Supplementary Information

An integrated microwell array platform for cell lasing analysis

Qiushu Chen,^a Yu-Cheng Chen,^a Zhizheng Zhang,^b Biming Wu,^a

Rhima Coleman^a and Xudong Fan^{a*}

a. Department of Biomedical Engineering, University of Michigan,

1101 Beal Ave, Ann Arbor, Michigan, 48109, United States

 b. Department of Electrical Engineering and Computer Science, University of Michigan, 1301 Beal Avenue, Ann Arbor, Michigan, 48109, United States

*xsfan@umich.edu



Figure S1. Simulation of the scattered field of the incident Gaussian beam

(a) The simulation geometry. $n_{water}=1.33$. $n_{mirror}=1.45$. A Gaussian beam along y-axis is set as the background field with an FWHM of 60 µm and the focal plane at the top interface between the mirror and water. (b) The distribution of the scattered field when there is no microwell structure (the SU-8 domain is set to be the same as the water domain, $n_{SU-8}=1.33$). (c) The distribution of the scattered field when the microwell is present ($n_{SU-8}=1.60$). The integration of the scattered field energy density over the water domain is 2.45e-19 and 2.60e-19 J/m³ without and with the SU-8 microwell structure, respectively. The discrepancy is less than 10%, thus we conclude that the microwell structure does not have any significant effect on the excitation profile.



Figure S2. Lasing stability of a cell captured in a microwell

Lasing stability of a cell captured in a microwell. The cell was continuously excited for 30 seconds with a pump intensity of 100 μ J/mm² per pulse. The cell underwent up to 600 excitation pulses during the test, about twice the number of pulses that a cell might receive when it was scanned for 6 times in our work. As shown in Figure S2, no wavelength shift is observed in those lasing peaks.

The thermal effect on the microwell-integrated cell lasing array

Given that there is environmental temperature fluctuation during the experiment, the resulting lasing wavelength shift is examined as follows. The dependence of the lasing wavelength λ on

temperature can be written as
$$\frac{\Delta \lambda}{\lambda} = \left(\frac{1}{n_{eff}} \frac{dn_{eff}}{dT} + \frac{1}{L}\frac{dL}{dT}\right)\Delta T$$
, where n_{eff} is the effective refractive $\frac{dL}{dL}$

index, \overline{dT} the thermal-optic coefficient, L the cavity length and \overline{dT} the linear thermal expansion coefficient. A cell laser cavity consists of mainly water, DNAs and proteins, with an effective thermal-optic coefficient estimated to be $-1 \sim 4 \times 10^{-4}$ /°C.¹⁻³ Since the SU-8 layer acts as the spacer for the laser cavity, the linear thermal expansion coefficient of SU-8 (52×10⁻⁶/°C, MicroChem $\frac{1dL}{|\Delta\lambda|} < 0.05\%$

Corp.) is used to estimate $\frac{-\lambda^2}{LdT}$. Thus, for a temperature drift of 1°C, $\left|\frac{\Delta\lambda}{\lambda}\right| < 0.05\%$ and the corresponding lasing wavelength shift $\Delta\lambda$ around 540 nm is about 0.2 nm.

We also examine the thermal effect of the pump laser. At 120 μ J/mm² pump fluency, the total energy impinged on an area of 2×10⁻⁴ mm² (the area of a cell) is 2.4×10⁻⁸ J per excitation pulse. Typically, less than 10% of the total energy can be absorbed (absorption cross section of dyes at excitation wavelength $\sigma_a \sim 1 \times 10^{-16} cm^2$, dye concentration C ~1 mM, gain medium length L~ 10 μ m, thus absorbance A = $\sigma_a CL = 0.06$) and only a fraction that non-radiatively dissipates turns into heat (assume to be 40%, since quantum yield of SYTO9=0.6). The resulting fluctuation in F

$$\Delta T = \frac{E_{heat}}{C_{m}}$$

local temperature is estimated to be around 0.3 °C ($C_p m$, E_{heat} the energy that turns into heat, C_p the specific heat (water: 4.184 J/(g·°C)), m the mass). The lasing wavelength might have a drift of 0.06 nm accordingly. However, no build-up effect is expected under continuous pulse excitation due to the small duty cycle (5 ns/50 ms) of the 20 Hz OPO laser.

References

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