

Supporting information

A Recyclable AgI/OAc⁻ Catalytic System for the Efficient Synthesis of α -Alkylidene Cyclic Carbonates: Carbon Dioxide Conversion at Atmospheric Pressure

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1. General information

All the compounds involved including the substrates, silver salts and ionic liquids were commercially purchased from Sigma-Aldrich, Aladdin, TCI, Alfa, Macklin in China and used without further dryness and purification. The purity of CO₂ used for purging and reacting was 99.999%. NMR spectra were recorded on a Bruker 500 MHz NMR (¹H NMR, 500 MHz; ¹³C NMR, 126 MHz) spectrometer. Their peak frequencies were referenced versus an internal standard (TMS) shifts at 0 ppm for ¹H NMR and against the solvent (CDCl₃, 77.0 ppm; DMSO-*d*₆, 39.9 ppm) for ¹³C NMR, respectively. Multiplicity abbreviations: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet. The coupling constants, *J*, were reported in Hertz (Hz).

2. Experimental section

2.1 Synthesis of α -alkylidene cyclic carbonates by carboxylative cyclization of propargylic alcohols with CO₂ catalyzed by AgI/IL₁ system.

AgI (11.7 mg, 0.05 mmol, 1 mol%), IL₁ (1 mL, 1.102 g, 6.474 mmol), a propargylic alcohol (5 mmol) were added to a Schlenk tube equipped with a magnetic stirring bar. The system was quickly purged 3 times with CO₂. Then the mixture was stirred at 45 °C, 1 bar of CO₂ for the required time. Upon completion, the mixture was extracted with hexane (4 × 15 mL). The upper layer was combined and the solvent was removed to obtain the crude product, which could be further purified by column chromatography on silica gel using petroleum ether/ethyl acetate (100:1-20:1) as an eluent. When the reusability of the catalytic system was investigated, the lower layer (recovered IL₁ + AgI) was dried under vacuum for 10-15 minutes to evaporate the residual hexane. After dryness, the recovered IL₁ and AgI were directly reused as the catalyst for the next round.

2.2 NMR yield calculation for the reaction system of CO₂ with propargylic alcohols.

The ¹H NMR spectra of the crude reaction mixture generated by 2-methylbut-3-yn-2-ol and CO₂ (Figure S1) was used as an example for the quantitative determination of products employing 1,1,2,2-tetrachloroethane as the internal standard. The deuterated solvent is CDCl₃.

Two protons that belongs to the double bonds of α -alkylidene cyclic carbonates (red and green marked), which are far away from the alkyl and aryl protons and show two clear individual peaks, could be treated as characteristic protons for quantitative determination. The sharp peak at $\delta = 5.98$ ppm (blue marked) represents two protons of 1,1,2,2-tetrachloroethane. Therefore, the exact amount and the NMR yield of the product could be calculated by the ratio of double bond proton and internal standard proton based on the exact amounts of 1,1,2,2-tetrachloroethane and the substrate.

Example: is = internal standard, p = product, s = substrate.

$m(\text{is}) = 0.8668 \text{ g}$, $M(\text{is}) = 167.86 \text{ g/mol}$. $\implies n(\text{is}) = 5.164 \text{ mmol}$.

ratio (is/p) = 2.23/2 : 1 = 1.115. $\implies n(\text{p}) = n(\text{is}) / \text{ratio (is/p)} = 4.631 \text{ mmol}$.

$m(\text{s}) = 0.4187 \text{ g}$, $M(\text{s}) = 84.12 \text{ g/mol}$. $\implies n(\text{s}) = 4.977 \text{ mmol}$.

Therefore, NMR yield = $n(\text{p}) / n(\text{s}) = 93\%$.

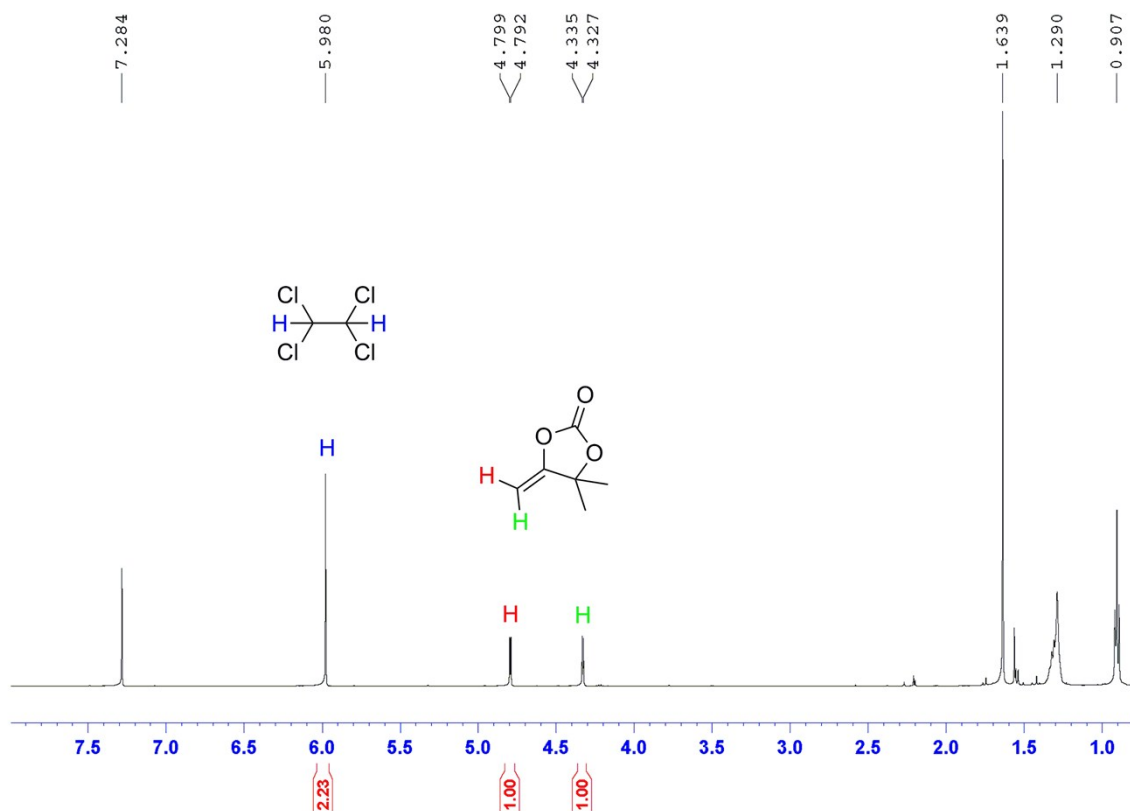
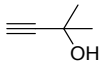
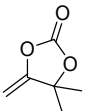
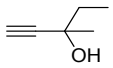
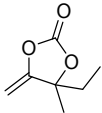
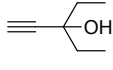
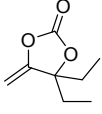
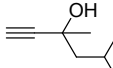
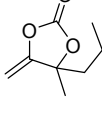
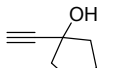
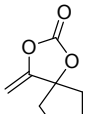
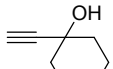
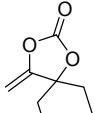
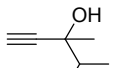
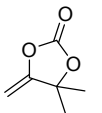
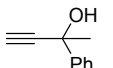
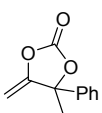


Figure S1. ^1H NMR of crude product generated by 2-methylbut-3-yn-2-ol and CO_2

2.3 The isolated yields of 2a-2h

Table S1. The isolated yields of 2a-2h.^a

Substrate	Product	Time	Yield ^b	Yield ^c
1a 	2a 	3 h	93%	90%

1b		2b		4 h	96%	93%
1c		2c		3 h	91%	87%
1d		2d		6 h	99%	94%
1e		2e		8 h	88%	83%
1f		2f		6 h	99%	95%
1g		2g		6 h	97%	93%
1h		2h		12 h	44%	40%

^a Reaction conditions: AgI (0.05 mmol), IL₁ (1 mL, 1.102 g, 6.474 mmol), substrate (5 mmol), CO₂ (1 bar) at 45°C.

^b NMR Yields were determined by ¹H NMR spectroscopy using 1,1,2,2-tetrachloroethane as the internal standard.

^c Isolated yields

2.4 The mass and the amount of ILs of Table 1.

Table S2. The mass and the amount of ILs of Table 1

	V	m	n
IL₁	1 mL	1.102 g	6.474 mmol
IL₂	1 mL	1.243 g	6.270 mmol
IL₃	1 mL	1.28 g	6.466 mmol
IL₄	1 mL	1.5 g	3.833 mmol
IL₅	/	1.5 g	6.301 mmol

2.5 Comparison of AgI/IL₁ system (this work) with the previously reported reusable silver systems.

Table S3. Comparison of AgI/IL₁ system (this work) with the previously reported reusable silver systems

Ref.	Year	Authors	Catalytic systems	General reaction condition	CO ₂ pressure	Recycle rounds
1	2013	Yuan <i>et al.</i>	Polystyrene-supported -NHC-Ag	2 mol% of Polystyrene - supported-NHC-Ag	5 MPa	15
2	2014	He <i>et al.</i>	Ag ₂ WO ₄ PPh ₃	1 mol% of Ag ₂ WO ₄ 2 mol% of PPh ₃	0.1 MPa	4
3	2014	Liu <i>et al.</i>	Fluorinated-MOP-Ag DBU	10 mol% of Ag 1 equiv of DBU	1 MPa	5
4	2015	Han <i>et al.</i>	Ag nanoparticles- sulfonated macroreticular resin DBU	10 mol% of Ag 1 equiv of DBU	0.1 MPa	5
5	2015	He <i>et al.</i>	[(Ph ₃ P) ₂ Ag] ₂ CO ₃	1 mol% of [(Ph ₃ P) ₂ Ag] ₂ CO ₃	0.1 MPa	>2
6	2016	Wang <i>et al.</i>	AgOAc [P ₆₆₆₁₄][DEIm]	1 mol% of AgOAc 10 mol% of [P ₆₆₆₁₄][DEIm]	0.1 MPa	>5
7	2016	Liu <i>et al.</i>	poly(PPh ₃)-azo-Ag DBU	0.064 mol% of Ag 1 equiv of DBU	1 MPa	5
8	2016	Wang <i>et al.</i>	AgI@C DBU	3 mol% of Ag 20 mol% of DBU	0.1 MPa	10
		This Work	AgI IL ₁	1 mol% of AgI 1.3 equiv of IL ₁	0.1 MPa	20

3. Investigations about the activation mechanism

In this investigation, all the ¹H NMR spectra applied were performed in DMSO-*d*₆.

