1 Electronic Supplementary Information (ESI)

2

3 Tyrosol and its metabolites as antioxidative and anti-inflammatory molecules

4 in human endothelial cells

- 5
- 6 Francisco J.G. Muriana,*^a Sergio Montserrat-de la Paz,^a Ricardo Lucas.^b Beatriz Bermudez,^c
- 7 Sara Jaramillo,^d Juan C. Morales,^b Rocio Abia^a and Sergio Lopez^{*a}
- 8
- 9 ^aLaboratory of Cellular and Molecular Nutrition, Instituto de la Grasa (CSIC). Seville, Spain.
- 10 bDepartment of Bioorganic Chemistry, Instituto de Investigaciones Quimicas (CSIC-
- 11 Universidad de Sevilla). Seville, Spain. ^cDepartment of Cell Biology, School of Biology
- 12 (University of Seville). 41012 Seville, Spain. ^dPhytochemicals and Food Quality Group,
- 13 Instituto de la Grasa (CSIC). Seville, Spain.

14 ESI Materials and Methods

15 Synthesis of Tyr-GLU metabolite

16 To a solution of Tyr acetate 4 [1] (200 mg, 1.02 mmol) in anhydrous CH_2Cl_2 (6 mL) and 17 trichloroacetimidate acetylate glucuronosyl donor 5 [2] (366 mg, 0.76 mmol) at -10 °C, BF3·OEt₂ (25 μL, 0.19 mmol) was added drop wise. After 2 h, TLC (hexane-EtOAc 2:1) showed 18 19 the formation of a new product and complete consumption of the glycosyl donor. The 20 reaction was neutralized with NEt₃ and concentrated in vacuum. The resulting residue was 21 purified by flash column chromatography (hexane-EtOAc from 3:1 to 1:1) to afford 2-[4'-22 (methyl 2,3,4-tri-O-acetyl- β -D-glucopyranosyluronate)phenyl] EtOAc, a solution of which (60 23 mg, 0.11 mmol) in MetOH (2 mL) was stirred at room temperature with a solution of Na₂CO₃ 24 (22 mg, 0.204 mmol) in H_2O (0.5 mL). After 16 h, water (1 mL) was added, followed by 25 addition of glacial acetic acid to adjust the pH to 6.2. The solvents were then removed and 26 residue was purified by Sephadex G-25 eluting with H₂O-MeOH (9:1). Fractions containing the desired product were freeze-dried affording compound **2**, tyrosol 4'-O- β -D-glucuronide 27 28 (Tyr-GLU) (25 mg, 98%); ¹H NMR (500.13 MHz, CD₃OD) δ : 7.13 (d, J = 8.36 Hz, H-2', H6'), 7.04 (d, J = 8.43 Hz, H-3', H-5'), 4.87 (d, J = 7.10 Hz, H-1"), 3.76 (bs, H-5"), 3.70 (t, J = 7.12 Hz, H-1), 29 3.57-3.45 (m, H-2", H-3", H-4"), 2.76 (t, J = 7.12 Hz, H-2); ¹³C NMR (100.62 MHz, CD₃OD) δ: 30 31 176.6 (C-6"), 157.8 (C-4'), 134.3 (C-1'), 130.9 (C-2', C-6'), 118.0 (C-3', C-5'), 102.7 (C-1"), 77.8 32 (C-3"), 76.7 (C-5"), 74.8 (C-2"), 73.7 (C-4"), 64.4 (C-1), 39.5 (C-2). ESI-HRMS: Calcd for 33 C₁₄H₁₈O₈ (M⁻): 314.29. Found: 312.9.

