

Supplementary information

Applications of Surface-Enhanced Raman Spectroscopy on Nanoparticle Analysis in the Environment

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Table S1. Current analytical techniques for NP detection and their pros/cons.

Techniques		Information provided	Pros	Cons	Ref
Light scattering techniques	Dynamic light Scattering	Hydrodynamic diameter	Quick, no sample consumption	Matrix interference, disability to measure smaller particles in polydispersed samples	1,2
	Multi-angle light-scattering	Particle shape	Shape measurement, no sample consumption	Matrix interference	3,4
	Nanoparticle tracking analysis	Hydrodynamic diameter, particle number concentration	Quick, sensitive, suitable to measure polydispersed samples, minimal sample disturbance	Matrix interference, reduced sensitivity for small particles and/or materials of low refractive index.	5
Electron microscopy	Transmission electron microscopy (TEM)	Size, shape, elemental composition (+EDS ^a), crystal structure (+SAED ^b)	Accurate size and shape measurement	Complex sample preparation, matrix interference, highly localized sampling area	1,6
	Scanning electron microscopy	Size, shape, elemental composition	Accurate size and shape measurement	Complex sample preparation, matrix	1,6

	(SEM)	(+EDS)		interference, highly localized sampling area	
Atomic spectrometry	Inductively coupled plasma mass spectrometry (ICP-MS)	Elemental concentration	High sensitivity (ng/L)	Laborious sample digestion and dilution, speciation disability	^{4,6}
	Single particle ICP- MS	Size and concentration (number and mass) of NPs	High sensitivity (ng/L or $10^6/L$), size discrimination	Matrix interference, high size detection limit (>40 nm) for Ti, Zn, Cu and their oxide NPs, inaccurate measurement upon incomplete vaporization	⁷⁻⁹
	Field flow fractionation ICP-MS (FFF-ICP- MS)	Size and concentration (number and mass) of NPs	Separation and detection of a wide size range of NPs, protection of the native states of NPs	Interference of the NPs elution by biomolecules, low recovery ($<80\%$), complicated optimization of working conditions	¹⁰⁻¹²
	Graphite furnace atomic absorption spectrometry (GFAA)	Elemental concentration	High sensitivity, efficient, no laborious digestion, low maintenance	Narrow linear range, one element each time, Complex sample preparation,	¹³

			and cost	speciation disability	
	X-ray absorption spectroscopy	Speciation and element distribution, geometry and oxidation states	Minimal sample preparation, little alteration of the physical and chemical original states of NPs	Low sensitivity, limited access and complicated data analysis	¹⁴
Electroanalytical techniques	Voltammetry of immobilized particles (VIP)	Chemical composition, oxidation state, size, mass concentration	Efficient, cost-effective	Unpractical for detecting trace levels of NPs in the environment due to the low sensitivity	^{15,16}
	Particle collision coulometry (PCC)	Size, number concentration, mass concentration	High sensitivity, portability	Problematic to detect NPs in mixtures with their ionic species	¹³

^aEDS: energy dispersive X-ray spectroscopy. ^bSAED: selected-area electron diffraction.

Table S2. Summary of potential nanomaterial analytes by SERS.

Category	Nanomaterials	Potential indicators	Enhancement mechanisms	Ref
Metal	Nickel nanowires	Crystal violet	Electromagnetic enhancement	¹⁷
	Copper nanoparticles	1,10-phenanthroline (phen), 4,4'-bipyridine (bipy)	ND ^a	¹⁸
	Copper nanoparticles	4-mercaptopypyridine (4-Mpy)	ND	¹⁹
	Palladium	4-Mpy	Electromagnetic	²⁰