It has been reported that the activation of the hydroxyl proton of the substrate was the pivotal process to initiate the catalytic cycle, which could be monitored by ¹H NMR.¹

Consequently, in order to recognize the species that activated the hydroxyl proton, substrate **1a** (5 mmol), and the mixture of **1a**/AgI (5 mmol of **1a**, 0.05 mmol of AgI), **1a**/IL₁ (5 mmol of **1a**, 1 mL of IL₁) were respectively added to a Schlenk tube and directly analyzed by ¹H NMR. The obtained spectra were supplied in Figure S2.

In Figure S2(a), a peak at $\delta = 5.29$ ppm was considered as the hydroxyl proton of

the substrate. When AgI was added into **1a**, no significant difference was observed, indicating that AgI individually couldn't activate the hydroxyl proton. Nevertheless, when IL₁ was mixed with **1a**, the peak of hydroxyl proton was extremely extended as a quite broad peak at $\delta = 6.45$ ppm (Figure S3). These results implied that the activation was accomplished by the IL component of the catalytic system.

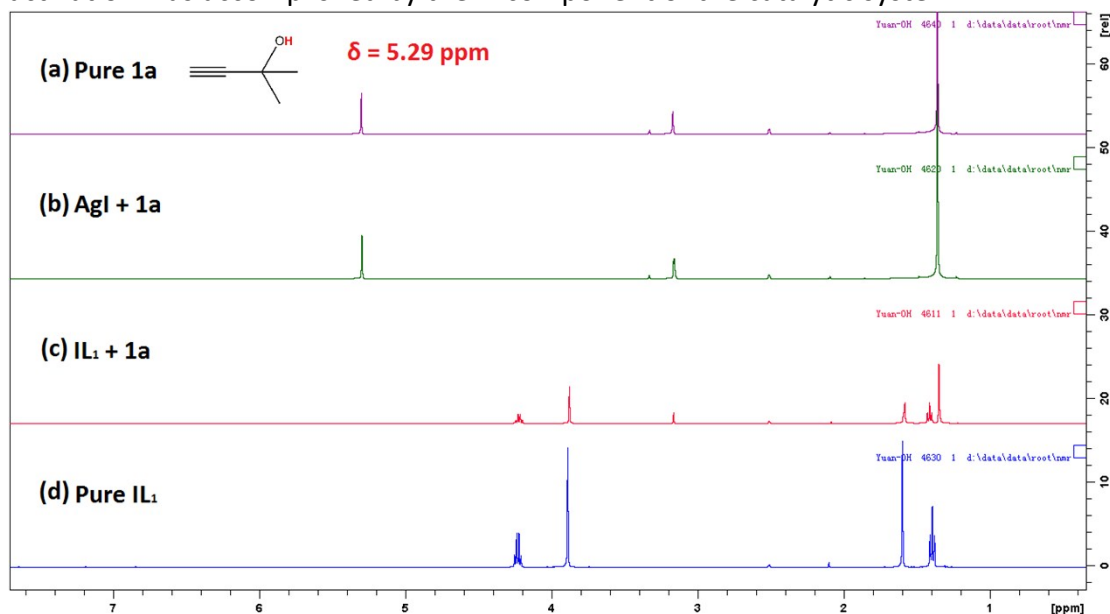


Figure S2. ¹H NMR of (a) pure **1a** in DMSO-*d*₆, (b) AgI+**1a** in DMSO-*d*₆, (c) IL₁+**1a** in DMSO-*d*₆, (d) pure IL₁ in DMSO-*d*₆

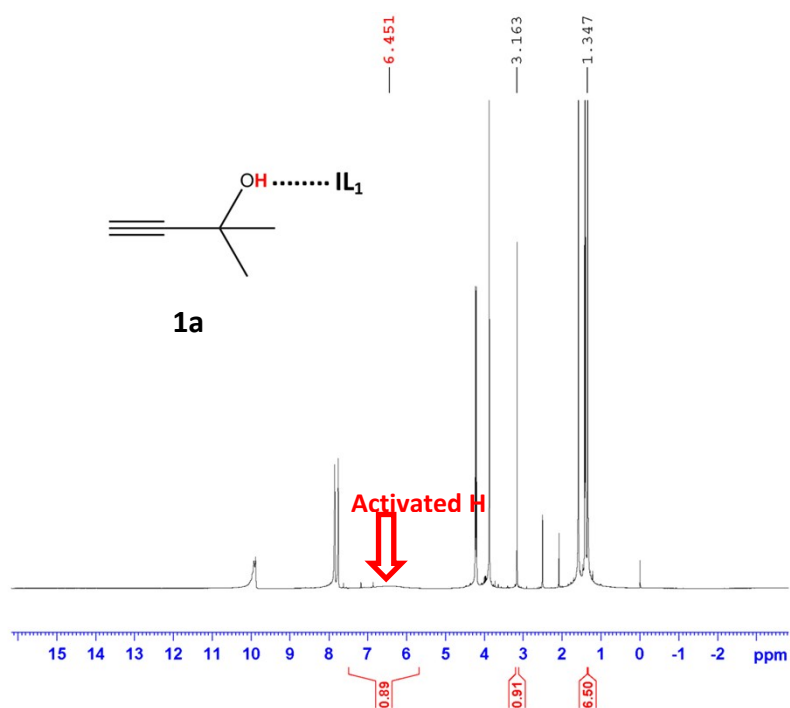
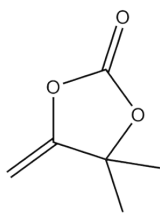


Figure S3. ¹H NMR of IL₁+**1a** in DMSO-*d*₆ (peaks marked with chemical shifts belong to **1a**)

4. Characterization of products



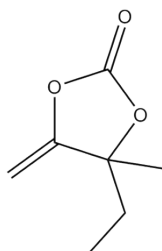
2a

4, 4-Dimethyl-5-methylene-[1,3]dioxolan-2-one (2a)

^1H NMR (500 MHz, CDCl_3) δ = 4.77 (d, J = 4.0 Hz, 1H), 4.33 (d, J = 4.0 Hz, 1H), 1.62 (s, 6H).

^{13}C NMR (126 MHz, CDCl_3) δ = 158.74, 151.27, 85.30, 84.65, 27.56.

The spectroscopic data matched those reported in the literature.⁹



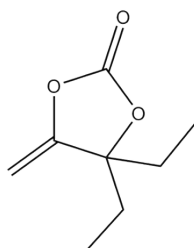
2b

4-Ethyl-4-methyl-5-methylene-[1,3]dioxolan-2-one (2b)

^1H NMR (500 MHz, CDCl_3) δ = 4.83 (d, J = 4.0 Hz, 1H), 4.28 (d, J = 3.9 Hz, 1H), 1.93 (dq, J = 14.6 Hz, 7.4 Hz, 1H), 1.78 (dq, J = 14.7 Hz, 7.4 Hz, 1H), 1.60 (s, 3H), 1.01 (t, J = 7.4 Hz, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ = 157.48, 151.53, 87.56, 85.53, 33.40, 25.95, 7.32.

The spectroscopic data matched those reported in the literature.⁹



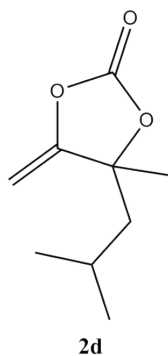
2c

4, 4-Diethyl-5-methylene-[1, 3] dioxolan-2-one (2c)

^1H NMR (500 MHz, CDCl_3) δ = 4.88 (d, J = 3.8 Hz, 1H), 4.24 (d, J = 3.9 Hz, 1H), 1.95 (dq, J = 14.6 Hz, 7.3 Hz, 2H), 1.72 (dt, J = 14.8 Hz, 7.4 Hz, 2H), 0.99 (t, J = 7.4 Hz, 6H).

^{13}C NMR (126 MHz, CDCl_3) δ = 155.83, 151.85, 90.83, 85.78, 31.90, 7.10.

The spectroscopic data matched those reported in the literature.⁹

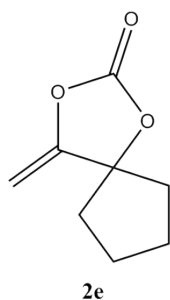


4-Isobutyl-4-methyl-5-methylene-[1,3] dioxolan-2-one (2d)

^1H NMR (500 MHz, CDCl_3) δ = 4.81 (d, J = 3.9 Hz, 1H), 4.29 (d, J = 3.9 Hz, 1H), 1.90 – 1.80 (m, 2H), 1.70 – 1.66 (m, 1H), 1.60 (s, 3H), 1.00 – 0.98 (m, 6H).

^{13}C NMR (126 MHz, CDCl_3) δ = 158.37, 151.45, 87.30, 85.54, 48.57, 27.02, 24.29, 23.98, 23.68.

The spectroscopic data matched those reported in the literature.⁹

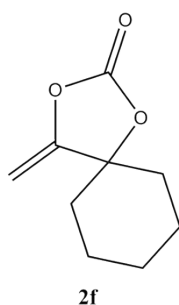


4-Methylene-1,3-dioxaspiro [4.4] nonan-2-one (2e)

^1H NMR (500 MHz, CDCl_3) δ = 4.81 (d, J = 3.9 Hz, 1H), 4.36 (d, J = 3.9 Hz, 1H), 2.28 – 2.23 (m, 2H), 1.98 – 1.83 (m, 6H).

^{13}C NMR (126 MHz, CDCl_3) δ = 157.79, 151.46, 94.21, 85.31, 40.65, 24.25.

The spectroscopic data matched those reported in the literature.⁵

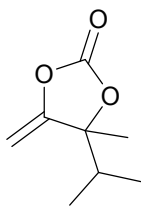


4-Methylene-1,3-dioxaspiro [4.5] decan-2-one (2f)

^1H NMR (500 MHz, CDCl_3) δ = 4.78 (d, J = 3.8 Hz, 1H), 4.30 (d, J = 3.8 Hz, 1H), 2.04 – 2.02 (m, 2H), 1.79 – 1.59 (m, 7H), 1.38 – 1.28 (m, 1H).

^{13}C NMR (126 MHz, CDCl_3) δ = 158.79, 151.48, 86.38, 85.46, 36.53, 24.37, 21.62.

