

Electronic Supplementary Information

Dissecting binding of a β -barrel outer membrane protein by phage display

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PCR conditions for generating the ShuA loop deletion or alanine-substitution variants

One of the following DNA templates (~50 ng) was used to generate the ShuA loop deletion or alanine-substituted variants for phage display or protein expression: pEShuA¹⁸, PCR products containing overlapping regions of the ShuA gene with the mutation of interest, or phagemid DNA encoding a ShuA extracellular variant. Platinum DNA Polymerase (Invitrogen) was used according to the manufacturer's specifications with the oligonucleotides listed in Table SI 1 and the following PCR thermal cycling conditions: 1 cycle of 2 min at 98 °C, followed by 30 cycles of 1 min at 98 °C, 1 min at 60 °C, and 2 min at 68 °C, and finishing with 5 min incubation at 68 °C.

Table SI 1. Oligonucleotide sequences

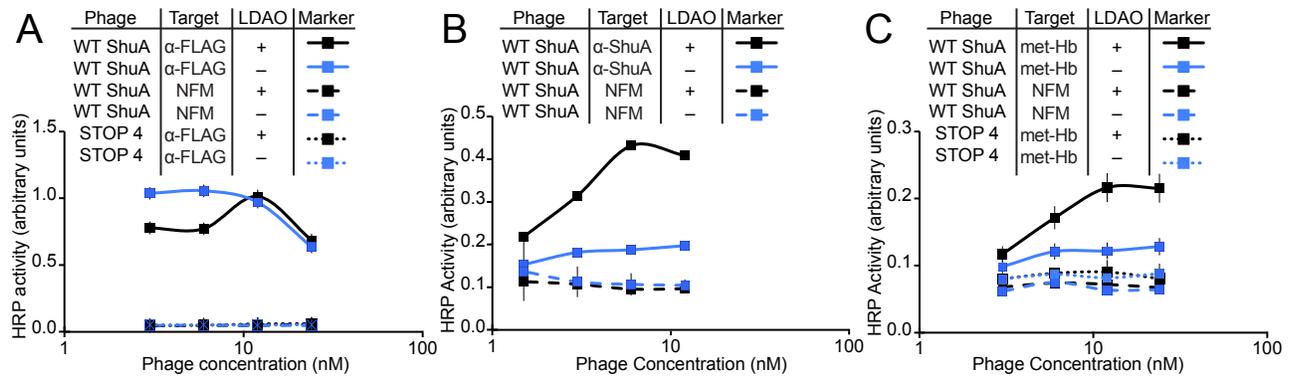
Oligonucleotide Label	DNA Sequence (5' to 3')	Resulting ShuA variant
OL_L1_Sb_F	TCG TGT CTT TGG TAC TGG CGG CAC GGG GGC CGC GAG CCT GGG ATT AGG CGC GAG CGC GTT	Ala-L1
OL_L1_Sb_R	AAC GCG CTC GCG CCT AAT CCC AGG CTC GCG GCC CCC GTG CCG CCA GTA CCA AAG ACA CGA	Ala-L1
OL_L2_Sb_F	TTG TGG CCT GGT CCA GTC GCG ATC GGG GTG ATG CGG CCG CGG CCG CTG CAG CAG CCG CGG CGA ATG ACG AGT CCA TTA ATA ACA TGC T	Ala-L2
OL_L2_Sb_R	AGC ATG TTA TTA ATG GAC TCG TCA TTC GCC GCG GCT GCT GCA GCG GCC GCG GCC GCA TCA CCC CGA TCG CGA CTG GAC CAG GCC ACA A	Ala-L2
OL_L3_Sb_F	TAC AAC AAC GAC GCG CGT GAA CCA AAA AAT GCG GCG GCC GCA GCG GCT GCG GCA GCC GCC AAC CCG ATG GTT GAT CGT TCA ACA ATT CAA	Ala-L3
OL_L3_Sb_R	TTG AAT TGT TGA ACG ATC AAC CAT CGG GTT GGC GGC TGC CGC AGC CGC TGC GGC CGC CGC ATT TTT TGG TTC ACG CGC GTC GTT GTT GTA	Ala-L3
OL_L4_Sb_F	GAA GTC CGT ATT AAT GCG CAA AAC GCA GCG GCA GCC GGC GAG TAT CGT GAA CAG ATA ACA	Ala-L4

