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Evaluation of affibody charge modification identified by synthetic consensus design in molecular PET imaging of epidermal growth factor receptor

Brett A. Case, Max A. Kruziki, Lawrence A. Stern, and Benjamin J. Hackel Department of Chemical Engineering and Materials Science University of Minnesota – Twin Cities Minneapolis, MN

Supplementary Figures and Tables

Variant	Yield (mg/L)	K _d (nM)	T _m (°C)	2° Structure	Specificity (A431:MCF7 Binding Signals)
EA68	3.5 ± 0.3	5.3 ± 1.7	71	α-helical	109 ± 63
EA26S	0.62 ± 0.02	1.6 ± 1.2	68	α-helical	214 ± 134
EA62S	1.9 ± 0.3	2.5 ± 0.7	59	α-helical	342 ± 209
EA35S	7.0 ± 0.5	1.7 ± 0.5	68	α-helical	529 ± 297
EA35C	12.7 ± 0.9	6.9 ± 1.4	71	α-helical	74 ± 29

Supplemental Table 1. Biophysical properties of affibody clones used for PET/CT imaging.



Supplemental Figure 1. Matrix-assisted laser desorption ionization mass spectrometry (MALDI-MS) of NODAGA-Affibody charge variants with 0, 1, 2, or 3 degrees of conjugation



Supplemental Figure 2. *NODAGA-Affibody target-affinity characterization.* EGFR-overexpressing A431 cells were mixed with the indicated concentration of NODAGA-conjugated affibody. Binding was detected with fluorophore-conjugated anti-His6 antibody via flow cytometry. Equilibrium dissociation constants were calculated assuming a 1:1 binding model. n = 3 measurements per variant. NODAGA-conjugated versions of EA35S and EA26 were nominally weaker binders to EGFR than their unconjugated counterparts (K_d = 3.9 ± 0.4 nM vs. 1.7 ± 0.5 nM and 4.6 ± 0.3 vs. 1.6 ± 1.2 nM respectively). However, these variances are slight and should not meaningfully impact *in vivo* biomarker localization. EA68 and EA62 showed no difference in binding affinity upon NODAGA conjugation and EA35C presented a slight improvement (4.3 ± 0.3 nM vs. 6.9 ± 1.4 nM).



Supplemental Figure 3. Matrix-assisted laser desorption ionization mass spectrometry (MALDI-MS) of DOTA-Affibody charge variants with 0, 1, 2, or 3 degrees of conjugation



Supplemental Figure 4. Radio chromatography of affibody clones left untreated, in serum, or in trypsin.