The spectroscopic data matched those reported in the literature.⁹



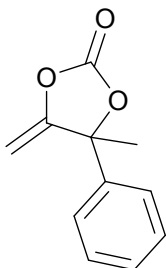
2g

4-Isopropyl-4-methyl-5-methylene-1,3-dioxolan-2-one (2g)

^1H NMR (500 MHz, CDCl_3) δ = 4.84 (d, J = 3.8 Hz, 1H), 4.28 (d, J = 3.8 Hz, 1H), 1.99-1.93 (m, 1H), 1.59 (s, 3H), 1.03 (dd, J = 13.4 Hz, 13.4 Hz, 6H).

^{13}C NMR (126 MHz, CDCl_3) δ = 157.14, 151.71, 89.82, 86.21, 36.99, 24.04, 16.34, 16.04.

The spectroscopic data matched those reported in the literature.⁵



2h

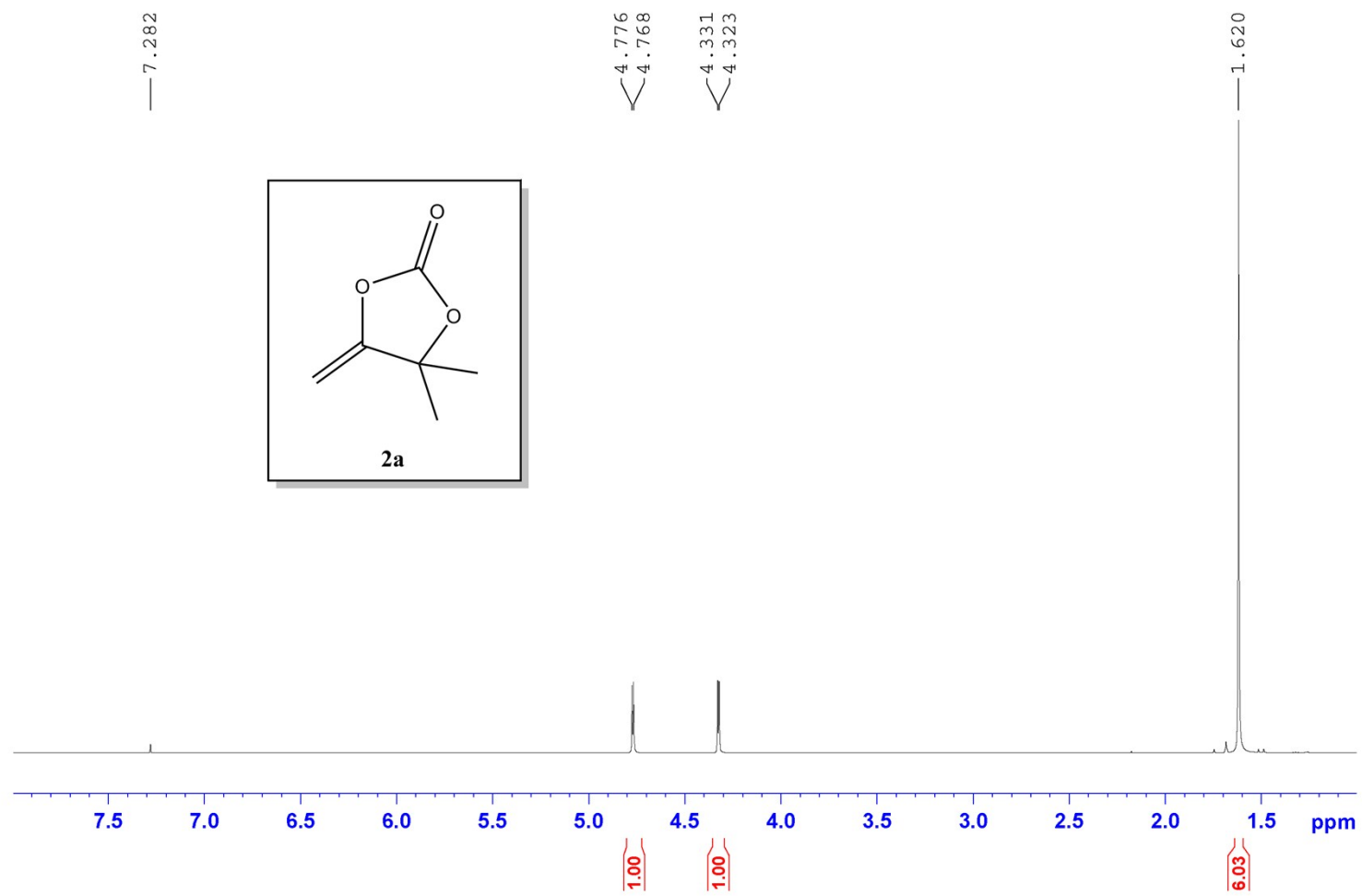
4-Methyl-5-methylene-4-phenyl-1,3-dioxolan-2-one (2h)

^1H NMR (500 MHz, CDCl_3) δ = 7.51 – 7.50 (m, 2H), 7.47 – 7.40 (m, 3H), 4.98 (d, J = 4.0 Hz, 1H), 4.49 (d, J = 4.0 Hz, 1H). 2.00 (s, 3H)

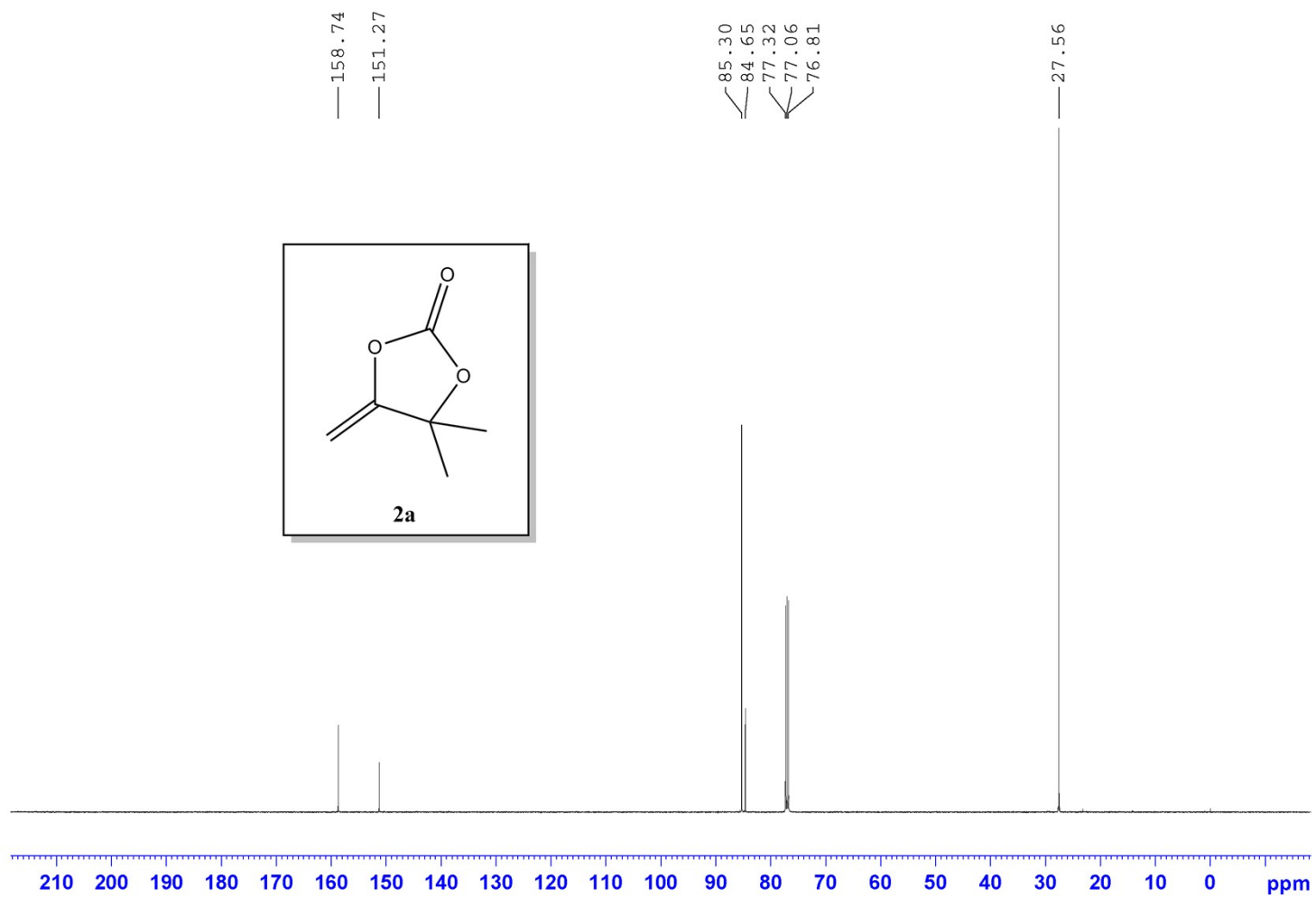
^{13}C NMR (126 MHz, CDCl_3) δ = 157.49, 151.16, 139.30, 129.19, 128.93, 124.70, 88.16, 87.16, 27.47