OL_L4_Sb_R	TGT TAT CTG TTC ACG ATA CTC GCC GGC TGC	Ala-L4
OL_L5_Sb_F	CGC TGC GTT TTG CGC ATT AAT ACG GAC TTC TGA GTA TTA TCG TCA GGA ACA ACA TCC	Ala-L5
OL_L5_Sb_R	GGG CGG CGC GGC GGC GGC CGC ACC GCA AGC AAA AAT CGA TTT TAG CTC GAG CTA AAA TCG ATT TTT GCT TGC GGT	Ala-L5
OL_L6_Sb_F	GCG GCC GCC GCC GCG CCG CCC GGA TGT TGT TCC TGA CGA TAA TAC TCA	Ala-L6
OL_L6_Sb_R	AGT TAT CGC GGT AGC AGT GAC GGT GCG AAA GAT GTT GAT GCC GAC AAA TGG TCA TCT	Ala-L6
OL_L7A_Sb_F	AGA TGA CCA TTT GTC GGC ATC AAC ATC TTT CGC ACC GTC ACT GCT ACC GCG ATA ACT	Ala-L6
OL_L7A_Sb_R	TGC CCA GGC ATT CCG CGC CCC GAC GGC GGC CGC AGC GGC AGC CGC GGC TGC GGC CGC CGC GGC GGG TCG CTT CTA TAC CAA CTA TTG GGT	Ala-L7A
OL_L7A_Sb_R	ACC CAA TAG TTG GTA TAG AAG CGA CCC GCC GCG GCG GCC GCA GCC GCG GCT GCC GCT GCG GCC GCC GTC GGG GCG CGG AAT GCC TGG GCA	Ala-L7A
OL_L7B_Sb_F	ACG ATT CTA AGC ACT TCT CGA TTG CGG CCG CCG CAG CCG CCG CGG CGG CGG CAG CCG CGG CCT TAC GTC CGG AAA CTA ACG AAA CTC AGG	Ala-L7B
OL_L7B_Sb_R	CCT GAG TTT CGT TAG TTT CCG GAC GTA AGG CCG CGG CTG CCG CCG CCG CGG CGG CTG CGG CGG CCG CAA TCG AGA AGT GCT TAG AAT CGT	Ala-L7B
OL_L8_Sb_F	AAG GAT TAC ATC TCC ACG ACC GTC GAT GCC GCG GCG GCG ACG ACT ATG TCG TAT AAC GT	Ala-L8
OL_L8_Sb_R	ACG TTA TAC GAC ATA GTC GTC GCC GCC GCG GCA TCG ACG GTC GTG GAG ATG TAA TCC TT	Ala-L8
OL_L9_Sb_F	TAA CCG TAC CCG CGG CAA AGA CAC CGA TGC CGC GGC AGC GGC GGC CAG CAT TAA CCC GGA TAC CGT TAC CA	Ala-L9
OL_L9_Sb_R	TGG TAA CGG TAT CCG GGT TAA TGC TGG CCG CCG CTG CCG CGG CAT CGG TGT CTT TGC CGC GGG TAC GGT TA	Ala-L9
OL_L10_Sb_F	TTC TCT GTT GGG TGG GTT GGT ACG TTT GCC GAT CGC GCA GCA GCG GCC GCC GCG GCT GCC GCG GCG GCA CCA GGC TAT GGC GTG AAT GAT	Ala-L10
OL_L10_Sb_R	ATC ATT CAC GCC ATA GCC TGG TGC CGC	Ala-L10