34

35 Synthesis of Tyr-SUL metabolite

36 Candida antarctica lipase (Novozym 435[®]) (180 mg) was added to a mixture of Tyr (1 equiv)
37 and the acylating agent methyl butyrate (20 equiv) in 45 mL of *t*-butyl methyl ether using a

38 dry round-bottom flask, and the mixture was stirred for 60 min at 40 °C. The enzyme was decanted and separated. The solvent was evaporated, and the product Tyr butyrate 6 was 39 40 purified by flash column chromatography [1, 3]. Tyr butyrate 6 (70 mg, 0.33 mmol) and 41 $SO_3 \cdot NMe_3$ (233 mg, 1.7 mmol) were subjected to sulphation conditions for 20 min. 42 Microwave based sulphation reaction was performed using a microwave synthesizer in 43 sealed reaction vessels [4]. TLC (ethyl acetate:MeOH; 10:1) showed the formation of a major 44 product and complete consumption of the starting material. Solvents were removed and the crude extract was purified by using Sephadex LH-20 in a solvent mixture of CH₂Cl₂:MeOH 45 (1:1) to afford triethylammonium, 4-(2-(butyryloxy)ethyl)phenyl sulphate salt, a solution of 46 47 which (97 mg, 0.32 mmol) and potassium carbonate (K_2CO_3 , 90 mg, 0.66 mmol) were prepared in MeOH (10 mL). The reaction mixture was stirred at room temperature for 24 h, 48 49 neutralized with IR 120 H⁺ resin, and the solvent was then removed in a vacuum. The crude 50 extract was purified by column chromatography with RP-C18 silica gel eluting with 51 H₂O:MeOH (from 100:0 to 70:30). Fractions containing the desired product were concentrated and freeze-dried affording compound 3, tyrosol 4-sulfate (Tyr-SUL) (68 mg, 52 53 94%, white powder). ¹H-NMR (400 MHz, D₂O) δ: 7.18, 6.86 (2d, 4H, J = 8.4 Hz, H_{arom}), 3.78, 2.78 (2t, 4H, J = 6.7 Hz, CH₂OH, CH₂Ar); ¹³C-NMR (75 MHz, D₂O) δ : 157 (Cq), 130.3 (2 × 54 55 CH_{arom}), 129.8 (Cq), 117.1 (2 × CH_{arom}), 63.0 (CH₂OH), 36.9 (CH₂Ar). ESI-HRMS (ES⁻) Calcd for 56 C₈H₉O₅S (M – H) 217.0171, Found: 217.0171.

57

58 SUPPLEMENTARY REFERENCES

S. Grasso, L. Siracusa, C. Spatafora, M. Renis and C. Tringali, Hydroxytyrosol lipophilic
 analogues: enzymatic synthesis, radical scavenging activity and DNA oxidative damage
 protection, *Bioorg Chem*, 2007, **35**, 137-52.

3

B. Fischer, A. Nudelman, M. Ruse, J. Herzig, H.E. Gottlieb and E. Keinan, A novel
method for stereoselective glucuronidation, *J Org Chem*, 1984, **49**, 4988-93.

3. 64 R. Lucas, F. Comelles, D. Alcantara, O. S. Maldonado, M. Curcuroze, J. L. Parra, and J. C. Morales, Surface-active properties of lipophilic antioxidants tyrosol and hydroxytyrosol 65 fatty acid esters: a potential explanation for the nonlinear hypothesis of the antioxidant 66 67 activity in oil-in-water emulsions, J Agric Food Chem, 2010, 58, 8021-6. 68 4. A. Atzeri, R. Lucas, A. Incani, P. Penalver, A. Zafra-Gomez, M. P. Melis, R. Pizzala, J. C. Morales and M.Deiana, Hydroxytyrosol and tyrosol sulfate metabolites protect against the 69 70 oxidized cholesterol pro-oxidant effect in Caco-2 human enterocyte-like cells, Food Funct, 71 2016, 7, 337-46.

ESI Table 1. Sequences of primers for gene expression analysis.