The spectroscopic data matched those reported in the literature.⁵

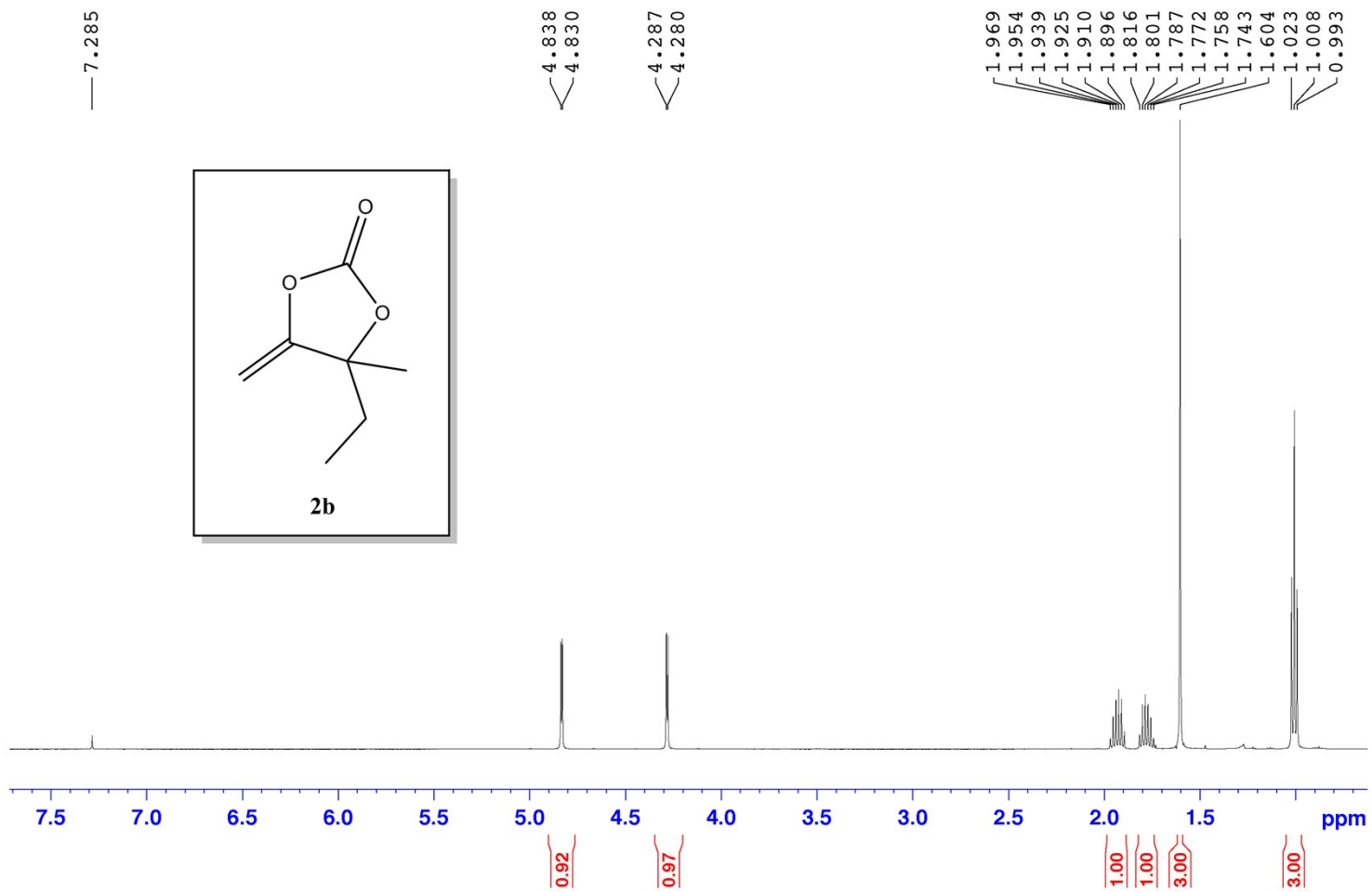
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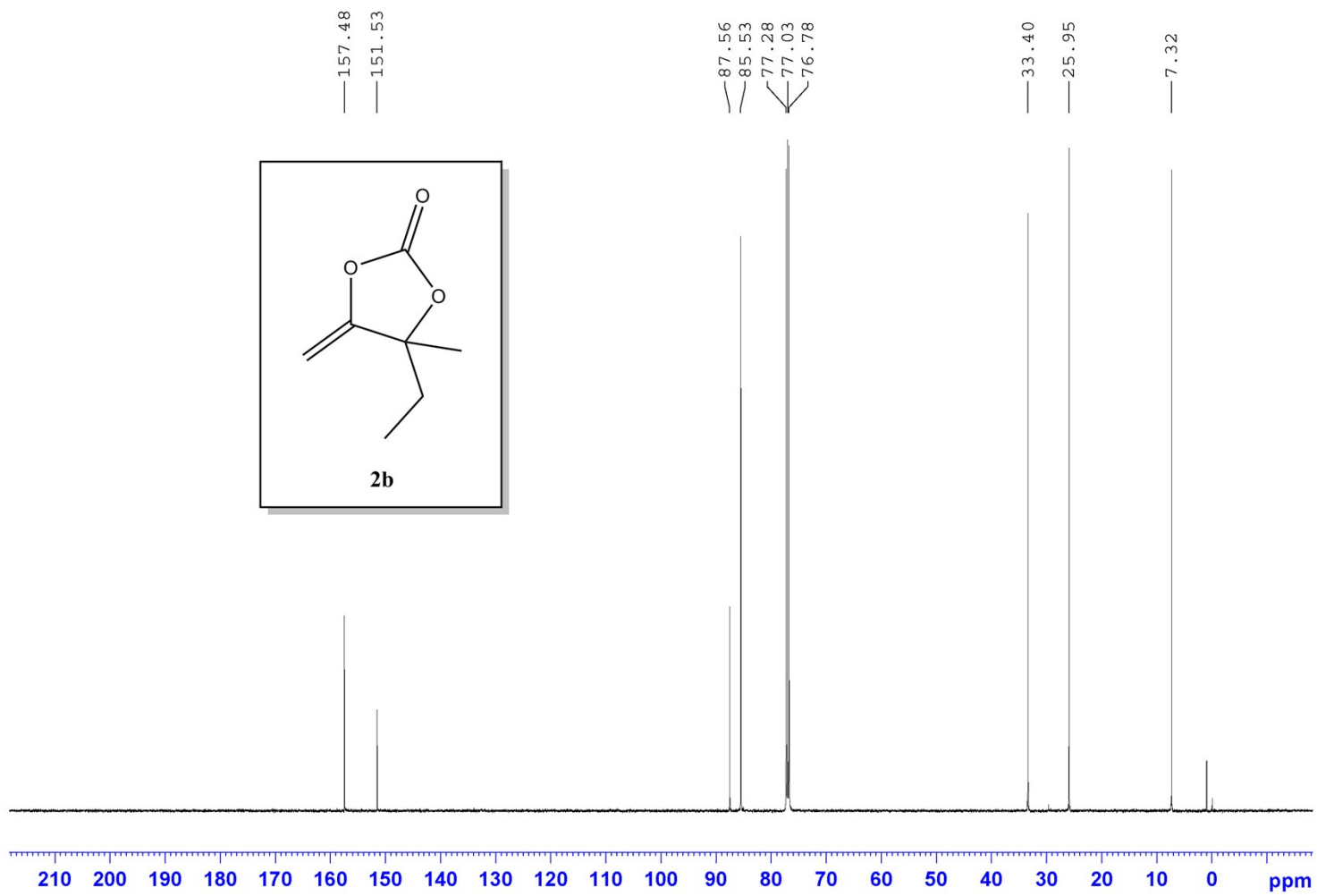
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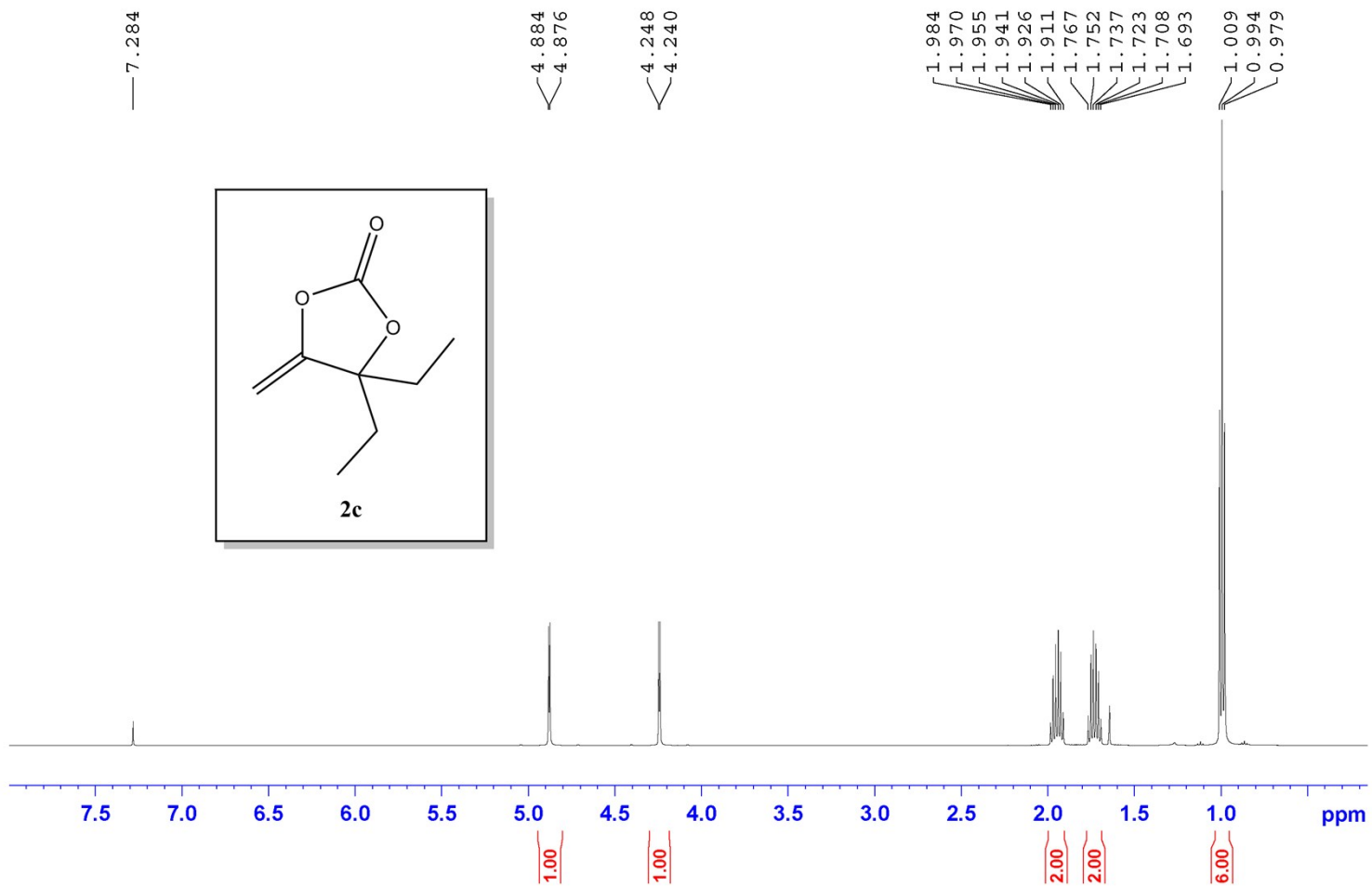