	CGC GGC AGC CGC GGC GGC CGC TGC TGC GCG ATC GGC AAA CGT ACC AAC CCA CCC AAC AGA GAA	
OL_L11_Sb_F	ACT ACT TTG GTG TTG GGT AAC GCT GCC GCC GCG GCG GCC GCG GCG GCG GCA GCC GCC GCA GCG GCT GGT CGT AAC GGA AAA ATT TTC GTG	Ala-L11
OL_L11_Sb_R	CAC GAA AAT TTT TCC GTT ACG ACC AGC CGC TGC GGC GGC TGC CGC CGC CGC GGC CGC CGC GGC GGC AGC GTT ACC CAA CAC CAA AGT AGT	Ala-L11
OL_L1_Del_F	T CGT GTC TTT GGT ACT GGC GGC ACG GGG AGC CTG GGA TTA GGC GCG AGC GCG TTT	ΔL1
OL_L1_Del_R	AAA CGC GCT CGC GCC TAA TCC CAG GCT CCC CGT GCC GCC AGT ACC AAA GAC ACG AA	ΔL1
OL_L2_Del_F	TTG TGG CCT GGT CCA GTC GCG ATC GGG GTG ATA ATG ACG AGT CCA TTA ATA ACA TGC T	ΔL2
OL_L2_Del_R	AGC ATG TTA TTA ATG GAC TCG TCA TTA TCA CCC CGA TCG CGA CTG GAC CAG GCC ACA A	ΔL2
OL_L3_Del_F	TAC AAC AAC GAC GCG CGT GAA CCA AAA AAT AAC CCG ATG GTT GAT CGT TCA ACA ATT CAA	ΔL3
OL_L3_Del_R	TTG AAT TGT TGA ACG ATC AAC CAT CGG GTT ATT TTT TGG TTC ACG CGC GTC GTT GTT GTA	ΔL3
OL_L4_Del_F	TTG GTC GGA AGT CCG TAT TAA TGC GCA AAA CGG CGA GTA TCG TGA ACA GAT AAC A	ΔL4
OL_L4_Del_R	TGT TAT CTG TTC ACG ATA CTC GCC GTT TTG CGC ATT AAT ACG GAC TTC CGA CCA A	ΔL4
OL_L5_Del_F	TGA GTA TTA TCG TCA GGA ACA ACA TCC GGG CGG CCC GCA AGC AAA AAT C	ΔL5
OL_L5_Del_R	GAG CTA AAA TCG ATT TTT GCT TGC GG G CCG CCC GGA TGT TGT TCC TGA CGA	ΔL5
OL_L6_Del_F	GAC AGT TAT CGC GGT AGC AGT GAC GGT AAA GAT GTT GAT GCC GAC AAA TGG TCA TCT CGT	ΔL6
OL_L6_Del_R	ACG AGA TGA CCA TTT GTC GGC ATC AAC ATC TTT ACC GTC ACT GCT ACC GCG ATA ACT GTC	ΔL6
OL_L7A_Del_F	TAT TTG GCT CAT ATG CCC AGG CAT TCC GCG GTC GCT TCT ATA CCA ACT ATT GGG TGC CA	ΔL7A
OL_L7A_Del_R	TGG CAC CCA ATA GTT GGT ATA GAA GCG ACC GCG GAA TGC CTG GGC ATA TGA GCC AAA TA	ΔL7A