	nanoplates		enhancement	
Metal oxide	Pt, Ru, Rh, Pd, Fe, Co, Ni, and their nanoalloys	Pyridine	Electromagnetic enhancement and charge transfer	21
	Cu ₂ O nanocrystals	4-Mpy	ND	22
	Cu ₂ O porous nanostructures	4- aminothiophenol (4-ATP) and dimercaptoazobenzene (DMAB)	Charge transfer and electromagnetic enhancement	23
	Cu ₂ O porous nanowires	4-aminobenzenethiol	ND	24
	Cu ₂ O nanospheres	4-mercaptobenzoic acid (4-MBA) and 4-Mpy	Charge transfer and electromagnetic enhancement	25
	One-dimensional ZnO nanostructures, nanowires and nanocones	4-Mpy	Cavity-like structural resonance of the electric field	26
	Porous ZnO nanosheets	4-MBA	Charge transfer	27
	ZnO nanorods	4-Mpy, 4-aminothiophenol (PATP), (Bu ₄ N) ₂ [Ru(dcbpyH) ₂ (NCS) ₂] (N719) and acetaminophen	Charge transfer	28
	ZnO nanorods	Methyl orange (MO)	Charge transfer	29
	ZnO nanorods	4-Aminobenzenethiol	Charge transfer	30
	ZnO nanoparticles	4-Mpy and 4-MBA	Charge transfer	31

ZnO nanocrystals	4-Mpy	Charge transfer	³²
Magnetite (Fe_3O_4) nanoparticles	Oxalate, pyrocatechol, cysteine	Electromagnetic enhancement	³³
$\alpha\text{-Fe}_2\text{O}_3$ colloids	Pyridine, 1,4-dioxane and 1-ethyl-3'-methyl-2-thiacyanine iodide	Electromagnetic enhancement for pyridine and 1-ethyl-3'-methyl-2-thiacyanine iodide; electromagnetic enhancement and surface-induced molecular resonance enhancement for 1,4-dioxane	³⁴
$\alpha\text{-Fe}_2\text{O}_3$ nanospheres	4-Mpy	ND	³⁵
$\alpha\text{-Fe}_2\text{O}_3$ nanoparticles	4-Mpy and 2-mercaptobenzothiazole (2-MBT)	Charge transfer	³⁶
$\alpha\text{-Fe}_2\text{O}_3$ nanocrystals (sphere, spindle, cube)	4-Mpy	Charge transfer	³⁷
TiO_2 nanoparticles	4-MBA	Charge transfer	^{38,39}
Anatase, TiO_2 nanoparticles	Polymethoxyflavones	Charge transfer	⁴⁰
Anatase, rutile and mixed TiO_2 nanoparticles	4-MBA	Charge transfer and the mixed crystal effect	⁴¹
TiO_2 nanoparticles	Nitrothiophenol isomers	Charge transfer	⁴²
Rutile, TiO_2	L-DOPA	Charge transfer	⁴³

	nanoparticles			
<i>Graphene family</i>	TiO ₂ nanoparticles	Enediol	Charge transfer	⁴⁴
	TiO ₂ , Fe ₂ O ₃ , ZrO ₂ , and CeO ₂ nanoparticles	Catechol	Charge transfer	⁴⁵
	TiO ₂ , SnO ₂ and α-Fe ₂ O ₃ nanoparticles	Catechol and dopamine	Charge transfer	⁴⁶
	Graphene	Phthalocyanine (Pc), rhodamine 6G (R6G), protoporphyrin IX (PPP), and crystal violet (CV)	Charge transfer	⁴⁷
	Graphene	Copper phthalocyanine (CuPc)	Charge transfer	^{48–50}
	Graphene	Metal phthalocyanine (M-Pc) molecules (M = Mn, Fe, Co, Ni, Cu, Zn)	Charge transfer	⁵¹
<i>Composite</i>	Graphene oxide nanosheets	Rhodamine B (RhB)	Charge transfer	⁵²
	Graphene	R6G, melamine, and cephalexin	Charge transfer	⁵³
	Graphene, oxidized graphene	R6G	Charge transfer	⁵⁴
	Silver-deposited TiO ₂ (Ag-TiO ₂) nanoparticles	4-MBA	Charge transfer; Ag-induced additional electron injection into the molecules adsorbed on the TiO ₂ surface through the conduction band of TiO ₂ NPs	⁵⁵
	Zn doped TiO ₂	4-MBA	Charge transfer	⁵⁶