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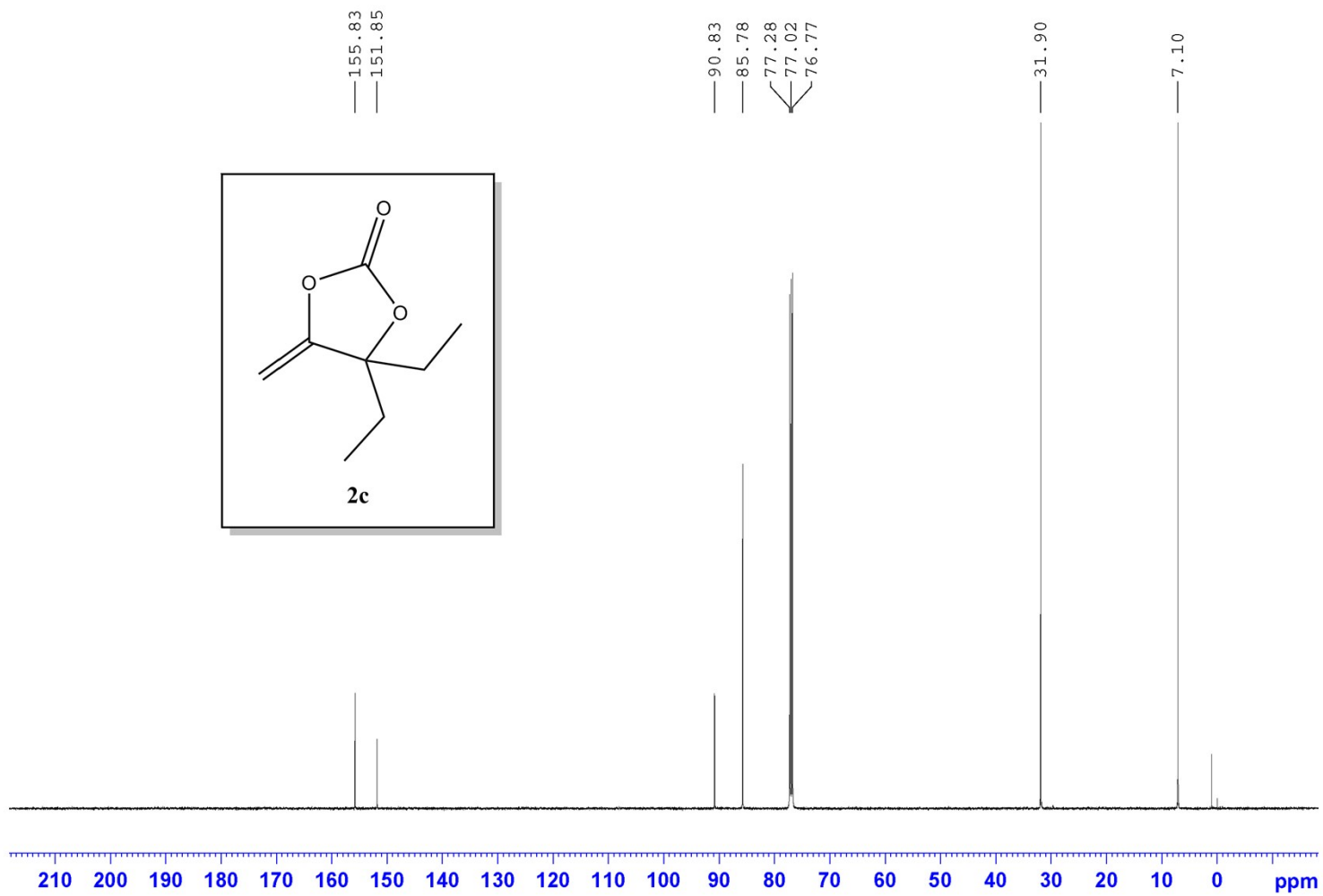
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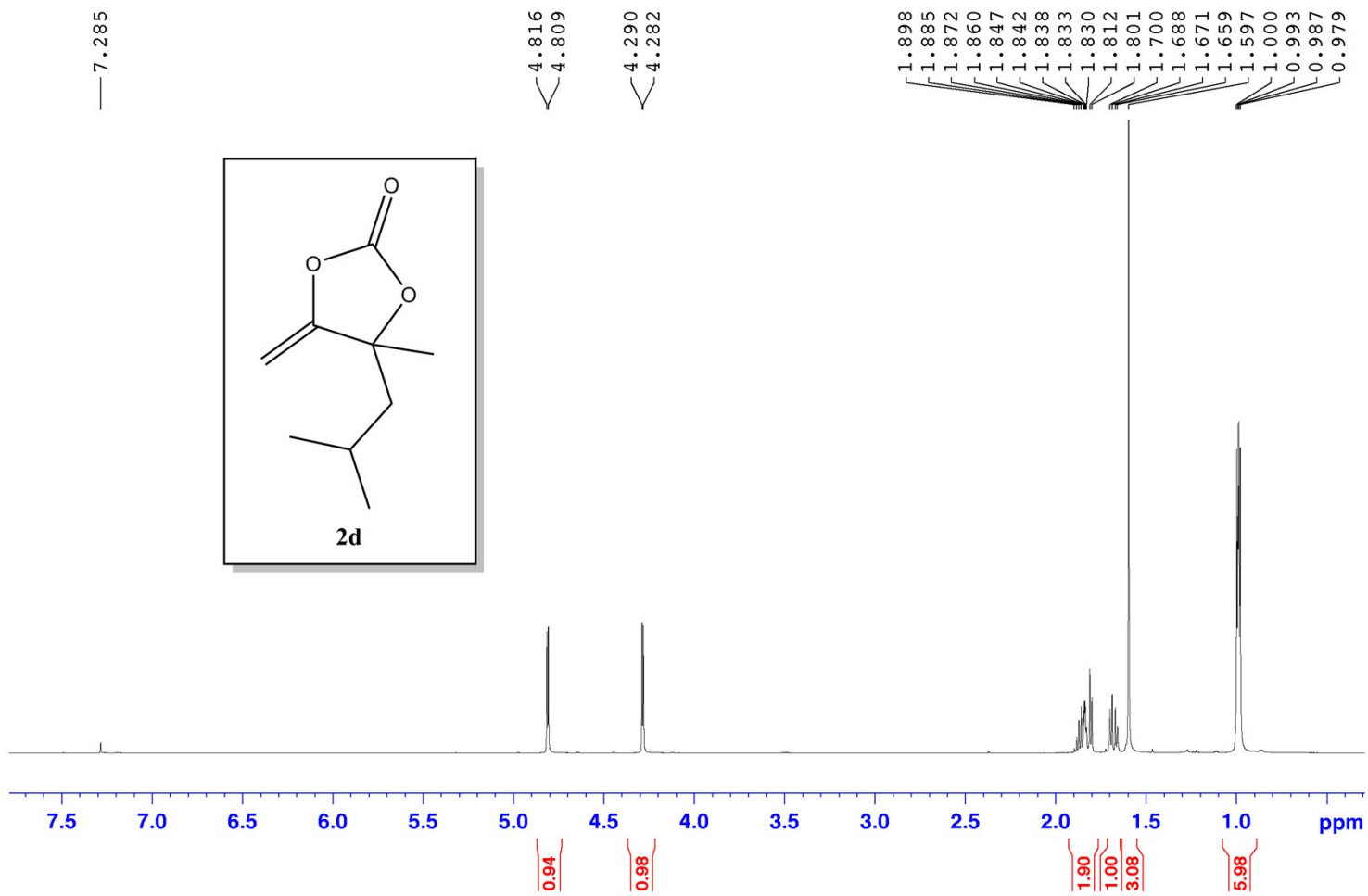
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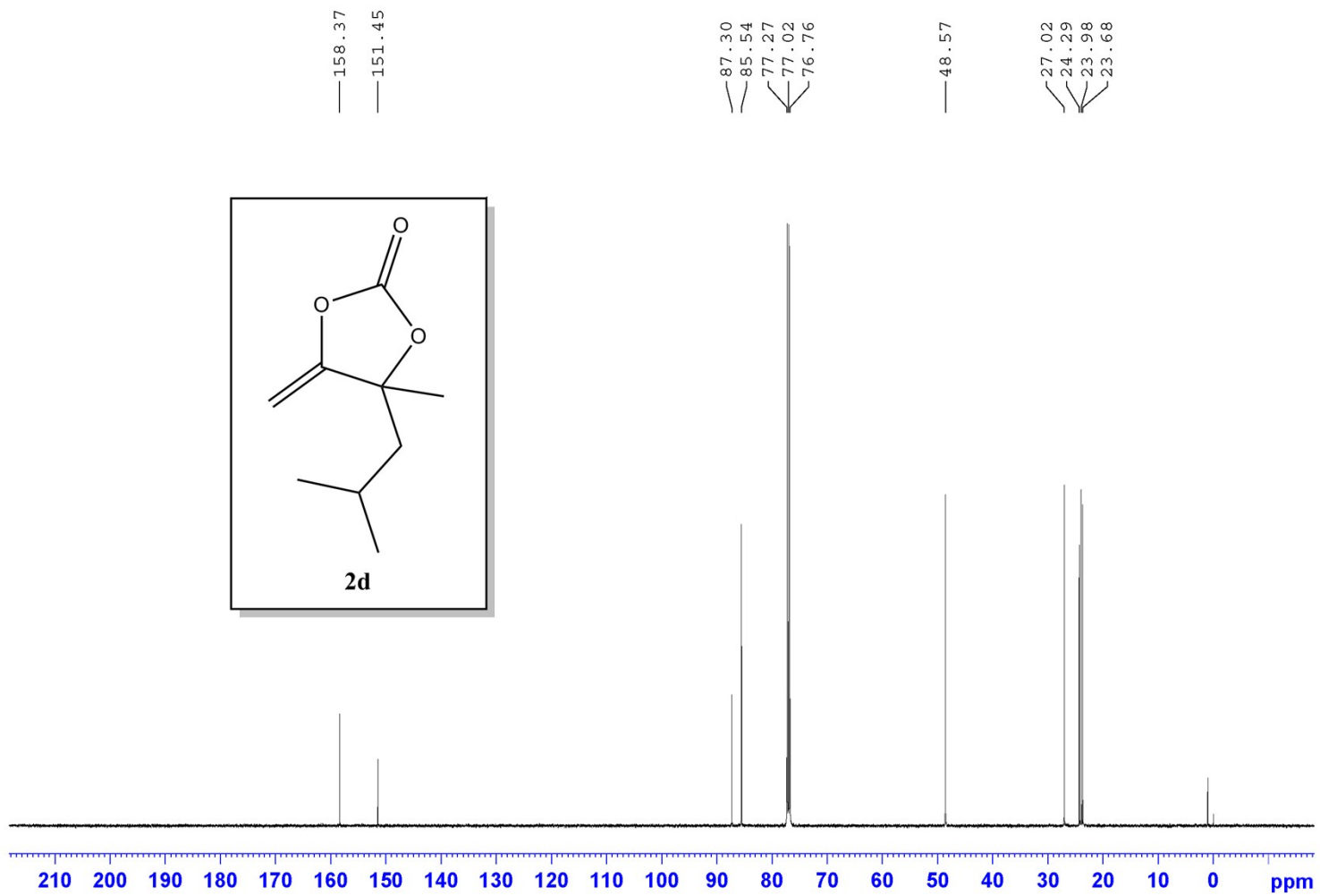