OL_L7B_Del_F	TG TAT AAC GAT TCT AAG CAC TTC TCG ATT GAA ACT AAC GAA ACT CAG GAG TAC GGT TT	ΔL7B
OL_L7B_Del_R	AAA CCG TAC TCC TGA GTT TCG TTA GTT TCA ATC GAG AAG TGC TTA GAA TCG TTA TAC A	ΔL7B
OL_L8_Del_F	AAG GAT TAC ATC TCC ACG ACC GTC GAT GCG GCG GCG ACG ACT ATG TCG TAT AAC GT	ΔL8
OL_L8_Del_R	ACG TTA TAC GAC ATA GTC GTC GCC GCC GCA TCG ACG GTC GTG GAG ATG TAA TCC TT	ΔL8
OL_L9_Del_F	TAA CCG TAC CCG CGG CAA AGA CAC CGA TAG CAT TAA CCC GGA TAC CGT TAC CA	ΔL9
OL_L9_Del_R	TGG TAA CGG TAT CCG GGT TAA TGC TAT CGG TGT CTT TGC CGC GGG TAC GGT TA	ΔL9
OL_L10_Del_F	TTC TCT GTT GGG TGG GTT GGT ACG TTT GCC GAT CGC CCA GGC TAT GGC GTG AAT GAT T	ΔL10
OL_L10_Del_R	AAT CAT TCA CGC CAT AGC CTG GGC GAT CGG CAA ACG TAC CAA CCC ACC CAA CAG AGA A	ΔL10
OL_L11_Del_F	ATG ACC ACT ACT TTG GTG TTG GGT AAC GCT GGT CGT AAC GGA AAA ATT TTC GTG AGT	ΔL11
OL_L11_Del_R	ACT CAC GAA AAT TTT TCC GTT ACG ACC AGC GTT ACC CAA CAC CAA AGT AGT GGT CAT	ΔL11
pM1155a_NsiI_F	AGC TTC ATG CAT GCG ATT ACA AGG ATG ACG ACG AT	For sub-cloning into the phage display vector
pM1155a_NcoI_R	ATC CTC CAC CAC TAG TAC CAT GGT ACC AT	For sub-cloning into the phage display vector
pET22b_MscI_F	5'-AGC CTG TGG CCA TGG CTA CTG AAA CCA TGA CCG TTA CGG CAA	For sub-cloning into the protein expression vector
pET22b_XhoI_R	5'-TTG GCT CTC GAG CCA TTG ATA ACT CAC GAA AAT TTT TCC GTT ACG A	For sub-cloning into the protein expression vector

*F and R designate forward and reverse primers, respectively.



SI Figure 1. Functional phage-displayed wild-type ShuA requires the detergent LDAO.

Phage-displayed wild-type ShuA or STOP4 (negative control phage) were incubated in the presence and absence of the detergent LDAO with immobilized **A**) anti-FLAG antibody (α -FLAG), **B**) anti-ShuA antibody (α -ShuA), or **C**) met-hemoglobin (met-Hb). Relative levels of the bound phage-displayed ShuA variants were quantified by anti-M13 antibody conjugated to HRP. Throughout this report, each data point represents the average of three replicates, and error bars indicate standard deviation around the mean.

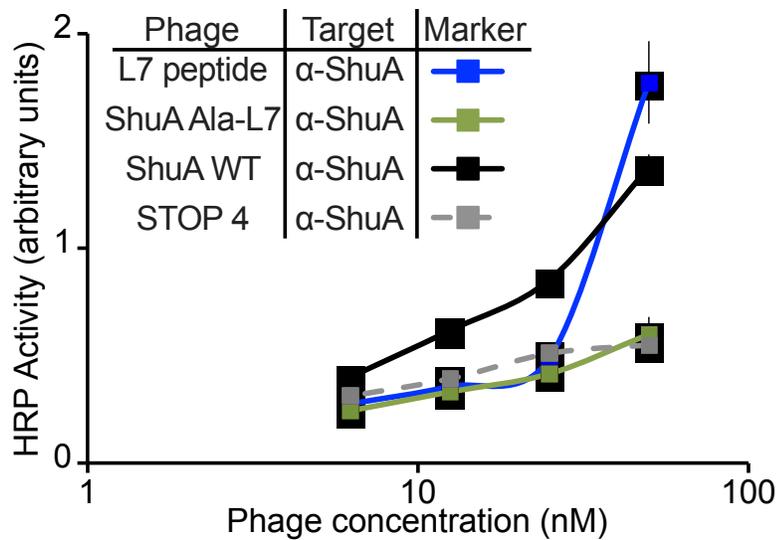


Figure SI 2. Phage-based ELISAs of a phage-displayed peptide, ShuA Loop 7. Phage-displayed ShuA L7 peptide, full-length ShuA alanine-substituted L7 variant (Ala-L7), wild-type ShuA (positive control phage), or STOP4 (negative control phage) were incubated with immobilized anti-ShuA antibody coated on microtiter plate wells. Relative levels of the bound phage-displayed ShuA variants and peptide were quantified by anti-M13 antibody conjugated to HRP. No binding was observed for phage-displayed ShuA Ala-L7 to the anti-ShuA antibody, but the phage-displayed peptide ShuA Loop 7 bound the immobilized target. Throughout this report, each data point represents the average of three replicates, and error bars indicate standard deviation around the mean.

