

1 Chromatographic and mass spectrometry analysis of wheat flour prolamins, the

2 causative compounds of Celiac Disease

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SUPPLEMENTARY INFORMATION

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This paper aims to contribute for the better knowledge about wheat prolamins, the main responsible in celiac disease (CD) onset. After prolamin extraction from commercial wheat flour, the extracts were characterized by MALDI-TOF in order to analyze the intact proteins profile. After that, and given the high complexity of each extract, the sample was fractionated before analysis. The study was focused on gliadin extract (GLI) given its strong relationship with CD, and it was fractionated by chromatographic methods. Each GLI fraction was further characterized by MALDI-TOF to establish the proteomic profile. The composition of proteins of each fraction was further verified by nano-LC-MS/MS following a shotgun proteomics approach. This supplementary information comprises the results obtained after digestion of the richest GLI fractions, further analysis by nano-LC-MS/MS and MS/MS spectra treatment by using the Proteome Discoverer software as described in the experimental section. Some additional information was included in this document to better understand the results presented herein.

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25 MATERIALS AND METHODS

26 **Wheat Gluten extraction.** 1 g of wheat flour was sequentially extracted with buffered salt
 27 solution, 0.4 M NaCl (Sigma, UK), 50mL 70% (v/v) aqueous ethanol (Sigma, UK) and 50%
 28 (v/v) 1-propanol (Merck, Germany) with 1% (w/v) dithiothreitol (DTT) (Sigma, UK) and Tris–
 29 HCl at pH 7.5, in order to extract the water-soluble albumins (ALB) and salt-soluble globulins
 30 (GLO), ethanol-soluble prolamins (GLI) and ethanol-insoluble prolamins (GLU), respectively.
 31 A 20:1 (w/v) solids-to-liquid ratio was used for the extractions, which were initiated by

32 vortexing for 2 min at room temperature and continued with magnetic stirring during 1h.
33 Ethanol-insoluble prolamins were extracted at 60°C under nitrogen. Extractions for each sample
34 were done in duplicate and the supernatants were collected by centrifugation (10000×g for 15
35 min). In the case of sodium chloride, an additional wash with deionised water was performed in
36 order to remove the residual salt.

37 **Fractionation of gliadins fractions.** GLI fraction was submitted to a dialysis cleanup process
38 before fractionation. Two different cut-off membranes (Spectrum Laboratories, Inc., Canada)
39 were assayed, 3.5kDa and 12-14kDa. The dialyzed GLI at 12-14kDa were further separated into
40 a preparative chromatographic system (Krauer K-1001 pump coupled to an Elite Lachrom L-
41 2420 UV vis detector, Agilent Technologies). Sample was pumped at 4mL/min into a Purospher
42 C18 (250x25mm, 5µm). The mobile phases used were (A) water (0.1%TFA) and (B)
43 acetonitrile (0.1%TFA) from 24%B to 56%B in 140min. The chromatograms were obtained at
44 214nm through the EZChrom Elite software (Agilent Technologies). The protein elution was
45 monitorized and collected by measuring absorbance at 214nm. After isolated proteins from GLI
46 (12-14kDa) were collected, these fractions were lyophilized and further characterized by mass
47 spectrometry.

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49 RESULTS

50 **Identification of gliadin sub-fractions by nano-LC-MS/MS.** Protein fractions resulting
51 from GLI fractionation were submitted to enzymatic digestion. The hydrolysates were
52 further analyzed by tandem mass spectrometry. Each fingerprint peptide was, therefore,
53 used to sequence the peptides and further identify their precursor protein. The results are
54 described in the following table.

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56 **Table S1.** Identification of the richest gliadin sub-fractions from FIII by nano-LC-
57 MS/MS and further data treatment in Proteome Discoverer

