6 \pm 11.3$	$1.87 \pm 11.70$
	2000	$186.5\pm21.4$	$-1.28\pm4.59$
	500	$264.5\pm13.7$	$-10.49 \pm 3.65$
10	1000	$168.2\pm25.8$	$8.73 \pm 6.83$
	2000	$65.6\pm7.2$	$19.45\pm6.64$
	500	$325.3 \pm 23.5$	$-12.86 \pm 1.35$
20	1000	$174.1 \pm 11.8$	$1.68\pm7.67$
	2000	$99.3 \pm 16.8$	$5.71\pm7.85$

Table S2. Hydrodynamic diameter and zeta potential of  $TiO_2$  NPs as affected by TC



**Figure S5**. Particle distribution of  $TiO_2$  NPs in different concentrations of  $TiO_2$  NPs amended 1/2x Hoagland's solution as affected by TC.

Antioxidant enzyme	Extraction buffer	<b>Reaction buffer</b>	Reaction time (min)	Wavelength (nm)
SOD	50 mM phosphate (pH 7.8) containing 0.1% ( $w/v$ ) ascorbate, 0.1 % ( $w/v$ ) bovine serum albumin (BSA), and 0.05% ( $w/v$ ) $\beta$ - mercaptoethanol	100 $\mu$ L of enzyme extract + 1900 $\mu$ L of 50 mM phosphate buffer (pH 7.8) containing 9.9 mM L-methionine, 57 $\mu$ M NBT, 0.0044% ( <i>w</i> / <i>v</i> ) riboflavin and 0.025% ( <i>w</i> / <i>v</i> ) Triton X-100	20	560
POD	50 mM phosphate (pH 7.0) containing 1% ( <i>w/v</i> ) polyvinylpyrrolidone	50 µL of enzyme extract + 1.75 mL of 50 mM sodium phosphate buffer (pH 7.0) +0.1 mL of 4% guaiacol in cuvette + 0.1 mL of 1% ( $v/v$ ) H <sub>2</sub> O <sub>2</sub> to initiate the reaction	2	470
CAT	25 mM KH <sub>2</sub> PO <sub>4</sub> (pH 7.4)	100 $\mu$ L of enzyme extract + 1900 $\mu$ L of 10 mM H <sub>2</sub> O <sub>2</sub>	3	240

 Table S3. Extraction and reaction buffer of antioxidant enzymes

Method	Equation	Description	Assessment
Toxicity Unit (TU)	$TUi = Ci/IC_{50}i$ $TU = \Sigma TUi$ $TU_0 = TU/max(TUi)$	Where <i>TUi</i> is the toxicity unit of the contaminant <i>i</i> ; <i>Ci</i> represents the	TU = 1, additive; $TU > TU_0$ , antagonistic; TU < 1, synergistic; $TU = TU_0$ , independent; $TU_0 > TU > 1$ , partially additive
Additional Index (AI)	When $TU \le 1$ , AI = (1/TU)-1; When $TU \ge 1$ , AI = TU(-1)+1	contaminant <i>i</i> caused 50% reduction of fresh biomass in the scenario of single exposure; $IC_{50i}$ represents the concentration of	AI > 0, synergistic; AI = 0, additive; AI < 0, antagonistic
Mixture Toxicity Index (MTI)	$MTI = 1 - \lg TU / \lg TU_0$	contaminant <i>i</i> that caused 50% reduction of fresh biomass in the scenario of co- exposure;	MTI = 0, independent; 0 < MTI <1, partially additive; MTI = 1, additive; MTI > 1, synergistic; MTI < 0, antagonistic

Table S4. Methods for joint toxicity evaluation of TC and TiO<sub>2</sub> NPs to rice seedlings

TiO <sub>2</sub> NPs TC (mg/L)	A1 (500)	A2 (1000)	A3 (2000)
<b>B1</b> (5)	4.013	3.157	1.903
<b>B2</b> (10)	8.859	7.759	5.736
<b>B3</b> (20)	18.775	17.561	15.174

Table S5. Freely dissolved concentration of TC in the co-exposure treatments

**Note**: The freely dissolved concentration of TC was calculated based on the difference between the addition amount of TC and the portion adsorbed onto  $TiO_2$  NPs, which was calculated by the fitting result of Langmuir model.

Treatment	<b>BAF</b> in shoot	BAF in root
5 mg/L TC	9.618	119.318
10 mg/L TC	10.397	214.879
20 mg/L TC	10.860	663.690
5 mg/L TC×500 mg/L TiO <sub>2</sub> NPs	4.030	4.780
5 mg/L TC×1000 mg/L TiO <sub>2</sub> NPs	3.266	2.570
5 mg/L TC×2000 mg/L TiO <sub>2</sub> NPs	3.424	1.770
10 mg/L TC×500 mg/L TiO <sub>2</sub> NPs	2.289	7.709
10 mg/L TC×1000 mg/L TiO <sub>2</sub> NPs	1.485	3.042
10 mg/L TC×2000 mg/L TiO <sub>2</sub> NPs	1.524	1.712
20 mg/L TC×500 mg/L TiO <sub>2</sub> NPs	2.059	47.667
20 mg/L TC×1000 mg/L TiO <sub>2</sub> NPs	2.007	21.302
20 mg/L TC×2000 mg/L TiO <sub>2</sub> NPs	0.915	3.377

Table S6. Bioaccumulation factor of TC in rice shoots and root

**Note:** BAF=  $C_{TC}$  in plant tissues/ $C_{TC}$  in Hoagland's solution



**Figure S6**. Interactions between TC and TiO<sub>2</sub> NPs in 1/2x Hoagland's solution in the absence of rice. (**A**) is TC reduction rate at ambient temperature over 5 day; (**B**) is TC adsorption amount on TiO<sub>2</sub> NPs.



**Individual treatments** 

**Co-exposure treatments** 

**Figure S7**. Phenotypic images of rice seedlings co-exposed to different concentrations of TC and TiO<sub>2</sub> NPs in 1/2x Hoagland's solution. (A) shows the images of TC alone and NPs alone treatments; (B) shows the images of co-contaminated treatments. In each image, two seedlings on the left-hand side are the control plants grown in 1/2x Hoagland's solution, and the ones on the right-hand side are the treated plants.



**Figure S8**. The contents of Ti in rice roots treated with different concentrations of  $TiO_2$  NPs and TC.



**Figure S9**. Bioaccumulation factor of tetracycline in rice shoots (**A**) and roots (**B**) *versus* freely dissolved concentration of TC in 1/2X Hoagland's solution.



**Figure S10**. The contents of other nutrient elements in rice shoots co-treated with TC and TiO<sub>2</sub> NPs. (A) - (C) represent the contents of Ca, Mg, and Mo, respectively.



**Figure S11**. The total protein contents in rice shoots (**A**) and roots (**B**) upon exposure to TC and TiO<sub>2</sub> NPs.



**Figure S12**. Inhibition rate of rice biomass among all the treatments. (A) and (B) represent biomass inhibition rate in the TC alone and TiO<sub>2</sub> NPs alone treatment, respectively; (C) – (E) represent biomass inhibition rate in the co-treatments when the TC concentration is at 5, 10, and 20 mg/L, respectively.