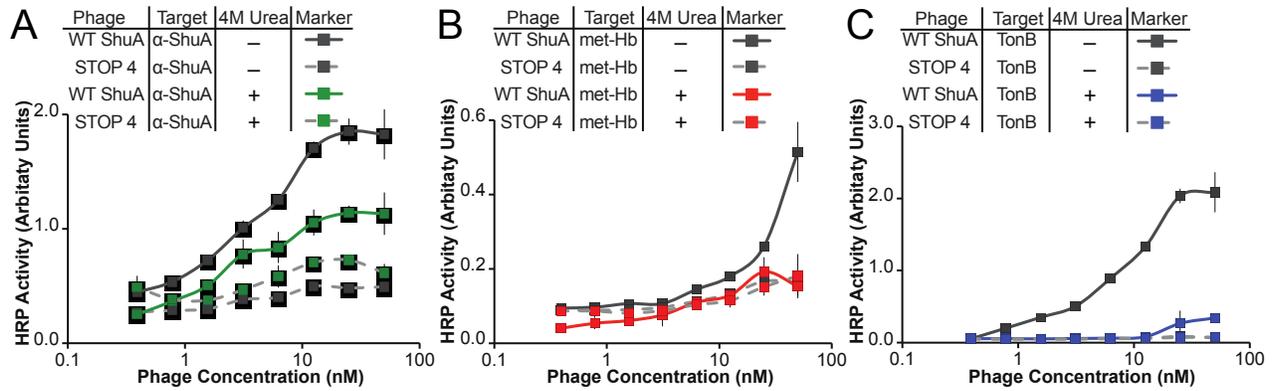


Figure SI 3. Phage-based ELISAs of phage-displayed wild-type ShuA treated with 4M urea.

Phage-displayed wild-type ShuA and STOP4 (negative control phage) were assayed for binding in the presence or absence of 4M urea. Following treatment with urea, the phage-displayed wild-type ShuA or STOP4 were purified from excess urea through a second PEG precipitation and incubated with the following immobilized targets: **A)** anti-ShuA antibody, **B)** met-Hemoglobin, or **C)** TonB. Relative levels of the bound phage-displayed ShuA were quantified by anti-M13 antibody conjugated to HRP. Phage-displayed wild-type ShuA treated with 4 M urea did not bind to the met-hemoglobin or TonB protein. However, binding is observed by urea-denatured, phage-displayed wild-type ShuA to the immobilized anti-ShuA antibody. Throughout this report, each data point represents the average of three replicates, and error bars indicate standard deviation around the mean.