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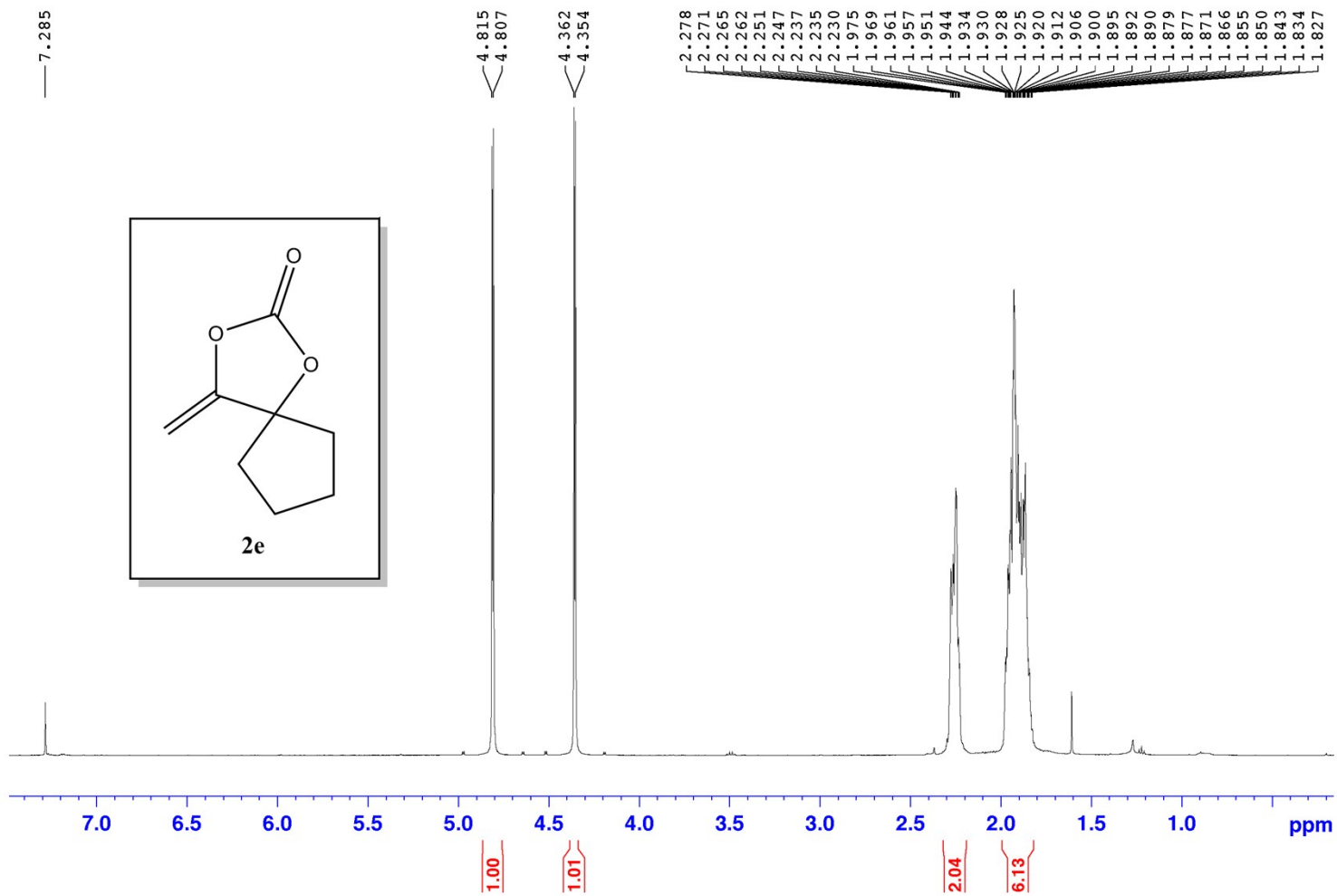
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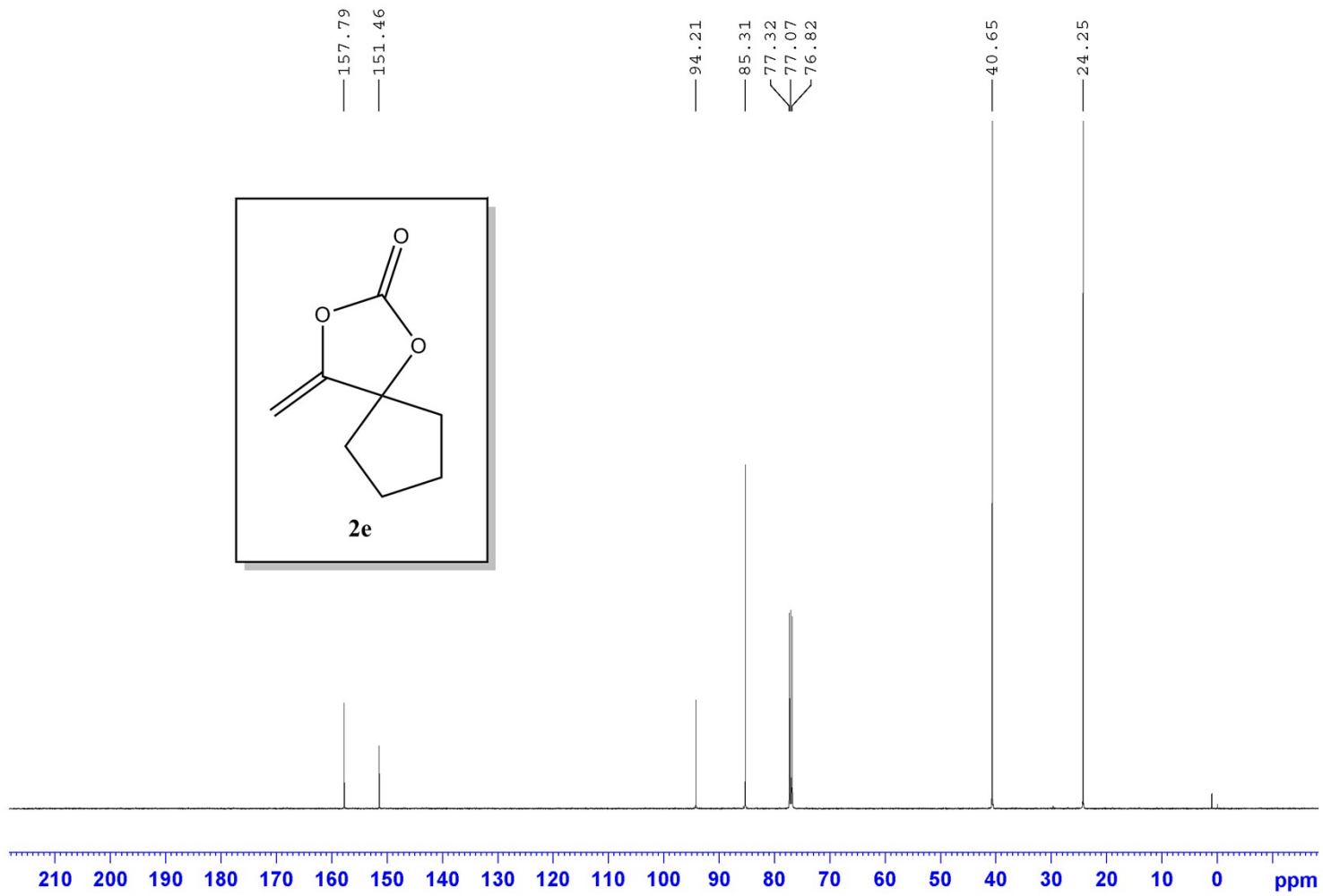
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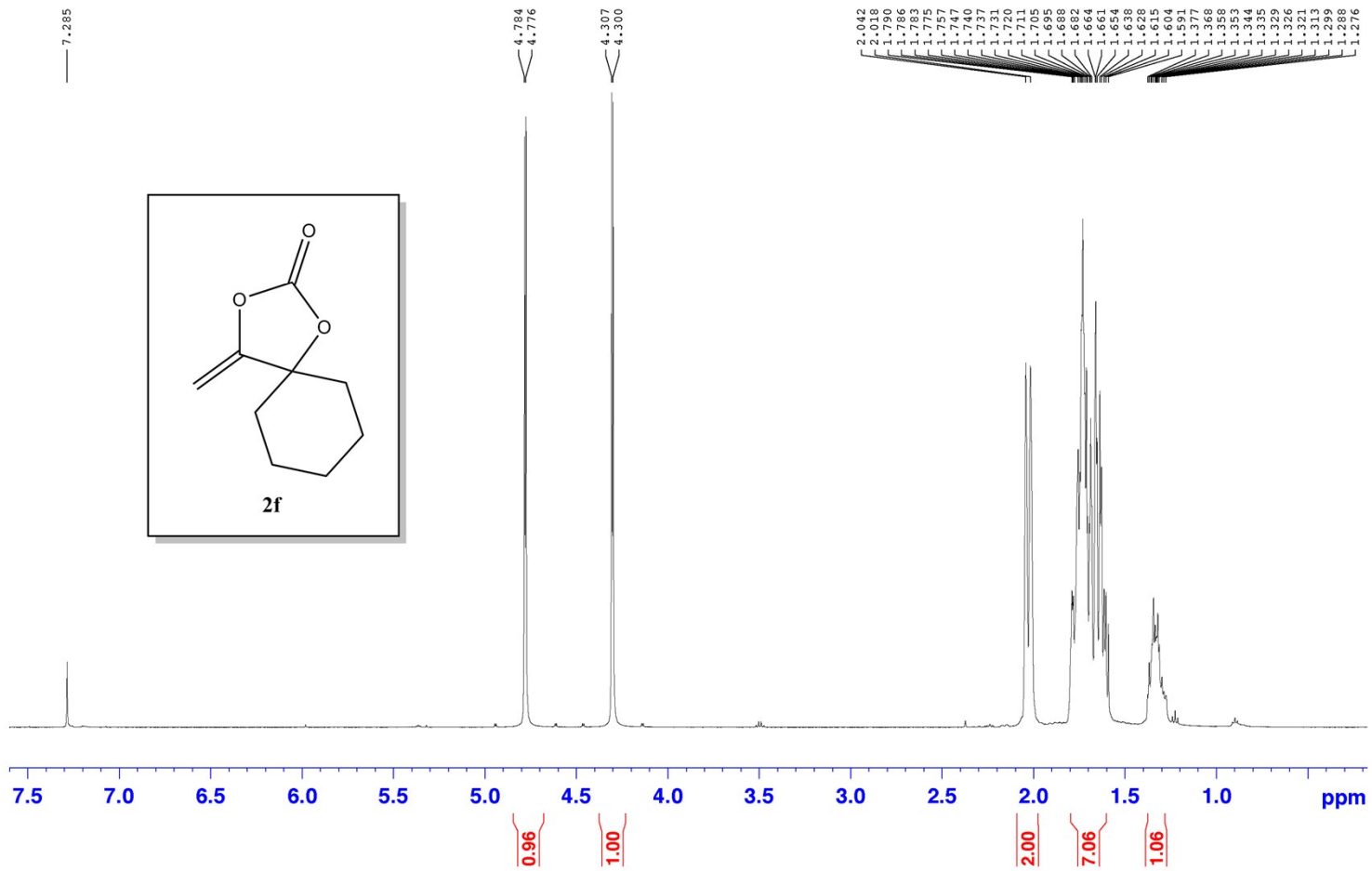