Target	GenBank accession number	Direction	Sequence $(5' \rightarrow 3')$
GPX1	NM_000581	Forward Reverse	AGAATGTGGCGTCCCTCTGA ACCGTTCACCTCGCACTTCT
GCLC	NM_001498	Forward Reverse	TCCAGGTGACATTCCAAGCC GAAATCACTCCCCAGCGACA
HO-1	NM_002133	Forward Reverse	TCTTGGCTGGCTTCCTTACC GGATGTGCTTTTCGTTGGGG
E-selectin	NM_000450	Forward Reverse	AGCCCAGAGCCTTCAGTGTA AACTGGGATTTGCTGTGTCC
ICAM-1	NM_000201	Forward Reverse	CAGTCACCTATGGCAACGAC ATTCAGCGTCACCTTGGCTC
VCAM-1	NM_001078	Forward Reverse	TCCGTCTCATTGACTTGCAG CACCTGCATTCCTTTTTCCA
CCL2	NM_002982	Forward Reverse	CCCCAGTCACCTGCTGTTAT TGGAATCCTGAACCCACTTC
PTGS2	NM_000963	Forward Reverse	TGAGCATCTACGGTTTGCTG TGCTTGTCTGGAACAACTGC
GAPDH	NM_001289746	Forward Reverse	TCGACAATGGCAGCATCTAC ATCCGTCTCCACAGACAAGG
HPRT	NM_000194	Forward Reverse	ACCCCACGAAGTGTTGGATA AAGCAGATGGCCACAGAACT

Concentration (μM)	Tyr	Tyr-GLU	Tyr-SUL
0	100 ± 3.1ª	100 ± 4.4ª	100 ± 10.7ª
1	101 ± 2.7ª	97 ± 5.6ª	97 ± 4ª
5	99 ± 4.8ª	95 ± 6.2ª	96 ± 4.8ª
10	96 ± 4ª	95 ± 3.8ª	95 ± 6.5ª
15	98 ± 2.9ª	96 ± 7.8ª	97 ± 4.7ª
50	97 ± 2.5ª	96 ± 2.4ª	95 ± 2.9ª
100	96 ± 6ª	95 ± 9ª	95 ± 5ª
200	93 ± 4.3ª	71 ± 2.3 ^b	58 ± 11 ^b

ESI Table 2. Effects of Tyr, Tyr-GLU, and Tyr-SUL on hEC viability.

hECs were cultured in the presence of Tyr or Tyr metabolites (0-200 μ M) for 48 h. Values are expressed in % of cells alive and are shown as mean \pm SD of eight samples repeated in three separate experiments. Means within columns sharing the same letter are not significantly different from each other (p < 0.05).

Treatment	sE-selectin	sICAM-1	sVCAM-1
Control	$2.9\pm1.5^{\text{a}}$	$3.0\pm0.7^{\text{a}}$	3.2 ± 1.7ª
TNF-α	$7.9\pm1.0^{\text{b}}$	$7.4\pm1.3^{ ext{b}}$	$19.6\pm3.0^{\text{b}}$
Tyr + TNF- α	$3.4\pm1.8^{\text{a}}$	$5.9\pm1.9^{\text{a}}$	$15.7\pm5^{ m b}$
Tyr-GLU + TNF- α	$2.6\pm2.5^{\text{a}}$	$3.1\pm1.2^{\text{a}}$	$2.6\pm2.5^{\text{a}}$
Tyr-SUL + TNF- α	3.5 ± 2.8^{a}	$2.9\pm0.7^{\circ}$	4.1 ± 2.8^{a}

ESI Table 3. Concentration of soluble forms of E-selectin, ICAM-1, and VCAM-1 in the medium of hECs.

hECs were untreated (control) or exposed to Tyr or its metabolites (100 μ M) for 16 h and then with TNF- α (10 ng/mL) for additional 16 h. Values are expressed in pg/mL and are shown as mean ± SD of three independent experiments. Means within columns sharing the same letter are not significantly different from each other (p < 0.05).