SUPPLEMENTAL INFORMATION

MyD88 in antigen-presenting cells is not required for CD4+

T-cell responses during peptide nanofiber vaccination

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Figure S1





E α **Q11 nanofibers and uptake by DCs and macrophages**. Peptides (1.33 mM E α Q11 and 0.67 mM Q11) were allowed to assemble at a final concentration of 2 mM in PBS. Nanofibers were diluted in PBS to a final concentration of 0.2 mM for TEM (scale bar: 200 nm). Like its OVAQ11 counterpart, E α Q11 was able to form nanofibers with Q11 (a). When injected intraperitoneally, E α Q11 nanofibers were internalized by both DCs and macrophages (b and c) when analyzed 24 h post administration, comparable to OVAQ11 uptake (as shown in Figure 2).

Figure S2



Deletion of *MyD88*^{fl} **allele in CD11c⁺ DCs.** DCs were sorted from CD11c-MyD88 KO mice based on CD11c expression, and MyD88 was measured by quantitative real-time (qRT) PCR on genomic DNA. Higher threshold cycle numbers indicate lower MyD88 levels.



Enrichment of OTII cells. The percentage of CD4⁺ T cells from OTII mice was ~95%, with <1% CD11c⁺ DCs, following enrichment using a CD4⁺ T cell enrichment kit.



T cell enrichment for adoptive transfer. The percentage of CD3⁺ T cells from WT or MyD88 KO mice was ~98%, following enrichment using a Pan T Cell Isolation Kit II.

Figure S5



Trif and IFN $_{\alpha}$ **R are not required for antibody responses**. Mice were immunized (day 0) and boosted (day 28) with OVAQ11 nanofibers subcutaneously. IgG titer in sera was analyzed by ELISA 1 week after boost (day 35). Data shown were from one representative study and analyzed by t test. ns: not significant.

Figure S6



OVAQ11 nanofibers do not induce the production of IL-6 and IL-12 by BMDCs in vitro. FIt-3L-induced BMDCs were treated with 0.1 mM OVAQ11 nanofibers overnight, and concentrations of IL-6 and IL-12 in the supernatant were measured by ELISA. CpG was used as a positive control. ***p>0.001 by one-way ANOVA with Tukey's post-hoc test.