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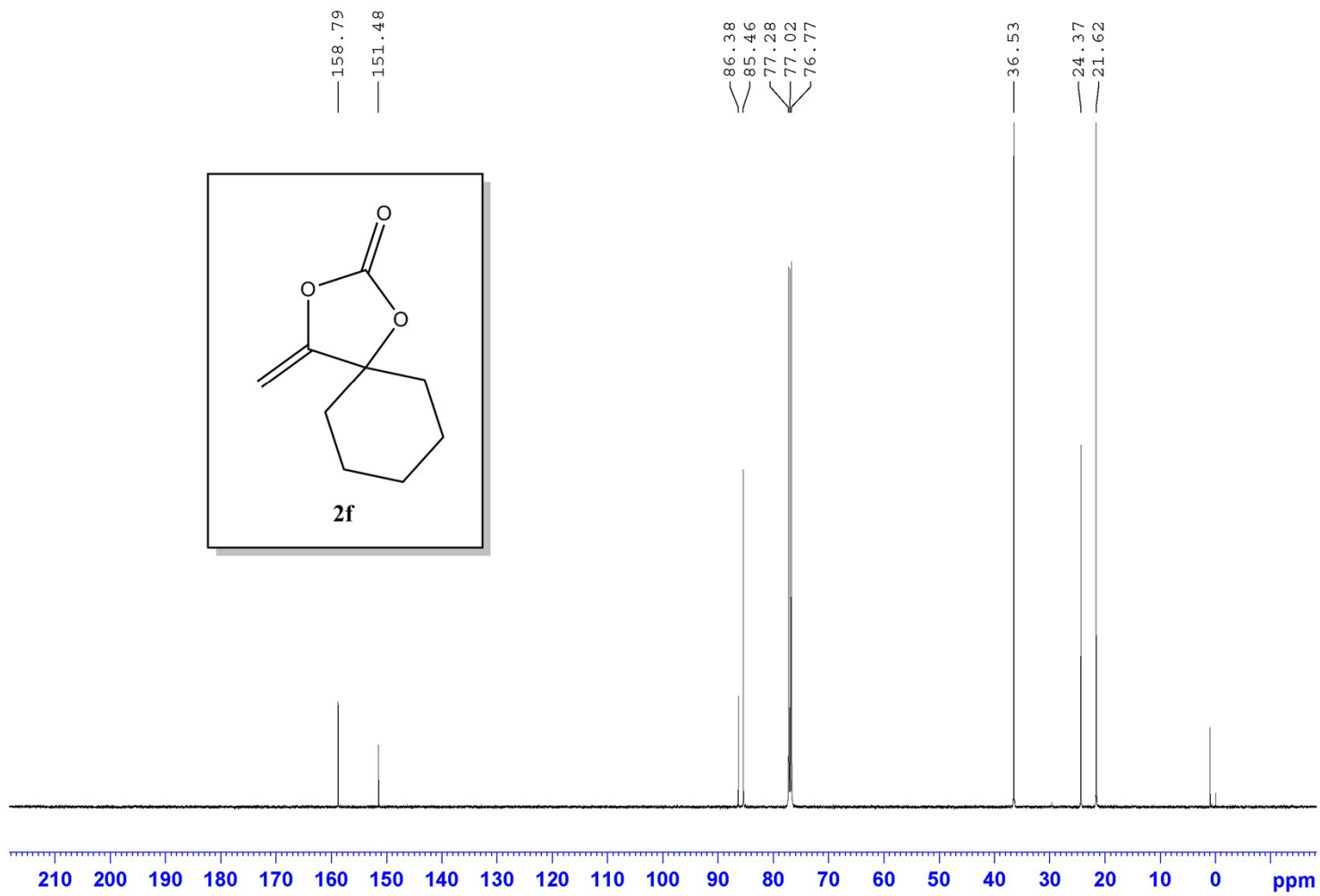
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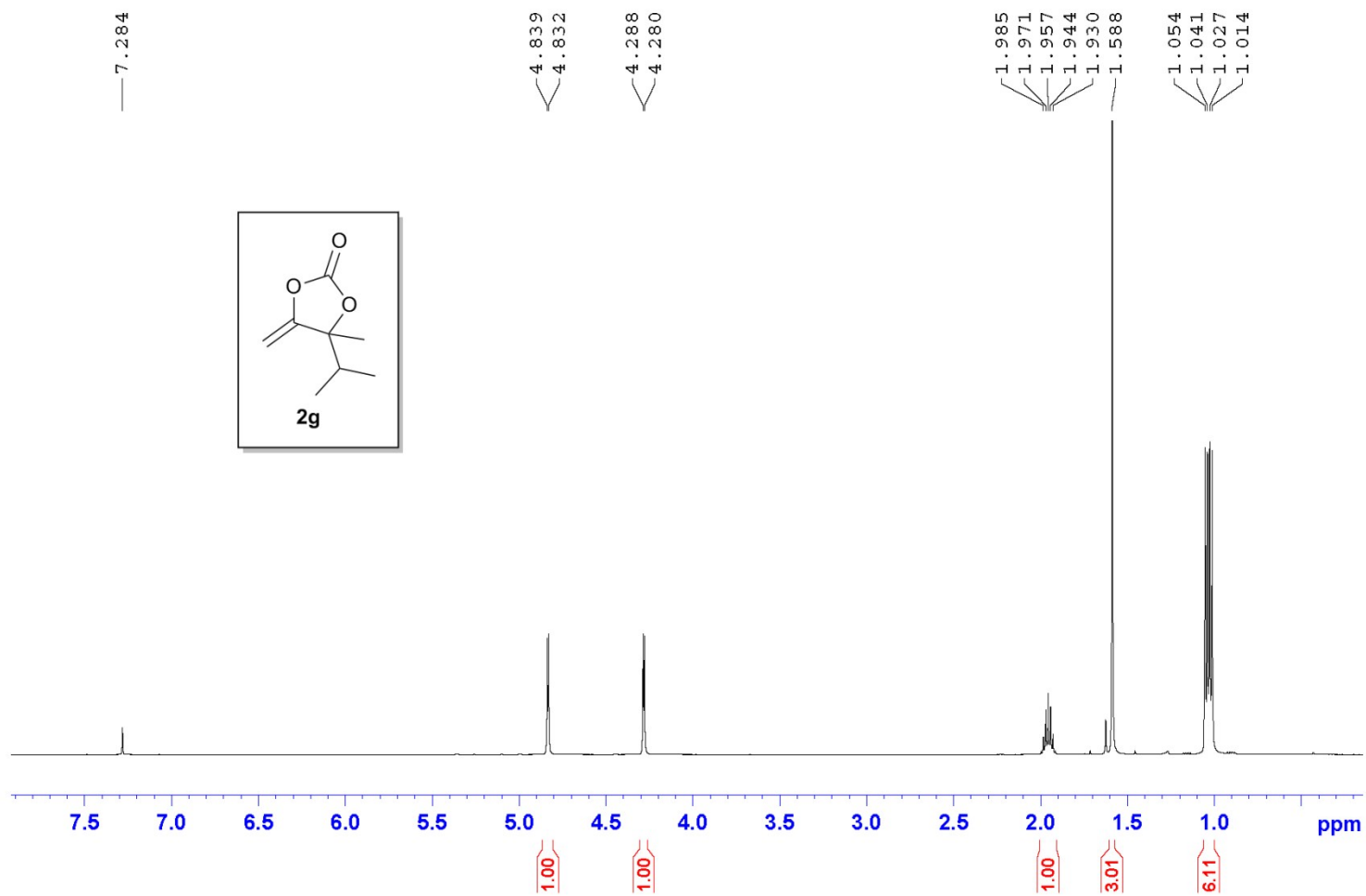
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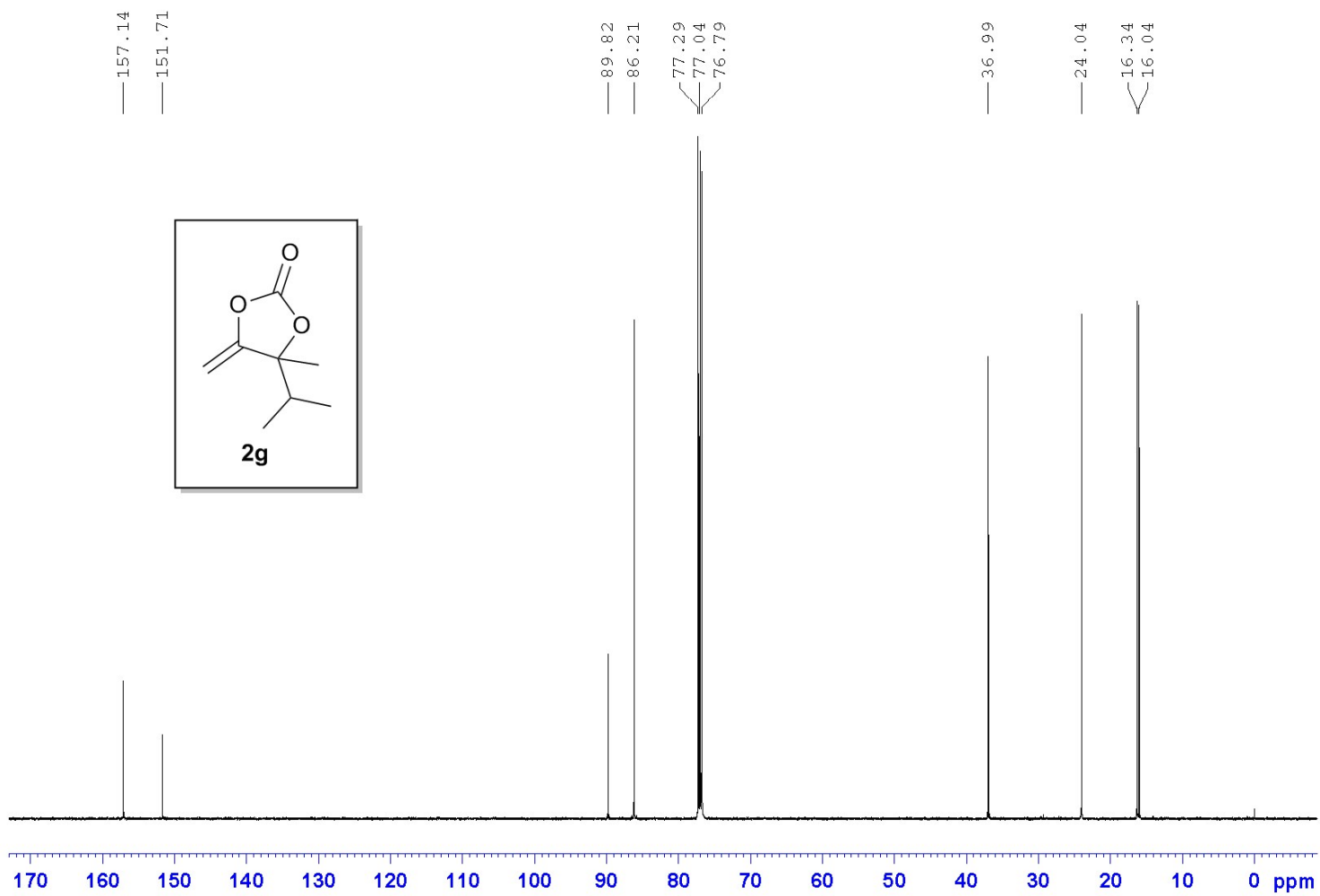