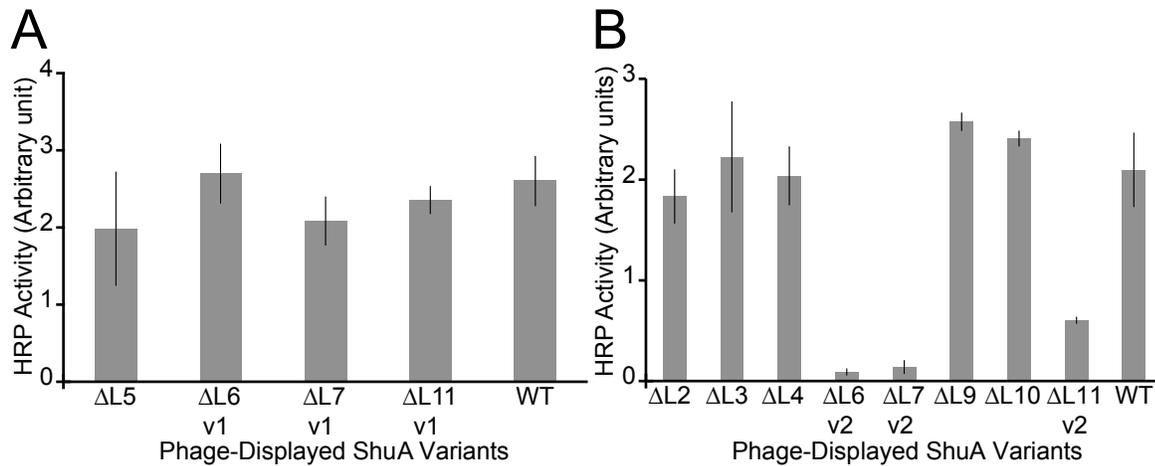


Figure SI 4. Phage-based ELISAs of ShuA extracellular loop deletion variants. To evaluate display levels, 50 nM of phage-displayed ShuA extracellular loop deletion variants, $\Delta L2$ through $\Delta L11$, or wild-type ShuA (positive control) were assayed for binding to immobilized anti-FLAG antibody, which can recognize a FLAG epitope fused to the N-terminus of the ShuA variants. **A)** Phage-displayed ShuA $\Delta L5$, $\Delta L6v1$, $\Delta L7v1$, and $\Delta L11v1$ demonstrated display levels similar to wild-type ShuA (WT) **B)** No or low display levels were observed for ShuA $\Delta L6v2$, $\Delta L7v2$, and $\Delta L11v2$ when one or two native amino acid residue(s) were removed as compared to wild-type ShuA (WT) and its loop deletion variants, ShuA $\Delta L2$, $\Delta L3$, $\Delta L4$, $\Delta L9$, and $\Delta L10$. Thus, this report focuses on the loop deletion variants that demonstrated similar display levels.