	nanoparticles			
<i>Others</i>	Au-ZnO NP hybrids	PATP molecules	Charge transfer	⁵⁷
	InAs/GaAs quantum dots	pyridine	Charge transfer	⁵⁸
<i>Others</i>	ZnS nanocrystals	4-Mpy	ND	⁵⁹
	PbS quantum dots	4-Mpy	Charge transfer	⁶⁰
	CdTe quantum dots	4-Mpy	Charge transfer	⁶¹

^a ND: not determined.

References

- 1 K. Tiede, A. Boxall, S. Tear, J. Lewis, H. David and M. Hasselov, *Food Addit. Contam. Part A*, 2008, **25**, 795–821.
- 2 B. J. Marquis, S. A. Love, K. L. Braun and C. L. Haynes, *Analyst*, 2009, **134**, 425.
- 3 F. V. D. Kammer, M. Baborowski and K. Friese, *Anal. Chim. Acta*, 2005, **552**, 166–174.
- 4 F. Laborda, E. Bolea, G. Cepriá, M. T. Gómez, M. S. Jiménez, J. Pérez-Arantegui and J. R. Castillo, *Anal. Chim. Acta*, 2016, **904**, 10–32.
- 5 J. A. Gallego-Urrea, J. Tuoriniemi and M. Hassellöv, *TrAC - Trends Anal. Chem.*, 2011, **30**, 473–483.
- 6 H. Guo, Z. Zhang, B. Xing, A. Mukherjee, C. Musante, J. C. White and L. He, *Environ. Sci. Technol.*, 2015, **49**, 4317–4324.
- 7 F. Laborda, E. Bolea and J. Jiménez-Lamana, *Anal. Chem.*, 2014, **86**, 2270–2278.
- 8 S. Lee, X. Bi, R. B. Reed, J. F. Ranville, P. Herckes and P. Westerhoff, *Environ. Sci. Technol.*, 2014, **48**, 10291–10300.
- 9 W.-W. Lee and W.-T. Chan, *J. Anal. At. Spectrom.*, 2015, **30**, 1245–1254.
- 10 A. D. Hawkins, A. J. Bednar, J. V. Cizdziel, K. Bu, J. A. Steevens and K. L. Willett, *RSC Adv.*, 2014, **4**, 41277–41280.
- 11 K. Loeschner, J. Navratilova, R. Grombe, T. P. J. Linsinger, C. Købler, K. Mølhav and E. H. Larsen, *Food Chem.*, 2015, **181**, 78–84.
- 12 V. Nischwitz and H. Goenaga-Infante, *J. Anal. At. Spectrom.*, 2012, **27**, 1084.
- 13 S. M. Majedi and H. K. Lee, *TrAC - Trends Anal. Chem.*, 2016, **75**, 183–196.