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9	A4URY8	70,0	59.4	1.0	R4JB56	36,4	82.8	100	J7I026	33,9	93.2	67	Q9FS73	32,	98.7	43	D2KFH2	11,3	76.5	14						
	A0A059U	38	0	5		58	9			58	3		Q41547	576	6			25	5							
10	HD1	89,8	58.8	1.0	R4JAP5	39,9	93.9	45						36,	93.2	42										
		35	2	2		50	0			50				759	4											
11					Q9XGF0	42,2	97.3	102																		
12	A4URY8	70,0	69.1	1.1	R4JAP5	39,9	98.2	34	A5JSAB	33,2	98.0	75	IOIT5	33,3	100.	56	Q9S8M8	2,2	95.6	56	D2KFH2	11,3	67.8	97		
		38	1	6		50	0			13	0		7	92	00			25	9							
13	Q03872	89,8	58.1	1.3	R4JDM1	36,3	100.	116	J7I026	33,9	90.3	15					B8XU40	34,	99.1	96	G9IOS9	32,3	56.4	54		
		19	9	2		56	00			58	0						900	0				52	3			
14	Q1KL95	84,6	58.0	1.1	Q9XGF0	42,2	92.4	89	F6M8E7	17,1	93.6	91														
		77	1	3		42	7			51	1															
15					A0A068F6	33,3	82.8	115	K7XE68	33,5	80.2	92														
						W7	02	9		52	3															
16					V9P737	34,2	93.9	100	K7X1L1	32,1	100.	71														
						16	0			13	00															
17									Q3YFI1	32,8	57.1	84														
									Q9M4L8	32,3	98.5	61														
18	A4URY8	70,0	56.0	57	Q9XGF0	42,2	92.6	101	F6M8E7	17,1	100.	60	IOIT5	34,1	100.	93	R9XUT4	37,	95.4	88	D2KFH2	11,3	99.7	79		
		38	0			42	1			51	00		9	14	00			225	3							
19	Q03872	89,8	54.6	28	A0A089VM	37,5	95.8	121	X2KWL1	35,7	98.6	82					B6DQB2	31,	73.9	42	G9IOS9	32,3	97.6	45		
		19	7			B3	48	2		09	6						847	8			D2KFG9	22,3	100.	67		
20					D3UAL6	36,2	98.9	144	K7WV42	32,8	100.	19														
						83	5			31	00															
21					B2Y2Q6	42,1	95.5	108	X2KS61	34,5	79.7	18														
						09	9			17	4															
22					Q0GNF9	32,4	91.1	1.3																		
						24	9	9																		
23					Q5MFH4	34,4	98.3	55	K7WV42	32,8	100.	89	M7ZZ	23,5	100.	56	A7XDG3	16,	100.	50	D2KFH2	11,3	94.1	23		
						85	4			31	00		V2	58	00			206	00							
24	Q94IJ6	77,3	51.9	18	R4JAP5	39,9	96.3	45	Q3YFI1	32,8	82.8	11	Q3S4	32,5	98.6	21	B6DQB8	32,	98.7	96	V5M290	32,3	100.	45		
		71	7			50	8			93	9		V7	68	5			449	6				09	00		
25	B5TM09	79,2	56.6	12	Q00M55	40,0	97.3	54	R9XV20	34,4	93.9	75	IOIT5	33,3	75.9	23	USUA50	32,	99.4	83						
		78	0			31	9			04	0		7	92	6			691	3							
26	A0MZ38	88,4	55.3	15	F8SGN3	40,0	93.2	102	R9XW75	34,4	97.3	17	P047	33,6	99.1	65	F2X0K8	33,	100.	31						
		61	9			71	3			18	3		22	61	2			083	00							
27					B2Y2Q7	41,9	100.	118	X2KS61	34,5	98.2	20														
						81	00			17	0															
28					Q9XGF0	42,2	97.8	177																		
29	A5HMG2	79,4	65.7	1 8	R4JAN5	37,7	98.0	104	F6M8E7	17,1	92.4	91	Q3S4	33,4	100.	47	B6DQB4	32,	96.7	97						
		67	9			63	0			51	7		V8	85	00			295	8							
30	G3K725	93,6	78.9	110					K7X1L1	32,1	100.	8 1					B6UKS0	34,	78.9	43						
		48	8							13	00							439	8							
31									K7WV42	32,8	92.6	16					R9XWD0	37,	98.3	68						
										31	1							229	4							
32					I3QPI2	17,3	89.7	142	Q41528	33,1	98.9	32	IOIT5	33,3	91.6	95	P08453	37,	97.3	17	D2KFH2	11,3	65.7	90		
						99	6			93	5		7	92	5			122	9				25	6		
33					B2Y2S2	38,9	97.6	40	Q9M4M3	30,6	100.	76	IOIT5	30,9	97.6	56	L7R4Z9	31,	76.5	89	A0A0B5JD	20	39,2	100.	9	
						77	5			25	00		8	59	5			294	4				42	00	5	
34					Q8W3X0	38,8	99.0	43	X2KWE1	35,5	100.	89					Q6EEX0	31,	69.8	98						
						18	0			67	00							451	7							