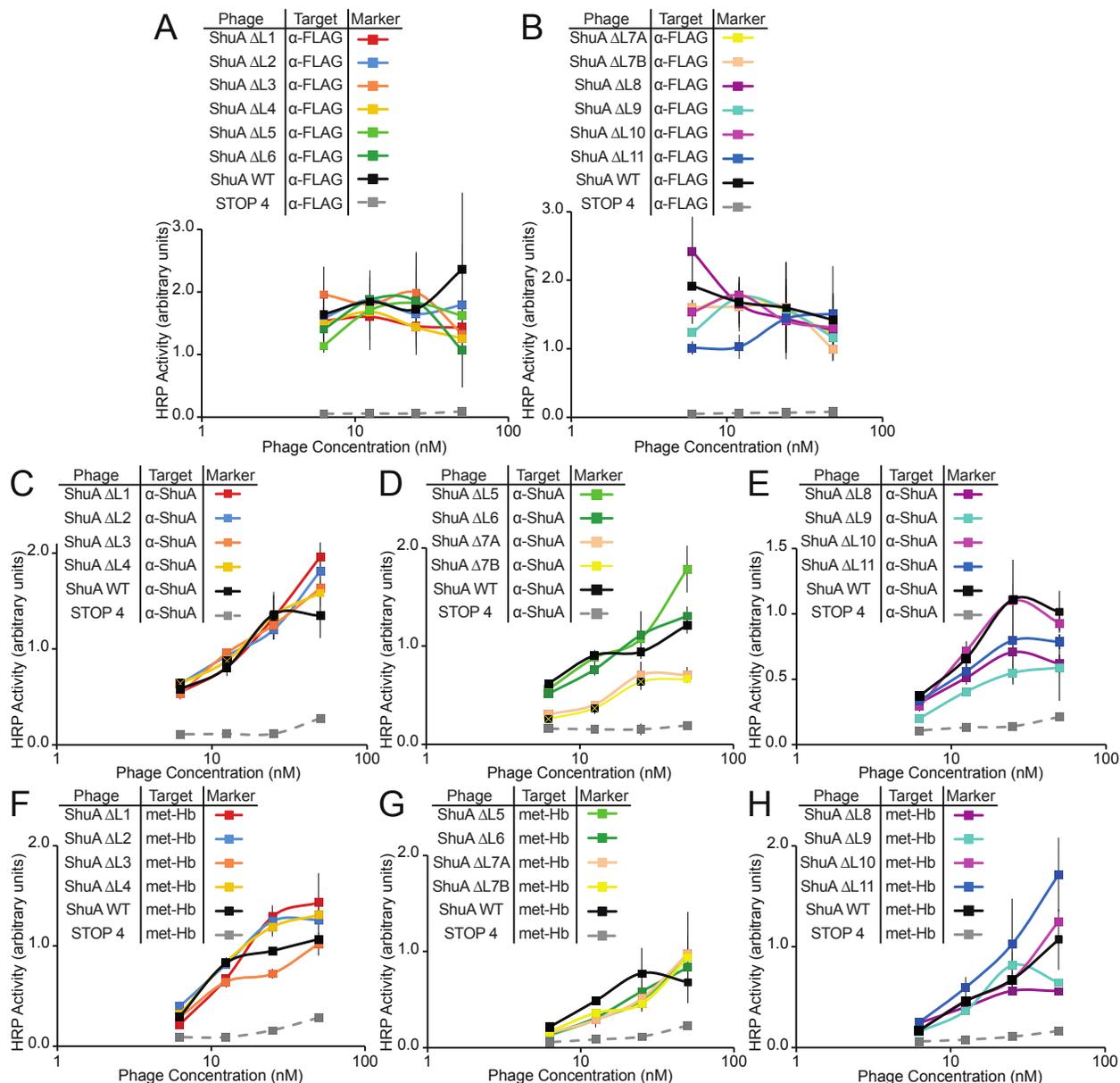


Figure SI 5. Functional ShuA extracellular loop deletion variants displayed on the phage surface. Phage-displayed ShuA loop deletion variants, Δ L1 through Δ L11, wild-type ShuA (positive control phage) or STOP4 (negative control phage) were incubated with immobilized **A** and **B**) anti-FLAG antibody (α -FLAG), **C**, **D**, and **E**) anti-ShuA antibody (α -ShuA), or **F**, **G**, **H**) methemoglobin (metHb). Relative levels of the bound phage-displayed ShuA variants were quantified by anti-M13 antibody conjugated to HRP. Throughout this report, each data point represents the average of three replicates, and error bars indicate standard deviation around the mean.

Figure SI 6. SDS-PAGE analysis of TonB.

Eluted fractions of a 92 residue C-terminal fragment of TonB (residue 142 – 239) from *S. dysenteriae* were purified by size exclusion chromatography, and visually examined in a 12% Tris-glycine SDS-PAGE gel stained by Coomassie blue. **Lane L.** PageRuler Plus pre-stained protein ladder (ThermoFisher Scientific). **Lanes 1 – 9.** Samples (15 μ L) of the 2 mL fractions were collected through size exclusion chromatography. This purified TonB protein was coated on microtiter plates for TonB binding ELISAs with phage-displayed and detergent-solubilized ShuA variants, using conditions described above.

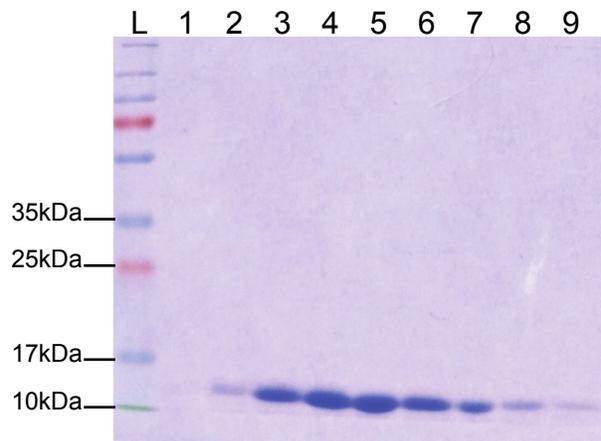


Figure SI 7. SDS-PAGE analysis of met-hemoglobin. A

sample of met-hemoglobin purified from human red blood cells was visually examined in a 12% Tris-glycine SDS-PAGE gel stained by Coomassie blue. **Lane L.** PageRuler Plus pre-stained protein ladder **Lane 2.** Sample (5 μ L) of 1 mg/mL met-hemoglobin. This protein sample was coated on microtiter plates for hemoglobin binding ELISAs with phage-displayed and detergent-solubilized ShuA variants, using conditions described above.