- 14 M. Gräfe, E. Donner, R. N. Collins and E. Lombi, *Anal. Chim. Acta*, 2014, **822**, 1–22.
- 15 L. K. Mudashiru, a. C. Aplin and B. R. Horrocks, *Anal. Methods*, 2011, **3**, 927.
- 16 G. Cepriá, W. R. Córdova, J. Jiménez-Lamana, F. Laborda and J. R. Castillo, *Anal. Methods*, 2014, **6**, 3072.
- 17 K. R. Krishnadas, P. R. Sajanlal and T. Pradeep, *J. Phys. Chem. C*, 2011, **115**, 4483–4490.
- 18 M. Muniz-Miranda, C. Gellini and E. Giorgetti, *J. Phys. Chem. C*, 2011, **115**, 5021–5027.
- 19 Y. Wang and T. Asefa, *Langmuir*, 2010, **26**, 7469–7474.
- 20 Y. Xiong, J. M. McLellan, J. Chen, Y. Yin, Z. Y. Li and Y. Xia, *J. Am. Chem. Soc.*, 2005, **127**, 17118–17127.
- 21 Z. Tian, B. Ren and D. Wu, *J. Phys. Chemsitry*, 2002, **106**, 9463–9483.
- 22 Y. Wang, H. Hu, S. Jing, Y. Wang, Z. Sun, B. Zhao, C. Zhao and J. R. Lombardi, *Anal. Sci.*, 2007, **23**, 787–791.
- 23 C. Qiu, L. Zhang, H. Wang and C. Jiang, *J. Phys. Chem. Lett.*, 2012, **3**, 651–657.
- 24 R. C. Wang and H. Y. Lin, *Mater. Chem. Phys.*, 2012, **136**, 661–665.
- 25 L. Jiang, T. T. You, P. G. Yin, Y. Shang, D. F. Zhang, L. Guo and S. H. Yang, *Nanoscale*, 2013, **5**, 2784–2789.
- 26 H.-Y. Shin, E.-L. Shim, Y.-J. Choi, J.-H. Park and S. Yoon, *Nanoscale*, 2014, **6**, 14622–14626.
- 27 Q. Liu, L. Jiang and L. Guo, *Small*, 2014, **10**, 48–51.
- 28 X. Wang, G. She, H. Xu, L. Mu and W. Shi, *Sensors Actuators B Chem.*, 2014, **193**, 745–751.
- 29 S. Lee, J. W. Peng and C. S. Liu, *Appl. Surf. Sci.*, 2013, **285**, 748–754.
- 30 K. Kim, K. L. Kim and K. S. Shin, *Phys. Chem. Chem. Phys.*, 2013, **15**, 9288–94.
- 31 Z. Sun, B. Zhao and J. R. Lombardi, *Appl. Phys. Lett.*, 2007, **91**, 89–92.
- 32 Y. Wang, W. Ruan, J. Zhang, B. Yang, W. Xu, B. Zhao and J. R. Lombardi, *J. Raman Spectrosc.*, 2009, **40**, 1072–1077.
- 33 N. Lee, P. J. Schuck, P. S. Nico and B. Gilbert, *J. Phys. Chem. Lett.*, 2015, **6**, 970–974.
- 34 H. . L. Zhang, P.; Wang, Y.; He, T.; Zhang, B.; Wang, X.; Xin and F., *Chem. Phys. Lett.*, 1988, **153**, 215–222.
- 35 X. Fu, F. Bei, X. Wang, X. Yang and L. Lu, *Mater. Lett.*, 2009, **63**, 185–187.
- 36 X. Fu, S. Wang, Q. Zhao, T. Jiang and H. Yin, *Front. Chem. China*, 2011, **6**, 206–212.
- 37 X. Fu, F. Bei, X. Wang, X. Yang and L. Lu, *J. Raman Spectrosc.*, 2009, **40**, 1290–1295.
- 38 X. Xue, W. Ji, Z. Mao, H. Mao, Y. Wang, X. Wang, W. Ruan, B. Zhao and J. R. Lombardi, *J. Phys. Chem. C*, 2012, **116**, 8792–8797.
- 39 L. Yang, X. Jiang, W. Ruan, B. Zhao, W. Xu and J. R. Lombardi, *J. Phys. Chem.*, 2008, **112**, 20095–20098.