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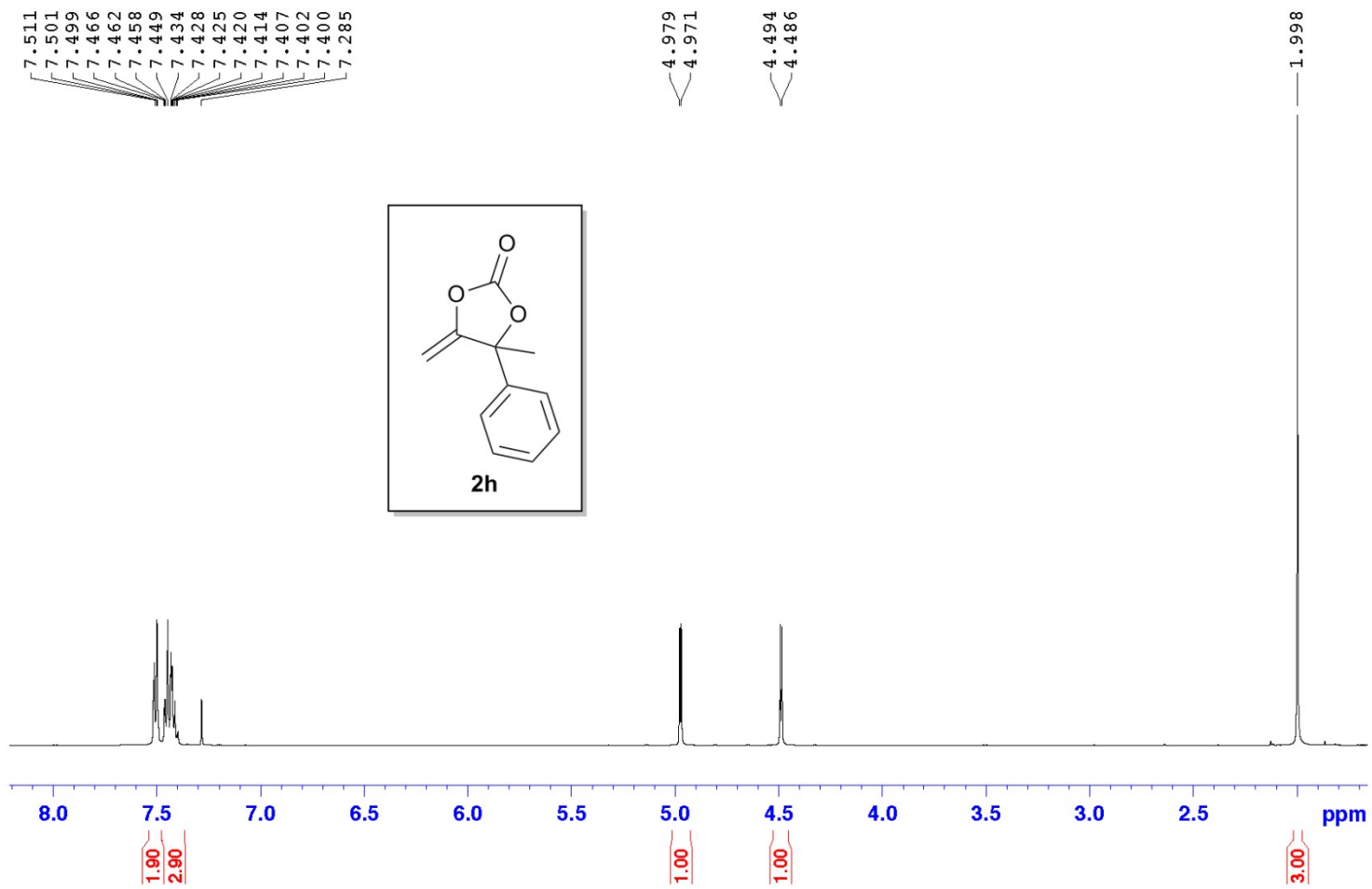
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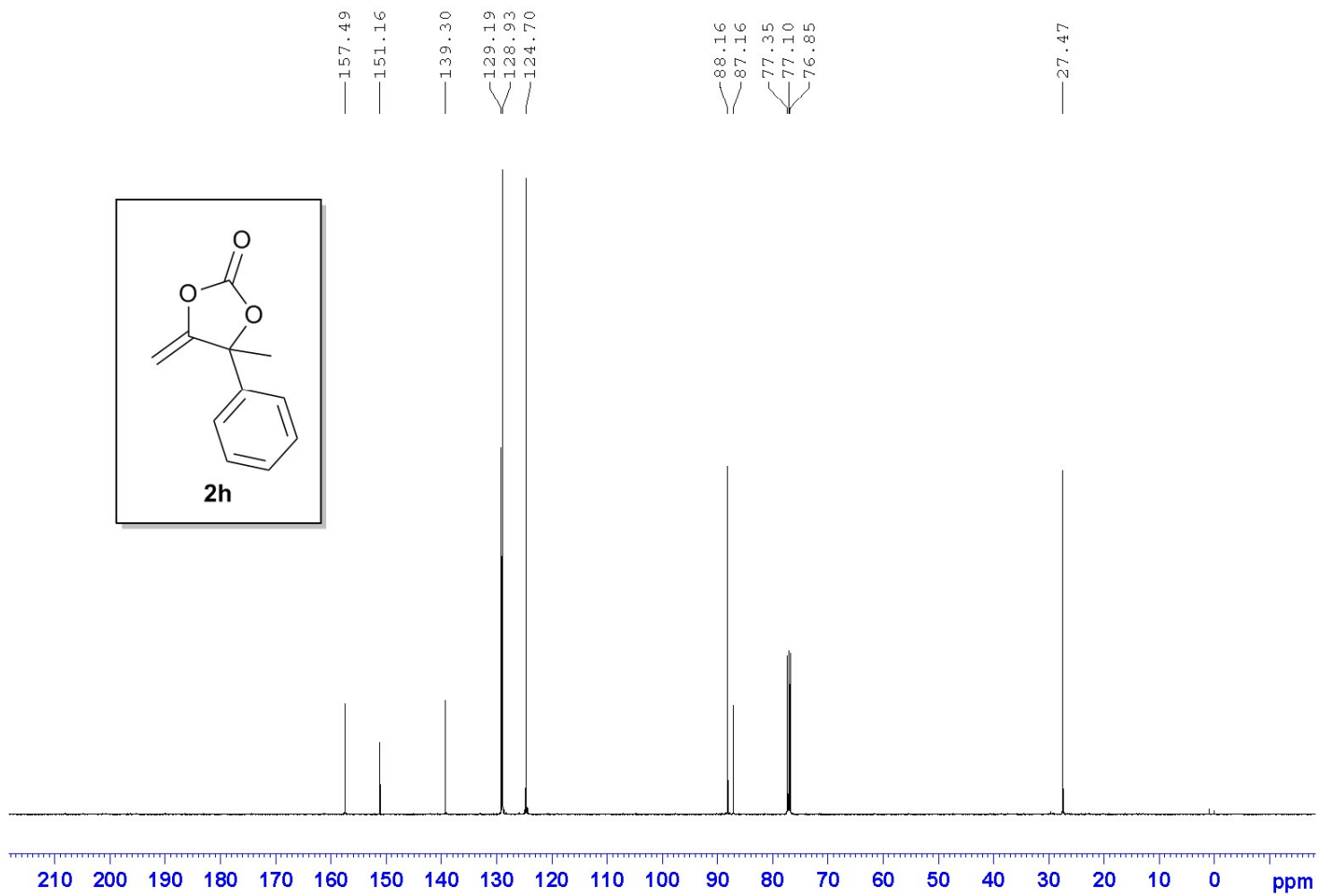
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¹H NMR



¹³C NMR



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