- 40 X. Cao, C. Ma, Z. Gao, J. Zheng, L. He, D. J. McClements and H. Xiao, *J. Agric. Food Chem.*, 2016, **64**, 9436–9441.
- 41 L. Yang, M. Gong, X. Jiang, D. Yin, X. Qin, B. Zhao and W. Ruan, *J. Raman Spectrosc.*, 2015, **46**, 287–292.
- 42 J. S. Teguh, F. Liu, B. Xing and E. K. L. Yeow, *Chem. - An Asian J.*, 2012, **7**, 975–981.
- 43 N. Lee, D. R. Hummer, D. A. Sverjensky, T. Rajh, R. M. Hazen, A. Steele and G. D. Cody, *Langmuir*, 2012, **28**, 17322–17330.
- 44 A. Musumeci, D. Gosztola, T. Schiller, N. M. Dimitrijevic, V. Mujica, D. Martin and T. Rajh, *J. Am. Chem. Soc.*, 2009, **131**, 6040–6041.
- 45 S. J. Hurst, H. C. Fry, D. J. Gosztola and T. Rajh, *J. Phys. Chem. C*, 2011, **115**, 620–630.
- 46 P. Tarakeshwar, D. Finkelstein-Shapiro, S. J. Hurst, T. Rajh and V. Mujica, *J. Phys. Chem. C*, 2011, **115**, 8994–9004.
- 47 X. Ling, L. Xie, Y. Fang, H. Xu, H. Zhang, J. Kong, M. S. Dresselhaus, J. Zhang and Z. Liu, *Nano Lett.*, 2010, **10**, 553–561.
- 48 X. Ling, J. Wu, W. Xu and J. Zhang, *Small*, 2012, **8**, 1365–1372.
- 49 X. Ling, L. G. Moura, M. A. Pimenta and J. Zhang, *J. Phys. Chem. C*, 2012, **116**, 25112–25118.
- 50 X. Ling, W. Fang, Y. H. Lee, P. T. Araujo, X. Zhang, J. F. Rodriguez-Nieva, Y. Lin, J. Zhang, J. Kong and M. S. Dresselhaus, *Nano Lett.*, 2014, **14**, 3033–3040.
- 51 H. Xu, L. Xie, H. Zhang and J. Zhang, *ACS Nano*, 2011, **5**, 5338–5344.
- 52 X. Yu, H. Cai, W. Zhang, X. Li, N. Pan, Y. Luo, X. Wang and J. G. Hou, *ACS Nano*, 2011, **5**, 952–958.
- 53 W. Liang, X. Chen, Y. Sa, Y. Feng, Y. Wang and W. Lin, *Appl. Phys. A Mater. Sci. Process.*, 2012, **109**, 81–85.
- 54 L. Kang, J. Chu, H. Zhao, P. Xu and M. Sun, *J. Mater. Chem. C*, 2015, **3**, 9024–9037.
- 55 L. Yang, X. Jiang, W. Ruan, J. Yang, B. Zhao, W. Xu and J. R. Lombardi, *J. Phys. Chem. C*, 2009, **113**, 16226–16231.
- 56 L. Yang, Y. Zhang, W. Ruan, B. Zhao, W. Xu and J. R. Lombardi, *J. Raman Spectrosc.*, 2010, **41**, 721–726.
- 57 H. Wang, Y. Liu, M. Li, H. Huang, H. M. Xu, R. J. Hong and H. Shen, *Optoelectron. Adv. Mater. Rapid Commun.*, 2010, **4**, 1166–1169.
- 58 L. G. Quagliano, *J. Am. Chem. Soc.*, 2004, **126**, 7393–7398.
- 59 W. Kiefer, A. P. Mazzolini and P. R. Stoddart, *J. Raman Spectrosc.*, 2007, **38**, 1538–1553.
- 60 X. Fu, Y. Pan, X. Wang and J. R. Lombardi, *J. Chem. Phys.*, 2011, **134**, 24707.
- 61 Y. Wang, J. Zhang, H. Jia, M. Li, J. Zeng, B. Yang, B. Zhao, W. Xu and J. R. Lombardi, *J. Phys. Chem. C*, 2008, **112**, 996–1000.